Diabetes Dataset Testing

2023-09-24

Functions

```
count_na_per_column <- function(df) {</pre>
    sapply(df, function(x) sum(is.na(x)))
}
# Create a function to calculate metrics
calc_metrics <- function(pred, true) {</pre>
  confusion <- table(pred, true)</pre>
 TP <- confusion[2, 2]
 FP <- confusion[2, 1]
  TN <- confusion[1, 1]
  FN <- confusion[1, 2]
  Sensitivity <- TP / (TP + FN)
  Specificity <- TN / (TN + FP)
  Accuracy <- (TP + TN) / (TP + FP + TN + FN)
  Precision <- TP / (TP + FP)
  F1 <- 2 * (Precision * Sensitivity) / (Precision + Sensitivity)
  pred_obj <- prediction(as.numeric(pred), as.numeric(true))</pre>
  perf <- performance(pred_obj, "auc")</pre>
 AUC <- as.numeric(perf@y.values)
 return(c(AUC, Sensitivity, Precision, F1, Accuracy))
### A string like "Logistic" has to be put in. <--- from the colnames of df_coef
importance_plot <- function(model_string) {</pre>
# Create a data frame for plotting
coef_df <- data.frame(</pre>
  Variable = rownames(df_coef)[-1], # Exclude the intercept
  Importance = abs(df_coef[-1, model_string]) # Exclude the intercept
# Order the variables by importance
coef_df <- coef_df[order(coef_df$Importance), ]</pre>
ggplot(coef_df, aes(x = reorder(Variable, Importance), y = Importance, fill = Importance)) +
 geom bar(stat = "identity") +
  scale_fill_gradient(low = "lightblue2", high = "lightblue3") +
 theme minimal() +
 theme(axis.text.y = element_text(size = 12),
        axis.text.x = element text(size = 10),
        title = element_text(size = 15),
```

```
axis.title.y = element_blank(),
    axis.title.x = element_blank(),
    legend.position = "none") +
coord_flip() +
ggtitle("Variable Importance")
}
```

5. Data Processing

```
# Loading the data
data("PimaIndiansDiabetes2", package = "mlbench")
diabetes <- PimaIndiansDiabetes2
diabetes$diabetes <- as.factor(ifelse(diabetes$diabetes == "pos", 1, 0))
#diabetes$diabetes <- ifelse(diabetes$diabetes == "pos", 1, 0)
#kable(t(count_na_per_column(diabetes)))</pre>
```

Removing NA's

```
diabetes <- na.omit(diabetes)
#kable(t(count_na_per_column(diabetes)))</pre>
```

Removing Outliers

```
outliers <- check_outliers(diabetes, method = "mahalanobis")
#plot(outliers)
outliers <- as.vector(outliers)
#diabetes <- diabetes[!outliers, ]</pre>
```

Data for models

```
# Split
set.seed(222)
n <- nrow(diabetes)</pre>
training.samples \leftarrow sample(1:n, size = 0.75 * n)
train.data <- diabetes[training.samples, ]</pre>
scaled_train.data <- scale(train.data[, 1:8])</pre>
train.data[, 1:8] <- scaled_train.data</pre>
#train.data <- smote(diabetes ~ ., train.data, perc.over = 1)</pre>
test.data <- diabetes[-training.samples, ]</pre>
scaled_test.data <- scale(test.data[, 1:8])</pre>
test.data[, 1:8] <- scaled_test.data</pre>
#test.data <- na.omit(test.data)</pre>
# Handle NA's in training set
\#mice <- complete(mice(subset(train.data, select = -c(triceps, insulin)), method='rf', seed = 123))
#mice <- complete(mice(train.data, method='rf', seed = 123))</pre>
#train.data$glucose <- mice$glucose</pre>
```

```
#train.data$pressure <- mice$pressure</pre>
#train.data$mass <- mice$mass</pre>
#train.data <- na.omit(train.data)</pre>
#mice.triceps <- complete(mice(subset(train.data, select = -insulin), method='rf', seed = 123))</pre>
#train.data$triceps <- mice.triceps$triceps</pre>
#mice.insulin <- complete(mice(train.data, method='rf', seed = 123))</pre>
#train.data$insulin <- mice.insulin$insulin</pre>
X.train <- model.matrix(diabetes~., data = train.data)[,-1]</pre>
#X.train <- scale(X.train)</pre>
y.train <- train.data$diabetes
# Test
X.test <- model.matrix(diabetes ~., data = test.data)[,-1]</pre>
#X.test <- scale(X.test)</pre>
y.test <- test.data$diabetes</pre>
Logistic Regression
set.seed(123)
log.model <- glm(diabetes ~., data = train.data, family = "binomial")</pre>
# Make predictions
```

```
probabilities <- predict(log.model, newdata = test.data, type = "response")</pre>
predicted.classes.log <- as.factor(ifelse(probabilities > 0.5, 1, 0))
# Accuracy
# mean(predicted.classes == y.test)
log.conf <- confusionMatrix(predicted.classes.log, y.test, positive = "1")</pre>
log.conf
## Confusion Matrix and Statistics
##
##
             Reference
## Prediction 0 1
            0 61 10
##
##
            1 6 21
##
##
                  Accuracy: 0.8367
##
                    95% CI: (0.7484, 0.9037)
##
       No Information Rate: 0.6837
##
       P-Value [Acc > NIR] : 0.0004538
##
##
                     Kappa : 0.609
##
## Mcnemar's Test P-Value: 0.4532547
##
##
               Sensitivity: 0.6774
##
               Specificity: 0.9104
```

```
##
            Pos Pred Value: 0.7778
##
            Neg Pred Value: 0.8592
##
                Prevalence: 0.3163
##
           Detection Rate: 0.2143
##
      Detection Prevalence: 0.2755
##
         Balanced Accuracy: 0.7939
##
##
          'Positive' Class : 1
##
LASSO
set.seed(123)
cv.lasso.model <- cv.glmnet(X.train, y.train, alpha = 1, family = "binomial",
                            intercept = T)
#plot(cv.lasso.model)
cbind(coef(cv.lasso.model, s = cv.lasso.model$lambda.min), coef(cv.lasso.model, s = cv.lasso.model$lamb
## 9 x 2 sparse Matrix of class "dgCMatrix"
                       s1
## (Intercept) -0.9087548 -0.79367862
## pregnant
                0.2500270 0.04713706
## glucose
                1.0571306 0.80577102
## pressure
## triceps
                0.0626410
## insulin
## mass
                0.4184043 0.19426613
## pedigree
                0.3622318 0.10361089
## age
                0.2235214 0.17759602
# Make predictions
probabilities <- predict(cv.lasso.model, newx = X.test, s = cv.lasso.model$lambda.min, type = "response"
predicted.classes.lasso.min <- as.factor(ifelse(probabilities > 0.5, 1, 0))
probabilities <- predict(cv.lasso.model, newx = X.test, s = cv.lasso.model$lambda.1se, type = "response"
predicted.classes.lasso.1se <- as.factor(ifelse(probabilities > 0.5, 1, 0))
# Accuracy
#mean(predicted.classes.lasso.min == y.test)
lasso.min.conf <- confusionMatrix(predicted.classes.lasso.min, y.test, positive = "1")</pre>
lasso.min.conf
## Confusion Matrix and Statistics
##
##
            Reference
## Prediction 0 1
           0 61 10
##
            1 6 21
##
##
##
                  Accuracy : 0.8367
##
                    95% CI: (0.7484, 0.9037)
##
       No Information Rate: 0.6837
##
       P-Value [Acc > NIR] : 0.0004538
##
```

