

# Probabilistic mapping of the sub-cellular proteome

Slides available at: <http://bit.ly/ABLS2020>

**Laurent Gatto**  
13 February 2020

**Abstract:** In biology, localisation is function - understanding the sub-cellular localisation of proteins is paramount to comprehend the context and full extend of their functions. Shotgun mass spectrometry-based spatial proteomics method are orthogonal to widely used targeted microscopy-based assay. In conjunction with contemporary machine learning, the former enable to build proteome-wide protein localisation maps, informing us on the location of thousands of proteins. When studying these proteome-wide spatial maps, one can learn that while some proteins can be found in a single location within a cell, up to half of the proteins may reside in multiple locations, can dynamically re-localise, or reside within an unknown functional compartment, leading to considerable uncertainty in associating proteins to their sub-cellular location. Recent Bayesian modelling approaches enable us to mine these data, and in particular the dynamic fraction of the spatial proteome, in much greater depth. We are now in a position to (1) probabilistically model protein localisation as well as quantify the uncertainty in the location assignments, and (2) compute a probability for, and quantify uncertainty in, whether a protein is differentially localised upon cellular perturbation. These computational approaches lead to better and more trustworthy biological interpretation of these rich spatial proteomics data.

[Dr Lisa Breckels](#), Cambridge: novelty detection, transfer learning, pRoloc and pRolocdata.

[Mr Oliver Crook](#), Cambridge: Bayesian spatial proteomics, pRoloc and pRolocdata.

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# Outline

Spatial proteomics

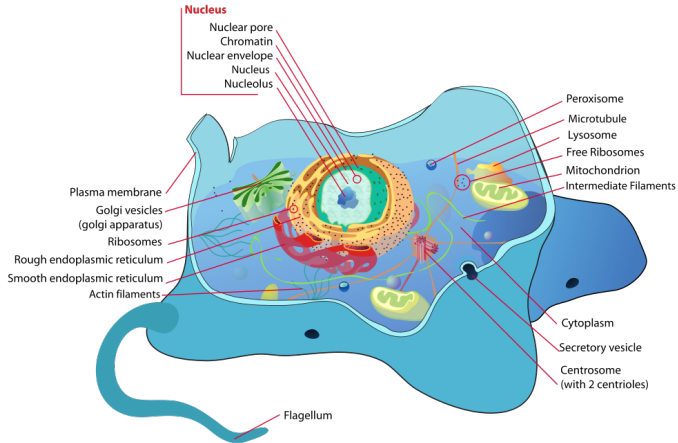
Data analysis (1)

Computational spatial proteomics (2)

Behind the scenes

Conclusions

# Cell organisation - localisation is function



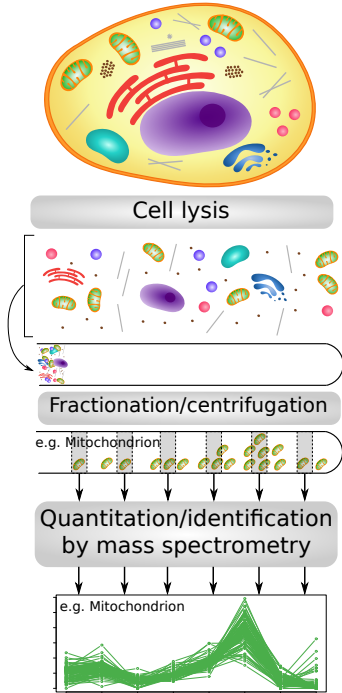
**Spatial proteomics** is the systematic study of protein localisations.

**Localisation – re-localisation – mis-localisation**

Image from Wikipedia [http://en.wikipedia.org/wiki/Cell\\_\(biology\)](http://en.wikipedia.org/wiki/Cell_(biology)).

**Explorative/discovery approaches, steady-state global localisation maps** (as opposed to microscopy-based targeted approaches).

**Density gradient:** PCP (Dunkley et al., 2006), LOPIT (Foster et al., 2006), hyperLOPIT (Christoforou et al., 2016; Mulvey et al., 2017) and **Differential centrifugation** Itzhak et al. (2016), LOPIT-DC (Geladaki et al., 2019).



