

aneur: comparando stan y cgam

1. Aortic Aneurysm Progression Data

This dataset contains longitudinal measurements of grades of aortic aneurysms, measured by ultrasound examination of the diameter of the aorta.

A data frame containing 4337 rows, with each row corresponding to an ultrasound scan from one of 838 men over 65 years of age.

- ptぬm (numeric) Patient identification number
- age (numeric) Recipient age at examination (years)
- diam (numeric) Aortic diameter
- state (numeric) State of aneurysm.

The states represent successive degrees of aneurysm severity, as indicated by the aortic diameter.

- State 1 Aneurysm-free < 30 cm
- State 2 Mild aneurysm 30-44 cm
- State 3 Moderate aneurysm 45-54 cm
- State 4 Severe aneurysm > 55 cm

683 of these men were aneurysm-free at age 65 and were re-screened every two years. The remaining men were aneurysmal at entry and had successive screens with frequency depending on the state of the aneurysm. Severe aneurysms are repaired by surgery.

```
data(aneur)
attach(aneur)
head(aneur)

##   ptぬm      age diam state
## 1     1 60.00000  29     1
## 2     1 65.47671  29     1
## 3     1 67.50411  29     1
## 4     1 70.04384  29     1
## 5     1 72.07671  29     1
## 6     1 74.08767  29     1

tail(aneur)
```

```

##      ptnum      age diam state
## 4332   838 73.40822    43     2
## 4333   838 73.61644    43     2
## 4334   838 73.87671    42     2
## 4335   838 74.05753    43     2
## 4336   838 74.31507    41     2
## 4337   838 74.56712    40     2

#help(aneur)
dim(aneur)

## [1] 4337     4

(N = n_distinct(aneur$ptnum))    # subjects

## [1] 838

(K = max(table(aneur$ptnum)))    # times

## [1] 21

table(table(aneur$ptnum))

##      2      3      4      5      6      7      8      9      10      11      12      14      15      16      17      18      19      21 
## 121 107  99  96 260  97  12  12  9  5  2  5  5  3  1  2  1  1 

J = 4    # categories
Y_diam = array(NA,dim=c(N,K))
Y_state = array(NA,dim=c(N,K))
X_age = array(NA,dim=c(N,K))
Ki = table(aneur$ptnum)
Ni = c(0,cumsum(Ki))+1
for(i in 1:N){
  aneur_i = aneur[aneur$ptnum==i,]
  for(k in 1:Ki[i]){
    Y_diam[i,k] = aneur_i$diam[k]
    Y_state[i,k] = aneur_i$state[k]
    X_age[i,k] = aneur_i$age[k]
  }
}
(Y_diam[11:18,1:8])

##      [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8]
## [1,]    29    29    29    29    29    29    29   NA
## [2,]    29    29    29    29    29    29    29   NA
## [3,]    29    29    29    29    29    29    29   NA
## [4,]    29    29    NA    NA    NA    NA    NA   NA
## [5,]    29    29    29    29    29    29    29   NA
## [6,]    29    29    29    29    29    29    29   NA
## [7,]    29    29    29    29    NA    NA    NA   NA
## [8,]    29    29    34    NA    NA    NA    NA   NA

```

```
(Y_state[11:18,1:8])
```

```
##      [,1] [,2] [,3] [,4] [,5] [,6] [,7] [,8]
## [1,]    1    1    1    1    1    1    1    NA
## [2,]    1    1    1    1    1    1    1    NA
## [3,]    1    1    1    1    1    1    1    NA
## [4,]    1    1   NA   NA   NA   NA   NA   NA
## [5,]    1    1    1    1    1    1    1    NA
## [6,]    1    1    1    1    1    1    1    NA
## [7,]    1    1    1    1   NA   NA   NA   NA
## [8,]
```

```
(X_age[11:18,1:8])
```

```
##      [,1]      [,2]      [,3]      [,4]      [,5]      [,6]      [,7]      [,8]
## [1,] 60 65.45205 67.45205 69.92877 72.01096 74.01096 76.00000  NA
## [2,] 60 65.44932 67.46301 69.92603 71.96986 73.96986 75.92055  NA
## [3,] 60 65.45753 67.44658 69.92329 71.96712 73.96712 75.91781  NA
## [4,] 60 65.44384       NA       NA       NA       NA       NA  NA
## [5,] 60 65.43836 67.42192 69.93699 71.94247 73.94247 75.89315  NA
## [6,] 60 65.40822 67.40822 70.04932 72.07123 74.06575 76.09041  NA
## [7,] 60 65.38082 67.38082 70.02192       NA       NA       NA  NA
## [8,]
```

```
(Ki[11:18])
```

```
##
## 11 12 13 14 15 16 17 18
## 7 7 7 2 7 7 4 3
```

```
### Considering only data having more than one screen (state>1)
idx2 = c()
for(i in 1:N){
  if( sum(Y_state[i,1:Ki[i]])>Ki[i]){
    idx2 = c(idx2,i)
  }
}
Y2_diam = Y_diam[idx2,]
Y2_state = Y_state[idx2,]
X2_age = X_age[idx2,]
N2 = length(idx2)
Ki2 = Ki[idx2]

### Considering only data having more than one screen (diam!=29, or diam<29 & dim>29)
idx3 = c()
for(i in 1:N){
  if( min(Y_diam[i,1:Ki[i]])!=max(Y_diam[i,1:Ki[i]])){
    idx3 = c(idx3,i)
  }
}
Y3_diam = Y_diam[idx3,]
```

```

Y3_state = Y_state[idx3,]
X3_age = X_age[idx3,]
N3 = length(idx3)
Ki3 = Ki[idx3]

aneur2 = aneur%>%filter(aneur$ptnum%in%idx2)
aneur3 = aneur%>%filter(aneur$ptnum%in%idx3)
### Creo que es mejor trabajar con aneur3