Kappa: 0.609

##

```
##
    Mcnemar's Test P-Value: 0.4532547
##
##
##
               Sensitivity: 0.6774
##
               Specificity: 0.9104
            Pos Pred Value: 0.7778
##
##
            Neg Pred Value: 0.8592
##
                Prevalence: 0.3163
##
            Detection Rate: 0.2143
##
      Detection Prevalence: 0.2755
##
         Balanced Accuracy: 0.7939
##
##
          'Positive' Class: 1
##
lasso.1se.conf <- confusionMatrix(predicted.classes.lasso.1se, y.test, positive = "1")
lasso.1se.conf
  Confusion Matrix and Statistics
##
##
             Reference
##
  Prediction 0 1
            0 62 12
##
            1 5 19
##
##
##
                  Accuracy: 0.8265
##
                    95% CI: (0.7369, 0.8956)
       No Information Rate: 0.6837
##
       P-Value [Acc > NIR] : 0.001064
##
##
##
                     Kappa: 0.573
##
    Mcnemar's Test P-Value: 0.145610
##
##
##
               Sensitivity: 0.6129
##
               Specificity: 0.9254
            Pos Pred Value: 0.7917
##
##
            Neg Pred Value: 0.8378
##
                Prevalence: 0.3163
            Detection Rate: 0.1939
##
##
      Detection Prevalence: 0.2449
##
         Balanced Accuracy: 0.7691
##
##
          'Positive' Class: 1
##
in data: set.seed(123) and train 0.75 all others see(123) i like it a lot. min and 1se very sparse and good
```

in data: set.seed(123) and train 0.75 all others see(123) i like it a lot. min and 1se very sparse and good pred, just not much difference in amount of sparsity and amount of FN

in data: set.seed(2) and train 0.75 all others see(123) i like it a lot. 1se sparse and more FN but same pred

in data: set.seed(222) and train 0.75 all others see(123) pretty good similar too above

in data: set.seed(6) and train 0.75 all others see(123) gives very sparse 1se lasso and okay pred

in data: set.seed(13) and train 0.75 all others see(123) pretty nice! sparse 1se, but pred is same

in data: set.seed(42) and train 0.75 all others see(123) pretty nice! very very sparse 1se, more FN in 1se

some things we see:

• the more conservative model (1se) usually has more FN —> it makes the safe/conservative choice of going for the class that has 2/3 of observations

Ridge

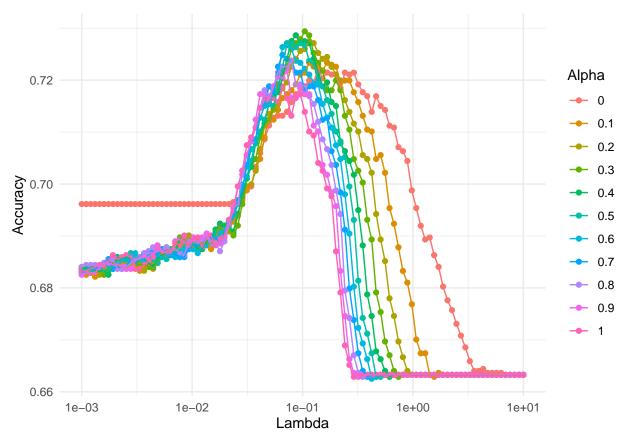
```
set.seed(123)
cv.ridge.model <- cv.glmnet(X.train, y.train, alpha = 0, family = "binomial", intercept = T)</pre>
#plot(cv.ridge)
# Make predictions
probabilities <- predict(cv.ridge.model, newx = X.test, s = cv.ridge.model$lambda.min, type = "response"
predicted.classes.ridge <- as.factor(ifelse(probabilities > 0.5, 1, 0))
# Accuracy
\#mean(predicted.classes.ridge == y.test)
ridge.conf <- confusionMatrix(as.factor(predicted.classes.ridge), y.test, positive = "1")
ridge.conf
## Confusion Matrix and Statistics
##
##
             Reference
## Prediction 0 1
##
            0 62 10
            1 5 21
##
##
##
                  Accuracy : 0.8469
                    95% CI: (0.7601, 0.9117)
##
##
       No Information Rate: 0.6837
##
       P-Value [Acc > NIR] : 0.0001806
##
##
                     Kappa : 0.6301
##
   Mcnemar's Test P-Value: 0.3016996
##
##
               Sensitivity: 0.6774
##
##
               Specificity: 0.9254
##
            Pos Pred Value: 0.8077
##
            Neg Pred Value: 0.8611
##
                Prevalence: 0.3163
            Detection Rate: 0.2143
##
##
      Detection Prevalence: 0.2653
         Balanced Accuracy: 0.8014
##
##
          'Positive' Class: 1
##
##
```

