



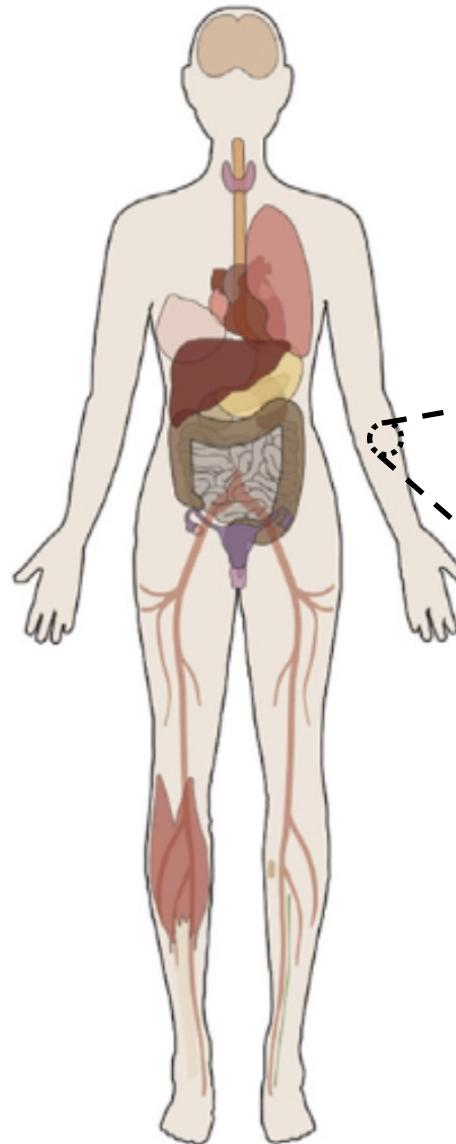
# Tilted-CCA: Quantifying common and distinct information in multi-modal single-cell data via matrix factorization

Kevin Lin  
 : linnykos

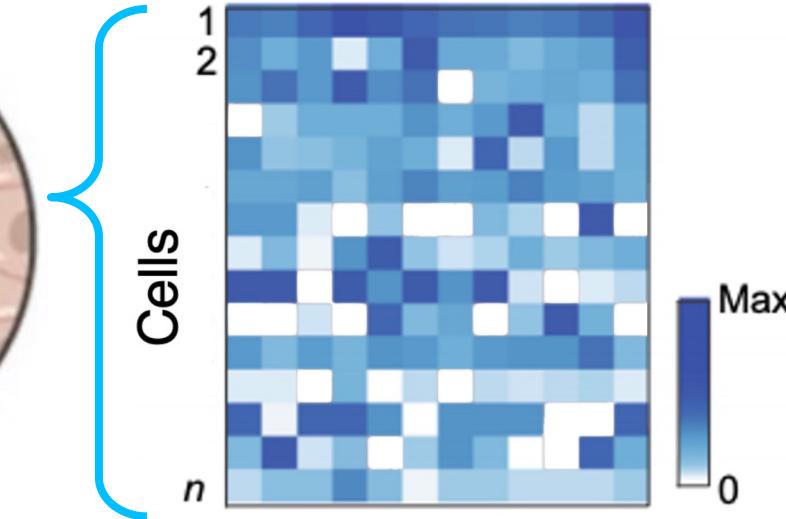
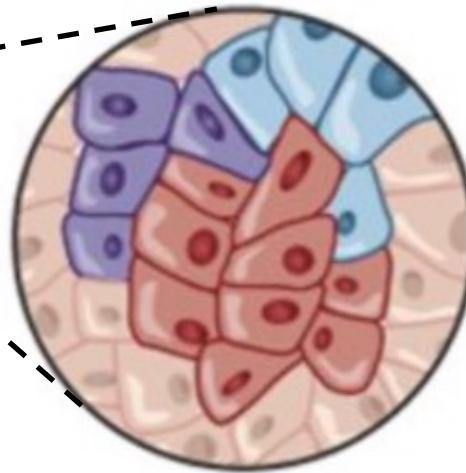
January 3,  
2023



My body of work focuses on advancing statistical ideas that are inspired by advancements in biomedical technology.



Single-cell RNA-seq  
(circa 2015)



Measuring “how active a gene is in a particular cell”

## Previous statistical methods & theory I've developed for single-cell RNA-seq:

- Relevant for denoising single-cell data (Matrix factorization)
  - K. Lin, H. Liu, K. Roeder (JASA 2021)
  - K. Lin, J. Lei, K. Roeder (JASA 2022)
- Relevant for answering biological questions (Network methods and changepoint detection)
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However, our talk today is inspired by newer biomedical technology.

Our story today starts with a newer technology:

FOCUS | EDITORIAL

## Method of the Year 2019: Single-cell multimodal omics

Multimodal omics measurement offers opportunities for gaining holistic views of cells one by one.

Teichmann, Efremova. Nature Methods (2020)

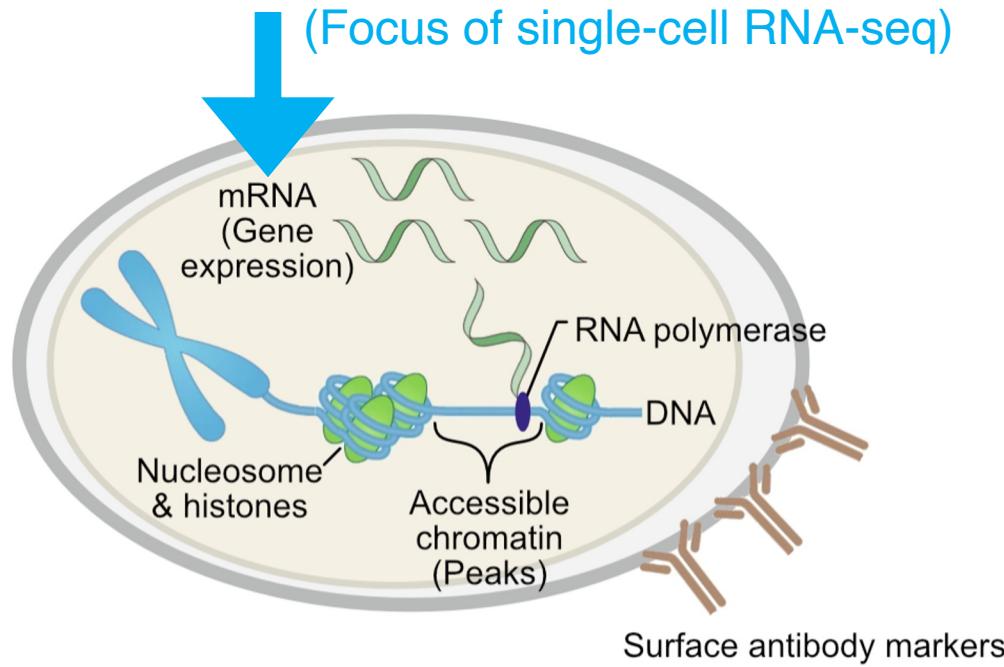
FOCUS | COMMENT

## Single-cell multimodal omics: the power of many

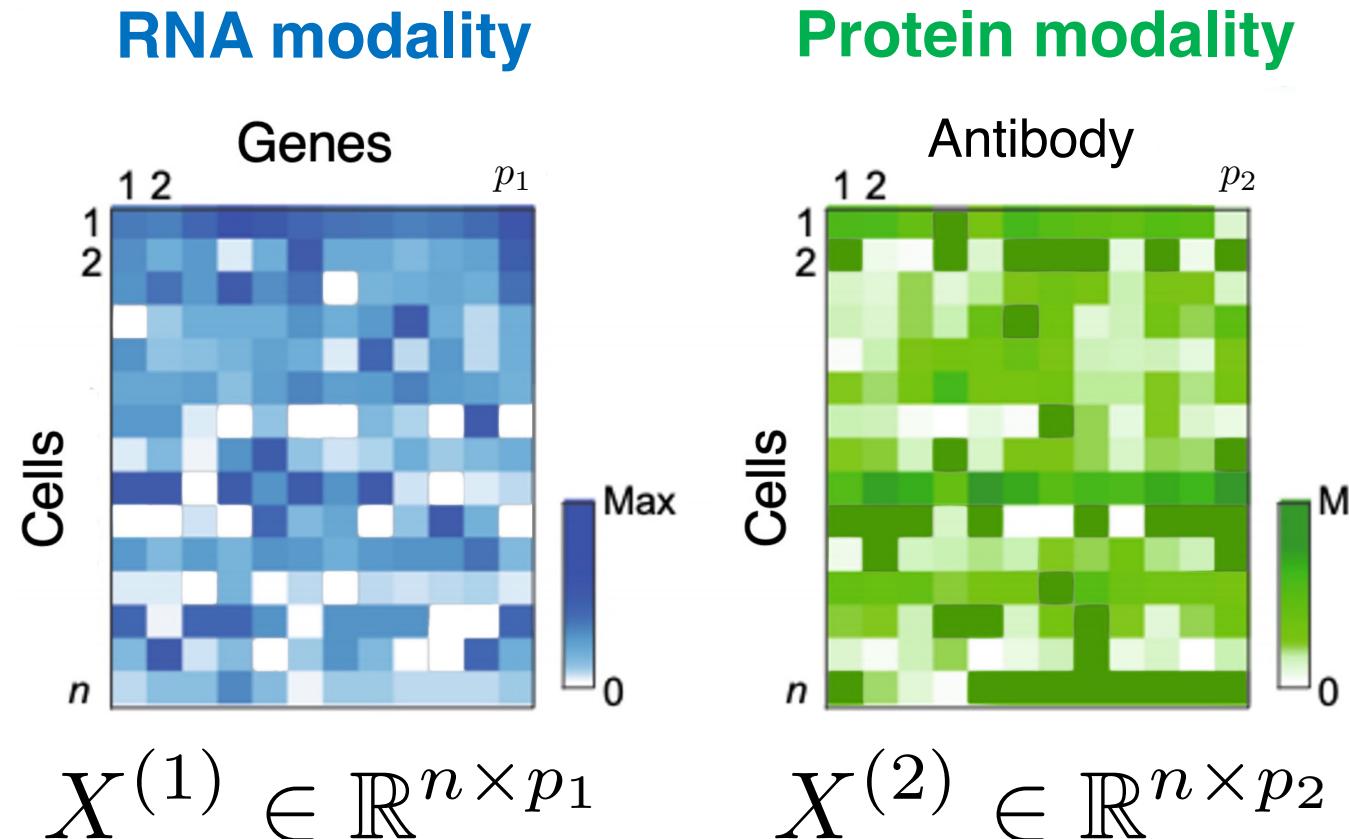
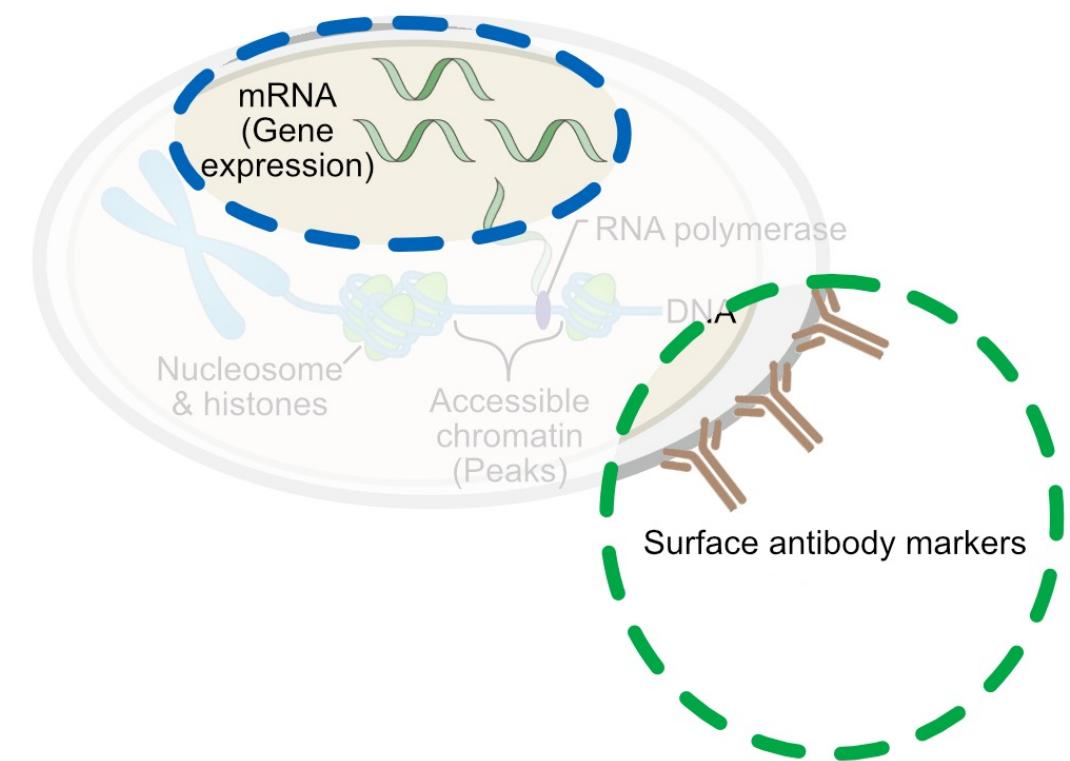
Advances in single-cell genomics technologies have enabled investigation of the gene regulation programs of multicellular organisms at unprecedented resolution and scale. Development of single-cell multimodal omics tools is another major step toward understanding the inner workings of biological systems.

Zhu, Preissl, Ren. Nature Methods (2020)

Our story today starts with a newer technology: Single-cell multi-modal (i.e., “multi-view”) sequencing (circa 2020).



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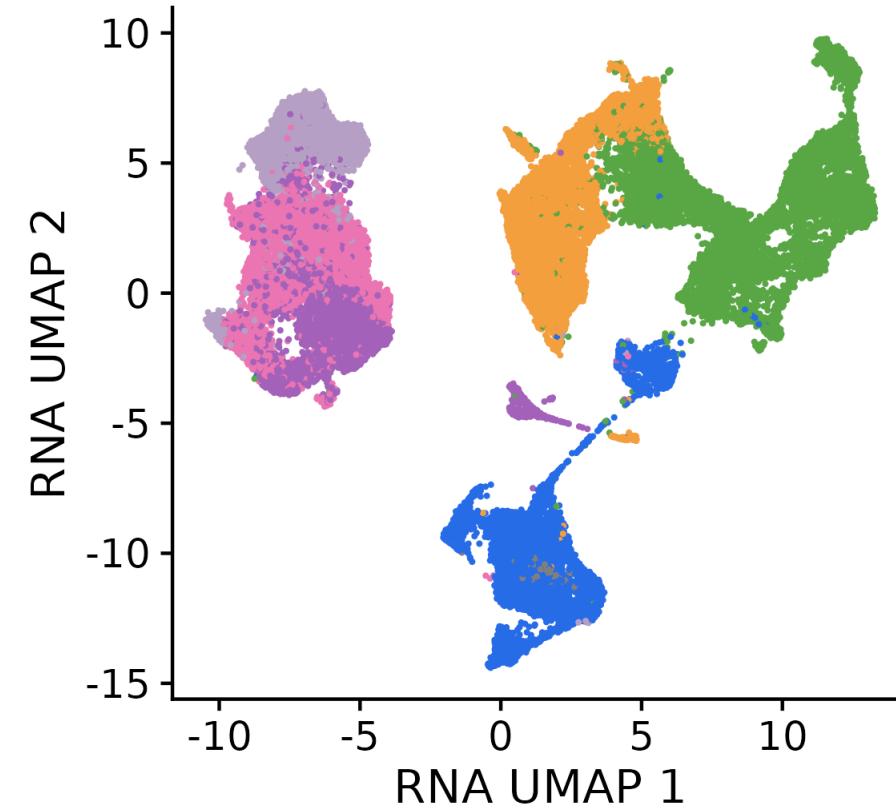


**Central question:** What “information” is unique to a modality, or represents the **coordination** between both modalities?

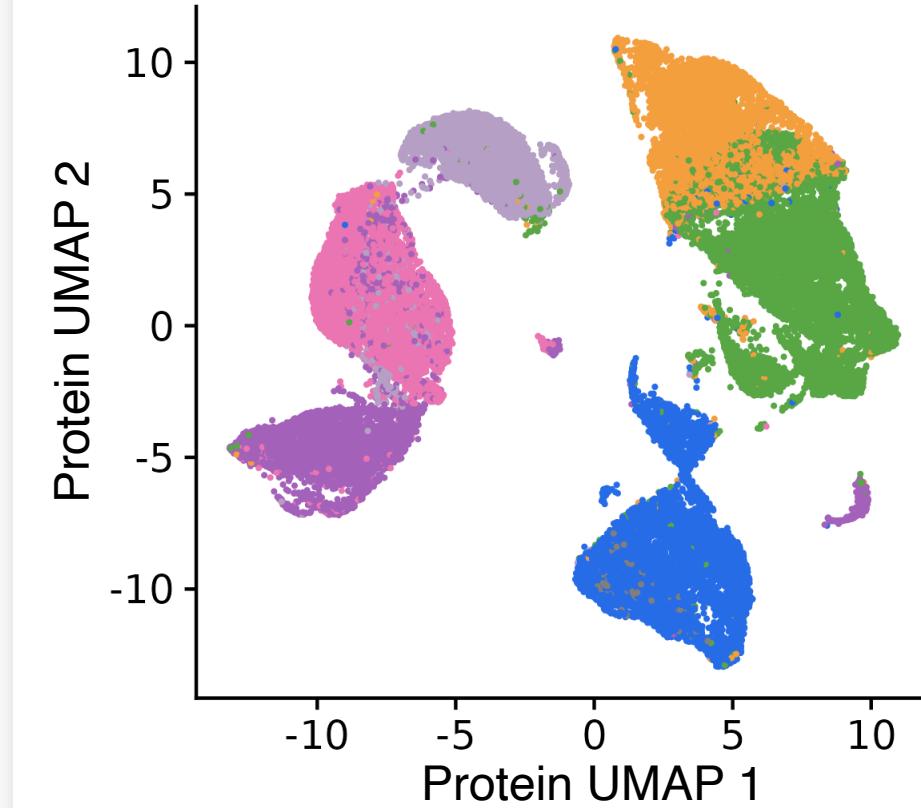
What “information” is **unique** to a modality, or represents the **coordination** between both modalities?

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**RNA modality (400+ genes)**



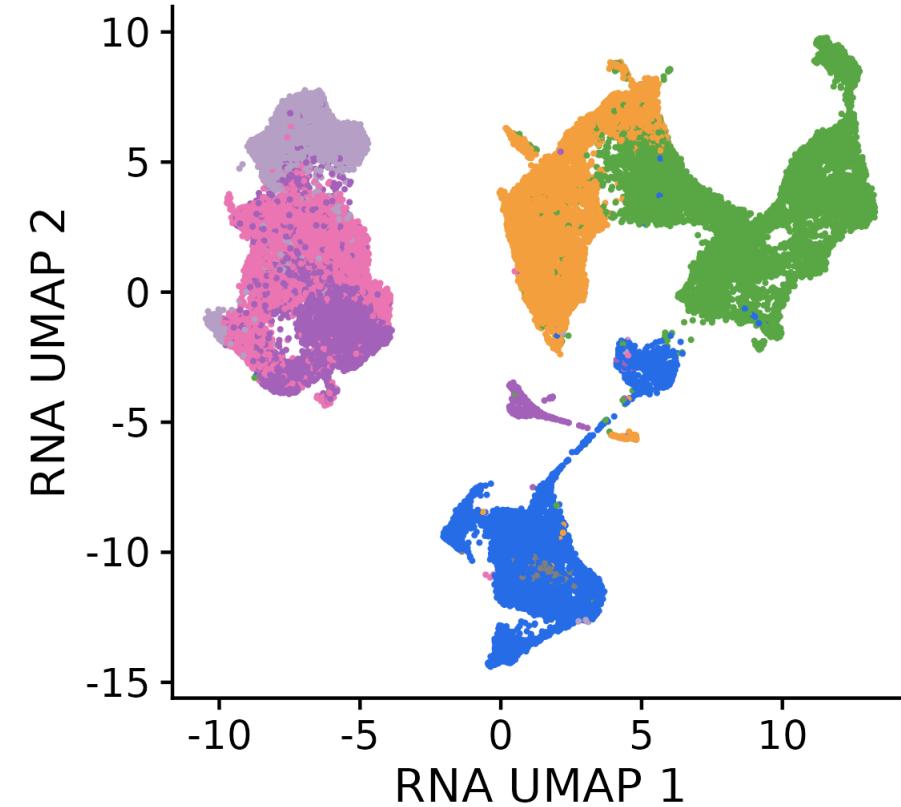
**Protein modality (97 antibodies)**



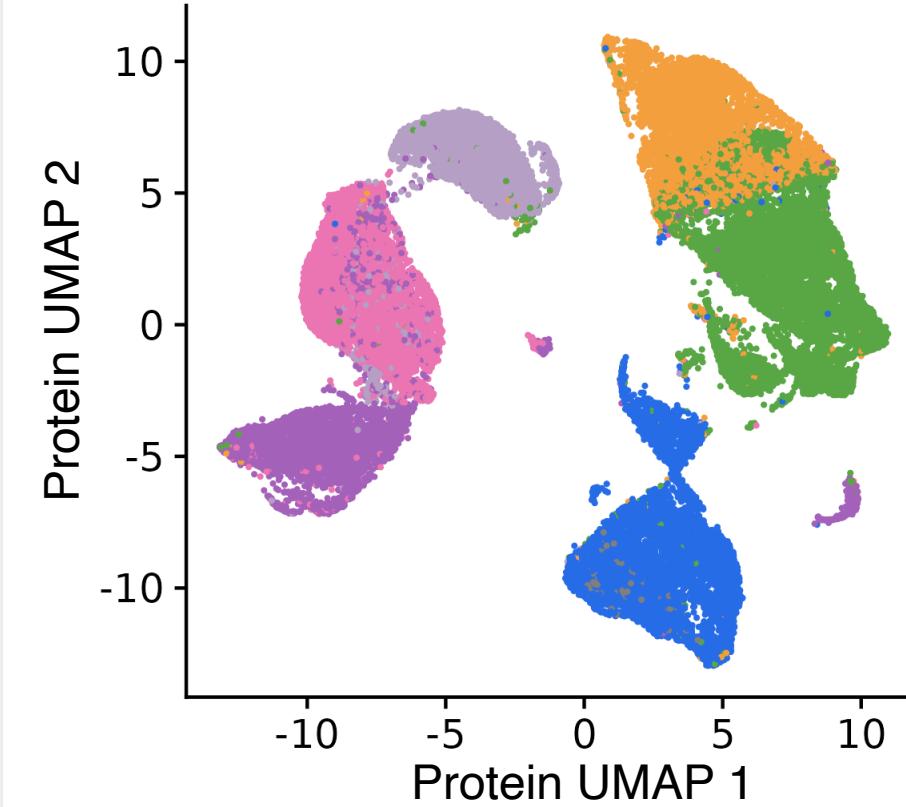
**Human bone marrow** (AbSeq, Triana et al., 2021), 49000+ cells (colored by annotated cell type)

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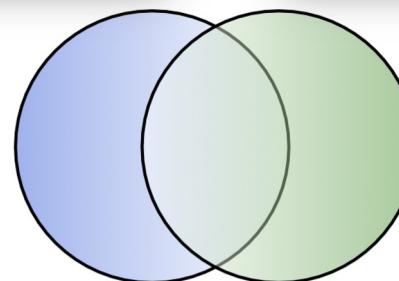
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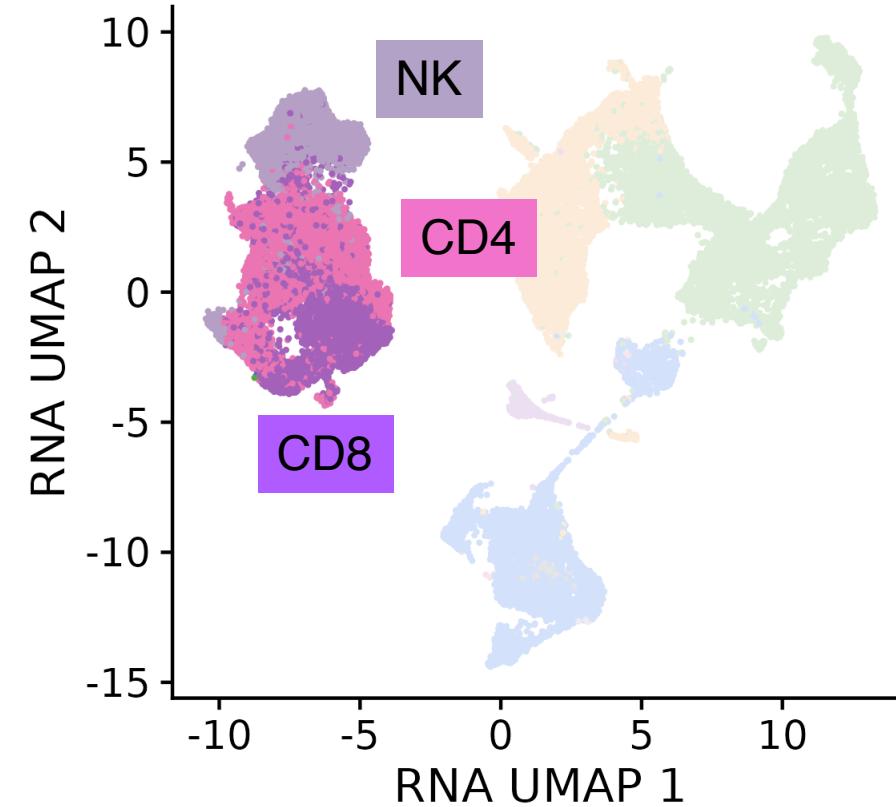


Venn diagram of geometry (“information”):

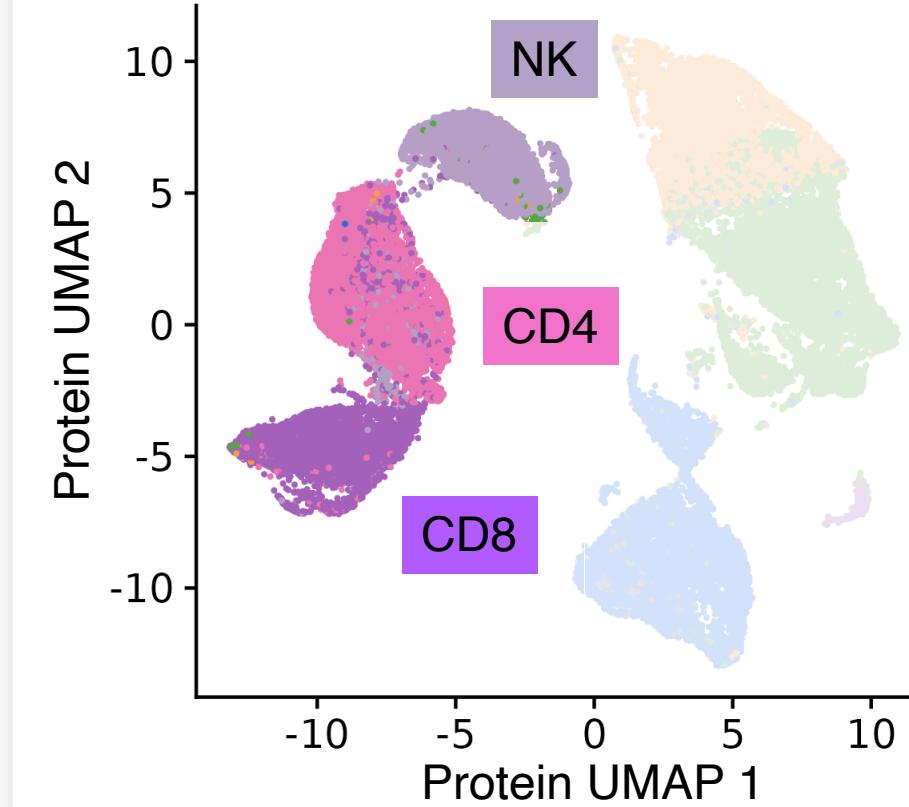


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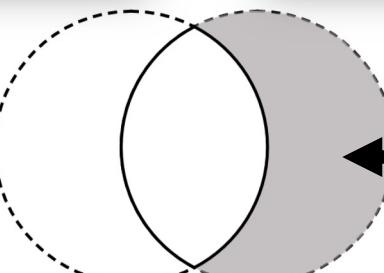
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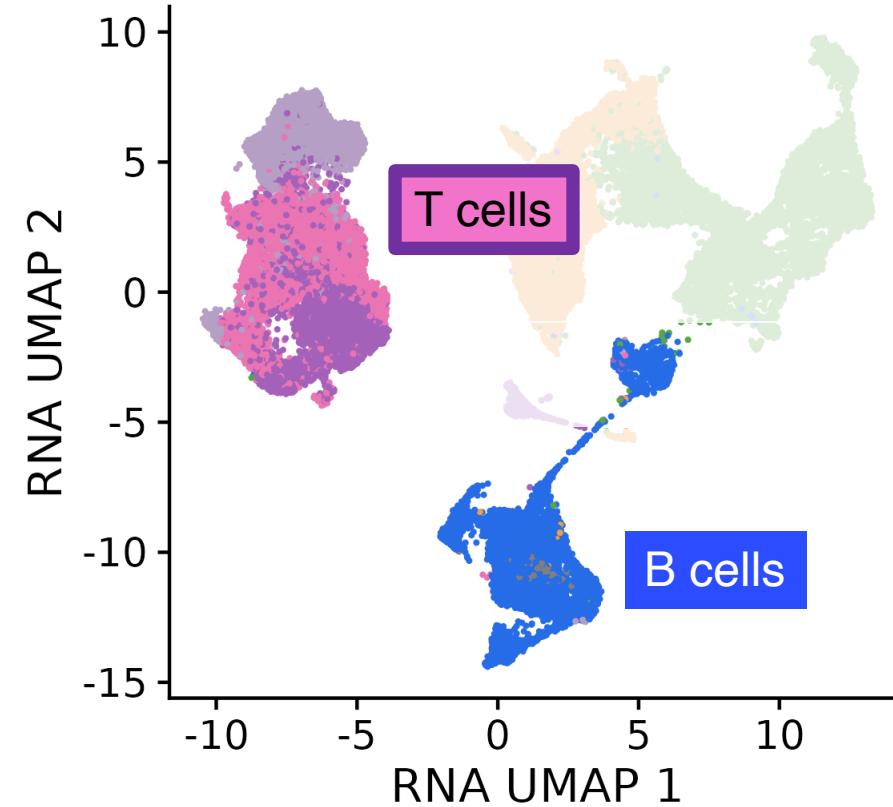
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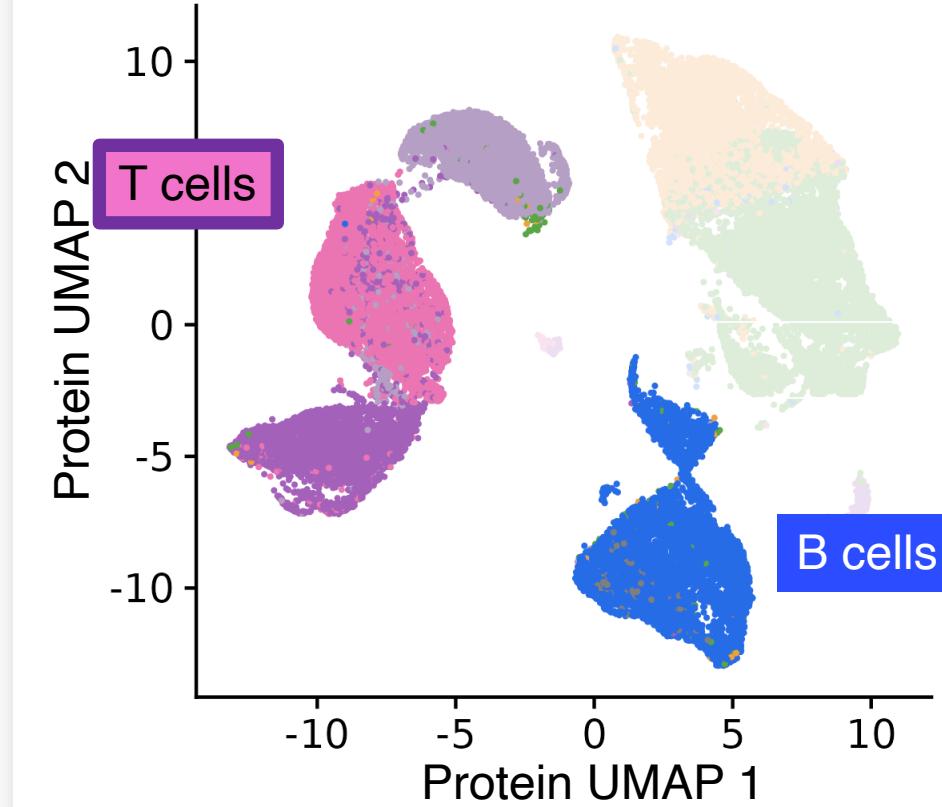
Cell-type separation  
unique to proteins

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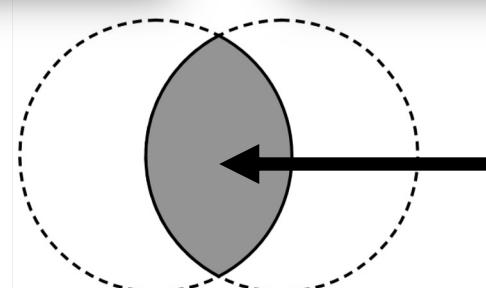
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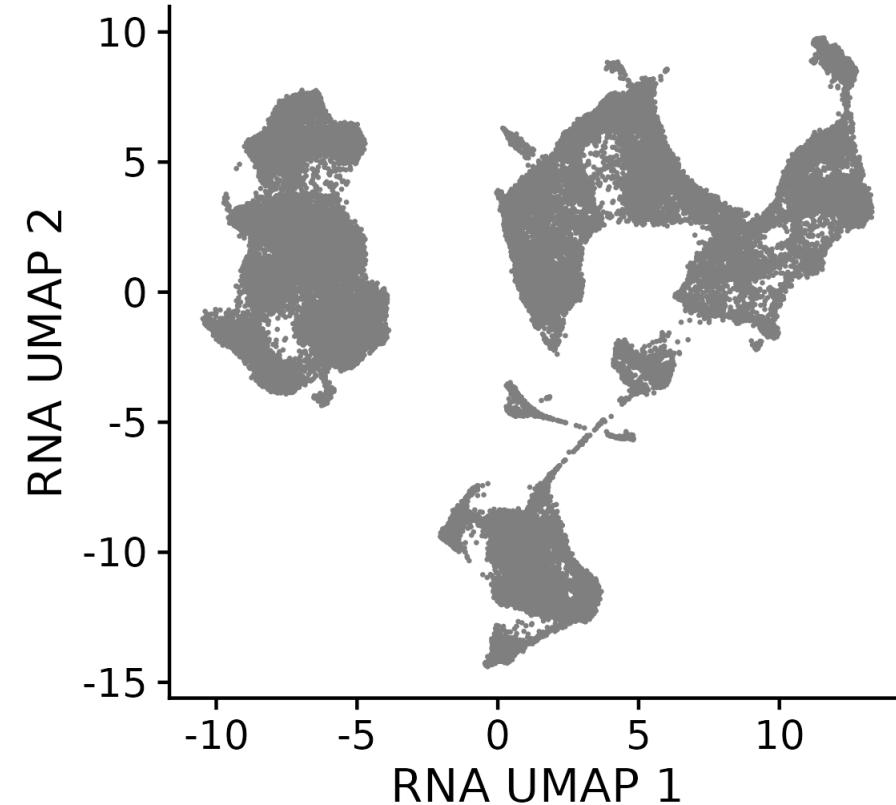
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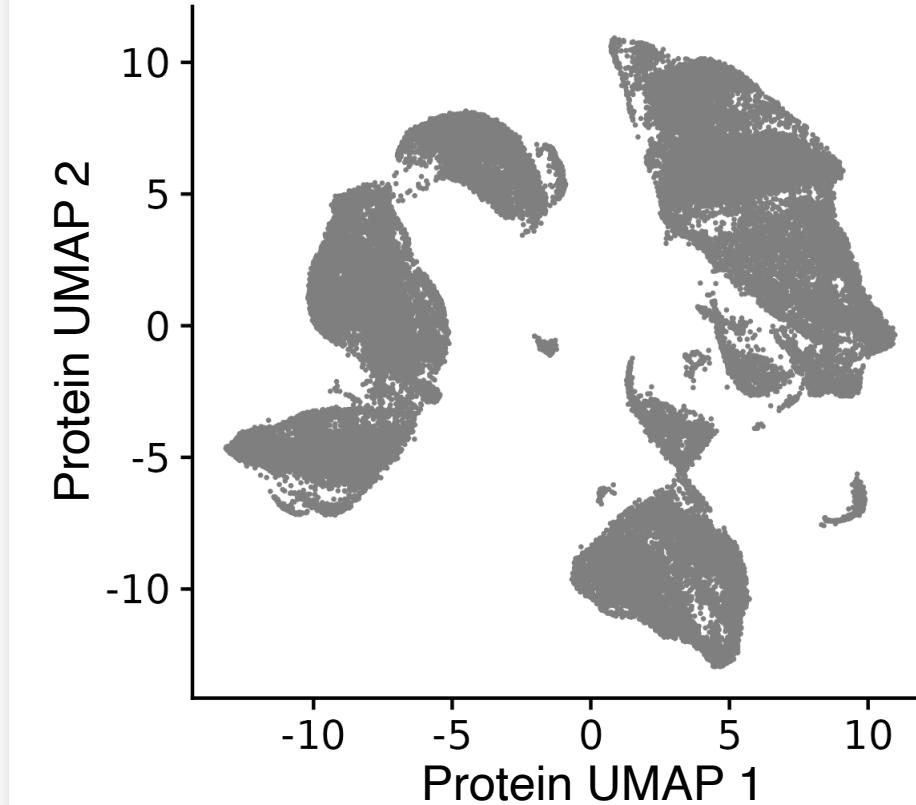
Between-modality coordination

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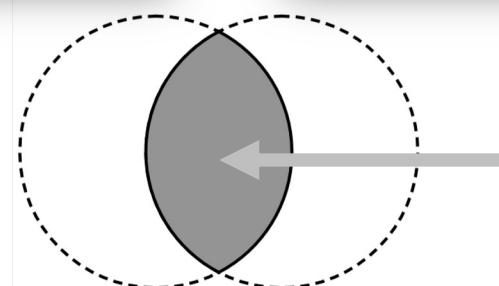
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Venn diagram of  
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(No cell-type information)



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**Statistical goal:** Develop a new matrix factorization framework for multi-modal data based on shared/unique geometry to answer the following biological questions.

