PAPILLOMAVIRUSES AND CANCER: FROM BASIC STUDIES TO CLINICAL **APPLICATION**

Harald zur Hausen

Links between human papillomaviruses (HPVs) and cervical cancer were first suspected almost 30 years ago. DNA of specific HPV types has since been found in almost all cervical cancer biopsies. HPV oncogenes that are expressed in these cells are involved in their transformation and immortalization, and are required for the progression towards malignancy. Epidemiological studies have underlined that HPVs are the main aetiological factor for cervical cancer. But how has this knowledge been translated into the clinic to allow the prevention, screening and treatment of cervical cancer?

KOILOCYTE A papillomavirus particleproducing cell that acquires an 'owl-eye' shape due to the shrinkage of the nucleus, and a translucent halo that surrounds the nucleus

DYSPLASIA An early stage in cancer progression, which is characterized by increased cell proliferation and architectural disarray at the tissue level.

LARYNGEAL PAPILLOMA A benign tumour of the larynx.

Deutsches Krebsforschungszentrum, Im Neuenheimer Feld 280, 69120 Heidelberg, Germany. e-mail: zurhausen@dkfzheidelberg.de doi:10.1038/nrc798

Carcinomas of the anogenital tract — particularly cancer of the cervix — account for almost 12% of all cancers in women, and so represent the second most frequent gynaecological malignancy in the world1. Cervical cancer is caused by specific human papillomavirus (HPV) infections (reviewed in REF. 2). Even ~25% of oral cancers contain anogenital HPV DNA³. These HPV types have therefore emerged as one of the most important identified risk factors for widespread human cancers.

Here, cervical cancer will be used as a 'test case' to reveal some of the properties of specific HPV types in human carcinogenesis, and to show the development of clinical approaches that interfere with these infections.

Between 1974 and 1976, researchers started to postulate and analyse a possible role of HPV in cervical cancer⁴⁻⁶. In 1976, Meisels and Fortin^{7,8} published two reports outlining that the appearance of KOILOCYTES in cervical smears indicates the presence of a papillomavirus infection. They also suggested that it might be possible to differentiate between 'benign, warty' lesions that do not progress to cervical cancer and — as they suspected initially — 'nonviral' precursor lesions that do progress to cervical cancer. This idea was supported by the identification of typical papillomavirus particles in mild DYSPLASTIC LESIONS of the cervix 8-10.

The demonstration of heterogeneity within the papillomavirus family^{11–13}, and the subsequent isolation of specific types from genital warts and LARYNGEAL PAPILLOMAS 14,15, as well as the application of nucleic-acid hybridization procedures at reduced stringency — to screen for related but not identical HPV types¹⁶ allowed a fresh look into the possible involvement of papillomavirus infections in anogenital cancers. The first HPV types were isolated directly from cancer biopsies of the cervix — HPV16 and HPV18 were cloned in 1983 and 1984, respectively 17,18 — and this initiated a rapid expansion of the field.

Within four years, the basic experimental details that explained the role of HPVs in cervical cancer aetiology had been described: the expression of specific viral genes (such as E6 and E7) was shown in cervical cancer cell lines and cancer biopsies19; a specific opening within the viral ring molecule was shown for integrated genome copies¹⁷; and the immortalization property of viral DNA^{20,21} and the encoded viral oncogenes²² supported the initial suspicions. An early epidemiological study pointed to a high rate of infection in younger women and to a decreasing rate of infection with age23, although persisting infections still represent a high risk factor for cervical cancer development in older women. Shortly after, the requirement for viral oncogene expression

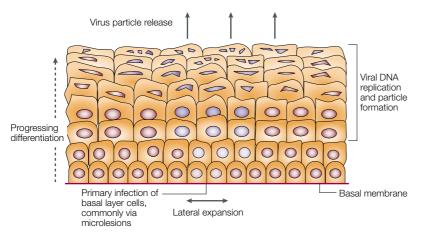


Figure 1 | **The human papillomavirus life cycle.** Infection requires the availability of a basal layer cell. This usually occurs in microlesions of skin or mucosa. The infected cell divides and the population spreads laterally. Some of the progeny migrate into the suprabasal differentiating cell layers, where viral genes are activated, viral DNA is replicated and capsid proteins are formed. Viral particle formation ensues. Particles are released at the surface and might then infect additional tissues.

EPISOME

An independent DNA element, such as a plasmid, that can replicate extrachromosomally or that can be maintained by integration into the genome of the host.

for the maintenance of the malignant phenotype in specific cervical cancer cell lines was shown^{24,25}.

The subsequent 14 years have resulted in a better understanding of viral oncogene functions and in a more detailed knowledge of the natural history of HPV infection. A substantial number of epidemiological studies have also been performed, which point to high-risk HPV as the primary risk factor for cervical cancer. Large case—control and prospective epidemiological studies supported this idea, and indicated that persisting HPV infections were the most significant risk factor for cervical cancer^{26–28}.

HPV pathogenesis

The papillomavirus life cycle differs from all other virus families: infection requires the availability of epidermal or mucosal epithelial cells that are still able to proliferate (basal layer cells) (reviewed in REF. 2). In these cells, viral gene expression is largely suppressed,

Summary

- High-risk human papillomavirus (HPV) genotypes cause cervical cancer. The same types also seem to be responsible for other anogenital, and a subset of head and neck, cancers.
- \bullet Viral oncogene transcription and oncoprotein expression are controlled by cellular signalling cascades.
- Expression of specific viral oncoproteins, E6 and E7, is required for maintaining the malignant growth of cervical cancer cells, specifically by inhibiting the tumour suppressors p53 and RB.
- The detection of viral DNA and cellular proteins that are induced by high-risk HPV allow new approaches to cervical cancer screening to be taken.
- Preventive and therapeutic vaccines against HPV infections are, at present, in clinical trials. The prospects for efficient prevention are excellent.
- Global application of such vaccines could contribute significantly to reducing the human cancer burden.

although the limited expression of specific 'early' viral genes (such as E5, E6 and E7) results in enhanced proliferation of the infected cells and their lateral expansion. Following entry into the suprabasal layers, 'late' viral gene expression is initiated; the circular viral genome is then replicated and structural proteins form. In the upper layers of the epidermis or mucosa, complete viral particles are assembled and released (FIG. 1).

Three genes possess proliferation-stimulating activity: E5, E6 and E7 (FIG. 2). E5 seems to be important in the early course of infection. It stimulates cell growth by forming a complex with the epidermal-growth-factor receptor, the platelet-derived growth-factor-β receptor and the colony-stimulating factor-1 receptor²⁹. Recently, E5 has also been shown to prevent apoptosis following DNA damage³⁰. However, as HPV-infected lesions progress to cervical cancer, the EPISOMAL viral DNA frequently becomes integrated into host-cell DNA (FIG. 2), and a substantial part of the genome, commonly including the E5 coding sequence, is deleted¹⁹. So, E5 is not obligatory in late events of HPV-mediated carcinogenesis.

