



NGS Techniques

Practical workshop on Large-Scale Genomic Data Analyses: GWAS in structured populations

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Learning objectives

In this class we will learn

- What are model and non-model organisms
- Why and how can we study non-model organisms
- How sequencing technologies have enabled the study of natures biodiversity
- What a reference genome is
- How to sequence, assemble and annotate a genome
- How to sequence and assemble a transcriptome
- Among other things





The sequencing revolution

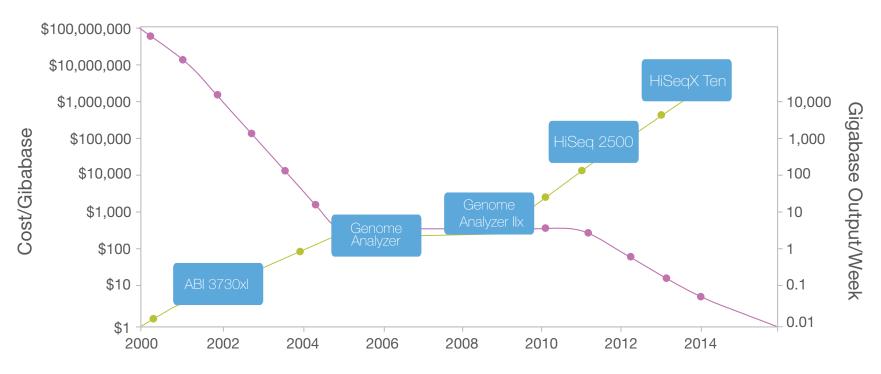
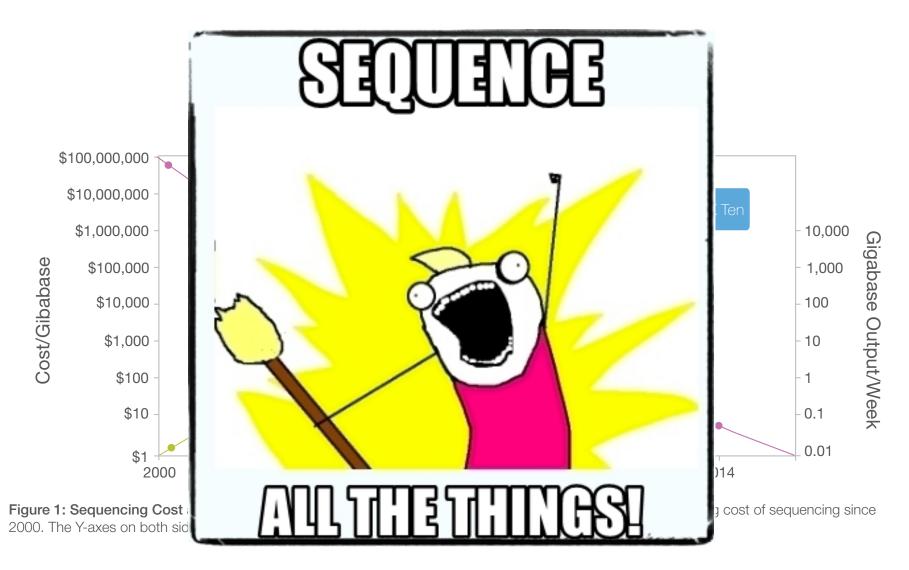


Figure 1: Sequencing Cost and Data Output Since 2000—The dramatic rise of data output and concurrent falling cost of sequencing since 2000. The Y-axes on both sides of the graph are logarithmic.









High-throughput sequencing techniques

- Pyrosequencing
- Sequencing by synthesis
- Sequencing by ligation
- Ion semiconductor
- Nanopore sequencing
- Single Molecule Real Time Sequencing (SMRT)