Elastic Net

```
set.seed(123)
# CV and tuning grid
```

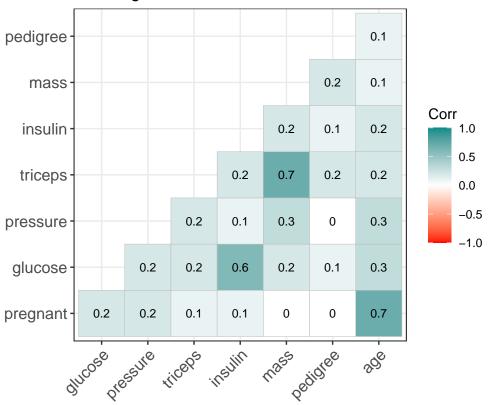
```
myFolds <- createFolds(y.train, k = 10, list = TRUE)</pre>
myControl <- trainControl(index = myFolds)</pre>
#cvControl <- trainControl(method = "cv", number = 10)
tuneGrid <- expand.grid(alpha = seq(0, 1, by = 0.05), lambda = 10^seq(1, -3, length=100))
# Train model
elasticnet.model <- train(X.train, y.train, method = "glmnet", trControl = myControl,
                          tuneGrid = tuneGrid, intercept = T, family = "binomial")
# Make predictions
probabilities <- predict(elasticnet.model, newdata = X.test, type = "prob") # "raw" already outputs pre
predicted.classes.elasticnet <- ifelse(probabilities 1 > 0.5, 1, 0)
# Accuracy
# mean(predicted.classes == y.test)
elasticnet.conf <- confusionMatrix(as.factor(predicted.classes.elasticnet), y.test, positive = "1")
elasticnet.conf
## Confusion Matrix and Statistics
##
            Reference
##
## Prediction 0 1
##
           0 63 11
##
            1 4 20
##
##
                  Accuracy : 0.8469
                    95% CI: (0.7601, 0.9117)
##
##
       No Information Rate: 0.6837
       P-Value [Acc > NIR] : 0.0001806
##
##
##
                     Kappa: 0.6233
##
  Mcnemar's Test P-Value: 0.1213353
##
##
               Sensitivity: 0.6452
##
##
               Specificity: 0.9403
##
            Pos Pred Value: 0.8333
##
            Neg Pred Value: 0.8514
                Prevalence: 0.3163
##
##
           Detection Rate: 0.2041
##
     Detection Prevalence: 0.2449
##
         Balanced Accuracy: 0.7927
##
          'Positive' Class : 1
##
##
# Output coefficients
coef(elasticnet.model$finalModel, s = elasticnet.model$bestTune$lambda)
## 9 x 1 sparse Matrix of class "dgCMatrix"
##
                         s1
## (Intercept) -0.783644314
## pregnant
               0.124519132
## glucose
                0.616288277
```

```
## pressure
                0.003625147
## triceps
                0.073458775
## insulin
                0.067361284
## mass
                0.200698201
## pedigree
                0.158043114
## age
                0.188599082
# Filter data to only include specific alpha values
filtered_data <- subset(elasticnet.model$results, alpha %in% seq(0, 1, by=0.1))
# Create the plot
ggplot(filtered_data, aes(x=lambda, y=Accuracy, color=factor(alpha))) +
  geom_point() +
 geom_line() +
  scale_x_log10() +
  scale_color_discrete(name = "Alpha") +
  labs(x = "Lambda", y = "Accuracy") +
 theme_minimal()
```



```
colors = c("red", "white", "cyan4"),
title = "Correlogram of Diabetes Dataset",
ggtheme = theme_bw())
```

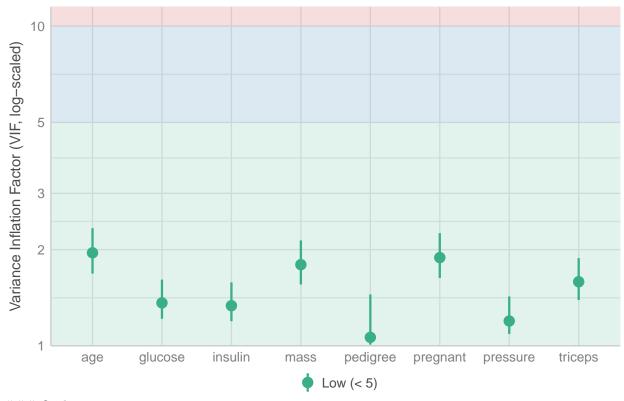
Correlogram of Diabetes Dataset



```
result <- check_collinearity(log.model)
plot(result) + ggtitle("")</pre>
```

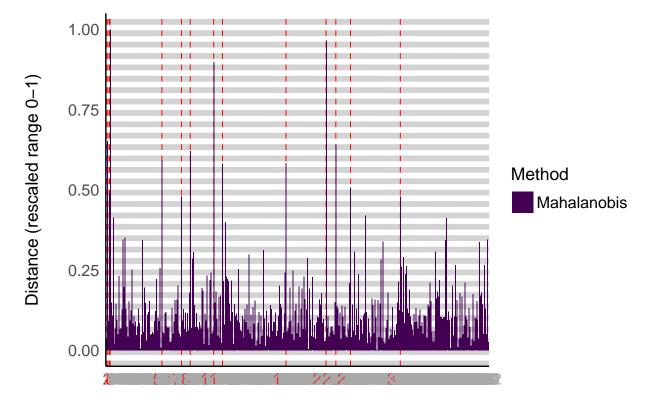
Variable 'Component' is not in your data frame :/

High collinearity (VIF) may inflate parameter uncertainty



Outliers

```
result <- check_outliers(diabetes, method = "mahalanobis")
plot(result, type = "dots")</pre>
```



Observation

```
result
```

```
## 13 outliers detected: cases 2, 4, 5, 58, 78, 87, 111, 120, 185, 226,
     236, 251, 302.
## - Based on the following method and threshold: mahalanobis (30).
```