A more significant role for malignant transformation can be assigned to the E6 and E7 genes and their respective proteins. They are consistently expressed in malignant tissue, and inhibiting their expression blocks the malignant phenotype of cervical cancer cells. They are independently able to immortalize various human cell types in tissue culture, but efficiency is increased when they are expressed together ^{22,31}.

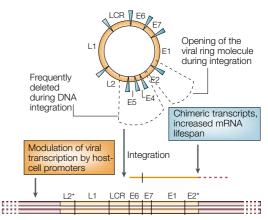


Figure 2 | The organization of circular HPV DNA and its integration into host-cell DNA. The human papillomavirus (HPV) genome contains between 6800 and 8000 base pairs and is divided into eight open reading frames — E6, E7, E1, E2, E4, E5, and L2 and L1 - coding for 'early' (E) or 'late' (L) functions. In the course of cancer development, the viral molecule frequently becomes integrated into host-cell DNA. The ring molecule is most often opened within the E2 open reading frame, disrupting the continuity of that gene. Part of E2 and open reading frames that are adjacent to E2 - E4, E5 and part of L2 — are regularly deleted after integration (partial genes are represented by an asterisk). Viral transcripts, which uniformly span the E6 and E7 region, and are often linked to flanking cellular sequences, are present and transcription might be modulated (enhanced) by flanking host-cell promoters. LCR, long control region.

REVIEWS REVIEWS

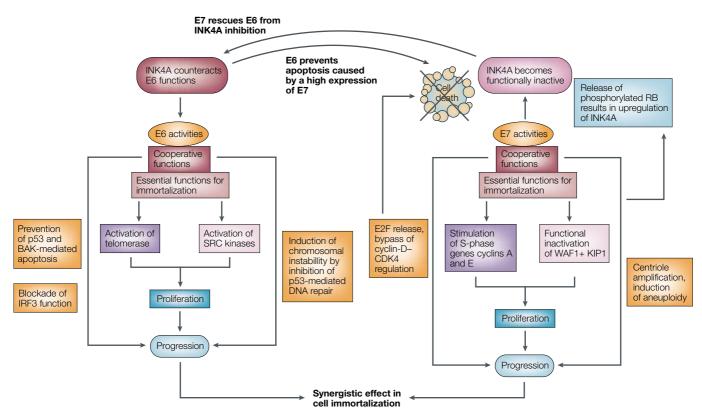


Figure 3 | Functions of the E6 and E7 oncoproteins, and their interaction with each other in steps that lead to cell immortalization. E6 functions to activate telomerase and the SRC kinases and inhibit p53 and BAK. E7 inhibits RB, which releases E2F and results in the upregulation of INK4A, but E7 also inactivates INK4A. In addition, E7 seems to stimulate cyclins A and E, and inactivate the cyclin-dependent kinase inhibitors WAF1 and KIP1. E6 and E7 synergize in cell immortalization and malignant transformation: E6 prevents apoptosis that is induced by high E2F levels, and E7 rescues E6 from inhibition by INK4A.

Several functions have been described for E6 and E7 (FIG. 3). Initial observations revealed that E6 interacts with p53 (REF. 32), and E7 interacts with RB³³ to block the activity of these tumour suppressors. Indeed, some of the prominent functions of the E6 protein originate from its interaction with, followed by degradation of, p53 and the pro-apoptotic protein BAK³⁴, which results in resistance to apoptosis and an increase in chromosomal instability. In addition, the activation of telomerase 35 and the postulated inhibition of degradation of SRC-family kinases by the E6 oncoprotein³⁶ seem to fulfil important functions in growth stimulation. It has been speculated that the stabilization of the activated forms of specific members of the SRC family of kinases could contribute to the HPV-transformed phenotype³⁶. The cyclin-dependent kinase inhibitor INK4A (also known as p16) seems to counteract these functions.

E7, however, interacts with and degrades RB, which releases the transcription factor E2F from RB inhibition and upregulates INK4A^{33,37}. The resulting high E2F activity might lead to apoptosis in E7-expressing cells. Moreover, E7 stimulates the S-phase genes cyclin A and cyclin E³⁸, and seems to block the function of the cyclin-dependent kinase inhibitors WAF1 (also known as CIP1 and p21) and KIP1 (also known as p27)^{39–41}. By inducing centriole amplification, E7 also induces aneuploidy of the E7-expressing cells, which contributes to tumorigenesis⁴².

E6 and E7 can independently immortalize human cells, but at reduced efficiency ^{43,44}; their joint function results in a marked increase in transforming activity. This seems to be due to an interesting complementary and synergistic effect. As mentioned previously, E6 seems to be impaired by INK4A, whereas E7 bypasses this inhibition by directly activating cyclins A and E. E6, in turn, prevents E7-induced apoptosis by degrading the apoptosis-inducing proteins p53 and BAK ^{38,45}.

At present, it is difficult to assign a role for other HPV early proteins (such as E1, E2 and E4) in the process of malignant conversion. The two structural proteins L1 and L2 are not expressed in precancerous and malignant cells, but they are important for vaccine development (see below).

High- and low-risk HPV infections

HPV types that are found preferentially in cervical and other anogenital cancers have been designated as 'high-risk' types⁴⁶. Conversely, those found primarily in genital warts and non-malignant lesions were labelled as 'low-risk' types. Subsequently, it was shown that only the E6 and E7 genes of high-risk types were able to immortalize human cells in tissue culture ^{22,47} and fulfil the functions outlined in FIG. 3.

High-risk HPV types (TABLE 1), particularly HPV16, are widespread within all human populations. Infection is commonly transmitted by sexual contact and results

Table 1 | Papillomavirus types in genital lesions

Type of genital lesion	HPV type Less prevalent	More prevalent
Condylomata acuminata	42,44,51,53,83	6,11
Intraepithelial neoplasias	6,11,18,26,30,31,33,34,35,39,40,42,43, 45,51,52,53,54,55,56,57,58,59,61,62,64, 66,67,68,69,70,71,73,74,79,81,82,83,84	16
Cervical and other anogenital cancers	(6,11),18,31,33,35,39,45,51,52,54,56, 58,59,66,68,69	16

Human papillomavirus (HPV) types in brackets indicate extremely rare prevalence.

HUMORAL IMMUNE REPONSE A specific immune response, directed against a pathogen, that is mediated by antibodies.

CELLULAR IMMUNE RESPONSE An adaptive immune response, directed against a pathogen, that is mediated by antigen-specific lymphocytes. initially in inconspicuous squamous intraepithelial lesions (SIL) in women (reviewed in REF. 48). Most of these lesions are cleared 6–12 months after appearance, probably due to immunological intervention. A small percentage, however, persists, progresses to high-grade SIL, carcinoma *in situ* and, without surgical interference, to squamous-cell carcinoma or adenocarcinoma of the cervix. The individual stages are outlined in FIG. 4.

