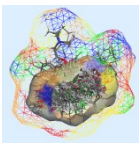


Computer Lab, Session 7



Virtual screening

Download Thrombin_VS.tar.gz from course website to Lab7 folder.

Unzip and untar file:

```
tar xzf Thrombin_VS.tar.gz
```

Change into Thrombin_VS folder:

```
cd Thrombin_VS
```

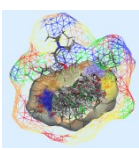
and open all mol2 files in PyMOL:

```
pymol *.mol2
```

Generate ligand library (as in Lab6):

```
AutoDockVina → Export ligand library and save as Thrombin_VS  
library
```

Next, perform docking with similar settings as in Lab6 (use again Lab6_0.pse file for protein template):



Virtual screening

AutoDockVina Plugin

Choose target protein and ligand library for AutoDock simulation

Project definition name

Base directory:

Project subdirectory:

Ligand library

Path to ligand library:

Protein selection

☒ Protein from file [PDB,MOL2,MOL]

Path to protein file:

☐ File containing multiple protein structures [NMR-PDB]

Path to protein file:

☐ Trajectory from Amber

Path to topology file:

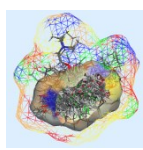
Path to trajectory file:

☒ Protein in current PyMol session

Objects to select from:

Protein preparation

☐ Let autodock change protonation states



Virtual screening

AutoDockVina Plugin

Search volume | Flexible residues | Output settings

Center of box

Definition based on x,y,z coordinates

Center (x): 17.1
-10.0 0.0 10.0 20.0 30.0

Center (y): -13.7
-30.0 -20.0 -10.0 0.0 10.0 20.0

Center (z): 23.1
-10.0 0.0 10.0 20.0 30.0 40.0

Definition based on center-of-mass of user selection

Selection sele Determine new box coordinates

Size of box

Definition based on x,y,z coordinates

Box length (x): 23.0
0.0 10.0 20.0 30.0 40.0 50.0 60.0 70.0 80.0 90.0

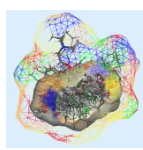
Box length (y): 19.0
0.0 10.0 20.0 30.0 40.0 50.0 60.0 70.0 80.0 90.0

Box length (z): 18.0
0.0 10.0 20.0 30.0 40.0 50.0 60.0 70.0 80.0 90.0

Definition based on user selection

Selection sele Radius around selection 5.0 Determine new box dimensions

OK Cancel



Virtual screening

After docking simulations are finished
import results:

AutoDockVina → Import results

select Monitor.aut in folder
project_VS_client

For statistical analysis: Open Excel

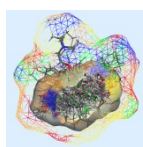
Create Excel table with the following
format using the AutoDock Results
dialog:

The PyMOL Molecular Graphics System

AutoDock results

Ligand name	Binding free energy	
decoy5	-10.10	Show details
active4	-9.80	Show details
decoy10	-9.70	Show details
active2	-9.60	Show details
decoy4	-9.40	Show details
active1	-8.90	Show details
decoy2	-8.90	Show details
active3	-8.80	Show details
decoy3	-8.70	Show details
decoy9	-8.60	Show details
decoy1	-8.50	Show details
active5	-8.40	Show details
decoy7	-7.20	Show details
decoy8	-7.20	Show details
decoy6	-7.20	Show details

Close

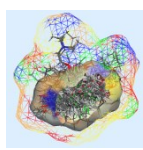


Virtual screening

Excel table:

Active or decoy in ranked list	Rank %	Actives found at rank %	Ideal	Random
	0	0	0	0
0	6.666666	=# of actives in first column*100/5	20	6.666666
1	13.33333	...	40	13.33333
0	20.00000	...	60	20.00000
...

Plot “Actives found at rank %”, “Ideal”, “Random” as a function of “Rank %” and compute enrichment factor at 0, 6.66666%, 13.3333%, ...



Discussion

Case study (**Use software from course**):

Based on the crystal structure of a thrombin-ligand complex (1mu6) you have virtually screened a library of 10000 compounds using Autodock Vina with a single solution as output. You want to identify possible new lead structures and optimize their binding affinity.

Please, discuss the following issues:

1. Propose ideas to validate the outcome of your screen; in particular, how can you decide how many compounds from the top-ranked list you should consider as possible active compounds? How can you refine the outcome of the screen?
2. Plan a possible strategy to optimize the affinity of the lead compound using discussed software. Take into consideration a reasonable balance between accuracy and efficiency.