Permutation Tests

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1 Introduction

1.1 Introduction

- Well established nonparametric approach to inference: Fisher, 1935; Pitman, 1937; Pitman, 1938.
- (In general) it requires less assumptions about the data generating process than the parametric counterpart.
- Very good inferential properties, typically:
 - exactness (i.e. exact control of the type I error)
 - asymptotically optimality and convergence to the parametric counterpart when it does exist.
- Fisher exact test is a prototypical example, but
- the general approach has restricted applicability without the support of a computer.

1.2 Renewed interest toward permutation testing

- A milestone: Westfall and Young (1993). Resampling-Based Multiple Testing: Examples and Methods for p-value Adjustment. Wiley.
- Many actives areas of research adopt these methods in their daily statistical analysis (e.g. genetics and neuroscience: Nichols and Holmes (2002); Pantazis et al. (2009); Winkler et al. (2014)).
- Permutation approach:
 - Ideal for randomized experimental design
 - deals with very complex models, without formal definition of the data generating process.

1.3 The package flip

It is on CRAN and on github (https://github.com/livioivil/flip)

To install the github version type (in R):

```
library(devtools)
install_github('livioivil/flip')
```

Before we start

```
#clean the memory
rm (list=ls ())

# We customize the output of our graphs a little bit
par.old=par ()
par (cex.main=1.5, lwd=2, col="darkgrey", pch=20, cex=3)
# par (par.old)
palette (c ("#FF0000", "#00A08A", "#FFCC00", "#445577", "#45abff"))

# customize the output of knitr
knitr :: opts_chunk$set (fig.align="center")#, fig.width=6, fig.height=6)
```

1.4 The Age vs Reaction Time Dataset

The reaction time of these subjects was tested by having them grab a meter stick after it was released by the tester. The number of centimeters that the meter stick dropped before being caught is a direct measure of the person's response time.

The values of Age are in years. The Gender is coded as F for female and M for male. The values of Reaction. Time are in centimeters.

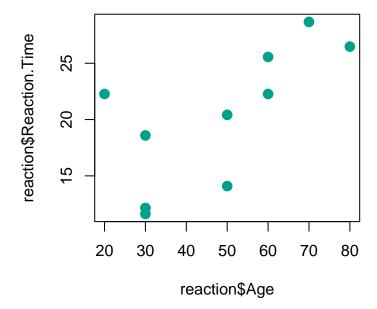
(data are fictitious)

To read the data

```
data(reaction,package = "flip")
# or download it from: https://github.com/livioivil/flip/tree/master/data
# str (reaction)
```

We plot the data

plot(x=reaction\$Age,y=reaction\$Reaction.Time,pch=20,col=2,cex=2)



1.5 Measuring the dependence between two variables

we define:

- X = Age
- Y = Reaction.Time

We review some famous index to measure the (linear) dependence among two variables

1.5.1 Covariance and Variance

Covariance between X and Y:

$$\sigma_{xy} = \frac{\sum_{i=1}^{n} (x_i - \bar{x})(y_i - \bar{y})}{n}$$

• values between $-\infty$ and ∞

- $\sigma_{xy} \approx 0$: there is no dependency between X and Y
- $\sigma_{xy} >> (<<)0$: there is a strong positive (negative) dependency between X and Y

Variance of X

$$\sigma_{xx} = \sigma_x^2 = \frac{\sum_{i=1}^n (x_i - \bar{x})^2}{n}$$

Standard Deviation of X:

$$\sigma_{xx} = \sqrt{\sigma_{xx}} = \sigma_x$$

1.5.2 Correlation

With the Covariance it is difficult to understand when the relationship between X and Y is strong/weak. We note that

$$-\sigma_x \sigma_y \leq \sigma_{xy} \leq \sigma_x \sigma_y$$
 is quivalent to $-1 \leq \frac{\sigma_{xy}}{\sigma_x \sigma_y} \leq 1$

Correlation between X and Y:

$$\rho_{xy} = \frac{\sigma xy}{\sigma_x \sigma_y} = \frac{\sum_{i=1}^{n} (x_i - \bar{x})(y_i - \bar{y})}{\sqrt{\sum_{i=1}^{n} (x_i - \bar{x})^2} \sqrt{\sum_{i=1}^{n} (y_i - \bar{y})^2}}$$

- values between -1 and 1
- $\rho_{xy} \approx 0$: there is no dependency between X and Y
- $\rho_{xy} \approx 1(-1)$: there is a strong positive (negative) dependency between X and Y

1.5.3 Linear Trend, the least squares method

We describe the relationship between

Reaction. Time and Age with a straight line.

$$E(Reaction.Time) \approx \beta_0 + \beta_1 Age$$

 $E(Y) = \beta_0 + \beta_1 X$

Let's draw a line 'in the middle' of the data.

The least-squares estimator

We look for the one that passes more 'in the middle', the one that minimizes the sum of the squares of the residues:

$$\hat{\beta}_0$$
 and $\hat{\beta}_1$ such that $\sum_{i=1}^n (y_i - (\hat{\beta}_0 + \hat{\beta}_1 x_i))^2$ is minimum.

Estimates:

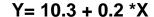
- Angular coefficient: $\hat{\beta}_1 = \frac{\sigma_{xy}}{\sigma_{xx}} = \rho_{xy} \frac{\sigma_y}{\sigma_x} = \frac{\sum_{i=1}^n (x_i \bar{x})(y_i \bar{y})}{\sum_{i=1}^n (x_i \bar{x})^2} = 0.2064719$
- Intercept: $\hat{\beta}_0 = \bar{y} \hat{\beta}_1 \bar{x} = 10.3013483$
- Response (estimated y): $\hat{y}_i = \hat{\beta}_0 + \hat{\beta}_1 x_i$
- Residuals (from the estimated response): $y_i (\hat{\beta}_0 + \hat{\beta}_1 x_i) = y_i \hat{y}_i$

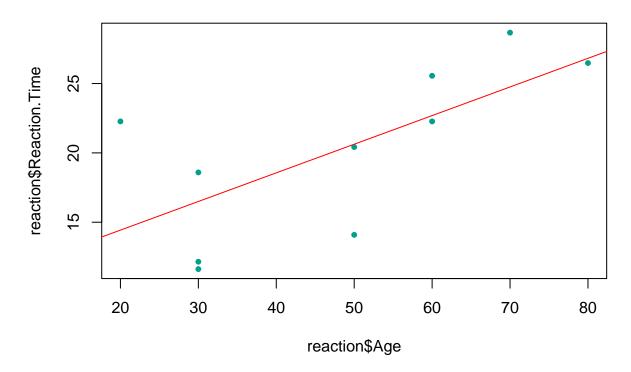
and therefore the least squares are the sum of the squared residuals: $\sum_{i=1}^{n}(y_i-\hat{\beta}_0+\hat{\beta}_1x_i)^2=\sum_{i=1}^{n}(y_i-\hat{y}_i)^2$

A graphical representation:

(Intercept) Age ## 10.3013483 0.2064719

```
plot(reaction$Age,reaction$Reaction.Time,pch=20,col=2,cex=1)
coeff=round(coefficients(model),1)
title(paste("Y=",coeff[1],"+",coeff[2],"*X"))
abline(model,col=1)
```





2 Permutation approach to Hypothesis Testing

2.0.1 Some remarks

Let's note that all the measures above does not make any assumptions on the random process that generate them.

Let now assume that Y - and possibly X - is generated by a random variable.

Further minimal assumptions will be specified later.