- For variables: pregnant, glucose, pressure, triceps, insulin, mass,

pedigree, age.

as.vector(result)

```
[1] FALSE TRUE FALSE TRUE FALSE FALS
##
                                                                         [13] FALSE F
                                                                         [25] FALSE F
##
                                                                         [37] FALSE F
##
                                                                         [49] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
##
                                                                         [61] FALSE F
                                                                         [73] FALSE FALSE FALSE FALSE
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               TRUE FALSE FALSE FALSE FALSE FALSE
##
##
                                                                         [85] FALSE FALSE
                                                                                                                                                                                                                                                                                                                                                                                     TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
                                                                         [97] FALSE F
                                                  [109] FALSE FALSE
                                                                                                                                                                                                                                                                                                                                                                                                 TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
                                                       [121] FALSE 
                                                     [133] FALSE 
                                                  [145] FALSE 
                                                  [157] FALSE FALSE
                                                     [169] FALSE 
                                                  [181] FALSE 
## [193] FALSE FALSE
```

```
## [205] FALSE FAL
## [217] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE TRUE FALSE FALSE
## [229] FALSE FALSE FALSE FALSE FALSE FALSE TRUE FALSE FALSE FALSE FALSE
## [241] FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [253] FALSE FALSE
## [265] FALSE FALSE
## [277] FALSE FALSE
## [289] FALSE FALSE
## [301] FALSE TRUE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE FALSE
## [313] FALSE FALSE
## [325] FALSE FALSE
## [337] FALSE FALSE
## [349] FALSE FALSE
## [361] FALSE FALSE
## [373] FALSE FALSE
## [385] FALSE FALSE FALSE FALSE FALSE FALSE FALSE
```

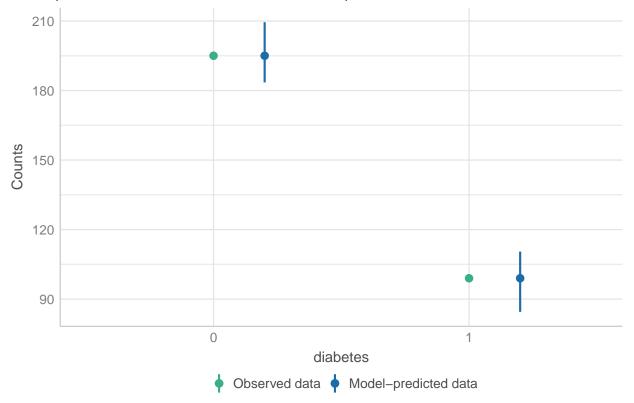
Check model

```
check <- check_model(log.model, panel = F)
plot(check)</pre>
```

\$PP_CHECK

Posterior Predictive Check

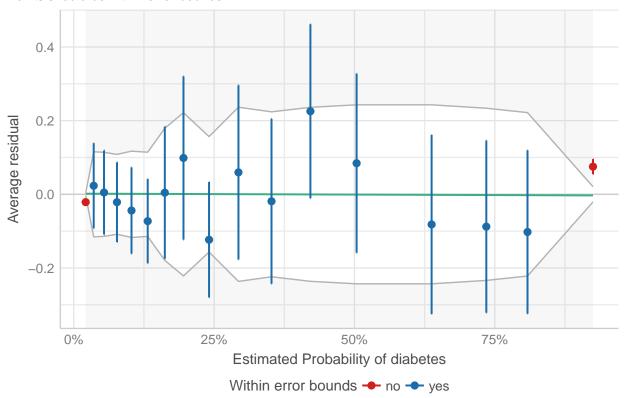
Model-predicted intervals should include observed data points



##
\$BINNED_RESID

Binned Residuals

Points should be within error bounds

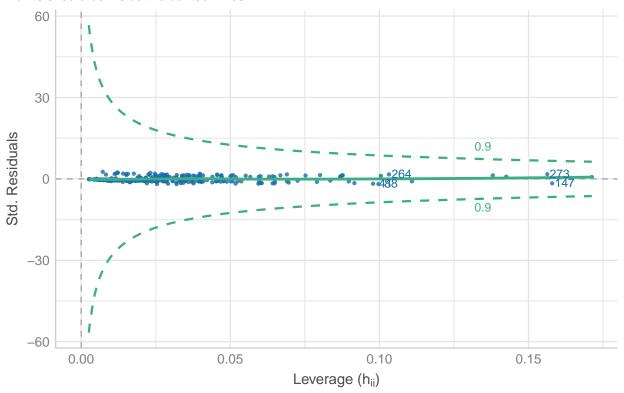


##

\$OUTLIERS

Influential Observations

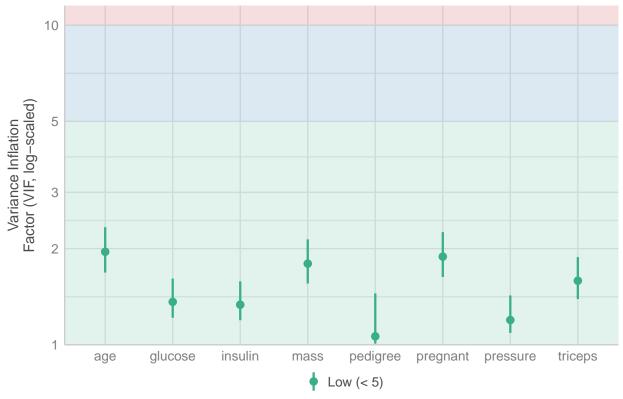
Points should be inside the contour lines



\$VIF

Collinearity

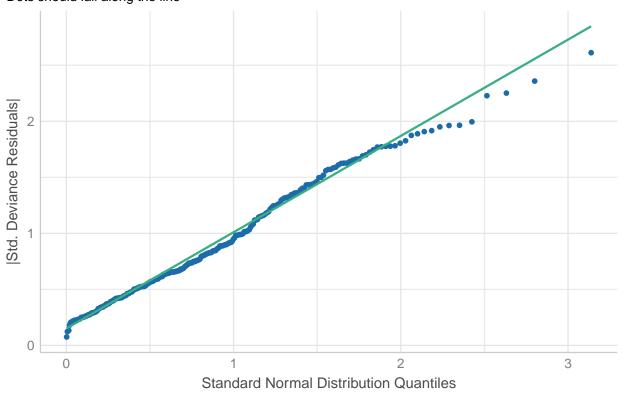
High collinearity (VIF) may inflate parameter uncertainty



##

\$QQ

Normality of Residuals Dots should fall along the line



binned residuals(log.model)

Warning: About 88% of the residuals are inside the error bounds (~95% or higher would be good).

