

Nelson, Curator, USDA-ARS, Soybean Germplasm Collection. These resistant accessions represent samples from five countries: Argentina, China, Japan, Russia and

South Koman Titin alingentible controls all ligher and

belled with ³²P using random priming reactions according to Feinberg and Vogelstein¹⁵.

On a marker basis we have calculated polymorphism

indan 1 Tuil minna u in the ellete formionant for i

Near-homogeneous populations of H. glycines were developed for each of the five races 1, 2, 3, 5 and 14 based on the methods described previously 10. Bioassays were performed for each of the five races based on the established procedures 3. In brief, each soybean seedling was grown in a single polypropylene micropot $(200 \times 25 \, \text{mm})$ filled with steam pasteurized Brosely fine sandy soil. Approximately 20 of these micropots were placed in a 20 cm diameter polypropylene container and maintained at 27 ± 1 °C in thermoregulated waterbaths (Forma Scientific Inc., Marietta, OH, USA).

White and light yellow females of a given race of *H. glycines* were chosen selectively and were crushed to release eggs and larvae. Each seedling was inoculated

index = $1 - \sum \sum pi^2$ where different polymorphic loci are summed.

Only the fragments that were polymorphic among accessions and could be clearly scored were used in the data analysis. All bands having equivalent migration distance were given the same letter score. Genetic distances ($\mathrm{GD_R}$) among all possible pairs of accessions were estimated from a modification of Nei's similarity equation¹⁶ as used by Keim *et al.*⁸ in soybean. The proportion of similar RFLP loci, Sxy, between pairs of PI lines was estimated as 2Nxy/(Nx+Ny), where Nxy is the number of RFLP loci for which PI lines X and Y possess the same allele, Nx is the number of alleles identified in line X and Ny is the number of alleles

Table 1.	Reaction*	and index	of parasitism**	of	soybean	ΡI	lines	to	Н.	glycines	race	isolates
			1. 2: 3.	5	and 14							

Accession	Race I	Race II	Race III	Race V	Race XIV
PI398680	MS (56.0)	S (78.6)	MR (26.2)	S (67.5)	MS (42.5)
PI209332	MS (47.0)	S (70.0)	R (4.0)	R (7.0)	R (5.0)
PI404166	R (1.0)	R (6.0)	R (0.6)	R (0.4)	MR (10.0)
PI399061	MS (38.0)	MS (41.7)	MR (33.3)	R (7.8)	S (63.7)
Peking	R (1.8)	MR (19.7)	R (1.5)	R (0.9)	MR (28.1)
PI339868B	R (2.0)	MR (20.0)	R (0.8)	R (1.7)	MR (12.6)
PI437654	R (0.2)	R (1.0)	R (0.3)	R (0.4)	R (0.3)
PI437679	MR (22.0)	MR (14.3)	R (4.0)	R (0.1)	R (2.2)
PI84751	R (3.2)	MR (28.0)	R (0.2)	R (1.4)	MR (14.0)
PI90763	R (7.9)	R (2.3)	R (0.4)	R (0.1)	MR (26.8)
PI438342	MS (38.0)	MR (21.6)	MS (41.7)	R (1.7)	MS (32.3)
PI438496B	MR (30.0)	S (96.0)	R (2.3)	MS (35.0)	MS (41.0)
PI876311	MS (38.0)	S (67.3)	R (9.0)	MR (16.6)	MR (10.9)
PI437725	R (0.6)	MR (18.7)	R (0.4)	R (2.4)	MS (45.7)
PI438497	R (5.0)	MR (17.0)	R (0.4)	R (2.2)	MR (29.0)
PI88788	MS (39.0)	S (77.2)	R (5.4)	MS (46.0)	R (2.4)
PI404198A	R (0.7)	R (8.5)	R (1.4)	R (1.0)	MS (54.6)
PI404198B	R (2.0)	MS (44.2)	R (1.7)	R (1.4)	MR (23.6)
PI438503A	MS (44.0)	MS (57.0)	R (2.9)	MR (22.1)	R (9.0)
PI89008	MS (35.0)	S (66.4)	MR (17.9)	MR (17.8)	MR (17.8)
PI437690	R (0.8)	R (7.1)	R (0.0)	R (1.7)	MR (25.0)
PI438489B	R (0.4)	R (4.5)	R (0.6)	R (1.1)	R (8.0)
PI89772	R (1.5)	R (3.5)	R (0.2)	R (0.8)	MR (13.6)
PI416762	MR (16.0)	MS (47.1)	R (6.5)	R (5.7)	R (8.1)
PI437655	R (1.0)	S (69.0)	R (0.6)	R (3.0)	MR (11.5)
PI407729	MR (11.0)	MS (46.5)	MR (14.5)	R (6.3)	R (4.5)
PI424137B	S (61.0)	S (88.0)	MR (34.9)	MR (11.8)	MR (27.5)
PI424595	MS (58.0)	MS (45.2)	S (95.2)	R (9.9)	S (64.9)
PI437908	S (74.0)	S (80.9)	S (95.2)	MS (54.8)	S (84.7)
cv. Hutcheson	S (100)	S (100)	S (100)	MS (100)	S (100)