```

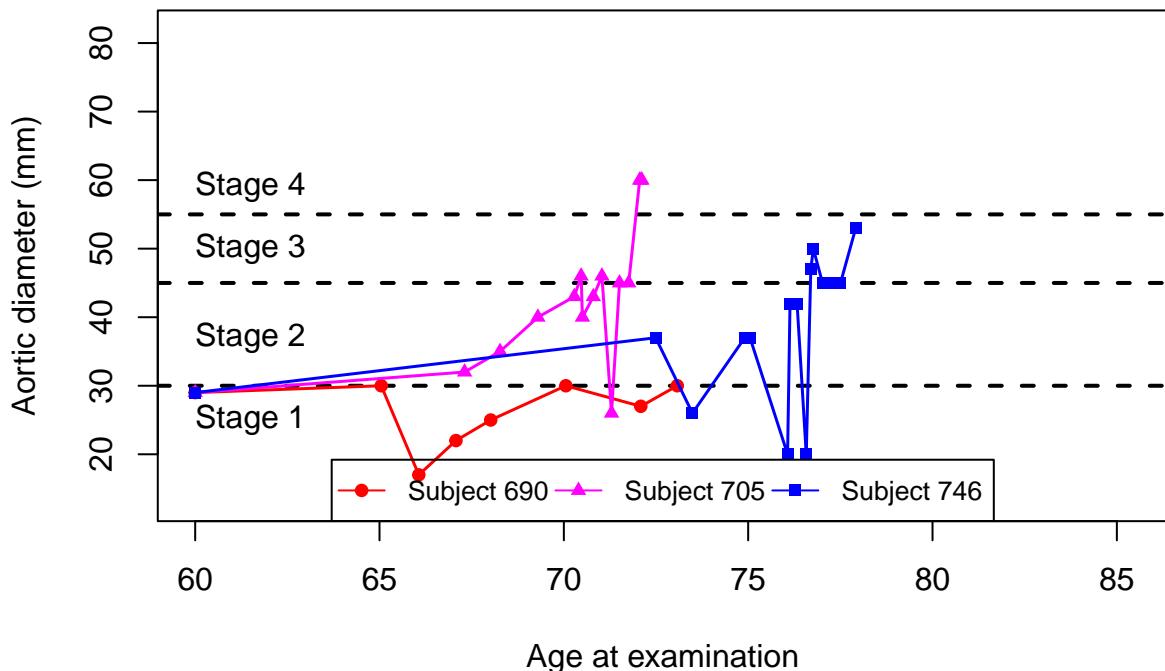
```

##  

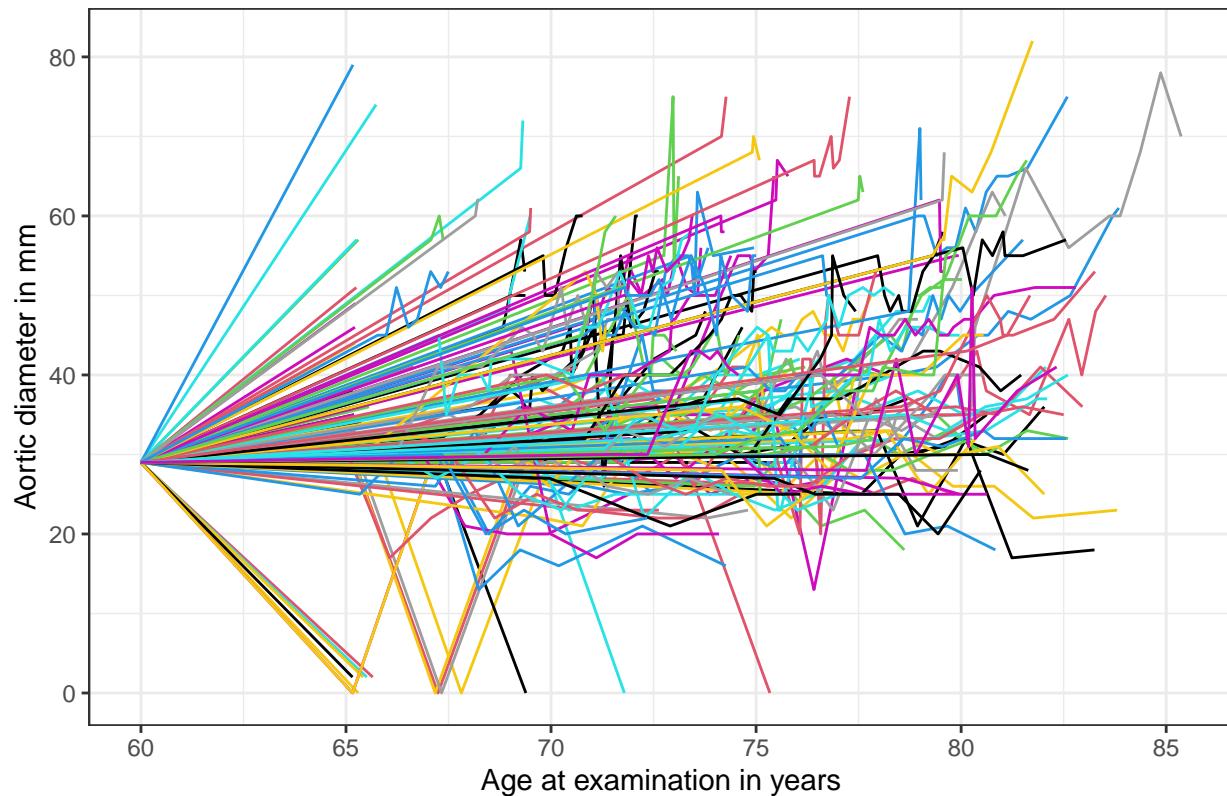