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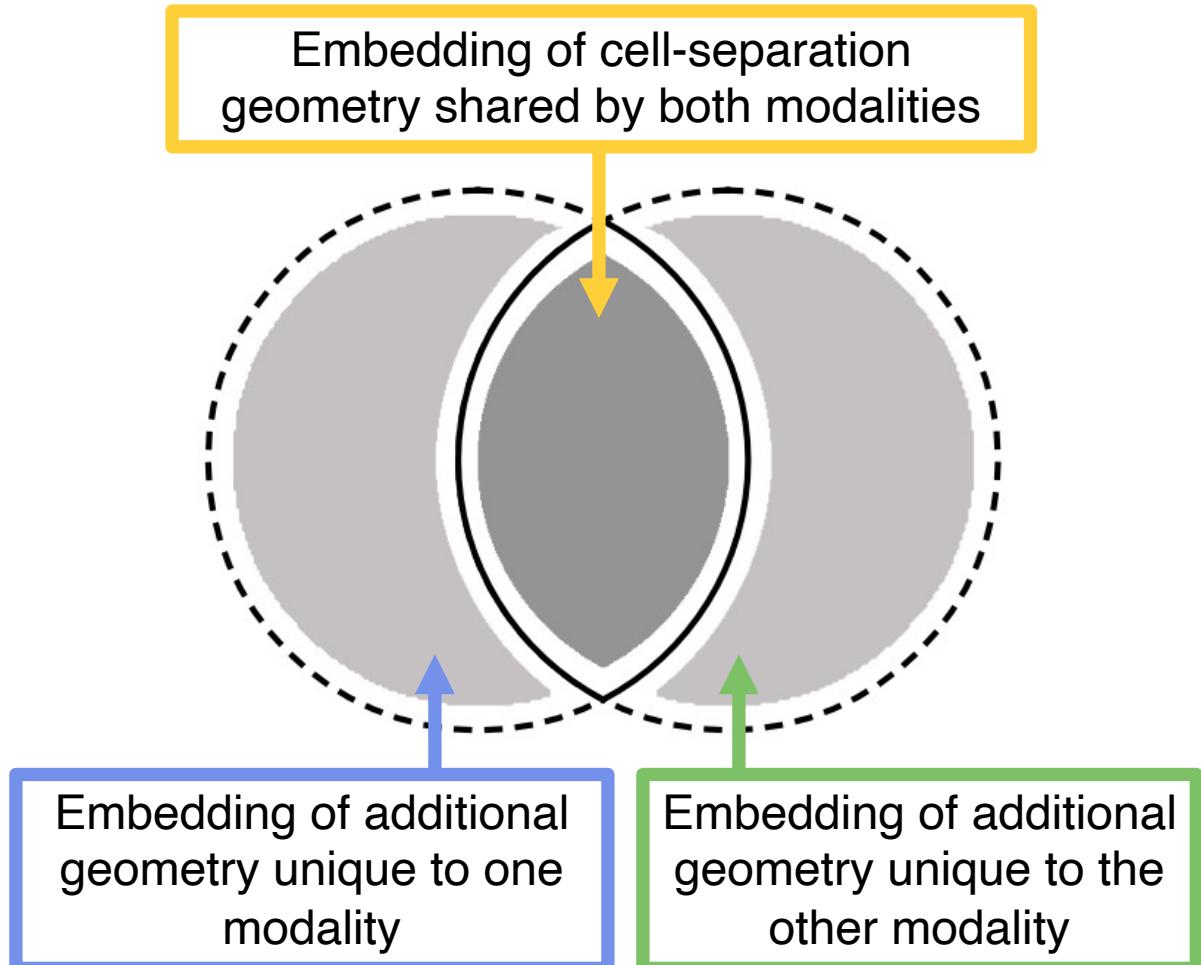
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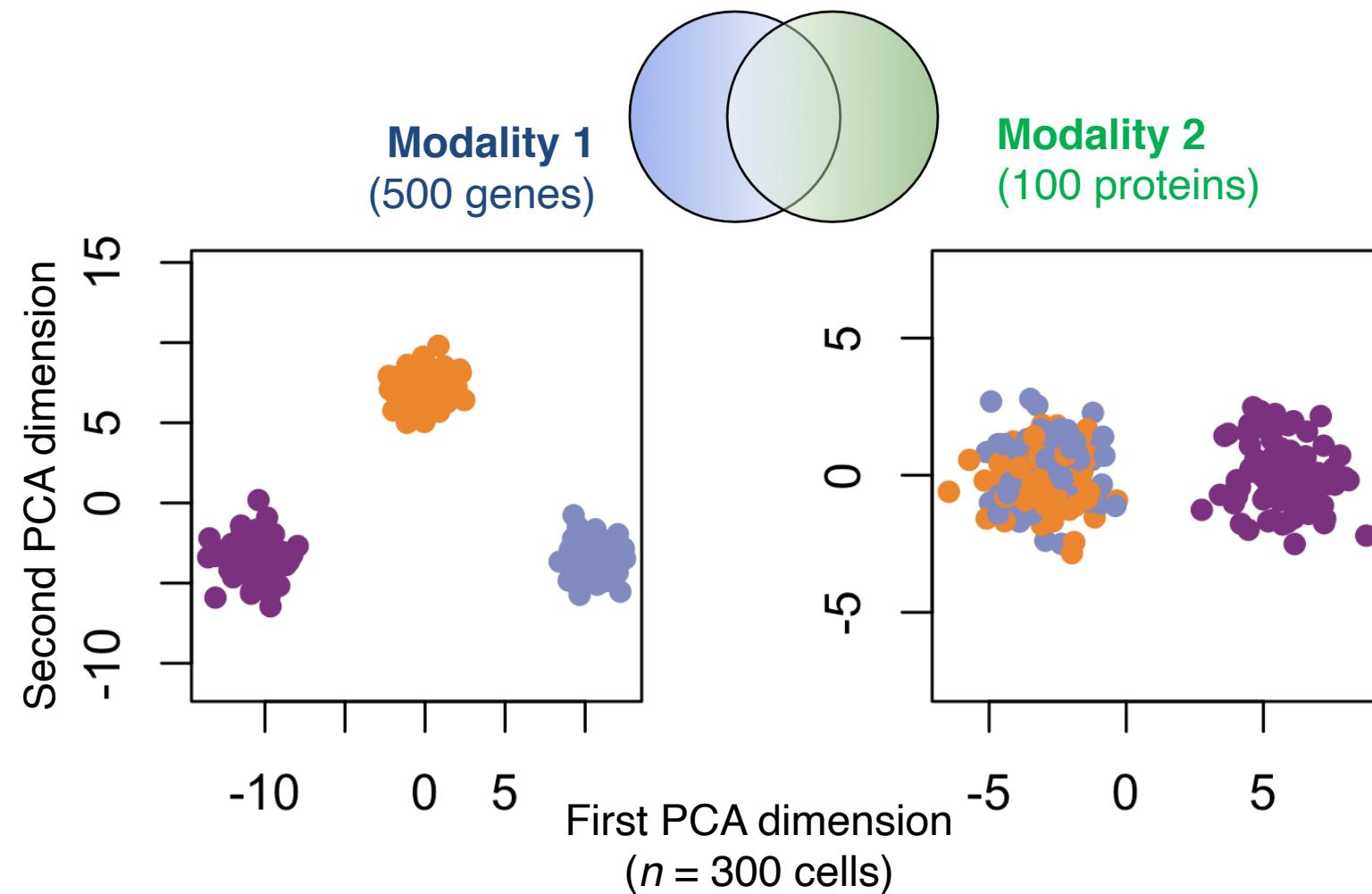
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Our method inspires new theoretical questions and is applicable to any multi-modal dataset.

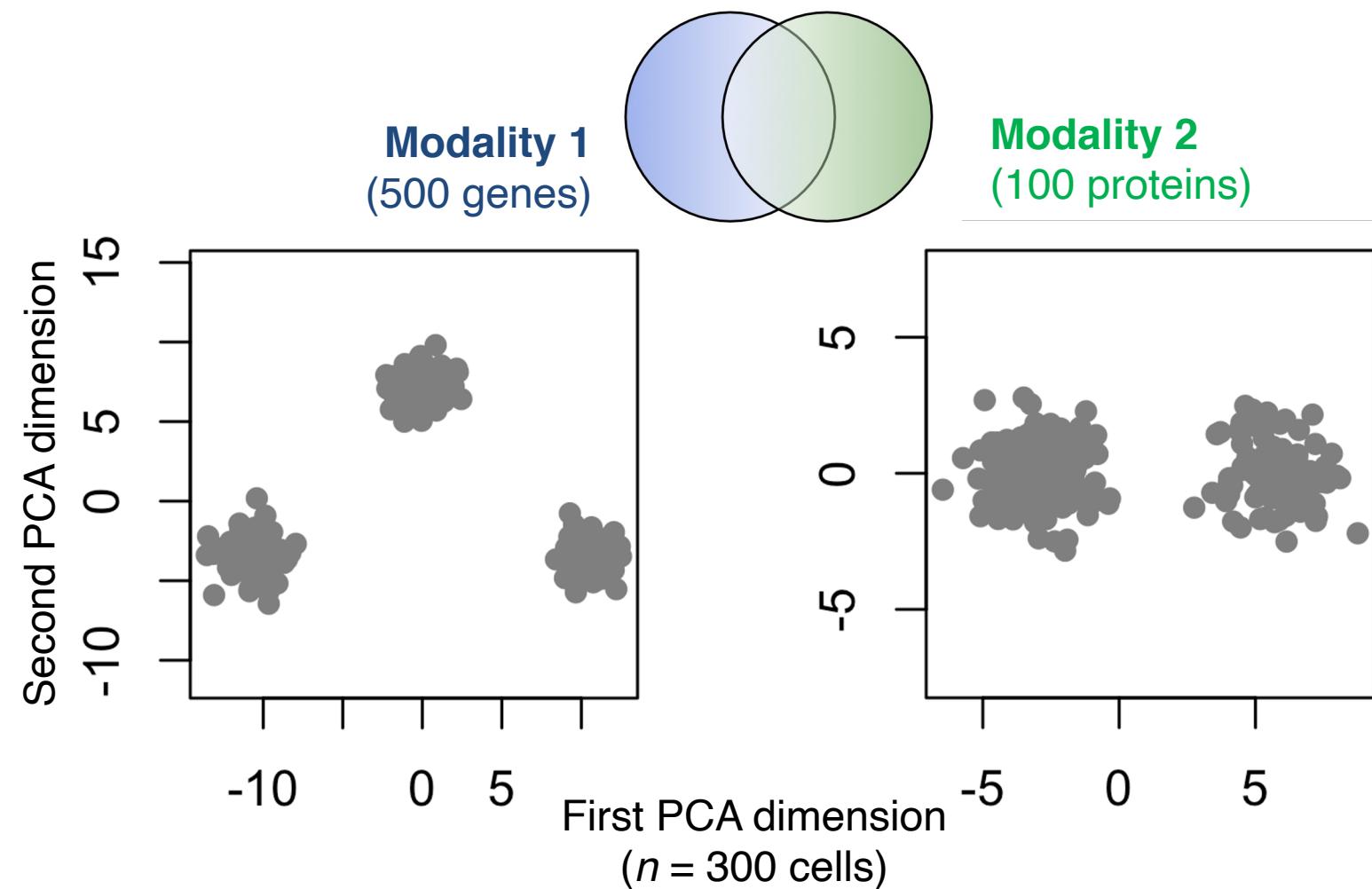
To start thinking about how to define geometry shared/unique to each modality, consider a toy example where one modality has more “information” than the other.

- Geometry = “Information”  $\approx$  “Density clustering”



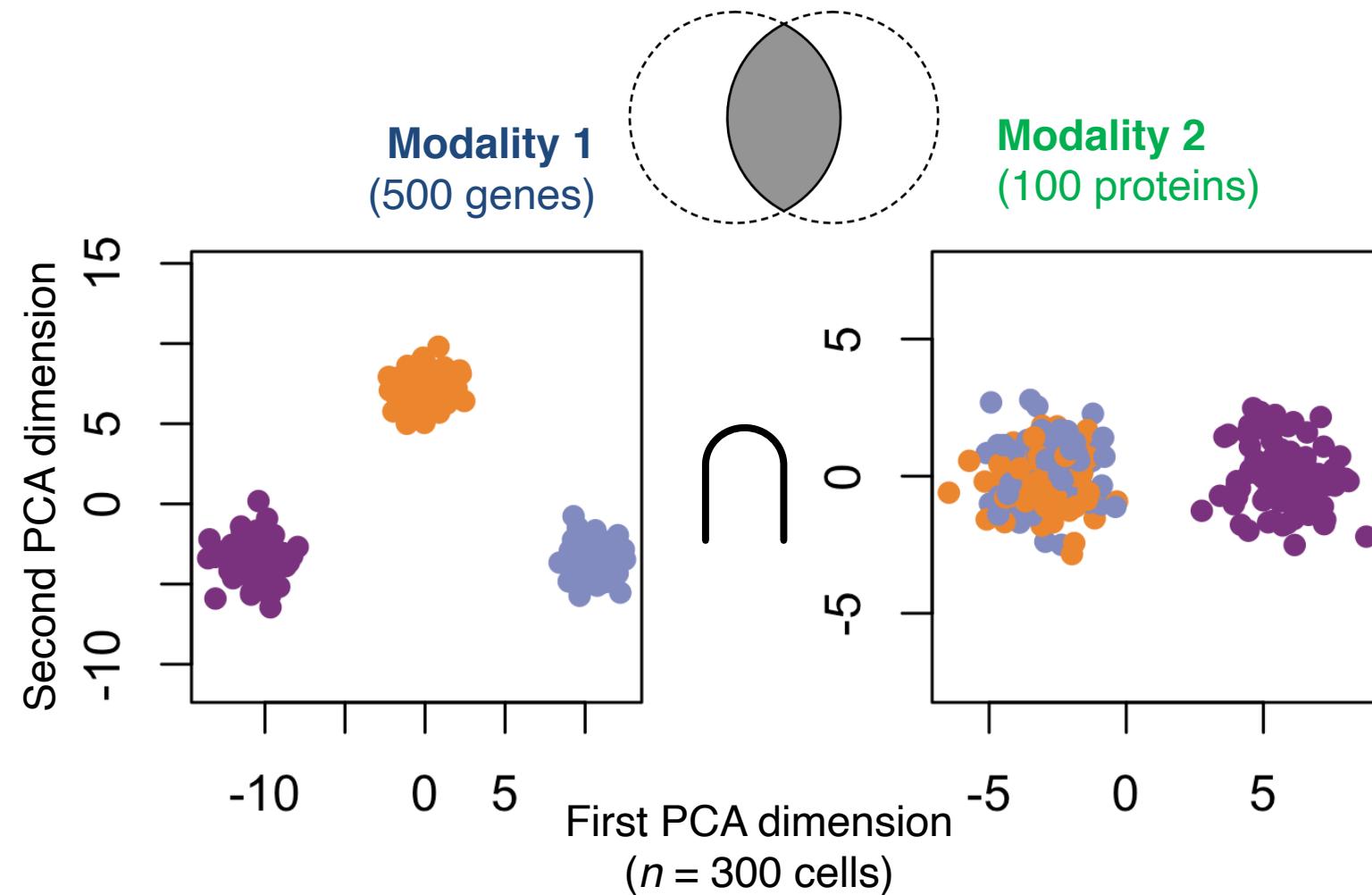
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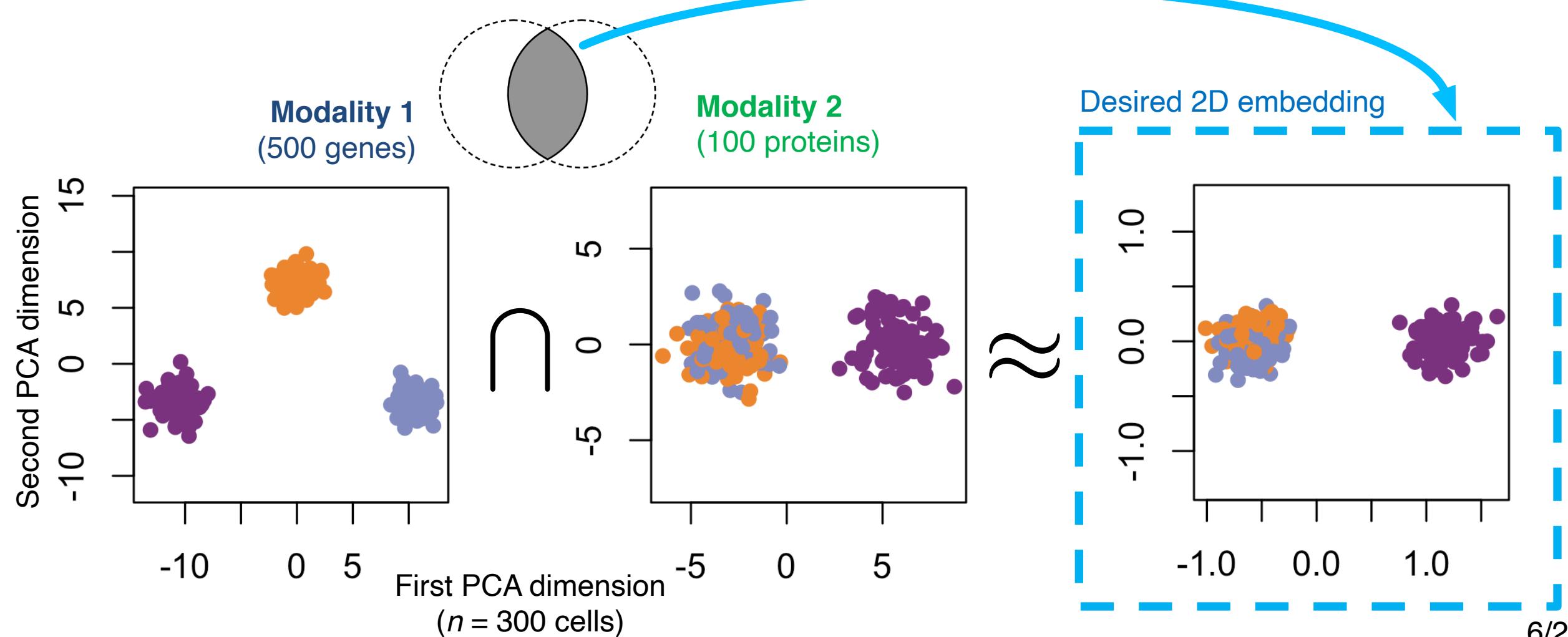
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(a phrase we coined for  
multimodal matrix factorization)

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Consider Consensus PCA (Wold et al., 1987):

$$\widehat{L} = \arg \min_{L \in \mathbb{R}^{(p_1+p_2) \times r}} \left\| [X^{(1)} ; X^{(2)}] - [X^{(1)} ; X^{(2)}] LL^\top \right\|_F^2$$

Resulting embedding via SVD:

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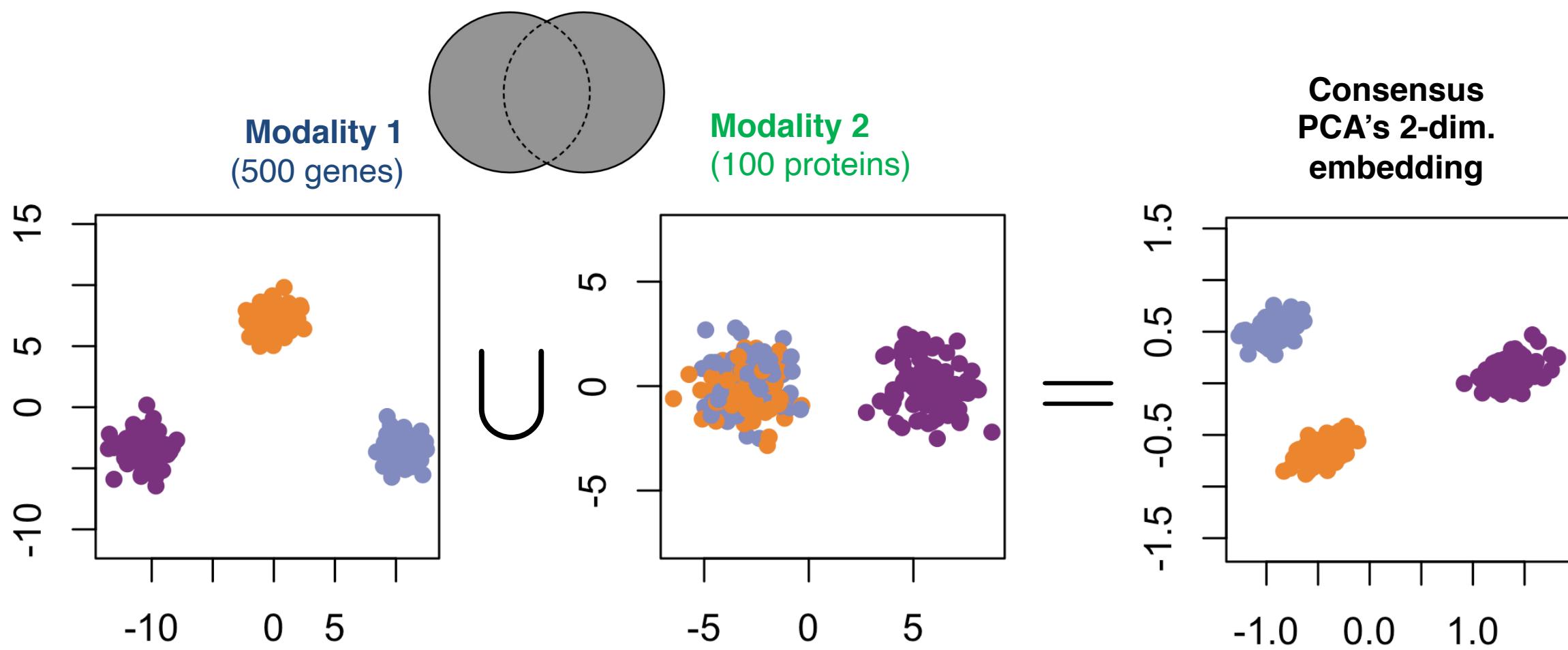
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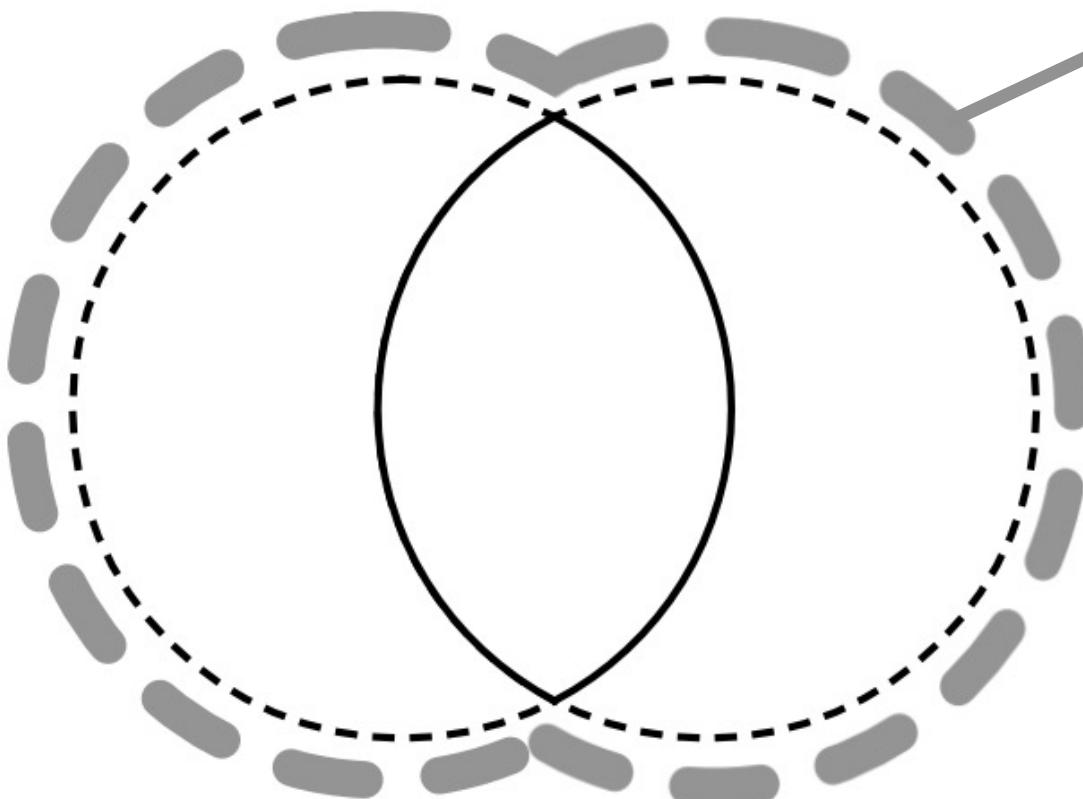
Low-dimensional embedding

**Observation:** The embedding combines the axes of variation from each modality (i.e. the “union”).

Let's see the “union of information” in toy example:



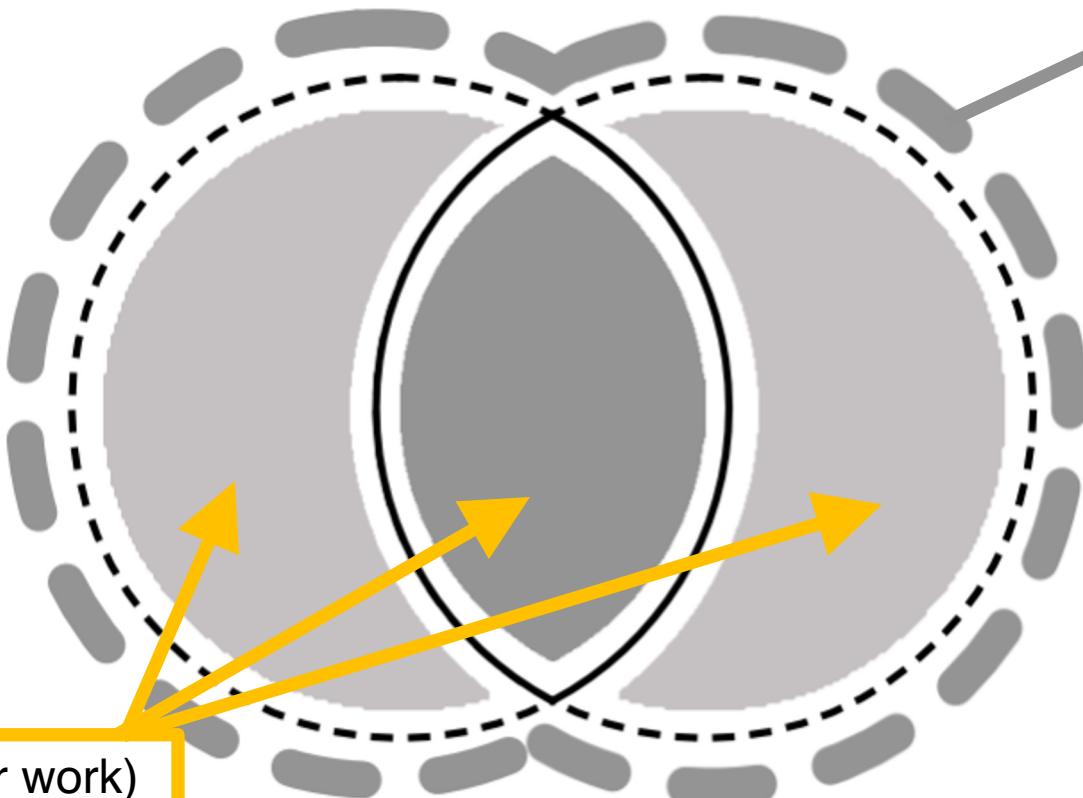
**Our contribution:** A matrix factorization for multi-modal data that separates of “intersection” and “unique” of information, as opposed many existing methods that represent the “union” of information.



- Consensus PCA (Wold et al., 1987)
- scAI (Nie et al., 2020)
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**Cell embeddings for the “union” geometry:** Useful for making an “atlas” across both modalities

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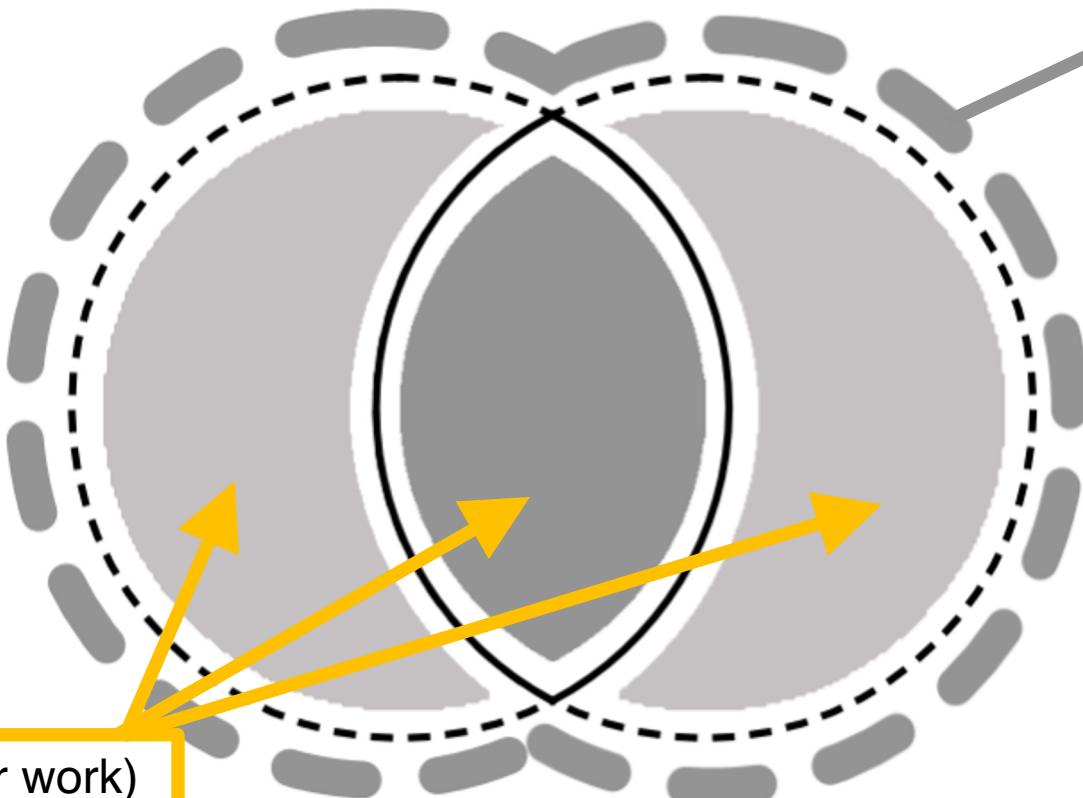
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Tilted-CCA (Our work)

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Useful to understand the coordination between modalities

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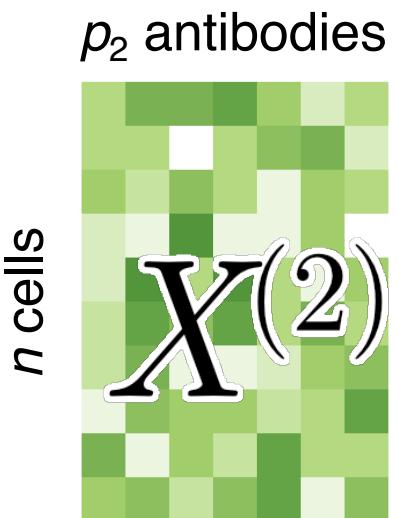
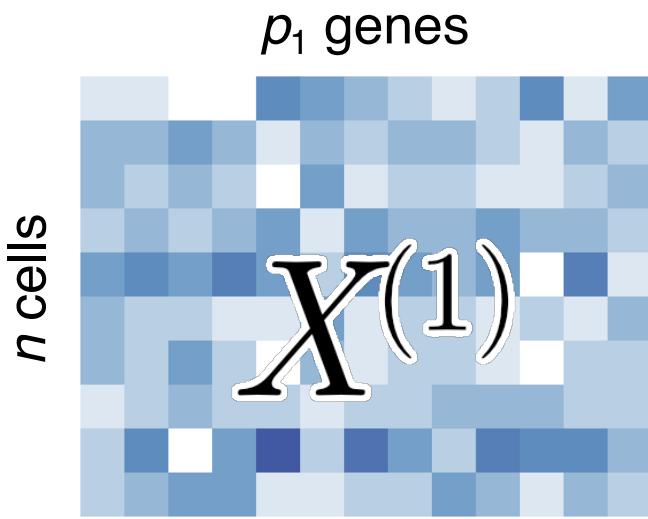
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## **Statistical method:**

New dimension-reduction framework for multi-modal data,  
not about denoising but instead the geometry

## Proposed model for multi-modal data: Capturing the “shared” geometry



(Assumed to be sufficiently preprocessed)

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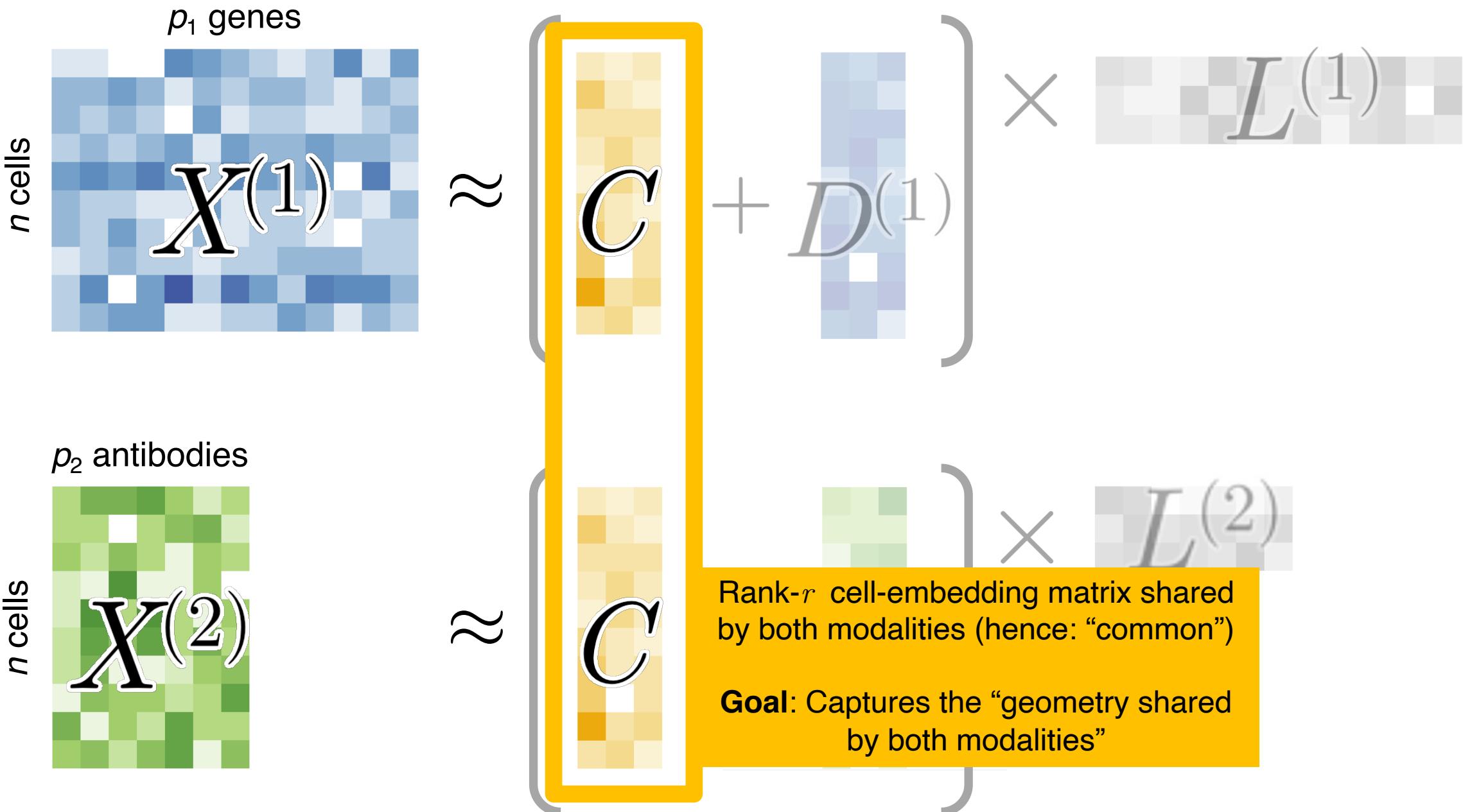
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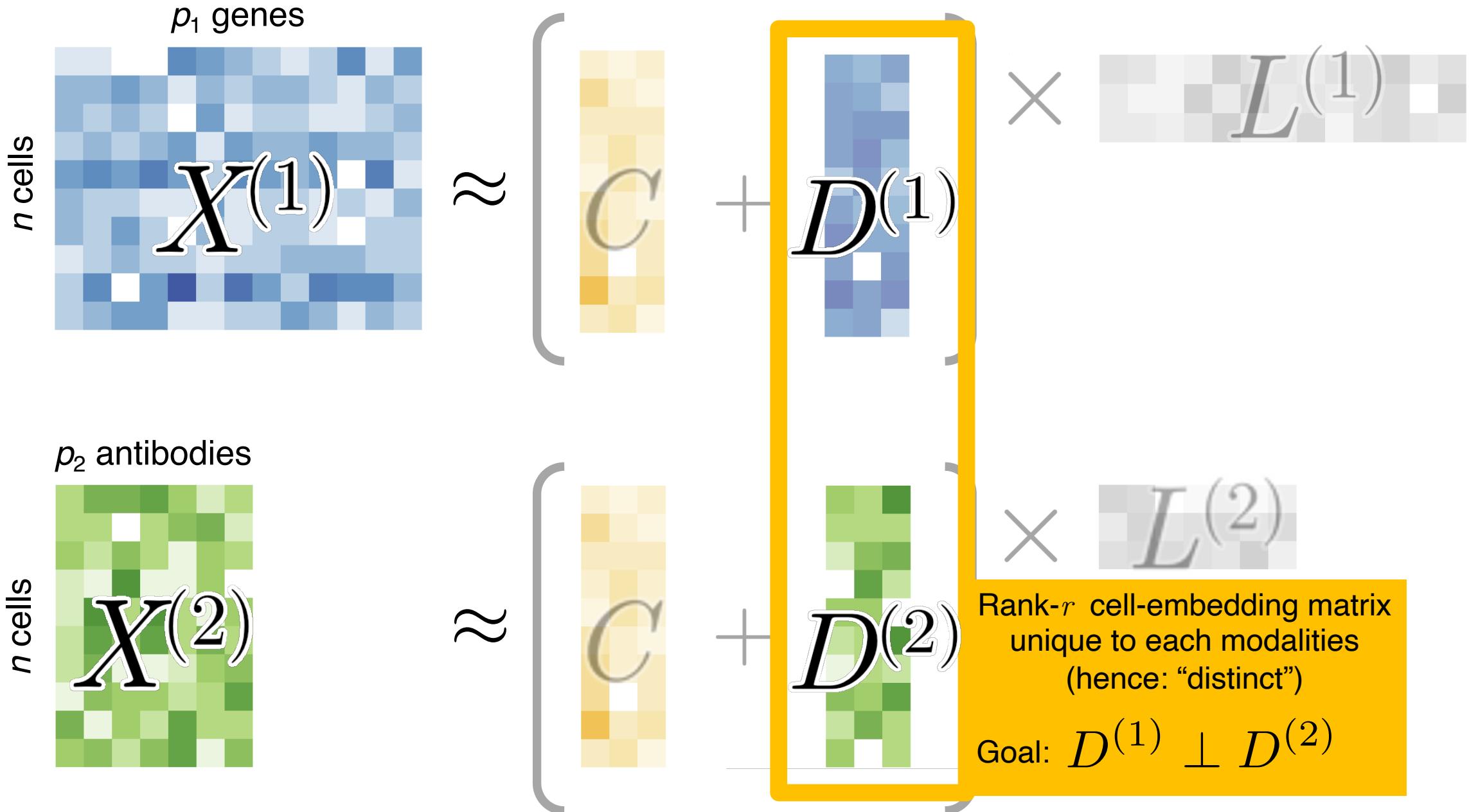
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Transformation of right-singular vectors

