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# A MULTIDIMENSIONAL ANALYSIS OF COMPRESSIBLE AND INCOMPRESSIBLE FLOW FLUID DYNAMICS PROTEIN SEPARATION BY POLYACRYLAMIDE GEL ELECTROPHORENSIS



by
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Brian Steven Brown

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A dissertation submitted to the faculty of The University of North Carolina at Charlotte in partial fulfillment of the requirements for the degree of Doctor of Philosophy in Applied Mathematics

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Charlotte
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2015

Approved by:	
Dr. Jane David	
Dr. Frost Kane	
Dr. Janice Dickey	
Dr. Randle Elle	
Dr. Renee Zack	



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# ISOLATION AND CHARACTERIZATION OF *TRITICUM AESTIVUM* ROOT cDNA SEQUENCES WHICH SHOW SIGNIFICANT HOMOLOGY TO THE GRAMNEGATIVE BACTERIAL TRANSPOSON TN*1721*

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by
1 single space
Jennifer Renee Brown

4 single spaces

A thesis submitted to the faculty of The University of North Carolina at Charlotte in partial fulfillment of the requirements for the degree of Master of Science in Biology

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Charlotte
1 single space
2015

Approved by:	
Dr. Jane David	
Dr. Frost Kane	
Dr. Janice Dickey	
Dr. Randle Elle	
Dr. Renee Zack	

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### **ABSTRACT**

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JENNIFER RENEE BALL. Isolation and characterization of *Triticum aestivum* root cDNA sequences which show significant homology to the gram-negative bacterial transposon Tn1721. (Under the direction of DR. HELEN W. JOHNSON)

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While previously screening a bread wheat (*Triticum aestivum* cv. Pavon) pollen embryo cDNA library, four clones were identified that showed high homology to the bacterial transposon Tn1721. Finding these clones led to studies to ascertain whether the bacterial sequence was actually present in the wheat genome or was simply an artifact of the cloning procedure. Using the transposon as a probe, a root cDNA library was screened. Two putative clones were isolated from this library and sequenced in both directions. Consensus sequences were used to search the nucleotide databases for homologies. This search revealed that both clones were highly homologous to the *tetR* gene of Tn1721. Northern hybridization using RNA isolated from roots confirmed that the Tn1721 sequence was present in genes expressed by this tissue. These results support the hypothesis that regions of the Tn1721 transposon are integrated into the wheat chromosomal DNA and that this may be an example of an interkingdom horizontal transfer of DNA from a bacterium to a plant.



# **ACKNOWLEDGMENTS**

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The important element in the Acknowledgments is simple courtesy in which there are usually two possible ingredients to consider. First you should acknowledge any significant help you received from any individual whether in your department or elsewhere. Specifically, you should acknowledge the source of special materials, documents, or equipment. Further, you should acknowledge the help of anyone who contributed significantly to the work or to the interpretation of the work. Second, you should acknowledge any outside source of financial assistance, such as grants, contracts, or fellowships. A word of caution is in order. Often it is wise to show the proposed wording of the Acknowledgments to the person whose help you are acknowledging. He or she might well believe that your acknowledgment is insufficient or (worse) that it is too effusive.

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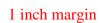
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# LIST OF ABBREVIATIONS

2 single spaces, start the next line on the 3<sup>rd</sup>

ABA abscisic acid

ABRE abscisic acid response element

ANOVA analysis of variance

BHT butylated hydroxytoluene

cDNA complementary DNA

cpm counts per minute

DS dextran sulfate

dATP deoxyATP

ddATP dideoxyATP

2,4-D 2,4-dichlorophenoxyacetic acid

EcMt early cysteine-labeled metallothionein

KN Kinetin

mAb monoclonal antibody

mRNA messenger RNA

#### **CHAPTER 1: INTRODUCTION**

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Transposons are genetic elements that are mobile within a genome, therefore, they are one of the major causes of genomic variation (Lewin 1994). However, evidence is growing that transposable elements are capable of horizontal transfer. That is, they can move across genomes of different species (Prins and Zadocks 1992). Our laboratory is interested in the identification and characterization of bacterial transposon-like nucleotide sequences found in the wheat genome that may be an example of horizontal DNA transfer.

Reynolds and Kitto (1992) screened a Mexican spring wheat (*Triticum aestivum* cv.Pavon) cDNA library to identify genes expressed specifically during pollen embryogenesis. After sequencing unique clones from this library, four sequences were found that showed high homology to the bacterial transposon, Tn1721. This transposon was derived from a gram-negative bacterium and is a Tn3-like transposon found in the Tn21 subgroup (Grinstead et al. 1990). It is a unique sequence since it contains a basic transposon (Tn1722) that is capable of independent transpososition. As shown in Figure 1, Tn1722 contains an open reading frame that encodes a 525 amino acid chemotaxic protein (Allmeier et al. 1992). The Tn1722 portion of the transposon contains the *tnpR* and *tnpA* genes which are utilized during the genetic resolution and integration of either the major or minor sequences. The entire transposable element also include three inverted repeats which function as the insertion and excisions sites for the transposon.

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## APPENDIX A: PCR OPTIMIZATION

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The optimization of the PCR reactions were conducted on the bacterial plasmid pJOE105, which contains the entire Tn1721 transposon. This optimization required three independent experiments to determine the best parameters for each set of primers: MgCl<sub>2</sub> concentration, target DNA concentration, and thermal cycling parameters.

Magnesium provides the divalent cations required by the DNA polymerase to function. The MgCl<sub>2</sub> concentrations were optimized by titration reactions ranging from 1.55 mM to 3.55 mM final concentration in each reaction tube.

The concentration of target DNA was optimized to ensure the highest possible primer specificity. DNA was diluted serially for each reaction to determine the lowest concentration of polynucleotide that still yielded visible bands on EtBr-stained agarose gels; for pJOE105 this was # 1 ng of DNA.

PCR cycle parameters were examined to reduce the so called plateau effect which results in the non-specific amplification of background products. Taking this into account, cycling parameters were set to allow efficient amplification with the lowest number of cycles. Conditions were set at 33 cycles of 1 min. at 94 C for denaturation, 1 min. at 56.5 C for annealing, and 2 min. at 72 C for synthesis, followed by 10 min. at 72 C for extension.