##   67   80  119  

##     6     6     6

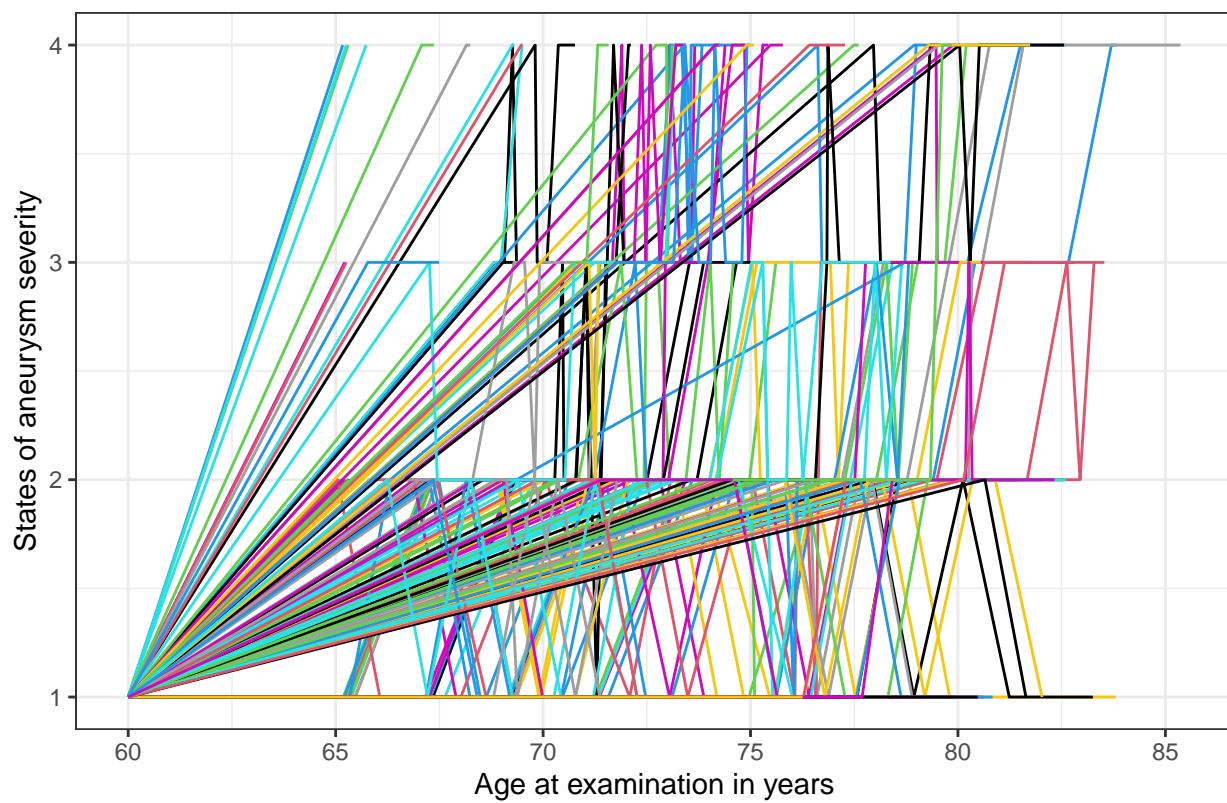
```