Host control of HPV infection

The immune system is important in the control of HPV infections. This can be indirectly deduced from the increased incidence and prolonged persistence of SIL in immunosuppressed women⁴⁹. There is also evidence for T-helper-cell involvement in regressing lesions, and a concomitant detectable HUMORAL and CELLULAR IMMUNE RESPONSE against HPV antigens during

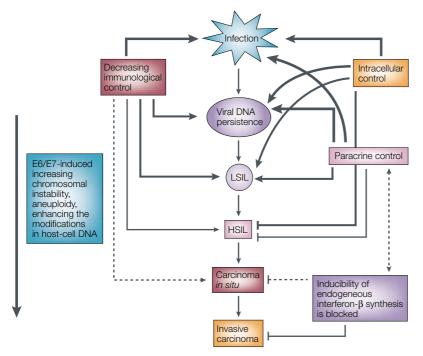


Figure 4 | Systemic and host-cell controls that interfere with HPV-induced progression towards malignant proliferation. The progression of human papillomavirus (HPV)-infected cells to low-grade squamous intraepithelial lesion (LSIL) and high-grade squamous intraepithelial lesion (HSIL), carcinoma in situ and invasive cancer is determined by failing control mechanisms. These include an intracellular control that is presumably exerted by cyclin-dependent kinase inhibitors, a paracrine signalling cascade and a decreasing immunological control. The paracrine control is triggered by macrophages and cytokines, such as tumour necrosis factor- α , and is underlied by loss of synthesis of interferon- β .

the course of regression^{50,51}. The escape of high-grade SIL and carcinomata *in situ* from immunological control seems to be based on different modifications of the cellular antigen-presenting system, which might involve the proteasome transportation system, the HLA receptors and the cellular recognition system for presented oligopeptides^{52–54}. The escape from immune-surveillance mechanisms emerges as one important step in the progression of HPV-linked tumours (FIG. 4).

In addition, two other modes of control exist in proliferating cells that are infected by HPV to protect them against malignant transformation: one involves inhibition of viral oncoprotein function, the other involves transcriptional control (FIG. 4). The evidence that host cells can impair the function of viral oncoproteins is still indirect and, so far, exists only for the E6 oncoprotein in cells that have been immortalized by E6 alone⁵⁵. In cell lines that have been transfected with E6 DNA, the CDKN2A gene — which encodes INK4A — is commonly inactivated by methylation, mutation or deletion. INK4A inactivates cyclin D1-CDK4 or cyclin D1-CDK6 complexes, which prevents expression of cyclin E and, therefore, progression through the cell cycle. The consistent interruption of CDKN2A gene function in E6-immortalized cells indicates that INK4A can functionally interfere with the transforming activity of E6 (REF. 55). As outlined in FIG. 3, expression of E7 can overcome this block, as it directly stimulates cyclins A and E through its interaction with RB38. In cells that are immortalized by both E6 and E7, the CDKN2A gene remains intact. It is still highly overexpressed, but seems to be functionally sequestered from its interference with the cell cycle⁵⁶. No firm evidence exists at present for a negative interference of other cyclin-dependent kinase inhibitors, such as WAF1 and KIP1, with the E7 protein. This could be relevant when E7 is expressed at low levels in the early stages of infection.

Another pathway — which involves blocking the transcription of HPV DNA and is known as the cellular interference factor (CIF) concept — has been analysed in more detail. It is triggered by paracrine stimulation of cervical epithelial cells by macrophages and tumour necrosis factor- α (TNF- α), and causes several effects in HPV-immortalized cells. These include a modification of the transcription factor AP1, which is essential for HPV gene expression, and the induction of endogenous synthesis of the antiviral interferon- β^{57} . It is suspected that the change in AP1 composition mediates the suppression of high-risk HPV transcription⁵⁸. These pathways do not function in cervical carcinoma cells, indicating that the TNF- α -mediated signalling cascade is interrupted during malignant transformation⁵⁹.

Factors that affect HPV malignancy

High-risk HPV infections result in progression to cervical cancer in only a small percentage of infected women, after a long latency period (reviewed in REF. 2). A high percentage of infected women clear the infection by immunological mechanisms. Lasting immunosuppression represents a risk factor for viral DNA persistence and lesional progression⁴⁹, but the factors that determine

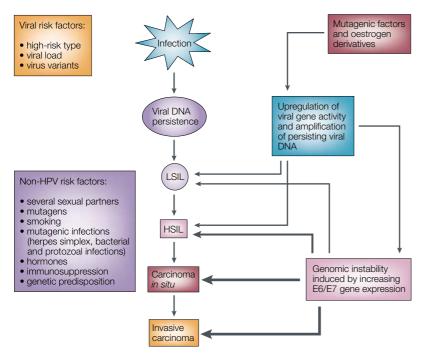


Figure 5 | HPV and non-HPV factors that contribute to HPV-induced malignant progression. Viral and non-viral risk factors that influence the progression of infected cells are listed on the left side. Hormonal factors (oestrogen and its derivatives) activate the human papillomavirus (HPV) promoter and facilitate immortalization of HPV-infected cells. Mutagenic agents amplify persisting HPV DNA. They enhance progression by modifying cellular signalling cascades that control HPV persistence, or lead to increased viral oncogene expression by a gene-dosage effect. A rising level of E6 and E7 oncogene expression, in turn, results in increasing genomic instability, which further facilitates progression of the infected cells towards invasive growth.

viral persistence in other women are not completely defined. Some result from modifications of cellular genes that influence antigen presentation, or from signalling cascades that are engaged in the suppression of viral oncogene transcription or viral oncoprotein function (reviewed in REF. 48; FIG. 4). Other factors directly affect the persisting viral DNA; for example, by upregulating viral oncogene transcription, by modifying the viral promoter region or by amplifying the persisting viral genomes (reviewed in REF. 2; FIG. 5).

The modification of specific cellular genes during the course of viral transformation has been postulated previously60 to account for long latency periods, monoclonality of the respective tumours and the SYNCARCINOGENIC EFFECTS of chemical and physical carcinogens. Gyllensten and co-workers elegantly showed the existence of genetic predispositions for cervical cancer by taking advantage of data from the Swedish Cancer Registry: there was a significantly higher risk for natural daughters in comparison with adopted daughters, for sisters in comparison with adopted sisters, and an approximately 50% reduced risk for half-sisters 61. The genes involved in this predisposition have not yet been identified.

Cancer screening

Cervical cancer screening is commonly based on cytological and colposcopic analyses. Screening performed by experienced cytologists and gynaecologists has

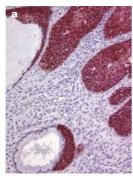
resulted in a dramatic reduction of cervical cancer incidence. In countries with less experienced cytologists, cytology nurses or gynaecologists, however, these tests run a high risk for false-negative or false-positive results. Simple, reliable and cheap tests for effective large-scale screening programmes are therefore required — particularly in tropical countries that do not have effective screening programmes, and which therefore have the highest rates of cervical cancer.