The question: Is there a relationship between Y and X?

We estimated $\hat{\beta}_1 = 0.2064719$

But the **true value** β_1 is really different from 0 (i.e. no relationship)? Otherwise, is the difference from 0 due to the random sampling?

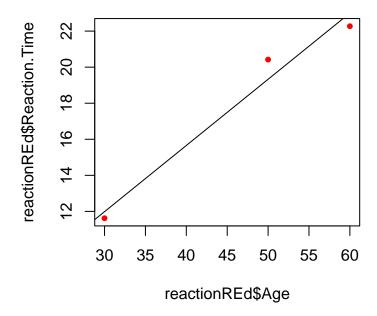
- Null Hypothesis H_0 : $\beta_1 = 0$ (the true β_1 , not its estimate $\hat{\beta}_1$!). There is no relationship between X and Y.
- Alternative Hypothesis $H_1: \beta_1 > 0$ The relationship is positive.

Other possible specifications of H_1 : $\beta_1 < 0$ and, more commonly, H_1 : $\beta_1 \neq 0$.

2.1 Permutation tests - in a nutshell

As a toy example, let use a sub-set of the data:

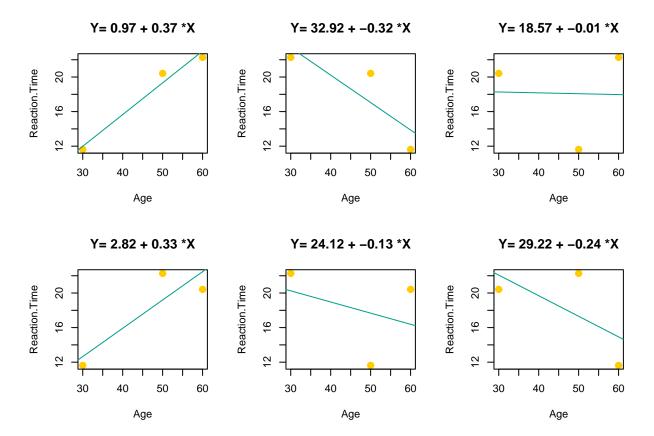
##		Age	Gender	Reaction.Time
##	2	50	F	20.42
##	3	30	M	11.62
##	4	60	F	22.27



- If H_0 is true: there is no linear relationship between X and Y
- Therefore, the trend observed on the data is due to chance.
- Any other match of x_i and y_i was equally likely to occur
- I can generate the datasets of other hypothetical experiments by exchanging the order of the observations in Y.
- How many equally likely datasets could I get with X and Y observed? 3*2*1=3!=6 possible datasets.

Remark: Here we only assume that y is a random variable. The only assumption here is the exchangeability of the observations: the joint density $f(y_1, \ldots, y_n)$ does not change when the ordering of y_1, \ldots, y_n is changed.

2.1.1 All potential datasets



2.1.1.1 In our data set We apply the same principle to the complete dataset...

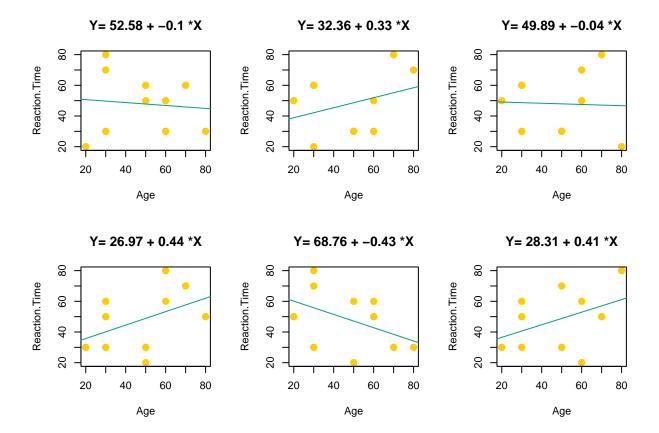
How many permutations of the vector y_1, \ldots, y_n are possible? n! = 10! = 3628800.

big, perhaps not too big . . . but what happen with, for example, n = 20? We got 20! = 2.432902e + 18. This is too big, definitely!

We calculate a smaller (but sufficiently large) B of random permutations.

here some example

Age vs a permutations of Reaction.Time



We repeat 10^4 times and we look at the histogram of the $\hat{\beta}_1$

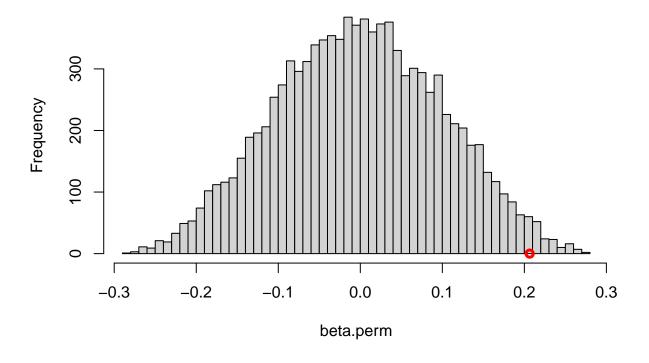
```
# beta_1 estimated on the observed data:
beta1=coefficients(lm(Reaction.Time~Age,data=reaction))[2]

# function that permutes the y values and calculates the coeff beta_1

my.beta.perm <- function(Y,X){
   model=lm(sample(Y)~X)
   coefficients(model)[2]
}

#replicate it B-1 times
beta.perm= replicate(B,my.beta.perm(reaction$Reaction.Time, reaction$Age ))</pre>
```

Histogram of beta.perm



2.1.2 How likely WAS $\hat{\beta}_1^{obs}$?

(before the experiment!)

How likely was it to get a $\leq \hat{\beta}_1^{obs}$ value among the many possible values of $\hat{\beta}_1^{*b}$ (obtained by permuting data)? Remarks:

- $\hat{\beta}_1^{*b} < \hat{\beta}_1^{obs}$ (closer to 0): less evidence against H_1 than $\hat{\beta}_1^{obs}$ $\hat{\beta}_1^{*b} \ge \hat{\beta}_1^{obs}$: equal or more evidence towards H_1 than $\hat{\beta}_1^{obs}$

2.1.3Calculation of the p-value

Over B=10⁴ permutations we got 9845 times a $\hat{\beta}_1^{*b} \leq \hat{\beta}_1^{obs}$.

The p-value (significance) is $p = \frac{\#(\hat{\beta}_1^{*b} \ge \hat{\beta}_1^{obs})}{B} = 0.0157$

 $(\hat{\beta}_1^{obs})$ counts as a random permutation

Interpretation

The probability of $p = P(\hat{\beta}_1^* \geq \hat{\beta}_1 = 0.206 | H_0)$ is equal to p = 0.0157, i.e. very small. So, it was unlikely to get a value like this **IF** H_0 **is true**.

Neyman-Pearson's approach has made common the use of a significance threshold for example $\alpha = .05$ (or = .01). When $p \le \alpha$ rejects the hypothesis that there is no relationship between X and Y (H_0) . If so, we are inclined to think that H_1 is true (there is a positive relationship).

• Type I error: False Positive the true hypo is H_0 (null correlation), BUT we accept H_1 (correlation is positive) • Type II error: False Negative the true hypo is H_1 (positive correlation), BUT we do not reject H_0 (null correlation)

2.2 To sum up

p-value: proportion of experiments providing equal or more evidence against H_0 with respect to observed data.