	Fraction <sub>1</sub>	Fraction <sub>2</sub>	...	Fraction <sub>L</sub>
<b>x</b> <sub>1</sub>	x <sub>1,1</sub>	x <sub>1,2</sub>	...	x <sub>1,L</sub>
<b>x</b> <sub>2</sub>	x <sub>2,1</sub>	x <sub>2,2</sub>	...	x <sub>2,L</sub>
<b>x</b> <sub>3</sub>	x <sub>3,1</sub>	x <sub>3,2</sub>	...	x <sub>3,L</sub>
⋮	⋮	⋮	⋮	⋮
<b>x</b> <sub>i</sub>	x <sub>i,1</sub>	x <sub>i,2</sub>	...	x <sub>i,L</sub>
⋮	⋮	⋮	⋮	⋮
<b>x</b> <sub>N</sub>	x <sub>N,1</sub>	x <sub>N,2</sub>	...	x <sub>N, L</sub>

# Quantitation data and organelle markers

	Fraction <sub>1</sub>	Fraction <sub>2</sub>	...	Fraction <sub>L</sub>	markers
<b>x<sub>1</sub></b>	x <sub>1,1</sub>	x <sub>1,2</sub>	...	x <sub>1,L</sub>	unknown
<b>x<sub>2</sub></b>	x <sub>2,1</sub>	x <sub>2,2</sub>	...	x <sub>2,L</sub>	<i>loc<sub>1</sub></i>
<b>x<sub>3</sub></b>	x <sub>3,1</sub>	x <sub>3,2</sub>	...	x <sub>3,L</sub>	unknown
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# Outline

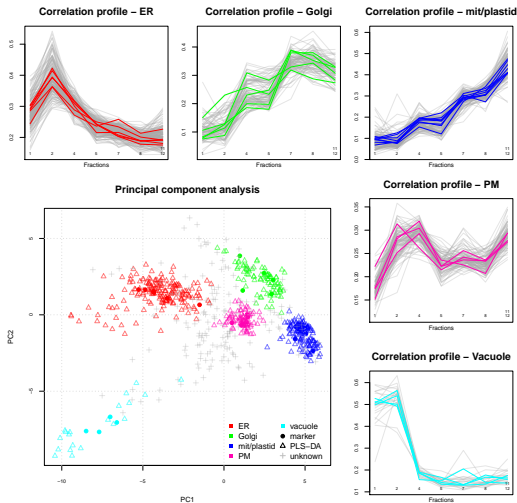
Spatial proteomics

Data analysis (1)

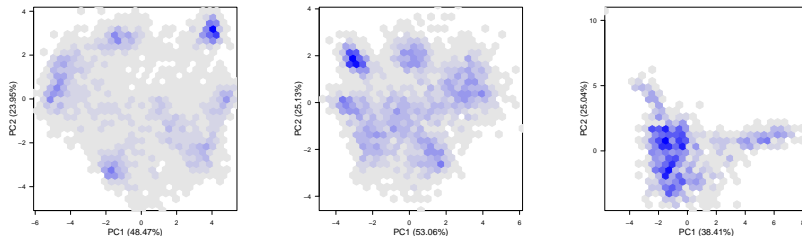
Computational spatial proteomics (2)

Behind the scenes

Conclusions

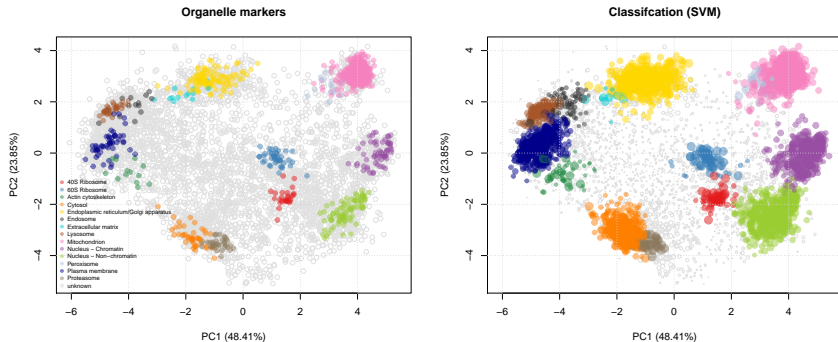


**Figure:** From Gatto et al. (2010), *Arabidopsis thaliana* data from Dunkley et al. (2006)



**Figure:** Assessing sub-cellular resolution in spatial proteomics experiments (Gatto et al., 2018)

# Problem statement: classification



**Figure:** Support vector machines classifier (after 5% FDR classification cutoff) on the embryonic stem cell data from Christoforou et al. (2016).