The identification of specific papillomavirus types as causative agents for cancer of the cervix and its precursor lesions allowed the development of a new method for cancer screening and early diagnosis. HPV genomes and viral oncoproteins, which are present in all affected cells, should represent convenient markers for a transient or persistent infection. Similarly, cellular proteins that are induced in response to these infections could have an analogous role. The presence of high-risk HPV markers per se should not, in itself, be cause for alarm, but rather should lead to the instigation of a careful clinical investigation and repeated testing for HPV persistence, which is a significant risk factor for the development of proliferative lesions and their progression⁶².

The detection of high-risk HPV DNA by hybridization procedures or by polymerase chain reaction (PCR) technology has been developed to some maturity, and enables a risk assessment to be made for low-grade squamous intraepithelial lesions (LSIL)63. This strategy – for which molecular probes are already available as kits — has also been recommended to replace or be combined with conventional cytological screening procedures⁶⁴. If competently performed, these techniques clearly improve the detection rate of high-risk HPV DNA in women, and also avoid some false-negative diagnoses 65 ; however, they are relatively expensive and therefore prohibitive for a large part of the world where cervical cancer is still a principal cause of death.

Another consequence of HPV infection might also be capitalized on to improve screening procedures. Papillomavirus infection induces cell proliferation within the differentiating layers of the epithelium that are otherwise devoid of replicating cells (FIG. 1). Theoretically, the abnormal expression, within these cells, of a set of proteins that are involved in progression through the cell cycle might serve as a marker for HPV infections. INK4A emerges as a particularly powerful candidate for such a marker. In high-risk HPV infection, E7 binds to and degrades RB (see above and FIG. 3), which results in substantial upregulation of INK4A synthesis — readily detectable in high-risk HPV-infected cells and tissues⁵⁶. In spite of the high level of INK4A synthesis, this protein remains functionally inactive, as E7 induces cyclin A and cyclin E expression, thereby functionally bypassing its interference with the cell cycle. Antibodies that are directed against INK4A allow selective staining of high-risk HPV-infected histological sections or of cytological smears (FIG. 6). It is anticipated that this procedure, if affordable, could be used in cervical screening and might significantly reduce the number of false-negative diagnoses.

SYNCARCINOGENIC EFFECT Synergism between two factors. each of them able to induce



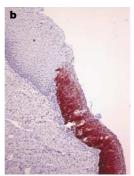


Figure 6 | **Cytological smears**. **a** | A cervical adenocarcinoma and **b** | a cervical neoplasia stained for the expression of INK4A. The intensive red staining pattern of the human-papillomavirus-infected tissue sharply demarcates it from non-infected cells. Photographs kindly provided by Magnus von Knebel Doeberitz, Heidelberg.

Tests for HPV DNA originate as a direct consequence of HPV identification; the presence of INK4A allows, by contrast, an indirect approach for detecting high-risk HPV.

Cancer prevention

Hygiene. The most visible benefit from HPV research is prevention of cervical cancer. A very early benefit originated from the recognition by clinicians that cervical, vulvar, vaginal and perianal squamous intraepithelial lesions are of infectious origin and produce large quantities of infectious virus (reviewed in REF.2). Specific standards of hygiene were therefore enforced in order to prevent latrogenic transmissions among gynaecological patients.

Conversely, the mechanical prevention of anogenital HPV infections that are transmitted by sexual contact is impractical. HPV infections are highly prevalent within the sexually active population and the use of condoms offers only limited protection in view of the common presence of virus-infected cells at external genital sites.

Vaccines. One of the main benefits of HIV research, however, would be vaccination against high-risk HPV types. It is widely accepted that in most cases, HPV infection is cleared by immunological intervention. Spontaneous regression of LSILs, or even HSILs, is commonly accompanied by humoral and cellular immune responses against virus-specific antigens⁵⁹⁻⁶¹. Although the surface localization of most viral antigens does not sufficiently stimulate the immune system, vaccination with viral structural proteins in animal systems and in humans regularly leads to the induction of an effective immune response⁶⁶⁻⁶⁹.

Initial experiments performed with purified papillomavirus structural proteins — that spontaneously assemble into 'VIRUS-LIKE PARTICLES' (VLPs) — of animal papillomaviruses (such as canine oral papillomavirus and cottontail rabbit papillomavirus) resulted in effective protection against the primary infection of dogs and rabbits, respectively ^{66,67}. These results provided the background for attempts to develop vaccines against human high-risk HPV types.

Several companies, but also academic research laboratories, are engaged, at present, in preclinical and clinical trials of vaccines against high-risk HPV. Most approaches are based on the use of VLPs that are derived from the structural proteins L1, or L1 and L2. Frazer and colleagues initially used L1 and L2 VLPs⁶⁸ and applied them to HPV-infected patients and patients with genital warts⁶⁹. The conclusion of a Phase I safety and immunogenicity trial in adult volunteers of an HPV16L1 VLP vaccine was recently reported⁷⁰. This vaccine was highly immunogenic and well tolerated, even without adjuvant, as was previously shown for an HPV6b VLP vaccine⁶⁹. The antibody titres seem to be, on average, at least 10 times higher than after clearing of a natural infection and persist for prolonged periods. Although the available data do not yet allow firm conclusions to be made about protective effects, animal papillomavirus vaccination studies do point in this direction. So, the prospects for this vaccination are remarkably promising. On the basis of previously mentioned studies in dogs and rabbits, an effective prevention can also be expected for the human vaccine. If this turns out to be true and if the vaccine was globally applied, prevention of infection by the most prevalent high-risk HPV types could theoretically prevent more than 300,000 cervical cancer cases per year on a global scale.

It is anticipated that POLYVALENT VACCINES will eventually reach the market. Vaccines that cover the four or five most prevalent high-risk HPV types should be able to prevent 80–90% of cervical cancer incidence. In view of some geographical differences in the prevalence of specific high-risk HPV types, polyvalent vaccines might need to be adapted slightly for use in different countries.

Attempts are, at present, being made to develop vaccines that do not require injections. Particularly in parts of the developing world, the repeated use of syringes without appropriate intervening sterilization bears the risk of transmitting other agents, such as the hepatitis B and C viruses, and human immunodeficiency virus (HIV). Some studies are testing the inhalation of VLPs or L1 genes in viral vectors^{71,72}. These vectored constructs seem to be particularly suited to intranasal inhalation, and are capable of inducing an immune response at cervical sites⁷³.

The discovery of infectious agents as causative factors for specific human cancers has already been shown to have important consequences for cancer prevention. The Taiwan vaccination programme of newborn children against hepatitis B virus (HBV) infections, introduced in 1986, not only drastically reduced the percentage of persistently HBV-infected children, but also resulted in the first measurable decrease in liver cancer incidence⁷⁴. Provided that the vaccines against human high-risk HPVs prove to be as effective as in animal experiments, it can be estimated that a global application of HPV vaccines could theoretically reduce the cancer risk in women by approximately 10–15%⁴⁸.