To compute it, we need the **Orbit** \mathcal{O} and a **Test statistic** $(T:\mathbb{R}^n\to\mathbb{R})$ quantifies the evidence against H_0

- higher values provide more evidence against H_0
- compute a test statistic for each element of the Orbit \mathcal{O} , this induces an ordering on \mathcal{O} .

In our example: $T = \hat{\beta}_1 = \hat{\sigma}_{xy}/\hat{\sigma}_{yy}$ is the (estimated) slope. Higher the slope, higher the evidence for H_1 .

Type I error control

We want to guarantee not to get false relationships (a few false positives), better to be conservative. To make this, we want to bound the probability to make a false discovery:

$$P(p-value \le \alpha|H_0) \le \alpha$$

We built a machinery that in the long run (many replicates of the experiment) finds false correlations with probability α (e.g. 0.05 = 5%).

2.2.1 We make it in flip

```
library(flip)
(res=flip(Reaction.Time~Age,data=reaction,tail=1))

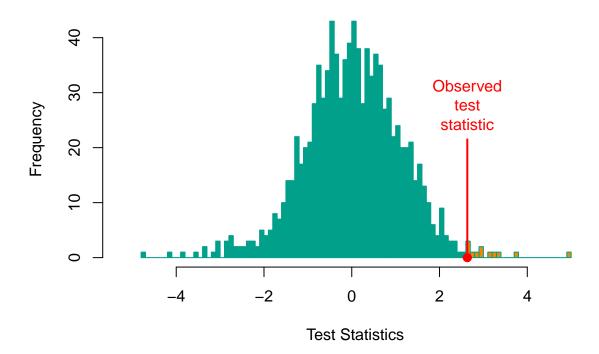
##

## Test Stat tail p-value
## Reaction.Time t 2.633 > 0.0110

## compare also with
# flip(Reaction.Time~Age,data=reaction,tail=1,statTest = "cor")
# flip(Reaction.Time~Age,data=reaction,tail=1,statTest = "coeff")

plot(res)
```

Reaction.Time



Type I error control

We want to guarantee not to get false relationships (a few false positives), better to be conservative. To make this, we want to bound the probability to make a false discovery:

$$P(p-value \le \alpha|H_0) \le \alpha$$

We built a machinery that in the long run (many replicates of the experiment) finds false correlations with probability α (e.g. 0.05 = 5%).

2.2.2 Composite alternatives (bilateral)

The hypothesis $H_1: \beta_1 > 0$ (the relation is positive) must be justified with a priori knowledge.

More frequently, the Alternative hypothesis is appropriate: $H_1: \beta_1 \neq 0$ (there is a relationship, I do not assume the direction)

I consider anomalous coefficients estimated as very small but also very large ('far from 0'). The p-value is

$$p = \frac{\#(|\hat{\beta}_1^{*b}| \ge |\hat{\beta}_1^{obs}|)}{B} = 0.0316$$

(remark: the observed test stat is included among the permuted one)

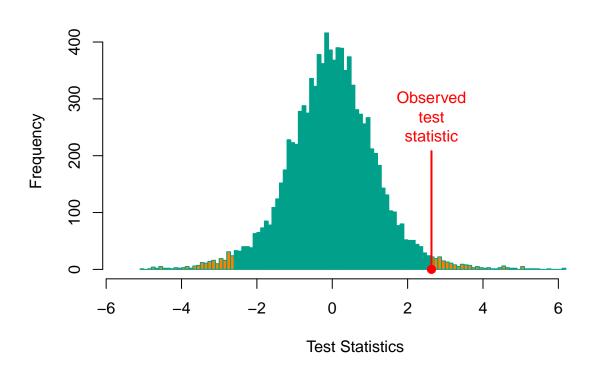
In flip:

```
library(flip)
(res=flip(Reaction.Time~Age,data=reaction,tail=0,perms=10000))
```

```
##
## Test Stat tail p-value
## Reaction.Time t 2.633 >< 0.0368</pre>
```

plot(res)





2.3 A more formal approach

(see also Pesarin, 2001; Hemerik & Goeman, 2017)

Let Y be data taking values in a sample space \mathcal{Y} . Let Π be a finite set of transformations $\pi : \mathcal{Y} \to \mathcal{Y}$, such that Π is a **group** with respect to the operation of composition of transformations, that is:

- it contains identity,
- every element has an inverse in the group,
- closure: if $\pi_1, \pi_2 \in \Pi$: $\pi_1 \circ \pi_2 \in \Pi$

(e.g. Π set of all possible permutations)

Null Hypothesis

 $H_0: Y \in \Omega_0$

Randomization Hypothesis Under the null hypothesis, the distribution of Y is invariant under the transformations in Π ; that is, for every π in Π , πY and Y have the same distribution whenever Y has distribution P in Ω_0 .

(See also Lehmann, E. L., & Romano, J. P. (2006). Testing statistical hypotheses. Springer Science & Business Media.)

Test statistic $T(Y): \mathbb{R}^n \to \mathbb{R}$

 $T^{(k)}(Y)$ is the $\lceil (1-\alpha)|\Pi| \rceil$ -th sorted value of $T(\pi Y)$

Define the test:

$$\phi(Y) = \begin{cases} 1 & \text{if } T(Y) \ge T^{(k)}(Y) \\ 0 & \text{if } otherwise \end{cases}$$
 (1)

Theorem: Under H_0 , $E_P(\phi(Y)) = \alpha$, that is $P(T(Y) \ge T^{(k)}) \le \alpha$.

Proof

By construction, $\sum_{\pi \in \Pi} \phi(\pi Y) = |\Pi|\alpha$. Therefore $|\Pi|\alpha = E_P(\sum_{\pi \in \Pi} \phi(\pi Y)) = \sum_{\pi \in \Pi} E_P(\phi(\pi Y))$

Next, by the null hypothesis: $E_P(\phi(Y)) = E_P(\phi(\pi Y))$, so that $|\Pi|\alpha = \sum_{\pi \in \Pi} E_P(\phi(Y)) = |\Pi|E_P(\phi(Y))$ gives $E_P(\phi(Y)) = \alpha$

(See also Lehmann, E. L., & Romano, J. P. (2006). Testing statistical hypotheses. Springer Science & Business Media.)

More about permutation testing

Orbit of \mathcal{O} :

$$\mathcal{O} = \{\pi Y : \pi \in \Pi\} \subseteq \mathcal{Y}.$$

(losely) the set of all samples having the same likelihood under H_0 .

$$\mathcal{O} = \{ \pi \mathbf{y} : \ f(\pi \mathbf{y}) = f(\mathbf{y}) \}$$

 $(|\mathcal{O}| \text{ number of elements of } \mathcal{O})$

If we assume exchangeability of observations, then:

$$\mathcal{O} = \{\text{all permutations of the observed data } \mathbf{y}\} = \{\mathbf{y}^* : \pi^* \circ \mathbf{y}\}$$

Remark: exchangeable observations: $f(y_1, y_2) = f(y_2, y_1)$.

It implies observations:

- are identically distributed:
 t-test and linear models assume normality, only asymptotic control of the tye I error
- have the same dependence: t-test and linear models assumes independence, which is just a special case $(f(y_1, y_2) = f(y_2)f(y_1) = f(y_2, y_1))$, i.e. more stringent assumptions

Intuition for an alternative proof of the control of the type I error

$$f(\mathbf{y}|\mathcal{O}) = \frac{f(\mathbf{y} \cap \mathcal{O})}{f(\mathcal{O})} = \frac{f(\mathbf{y})}{f(\mathcal{O})} = \frac{f(\mathbf{y})}{f(\cup_{y \in \mathcal{O}} y)} = \frac{1}{|\mathcal{O}|} \ \forall \ \mathbf{y} \in \mathcal{O}$$

i.e. each permutation is equally likely in the Orbit \mathcal{O} .