- Visualisation (cluster, unsupervised learning)
- Classification (supervised learning)
- **Novelty detection** (semi-supervised learning)
- Data integration (transfer learning)
- **Unvertainty quantification**
- **Multi-localisation**
- **Spatial dynamics**

To uncover and understand biology

# Outline

Spatial proteomics

Data analysis (1)

Computational spatial proteomics (2)

- Novelty detection

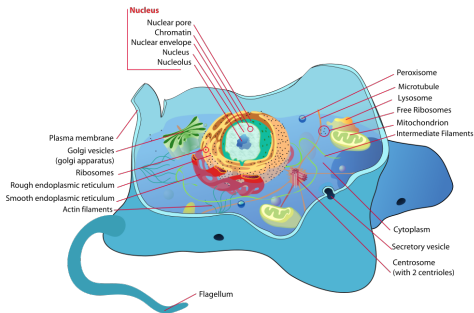
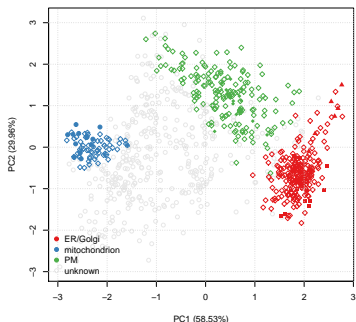
- Multi-localisation and uncertainly quantification

- Spatial dynamics

Behind the scences

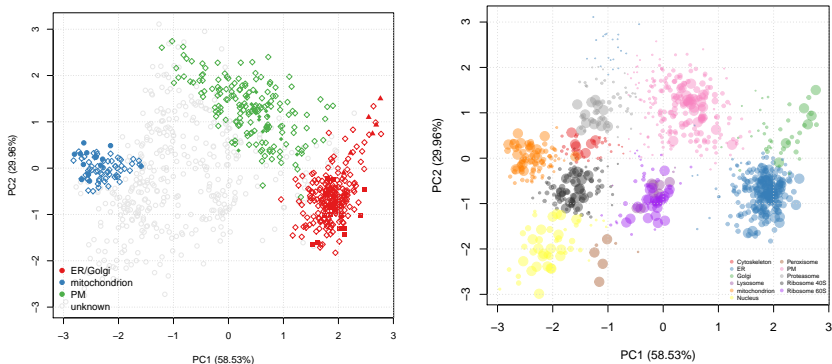
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# Importance of annotation



Incomplete annotation, and therefore lack of training data, for many/most organelles. *Drosophila* data from Tan et al. (2009).

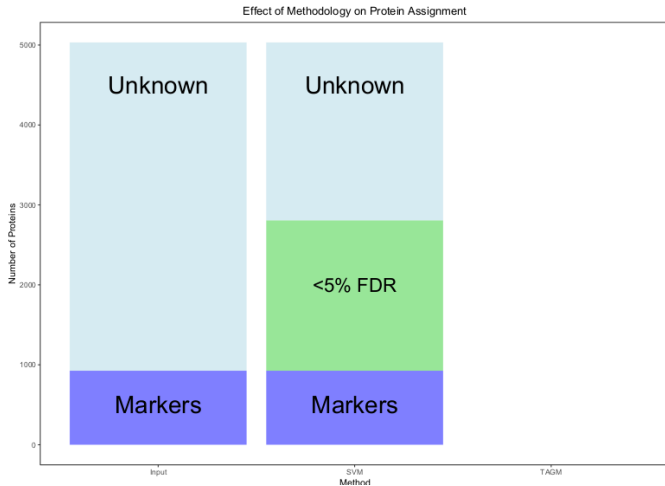
# Semi-supervised learning: novelty detection



**Figure:** Left: Original *Drosophila* data from Tan et al. (2009). Right: After semi-supervised learning and classification, Breckels et al. (2013). Under development: Bayesian novelty detection (Novelty-TAGM).