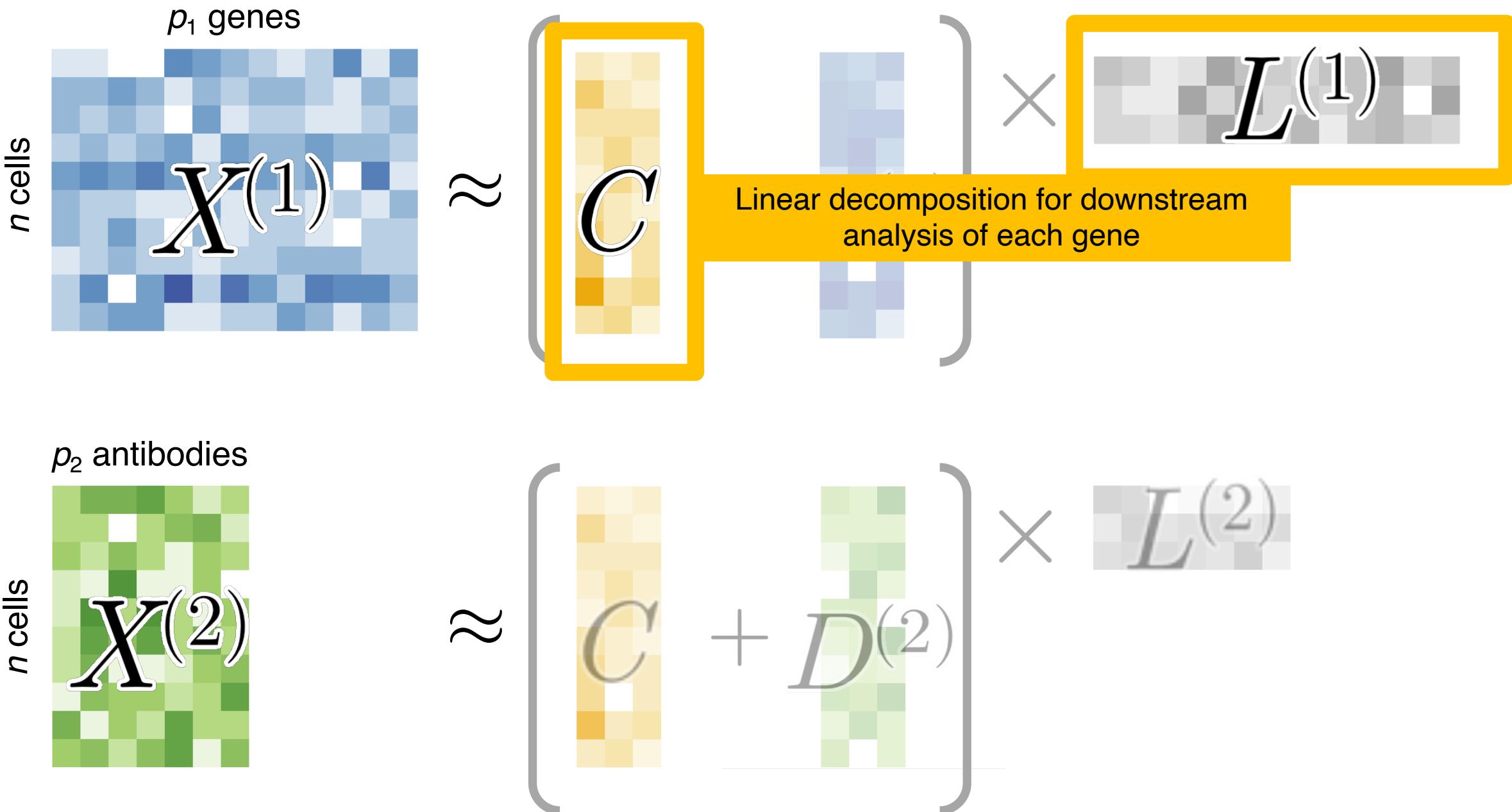
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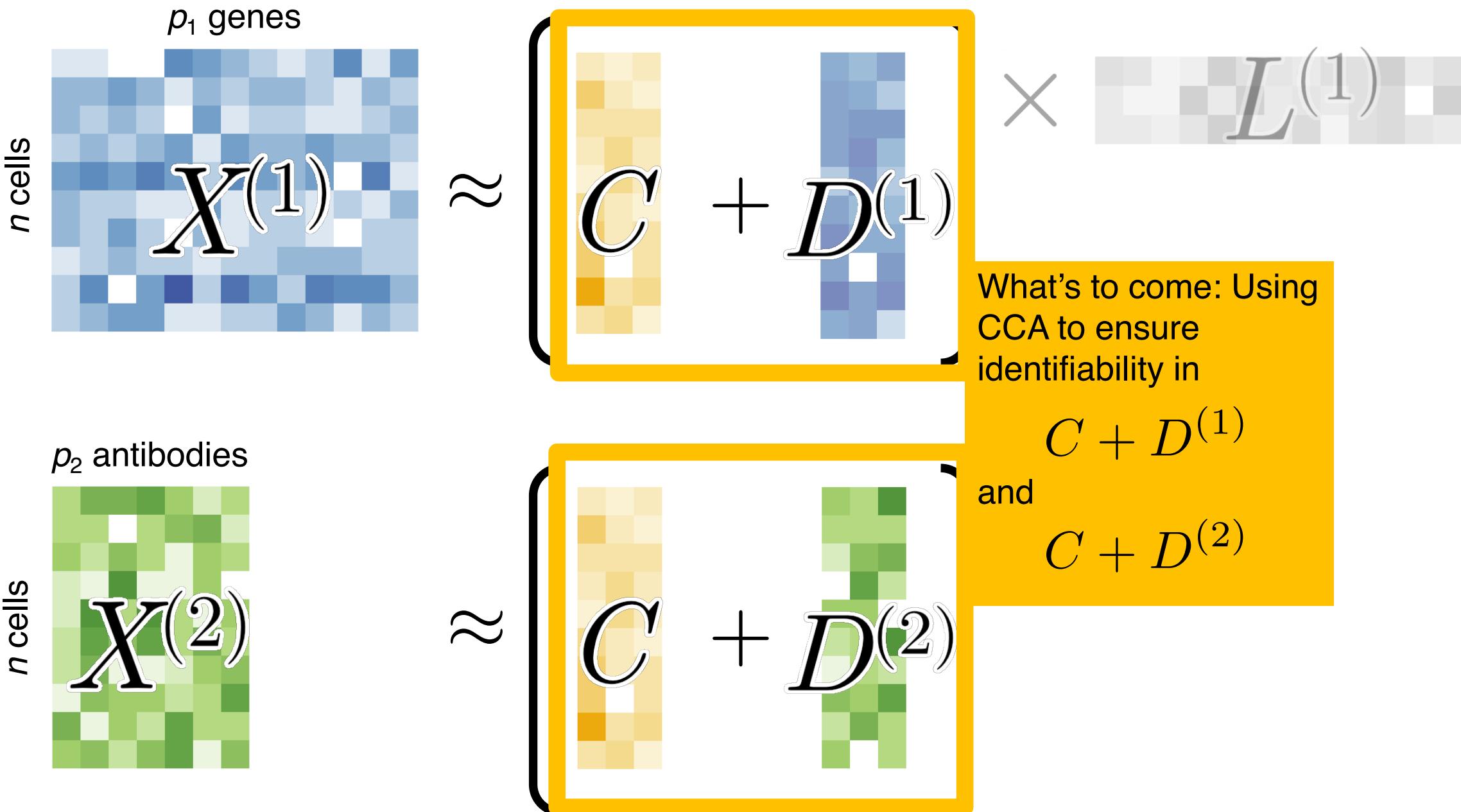
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## Brief aside: Review of Canonical Correlation Analysis

### CCA for the first latent dimension:

Given two modalities, find linear combination of variables that are highly correlated:

$$\{\hat{a}, \hat{b}\} = \arg \max_{\substack{a \in \mathbb{R}^{p_1} \\ b \in \mathbb{R}^{p_2}}} \text{Corr}(X^{(1)}a, X^{(2)}b)$$

### General multi-dimensional CCA:

$$\{\hat{A}, \hat{B}\} = \arg \max_{\substack{A \in \mathbb{R}^{p_1 \times r} : A^\top \Sigma^{(1)} A = I_r \\ B \in \mathbb{R}^{p_2 \times r} : B^\top \Sigma^{(2)} B = I_r}} \text{Tr}\left(A^\top (X^{(1)})^\top X^{(2)} B\right)$$

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Orthogonality of  
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At optimality:  
Orthogonality of  
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across modalities

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Let:  $Z^{(1)} = X^{(1)} \hat{A}$   
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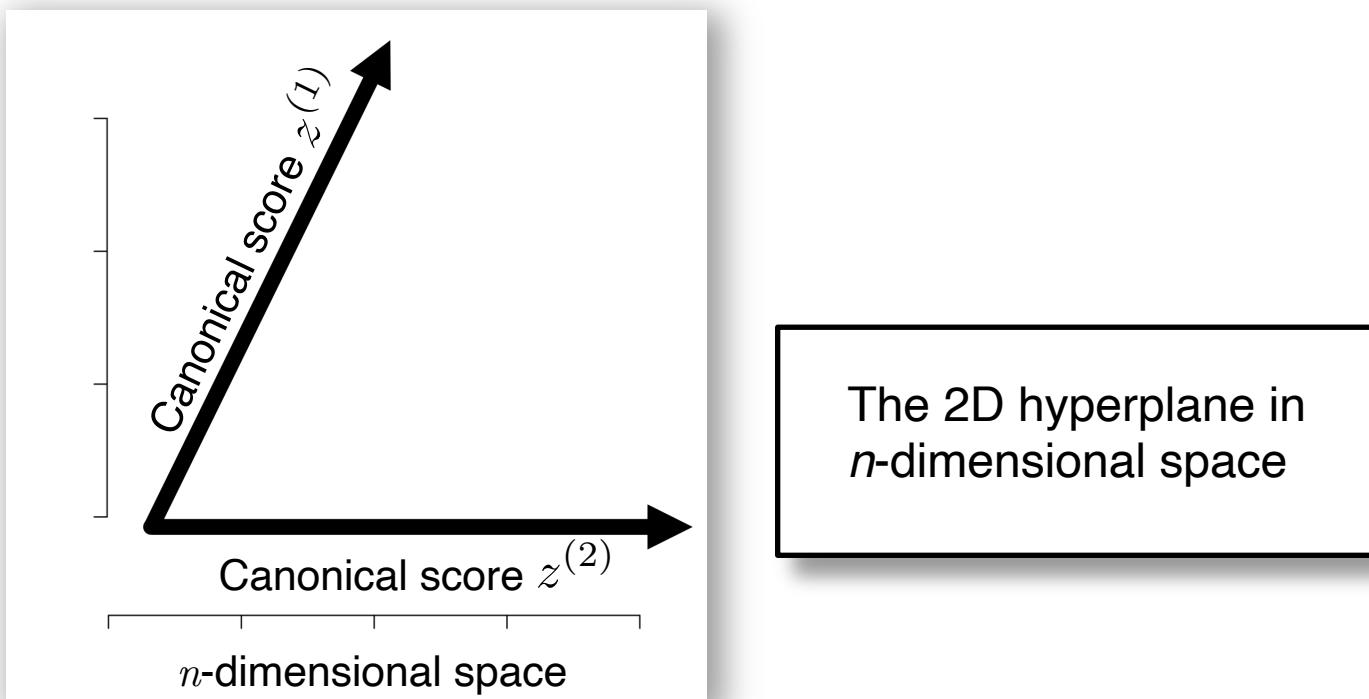
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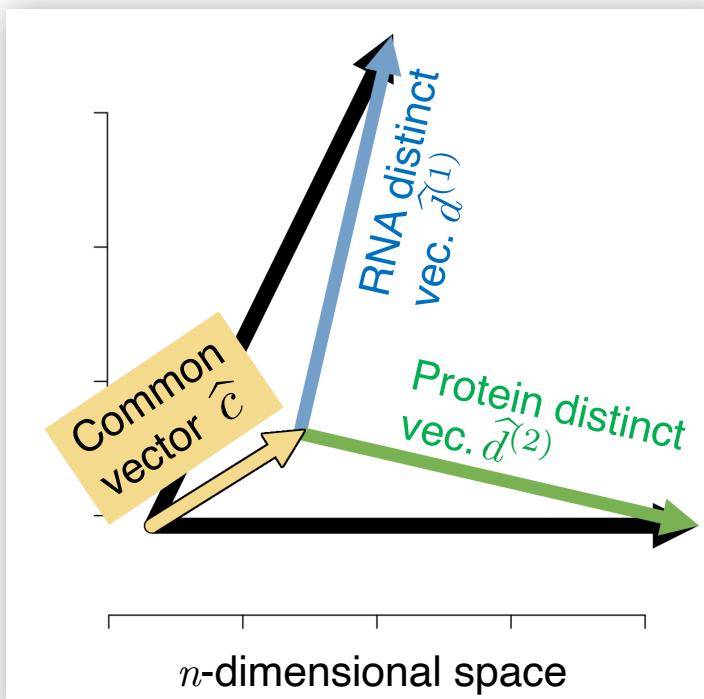
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Decomposition:

$$z^{(1)} = \hat{c} + \hat{d}^{(1)}$$

$$z^{(2)} = \hat{c} + \hat{d}^{(2)}$$

(quantifying what  
“common” means)

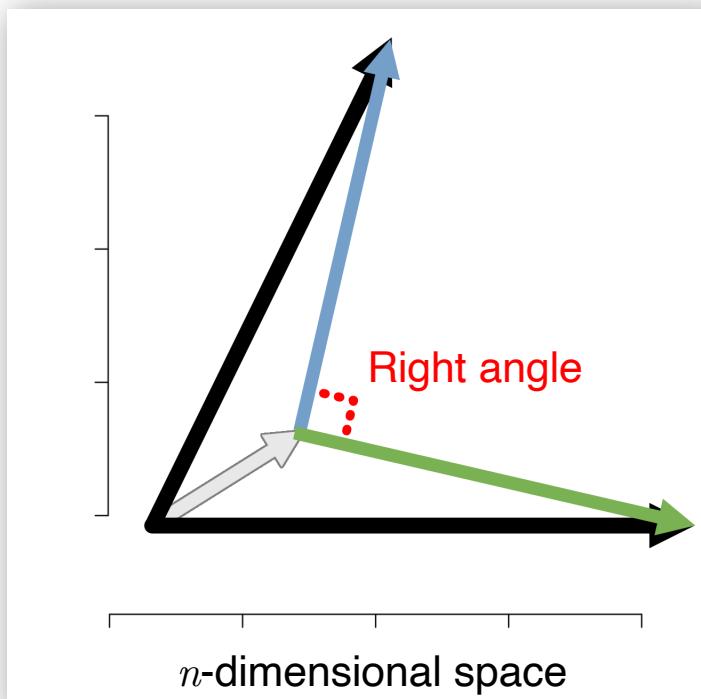
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Constrained to:

$$\hat{d}^{(1)} \perp \hat{d}^{(2)}$$

(quantifying what  
“distinct” means)

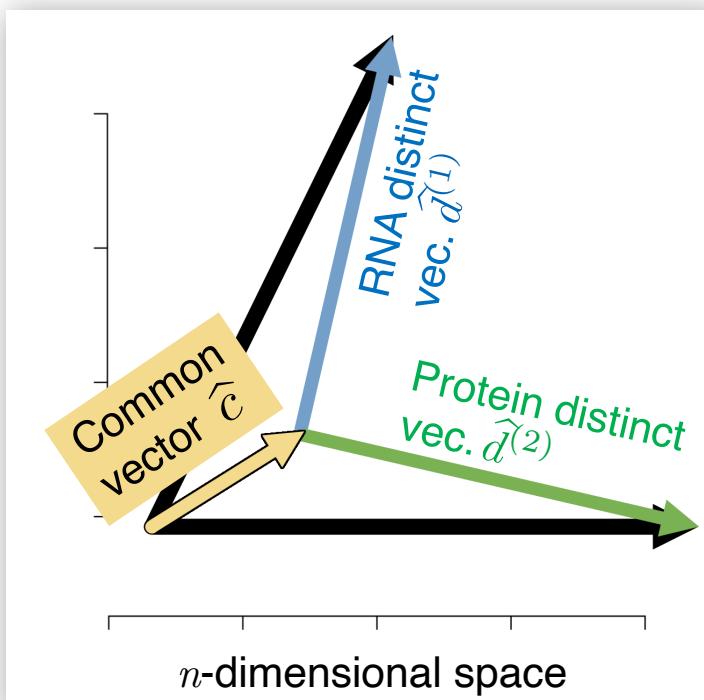
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$$A \in \mathbb{R}^{p_1 \times r} : A^\top \Sigma^{(1)} A = I_r$$

$$B \in \mathbb{R}^{p_2 \times r} : B^\top \Sigma^{(2)} B = I_r$$

Let:  $Z^{(1)} = X^{(1)} \hat{A}$   
 $Z^{(2)} = X^{(2)} \hat{B}$



$$X^{(1)} \approx \begin{bmatrix} C \\ C + D^{(1)} \end{bmatrix} \times L^{(1)}$$

$$X^{(2)} \approx \begin{bmatrix} C \\ C + D^{(2)} \end{bmatrix} \times L^{(2)}$$

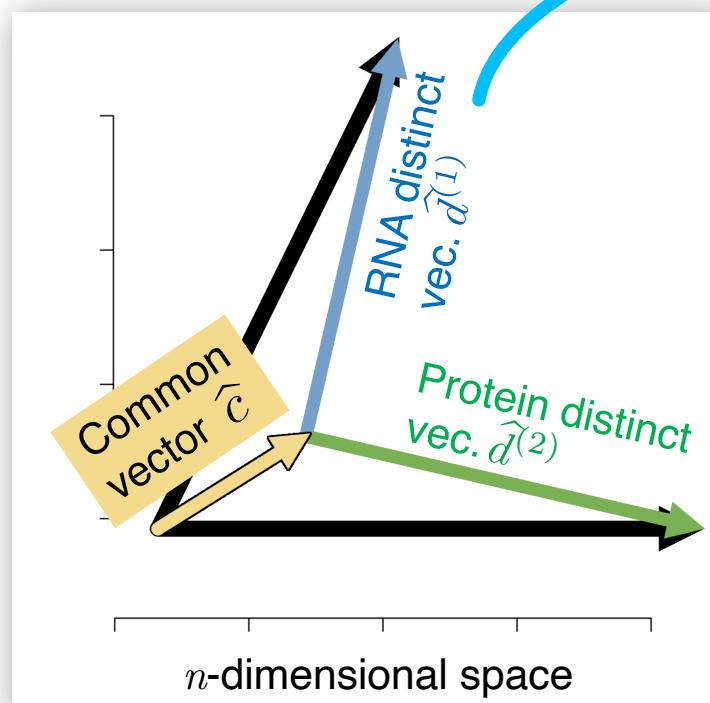
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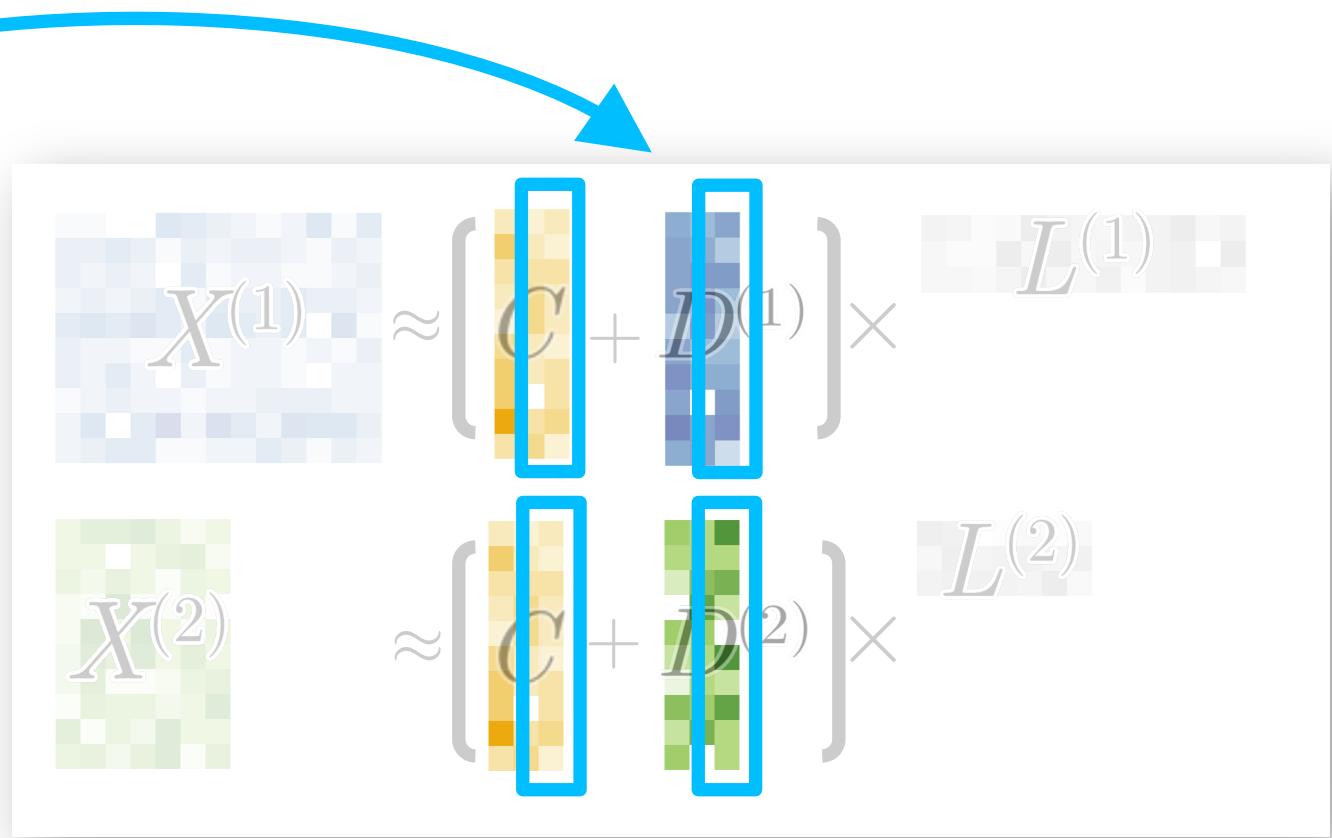
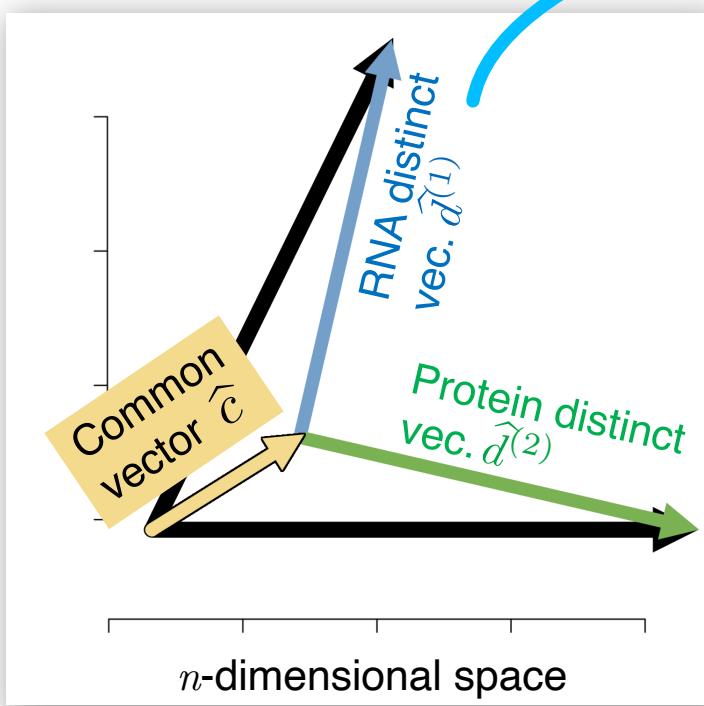
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Identifiability ensured thanks to CCA's properties

$$\text{Let: } Z^{(1)} = X^{(1)} \hat{A}$$

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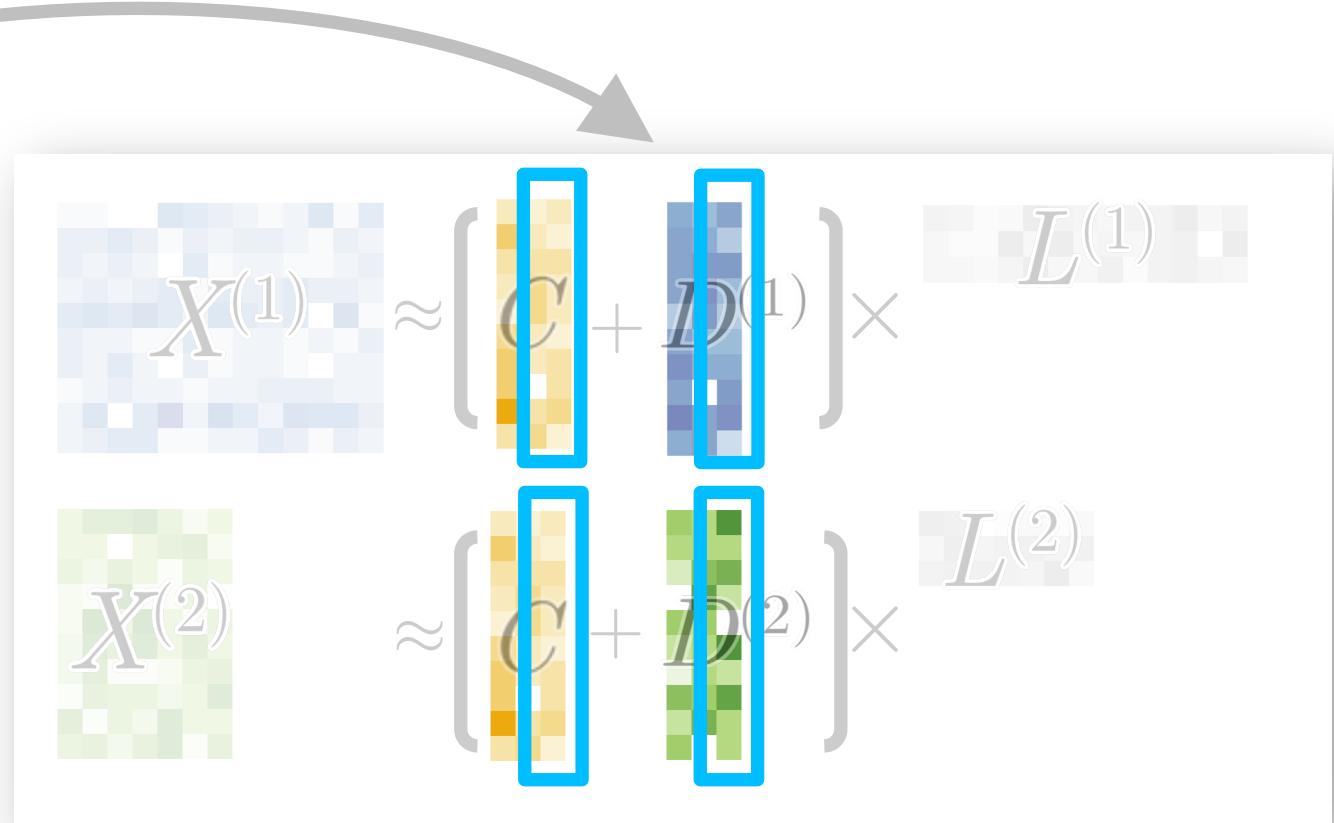
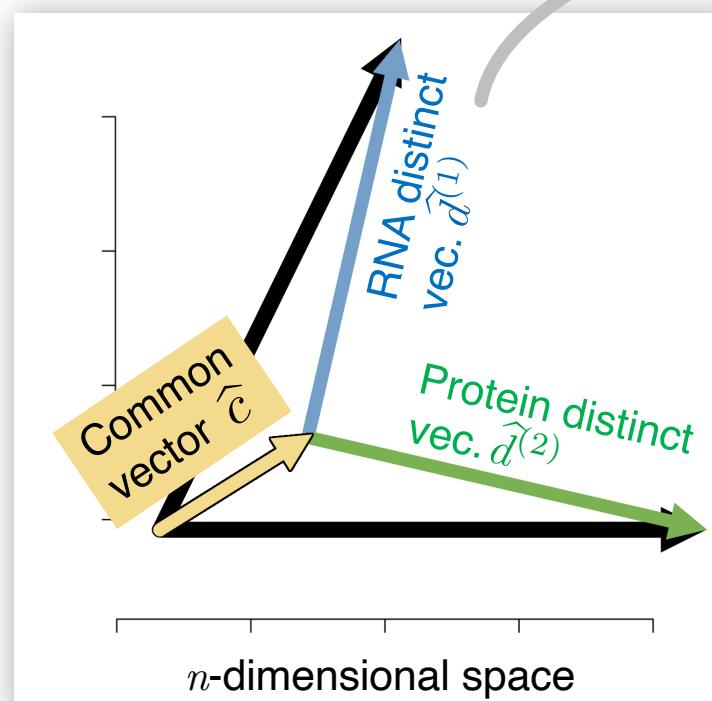
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$$Z^{(2)} = X^{(2)} \hat{B}$$



✓ Decomp. for each latent dim.

✓ Ortho. distinct vectors

✗ Common vector shared geometry

## Decomposition based on CCA

$$\{\hat{A}, \hat{B}\} = \arg \max \text{Tr} \left( A^\top (X^{(1)})^\top X^{(2)} B \right)$$

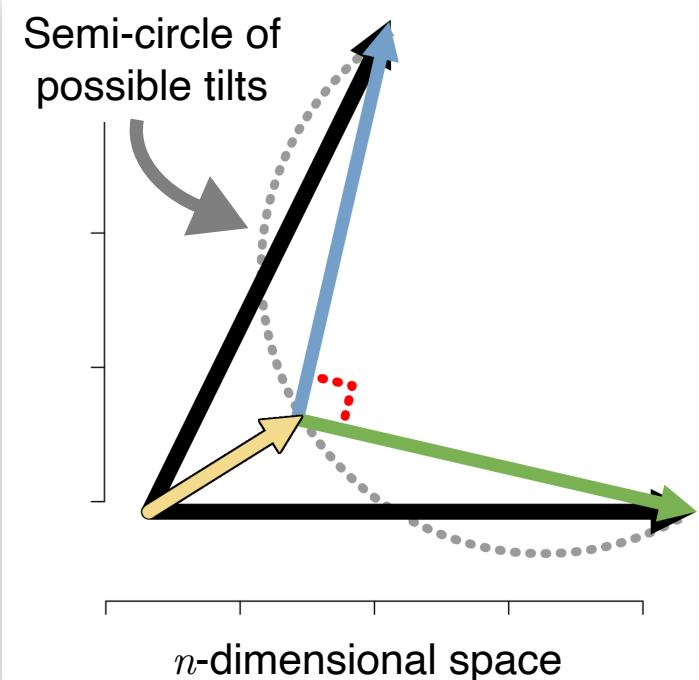
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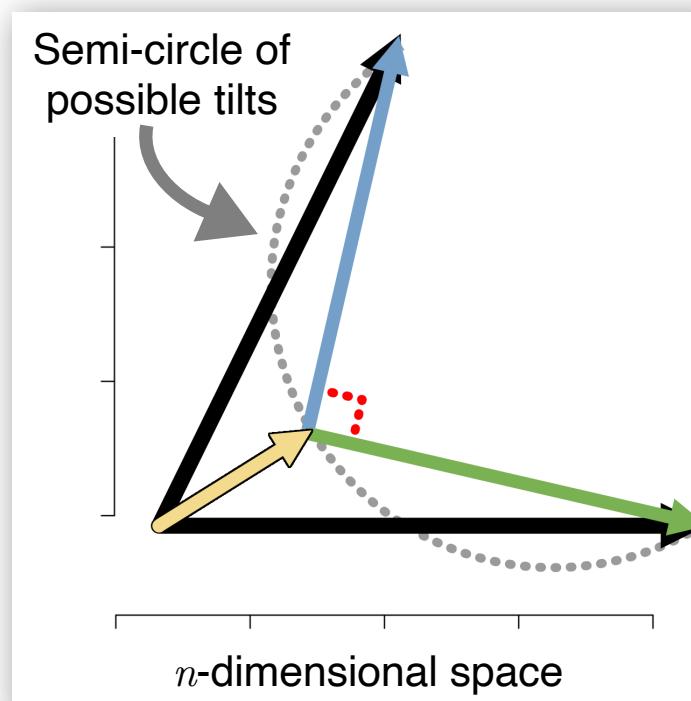


### Our insight:

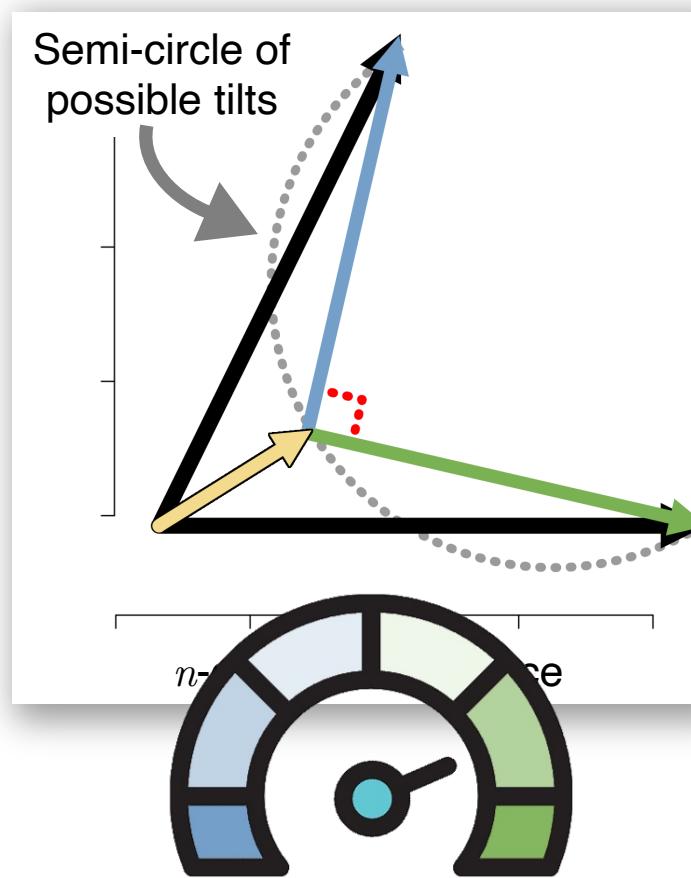
The common vector can fall anywhere along this **semi-circle** and still retain orthogonal distinct vectors.

(One extra degree of freedom per dimension)

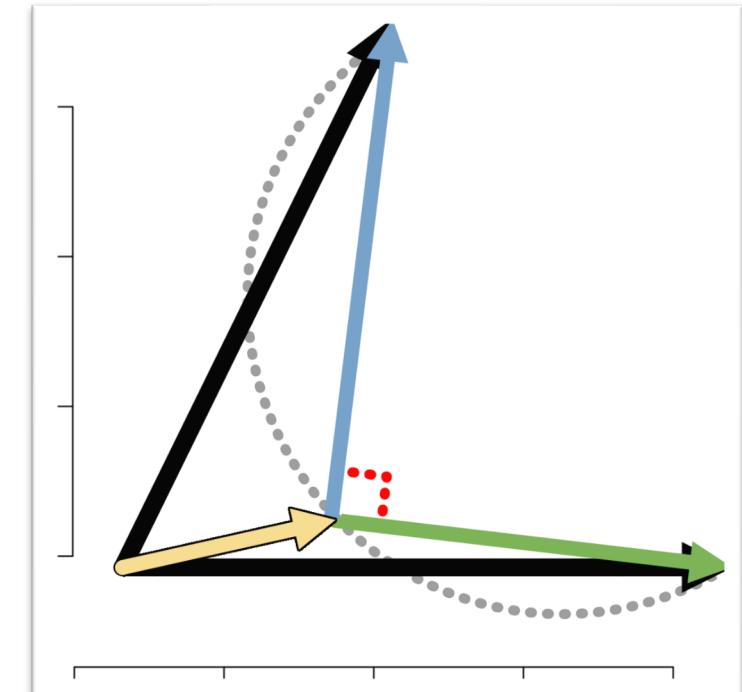
**Our novelty (combining ideas in CCA with geometry):** The tilt of the common vector (if chosen appropriately) allows the common embedding to reflect the shared geometry between both modalities.