Calculate model metrics

```
# Calculate metrics for each model
metrics.log <- calc_metrics(predicted.classes.log, y.test)</pre>
metrics.lasso.min <- calc_metrics(predicted.classes.lasso.min, y.test)</pre>
metrics.lasso.1se <- calc_metrics(predicted.classes.lasso.1se, y.test)</pre>
metrics.ridge <- calc_metrics(predicted.classes.ridge, y.test)</pre>
metrics.elasticnet <- calc_metrics(predicted.classes.elasticnet, y.test)</pre>
# Store in a dataframe
df_metrics <- as.data.frame(matrix(c(metrics.log, metrics.lasso.min, metrics.lasso.1se, metrics.ridge,
colnames(df_metrics) <- c("Logistic", "Lasso_min", "Lasso_1se", "Ridge", "ElasticNet")</pre>
rownames(df_metrics) <- c("AUC", "Recall", "Precision", "F1", "Accuracy")</pre>
df_metrics
##
              Logistic Lasso_min Lasso_1se
                                                 Ridge ElasticNet
## AUC
             0.7939336 0.7939336 0.7691382 0.8013962 0.7927299
## Recall
             0.6774194 0.6774194 0.6129032 0.6774194 0.6451613
## Precision 0.7777778 0.7777778 0.7916667 0.8076923 0.8333333
## F1
             0.7241379\ 0.7241379\ 0.6909091\ 0.7368421\ 0.7272727
## Accuracy 0.8367347 0.8367347 0.8265306 0.8469388 0.8469388
```

Latex Table Metrics

	Models				
	Logistic	Lasso λ_{\min}	Lasso λ_{1se}	Ridge	ElasticNet
Recall	0.677	0.677	0.613	0.677	0.645
Precision	0.778	0.778	0.792	0.808	0.833
$\mathbf{F1}$	0.724	0.724	0.691	0.737	0.727
Accuracy	0.837	0.837	0.827	0.847	0.847

Data Frame of all model coefficients

pregnant

0.348

0.25

```
coef.log <- coef(log.model)</pre>
coef.lasso.min <- (coef(cv.lasso.model, s = cv.lasso.model$lambda.min))</pre>
coef.lasso.1se <- coef(cv.lasso.model, s = cv.lasso.model$lambda.1se)</pre>
coef.ridge <- coef(cv.ridge.model)</pre>
coef.elasticnet <- coef(elasticnet.model$finalModel, s = elasticnet.model$bestTune$lambda)</pre>
# Create a data frame to store coefficients
df coef <- data.frame(</pre>
 Logistic = as.vector(coef.log),
 Lasso min = as.vector(as.matrix(coef.lasso.min)),
 Lasso_1se = as.vector(as.matrix(coef.lasso.1se)),
 Ridge = as.vector(as.matrix(coef.ridge)),
  ElasticNet = as.vector(as.matrix(coef.elasticnet))
rownames(df_coef) <- names(coef.log)</pre>
options(scipen = 999)
df_coef_sparse <- round(df_coef, 3)</pre>
df_coef_sparse[] <- apply(df_coef_sparse, 2, function(x) ifelse(x == 0, '.', x))</pre>
df_coef_sparse
               Logistic Lasso_min Lasso_1se Ridge ElasticNet
## (Intercept) -0.976
                            -0.909
                                    -0.794 -0.777
```

0.125

0.047 0.157

```
0.806 0.437
## glucose
                 1.197
                           1.057
                                                      0.616
## pressure
                -0.008
                                         . 0.084
                                                      0.004
## triceps
                 0.099
                           0.063
                                         . 0.121
                                                      0.073
## insulin
                                                      0.067
                -0.011
                                          0.157
## mass
                 0.527
                           0.418
                                     0.194 0.181
                                                      0.201
                 0.489
                           0.362
                                     0.104 0.177
                                                      0.158
## pedigree
## age
                  0.24
                           0.224
                                     0.178 0.186
                                                      0.189
```

Latex Table Coef

	Models				
	Logistic	Lasso λ_{\min}	Lasso λ_{1se}	Ridge	ElasticNet
(Intercept)	-0.976	-0.909	-0.794	-0.777	-0.784
pregnant	0.348	0.25	0.047	0.157	0.125
glucose	1.197	1.057	0.806	0.437	0.616
pressure	-0.008	•	•	0.084	0.004
${f triceps}$	0.099	0.063		0.121	0.073
insulin	-0.011		•	0.157	0.067
mass	0.527	0.418	0.194	0.181	0.201
$\mathbf{pedigree}$	0.489	0.362	0.104	0.177	0.158
age	0.24	0.224	0.178	0.186	0.189

Confusion matrix latex - log.model

```
# Convert the table to a data frame
log.conf.df <- as.data.frame(log.conf$table)

# Replace the numeric labels with text labels
log.conf.df$Prediction <- factor(log.conf.df$Prediction, levels = c("0", "1"), labels = c("Negative", "log.conf.df$Reference <- factor(log.conf.df$Reference, levels = c("0", "1"), labels = c("Negative", "Po")
# Rename columns as required by cums
names(log.conf.df) <- c('Target', 'Prediction', 'N')
# Plot the confusion matrix using cums
plot_confusion_matrix(log.conf.df, add_normalized = FALSE,</pre>
```

add_row_percentages = FALSE, add_col_percentages = FALSE, rotate_y_text = TRUE,

```
place_x_axis_above = TRUE, class_order = c("Positive", "Negative"))

## Warning in plot_confusion_matrix(log.conf.df, add_normalized = FALSE,
## add_row_percentages = FALSE, : 'ggimage' is missing. Will not plot arrows and
## zero-shading.

## Warning in plot_confusion_matrix(log.conf.df, add_normalized = FALSE,
## add_row_percentages = FALSE, : 'rsvg' is missing. Will not plot arrows and
## zero-shading.
```