IATROGENIC TRANSMISSION
Transmission of infections by a

VIRUS-LIKE PARTICLE (VLP). Empty shell of a virus particle, without genetic material, that is produced by genetic engineering.

POLYVALENT VACCINE
A vaccine that binds to, or
generates an immune response
against, more than one thing. In
this case, several human
papillomavirus types.

ACYCLIC NUCLEOSIDE PHOSPHONATE A drug that blocks the replication of several virus types.

IMMUNOMODULATORY DRUG A drug that modifies the immune system.

Cancer therapy

Immunotherapy. Concepts similar to those developed for immunoprevention of HPV infections have also been adapted for immunotherapy. The development of chimeric vaccines seems to be an appropriate step in this direction^{75,76}. For example, antigenic epitopes from the E7 oncoprotein have been added to the L1 protein to generate deformed VLPs. In animal experiments, these VLPs exert both a preventive and a therapeutic effect, as an immune response is induced against the viral E7 oncoprotein; a Phase I trial was initiated in 2001 (L. Gissmann, personal communication). This type of combined immunopreventive and immunotherapeutic vaccine might be particularly useful in the treatment of early lesions (cervical intraepithelial neoplasias), but it is more questionable whether it will still be effective in invasive HPV-caused cancers that have a large tumour mass. A Phase I clinical trial that vaccinated cervical cancer patients with E6 and E7 proteins — with the aim of generating an immune response against these viral oncoproteins — has also been reported77,78. Unfortunately, for the women vaccinated in this study — who had late-stage cervical cancer — the vaccine did not seem to be beneficial. Oligopeptide vaccines derived from the E7 oncoprotein might, however, show some clinical activity⁷⁹.

It remains to be seen whether vaccinations will be of benefit to those who have developed cervical cancer, as the cellular adaptations that take place during tumorigenesis regularly seem to result in the loss of appropriate viral antigen presentation. Immunotherapy might have a reasonable chance of success in patients with precursor lesions, but its effectiveness in invasive cancer is less likely.

Immunomodulatory therapy. Several cytokines have been shown to repress papillomavirus transcription, and this repression involves transforming growth factor- β (TGF- β)^{80,81} and interleukin-1 (REF. 82), as well as TNF- $\alpha^{59,83}$. The repression by TNF- α is lost during malignant conversion⁵⁹. None of the cytokines have been used in clinical trials so far.

The activation of retinoic-acid receptors by a tissue hormone, retinoic acid, can also suppress HPV gene activity 84,85 and has been shown to reveal some clinical effects in premalignant and malignant cervical lesions. Unfortunately, unpleasant side effects accompany its use86,87.

An acyclic nucleoside phosphonate, cidofovir, which has broad-spectrum activity against DNA viruses, also shows some activity against papillomaviruses, by inducing apoptosis, if locally applied in infected cells88. In addition, the IMMUNOMODULATORY DRUG imiquimod has proved to be effective in HPV-infected cells (reviewed in REF. 89) and seems to act by the stimulation of cytokines.

Molecular and chemotherapy. Several interesting in vitro studies might have an impact on future developments in chemotherapeutic approaches to HPV infections and HPV-linked human cancers. Because the malignant growth of cervical cancer cells depends on the presence and expression of the E6/E7 oncogenes, antisense constructs were generated against both the E6 and E7 message. This was shown to inhibit the tumorigenicity of cervical cancer cells in nude mice²⁴.

Another report revealed that transcription of high-risk HPV could be selectively inhibited by the antioxidant pyrrolidine-dithiocarbamate (PDTC) in HPV-immortalized, but not in malignant, cells⁵⁸. 2-Deoxyglucose also suppresses HPV transcription in both immortalized and malignant cells90. More recently, HPV16 E6-binding peptide aptamers have been shown to eliminate HPV16-positive cancer cells by inducing apoptosis⁹¹. In addition, histone deacetylase inhibitors, such as sodium butyrate and trichostatin A, arrest HPV-positive cells and induce apoptosis without affecting the expression of HPV oncogenes⁹². None of these studies has yet been clinically applied. It remains to be seen whether some of them turn out to be useful for treating patients.

Future directions

The present developments in preventive HPV vaccination look particularly promising. The next few years will reveal the protective and therapeutic potential of those preparations that are presently being tested in clinical studies. Although all available results seem to support the idea that interference with infections by a number of HPV types will be achieved, some important questions remain unanswered. Which vaccine formula will provide optimal results? Do we need different combinations of HPV types for different regions of the world? Will it be possible to develop a cost-effective vaccine that is suitable for global application? How do we reach people in those tropical and subtropical regions where cervical cancer is most prevalent? Appropriate answers to these questions and the respective actions have the potential to save the lives of more than 200,000 women per year.

Conclusions

Research in high-risk HPVs started actively in the second half of the 1970s. The concept of HPV heterogeneity and its proof, as well as the demonstration of specific HPV types in cervical and other anogenital cancers, substantially boosted this research. After partial elucidation of the viral functions that contribute to malignant conversion, clinical application of these results started to emerge. An important practical impact originates from the development of prophylactic and therapeutic vaccines. Theoretically, cervical cancer, which contributes to approximately 12% of the global cancer burden in women, should be preventable. In addition, however, new screening procedures that are based on the molecular understanding of high-risk HPV-host-cell interactions and targeted therapeutic approaches are emerging. These will, in all likelihood, contribute in the future to the control of this, as well as of other, HPV-linked human cancers. HPV research exemplifies the value of basic research for cancer prevention and shows the importance of infectious events in human carcinogenesis.

- Pisani, P., Bray, F. & Parkin, D. M. Estimates of the worldwide prevalence of cancer for 25 sites in the adult population. Int. J. Cancer 97, 72-81 (2002).
- zur Hausen, H. Papillomavirus infections: a major cause of human cancers. Biochem. Biophys. Acta 1288, F55-F78
- Gillison, M. L. & Shah, K. V. Human papillomavirusassociated head and neck squamous cell carcinoma: mounting evidence for an etiologic role for human papillomavirus in a subset of head and neck cancers. Curr. Opin. Oncol. 13, 183–188 (2001).
- zur Hausen, H., Meinhof, W., Scheiber, W. & Bornkamm, G. W. Attempts to detect virus-specific DNA sequences in human tumors: I. Nucleic acid hybridizations with complementary RNA of human wart virus. Int. J. Cancer 13, 650-656 (1974).

First experiments to search for papillomavirus DNA in cervical cancer.