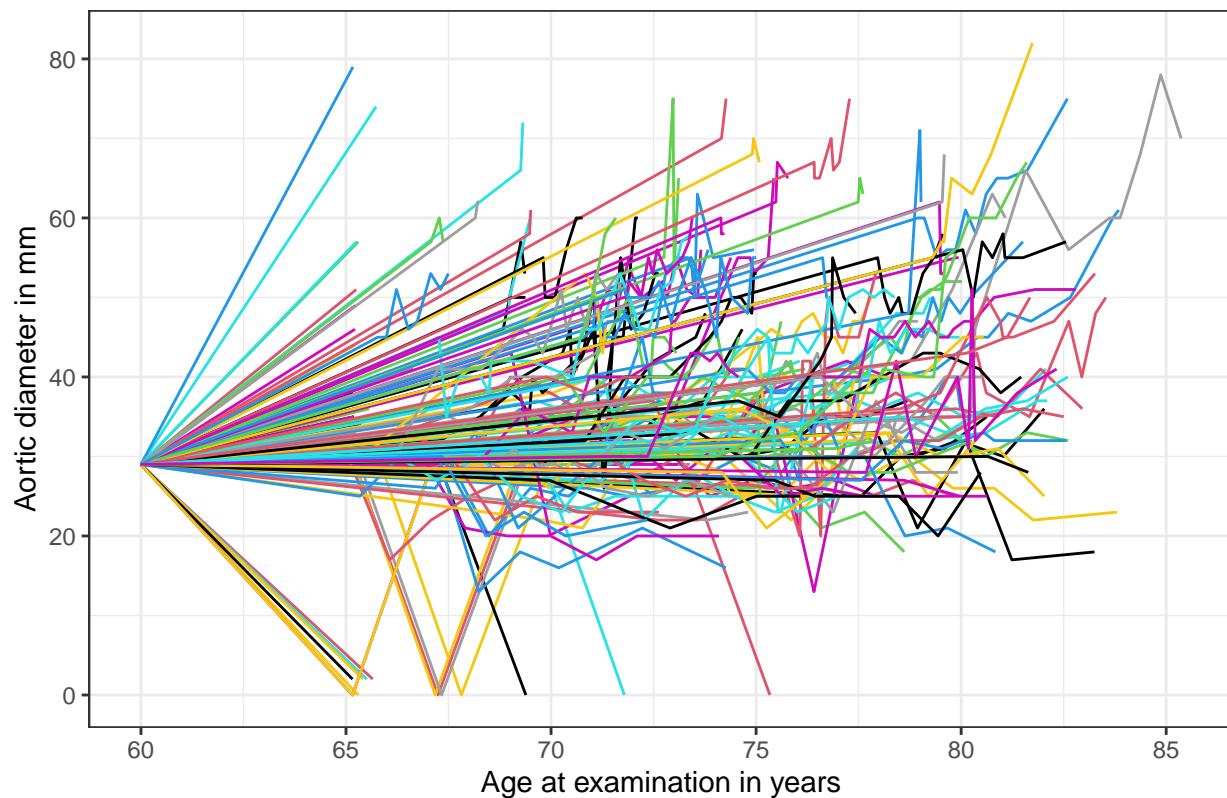
Profiles aortic diameter by patient



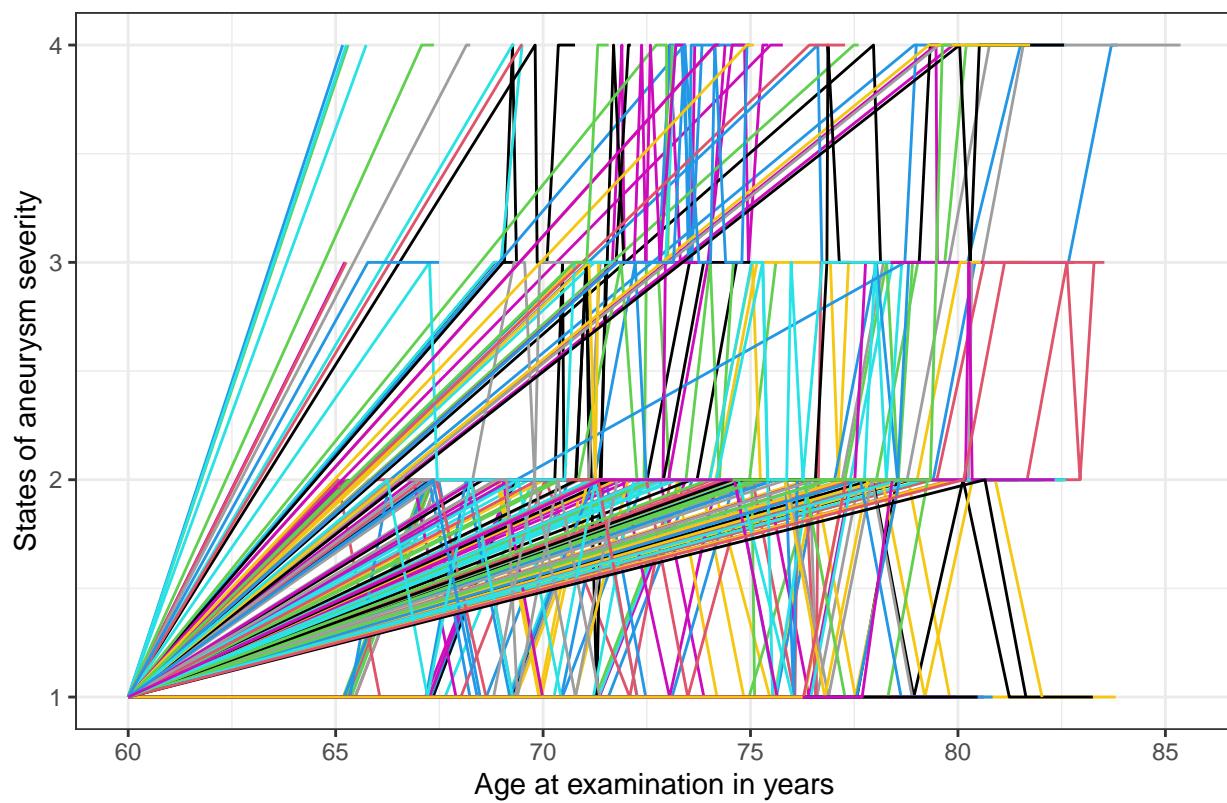
Profiles states of aneurysm severity by patient



Profiles aortic diameter by patient



Profiles states of aneurysm severity by patient



La variable respuesta puede ser continua ("diam") u ordinal ("state"), y la única covariable es la edad

(“age”) \

$$diam_{it} = \beta_0 + f_1(age_{it}) + b_{0i} + age_{it} \times b_{1i} + \varepsilon_{it}, \quad b_i \sim N(0, \psi), \quad \varepsilon_i \sim N(0, \Lambda\sigma^2),$$

where f_1 is a non-decreasing smoothing function and $b_{1i} > 0$.

Quiza solo debemos considerar intercepto fijo, pero NO intercepto aleatorio, y SI pendiente aleatorio

$$diam_{it} = \beta_0 + f_1(age_{it}) + age_{it} \times b_{1i} + \varepsilon_{it}, \quad b_{1i} \sim N(0, \psi), \quad \varepsilon_i \sim N(0, \Lambda\sigma^2),$$

The ordinal response $state_{it}$ is modelled in terms of the cumulative probabilities $P(state_{it} \leq j|b_i)$ by using the proportional odds model,

$$P(state_{it} \leq j|b_i) = \eta_{it,j},$$

subject to

$$\eta_{it,j} = \kappa_j + \beta_0 + f_1(age_{it}) + age_{it} \times b_{1i}, \quad b_{1i} \sim N(0, \psi),$$

where the constraints are such that f_1 is a non-decreasing smoothing function and $b_{1i} > 0$, and for the breakpoints $\kappa_j < \kappa_{j+1}$ with $j = 1, 2$.

```
y = aneur3$diam -29
x1 = aneur3$age -60
x2 = aneur3$age -60
id = as.numeric(as.factor(aneur3$ptnum))
```

```
(n = length(y))
```

```
## [1] 1387
```

```
(N = n_distinct(id))
```

```
## [1] 229
```

```
Ni = c(0,cumsum(table(id)))+1
k1 = 4 #knots
k2 = 4 #knots
knots1 = quantile(x1, c(0.33,0.67))
knots2 = quantile(x2, c(0.33,0.67))
```

2. Generar la matriz diseño X para los B-splines

Note que $f(x)$ se representa como:

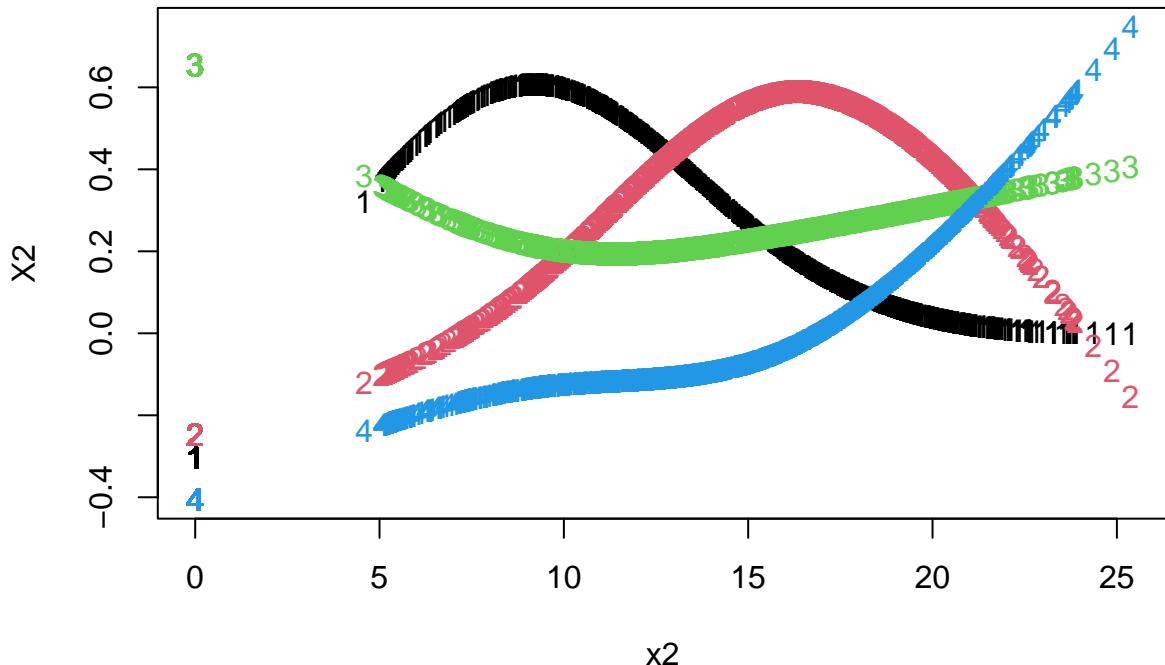
$$\begin{aligned} f(x) &= f_1(x_1) \\ &= \sum_{j=1}^{h_1} \beta_{1j} I_{1j}(x) \end{aligned}$$

para β_{1j} parámetros desconocidos, y para los $I_{1j}(x)$ se utilizarán I-splines y B-splines.

El número de knots se elige lo suficientemente grande para evitar **over-smoothing**, pero lo suficientemente pequeño para evitar excesivo costo computacional.

El número de knots K es considerado a priori.

```
# Generate a basis matrix for Natural Cubic Splines
X2 <- ns(x = x2, knots = knots2, intercept = TRUE)
####X2 = (X2-mean(X2))/sd(X2)
matplot(x2, X2)
```

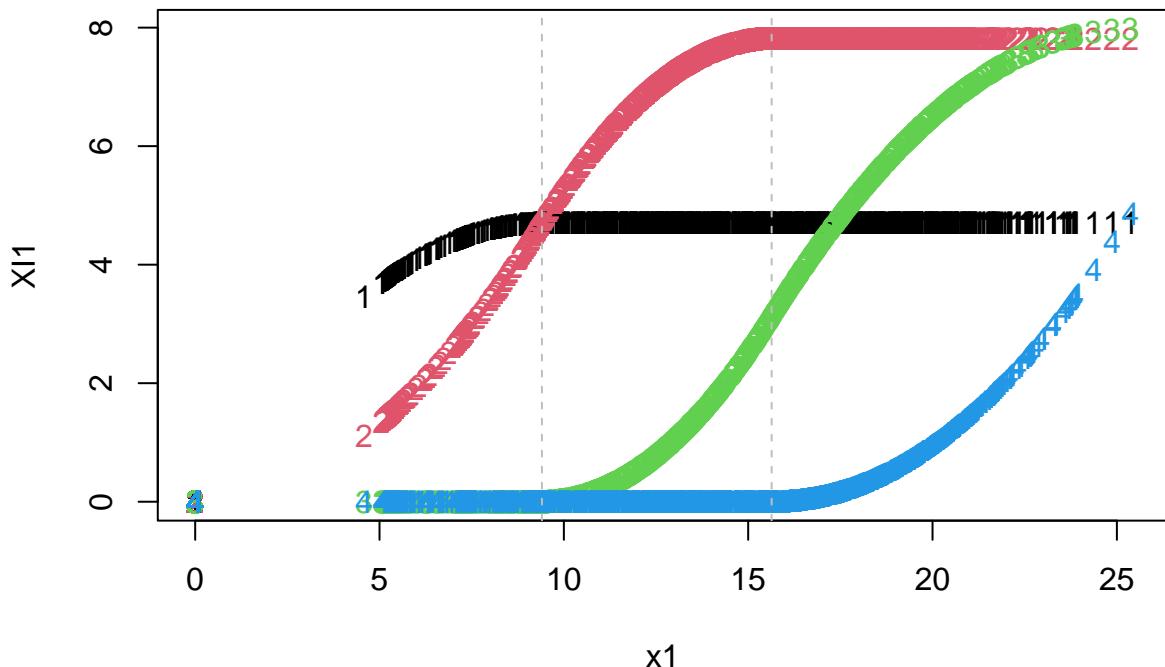


3. Generar la matriz diseño $XI1$ para los I-splines

$$f_1(x_1) = \sum_{j=1}^{h_1} \beta_{1j} I_{1j}(x_1)$$

$$I_{1j}(x_1) = \int_{x_0}^{x_1} B_{1j}(u) d_u$$

```
### ibs: integrated basis splines
### degree = 3 cubic splines
XI1 <- ibs(x1, knots = knots1, degree = 1, intercept = TRUE)
###XI1 = (XI1-mean(XI1))/sd(XI1)
matplot(x1, XI1)
abline(v = knots1, h = knots1, lty = 2, col = "gray")
```



4. Definir la penalización S_1 y S_2

La flexibilidad ajustada de f es controlada por K , a través de una penalización cuadrática de la forma:

$$\sum_j \lambda_j \beta^T S_j \beta$$

donde los S_j son matrices de coeficientes conocidos, y los λ_j son parámetros de suavizamiento estimados.