Confusion matrix latex - lasso.min.model

```
# For lasso.min.conf
lasso.min.conf df <- as.data.frame(lasso.min.conf$table)
lasso.min.conf.df$Prediction <- factor(lasso.min.conf.df$Prediction, levels = c("0", "1"), labels = c("lasso.min.conf.df$Reference, levels = c("0", "1"), labels = c("Nenames(lasso.min.conf.df) <- c('Target','Prediction','N')
plot_confusion_matrix(lasso.min.conf.df, add_normalized = FALSE, add_row_percentages = FALSE, add_col_p

## Warning in plot_confusion_matrix(lasso.min.conf.df, add_normalized = FALSE, :
## 'ggimage' is missing. Will not plot arrows and zero-shading.

## Warning in plot_confusion_matrix(lasso.min.conf.df, add_normalized = FALSE, :
## 'rsvg' is missing. Will not plot arrows and zero-shading.</pre>
```



Confusion matrix latex - lasso.1se.model

```
lasso.1se.conf.df <- as.data.frame(lasso.1se.conf$table)
lasso.1se.conf.df$Prediction <- factor(lasso.1se.conf.df$Prediction, levels = c("0", "1"), labels = c("lasso.1se.conf.df$Reference, levels = c("0", "1"), labels = c("Negrames(lasso.1se.conf.df) <- c('Target','Prediction','N')
plot_confusion_matrix(lasso.1se.conf.df, add_normalized = FALSE, add_row_percentages = FALSE, add_col_p

## Warning in plot_confusion_matrix(lasso.1se.conf.df, add_normalized = FALSE, :
## 'ggimage' is missing. Will not plot arrows and zero-shading.

## Warning in plot_confusion_matrix(lasso.1se.conf.df, add_normalized = FALSE, :
## 'rsvg' is missing. Will not plot arrows and zero-shading.</pre>
```



Confusion matrix latex - ridge.model

```
# For ridge.conf
ridge.conf.df <- as.data.frame(ridge.conf$table)
ridge.conf.df$Prediction <- factor(ridge.conf.df$Prediction, levels = c("0", "1"), labels = c("Negative ridge.conf.df$Reference <- factor(ridge.conf.df$Reference, levels = c("0", "1"), labels = c("Negative", names(ridge.conf.df) <- c('Target', 'Prediction', 'N')
plot_confusion_matrix(ridge.conf.df, add_normalized = FALSE, add_row_percentages = FALSE, add_col_percentages = FALSE, add_col_percentages = ridge.
## Warning in plot_confusion_matrix(ridge.conf.df, add_normalized = FALSE, :
## 'ggimage' is missing. Will not plot arrows and zero-shading.
## Warning in plot_confusion_matrix(ridge.conf.df, add_normalized = FALSE, :
## 'rsvg' is missing. Will not plot arrows and zero-shading.</pre>
```



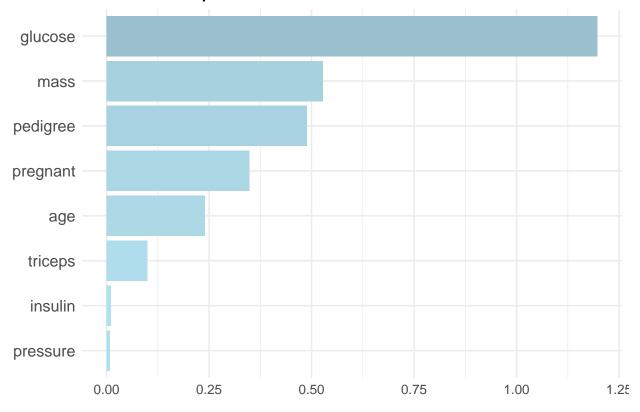
Confusion matrix latex - elasticnet.model

```
# For elasticnet.conf
elasticnet.conf.df <- as.data.frame(elasticnet.conf$table)
elasticnet.conf.df$Prediction <- factor(elasticnet.conf.df$Prediction, levels = c("0", "1"), labels = c
elasticnet.conf.df$Reference <- factor(elasticnet.conf.df$Reference, levels = c("0", "1"), labels = c("names(elasticnet.conf.df) <- c('Target','Prediction','N')
plot_confusion_matrix(elasticnet.conf.df, add_normalized = FALSE, add_row_percentages = FALSE, add_col_normalized = FALSE, add_row_percentages = FALSE, add_row_percentag
```

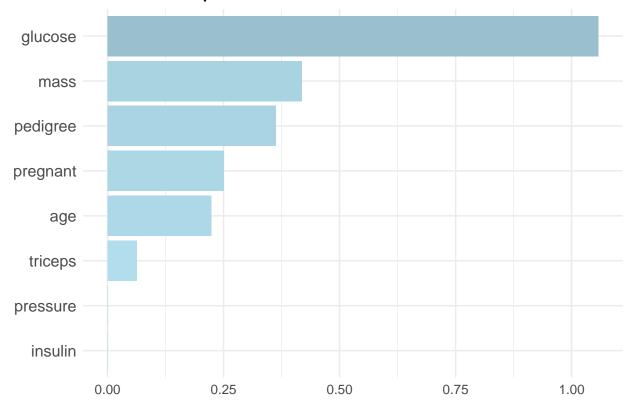


Variable Importance not working great because font size

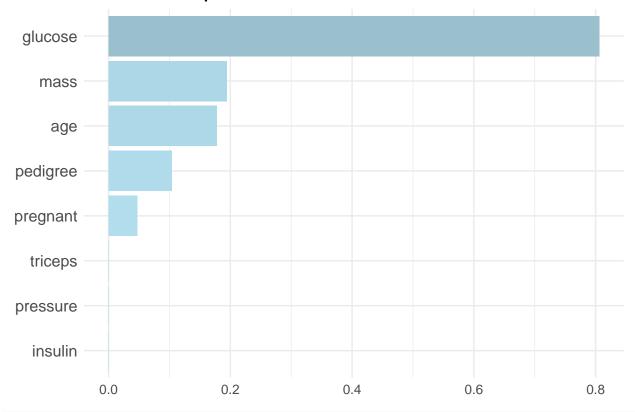
```
#png(filename = "varimp_log.png", width = 3000, height = 3000)
importance_plot("Logistic")
```