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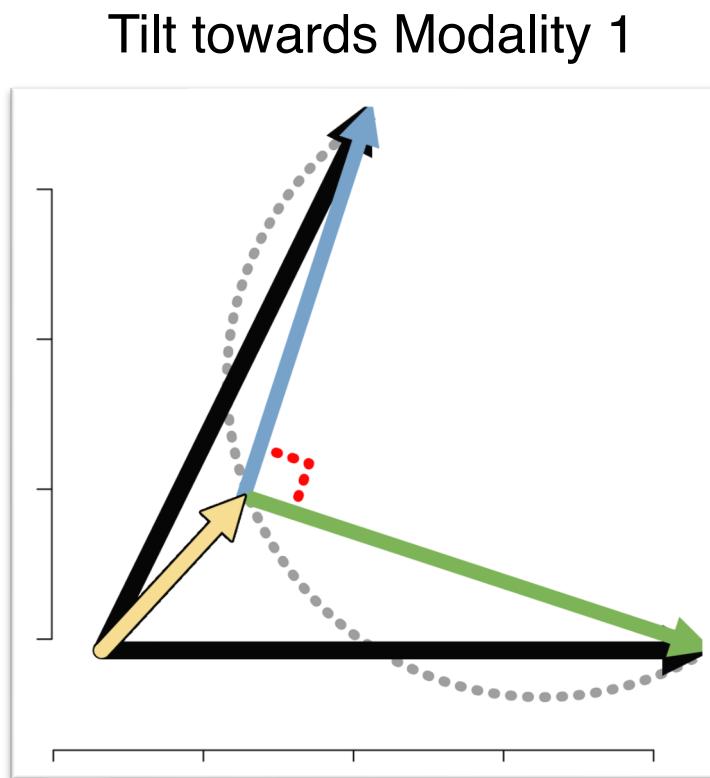


Tilt towards Modality 2

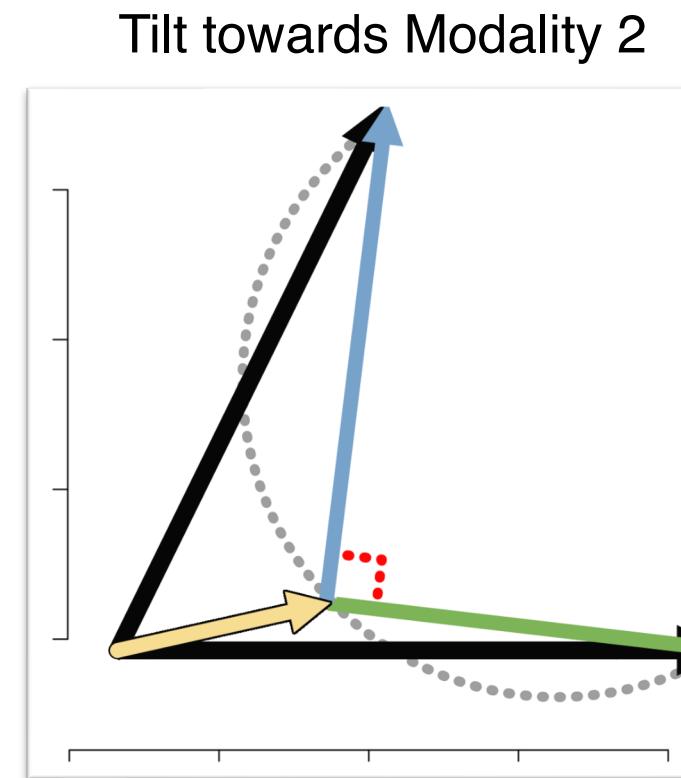
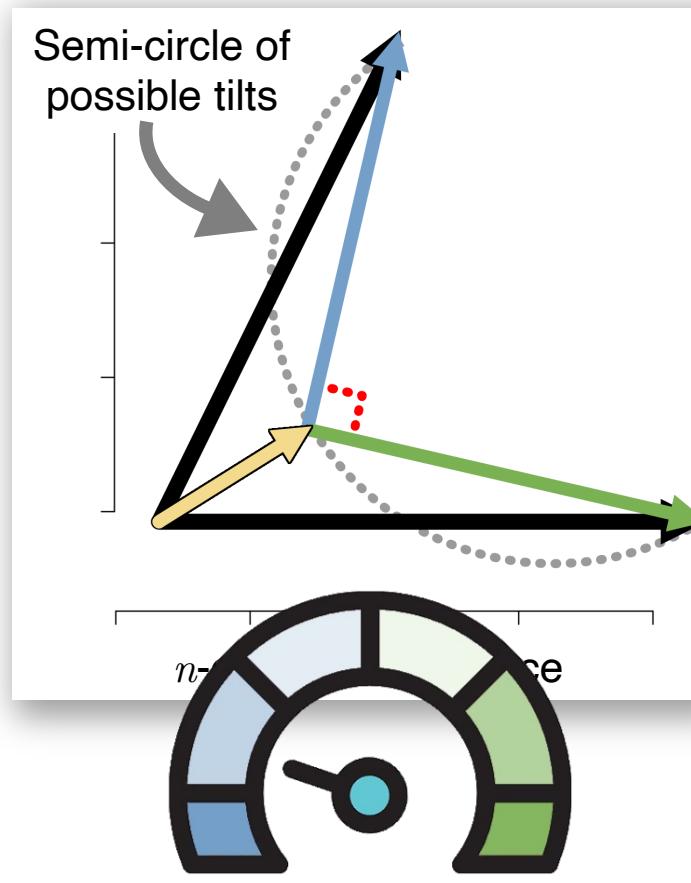


Common embedding has geometry similar to Modality 2

**Our novelty (combining ideas in CCA with geometry):** The tilt of the common vector (if chosen appropriately) allows the common embedding to reflect the shared geometry between both modalities.



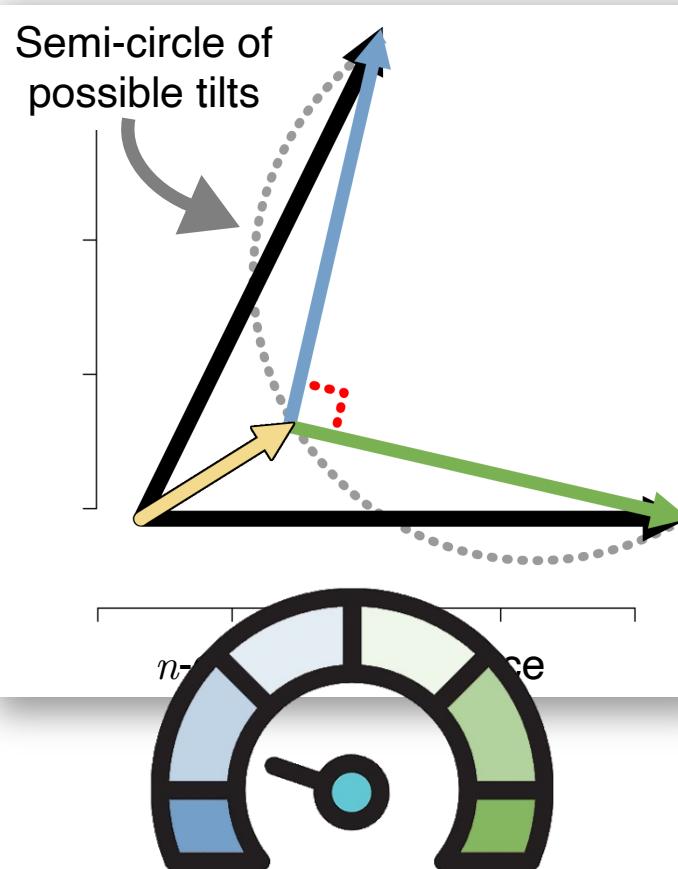
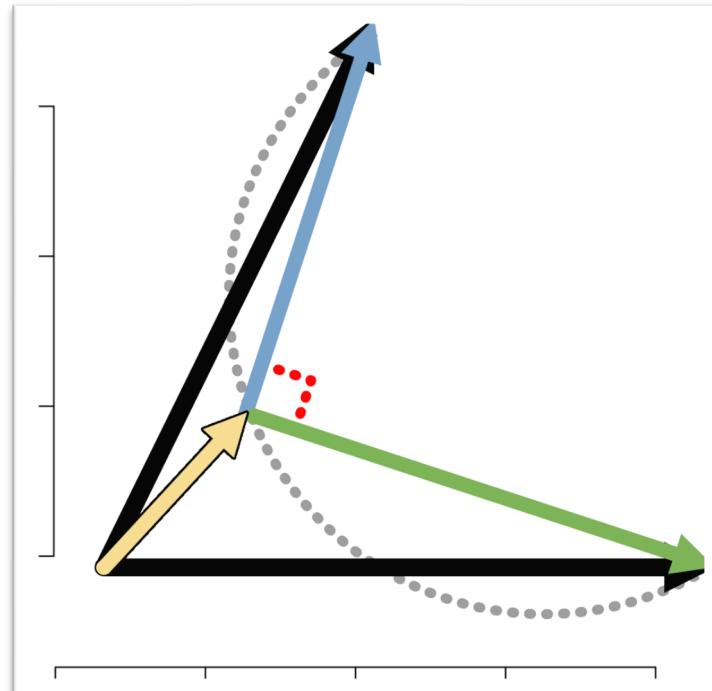
Common embedding has geometry  
similar to Modality 1



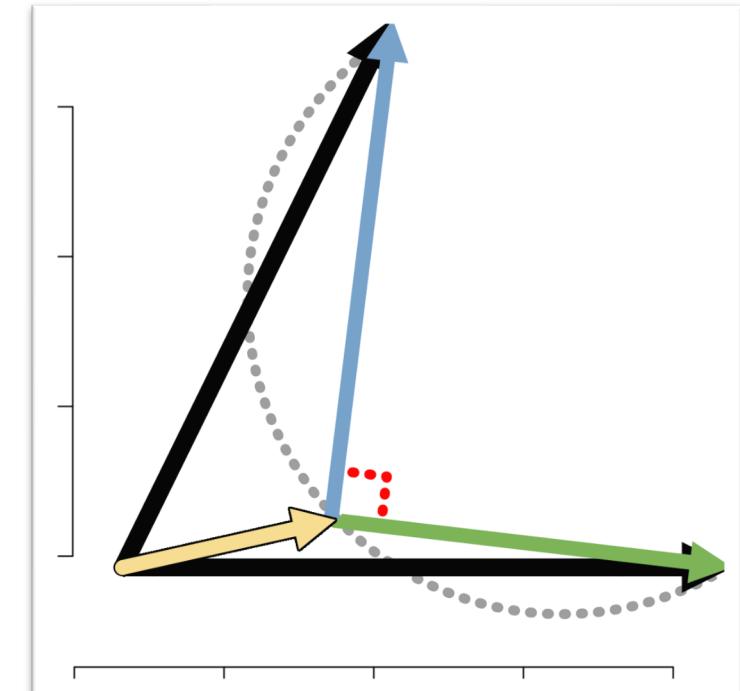
Common embedding has geometry  
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Tilt towards Modality 1



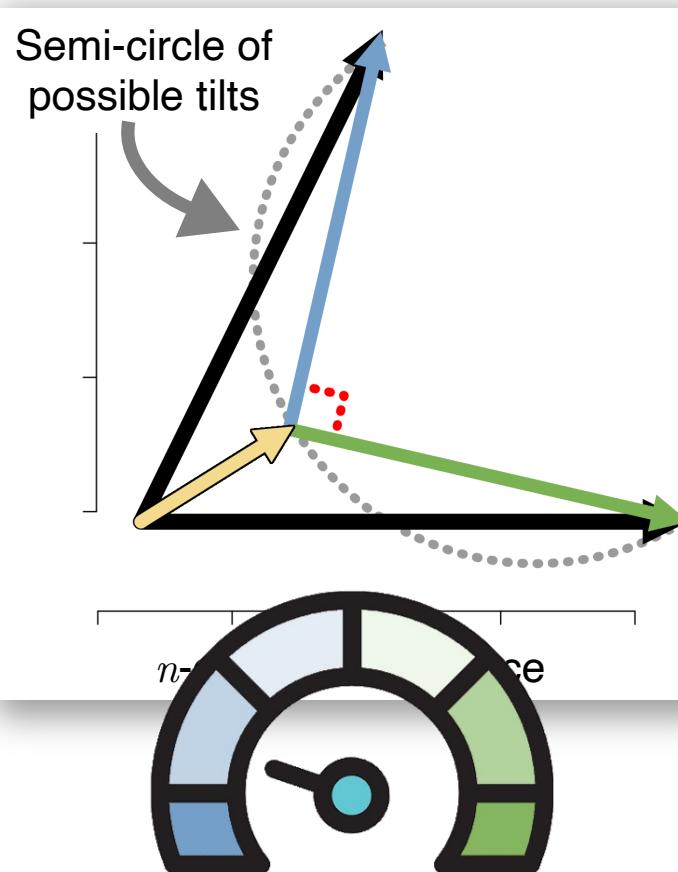
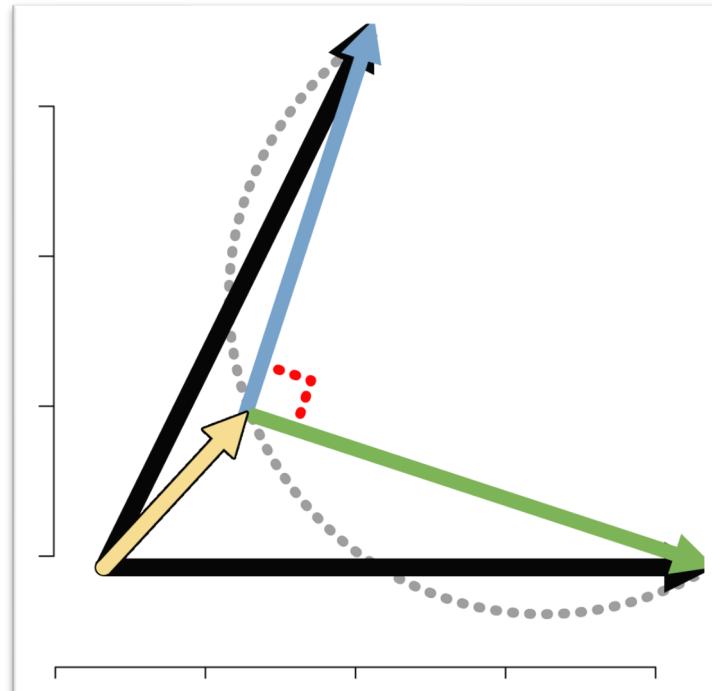
Tilt towards Modality 2



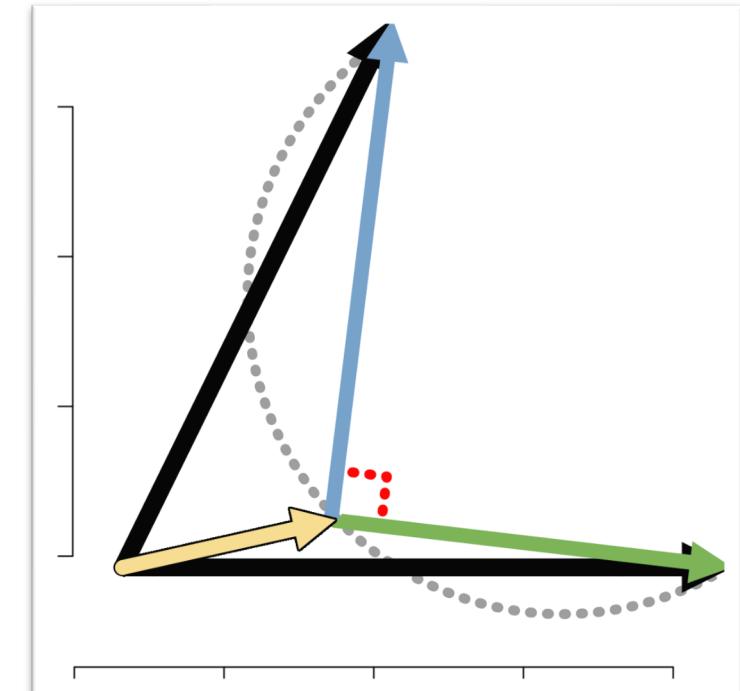
**Main Goal:** To estimate the tilt  $\tau_j \in [1, 2]$  in each latent dimension  $j$  to capture the desired shared geometry

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Tilt towards Modality 1



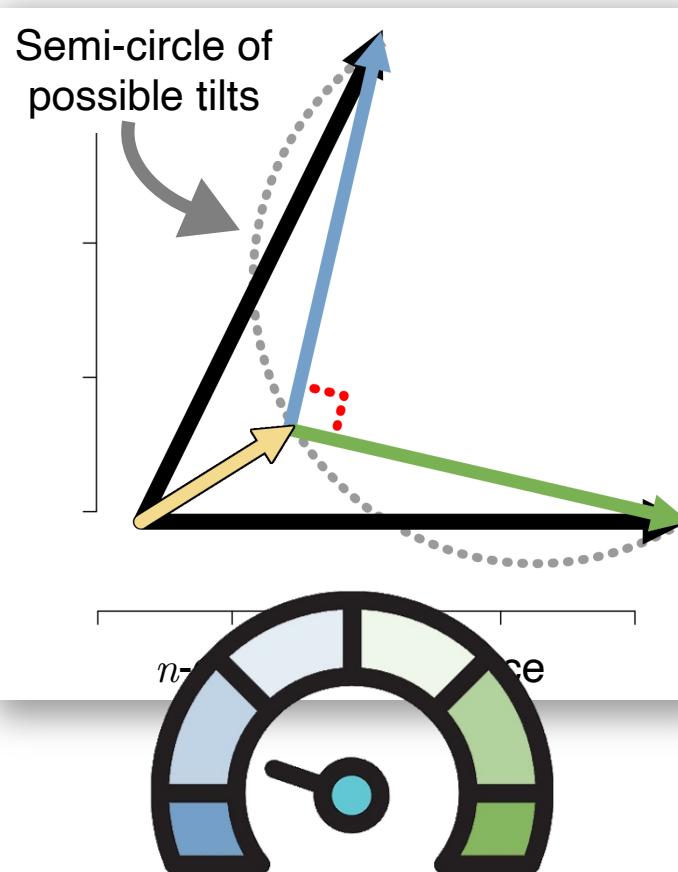
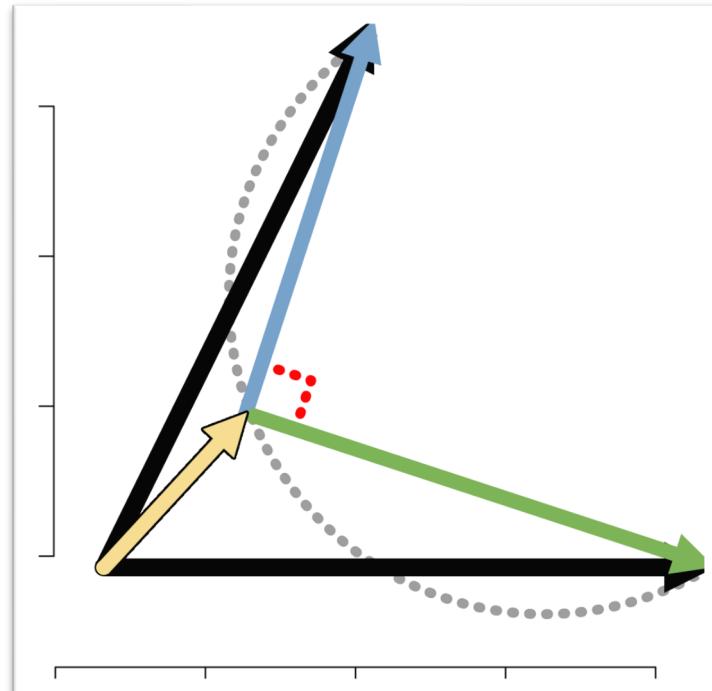
Tilt towards Modality 2



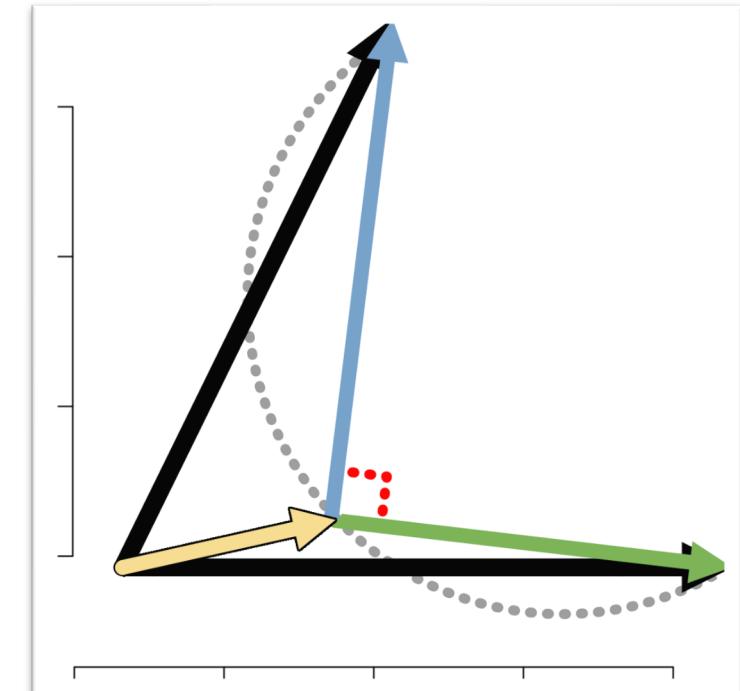
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Tilt towards Modality 2

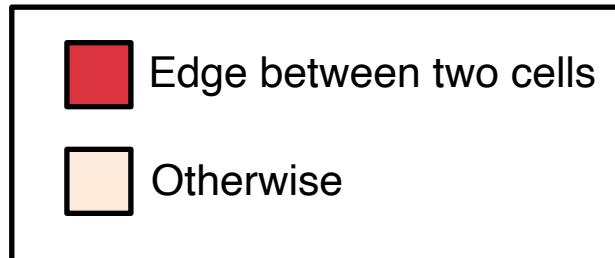


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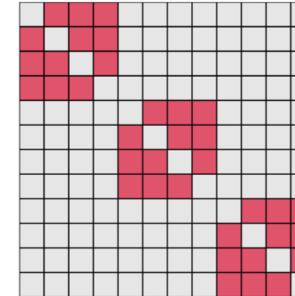
## (Unsupervised) Estimation: Part 1 – Defining the shared geometry

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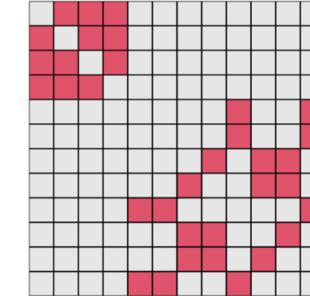
1



Cell-by-cell  
nearest-neighbor  
graph  
(RNA modality)



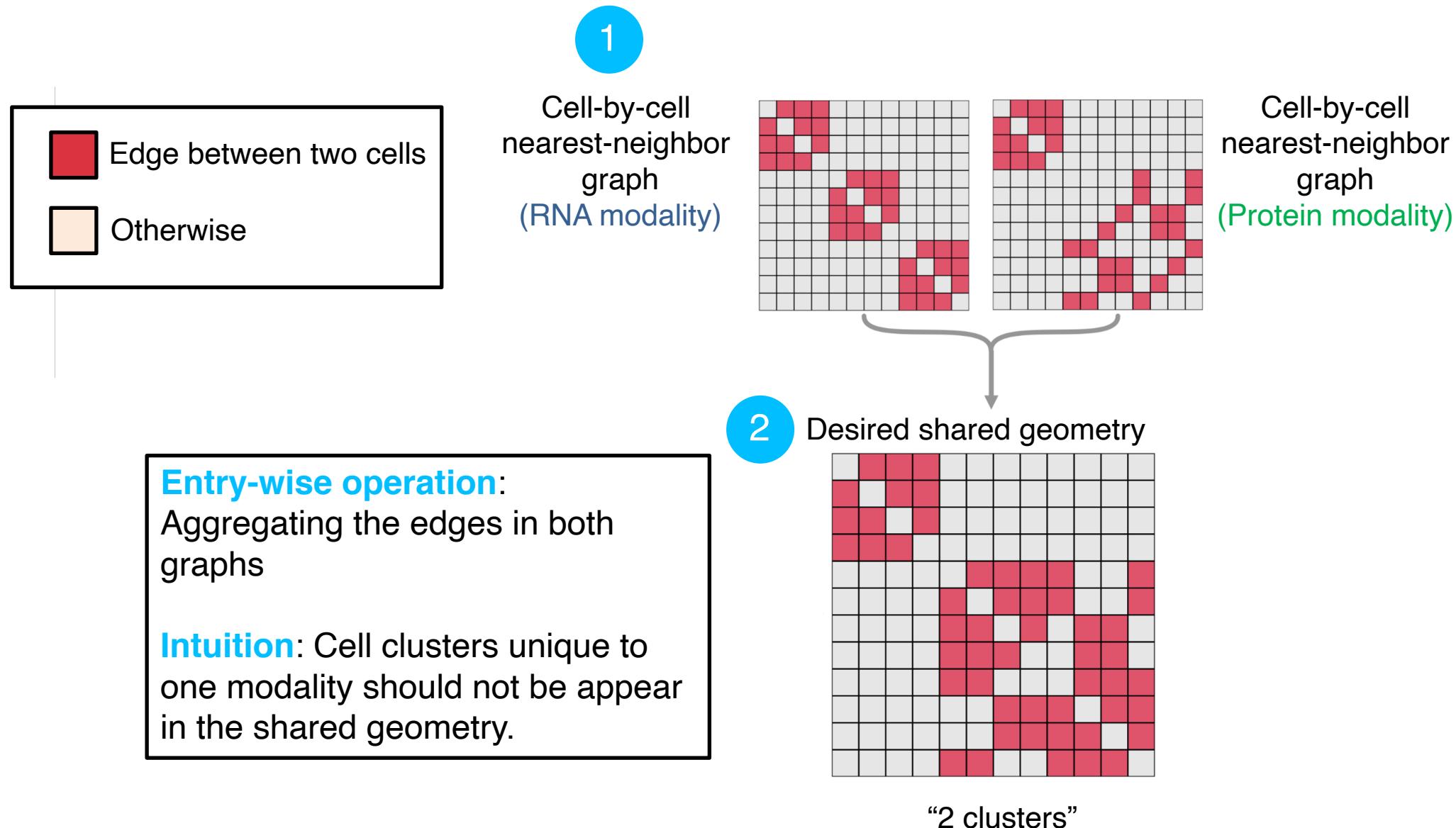
“3 clusters”



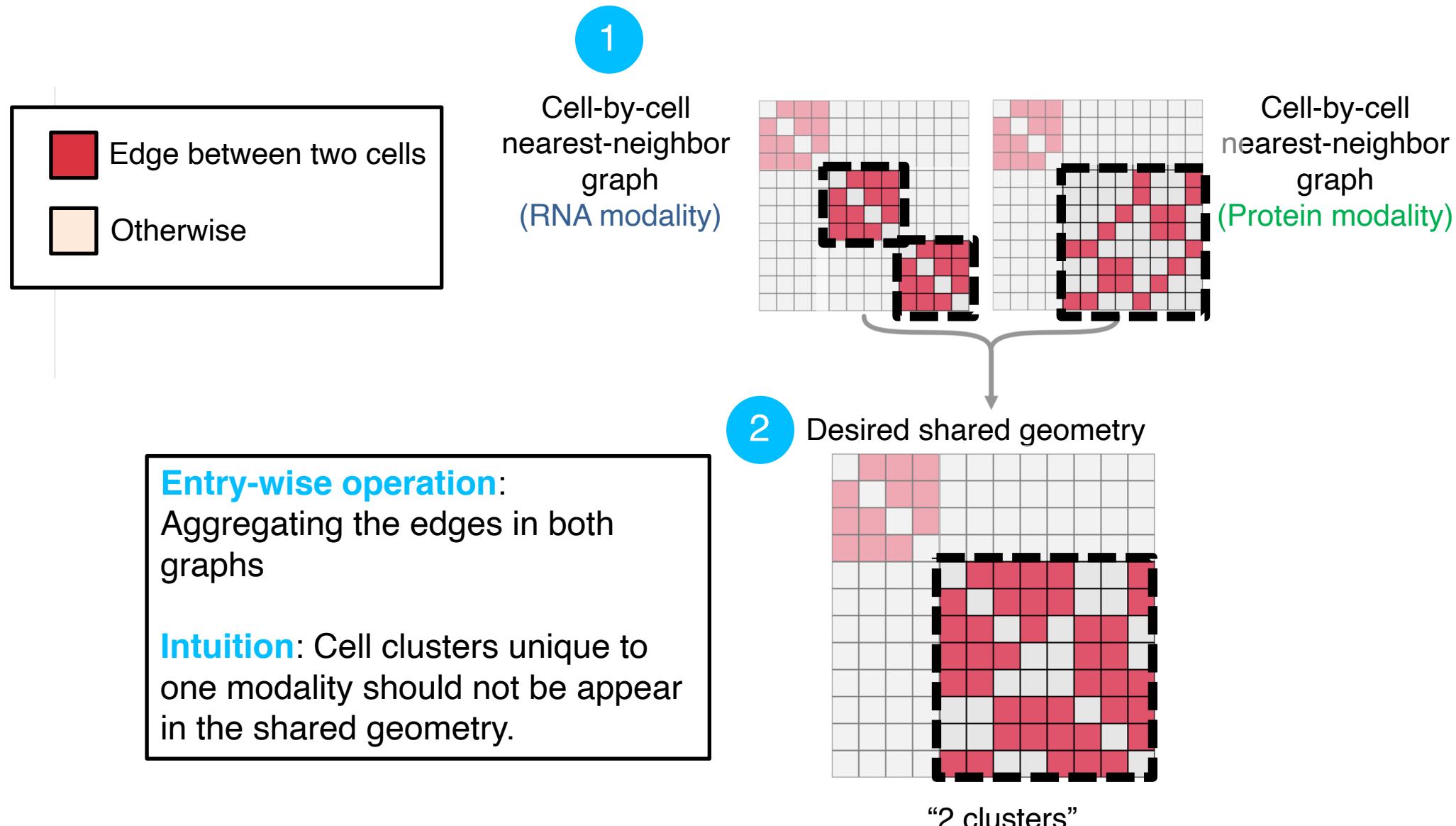
“2 clusters”

Cell-by-cell  
nearest-neighbor  
graph  
(Protein modality)

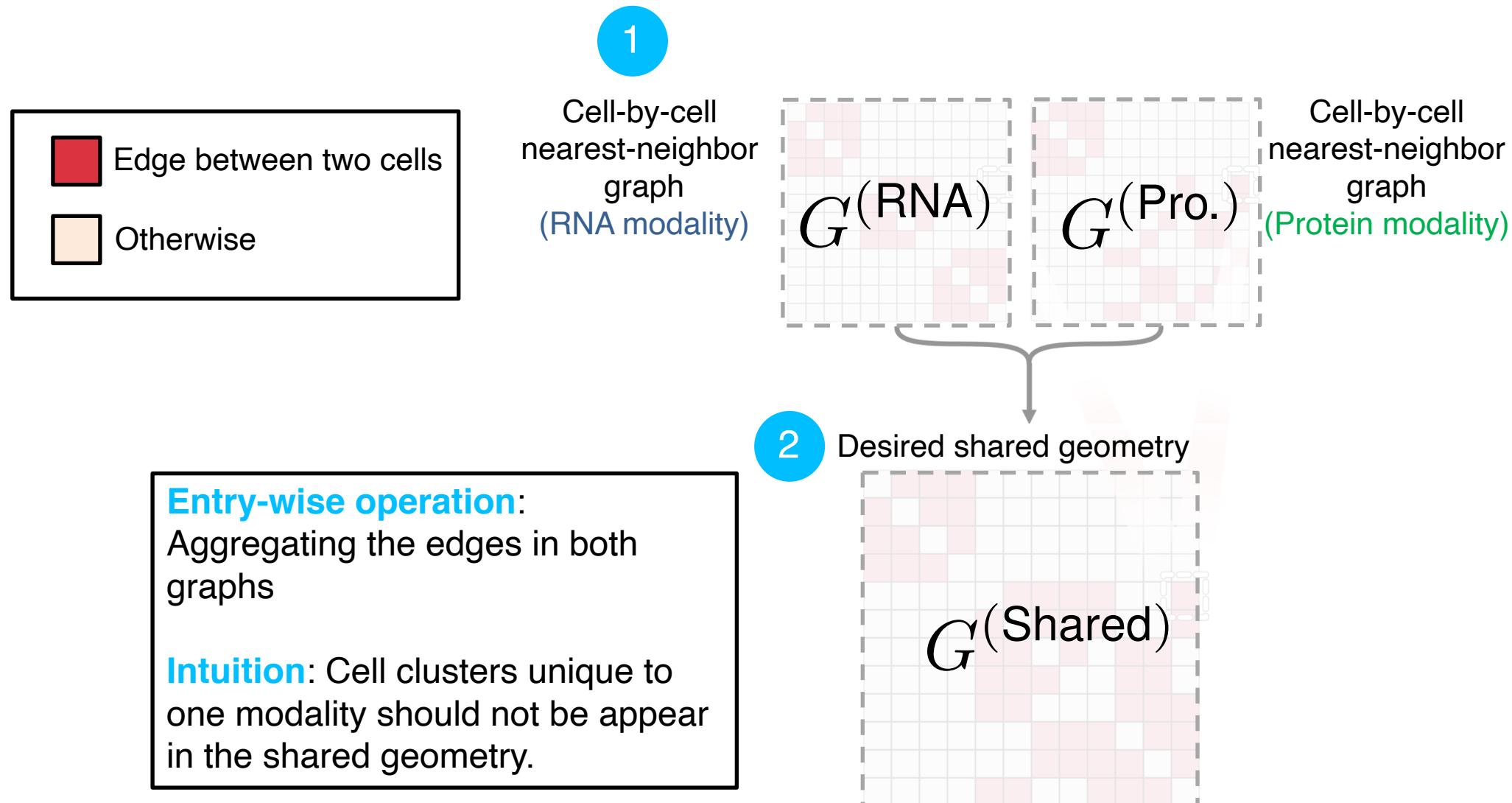
# (Unsupervised) Estimation: Part 1 – Defining the shared geometry



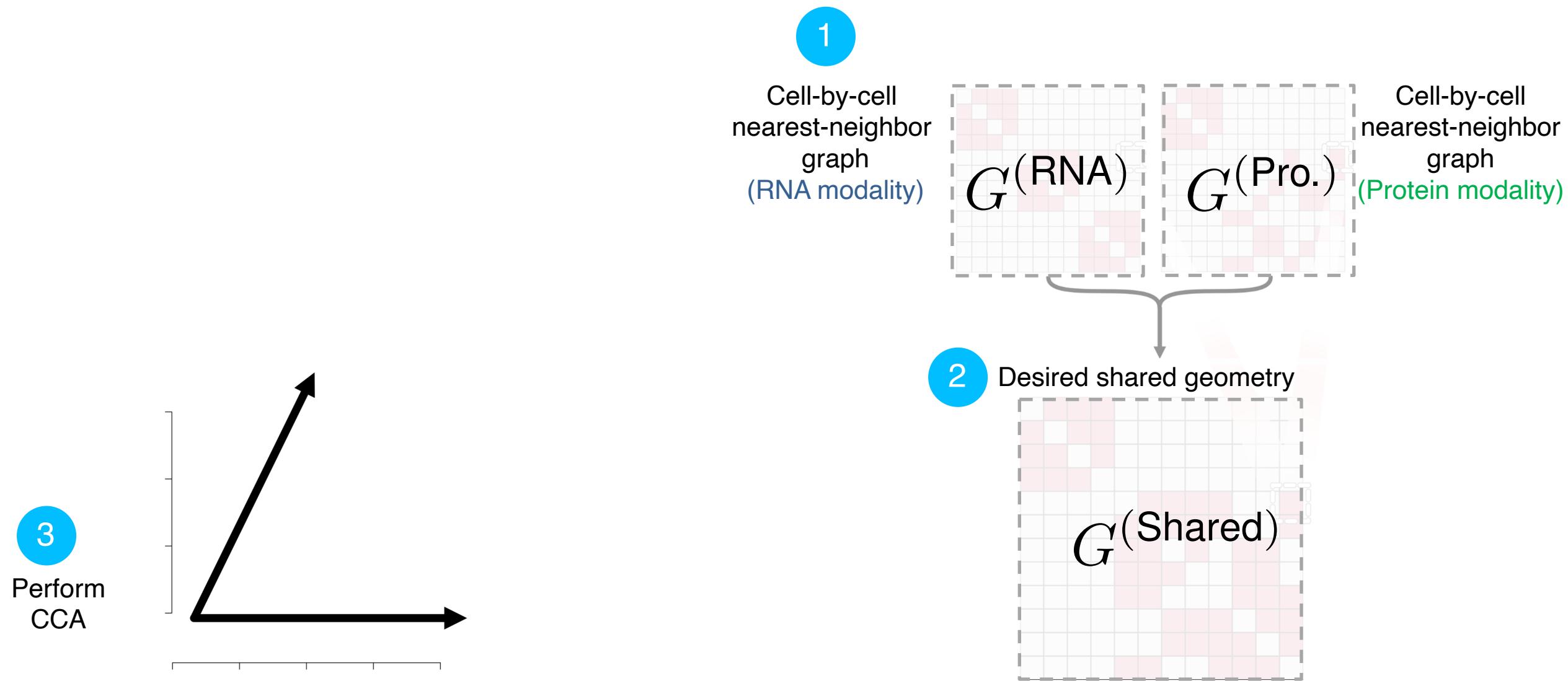
# (Unsupervised) Estimation: Part 1 – Defining the shared geometry



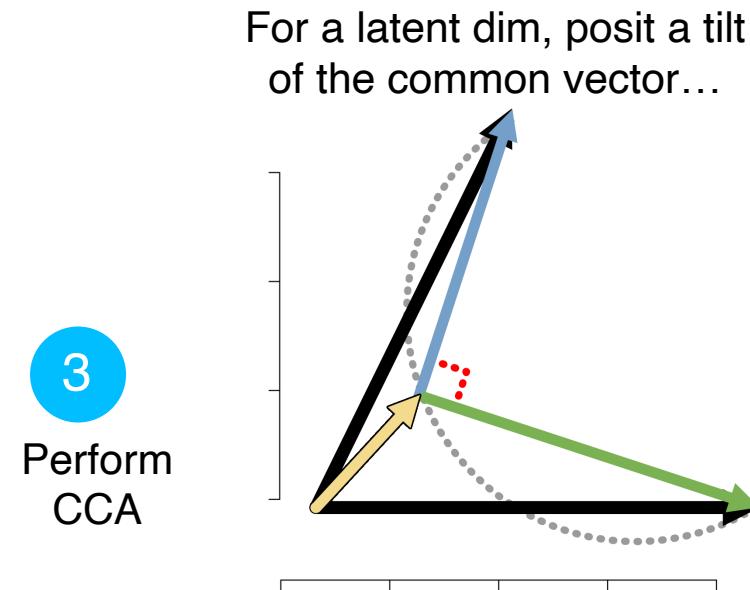
# (Unsupervised) Estimation: Part 1 – Defining the shared geometry



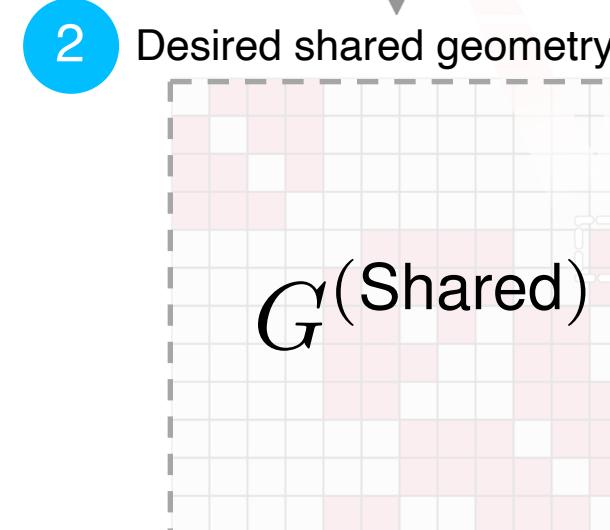
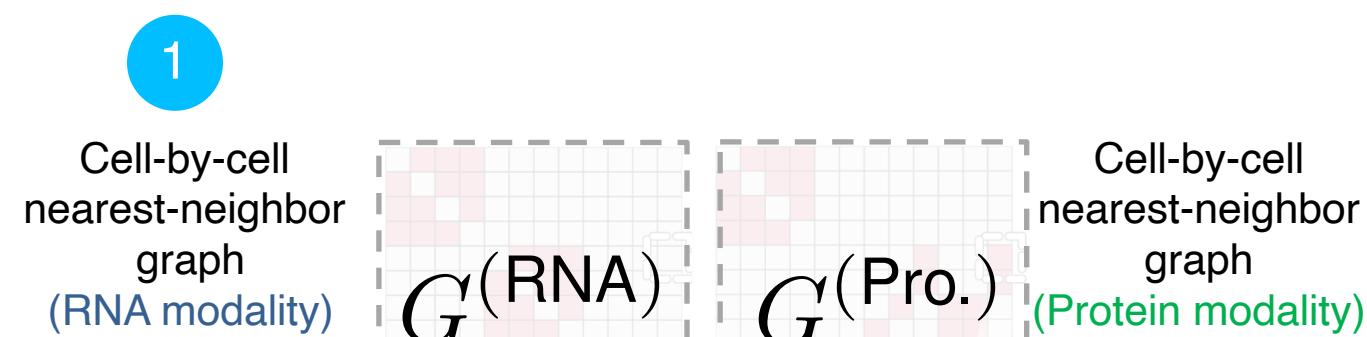
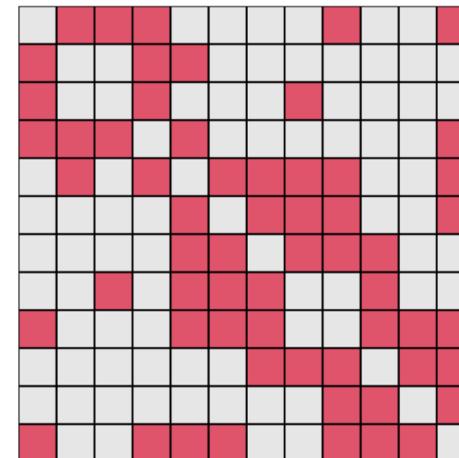
## (Unsupervised) Estimation: Part 2 – Estimating the corresponding matrix factorization