- zur Hausen, H. Condylomata acuminata and human genital cancer. Cancer Res. 36, 530 (1976).
- zur Hausen, H. Human papilloma viruses and their possible role in squamous cell carcinomas. Curr. Top. Microbiol. Immunol. **78**, 1–30 (1977).
- Meisels, A. & Fortin, R. Condylomatous lesions of the cervix and vagina. I. Cytologic patterns. Acta Cytol. 20, 505-509 (1976). The first study to point to a role of papillomavirus infections in cervical smears and mild dysplastic
- Meisels, A. et al. Human papillomavirus infection of the 8. cervix: the atypical condyloma. Acta Cytol. 25, 7-16 (1981).
- Della Torre, G., Pilotti, S., de Palo, G. & Rilke, F. Viral particles in 9
- cervical condylomatous lesions. *Tumori* **64**, 459–463 (1978). Laverty, C. R., Russel, P., Hills, E. & Booth, N. The significance of non-condyloma wart virus infection of the cervical transformation zone. A review with discussion of two illustrative cases. Acta Cytol. 22, 195-201 (1978).
- Gissmann, L. & zur Hausen, H. Human papilloma viruses: physical mapping and genetic heterogeneity. *Proc. Natl* Acad. Sci. USA 73, 1310-1313 (1976).
- Gissmann, L., Pfister, H. & zur Hausen, H. Human papilloma viruses (HPV): characterization of four different isolates. Virology **76**, 569–580 (1977). Orth, G., Favre, M. & Croissant, O. Characterization of a new
- 13 type of human papillomavirus that causes skin warts. J. Virol. 24, 108–120 (1977).
 Gissmann, L. & zur Hausen, H. Partial characterization of
- 14. viral DNA from human genital warts (condylomata acuminata). Int. J. Cancer 25, 605-609 (1980). Gissmann, L., Diehl, V., Schultz-Coulon, H. & zur Hausen, H.
- 15 Molecular cloning and characterization of human papillomavirus DNA from a laryngeal papilloma. J. Virol. 44, . 393–400 (1982).
- Law, M. F., Lancaster, W. D. & Howley, P. M. Conserved polynucleotide sequences among the genomes of papillomaviruses. J. Virol. 32, 199-207 (1979).
- Dürst, M, Gissmann, L., Ikenberg, H. & zur Hausen, H. A papillomavirus DNA from a cervical carcinoma and its prevalence in cancer biopsy samples from different geographic regions. *Proc. Natl Acad. Sci. USA* **80**, 3812–3815 (1983).

Isolation of the first cervical-cancer-linked human

- **papillomavirus type, HPV16.**Boshart, M. *et al.* A new type of papillomavirus DNA, its presence in genital cancer and in cell lines derived from genital cancer. *EMBO J.* **3**, 1151–1157 (1984).
- Schwarz, E. et al. Structure and transcription of human papillomavirus type 18 and 16 sequences in cervical carcinoma cells. *Nature* **314**, 111–114 (1985). Description of active HPV genes (E6/E7) in cervical
- cancer cells and demonstration of specific HPV deletions after integration into host-cell DNA. Dürst, M., Dzarlieva-Petrusevska, R. T., Boukamp, P. Fusenig, N. E. and Gissmann, L. Molecular and cytogenetic analysis of immortalized human primary keratinocytes obtained after transfection with human papillomavirus type 16 DNA. *Oncogene* **1**, 251–256 (1987).
- Pirisi, L., Yasumoto, S., Fellery, M., Doninger, J. K. & DiPaolo, J. A. Transformation of human fibroblasts and keratinocytes with human papillomavirus type 16 DNA. . Virol. 61, 1061–1066 (1987).
- Münger, K., Phelps, W. C., Bubb, V., Howley, P. M. & Schlegel, R. The E6 and E7 genes of human papillomavirus type 16 are necessary and sufficient for transformation of primary human keratinocytes. J. Virol. 63, 4417–4423 (1989).

 Description of the transforming properties of E6
- de Villiers, F.-M. et al. Human papillomavirus infection in women with and without abnormal cervical cytology. Lancet 2, 703-706 (1987)
 - The first epidemiological study of HPV infections.

von Knebel Doeberitz, M., Rittmüller, C., zur Hausen, H. & Dürst, M. Inhibition of tumorigenicity of cervical cancer cells in nude mice by HPV E6-E7 antisense RNA. Int. J. Cancer **51**, 831-834 (1992).

E6 and E7 genes are necessary factors for the malignant phenotype of HPV-positive cervical cancer

- von Knebel Doeberitz, M., Rittmüller, C., Aengeneyndt, F., Jansen-Dürr, P. & Spitkovsky, D. Reversible repression of papillomavirus oncogene expression in cervical carcinoma cells: consequences for the phenotype and E6-p53 and E7pRB interactions. J. Virol. 68, 2811–2821 (1994). Muñoz, N. et al. The causal link between human
- papillomavirus and invasive cervical cancer: a populationbased case-control study in Columbia and Spain. Int. J. Cancer **52**, 743–749 (1992).

A large-scale epidemiological study that provides convincing evidence that high-risk HPVs are the main risk factor for cervical cancer.

- 27. Bosch, F. X. et al. Prevalence of human papillomavirus in cervical cancer: a worldwide perspective. International biological study on cervical cancer (IBSCC) Study Group. J. Natl Cancer Inst. 87, 796–802 (1995).
- IARC Monograph on Evaluation of Carcinogenic Risks of Humans: Human Papillomaviruses Vol. 64 (IARC, Lyon, 1995).

HPV16 and HPV18 were defined as causative agents for cancer of the cervix.