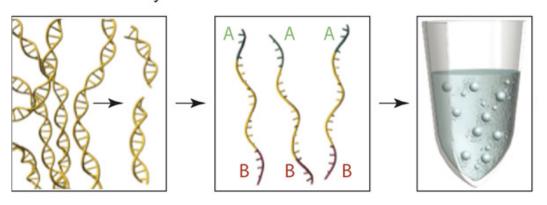


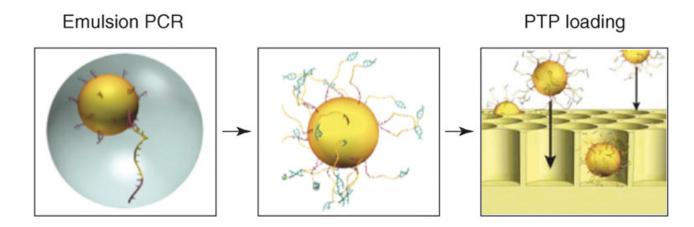




Pyrosequencing - 1

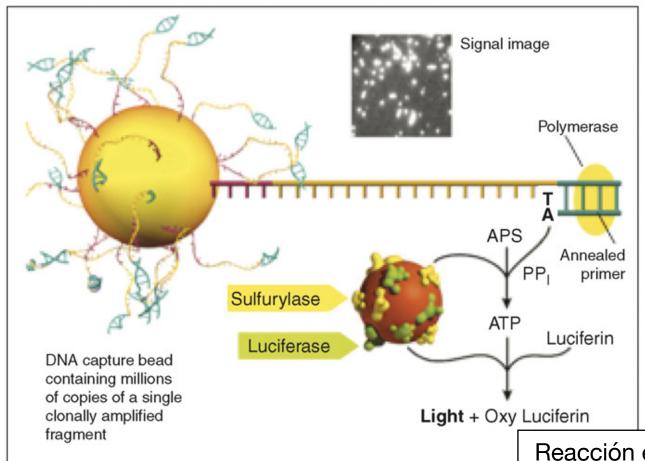
Roche (454) GSFLX Workflow: Library construction







Pyrosequencing - 2



Pyrosequencing reaction

Reacción enzimatica chemoluminiscente





Pyrosequencing

Advantages

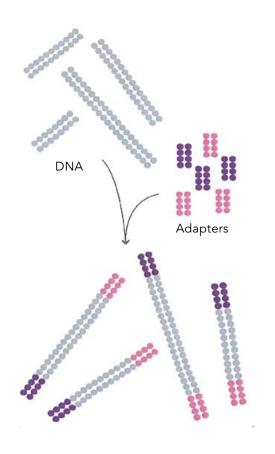
- Reasonable cost
- Long sequences (500 nts)

Disadvantages

- Few sequences produced
- High number of errors in regions with the same nucleotide (homopolymers)
- With the rise of other technologies and given its high level of errors it was ultimately discontinued



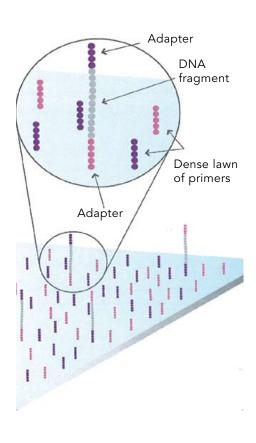




 The process starts by joining adapters to the DNA or RNA fragments that we want to sequence.



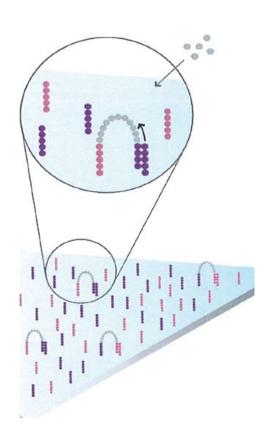




- The templates are immobilized on a flow cell
- In the case of RNA-Seq, complementarity with the adapter is used to synthesize a new cDNA chain in order to preserve information about the directionality of the transcript.

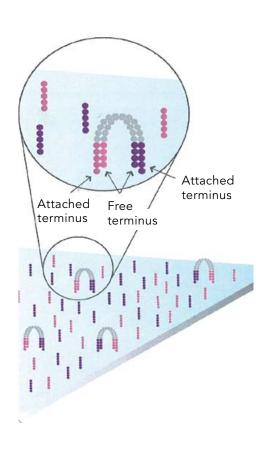






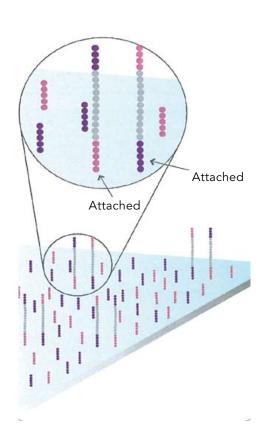
 A chain of DNA complementary to the DNA template is synthesized on the flow cell surface.