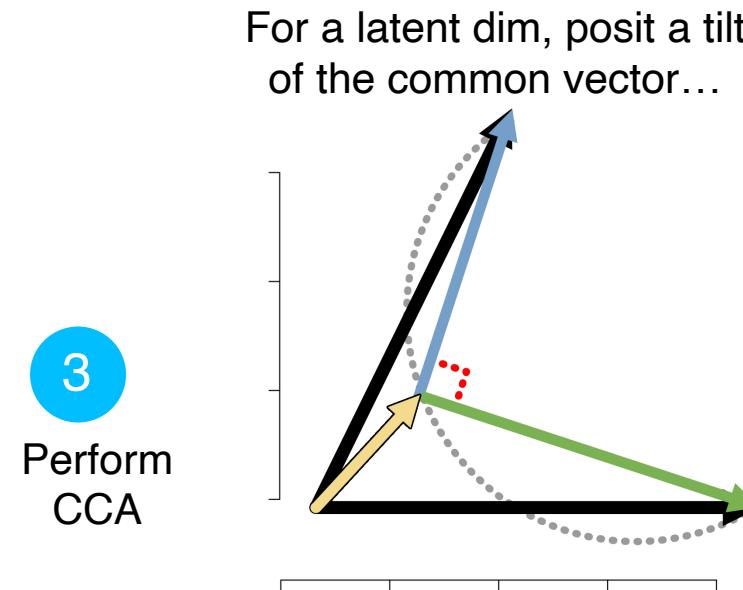
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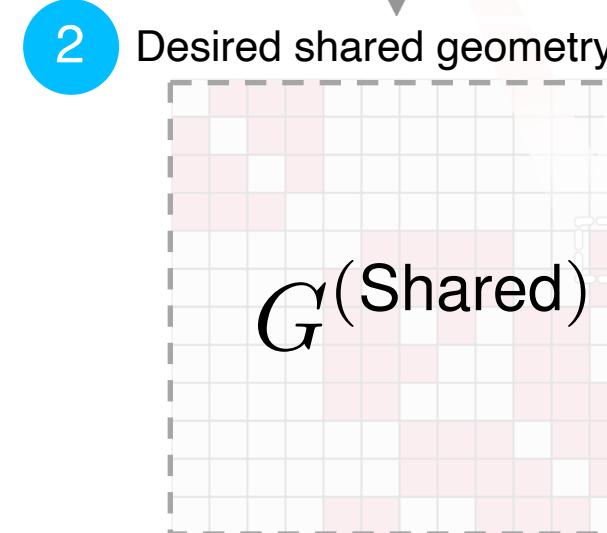
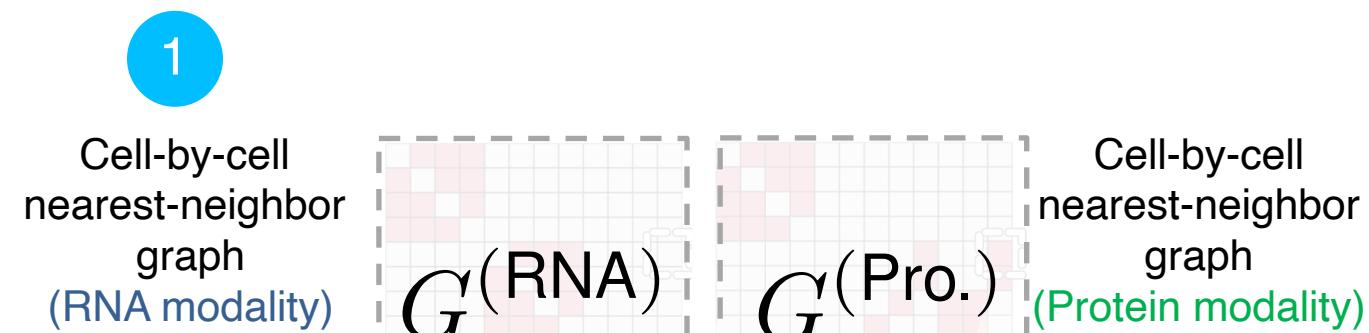
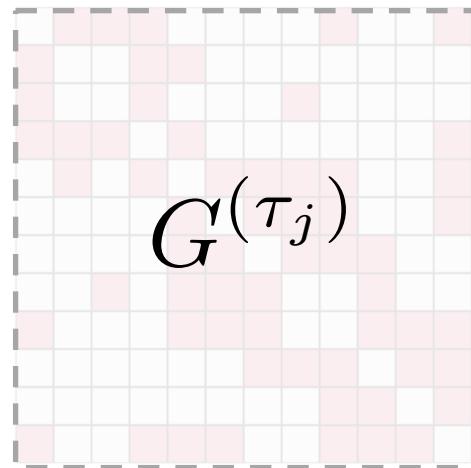
... and its corresponding cell-by-cell nearest-neighbor graph for the common embedding



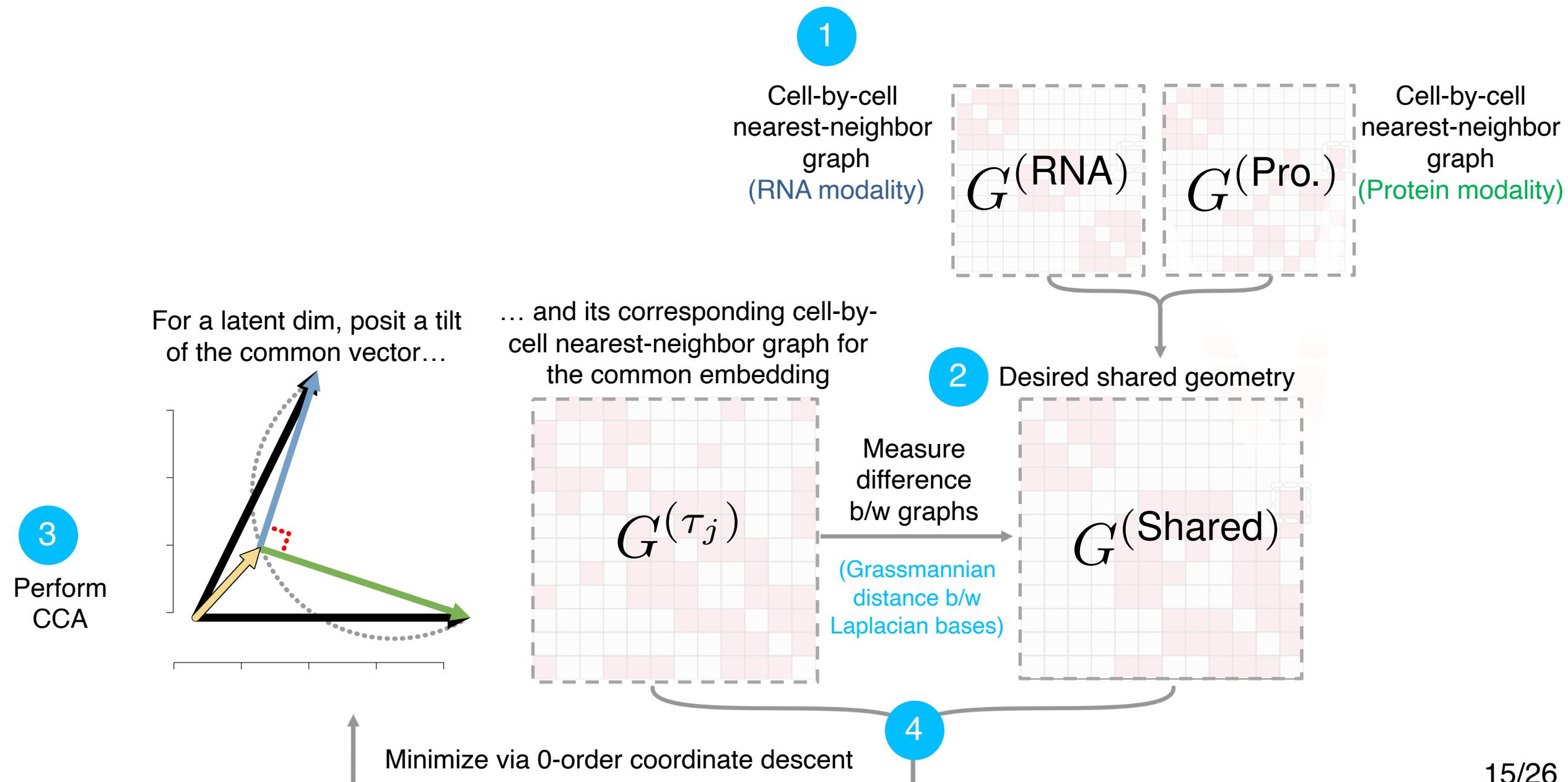
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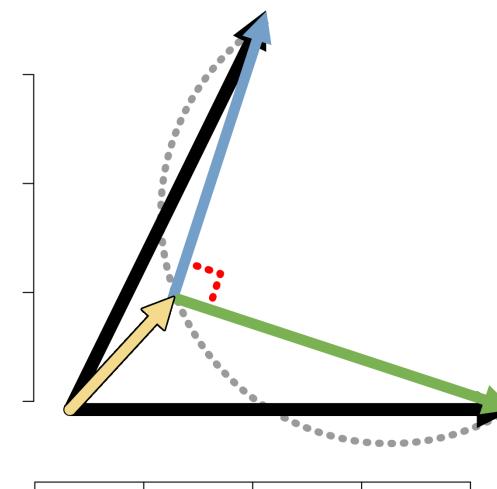
# (Unsupervised) Estimation: Part 2 – Estimating the corresponding matrix factorization



# (Unsupervised) Estimation: Part 2 – Estimating the corresponding matrix factorization

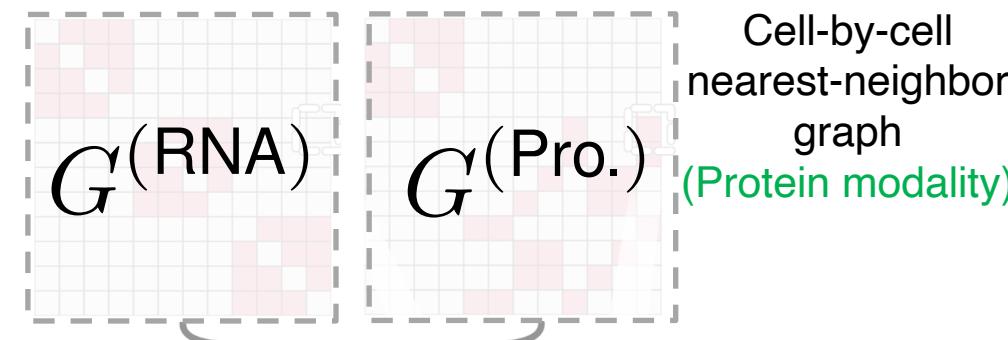
$$X^{(1)} \approx [C + D^{(1)}] \times L^{(1)}$$
$$X^{(2)} \approx [C + D^{(2)}] \times L^{(2)}$$
$$C + D^{(1)} = Z^{(1)}, C + D^{(2)} = Z^{(2)}$$

For a latent dim, posit a tilt  
of the common vector...

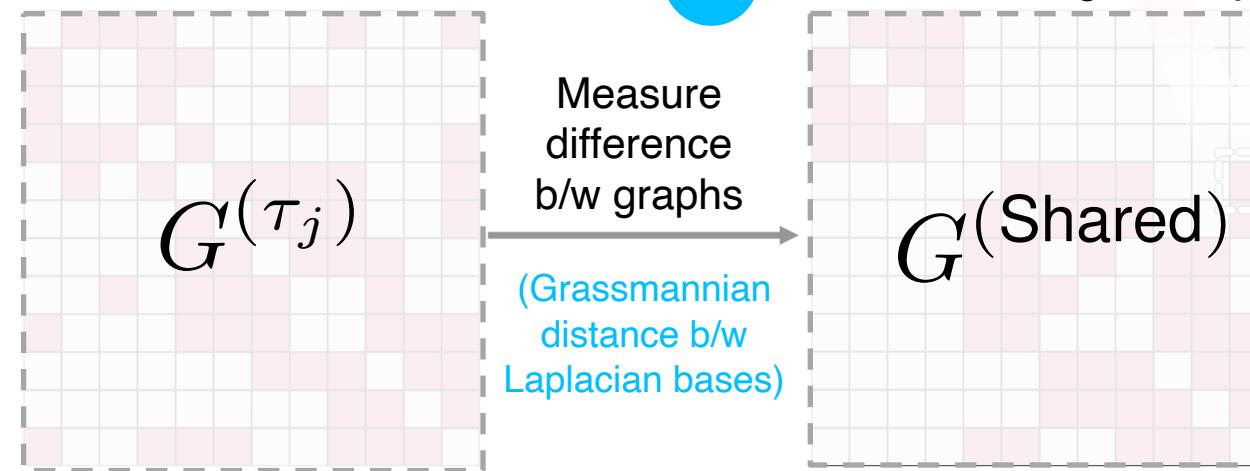


3  
Perform CCA

1  
Cell-by-cell  
nearest-neighbor  
graph  
(RNA modality)



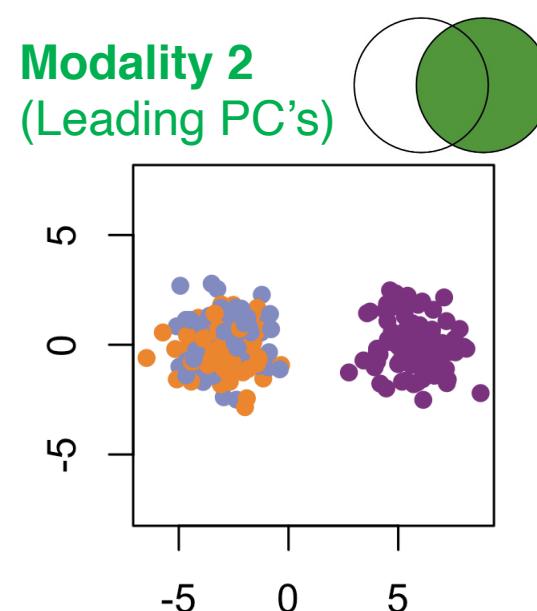
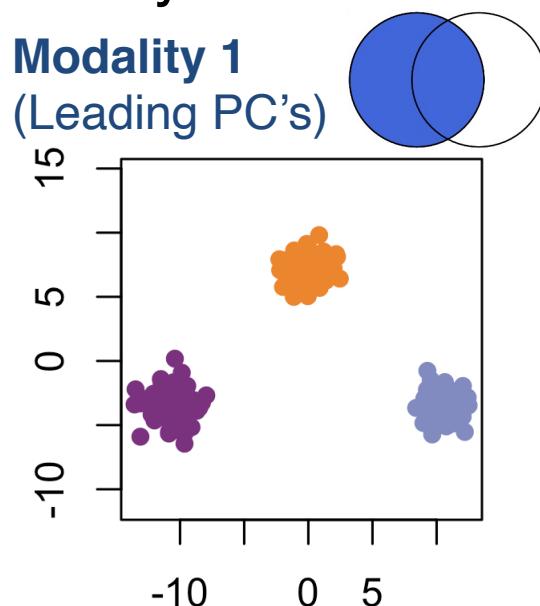
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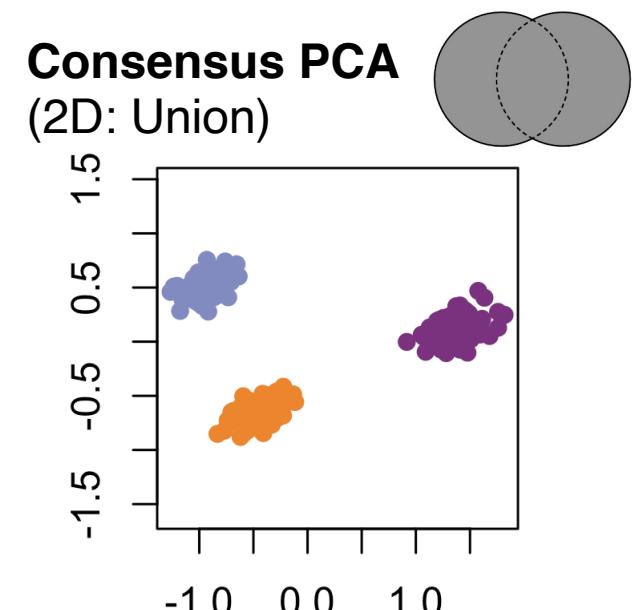
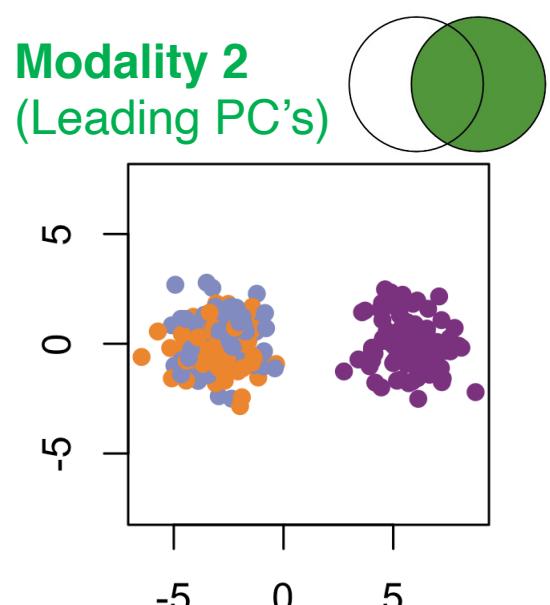
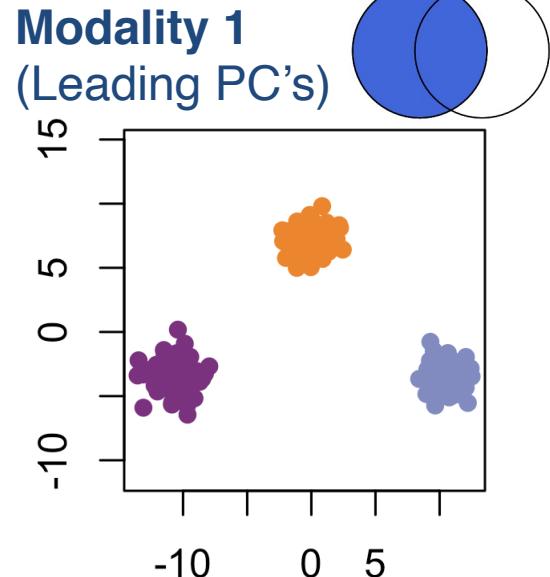
Minimize via 0-order coordinate descent

4

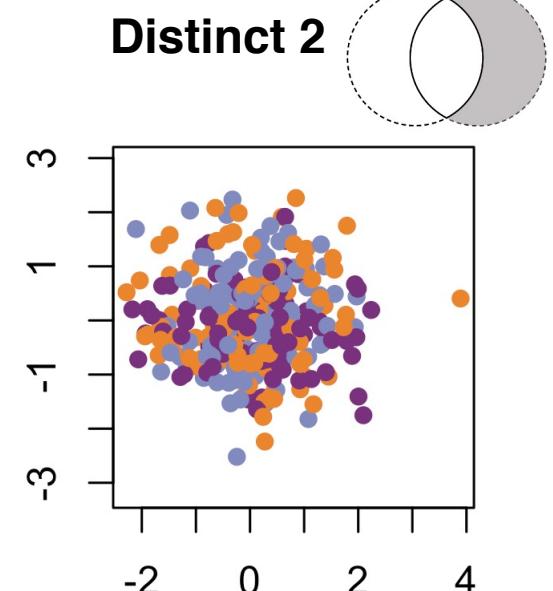
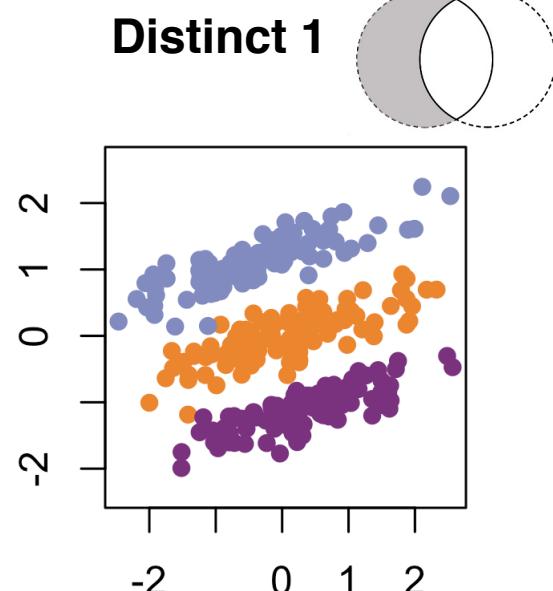
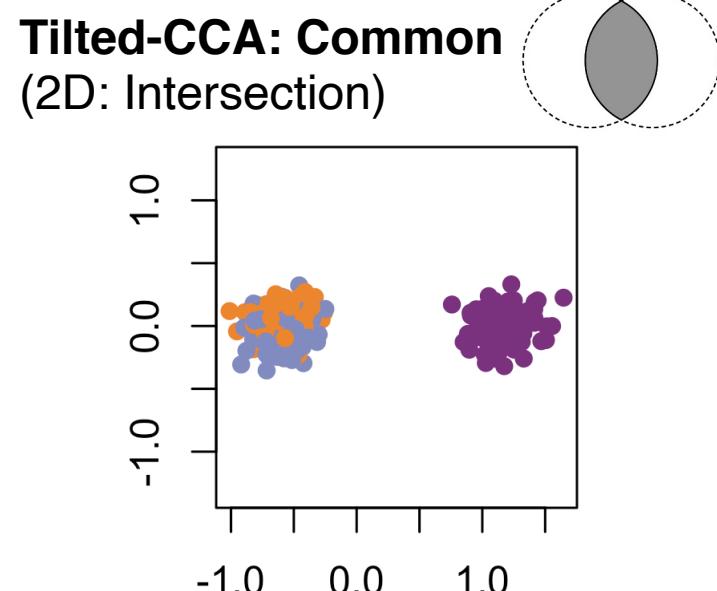
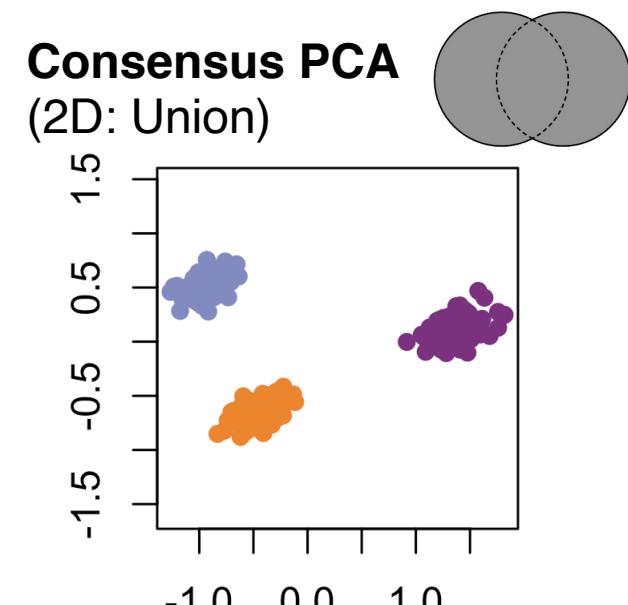
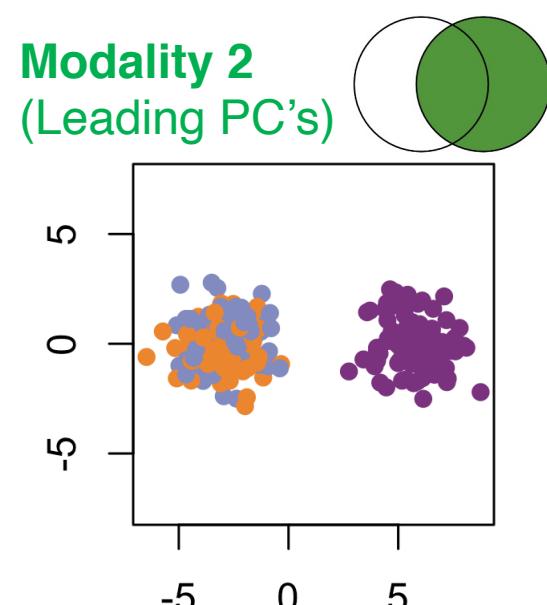
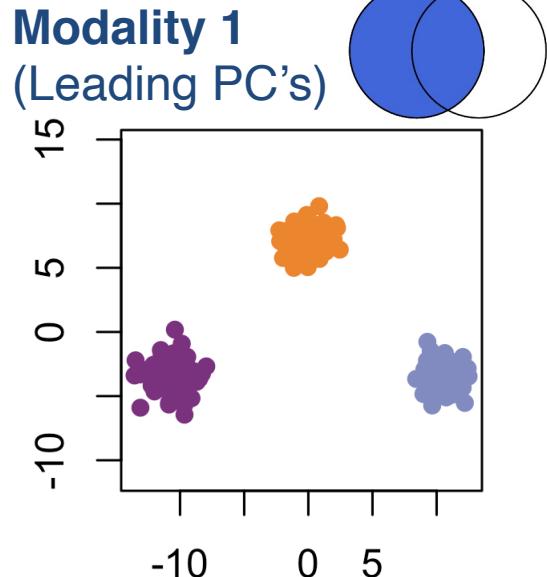
## Simulation 1: Toy



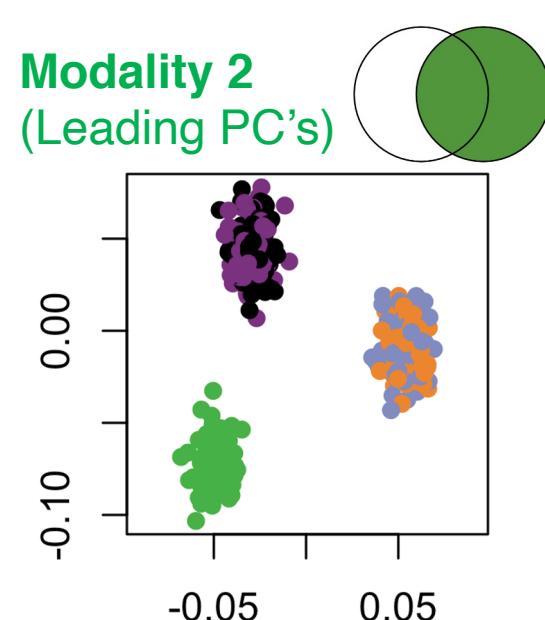
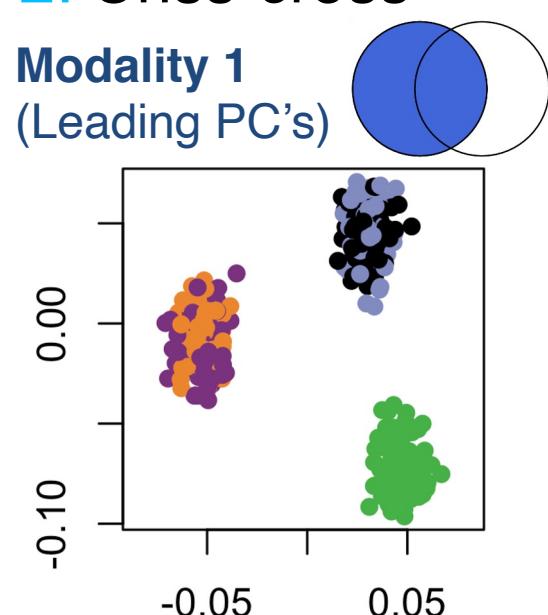
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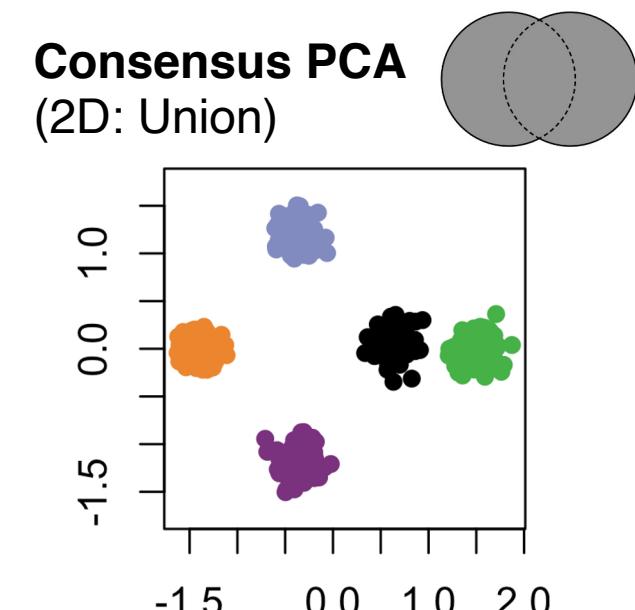
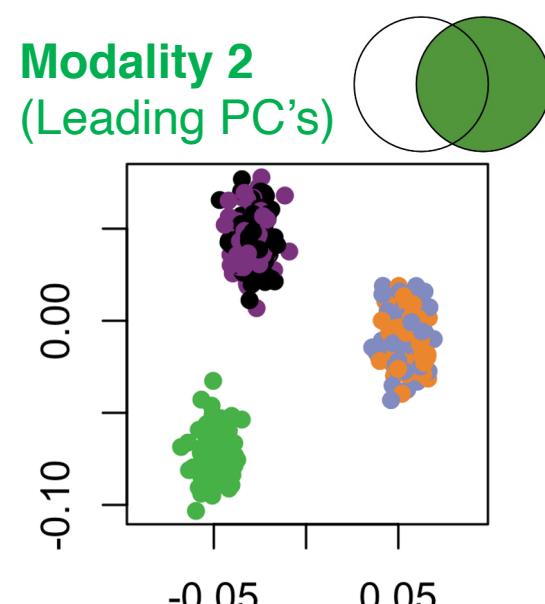
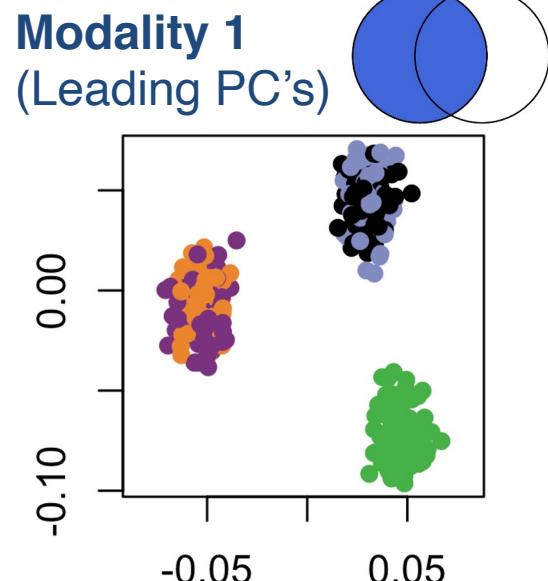
## Simulation 1: Toy



## Simulation 2: Criss-cross

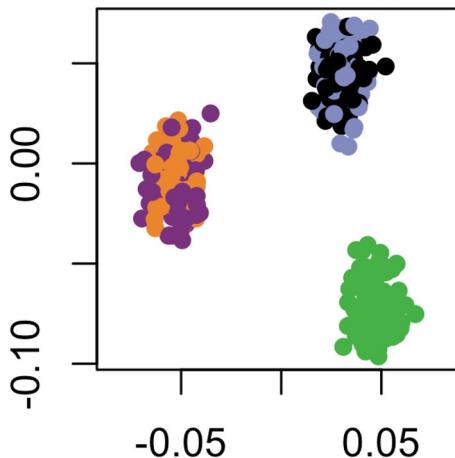


## Simulation 2: Criss-cross

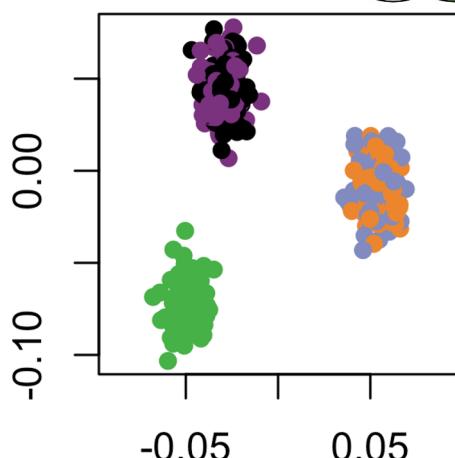


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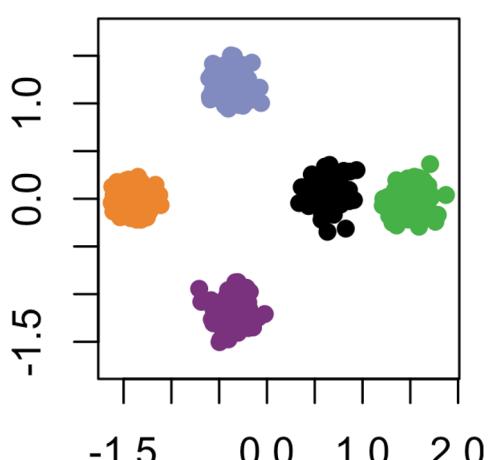
**Modality 1**  
(Leading PC's)



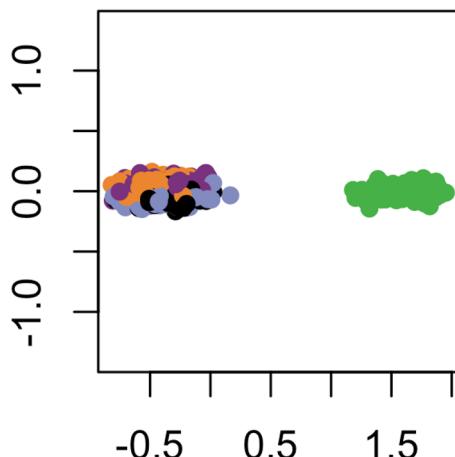
**Modality 2**  
(Leading PC's)



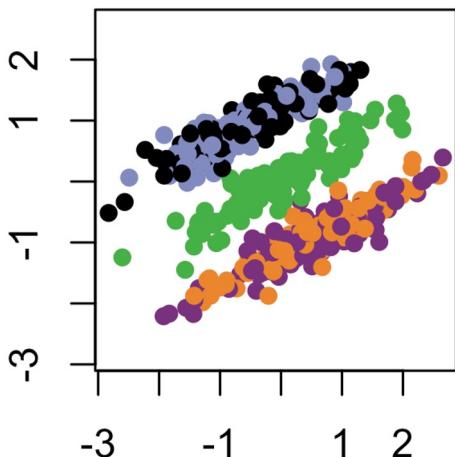
**Consensus PCA**  
(2D: Union)



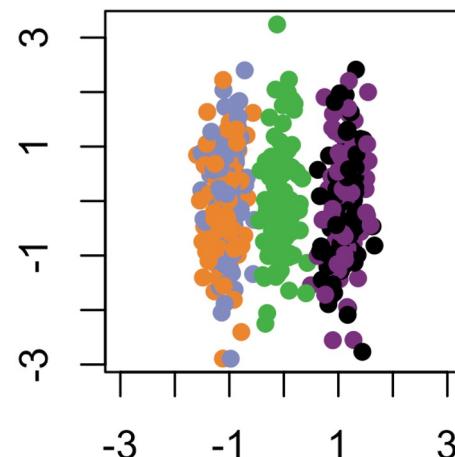
**Tilted-CCA: Common**  
(2D: Intersection)



**Distinct 1**



**Distinct 2**

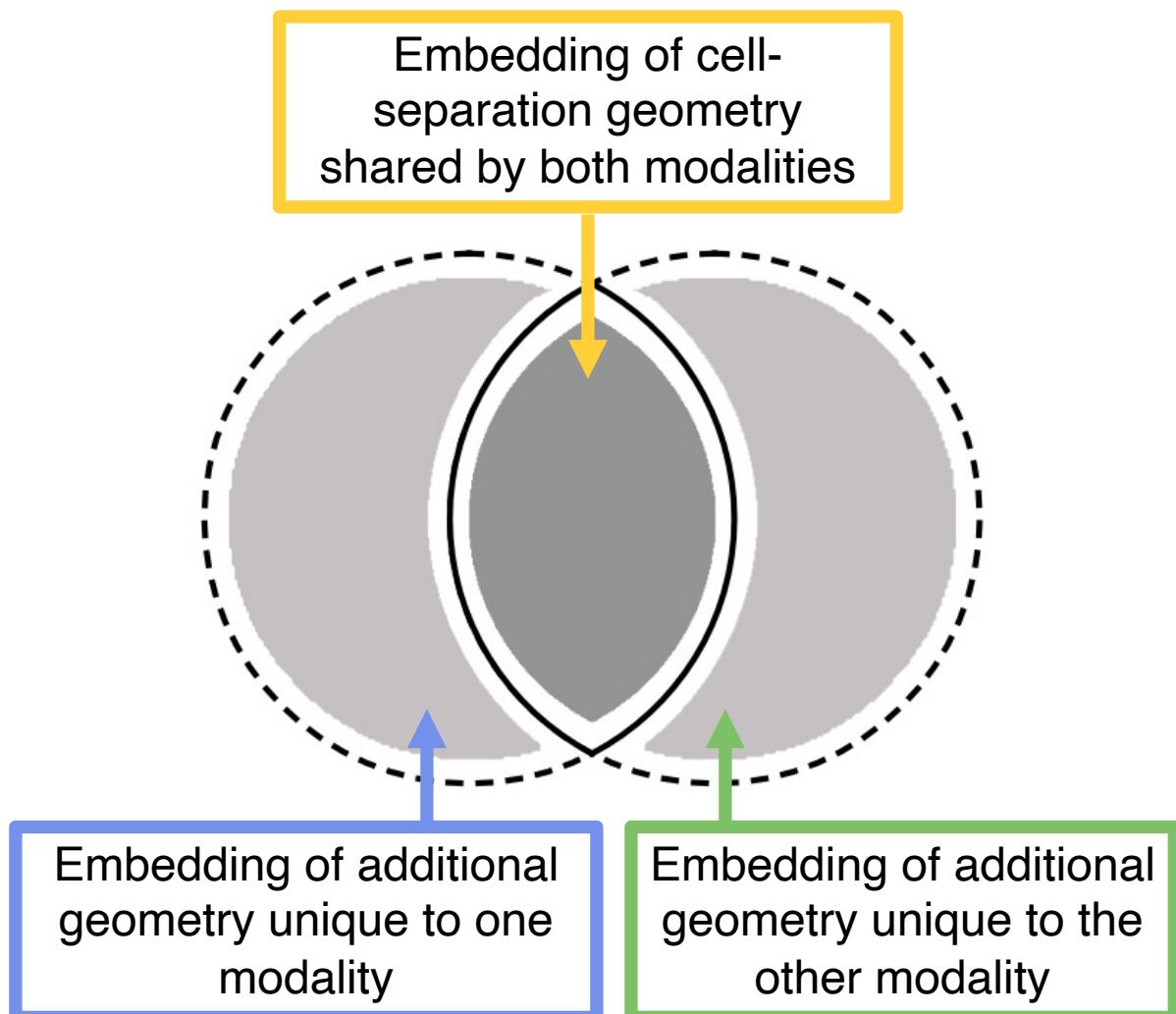


## **Single-cell investigation:**

New perspective of developmental biology, now that we have estimated the shared/unique geometry

## Recap of the biological goals:

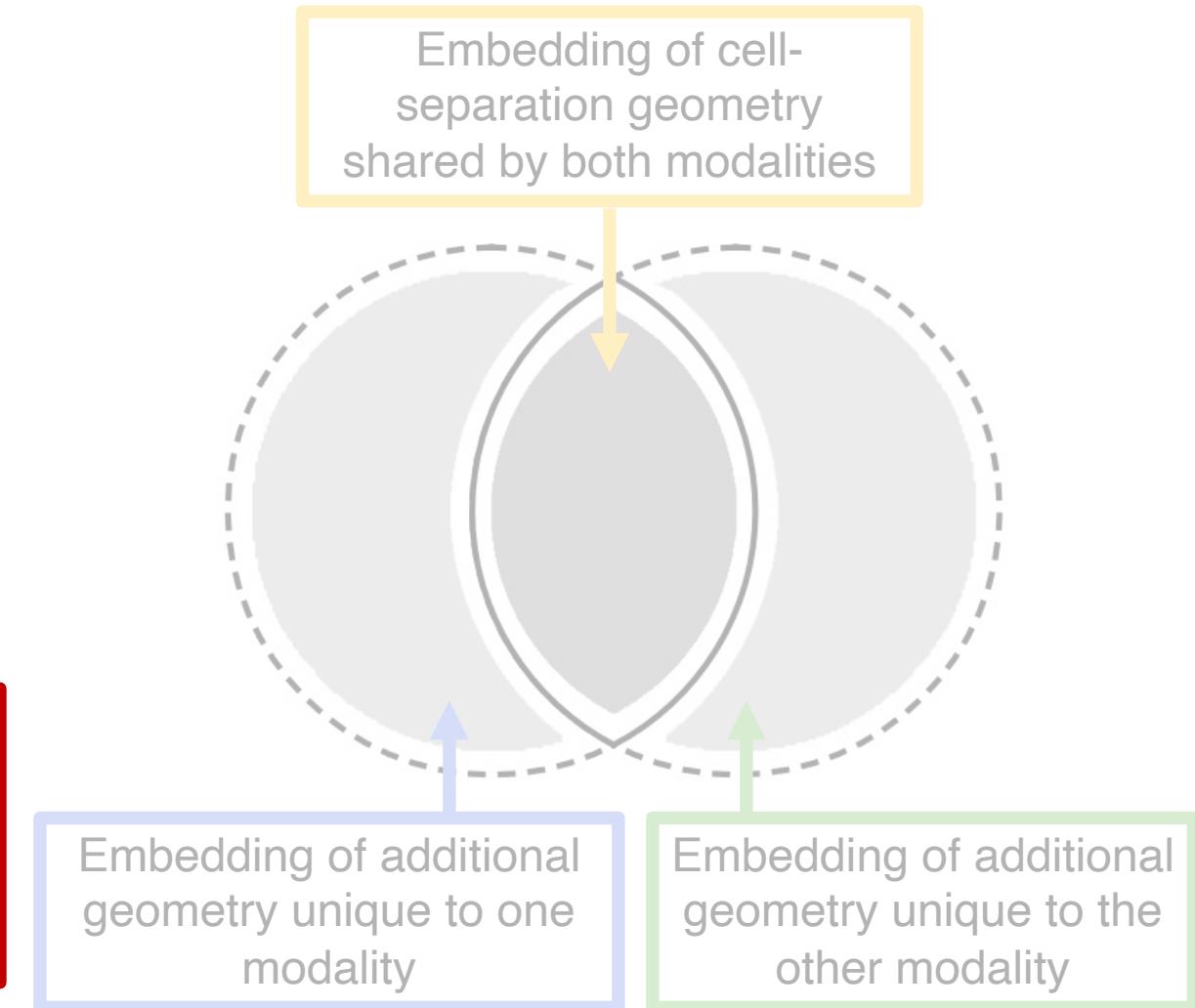
1. **(Experimental design)**: Which pair of modalities should biologist sequence to have the most comprehensive understanding?
2. **(Variable selection)**: For RNA-Protein data, how can we pick the antibodies that contribute the most additional information to the RNA modality?
3. **(Developmental biology)**: Can the amount of coordination between two modalities tell us if a cell is in a steady-state or is undergoing development?