^{*0-9%} = Resistant; 10-30% = moderately resistant; 31-60% = moderately susceptible; $\ge 60\%$ = susceptible.

cluster analysis (Statistical Analysis Systems, Cary, NC) on the distance matrices.

The principal component analysis (PCA) was done by first calculating a correlation matrix among the markers. Eigenvalues and eigenvectors were then obtained from the correlation matrix and these were used to calculate the coordinates of each accession. The accessions were then plotted on the basis of these coordinates (data not presented).

The reaction of PI lines to races of *H. glycines* is reported in Table 1. Results indicate that several PI lines with known resistance to some races were also found to be resistant to additional nematode races (Table 1). For example, PI438489B was resistant to races 3 and 5 and in this study it was found additionally resistant to *H. glycines* races 1, 2 and 14. Of the 32 probes examined, 13 were polymorphic producing two to nine restriction fragment bands. We found that 40% of the probes detected variation among the 29 resistant and two susceptible soybean genotypes.

Both cluster and principal component analyses separated the accessions and cultivars into several groups. Cluster analysis has grouped genotypes based on the

proportion of marker alleles that accessions and cultivars have in common. Genotypes that have a high proportion of alleles in common included 'Peking' and PI339868B; PI437679 and PI84751; PI438342 and PI438496B; and PI437725 and PI438497 (Figure 1). These pairs are distinguishable but very closely related based on the RFLP data.

Based on both the cluster analysis and the principal component analysis (data not shown) most widely used genetic sources for *H. glycines* resistance have 'tight' grouping or clustering. These included Peking, PI88788, PI90763, and PI437654. A few of the resistance genes in both the accessions PI437654 and Peking have been recently mapped using RFLPs^{17,18}. Genetic analyses have indicated that very few non-allelic genes exist among these sources of resistance¹⁹, and their indiscriminate use in cultivar breeding appeared to have contributed to their genetic vulnerability. More virulent race populations of *H. glycines* are being reproduced which continue to infest the resistant cultivars.

Some PI lines with multiple race resistance that were genetically distant from the previously used sources were identified in this study (Table 1). These included

^{**}Index of parasitism = $\frac{\text{Average number of females per PI line}}{\text{Average number of females per cv. Hutcheson}} \times 100$

PI438489B, PI404198B, PI438503A, PI89772, PI404166, PI437908, PI209332 and PI437655. The resistance genes in these PIs will be mapped.

The results showed that greater diversity for markers existed among the PIs than the two cultivars used in this study. The two cultivars Essex and Hutcheson were also clustered away from the PIs in the dendrogram (Figure 1). This is expected, because the PIs are in general a diverse group of accessions collected in Asia and South America, whereas the cultivars were derived from a limited number of PIs introduced in USA at the turn of the century. In this study, no relationship was found in general, between geographical origin of PIs and the clusters obtained based on the limited number of probes used, but some degree of relationship was observed for spectrum of resistance. Both PI424595 and PI437908 susceptible to races 3 and 14 which clustered at the lower end of dendrogram have originated from South Korea.

Our study has demonstrated that an analysis of RFLP markers can be used to determine genetic relationships among PI lines of soybean. In the absence of typical pedigree information, DNA fingerprinting should be most useful in establishing their genetic relationship to develop appropriate populations for gene mapping studies.