# How much do we learn? How much do we miss?



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- With the prior knowledge that many proteins are not captured by known sub-cellular niches, we augment our model with an **outlier component**. Outliers are often dispersed and thus this additional component is described by a heavy-tailed distribution: the multivariate Student's t-distribution, leading us to a *T Augmented Gaussian Mixture model* (Crook et al., 2018, 2019b).

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- This methodology allows proteome-wide **uncertainty quantification**, thus adding a further layer to the analysis of spatial proteomics.

We initially model the distribution of profiles associated with proteins that localise to the  $k$ -th component as multivariate normal with mean vector  $\boldsymbol{\mu}_k$  and covariance matrix  $\boldsymbol{\Sigma}_k$ , so that:

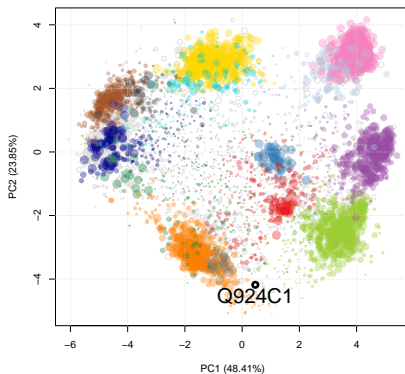
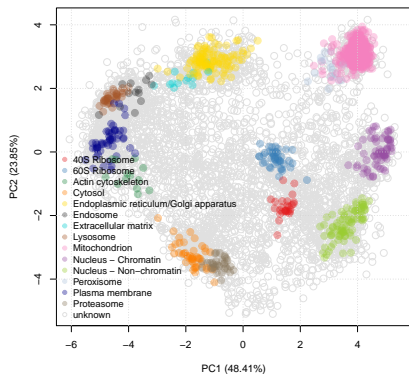
$$\mathbf{x}_i | z_i = k \sim \mathcal{N}(\boldsymbol{\mu}_k, \boldsymbol{\Sigma}_k) \quad (1)$$

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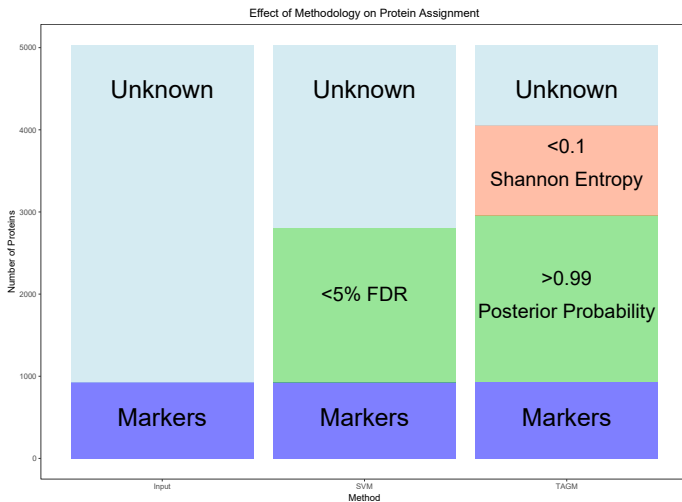
$$\mathbf{x}_i | z_i = k \sim \mathcal{N}(\boldsymbol{\mu}_k, \boldsymbol{\Sigma}_k) \quad (1)$$

We extend it by introducing an additional *outlier component*. To do this, we augment our model by introducing a further indicator latent variable  $\phi$ . Each protein  $\mathbf{x}_i$  is now described by an additional variable  $\phi_i$ , with  $\phi_i = 1$  indicating that protein  $\mathbf{x}_i$  belongs to a organelle derived component and  $\phi_i = 0$  indicating that protein  $\mathbf{x}_i$  is not well described by these known components. This outlier component is modelled as a multivariate T distribution with degrees of freedom  $\kappa$ , mean vector  $\mathbf{M}$ , and scale matrix  $V$ .