## Recap of the biological goals:

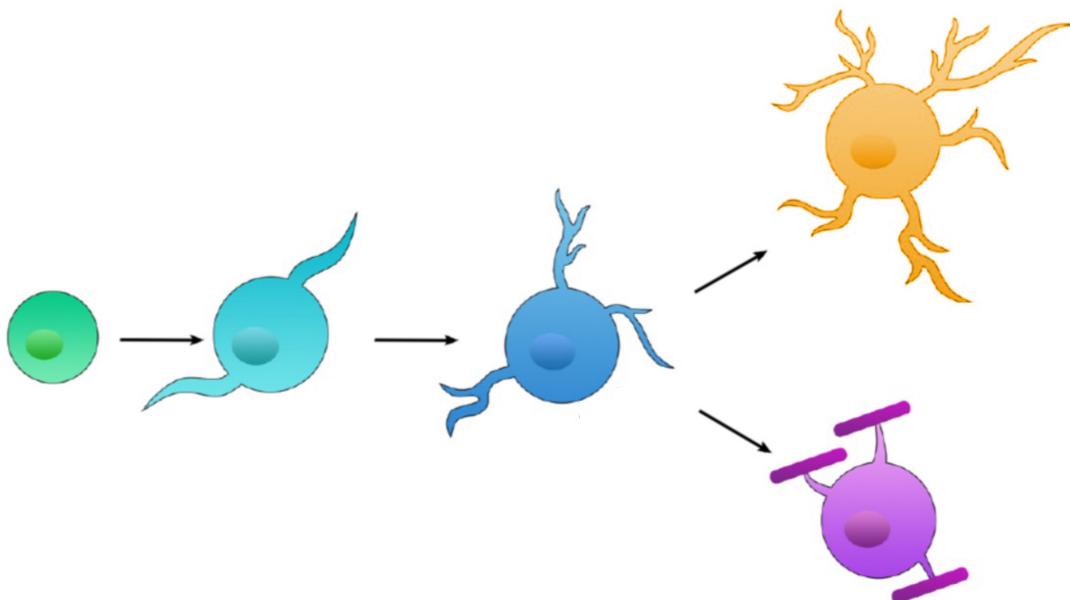
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Focus of the remainder of the talk

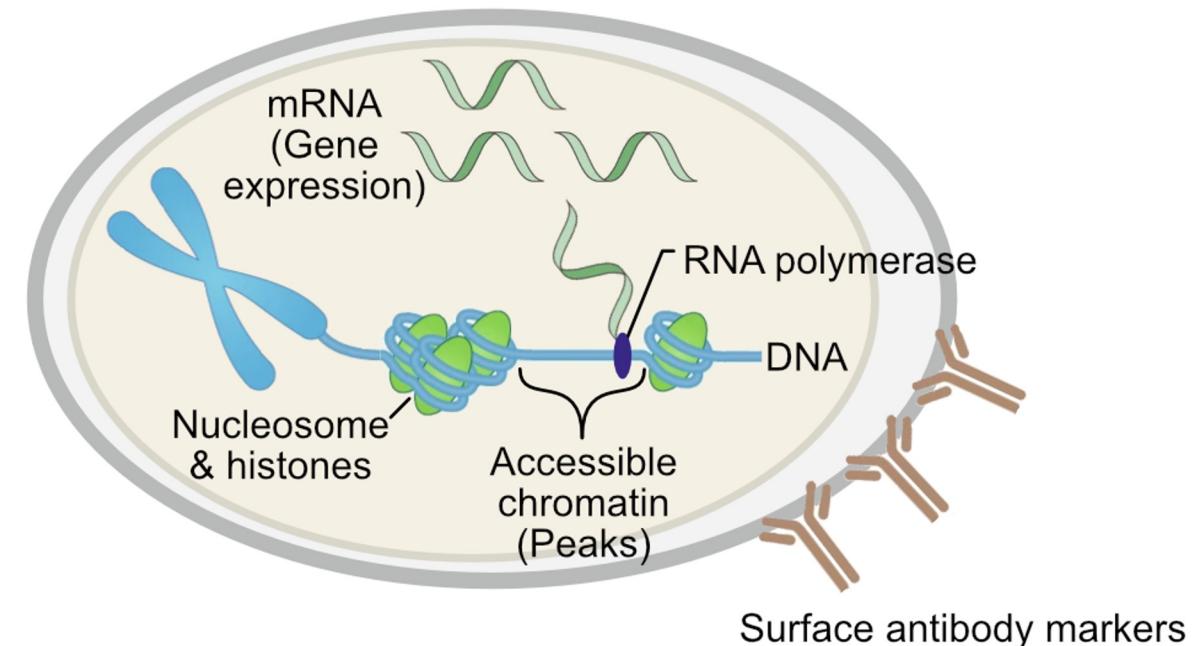


In developmental biology, we're interested in studying how cells continually specialize over time despite given static snapshots.

Progenitor cells  
(Youngest)



Mature neurons  
(Oldest)

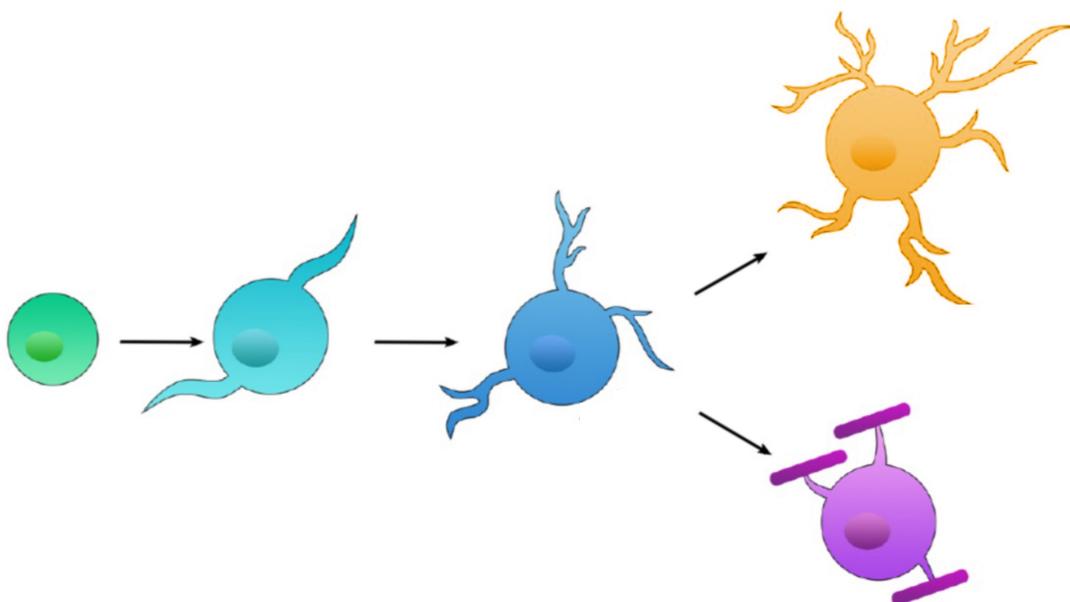


Existing methods:

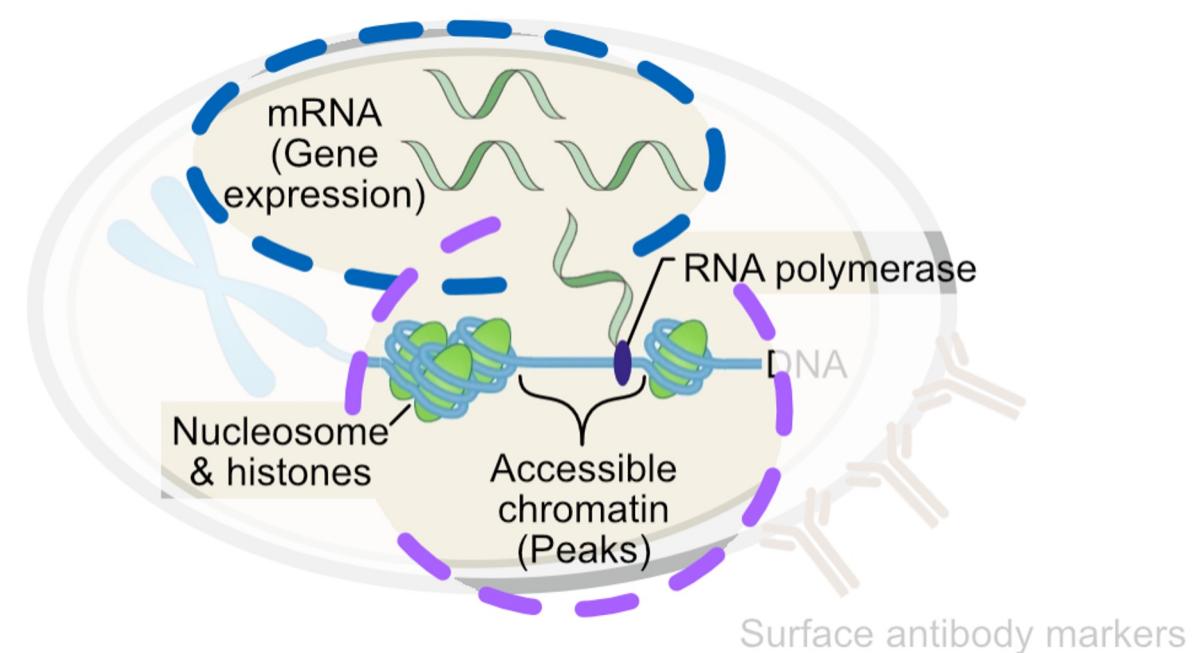
- Monocle (Trapnell et al., Nature Biotech., 2014),
- Slingshot (Dudoit et al., BMC Genomics, 2018),
- RNA velocity (Kharchenko et al., Nature, 2018)

In developmental biology, we're interested in studying how cells continually specialize over time despite given static snapshots.

Progenitor cells  
(Youngest)



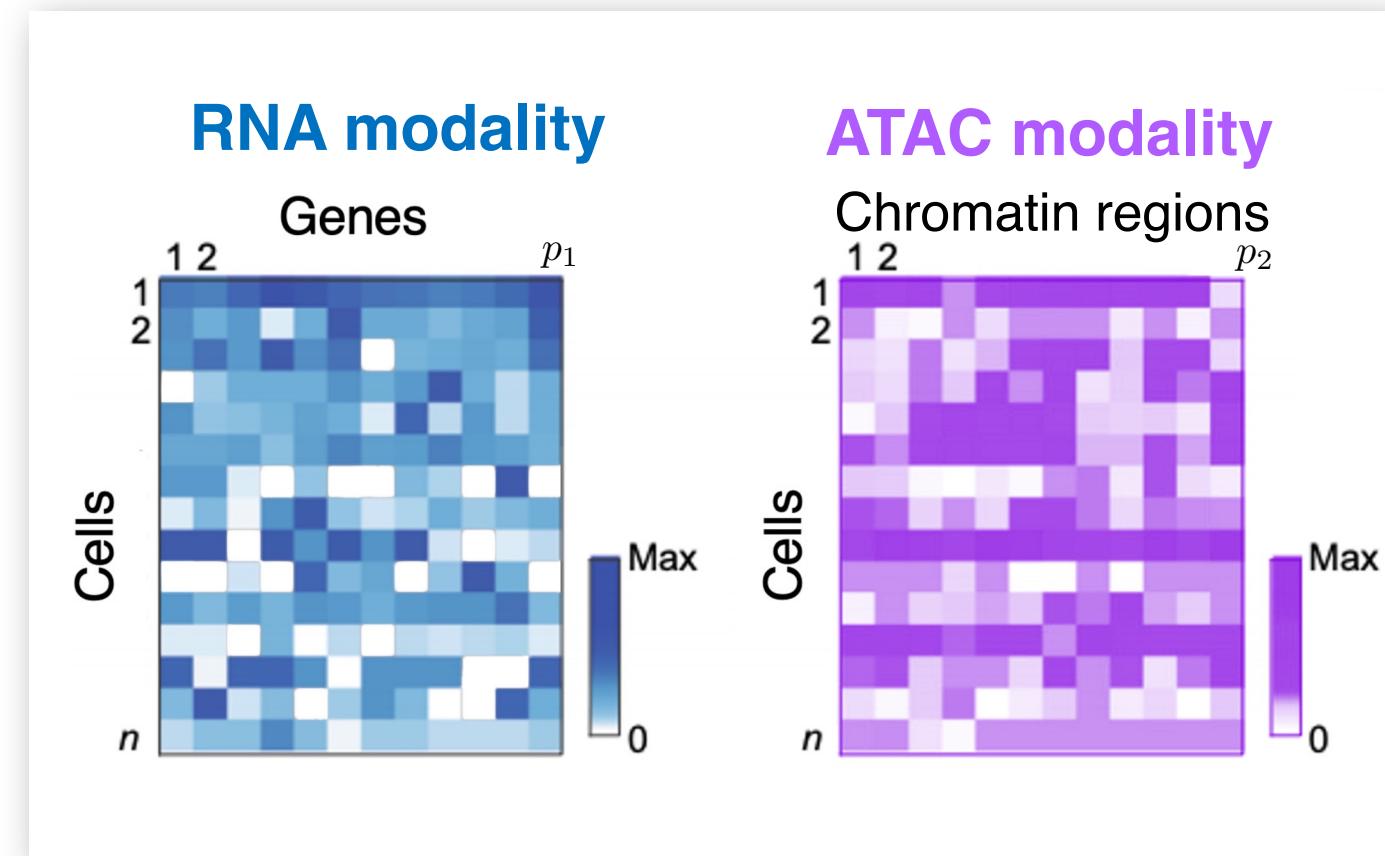
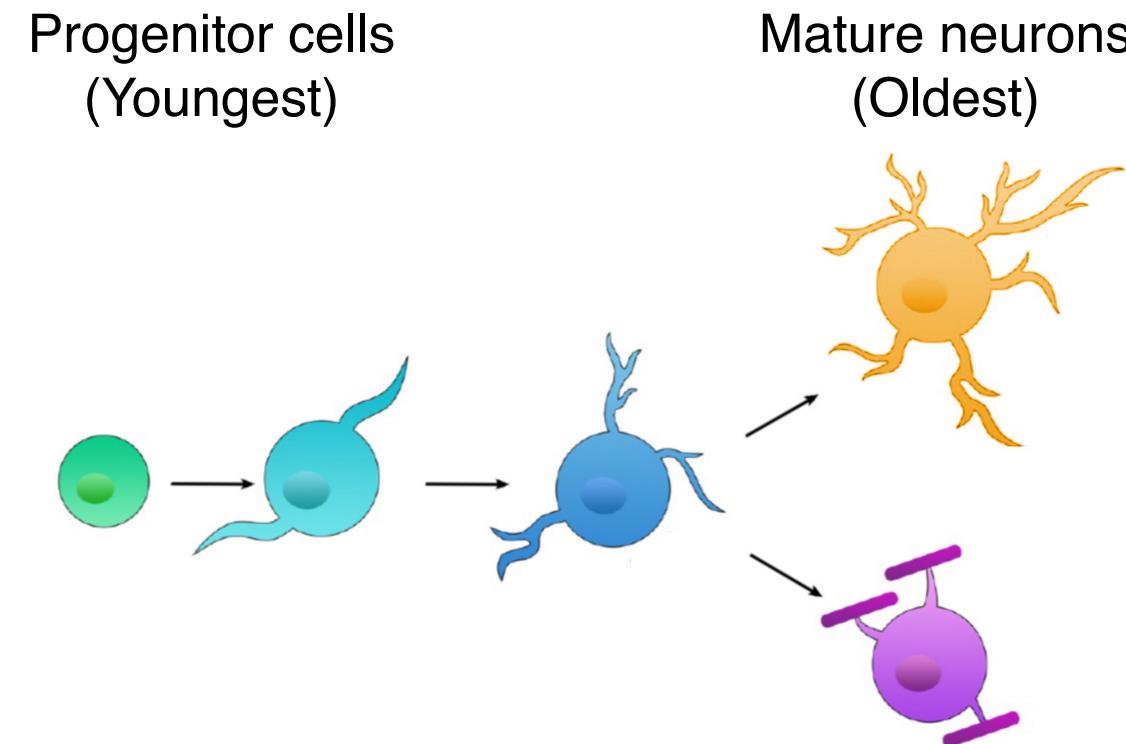
Mature neurons  
(Oldest)



Existing methods:

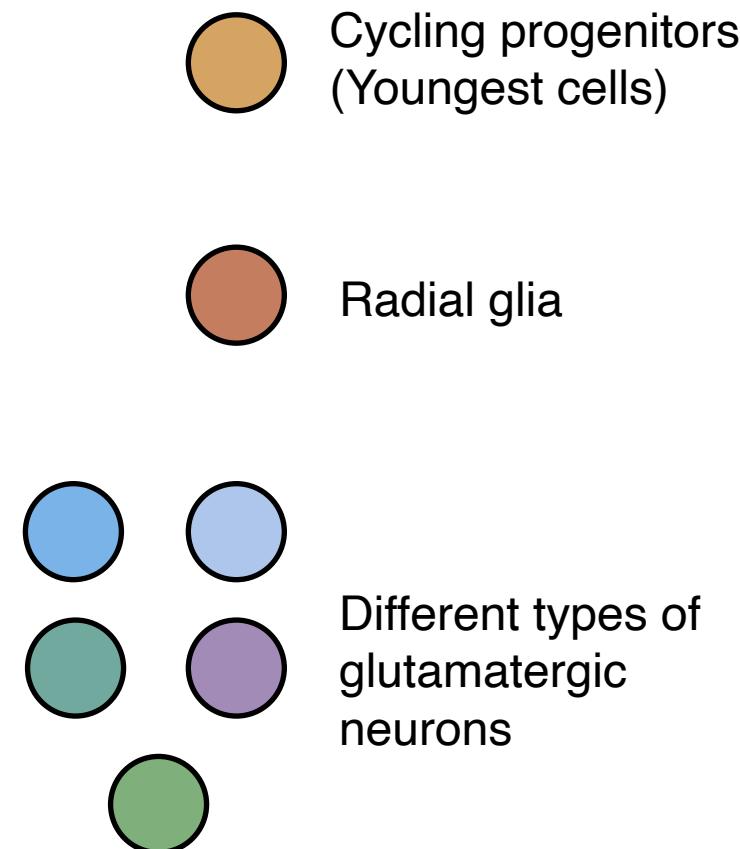
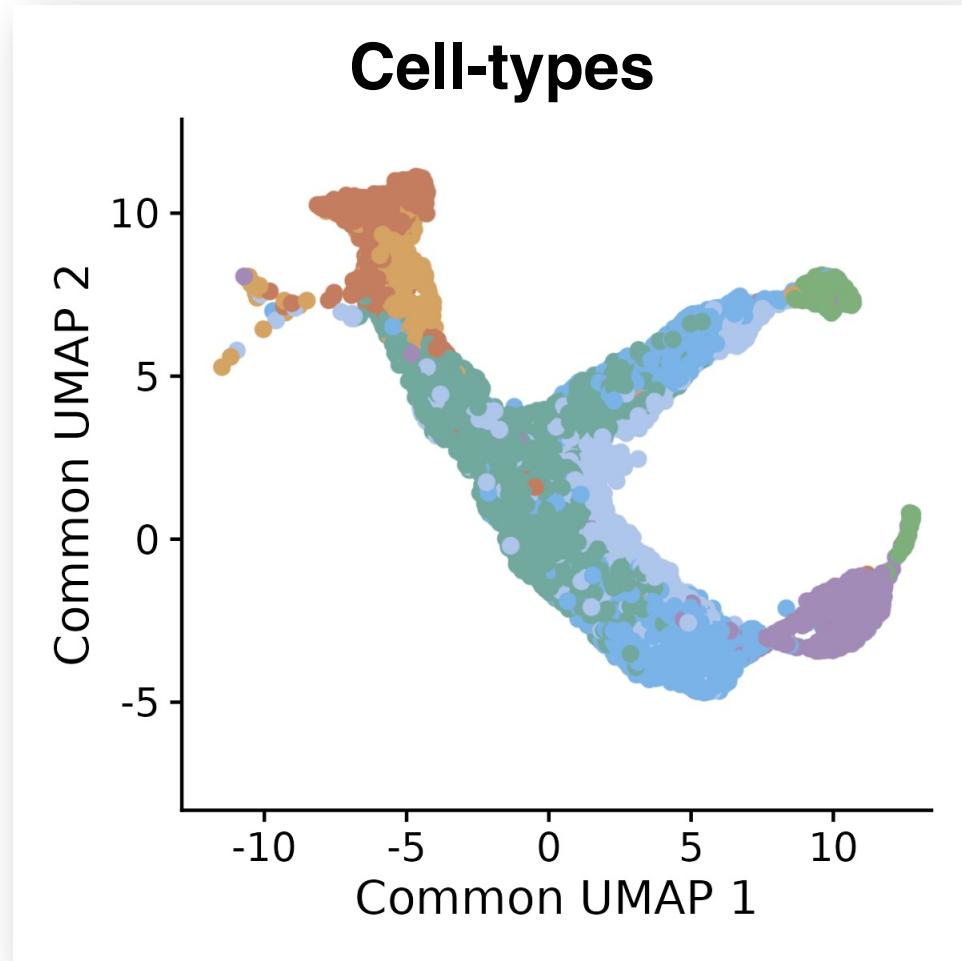
- Monocle (Trapnell et al., Nature Biotech., 2014),
- Slingshot (Dudoit et al., BMC Genomics, 2018),
- RNA velocity (Kharchenko et al., Nature, 2018)

In developmental biology, we're interested in studying how cells continually specialize over time despite given static snapshots.



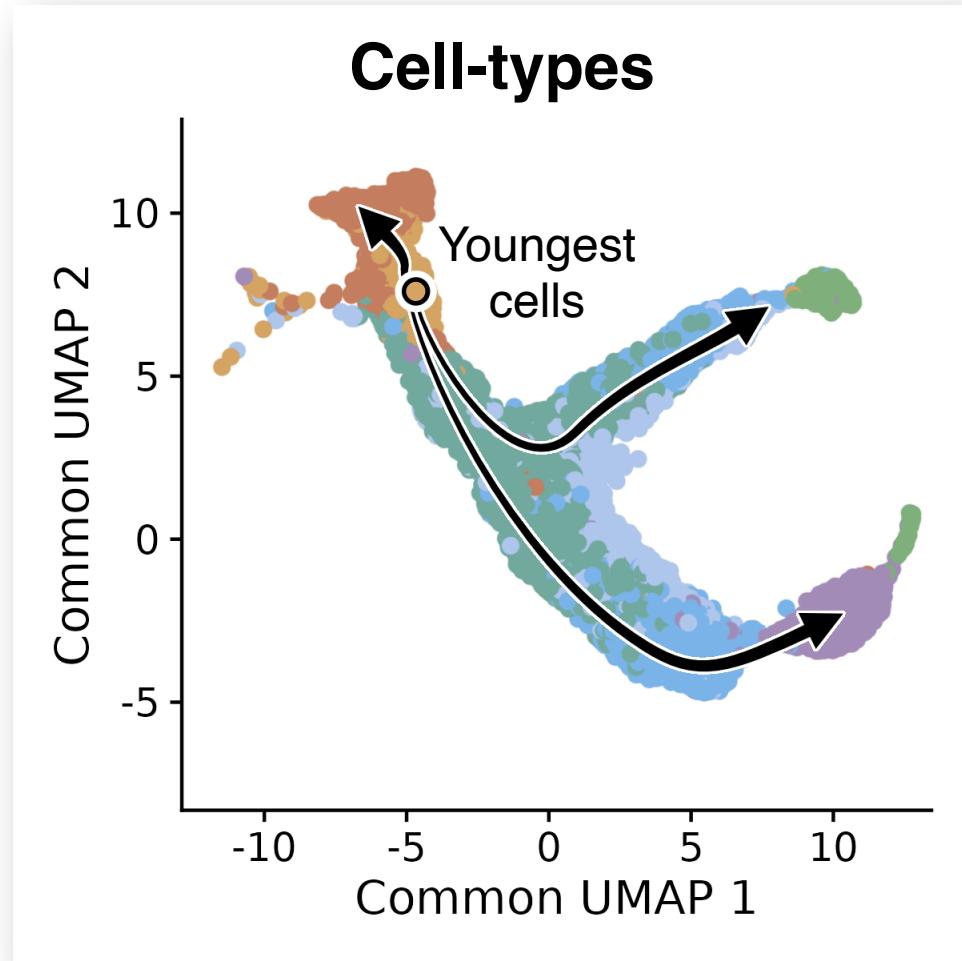
While existing methods focusing on one modality tell us how cells develop ("layout of the highway"), we want to know what status the cell is in ("speed of traffic").

Knowing the developmental trajectories is not the same as knowing if a cell undergoing development or is in steady-state.

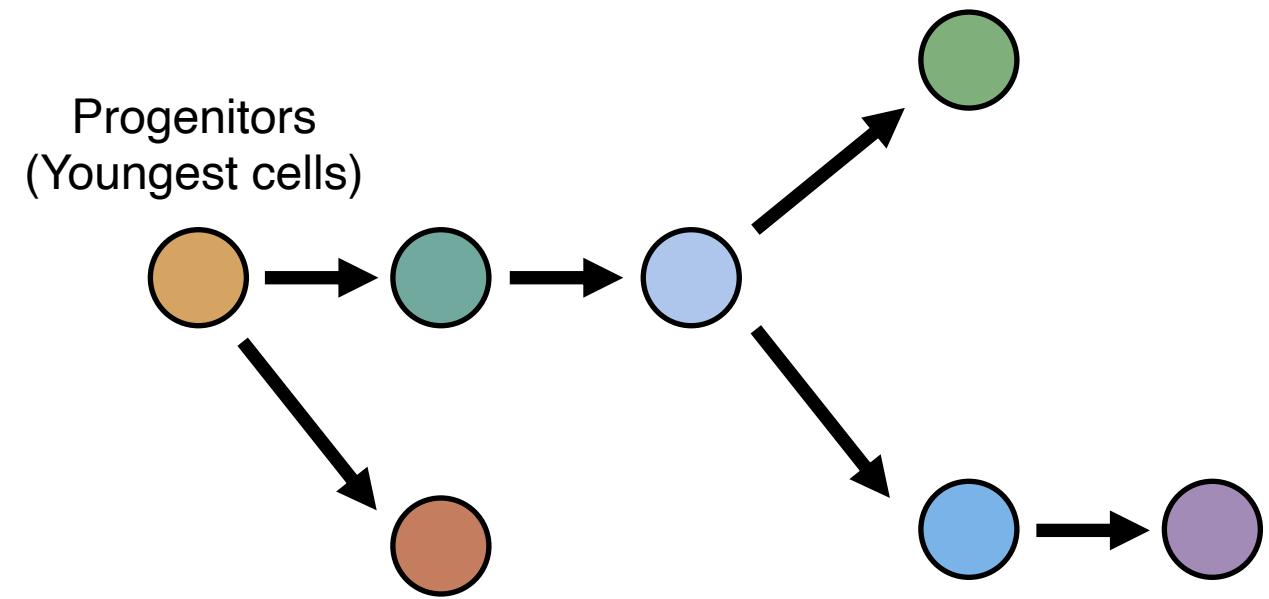


**Human brain development** (10x. RNA & ATAC.  
Greenleaf et al., Cell, 2021): 6000+ cells

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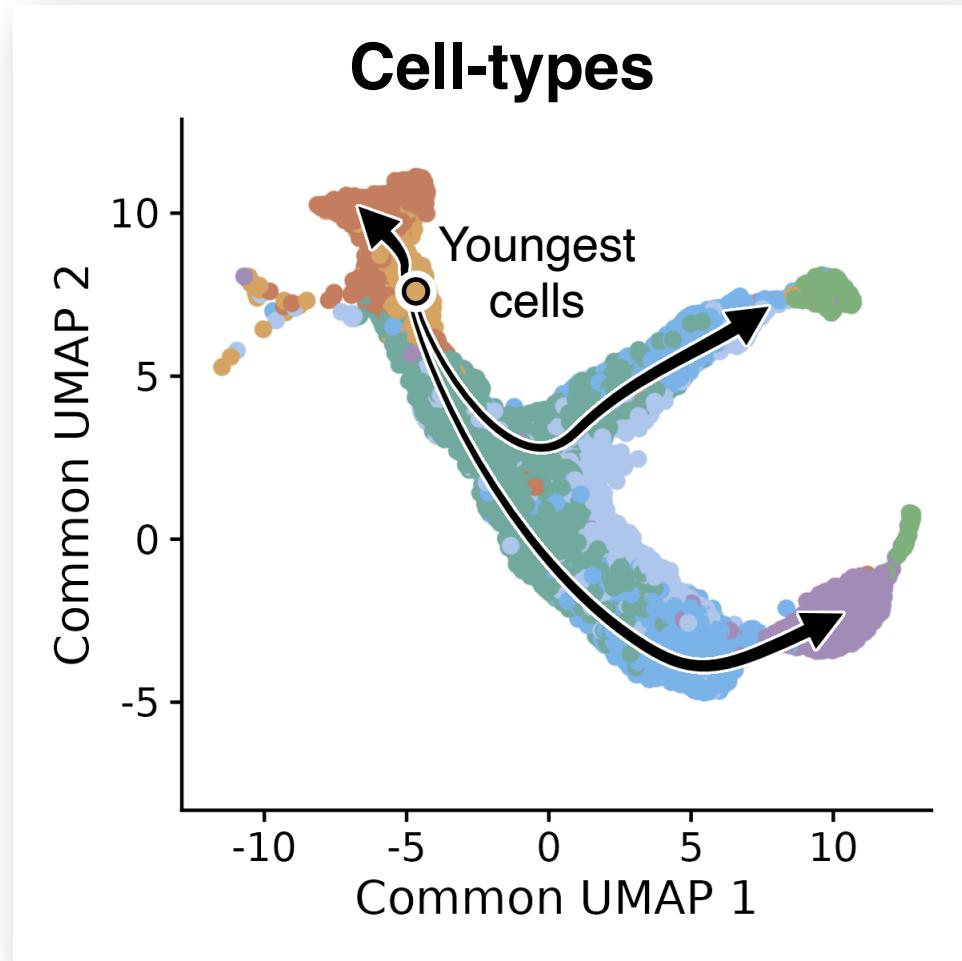


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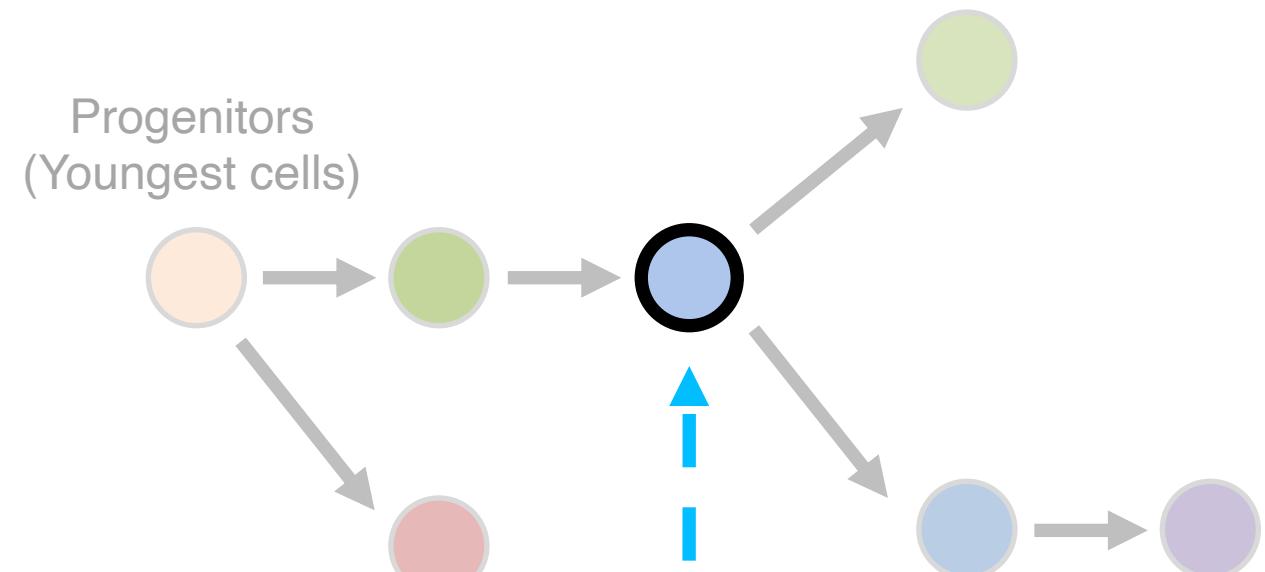


Existing methods:  
“Layout of the highway”

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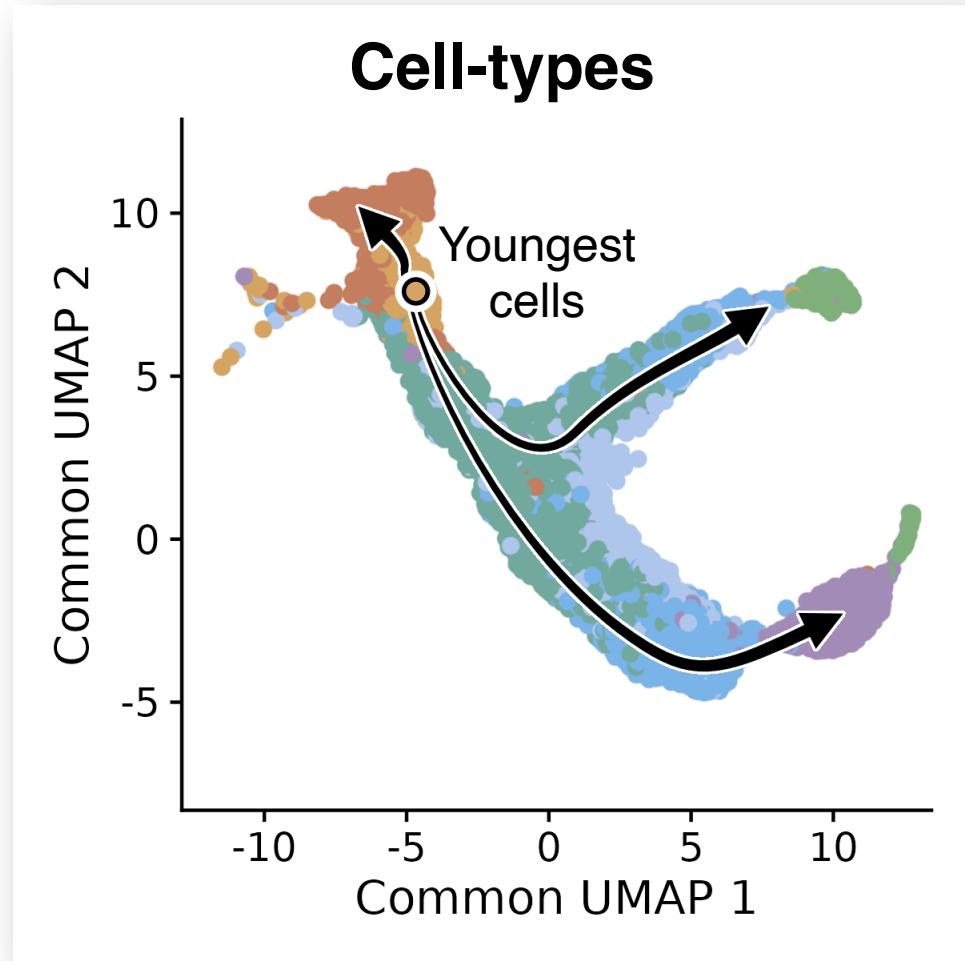


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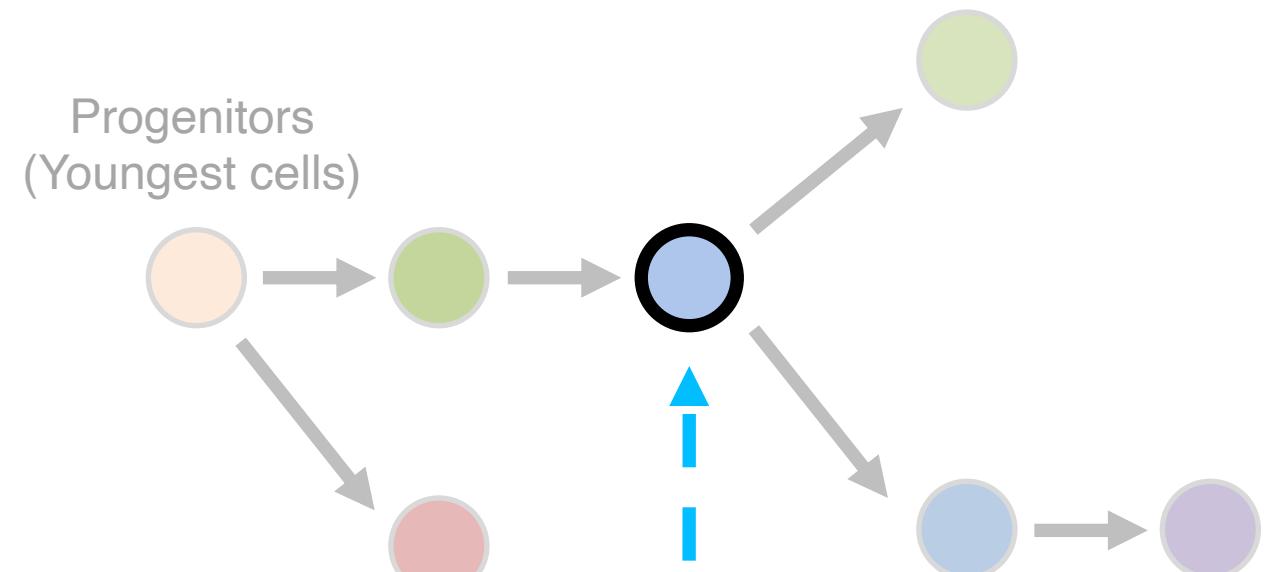


**“Speed of traffic”:**  
Are the neurons rapidly passing  
through OR idling around/waiting for  
an external stimulus?

Knowing the developmental trajectories is not the same as knowing if a cell undergoing development or is in steady-state.