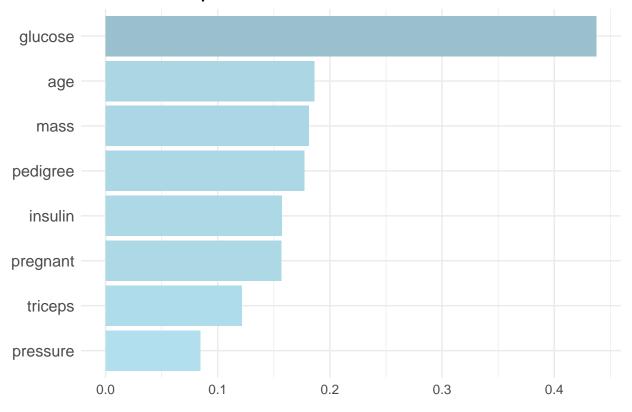
```
#dev.off()
#png(filename = "varimp_lasso_min.png", width = 3000, height = 3000)
importance_plot("Lasso_min")
```



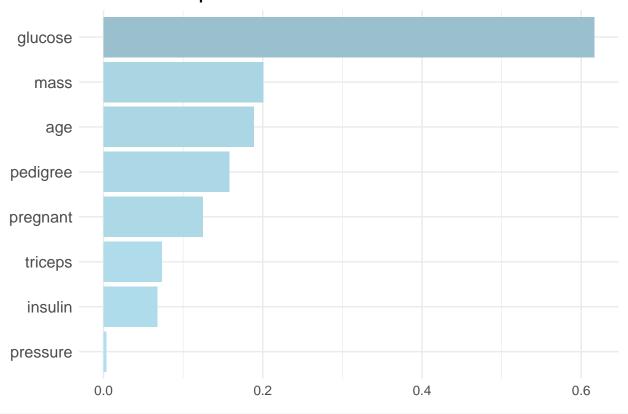
```
#dev.off()
#png(filename = "varimp_lasso_1se.png", width = 3000, height = 3000)
importance_plot("Lasso_1se")
```



```
#dev.off()
#png(filename = "varimp_ridge.png", width = 3000, height = 3000)
importance_plot("Ridge")
```



#dev.off()
#png(filename = "varimp_elasticnet.png", width = 3000, height = 3000)
importance_plot("ElasticNet")



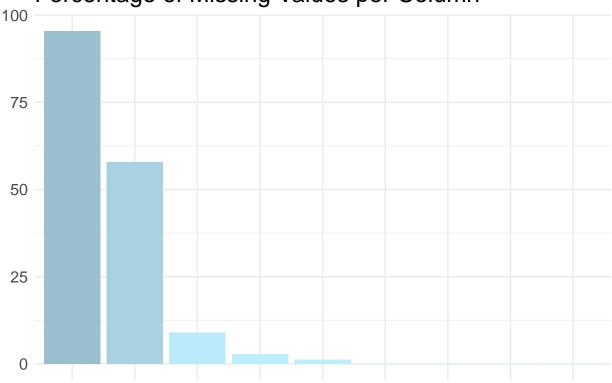
#dev.off()

Perc missing

```
library(ggplot2)
# Count of missing values per column
count_na <- c(pregnant = 0, glucose = 5, pressure = 35, triceps = 227, insulin = 374, mass = 11, pedigr</pre>
# Total number of rows in the data
n_rows <- nrow(diabetes)</pre>
# Convert to data frame and calculate percentages
df_na <- data.frame(Column = names(count_na), Count = as.numeric(count_na))</pre>
df_na$Percentage <- (df_na$Count / n_rows) * 100</pre>
# Plot
ggplot(df_na, aes(x = reorder(Column, -Percentage), y = Percentage, fill = Percentage)) +
  geom_bar(stat = "identity") +
  scale_fill_gradient(low = "lightblue1", high = "lightblue3") +
 theme_minimal() +
 theme(
   axis.text.y = element_text(size = 12),
    axis.text.x = element_text(size = 12),
   title = element_text(size = 15),
   axis.title.y = element_blank(),
    axis.title.x = element_blank(),
```

```
legend.position = "none"
) +
ggtitle("Percentage of Missing Values per Column")
```

Percentage of Missing Values per Column



insulin triceps pressure mass glucose age diabetes pedigreepregnant

log model summary table latex

```
# Extract coefficients from the model summary
coef summary <- summary(log.model)$coefficients</pre>
# Convert the matrix to a data frame for kable()
coef_summary <- round(as.data.frame(coef_summary)[, c(1, 4)], 3)</pre>
colnames(coef_summary) <- c("Estimate", "p-value")</pre>
# coef_summary$`p-value` <- cell_spec(coef_summary$`p-value`, "latex",
                                    bold = ifelse(coef_summary$`p-value` < 0.05, TRUE, FALSE))</pre>
# # Create the LaTeX table
# kable(coef_summary, "latex", booktabs = TRUE, align = "c", escape = F) %>%
   kable_styling(latex_options = c("striped", "scale_down", "hold_position")) %>%
    row spec(0, bold = TRUE)
# Add stars based on significance level and make significant p-values bold
coef_summary$`p-value` <- sapply(coef_summary$`p-value`, function(p) {</pre>
  stars <- ifelse(p < 0.001, "***", ifelse(p < 0.01, "**", ifelse(p < 0.05, "*", "")))
  value_with_stars <- paste0(round(p, 3), stars)</pre>
  cell_spec(value_with_stars, "latex", bold = p < 0.05)</pre>
})
```

```
# Create the LaTeX table
kable(coef_summary, "latex", booktabs = TRUE, align = "c", escape = F) %>%
kable_styling(latex_options = c("striped", "scale_down", "hold_position")) %>%
row_spec(0, bold = TRUE)
```