#Este es el código que produce la matriz de diferenciación.

#No es el óptimo, pero funciona.

#"k" es el número de b-splines y

#"d" el orden de la diferenciación.

#Adjunto el artículo donde discutimos esto (página 7).

```
diffMatrix = function(k, d = 2){
  if( (d<1) || (d %% 1 != 0) )stop("d must be a positive integer value");
  if( (k<1) || (k %% 1 != 0) )stop("k must be a positive integer value");
  if(d >= k)stop("d must be lower than k");
  out = diag(k);
  for(i in 1:d){
    out = diff(out);
  }
  return(out)
}
(D1 = diffMatrix(k=k1, d=2))
```

```
##      [,1] [,2] [,3] [,4]
## [1,]     1   -2     1     0
## [2,]     0     1   -2     1
```

```
(D2 = diffMatrix(k=k2, d=2))
```

```
##      [,1] [,2] [,3] [,4]
## [1,]     1   -2     1     0
## [2,]     0     1   -2     1
```

```
(S1 = t(D1) *% D1 + diag(1,k1)*10e-4)
```

```
##      [,1]     [,2]     [,3]     [,4]
## [1,] 1.001 -2.000  1.000  0.000
## [2,] -2.000  5.001 -4.000  1.000
## [3,]  1.000 -4.000  5.001 -2.000
## [4,]  0.000  1.000 -2.000  1.001
```

```
(S2 = t(D2) *% D2 + diag(1,k2)*10e-4)
```

```
##      [,1]     [,2]     [,3]     [,4]
## [1,] 1.001 -2.000  1.000  0.000
## [2,] -2.000  5.001 -4.000  1.000
## [3,]  1.000 -4.000  5.001 -2.000
## [4,]  0.000  1.000 -2.000  1.001
```

5. Lineal NO restricciones

5.1 Lineal fit without constraints:

5.2 LME: Lineal fit without constraints:

```
set.seed(123)
# mod.lme <- lme( y ~ x1 , random=~1/id )
mod.lme <- lme( y ~ x1 , random=list(id=~1) )
summary(mod.lme)

## Linear mixed-effects model fit by REML
##   Data: NULL
##       AIC      BIC    logLik
## 9776.121 9797.055 -4884.06
##
## Random effects:
##   Formula: ~1 | id
##             (Intercept) Residual
## StdDev:     8.473532 6.824656
##
## Fixed effects: y ~ x1
##                  Value Std.Error DF t-value p-value
## (Intercept) -0.4908194 0.6929494 1157 -0.708305 0.4789
## x1           0.6645373 0.0342989 1157 19.374878 0.0000
## Correlation:
##   (Intr)
## x1 -0.502
##
## Standardized Within-Group Residuals:
##   Min      Q1      Med      Q3      Max
## -4.39202484 -0.46543276 -0.01792227  0.51801631  4.26459889
##
## Number of Observations: 1387
## Number of Groups: 229

sigma(mod.lme)

## [1] 6.824656

set.seed(123)
mod.lme1 <- lme( y ~ x1 , random=~0+x1|id )
mod.lme1 <- lme( y ~ x1 , random=list(id=~0+x1) )
summary(mod.lme1)

## Linear mixed-effects model fit by REML
##   Data: NULL
##       AIC      BIC    logLik
## 9097.928 9118.862 -4544.964
##
## Random effects:
##   Formula: ~0 + x1 | id
##             x1 Residual
## StdDev: 1.273777 4.748012
##
```

```

## Fixed effects: y ~ x1
##                Value Std.Error DF   t-value p-value
## (Intercept) -0.8446225 0.2968668 1157 -2.845123 0.0045
## x1          0.7353480 0.0918708 1157  8.004154 0.0000
## Correlation:
##      (Intr)
## x1 -0.301
##
## Standardized Within-Group Residuals:
##       Min     Q1     Med     Q3     Max
## -5.6959071 -0.3607015  0.1205509  0.3598872  3.9137646
##
## Number of Observations: 1387
## Number of Groups: 229

sigma(mod.lme1)

## [1] 4.748012

# mod.lme3 <- lme( y ~ x1 , random=~x1/id )
mod.lme3 <- lme( y ~ x1 , random=list(id=~x1) )

## Error in lme.formula(y ~ x1, random = list(id = ~x1)): nlminb problem, convergence error code = 1
##   message = iteration limit reached without convergence (10)

# Error in lme.formula(y ~ x1, random = ~x1 / id) :
# nlminb problem, convergence error code = 1 message = iteration
# limit reached without convergence (10)

datos.lme <- list( y = y ,
                     n = length(y) , N = N , Ni = Ni,
                     x1 = x1 , id = id )
param.lme = c("b0","b1", "invtau2","tau2","tau", "invsig2","sig2","sigma")

fit.lme.non.reslope <- stan("jagam_9_aneur_lme_non_reslope.stan",
                             data=datos.lme,
                             chains=3,warmup=500,iter=1000,thin=2,cores=6 )

print(fit.lme.non.reslope, pars=param.lme, digits=5)

## Inference for Stan model: jagam_9_aneur_lme_non_reslope.
## 3 chains, each with iter=1000; warmup=500; thin=2;
## post-warmup draws per chain=250, total post-warmup draws=750.
##
##           mean se_mean    sd    2.5%    25%    50%    75%   97.5%
## b0      -0.86352 0.01219 0.28820 -1.44039 -1.05267 -0.85779 -0.66135 -0.33062
## b1       0.75458 0.01395 0.08488  0.58258  0.69883  0.75710  0.81077  0.92101
## invtau2  0.04417 0.00007 0.00179  0.04082  0.04290  0.04418  0.04540  0.04759
## tau2    22.67621 0.03807 0.92264 21.01414 22.02444 22.63652 23.30877 24.49930
## tau     4.76097 0.00399 0.09673  4.58412  4.69302  4.75779  4.82792  4.94968