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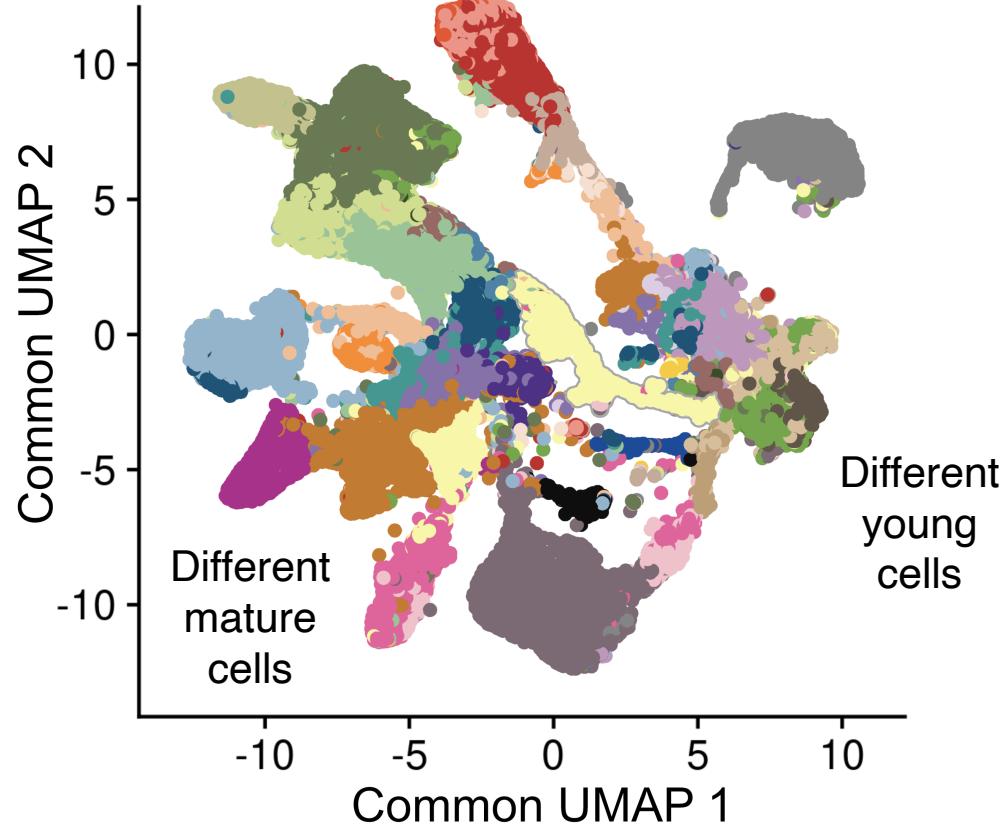


**“Speed of traffic”:**  
Are the neurons in-development  
**OR** in steady-state?

## Brief aside: More complex systems

### Mouse embryonic development

(10x. Reik et al., bioRxiv, 2022)

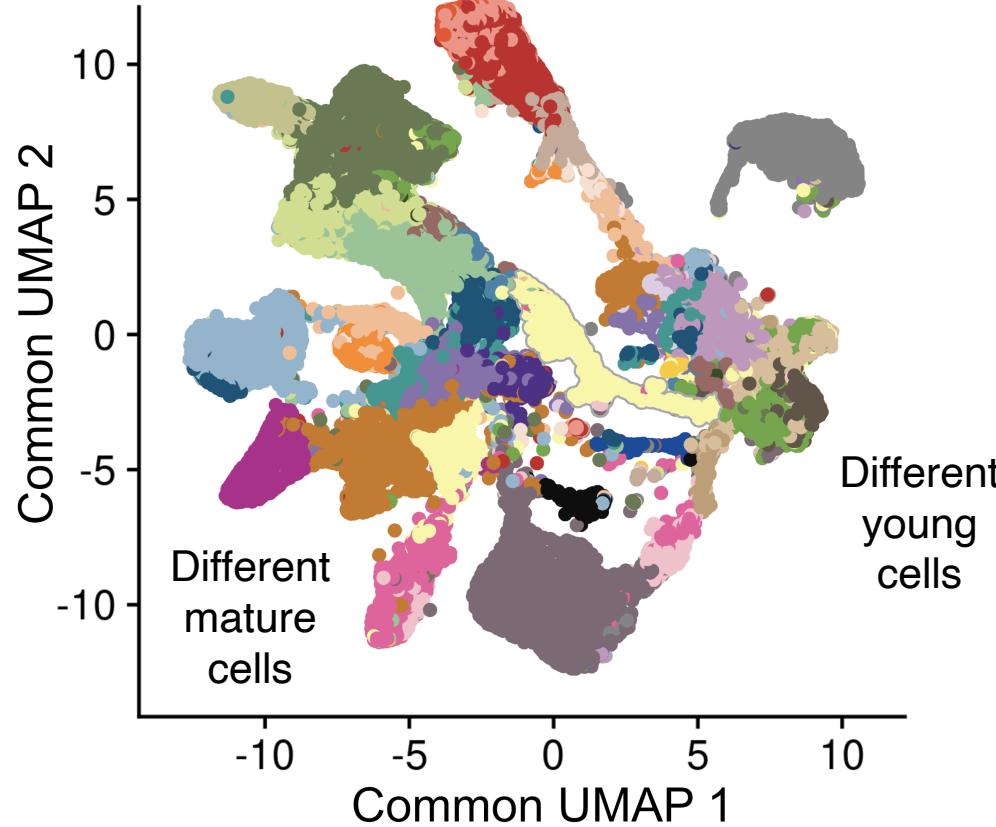


Difficult due to multiple trajectories

## Brief aside: More complex systems

### Mouse embryonic development

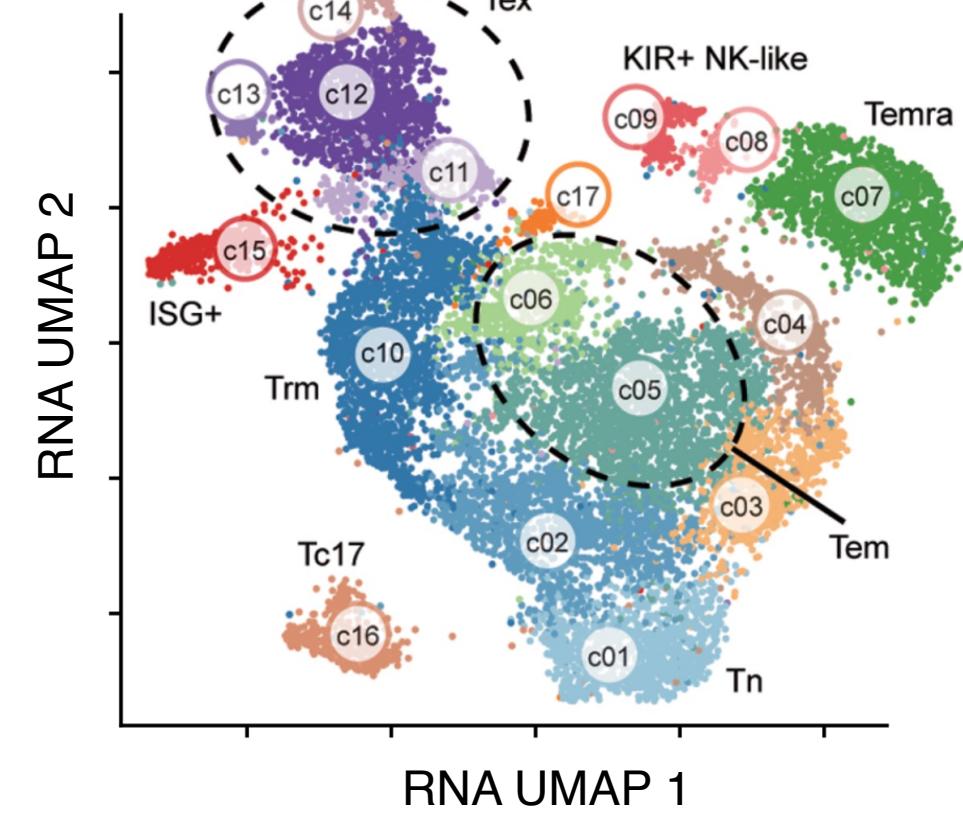
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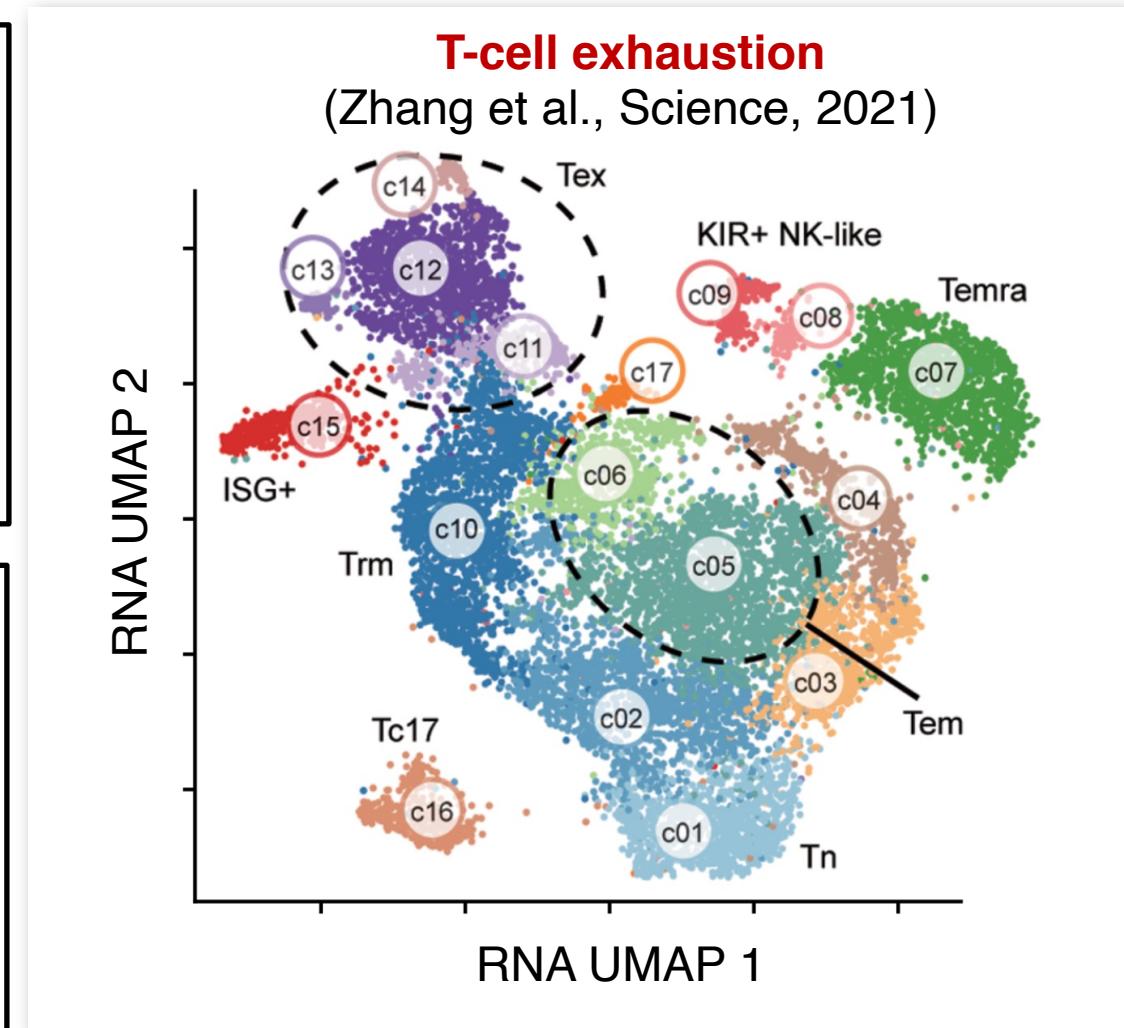
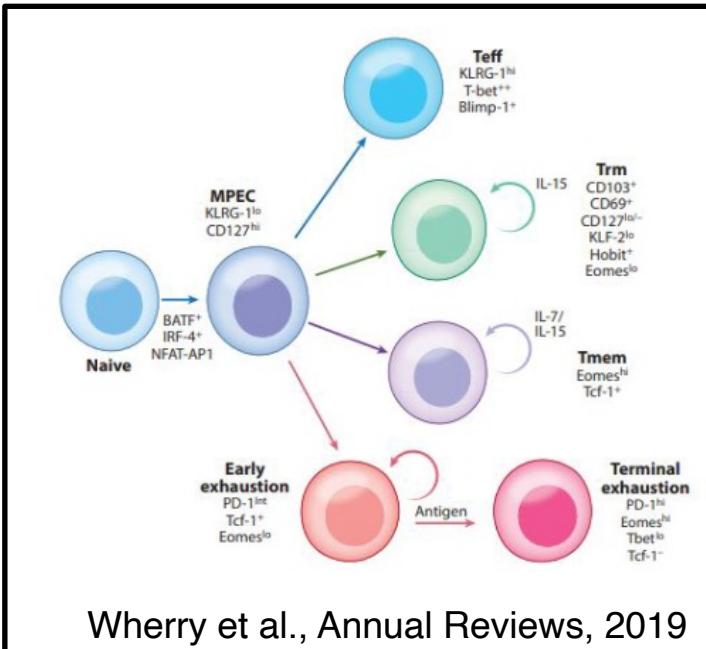
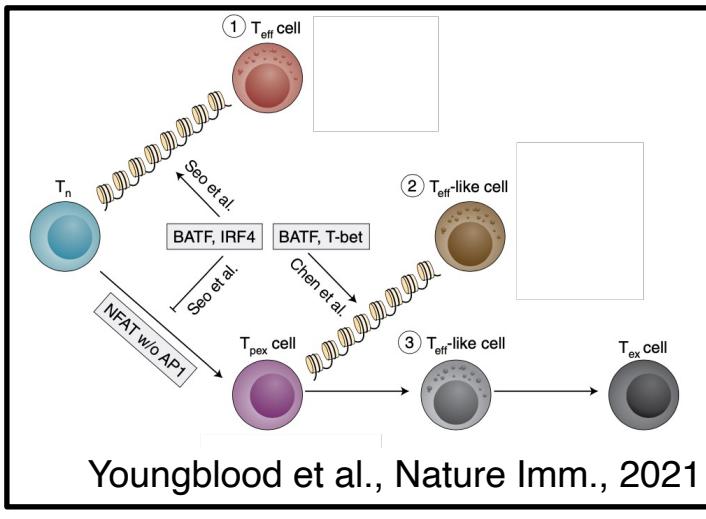
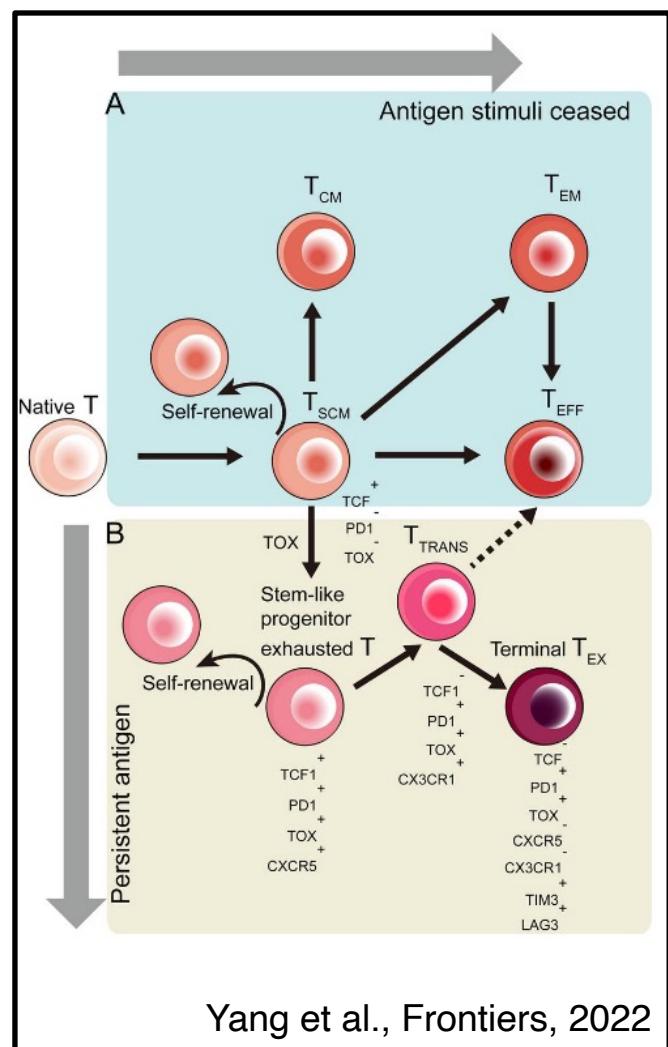
### T-cell exhaustion

(Zhang et al., Science, 2021)



Even more difficult since the biology is still anyone's guess

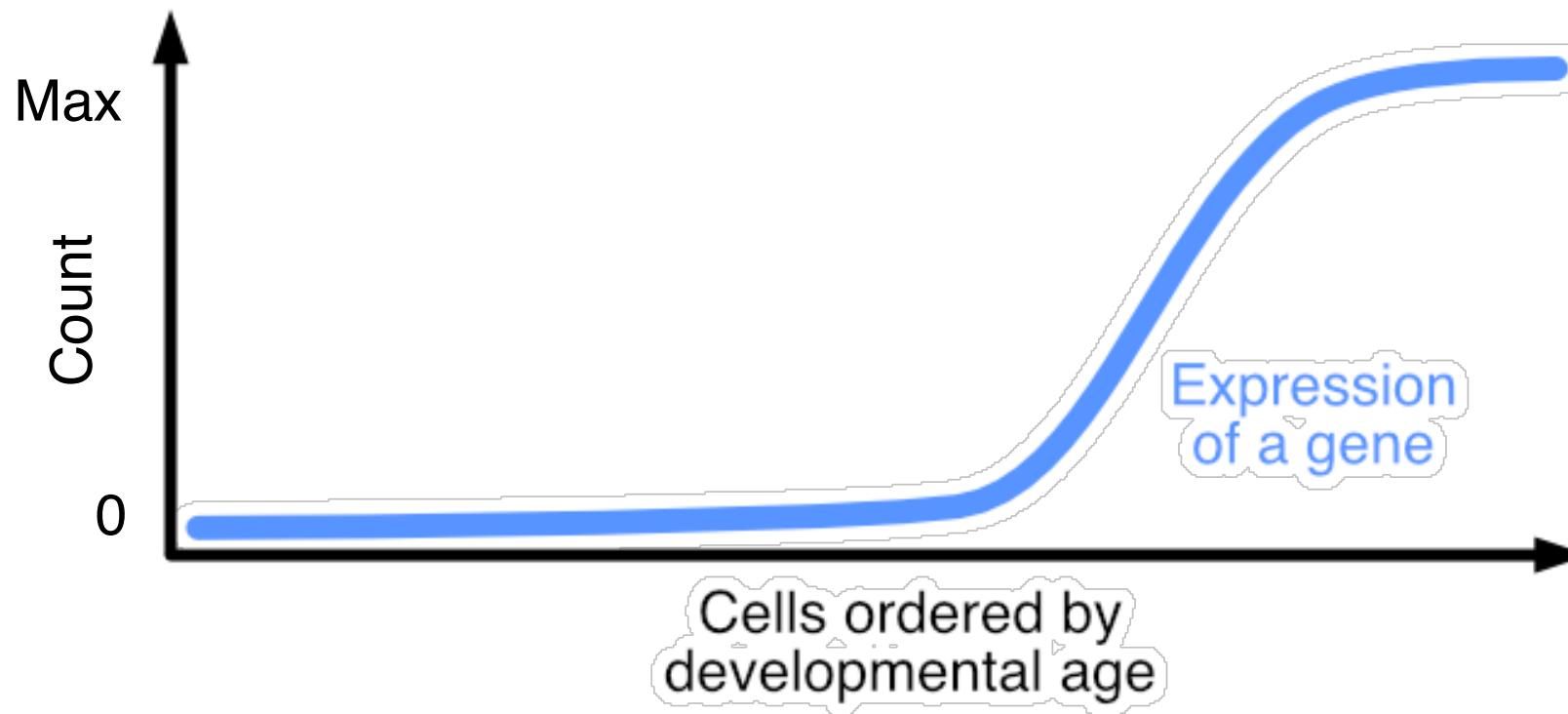
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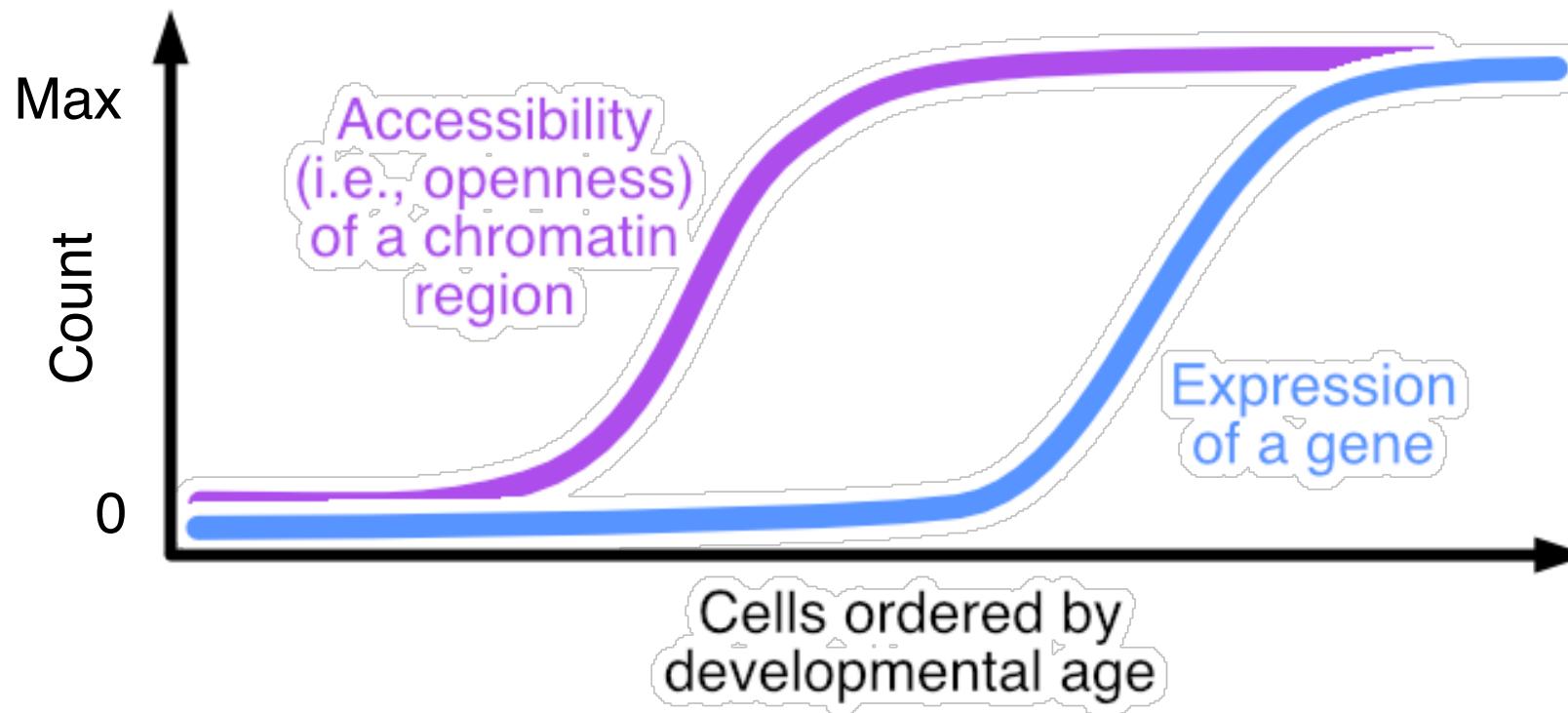
Tilted-CCA can learn which cells are in steady-state via RNA-ATAC relations thanks to principles discovered in previous work.

Schematic from SHARE-seq (Buenrostro et al., Cell, 2020)



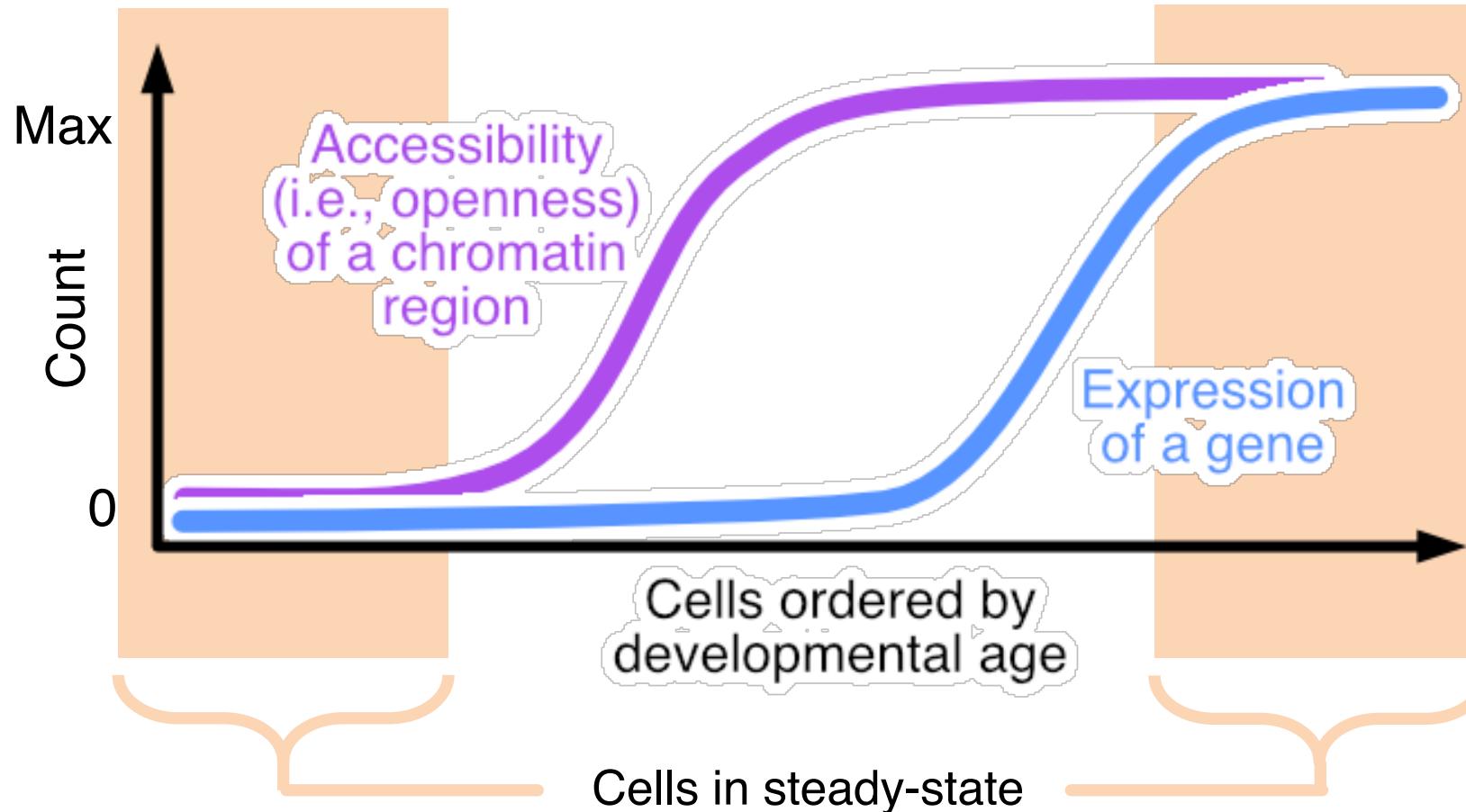
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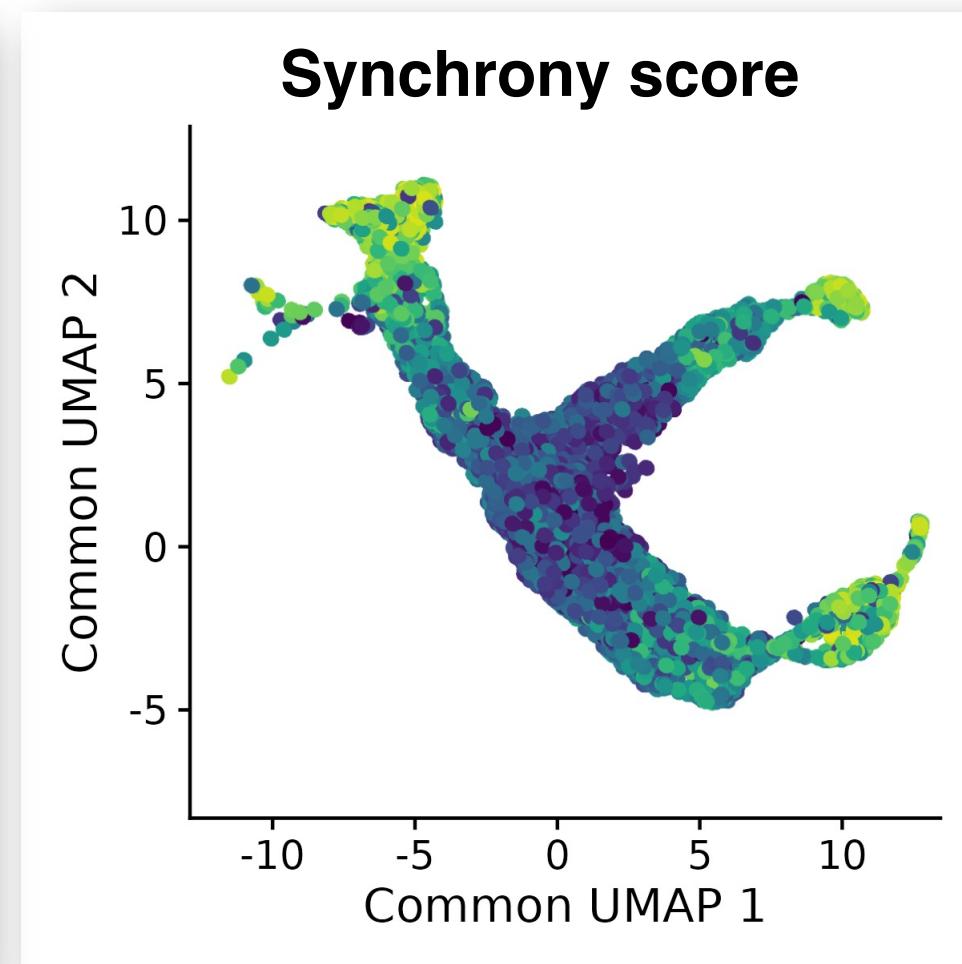
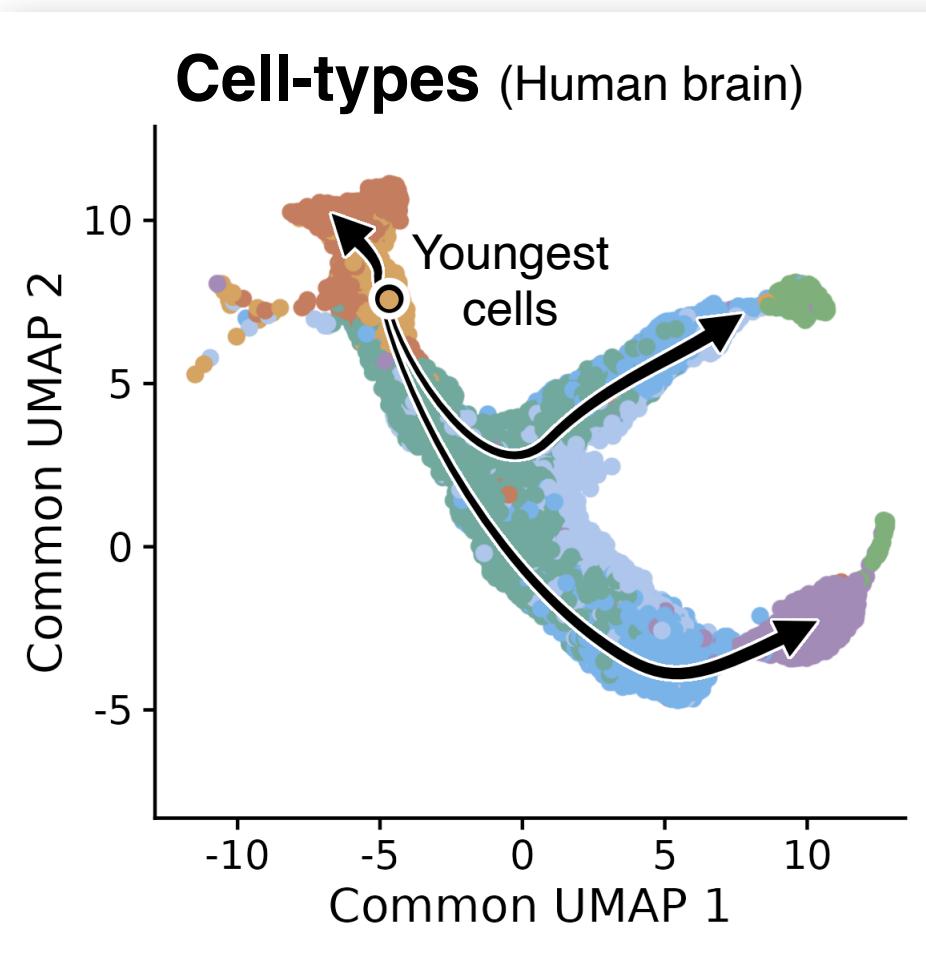
We want to learn which cells are in steady-state (w/o temporal ordering):  
Local geometries similar in both modalities → Small distinct components

**Our contribution:** The synchrony score, which measures the coordination b/w genes and chromatin accessibility within a cell, to reveal if a cell is in steady-state

After fitting Tilted-CCA, for each cell  $i$ : Correlation  $\begin{pmatrix} \text{common component among genes} & , \text{common + distinct components among genes} \end{pmatrix}$