(due to group structure)

$$E(\phi(Y)|\mathbf{y} \in \mathcal{O}, H_0) =$$

$$P(T(\mathbf{y}) \ge T^{(k)}|\mathbf{y} \in \mathcal{O}, H_0) =$$

$$= \int_{T^{(k)}}^{+\infty} f(T(\mathbf{y})) dT(\mathbf{y}) =$$

$$= \sum_{\mathbf{y} \in \mathcal{O}} I(T(\mathbf{y}) \ge T/|\mathcal{O}| \le \alpha \quad \forall \mathcal{O}$$

And now $E(\phi(\mathbf{y})) = \int_P E(\phi(\mathbf{y})|\mathbf{y} \in \mathcal{O}, H_0) d\mathbf{y}$

2.3.1 Properties (see Pesarin, 2001)

The theorem above proves that the permutation tests have **exact control of the type I error**, i.e. $P(p - value \le \alpha | H_0) = \alpha$ assuming $\alpha \in \{1/|\mathcal{O}|, 2/|\mathcal{O}|, \dots, 1\}$ - don't forget that the orbit \mathcal{O} is a finite set; if this is not the case, the test is (slightly) conservative.

Further properties:

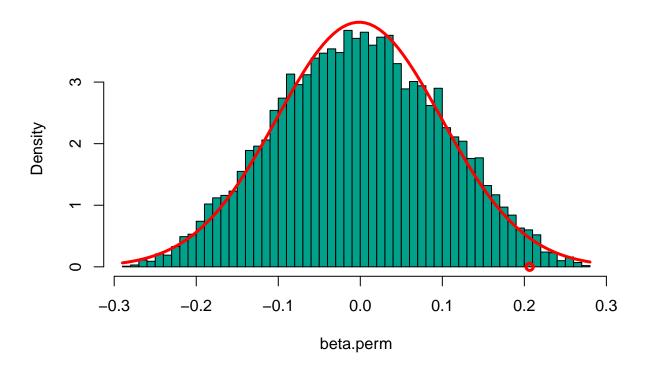
- The permutations tests are **Unbiased**: $P(p-value \leq \alpha|H_1) > \alpha$
- The test is **Consistent**: $P(p-value \leq \alpha|H_1) \to 1$ when $n \to \infty$
- The test converges to the parametric counterpart (when it exists)

2.4 A comparison (and relationships) with parametric linear model

We can see that the histogram of the statistical tests (calculated on the permuted data) is well described by a **Gaussian** (normal) curve.

```
hist(beta.perm,50,probability=TRUE,col=2)
curve(dnorm(x,mean(beta.perm),sd(beta.perm)),add=TRUE,col=1,lwd=3)
points(beta1,0,lwd=3,col=1)
```

Histogram of beta.perm



2.4.1 The (simple) linear parametric model

We assume that the observed values are distributed around true values $\beta_0 + \beta_1 X$ according to a Gaussian law: Y = linear part + normal error

$$Y = \beta_0 + \beta_1 X + \varepsilon$$

Assumptions of the linear model

- the $y_i = \beta_0 + \beta_1 x_i + \varepsilon_i$ the relationship between X and Y is truly linear, less than the error term ε_i
- $\varepsilon_i \sim N(0, \sigma^2)$, $\forall i = 1, ..., n$ errors have normal distribution with zero mean and common variance (homoschedasticity: same variance).

2.4.2 Hypothesis testing

If these assumptions are true,

$$\hat{\beta}_1 \sim N(\beta_1, \sigma^2 / \sum (x_i - \bar{x})^2)$$

We calculate the test statistic:

$$t = \frac{\hat{\beta_1}}{std.dev \ \hat{\beta_1}} = \frac{\hat{\beta_1}}{\sqrt{\sum_{i=1}^{n} (y_i - \hat{y_i})^2 / \sum (x_i - \bar{x})^2 / (n-2)}}$$

If
$$H_0: \beta_1 = 0, t \sim t(n-2)$$
 is true

On reaction data and $H_1: \beta_1 \neq 0$ (bilateral alternative)

han detto che era probabillmente uno dei partecipanti, e che bisogna disattivare la possibilità di scrivere sullo schermo, credo l'host possa risalire a chi sia stato han detto che era probabillmente uno dei partecipanti, e che bisogna disattivare la possibilità di scrivere sullo schermo, credo l'host possa risalire a chi sia stato

```
model=lm (Reaction.Time ~ Age, data=reaction)
summary (model)
```

```
##
## Call:
## lm(formula = Reaction.Time ~ Age, data = reaction)
##
## Residuals:
##
     Min
              1Q Median
                            3Q
  -6.535 -3.364 -0.272 2.676
##
##
## Coefficients:
##
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 10.30135
                           4.04407
                                     2.547
                                             0.0343 *
## Age
                0.20647
                           0.07841
                                     2.633
                                             0.0300 *
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 4.678 on 8 degrees of freedom
## Multiple R-squared: 0.4643, Adjusted R-squared: 0.3973
## F-statistic: 6.934 on 1 and 8 DF, p-value: 0.03003
```

Similar result, but much more assumptions!

2.4.3

What model do we assume in a permutation test?

Assumptions of a permutation test

Under the null hypo: $H_0: f(y) = f(y|x) \ \forall x$

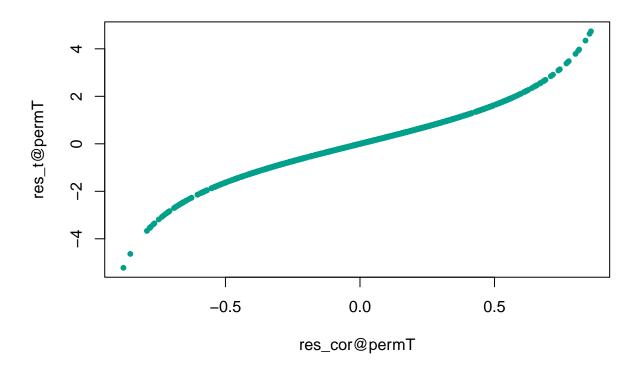
Under the alternative hypo no assumptions. in order to have power we hope that:

```
H_1: E(y|x) = \beta_0 + \beta_1 x; with \beta_1 \neq 0 and for some x that is: H_1: E(yx) \neq E(x)E(y)
```

No other assumptions on the distribution of f(y|x) (normality, nor finite moments)

2.5 Permutationally equivalent tests

```
set.seed(1)
(res_cor=flip(Reaction.Time~Age,data=reaction,statTest = "cor"))
##
##
                        Stat tail p-value
                 Test
## Reaction.Time cor 0.6814
                               >< 0.0410
set.seed(1)
(res_t=flip(Reaction.Time~Age,data=reaction,statTest = "t"))
##
##
                 Test Stat tail p-value
## Reaction.Time
                    t 2.633
                              >< 0.0410
plot(res_cor@permT,res_t@permT,pch=20,col=2)
```



2.5.1 Conclusion

The permutation tests:

• Different from bootstrap methods. The former are extractions without reintegration, the latter with. The former have almost optimal properties and have (almost always) an exact control of the first type errors.