	Estimate	p-value
(Intercept)	-0.976	0***
pregnant	0.348	0.087
glucose	1.197	0***
pressure	-0.008	0.963
triceps	0.099	0.622
insulin	-0.011	0.95
mass	0.527	0.016*
pedigree	0.489	0.004**
age	0.240	0.251

coef latex per model - lasso both

	Estimate	p value
(Intercept)	-0.976	0***
pregnant	0.348	0.087
glucose	1.197	0***
pressure	-0.008	0.963
${f triceps}$	0.099	0.622
insulin	-0.011	0.95
mass	0.527	0.016*
pedigree	0.489	0.004**
age	0.240	0.251

```
kable_styling(latex_options = c("striped", "scale_down", "hold_position"), position = "center") %>%
row_spec(0, bold = TRUE) %>%
column_spec(1, bold = TRUE) %>%
add_header_above(c(" " = 1, "Estimates" = 2), bold = TRUE)
print(coef_latex)
```

coef latex per model - log, ridge

	Estimates		
	Lasso λ_{\min}	Lasso λ_{1se}	
(Intercept)	-0.909	-0.794	
pregnant	0.25	0.047	
${f glucose}$	1.057	0.806	
pressure	•	•	
${f triceps}$	0.063	•	
insulin	•	•	
mass	0.418	0.194	
${f pedigree}$	0.362	0.104	
age	0.224	0.178	

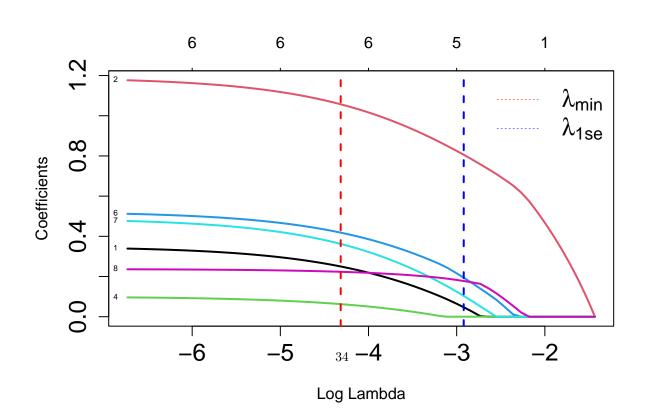
coef latex per model - log, elastic net

```
coef_latex <- df_coef_sparse[, c(1,5)] %>%
  kable(format = "latex", booktabs = TRUE, align = c('c'), escape = FALSE,
        col.names = c("Logistic", "ElasticNet")) %>%
  kable_styling(latex_options = c("striped", "scale_down", "hold_position"), position = "center") %>%
  row_spec(0, bold = TRUE) %>%
  column_spec(1, bold = TRUE) %>%
  add_header_above(c(" " = 1, "Estimates" = 2), bold = TRUE)
print(coef_latex)
# Plot the coefficient paths with thicker lines
plot(cv.lasso.model$glmnet.fit, xvar = "lambda", label = TRUE, cex.axis = 1.5, lwd = 2)
# Add vertical lines for lambda.min and lambda.1se with thicker lines
abline(v = log(cv.lasso.model$lambda.min), col = "red", lty = 2, lwd = 2)
abline(v = log(cv.lasso.model$lambda.1se), col = "blue", lty = 2, lwd = 2)
# Add a legend with LaTeX-style text and increased font size, and thicker lines
legend("topright",
      legend = expression(lambda["min"], lambda["1se"]),
```

	Estimates		
	Logistic	Ridge	
(Intercept)	-0.976	-0.777	
pregnant	0.348	0.157	
glucose	1.197	0.437	
pressure	-0.008	0.084	
${f triceps}$	0.099	0.121	
insulin	-0.011	0.157	
mass	0.527	0.181	
$\mathbf{pedigree}$	0.489	0.177	
age	0.24	0.186	

```
col = c("red", "blue"),
lty = 2,
cex = 1.5,
lwd = 0.5, bty = "n")
```

	Estimates		
	Logistic	ElasticNet	
(Intercept)	-0.976	-0.784	
pregnant	0.348	0.125	
${f glucose}$	1.197	0.616	
pressure	-0.008	0.004	
${f triceps}$	0.099	0.073	
insulin	-0.011	0.067	
mass	0.527	0.201	
${f pedigree}$	0.489	0.158	
age	0.24	0.189	



```
p value Acc > NIR
# Number of trials
n <- 98
# Number of successes (correctly classified instances)
\# Accuracy is 0.8367, so the number of successes would be 0.8367 * 392
n_success \leftarrow round(0.8367 * n)
# No Information Rate
nir < -0.6837
# Perform binomial test
binom_test_result <- binom.test(n_success, n, p = nir, alternative = "greater")</pre>
# Print the result
print(binom_test_result)
##
## Exact binomial test
##
## data: n_success and n
## number of successes = 82, number of trials = 98, p-value = 0.0004546
## alternative hypothesis: true probability of success is greater than 0.6837
## 95 percent confidence interval:
## 0.7626479 1.0000000
## sample estimates:
## probability of success
                0.8367347
log.conf$overall["Accuracy"]
Accuracy 0.8367347
log.conf$overall["AccuracyPValue"]
AccuracyPValue 0.0004537699
tibble(Accuracy = log.conf$overall["Accuracy"],
       `p-value [Acc > NIR]` = log.conf$overall["AccuracyPValue"])
```

A tibble: 1×2

Accuracy p-value [Acc > NIR]

0.8367

0.0005

Accuracy p-value [Acc > NIR]

0.8367

0.0005

Accuracy p-value [Acc > NIR]

0.8469

0.0002

Accuracy p-value [Acc > NIR]

0.8367

0.0005

Accuracy p-value [Acc > NIR]

0.8265

0.0011

Accuracy	p-value $[Acc > NIR]$
0.8469	0.0002