- Hwang, E. S., Nottoli, T. & Dimaio, D. The HPV 16 E5 protein: expression, detection, and stable complex formation with transmembrane proteins in COS cells. *Virology* **211**, 227–233 (1995).
- Zhang, B., Spandau, D. F. & Roman, A. S. E5 protein of human papillomavirus type 16 protects human foreskin keratinocytes from UV B-irradiation-induced apoptosis. Virol. 76, 220-231 (2002).
- McDougall, J. K. Immortalization and transformation of human cells by human papillomavirus. *Curr. Top. Microbiol.* Immunol. 186, 101-119 (1994).
- 32. Werness, B. A., Levine, A. J. & Howley, P. M. Association of human papillomavirus types 16 and 18 E6 proteins with p53. Science **248**, 76–79 (1990). Dyson, N., Howley, P. M., Münger, K. & Harlow, E. The
- human papillomavirus-16 E7 oncoprotein is able to bind to the retinoblastoma gene product. Science 243, 934-937 (1989).
- Jackson, S., Harwood, C., Thomas, M., Banks, L. & Storey, A. Role of Bak in UV-induced apoptosis in skin cancer and abrogation by HPV E6 proteins. *Genes Dev.* **14**, 3065–3073
- Veldman, T., Horikawa, I., Barrett, J. C. & Schlegel, R. Transcriptional activation of the telomerase hTERT gene by human papillomavirus type 16 E6 oncoprotein. J. Virol. 75, 4467-4472 (2001).
- Oda, H., Kumar, S. & Howley, P. M. Regulation of the Src family tyrosine kinase Blk through E6AP-mediated ubiquitination. Proc. Natl Acad. Sci. USA 96, 9557–9562
- Kiyono, T. et al. Both Rb/p16lNK4a inactivation and telomerase activity are required to immortalize human epithelial cells. Nature 396, 84-88 (1998).
- Zerfass, K. et al. Sequential activation of cyclin E and cyclin A gene expression by human papillomavirus type 16 E7 through sequences necessary for transformation. J. Virol. 69, 6389-6399 (1995).
- Jones, D. L., Alani, R. M. & Münger, K. The human papillomavirus E7 oncoprotein can uncouple cellular differentiation and proliferation in human keratinocytes by abrogating p21CIP1-mediated inhibition of CDK2. Genes
- Dev. 11, 2101–2111 (1997). Funk, J. O. et al. Inhibition of CDK activity and PCNAdependent DNA replication by p21 is blocked by interaction with the HPV-16 E7 protein. Genes Dev. 11, 2090-2100 (1997).
- Zerfass-Thome, K. et al. Inactivation of the CDK inhibitor p27KIP1 by the human papillomavirus type 16 E7 oncoprotein. Oncogene 13, 2323-2330 (1996).
- Duensing, S., Duensing, A., Crum, C. P. & Münger, K. Human papillomavirus type 16 E7 oncoprotein-induced abnormal centrosome synthesis is an early event in the evolving malignant phenotype. Cancer Res. 61, 2356-2360 (2001).
- Band, V., Zaychowski, D., Kulesa, V. & Sager, R. Human papillomavirus DNAs immortalize normal human mammary epithelial cells and reduce their growth factor requirements. Proc. Natl Acad. Sci. USA 87, 463–467 (1990). Halbert, C. L., Demers, G. W. & Galloway, D. A. The E7 gene
- of human papillomavirus type 16 is sufficient for immortalization of human epithelial cells. *J. Virol.* **65**, 473–478 (1991).

- Thomas, M. & Banks, L. Inhibition of BAK-induced apoptosis by HPV-18 E6. Oncogene 17, 2943–2954 (1998).
- zur Hausen, H. in Viruses and Cancer (eds Rigby, P. W. J. & Wilkie, N. M.) 83-90 (Cambridge Univ. Press, Cambridge, 1986).
- Hawley-Nelson, P., Vousden, K. H., Hubbert, N. L., Lowy, D. R. & Schiller, J. T. HPV 16 E6 and E7 proteins cooperate to immortalize human foreskin keratinocytes. EMBO J. 8, 3905-3910 (1989).
- zur Hausen, H. Papillomaviruses causing cancer: evasion from host cell control in early events in carcinogenesis. J. Natl Cancer Inst. **92**, 690–698 (2000).
 Petry, K. U. *et al.* Cellular immunodeficiency enhances the
- progression of human papillomavirus-associated cervical lesions. Int. J. Cancer 57, 836–840 (1994). Jenson, A. B., Kurman, R. J. & Lancaster, W. D. Tissue
- effects and host response to human papillomavirus infection. Dermatol. Clin. 9, 203-209 (1991).
- Höpfl, R. et al. Spontaneous regression of CIN and delayedtype hypersensitivity to HPV-16 oncoprotein E7. Lancet **356**, 1985–1986 (2000).
- Spataro, V., Norbury, C. & Harris, A. L. The ubiquitinproteasome pathway in cancer. Br. J. Cancer 77, 448-455 (1998).
- Bornstein, J. et al. Interferon- β and $-\gamma$, but not tumor necrosis factor- α , demonstrate immunoregulatory effects on carcinoma cell lines infected with human papillomavirus. Cancer 79, 924-934 (1997).
- Ghaderi, M. et al. Tumor necrosis factor α-11 and DR15-DQ (B*0602) haplotype increase risk for cervical intraepithelial neoplasia in human papillomavirus 16 seropositive women in Northern Sweden. Cancer Epidemiol. Biomarkers Prev. 9, 1067-1070 (2000).
- Reznikoff, C. A. et al. Long-term genome stability and minimal genotypic and phenotypic alterations in HPV16 E7-, but not E6-immortalized human uroepithelial cells. Genes Dev. **8**, 2227–2240 (1994). Klaes, R. *et al.* Overexpression of p16(INK4) as a specific
- marker for dysplastic and neoplastic epithelial cells of the cervix uteri. Int. J. Cancer 92. 276-284 (2001). Provides a new approach for the screening of highrisk HPV infections.
- Bachmann, A. et al. Disturbance of tumor necrosis factor-αmediated interferon-β signaling in cervical carcinoma cells
- J. Virol. **76**, 280–291 (2002). Rösl, F., Das, B. C., Lengert, M., Geletneky, K. & zur Hausen, H. Antioxidant-induced changes of the AP-1 transcription complex are paralleled by a selective suppression of human papillomavirus transcription. *J. Virol.* **71**, 362–370 (1997).
- Soto, U. et al. Conversion of HPV 18 positive non-tumorigenic HeLa-fibroblast hybrids to invasive growth involves loss of TNF- α mediated repression of viral transcription and modification of the AP-1 transcription complex. Oncogene 18, 3187-3198 (1999).
- zur Hausen, H. Papillomaviruses in anogenital cancer: a model to understand the role of viruses in human cancers. Cancer Res. 49, 4677-4681 (1989).
- Magnusson, P. K., Sparen, P. & Gyllensten, U. B. Genetic link to cervical tumours. *Nature* **400**, 29–30 (1999).

First evidence for a genetic component in cervical cancer development. Ho, G. Y. et al. Persistent genital human papillomavirus

- infection as a risk factor for persistent cervical dysplasia. J. Natl Cancer Inst. 87, 1345–1347 (1995). Schneider, A. et al. Screening for high grade cervical
- intraepithelila neoplasia and cancer by testing for high-risk HPV, routine cytology or colposcopy. Int. J. Cancer 89, 529-534 (2000).
- Cuzick, J. HPV typing in cervical screening. Pharmacoepidemiol. Drug Saf. **10**, 33–36 (2001).
- Schlecht, N. F. et al. Persistent human papillomavirus infection as a predictor of cervical intraepithelial neoplasia. JAMA 286, 3106–3114 (2001).
- Suzich, J. A. et al. Systemic immunization with papillomavirus L1 protein completely prevents the development of viral mucosal papillomas. Proc. Natl Acad. Sci. USA 92, 11553–11557 (1995). The first report on preventive large-scale

papillomavirus vaccination in dogs.

- Breitburd, F. et al. Immunization with virus-like particles from cottontail rabbit papillomavirus (CRPV) can protect against experimental CRPV infection. J. Virol. 69, 3959-3963 (1995). In a similar way to that described in reference 66, preventive vaccines were developed against
- cottontail rabbit papillomavirus infections. Zhou, J., Sun, X. Y., Stenzel, D. J. & Frazer, I. H. Expression of vaccinia recombinant HPV 16 L1 and L2 ORF proteins in epithelial cells is sufficient for assembly of HPV virion-like particles. *Virology* **185**, 251–257 (1991).