 A chain of DNA complementary to the DNA template is synthesized on the flow cell surface.

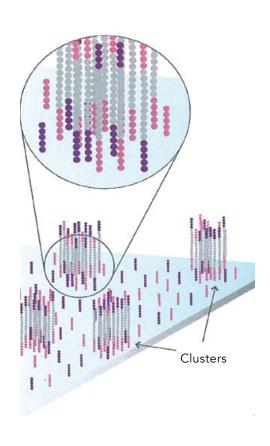




 The templates are separated using high temperature.



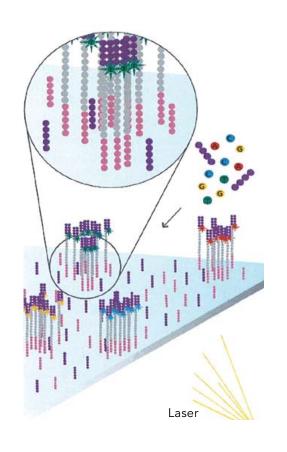


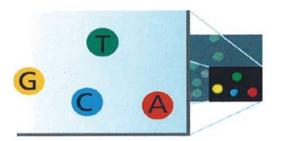


 This process is repeated hundreds of times until generating a "colony" or cluster of identical transcripts.





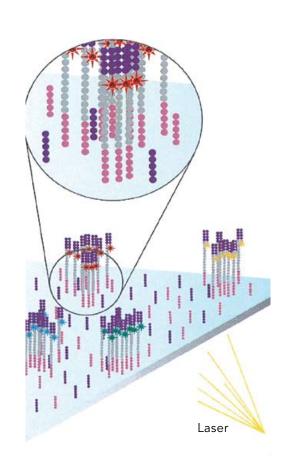


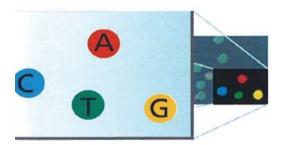


 Primers and fluorescent nucleotides (reversible terminators) are added in order (first A, then T, etc.) along with polymerase. When a nucleotide is incorporated a laser pulse coupled with imaging are used to identify which base was incorporated in each position.





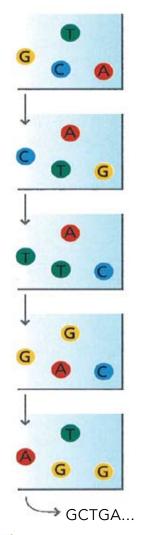


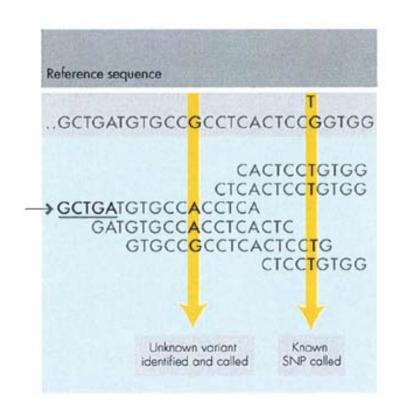


 This process is continued for all bases.









 The images are analyzed spatially to reveal each sequence.





Sequencing by Synthesis

Advantages

- Undoubtedly the leader in the market = strong scientific support network
- Produces large amounts of sequences
- Low error rate compared with other technologies

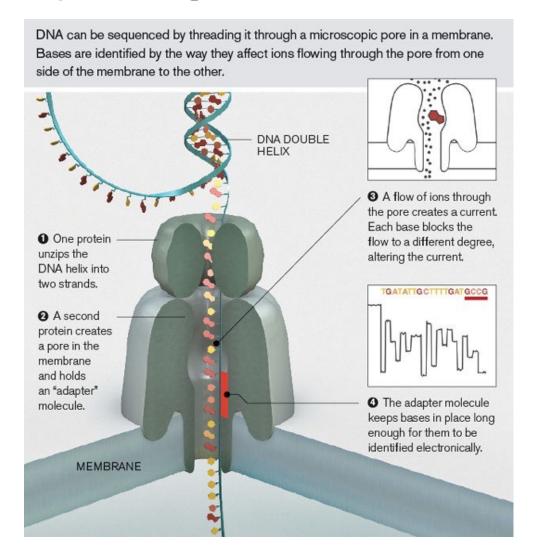
Disadvantages

- The sequences are short (150 to 250 bp)
- The cost is high
- Relatively slow sequencing