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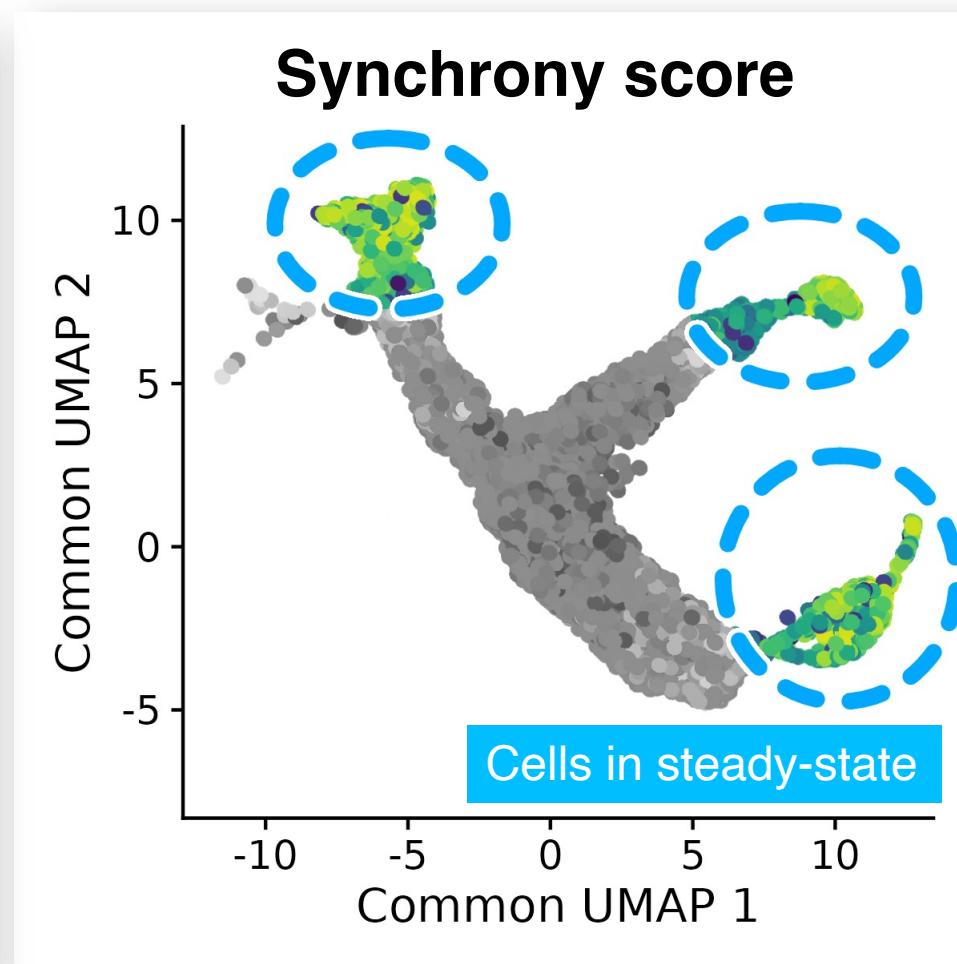
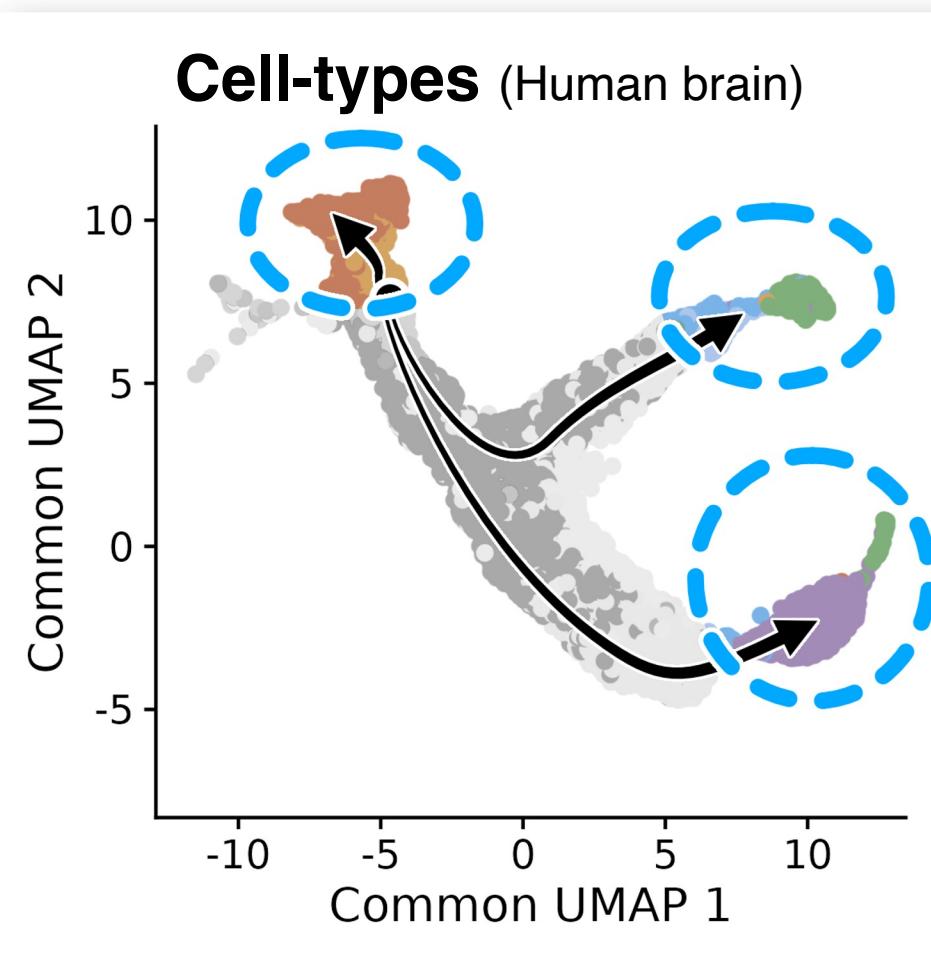
High synchrony score  
(i.e., steady-state)



Low synchrony score  
(i.e., undergoing development)

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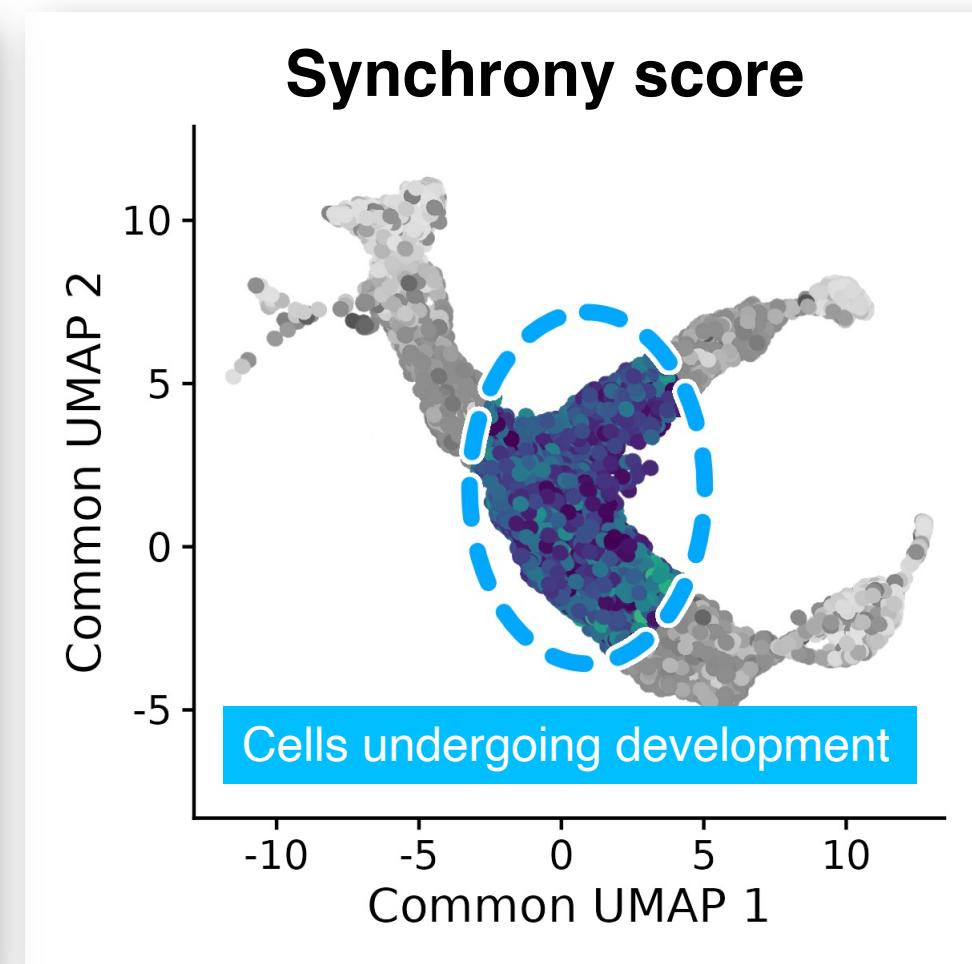
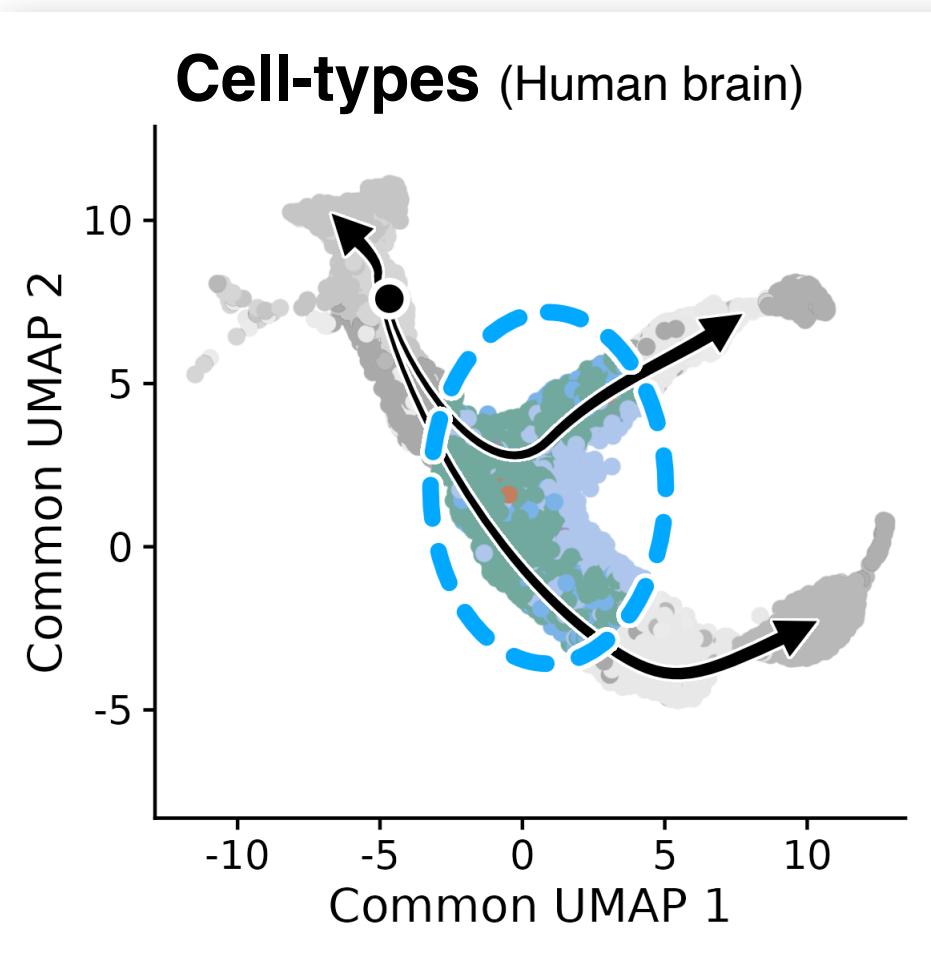
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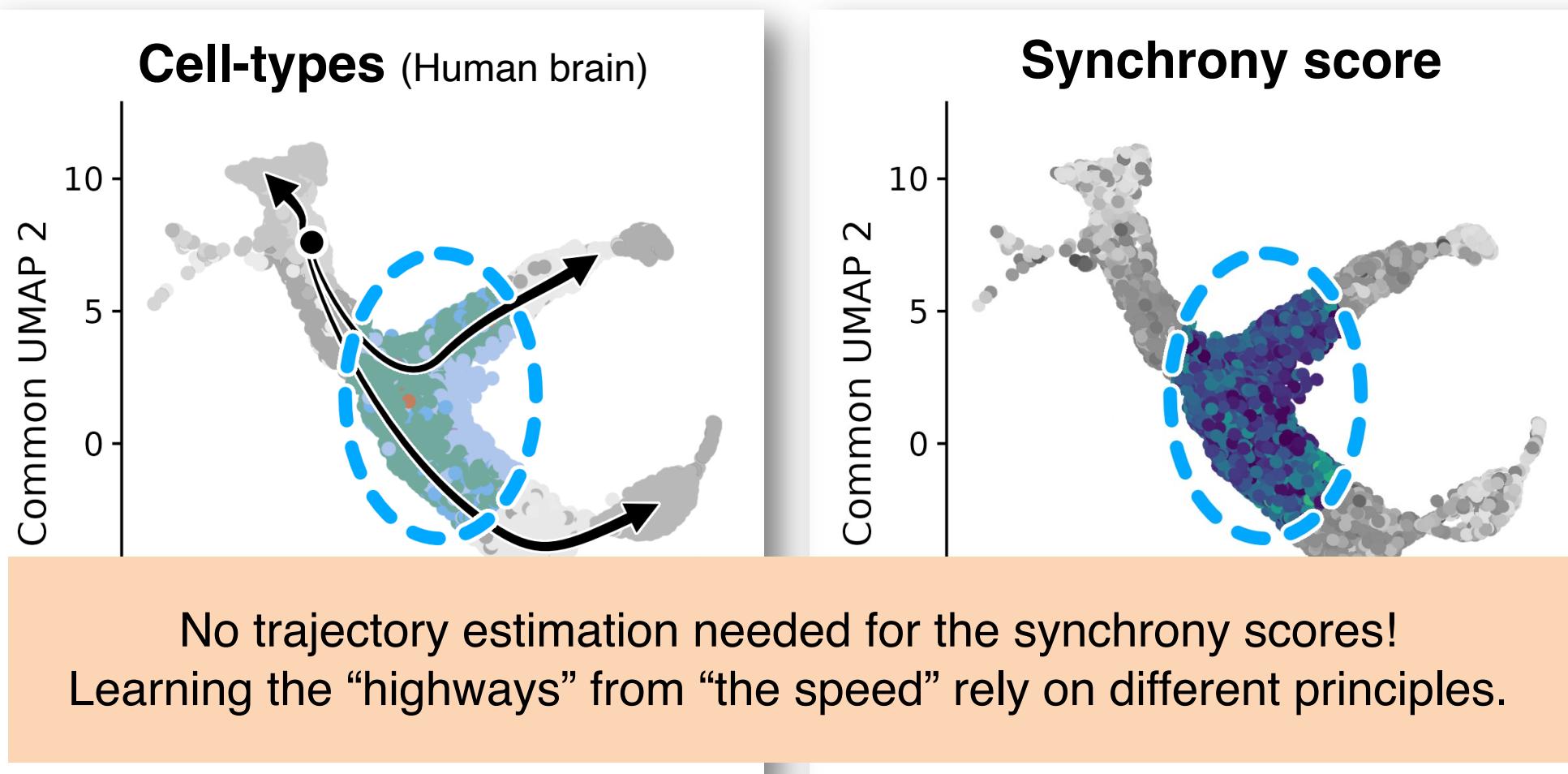


High synchrony score (i.e., steady-state)  
Low synchrony score (i.e., undergoing development)

A vertical color bar showing a gradient from dark purple at the bottom to bright yellow at the top, representing the range of synchrony scores. The text "High synchrony score (i.e., steady-state)" is positioned above the color bar, and "Low synchrony score (i.e., undergoing development)" is positioned below it.

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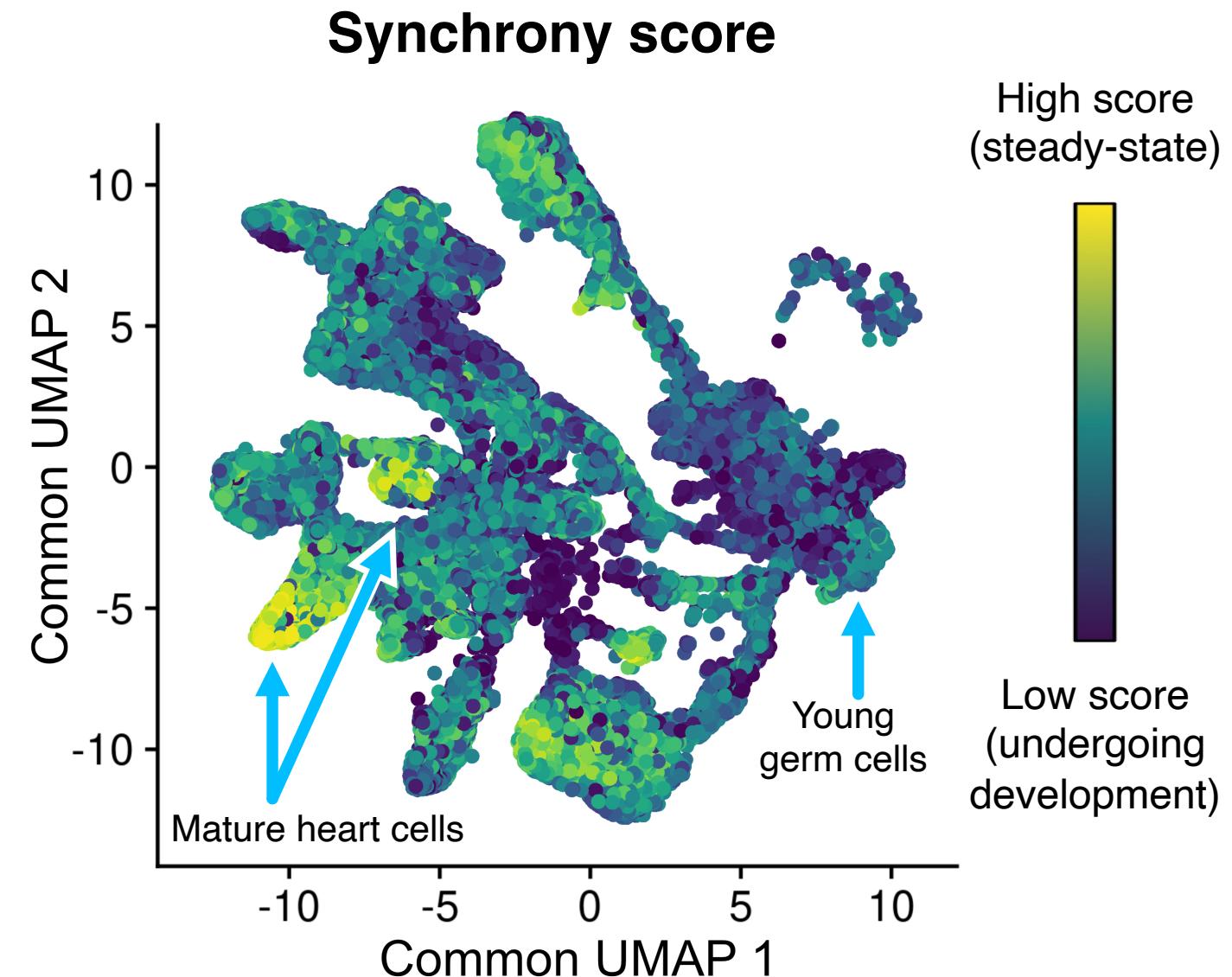
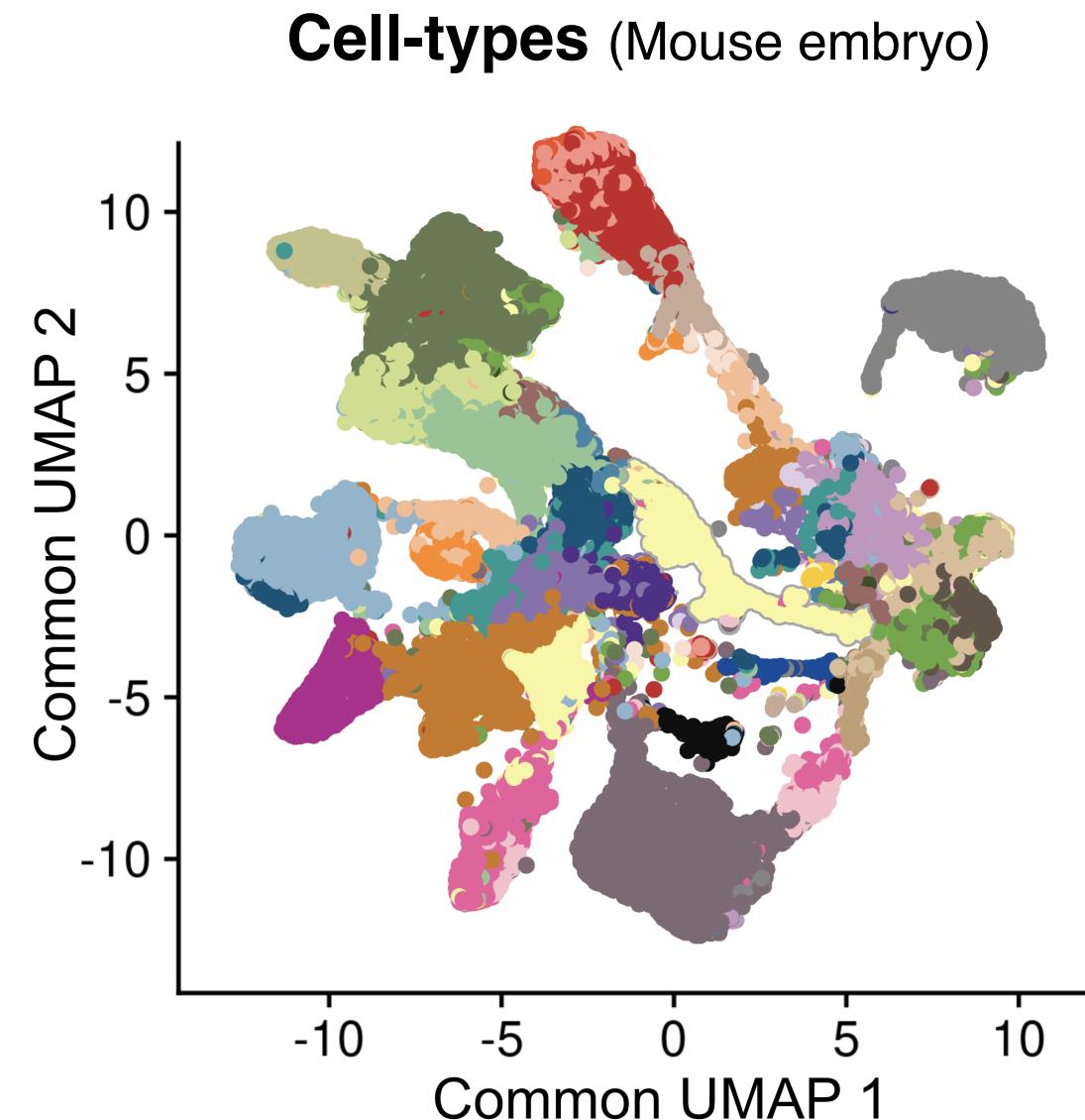
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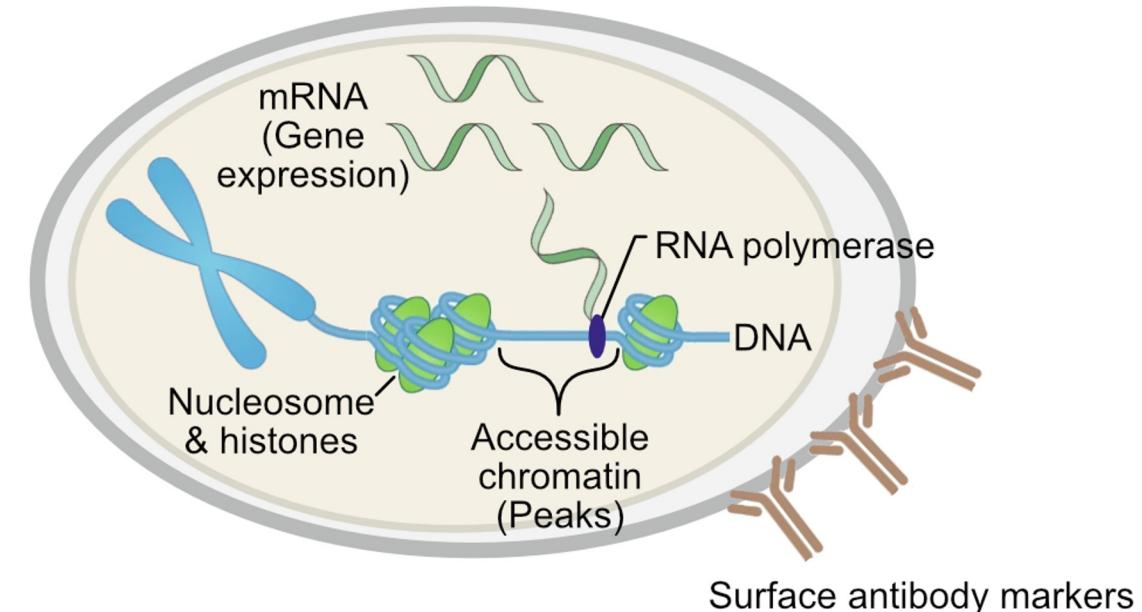
We demonstrate the utility of the synchrony scores on other harder systems.

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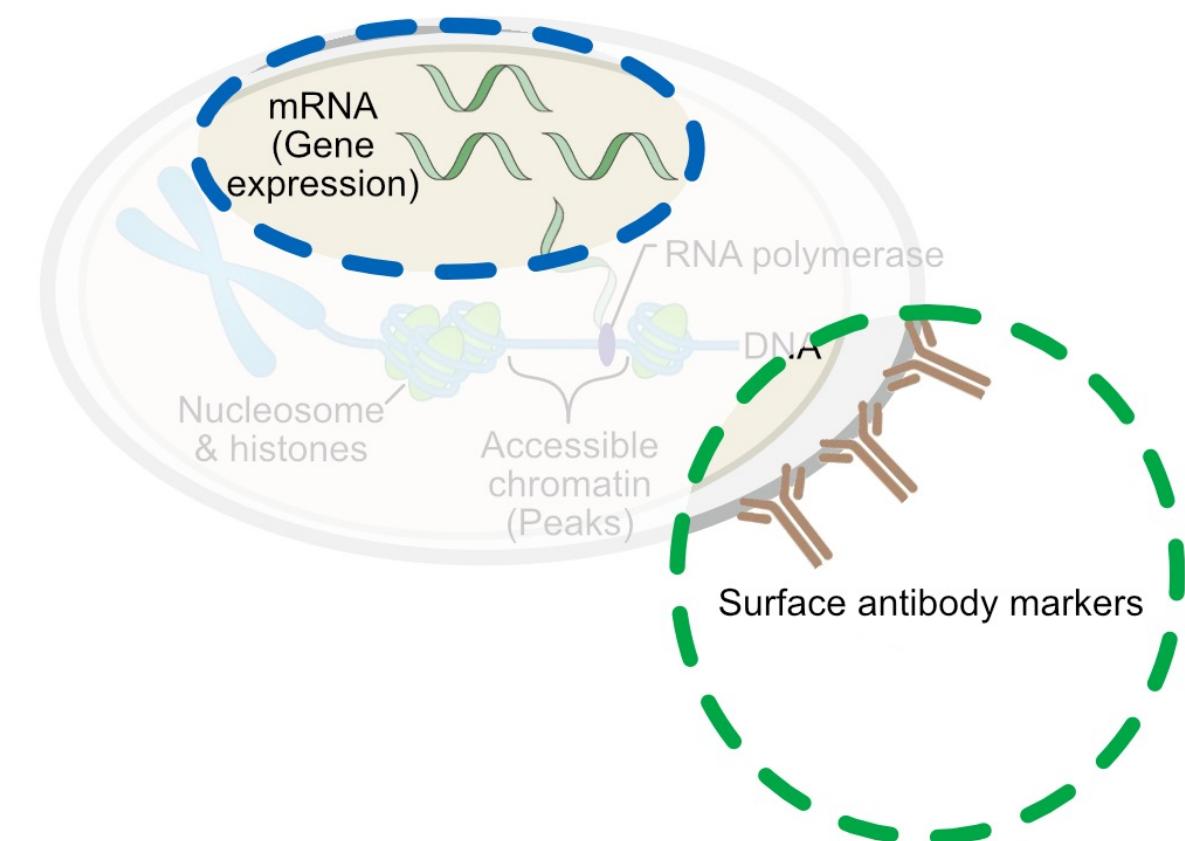
**Recap:** By matrix factorizing multi-modal data based on the common/distinct geometry, Tilted-CCA enables new perspectives to understand cell biology.

1. **(Experimental design):** Which pair of modalities should biologist sequence to have the most comprehensive understanding?
2. **(Variable selection):** For RNA-Protein data, how can we pick the antibodies that contribute the most additional information to the RNA modality?
3. **(Developmental biology):** Can the amount of coordination between two modalities tell us if a cell is in a steady-state or is undergoing development?



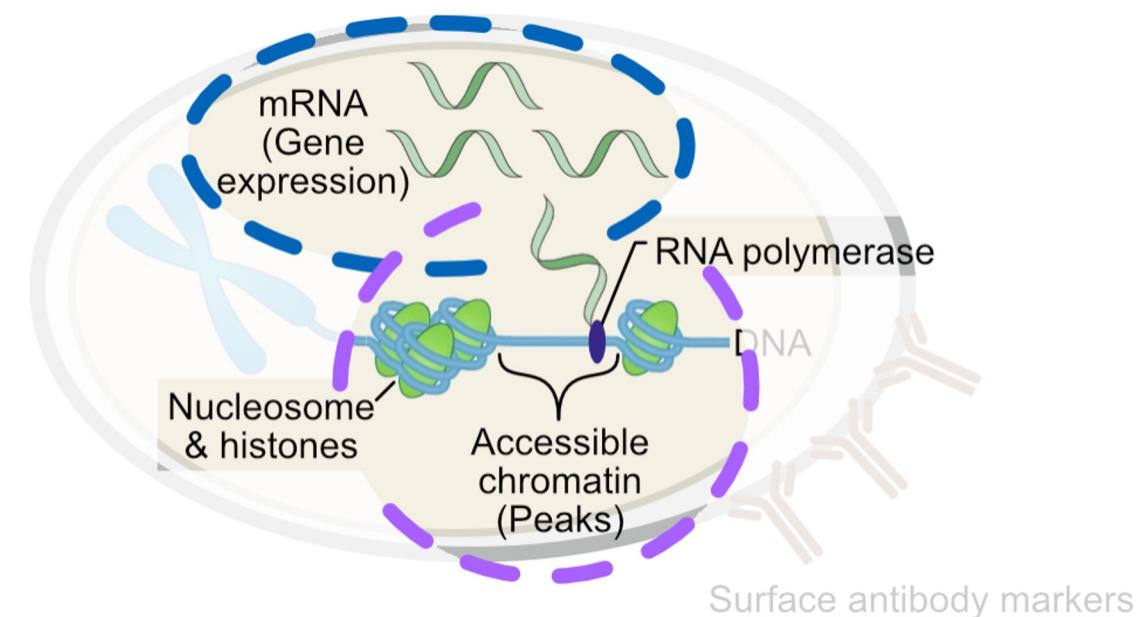
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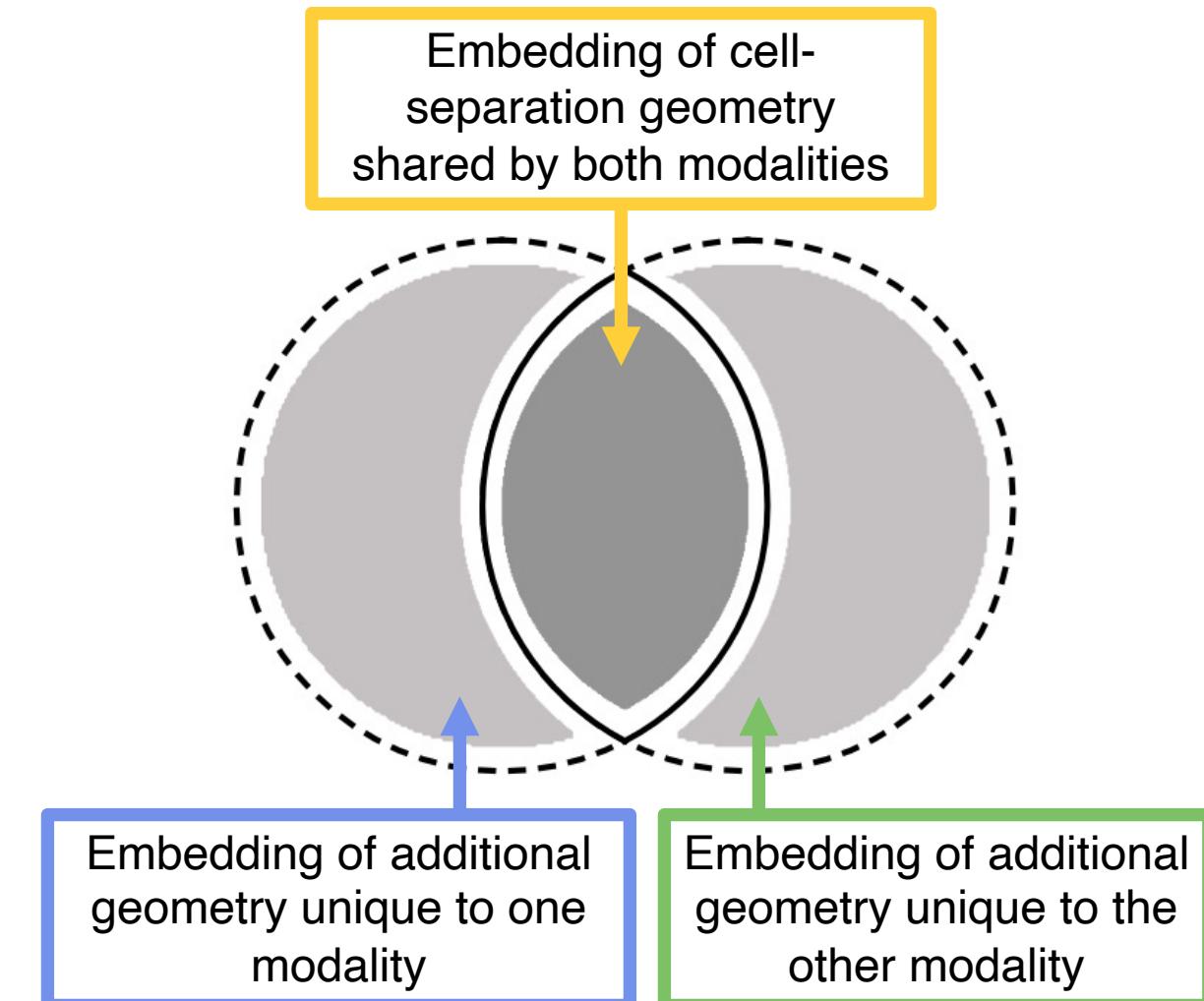
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Multi-modal analyses will only become more prevalent in biomedical research, and this will raise new biological theories that will require new matrix factorization approaches to study.

## Takeaways:

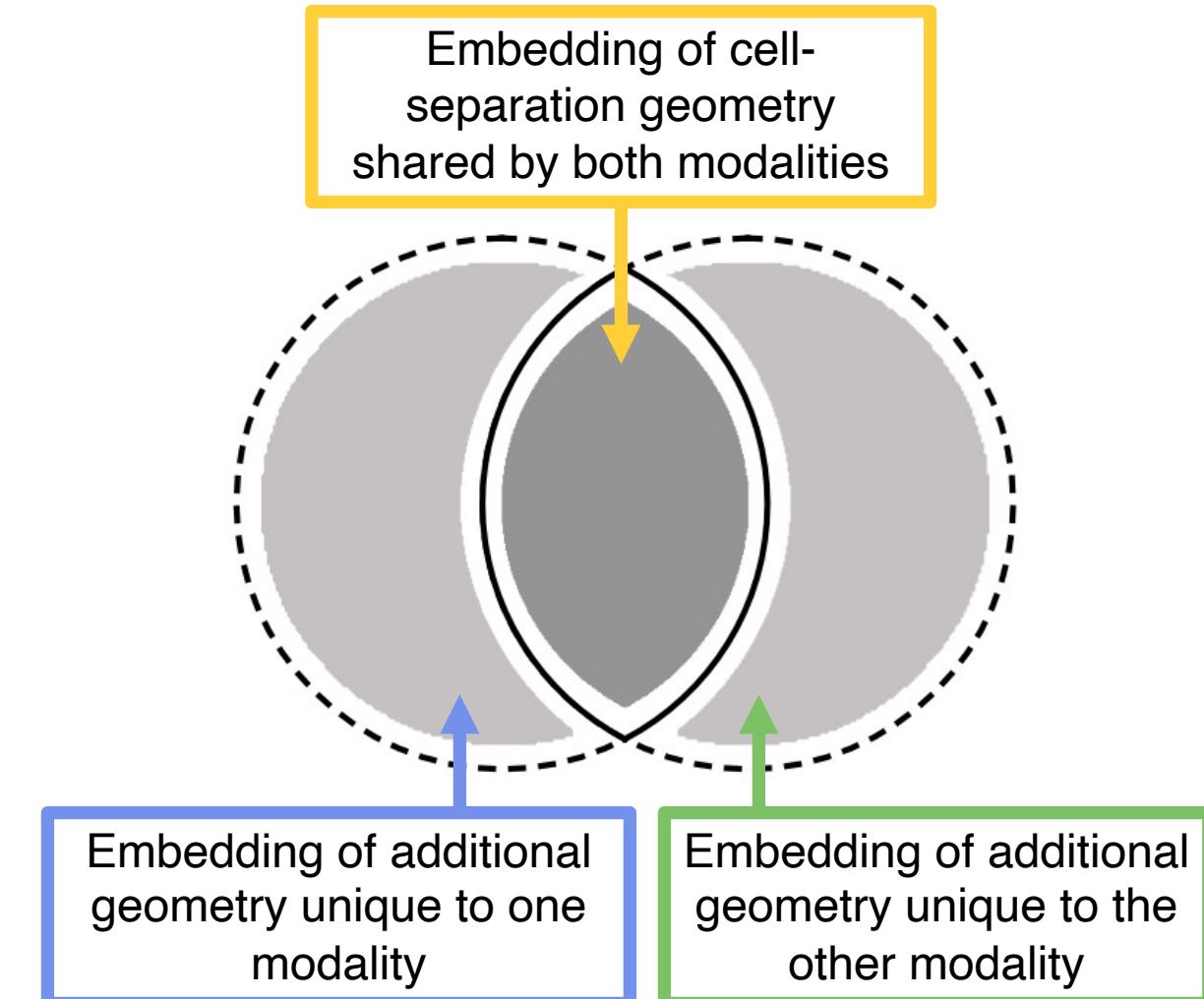
## Takeaways:

- **Novelty:** Estimating the “intersection of information” in multi-modal data, as opposed many existing methods for “union of information”
- **Strategy:** Matrix-factorization to estimate embeddings of the common and distinct geometries
- **Specifically:** Build upon CCA by “tilting” of the common vector to approximate the desired shared geometry



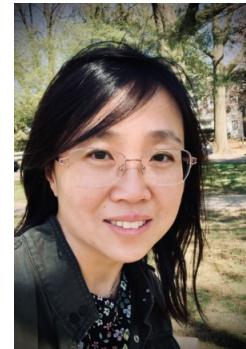
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- **Future theory:** What is the “population” shared geometry?
- **Future biology:** Cancer biology, where we need to understand the specifics of cellular mechanisms



# Thank you!

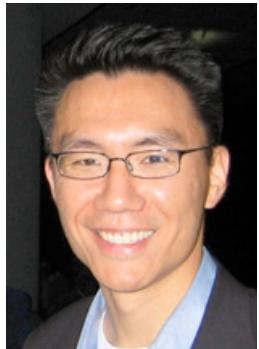
Tilted-CCA on Biorxiv: 2022.10.07.511320



Nancy  
Zhang



Sydney  
Shaffer



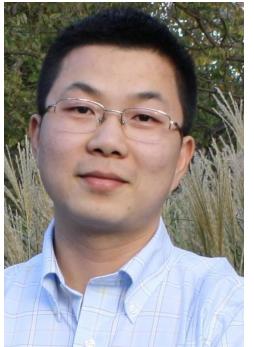
Andy  
Minn



E. John  
Wherry



Kathryn  
Roeder



Jing  
Lei



Ryan  
Tibshirani



Alessandro  
Rinaldo



Han  
Liu



Max  
G'Sell



James  
Sharpnack



Sangwon  
Hyun