- They constitute a general approach and are applicable in many contexts. Very few assumptions.
- some dedicated R packages:
 - coin http://cran.r-project.org/web/packages/coin/index.html
 - permuco https://cran.r-project.org/web/packages/permuco/index.html
 - flip http://cran.r-project.org/web/packages/flip/index.html (the development version is on github https://github.com/livioivil/flip)
 - flipscores http://cran.r-project.org/web/packages/flipscores/index.html (the development version is on github https://github.com/livioivil/flipscores)
 - multcomp https://cran.r-project.org/web/packages/multcomp/index.html
 - GFD https://cran.r-project.org/web/packages/GFD/index.html

3 Some special cases

3.1 Rank-correlation

- n observations from y, we are interested on F(y|x)
 - we don't need y_1 and y_2 do be continuous, we don't even neet to have finite moments (usual minimal assumption).
- Hypotheses

```
- H_0: F(y|x) = F(y|x') \ \forall x, x'
```

- $-H_1: \exists x < x': F(y|x) < F(y|x')$ or directional such as: $H_1: \exists x, x' F(y_1) \neq F(y_2)$
- Test Statistic: rank-correlation

```
(res=flip(Reaction.Time~Age,data=reaction,perms = 10000,statTest = "rank"))
##
##
                     Test Stat tail p-value
## Reaction.Time Wilcoxon 2.179
                                  >< 0.0210
# to see the rank correlation use the workaround:
(res=flip(rank(reaction$Reaction.Time)~rank(reaction$Age),perms = 10000,statTest = "cor"))
##
##
                                Test
                                       Stat tail p-value
## rank.reaction.Reaction.Time.
                                 cor 0.7153
                                              >< 0.0221
(cor.test(reaction$Reaction.Time,reaction$Age,method="spe"))
## Warning in cor.test.default(reaction$Reaction.Time, reaction$Age, method =
## "spe"): Cannot compute exact p-value with ties
##
##
   Spearman's rank correlation rho
##
## data: reaction$Reaction.Time and reaction$Age
## S = 46.983, p-value = 0.02005
## alternative hypothesis: true rho is not equal to 0
## sample estimates:
        rho
## 0.715256
plot(res)
```

rank.reaction.Reaction.Time.



3.2 The Two-independent-sample problem

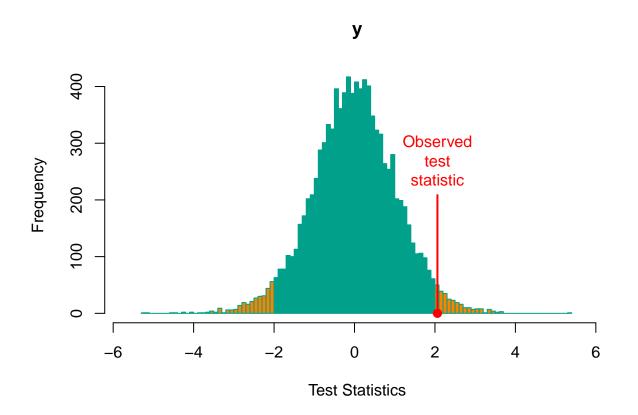
- Two samples:
 - $-n_1$ observations from y_1
 - $-n_2$ observations from y_2
 - we don't need y_1 and y_2 do be continuous, we don't even neet to have finite moments (usual minimal assumption)
- Hypotheses
 - $H_0 : F(y_1) = F(y_2)$
 - $H_1 : F(y_1) \neq F(y_2)$ (or directional such as: $H_1 : F(y_1) < F(y_2)$)
- Test Statistic:
 - Standardized mean difference (t-statistic)
 - Estimated slope coefficient (label of groups as dummy predictor)
 - other test statistic such as the (unstandardized) mean difference are permutationally equivalent

```
data("seeds")
seeds=na.omit(seeds)

(res=flip(y~grp,data=seeds,perms = 10000))
```

##
Test Stat tail p-value

```
## y
       t 2.061 >< 0.0511
(summary(lm(y~grp,data=seeds)))
##
## Call:
## lm(formula = y ~ grp, data = seeds)
##
## Residuals:
##
     Min
             1Q Median
                           3Q
                                 Max
## -7.331 -2.931 -1.651 4.663 7.863
##
## Coefficients:
              Estimate Std. Error t value Pr(>|t|)
##
## (Intercept)
                10.147
                            1.242
                                    8.168
                                             9e-09 ***
## grp
                 3.345
                            1.623
                                    2.061
                                             0.049 *
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 4.303 on 27 degrees of freedom
## Multiple R-squared: 0.136, Adjusted R-squared: 0.104
## F-statistic: 4.249 on 1 and 27 DF, p-value: 0.04903
plot(res)
```



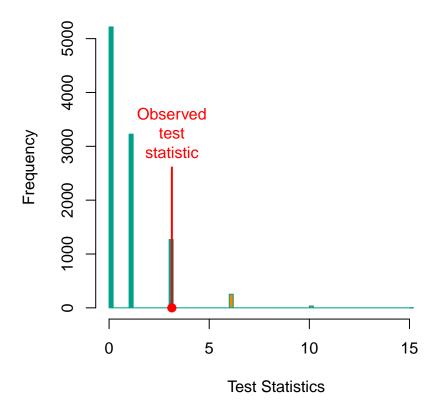
3.2.1 Rank test

Can we use rank-based statistics?

Yes, equivalent to rank-tests, we just rely on exact distribution instead of asymptotic one (and we have no limitations with ties).

```
(res=flip(y~grp,data=seeds,statTest = "rank",perms=10000))
##
##
        Test Stat tail p-value
## y Wilcoxon 2.13 >< 0.0317
(wilcox.test(y~grp,data=seeds))
## Warning in wilcox.test.default(x = c(12.54, 14.81, 16.71, 7.53, 7.02, 8.09, :)
## cannot compute exact p-value with ties
##
   Wilcoxon rank sum test with continuity correction
##
##
## data: y by grp
## W = 53.5, p-value = 0.03353
## alternative hypothesis: true location shift is not equal to 0
     Chi square and other cathegorical methods
data("seeds")
seeds$Germinated=!is.na(seeds$x)
seeds$Germinated=factor(seeds$Germinated)
seeds$grp=factor(seeds$grp)
table(seeds$grp,seeds$Germinated)
##
##
      FALSE TRUE
##
          8
              12
               17
chisq.test(seeds$grp,seeds$Germinated)
##
  Pearson's Chi-squared test with Yates' continuity correction
##
##
## data: seeds$grp and seeds$Germinated
## X-squared = 2.0063, df = 1, p-value = 0.1567
(res=flip(Germinated~grp,data=seeds,statTest = "Chisq",perms=10000))
##
##
                           Test Stat tail p-value
## grp_|_Germinated Chi Squared 3.135
                                        > 0.1557
plot(res)
```

grp_|_Germinated



... and the Fisher test:

fisher.test(seeds\$grp,seeds\$Germinated)\$p.value

```
## [1] 0.1551874
```

```
(flip(Germinated~grp, data=seeds, perms=10000))
```

```
##
## Test Stat tail p-value
## GerminatedFALSE t -1.798 >< 0.1542
## GerminatedTRUE t 1.798 >< 0.1542</pre>
```

3.4 ANOVA (C-sample)

e.g. 3 groups of Age: young [18-35), middle age [35-60), old [60-100)