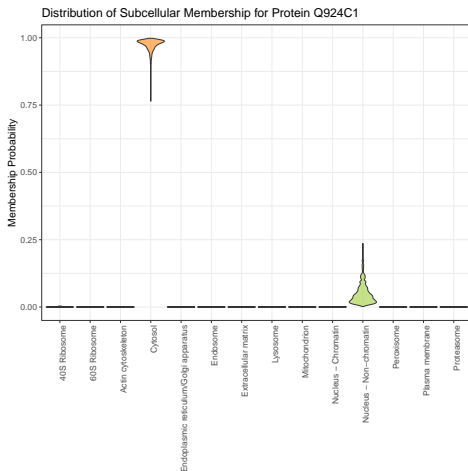
$$\mathbf{x}_i | z_i = k, \phi_i \sim \mathcal{N}(\boldsymbol{\mu}_k, \boldsymbol{\Sigma}_k)^{\phi_i} \mathcal{T}(\kappa, \mathbf{M}, V)^{1-\phi_i} \quad (2)$$



**Figure:** Assignment of proteins of *unknown* location to one of the annotated classes. The dots are scaled according to the protein assignment probabilities.

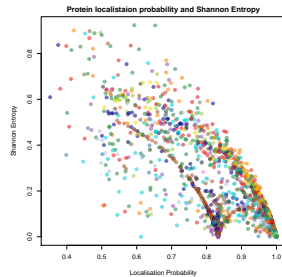
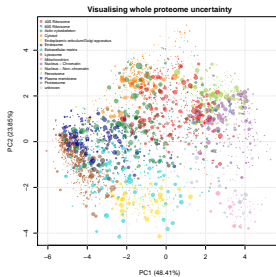
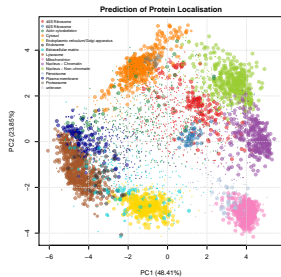




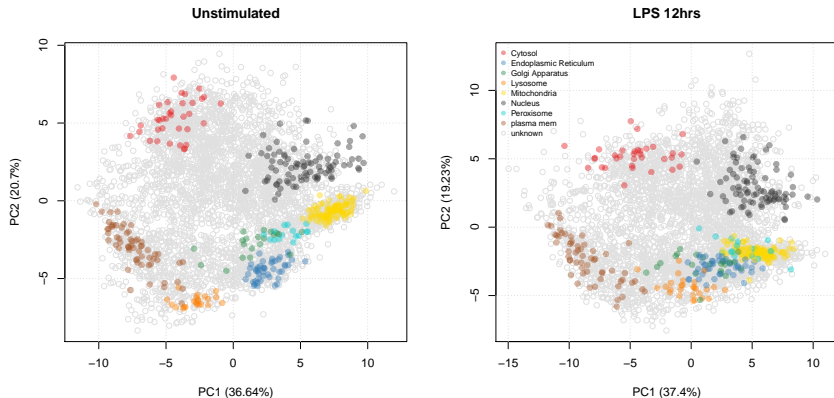


**Figure:** Exportin 5 (Q924C1) forms part of the micro-RNA export machinery, transporting miRNA from the nucleus to the cytoplasm for further processing. It then translocates back through the nuclear pore complex to return to the nucleus to mediate further transport between nucleus and cytoplasm. The model correctly infers that it most likely localises to the cytosol but there is some uncertainty with this assignment. This uncertainty is reflected in possible assignment of Exportin 5 to the nucleus non-chromatin and reflects the multi-location of the protein.