Expression of HPV structural proteins leads to spontaneous virus-like particle formation.

- Zhang, L. F. et al. HPV 6b virus like particles are potent immunogens without adjuvant in man. Vaccine 18, 1051-1058 (2000).
- Harro, C. D. et al. Safety and immunogenicity trial in adult volunteers of a human papillomavirus 16 L1 viruslike particle vaccine. J. Natl Cancer Inst. 93, 284-292 (2001).

The first report on immunogenicity of VLP vaccines in humans

- 71. Balmelli, C. et al. Nasal immunization of mice with human papillomavirus type 16 virus-like particles elicits neutralizing antibodies in mucosal secretions. J. Virol. 72, 8220-8229 (1998)
- Grimm, D. & Kleinschmidt, J. A. Progress in adenoassociated virus type 2 vector production: promises and prospects for clinical use. Hum. Gene Ther. 10, 2445-2450 (1999)
- Johansson, E. L., Wassen, L., Holmgren, J., Jertborn, M. & Rudin, A. Nasal and vaginal vaccinations have differential effects on antibody responses in vaginal and cervical secretions in humans. Infect. Immun. 69, 7481-7486 (2001).
- Chang, M. H. et al. Universal hepatitis B vaccination in Taiwan and the incidence of hepatocellular carcinoma in children. Taiwan childhood hepatoma study group. N. Engl. *J. Med.* **336**, 1855–1859 (1997).

The first report of an effective trend for a human tumour virus vaccine in preventing cancer.

- Greenstone, H. L. et al. Chimeric papillomavirus virus-like particles elicit antitumor immunity against the E7 oncoprotein in an HPV16 tumor model. Proc. Natl Acad.
- Sci. ÚSA **95**, 1800–1805 (1998). Jochmus, I., Schäfer, K., Faath, S., Müller, M. & Gissmann, L Chimeric virus-like particles of the human papillomavirus type 16 (HPV 16) as a prophylactic and therapeutic vaccine. *Arch. Med. Res.* **30**, 269–274 (1999).
- Borysiewicz, L. K. et al. A recombinant vaccinia virus encoding human papillomavirus types 16 and 18, E6 and E7 proteins as immunotherapy for cervical cancer. *Lancet* **347**, 1523–1527 (1996).

- 78. Adams, M. et al. Clinical studies of human papilloma virus vaccines in pre-invasive and invasive cancer. Vaccine 19. 2549-2556 (2001).
- Muderspach, L. et al. A phase I trial of human papillomavirus (HPV) peptide vaccine for women with high-grade cervical and vulvar intraepithelial neoplasia who are HPV 16 positive Clin. Cancer Res. 6, 3406-3416 (2000).
- Woodworth, C. D., Notario, V. & DiPaolo, J. A. Transforming growth factor-β1 and 2 transcriptionally regulate human papillomavirus (HPV) type 16 early gene expression in HPVimmortalized human genital epithelial cells. J. Virol. 64, 4767–4775 (1990). Braun, L., Dürst, M., Mikumo, R. & Gruppuso, P. Differential
- response of nontumorigenic and tumorigenic human papillomavirus type 16-positive epithelial cells to transforming growth factor- β I. Cancer Res. **50**, 7324–7332 (1990).
- Kyo, S. et al. Regulation of early gene expression of human papillomavirus type 16 by inflammatory cytokines. Virology **200**, 130–139 (1994).
- Malejczyk, J. et al. Increased tumorigenicity of human keratinocytes harbouring human papillomavirus type 16 is associated with resistance to endogenous tumor necrosis factor-α-mediated growth limitation. Int. J. Cancer 56, 593-598 (1994).
- Bartsch, D., Boye, B. Baust, C., zur Hausen, H. & Schwarz, E. Retinoic acid-mediated repression of human papillomavirus 18 transcription and different ligand regulation of the retinoic acid receptor-β gene in nontumorigenic and tumorigenic HeLa hybrid cells. *EMBO J.* **11**, 2283–2291 (1992).
- Pirisi, L., Batova, A., Jenkins, G. R., Hodam, J. R. & Creek, K. E. Increased sensitivity of human keratinocytes immortalized by human papillomavirus type 16 DNA to growth control by retinoids. Cancer Res. 52, 187-193 (1992).
- Surwit, E. A. et al. Evaluation of topically applied transretinoic acid in the treatment of cervical intraepithelial lesions. Am. J. Obstet. Gynecol. 143, 821–823 (1982).
- Lippman, S. M. et al. Retinoic acid and interferon combination studies in human cancer. Eur. J. Cancer 29A (Suppl. 5), S9-S13 (1993).
- Andrei, G., Snoek, R., Schols, D. & De Clerq, E. Induction of apoptosis by cidofovir in human papillomavirus (HPV)positive cells. Oncol. Res. 12, 397-408 (2001).

- 89. Baker, G. E. & Tyring, S. K. Therapeutic approaches to papillomavirus infections. Dermatol. Clin. 15, 331-340
- Maehama, T. et al. Selective down-regulation of human papillomavirus transcription by 2-deoxyglucose. Int. J.
- Butz, K., Denk, C., Ullmann, A., Scheffner, M. & Hoppe-Seyler, F. Induction of apoptosis in human papillomaviruspositive cancer cells by peptide aptamers targeting the viral E6 oncoprotein. Proc. Natl Acad. Sci. USA 97. 6693-6697 (2000).
- Finzer, P., Kuntzen, C., zur Hausen, H. & Rösl, F. Inhibitors of histone deacetylase arrest cell cycle and induce apoptosis in cervical carcinoma cells circumventing human papillomavirus oncogene expression. Oncogene 20, 4768-4776 (2001).

Online links

DATABASES

The following terms in this article are linked online to:

Cancer.gov: http://www.cancer.gov/cancer_information/ cervical cancer | liver cancer | oral cancer GenBank: http://www.ncbi.nih.gov/Genbank/ canine oral papillomavirus | cottontail rabbit papillomavirus | E1 | E2 | E4 | E5 | E6 | E7 | hepatitis B | hepatitis C | HIV | HPV16 | HPV18 | L1 | L2

LocusLink: http://www.ncbi.nlm.nih.gov/LocusLink/ BAK | CDK4 | CDK6 | colony-stimulating factor-1 receptor | cyclin A | cyclin D1 | cyclin E | E2F | epidermal-growth-factor receptor | INK4A | interferon- β | interleukin-1 | KIP1 | p53 | plateletderived growth-factor- β receptor | RB | SRC | telomerase | TGF- β | TNF-α | WAF1

Medscape DrugInfo:

http://promini.medscape.com/drugdb/search.asp cidofovir | imiguimod

FURTHER INFORMATION

Swedish Cancer Registry: http://www.sos.se/epc/cancereng.htm

Access to this interactive links box is free online.