- C samples:
 - n_i observations from y_i (i = 1, ..., C)
 - we don't need y_i do be continuous, we don't even neet to have finite moments (usual minimal assumption)
- Hypotheses
 - $H_0: F(y_i) = F(y_j) \ \forall (i,j)$
 - $-H_1: \exists (i,j): F(y_i) \neq F(y_j)$

```
    F-statistic

       -R^2
       - other test statistic such as the (unstandardized) mean difference are permutationally equivalent
       - Rank-based is also possible
reaction$AgeCateg=cut(reaction$Age,c(18,35,65,100),right = FALSE)
(res=flip(Reaction.Time~AgeCateg,data=reaction,perms = 10000,statTest = "ANOVA"))
##
##
                  Test Stat tail p-value
                     F 4.02
                               > 0.0838
## Reaction.Time
summary(lm(Reaction.Time~AgeCateg,data=reaction))
##
## Call:
## lm(formula = Reaction.Time ~ AgeCateg, data = reaction)
##
## Residuals:
##
      Min
              1Q Median
                             3Q
                                    Max
## -6.495 -3.279 0.465 2.246
                                 6.112
##
## Coefficients:
                     Estimate Std. Error t value Pr(>|t|)
##
## (Intercept)
                       16.157
                                    2.331
                                            6.932 0.000225 ***
## AgeCateg[35,65)
                        4.428
                                    3.296
                                            1.343 0.221144
## AgeCateg[65,100)
                                   4.037
                                            2.828 0.025478 *
                       11.418
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 4.662 on 7 degrees of freedom
## Multiple R-squared: 0.5346, Adjusted R-squared: 0.4016
## F-statistic: 4.02 on 2 and 7 DF, p-value: 0.06878
3.4.1 Stochastic Ordering
  • Same assumptions of ANOVA
  • Hypotheses
       - same null hypo H_0: F(y_i) = F(y_i) \ \forall (i,j)
       - BUT H_1 : \exists (i, j) : F(y_i) < F(y_i) \text{ (or >)}
(more details on NPC later)
(res=flip(Reaction.Time~AgeCateg,data=reaction,perms = 10000,tail=1))
##
##
                                        Test
                                               Stat tail p-value
## Reaction.Time_|_AgeCateg.[35,65).
                                           t 0.1423
                                                           0.4322
## Reaction.Time_|_AgeCateg.[65,100).
                                           t 2.2444
                                                        > 0.0259
```

• Test Statistic:

```
npc(res)
##
##
       comb.funct nVar Stat p-value
## V1
           Fisher
                      2 4.492 0.0210
3.5
       Stratified permutations (discrete nuisances)
What if we want to test x = \text{Age} also using z = \text{Gender} as nuisance in the reaction dataset?
Under the null hypothesis: f(y|x,z) = f(y|x',z) = f(y|z) \ \forall (x,x')
Therefore, even under the H_0, it holds f(y_i) = f(y_j) ONLY IF z_i = z_j (obs i and j have the same gender).
Can e permute same as in the previous cases? NO. We permute the observations only within the strata
defined by z.
Remark:
- we don't assume linear effect of the nuisance,
- we also allow heteroscedastic errors among strata.
(Test statistic remains the same)
(res=flip(Reaction.Time~Age,Strata=~Gender,data=reaction,perms=10000))
##
##
                   Test Stat tail p-value
## Reaction.Time
                      t 2.633
                                 >< 0.0684
Alternative model (more about NPC later):
(res=flip(Reaction.Time~Age*Gender,Strata=~Gender,data=reaction,perms=10000))
##
##
                                    Test
                                             Stat tail p-value
## Reaction.Time_|_Age
                                        t 2.4826
                                                     >< 0.0725
## Reaction.Time_|_Age:Gender.M.
                                        t -0.6518
                                                     >< 0.3402
npc(res)
##
##
       comb.funct nVar Stat p-value
## V1
           Fisher
                      2 3.702 0.1371
```

4 Multivariate Testing

4.1 Seeds data

```
# install.packages("flip")
library(flip)

omit the NAs:
data(seeds,package = "flip")
seeds=na.omit(seeds)
seeds

## grp x y
## 9 0 6.03 12.54
```

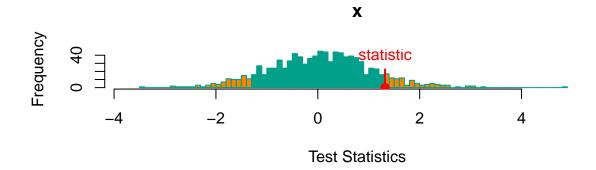
```
0 4.20 14.81
## 10
        0 4.49 16.71
## 11
        0 2.00 7.53
## 12
## 13
        0 2.84 7.02
##
  14
        0 3.88
               8.09
## 15
        0 2.04 5.76
## 16
        0 5.48 18.01
        0 2.31
## 17
                8.81
## 18
        0 1.90
                8.17
## 19
        0 1.75
               6.62
## 20
        0 3.02 7.69
## 24
        1 3.31 18.49
##
  25
        1 6.56 19.20
## 26
        1 3.16 9.85
## 27
        1 4.07 15.83
## 28
        1 2.09 6.16
## 29
        1 6.72 17.58
## 30
        1 3.93 19.29
## 31
        1 2.56 10.77
## 32
        1 8.30 18.31
## 33
        1 4.21 10.56
## 34
        1 1.86 9.48
## 35
        1 3.09 12.54
## 36
        1 5.09 18.35
## 37
        1 4.08 11.84
## 38
        1 3.63 11.44
## 39
        1 2.61 7.66
## 40
        1 5.21 12.00
```

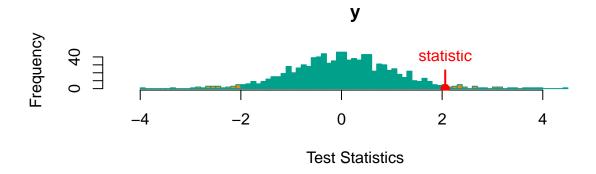
Use a permutation methods to test if there is any difference between the two groups in grp on the two variables x and y:

- perform the two tests for the two variables
- Combine the two p-values using the Fisher Combining Function to test the global hypothesis
- Use a closed testing procedure to adjust the 2 p-values.

4.2 Joint distribution

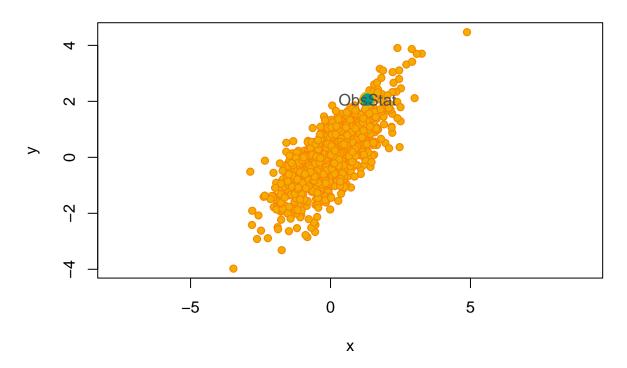
```
library(flip)
res=flip(.~grp,data=seeds)
hist(res)
```





plot(res)

Permutation Space



```
# Global p-value
npc(res, "Fisher")
##
##
      comb.funct nVar Stat p-value
                    2 4.628 0.0880
## V1
          Fisher
# adjusted p: Closed testing with Fisher combination
flip.adjust(res, "Fisher")
##
##
     Test Stat tail p-value Adjust:Fisher
                  >< 0.1810
## x
        t 1.320
                                     0.1810
                                     0.0880
        t 2.061
                  >< 0.0540
## y
```