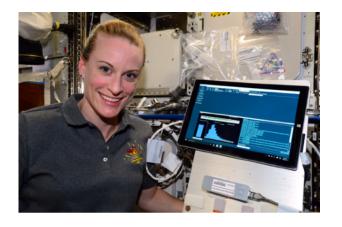
Nanopore sequencing

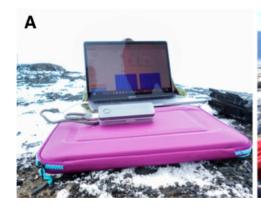




Nanopore sequencing













Nanopore whale watching



"Both jaws, like enormous shears, bit the craft completely in

- Nanopore is capable of generating very very long reads or "whales"
- The longest read detected to date has a length of 2,272,580 bases



Nanopore sequencing

Advantages

- Real-time sequencing
- You can stop sequencing when you have enough data
- Very portable useful for work in difficult areas
- Simple preparation
- Low cost \$ 80 USD per sample

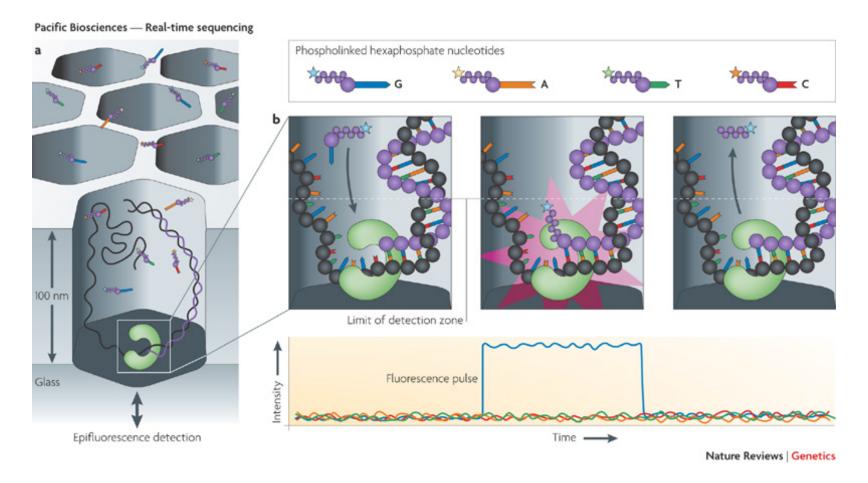
Disadvantages

- High number of errors although they have had a drastic increase in accuracy in the last year
- Pores failed sequence loss





Single Molecule Real Time Sequencing (SMRT)







SMRT sequencing

Advantages

- Does not require amplification reducing errors in abundance
- Very long sequences (860)
 - 1100 bp)

Disadvantages

- High number of errors such as the removal of CG bases among several others
- Extremely expensive





Sources of error

- There are two main sources of error:
 - Human error: mixing of samples (in the laboratory or when the files were received), errors in the protocol
 - Technical error: Errors inherent to the platform (e.g., mononucleotide sequences in pyrosequencing) - All platforms have some level of error that must be taken into account when designing the experiment.



Errors in sample preparation

- User error (e.g. mistakenly labeling a sample)
- DNA / RNA degradation by preservation methods
- Contamination with external sequences
- Low amount of DNA start





Errors in library preparation

- User error (e.g. polluting one sample with another, contaminate with previous reactions, errors in the protocol)
- PCR amplification errors
- Bias for primers (binding bias, methylation bias, primer dimers [first dimers])
- Bias for capture (Poly-A, Ribozero)
- Machine errors (misconfiguration, reaction interruption)
- Chimeras
- Index errors, adapter (contamination of adapters, lack of index diversity, incompatible codes (barcodes), overload)





Sequencing and image errors

- User error (e.g. cell overload)
- Delay (e.g., incomplete extension, addition of multiple nucleotides)
- Dead fluorophores, damaged nucleotides and overlapping signals
- Context of the sequence (e.g. high GC content, homologous) and low complexity sequences, homopolymers).
- Machine errors (e.g. laser, hard disk, programs)
- Chain biases





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The challenge - differentiate biological signals from noise/errors

- Negative and positive controls What do I expect?
- Technical and biological replicas help determine the noise rate
- Know the types of common errors in a certain platform





Now what?