# Whole sub-cellular proteome uncertainty



# Spatial dynamics



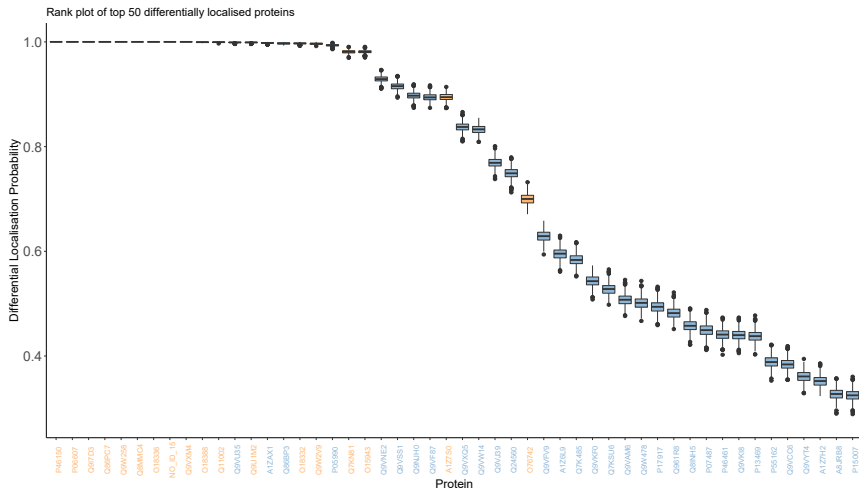
**Figure:** Spatial maps of unstimulated (**control**) and LPS-treated (**experimental condition**) cells (combined triplicates).

A probabilistic definition of differential localisation:

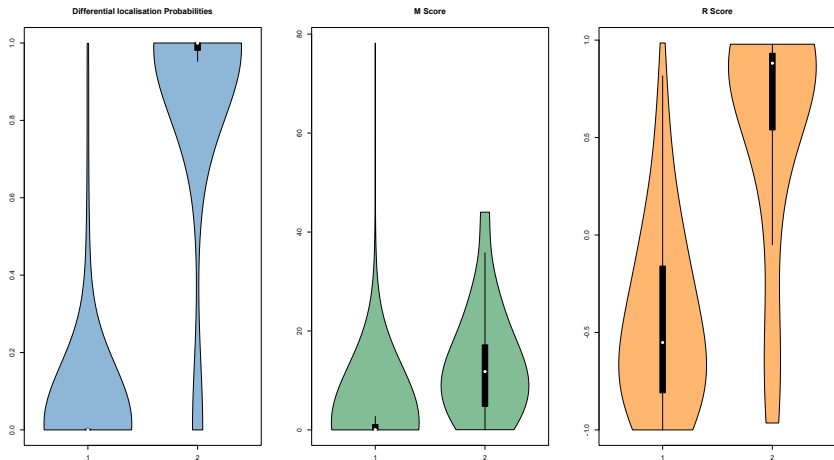
$$x_i = p(z_{i,1} \neq z_{i,2}) \quad (3)$$

with

- organelle-specific profiles modelled with mixtures of non-paramateric distributions (Crook et al., 2019a);
- explicit modelling of replicates and their variability;
- no assumption with regard to similarity of gradients between conditions;
- rigorous interpretation of the results with uncertainty quantification of differential localisation.



**Figure:** Proteins ranked based on their probability of being differentially localised, i.e. having been assigned different niches in the control and experimental condition.



**Figure: Differential localisation probabilities (left)** provide excellent discrimination between static (1) and differentially localised proteins (2).

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**Behind the scenes:** software/data structures and  
open research practice.



## Beyond the figures<sup>1</sup>

- Software: **infrastructure** (MSnbase, Gatto and Lilley (2012)), **dedicated machine learning** (pRoloc, Gatto et al. (2014)), **interactive visualisation**<sup>2</sup> (pRolocGUI, Breckels et al. (2017)) and **data** (pRolocdata, Gatto et al. (2014)) for spatial proteomics.
- The **Bioconductor** (Huber et al., 2015) ecosystem for high throughput biology data analysis and comprehension: **open source**, and **coordinated and collaborative**<sup>3</sup> **open development**, enabling **reproducible research**, enables understanding of the data (not a black box) and **drive scientific innovation**.

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<sup>1</sup>... which are all reproducible, by the way.

<sup>2</sup><https://lgatto.shinyapps.io/christoforou2015/>

<sup>3</sup>between and within domains/software

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## Applied Bioinformatics in Life Sciences.

- **Life Sciences:** Protein sub-cellular localisation, technologies (hyperLOPIT) and opportunities.
- **Bioinformatics:** Reliance on computational biology, statistics and dedicated software (pRoLoc *et al.*) to interpret data and acquire biological knowledge.
- Rigorous computational infrastructure and sound data analysis and interpretation is a **long term investment**.

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**Thank you for your attention**

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