4.3 Rejection regions

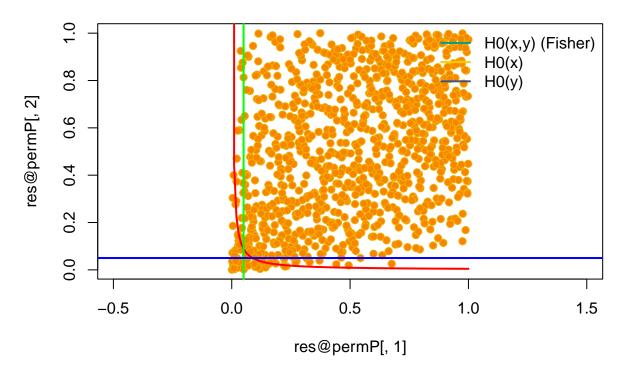
Ask for the multivariate distribution of the p-values:

```
res=flip(.~grp,data=seeds,flipReturn =list(permP=TRUE,permT=TRUE))
res.fisher=npc(res,"Fisher",flipReturn =list(permP=TRUE,permT=TRUE))
res.tippett=npc(res,"minP",flipReturn =list(permP=TRUE,permT=TRUE))
```

4.3.1 Fisher Combining Function

We inspect the rejection regions of the two univariate tests and the one of Fisher combination. The intersection of each univariate test with the Fisher region defines the rejection region of a closed testing - i.e. adjusted for multiple testing.

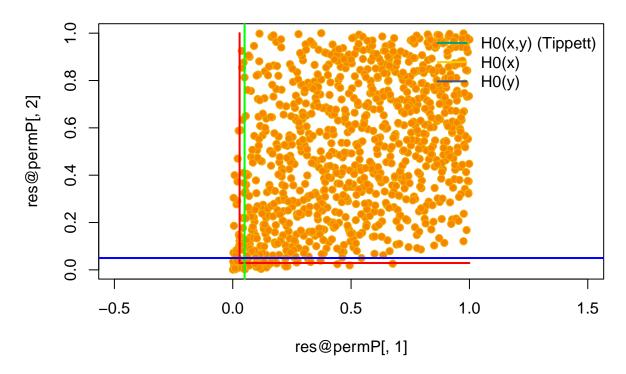
Alpha .05



4.3.2 Tippett (min-p) Combining Function

We ispect the rejection regions of the two univariate tests and the one of Fisher combination. The intersection of each univariate test with the Fisher region defines the rejection region of a closed testing - i.e. adjusted for multiple testing. This fall to be the same rejection region given by Wesfall & Young. Indeed, it is a closed testing with shortcut.

Alpha .05



5 FWER control via Permutations tests

5.1 Permutation Bonferroni

Bonferroni is conservative

- Bonferroni bound Reject for p-values at most α/m
- By Boole's inequality Guaranteed: FWER $\leq \alpha$, but often FWER $< \alpha$
- Can we improve? Reject for p-values at most $\tilde{\alpha} > \alpha/m$, while keeping FWER control
- Yes we can
 By permutations

5.2 Improved Bonferroni

- Reduced α Reject H_i if $p_i \leq \tilde{\alpha}$
- Control of FWER?

FWER =
$$P(p_i \le \tilde{\alpha} \text{ for at least one } i \text{ with } H_i \text{ true})$$

= $P(\bigcup_{i \in T} \{p_i \le \tilde{\alpha}\})$
= $P(\min_{i \in T} p_i \le \tilde{\alpha}) \le \alpha$

• How can we determine the value of $\tilde{\alpha}$?
Using permutations to find the distribution of the minimum p-value

5.3 Multiple testing using permutations

The single step min-P method

- Calculate the smallest p-value m for the real data
- Randomly permute the data
- Calculate new p-values for all tests based on permuted data
- Calculate the smallest p-value m^{π} for permuted data
- Repeat permutation many (say k=1000) times: $m_1^{\pi}, \ldots, m_k^{\pi}$
- Calculate $\tilde{\alpha}$ as the α -quantile of $m_1^{\pi}, \ldots, m_k^{\pi}$

Multiple testing result

Reject all hypotheses with (non-permuted) p-values at most $\tilde{\alpha}$

5.4 Correlation structure of p-values

Permutation

- Destroys correlation between covariates and response
- Retains correlation among covariates

Consequence

- P-values of correlated tests (i.e. data) remain correlated in permutations
- Distribution of minimum p-value correctly takes correlations into account

When the gain relative to Bonferroni is the gain large?

- Negatively correlated p-values: typically no gain
- Independent p-values: minimal gain
- Positively correlated p-values: gain can be large

5.5 Westfall & Young: permutation Holm

Westfall PH, Young SS (1993) Resampling-Based Multiple Testing: Examples and Methods for p-Value Adjustment. Wiley

Sequential permutation multiple testing

• Single step

Single step min-P is permutation equivalent of Bonferroni

• What about Holm?

Permutation equivalent of Holm's method: Westfall & Young

The min-P algorithm

- Start with all hypotheses
- Repeat
 - Do single step min-P to calculate $\tilde{\alpha}$
 - Reject hypotheses with p-value $\leq \tilde{\alpha}$
 - Remove rejected hypotheses
- Until no new rejections occur

5.6 Closed Testing

R Marcus, E Peritz, KR Gabriel (1976). On closed testing procedures with special reference to ordered analysis of variance. Biometrika 63: 655-660.

Test in each node: any multivariate permutation test

5.6.1 Closure Set

ABC

AB AC BC

A B

Adjusted $\tilde{p}_A = \max(p_A, p_{AB}, p_{AC}, p_{ABC})$

5.7 Conclusion

Accounting for dependencies

Adjusted p-value become lower (i.e. more rejections)

When?

- Negative correlation: generally no gain p-value Independents: little or no gain
- Positive correlation: big gain, usually

(NB: a test with bi-directional alternative and with negative correlation produce p-value positively correlated)

Real data

The variables of real datasets are often correlated then permutations are (often) convenient

How? R: library(flip); flip(); flip.adjust()

6 A case study: Pharmacokinetic Study of Carbidopa

Description:

http://webserv.jcu.edu/math//faculty/TShort/Bradstreet/part2/part2-table6.html

As part of a pharmacokinetic study, 12 healthy male subjects were allocated randomly to a three period crossover design receiving one of three graded doses (25, 50, 100 mg) of Carbidopa q8h in each treatment period. A seven day washout period separated the treatment periods. The pharmacokinetic variables AUC, Cmax, and Tmax were calculated for each subject from plasma concentrations assayed from blood samples taken at 0, 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7, and 8 hours postdosing following the second dose of carbidopa on the sixth day of each treatment period.

dataset:

http://webserv.jcu.edu/math//faculty/TShort/Bradstreet/part2/Bradp2t6.txt

Analyze the dataset without taking in account the Study Periods (which have been randomized in each subject, hence we can avoid to account for it in the analysis).

Research questions:

• Is there a dose response for AUC, Cmax, or Tmax? Overall?

\$ AUC : num 604 140 386 175 605 ...

\$ Tmax: num 1.5 1 1.5 1.5 0.5 1 1 1 2 2 ...

\$ Cmax: num 137 44.4 86.6 46.4 194 44.9 318 29 119 58.4 ...

• Can dose proportionality be established? (try to fit a linear model for each endpoint, then discuss the results)

6.1 A solution

We answer to both first and second question with a single analysis: we perform a linear model (accounting for individual variability) on log transformed end-points.