```

```

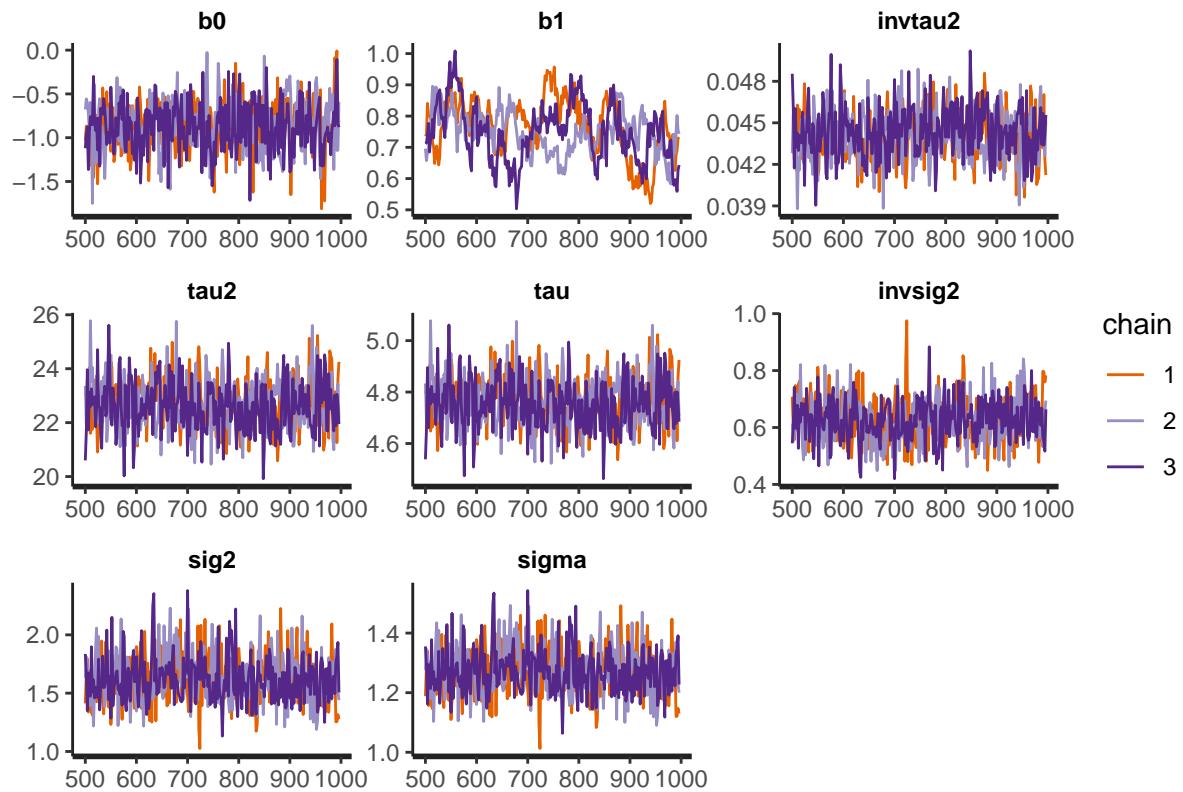
## invsig2  0.62446 0.00337 0.07751  0.48122  0.57091  0.62092  0.67642  0.79266
## sig2     1.62628 0.00881 0.20401  1.26157  1.47837  1.61052  1.75157  2.07805
## sigma    1.27279 0.00343 0.07937  1.12320  1.21588  1.26906  1.32347  1.44155
##          n_eff   Rhat
## b0        559 1.00039
## b1        37  1.03563
## invtau2   591 1.00259
## tau2      587 1.00226
## tau       588 1.00235
## invsig2   530 0.99946
## sig2      536 0.99961
## sigma     534 0.99956
##
## Samples were drawn using NUTS(diag_e) at Tue Jan  9 14:21:33 2024.