- 1. Wrather, J. A., Chambers, A. J., Fox, J. A., Moore, W. F. and Sciumbata, G. L., Plant Dis., 1995, 79, 1076.
- 2. Riggs, R. D. and Schmitt, D. P., J. Nematol., 1988, 20, 392.
- 3. Rao Arelli, A. P. and Anand, S. C., Crop Sci., 1988, 28, 650.
- 4. Rao Arelli, A. P., Anand, S. C. and Wrather, J. A., Am. Soc. Agron. Abst., 1991, 113.
- 5. Rao Arelli, A. P., Wrather, J. A. and Anand, S. C., Plant Dis., 1992, 76, 894.
- 6. Miller, J. C. and Tanksley, S. D., Theor. Appl. Genet., 1990, 80,
- 7. Kesseli, R., Ochoa, O. and Michelmore, R., Genome, 1991, 34,
- 8. Keim, P., Beavis, W., Schupp, J. and Freestone, R., Theor. Appl. Genet., 1992, 85, 205.
- 9. Rao Arelli, A. P. and Webb, D. M., Am. Soc. Agron. Abst., 1993,
- 10. Anand, S. C. and Rao Arelli, A. P., Crop Sci., 1989, 29, 1181.
- 11. Rao Arelli, A. P., Matson, K. W. and Anand, S. C., Plant Dis., 1991, 75, 594.
- 12. Keim, P. and Shoemaker, R. C., Soybean Genet. Newsl., 1988, 15. 147.
- 13. Sambrook, J., Fritsch, E. F. and Maniatis, T. (eds), Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, NY, 1989, pp. E.5.
- 14. Anonymous, Univ. of Missouri Maize RFLP Procedures Manual, 1995, p. 25.
- 15. Feinberg, A. P. and Vogelstein, B., Anal. Biochem., 1983, 137,
- 16. Nei, M. and Li, W. H., Proc. Natl. Acad. Sci. USA, 1979, 76,
- 17. Webb, D. M., Baltazar, B. M., Rao Arelli, A. P., Schupp, Jr., Clayton, K., Keim, P. and Beavis, W. D., Theor. Appl. Genet., 1995, 91, 574.
- 18. Qiu, B., Rao Arelli, A. P. and Sleper, D. A., Am. Soc. Agron. Abst., 1995, 177.

19. Rao Arelli, A. P., Anand, S. C. and Wrather, J. A., Crop Sci., 1992, 32, 862.

ACKNOWLEDGEMENTS. We thank Dr R. C. Shoemaker, USDA-ARS, Ames, IA for providing DNA probes used in this study and Dr Pat Byrne, USDA-ARS, Columbia, MO for scientific review. This research was supported by grants from Missouri Soybean Merchandising Council 9510176-1 and USDA-CSRS-93-34113-8498.

Received 12 January 1996; revised accepted 3 July 1996

Aeolian deposition of Arabia and Somalia sediments on the southwestern continental margin of India

Onkar S. Chauhan

Geological Oceanography Division, National Institute of Oceanography, Dona Paula, Goa 403 004, India

Kaolinite, smectite, illite and chlorite as major clay minerals and palygorskite and gibbsite in minor quantities have been recorded from the slope of southwestern continental margin of India. Contribution of kaolinite, smectite and gibbsite is from peninsular India through fluvial discharge. Since formation of palygorskite calls for an arid and hot climate and saline conditions, occurrence of this clay mineral in the sediments of the study area documents aeolian sediment contribution from Arabia and Somalia by the Arabian northwesterly winds.

Studies of marine clays are a significant tool to determine sources, sediment dynamics and environment of deposition1-4. The climate and geology of the source area3 largely dictates the type of clay species supplied. By and large, characteristic clay minerals of different climatic and geological settings have been identified 1,3.

Studies of clay mineral variations in the western continental margin of India mainly suggest two important sources of the clays. Illite and chlorite are reported to be mostly contributed by the Indus River and the low salinity Bay of Bengal Waters (BBW), intruding into the southwestern continental margin during November-January⁵⁻¹¹. Kaolinite, gibbsite and smectite (and minor amounts of illite) are produced due to intense chemical weathering of Indian subcontinent, and are contributed from the adjacent landmass⁵⁻⁷. Two contrasting opinions exist about the dispersal of these clay minerals. Ramaswamy and Nair9 have reported a lack of cross-shelf sediment transport and have suggested an along shelf transport of the sediments brought by the major fluvial