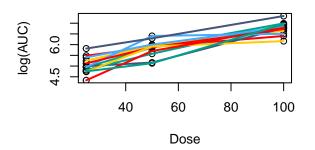
```
# transform all responses with log-transformed,
# so that a linear relationship between time and end-point indicates proportionality
dati[,3:5]=log(dati[,3:5])
#Descriptives and plots:
summary(dati[,-1])
                        AUC
        Dose
                                       Cmax
                                                      Tmax
## Min. : 25.00
                   Min. :4.337
                                  Min. :3.219
                                                Min. :-0.6931
  1st Qu.: 25.00
                   1st Qu.:5.156
                                  1st Qu.:3.966
                                                 1st Qu.: 0.0000
                   Median :5.886
                                  Median :4.485
## Median : 50.00
                                                 Median: 0.2027
## Mean : 58.33
                   Mean
                        :5.873
                                  Mean :4.547
                                                 Mean : 0.2474
## 3rd Qu.:100.00
                   3rd Qu.:6.539
                                  3rd Qu.:5.280
                                                 3rd Qu.: 0.6931
## Max.
        :100.00
                   Max. :7.335
                                  Max. :5.989
                                                 Max. : 1.0986
by(dati[,3:5],dati$Dose,summary)
## dati$Dose: 25
##
        AUC
                       \mathtt{Cmax}
                                      Tmax
## Min. :4.337
                 Min. :3.219 Min. :-0.6931
  1st Qu.:4.803
                 1st Qu.:3.390
                                 1st Qu.: 0.0000
## Median :4.972 Median :3.801
                                 Median : 0.0000
## Mean :5.051
                  Mean :3.783
                                 Mean : 0.2071
                  3rd Qu.:4.022
##
   3rd Qu.:5.289
                                 3rd Qu.: 0.6931
##
  Max. :5.818
                  Max. :4.464
                                 Max. : 0.6931
## dati$Dose: 50
##
       AUC
                      Cmax
                                     Tmax
## Min. :5.133 Min. :3.837 Min. :-0.6931
## 1st Qu.:5.670 1st Qu.:4.374 1st Qu.: 0.0000
## Median :5.886
                  Median :4.484
                                 Median: 0.2027
## Mean :5.815
                  Mean :4.479
                                 Mean : 0.1689
## 3rd Qu.:5.967
                  3rd Qu.:4.625
                                 3rd Qu.: 0.4055
## Max. :6.405
                  Max. :5.268
                                 Max. : 1.0986
## -----
## dati$Dose: 100
        AUC
                       Cmax
                                      Tmax
                  Min. :4.920
## Min. :6.164
                                 Min. :0.0000
## 1st Qu.:6.607
                  1st Qu.:5.229
                                 1st Qu.:0.0000
## Median :6.782 Median :5.412
                                 Median :0.4055
## Mean :6.751
                  Mean :5.378
                                 Mean :0.3662
## 3rd Qu.:6.922
                  3rd Qu.:5.515
                                 3rd Qu.:0.6931
## Max. :7.335
                  Max. :5.989
                                 Max. :1.0986
par(mfrow=c(2,2))
plot(dati$Dose,dati$AUC,ylab="log(AUC)",xlab="Dose",main="Dose vs log(AUC)")
r=sapply(unique(dati$Sub),function(s){
 d=subset(dati,Sub==s)
 d=d[order(d$Dose),]
 lines(d$Dose,(d$AUC),col=s,lwd=2)})
plot(dati$Dose,dati$Cmax,ylab="log(Cmax)",xlab="Dose",main="Dose vs log(Cmax)")
r=sapply(unique(dati$Sub),function(s){
```

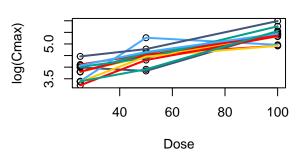
```
d=subset(dati,Sub==s)
  d=d[order(d$Dose),]
  lines(d$Dose,(d$Cmax),col=s,lwd=2)})

plot(dati$Dose,dati$Tmax,ylab="log(Tmax)",xlab="Dose",main="Dose vs log(Tmax)")
r=sapply(unique(dati$Sub),function(s){
  d=subset(dati,Sub==s)
  d=d[order(d$Dose),]
  lines(d$Dose,(d$Tmax),col=s,lwd=2)})
```

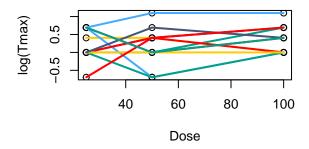
Dose vs log(AUC)

Dose vs log(Cmax)





Dose vs log(Tmax)



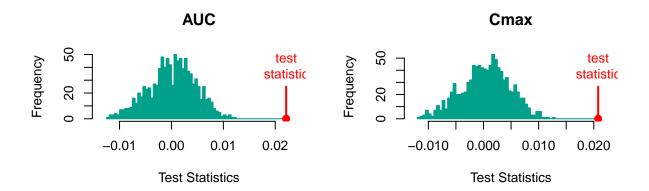
Now the analysis: A simple solution could be:

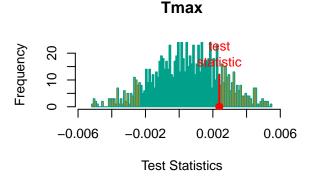
>< 0.2680

Tmax coeff 0.0024

```
library(flip)
res=flip(.~Dose,data=dati,Strata=~Sub,statTest = "coeff")
summary(res)

## Call:
## flip(Y = . ~ Dose, data = dati, statTest = "coeff", Strata = ~Sub)
## 999 permutations.
##
## Test Stat tail p-value sig.
## AUC coeff 0.0221 >< 0.0010 ***
## Cmax coeff 0.0208 >< 0.0010 ***</pre>
```





comb.funct nVar Stat p-value

Multivariate:

##

• Overall

```
res=flip.adjust(res)
npc(res,"Fisher")
##
```

V1 Fisher 3 15.13 0.0010 There is an effect of Dose, overall.

• By end-points (closed testing with max-t combining function). Try also different methods (e.g. method="Fisher") and compare the results of method="minP" with the one of method="Holm".

```
res=flip.adjust(res,method="holm")
res=flip.adjust(res,method="Fisher")
summary(res)
```

## Cmax coeff 0.0208	><	0.0010	0.0010	0.0030	0.0020	**
## Tmax coeff 0.0024	><	0.2680	0.2680	0.2680	0.2680	

AUC and Cmax show a significant effect after correction for multiplicity, while Tmax does not.

7 (minimal) Bibliography

The Grounding Theory:

- Pesarin (2001) Multivariate Permutation Tests: With Applications in Biostatistics by Fortunato, Wiley, New York

A flexible approach to General Linear Model based on the sign-flip score test:

- Hemerik, Goeman and Finos (2020) Robust testing in generalized linear models by sign flipping score contributions. Journal of the Royal Statistical Society Series B (Statistical Methodology) 82(3). DOI: 10.1111/rssb.12369

Implemented in R package flipscores:

https://cran.r-project.org/web/packages/flipscores/index.html

better to use the github develop version:

https://github.com/livioivil/flipscores

A nice review of the regression model within the permutation framework:

- Anderson M. Winkler, Gerard R. Ridgway, Matthew A. Webster, Stephen M. Smith, Thomas E. Nichols (2014) Permutation inference for the general linear model, NeuroImage, Volume 92, Pages 381-397, ISSN 1053-8119 https://doi.org/10.1016/j.neuroimage.2014.01.060