## For each parameter, n_eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).

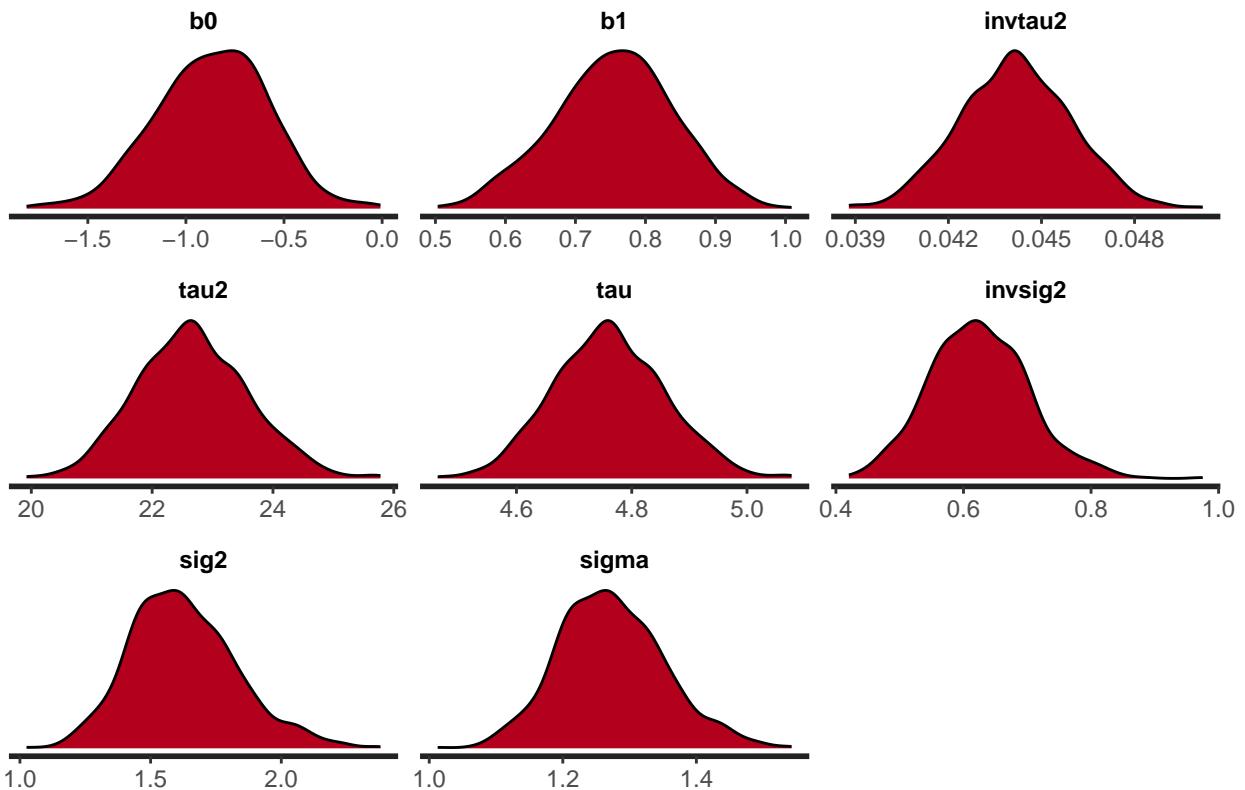
```

```

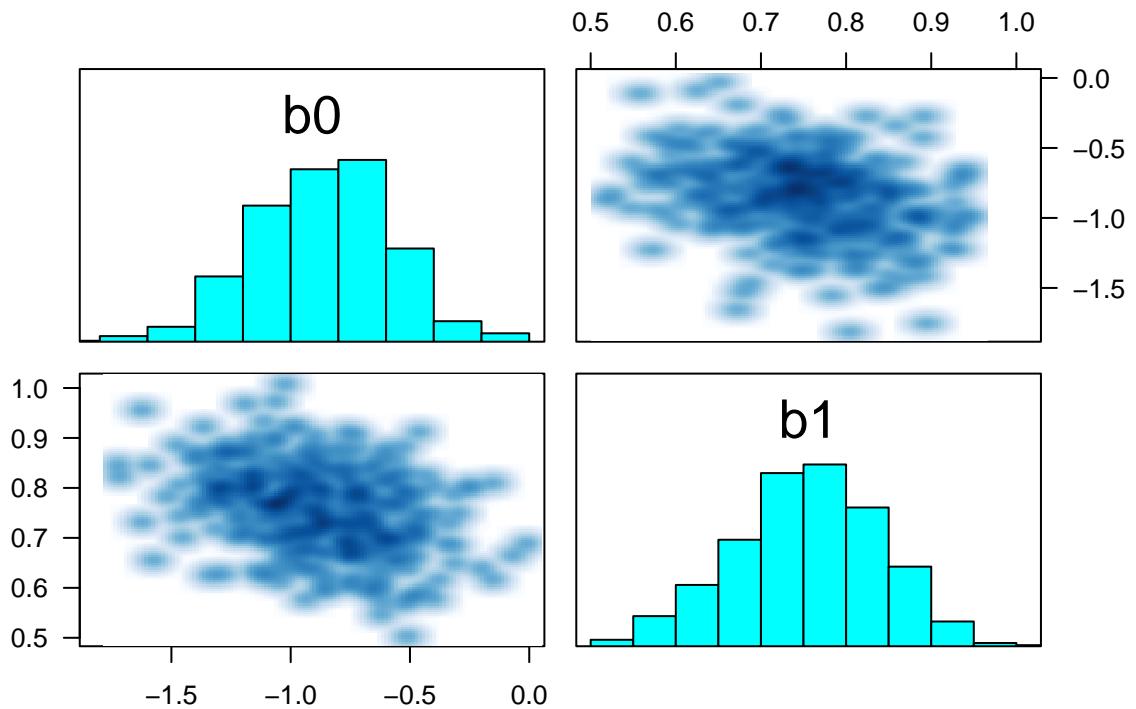
stan_trace(fit.lme.non.reslope,pars=param.lme)
stan_dens(fit.lme.non.reslope,pars=param.lme)

```

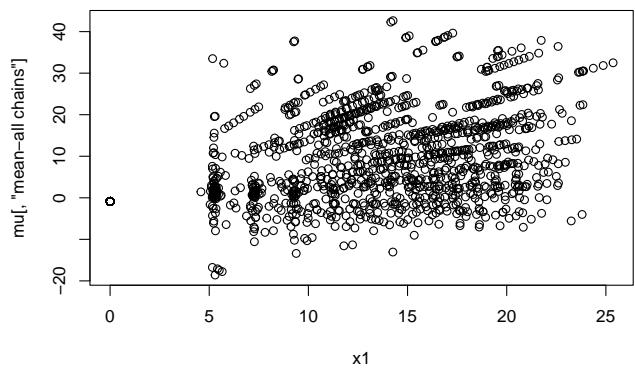




```
pairs(fit.lme.non.reslope, pars = c("b0", "b1"), las = 1)
```



```
mu=get_posterior_mean(fit.lme.non.reslope, "mu")
plot(x1,mu[, "mean-all chains"])
```



6. Lineal creciente

6.1. Lineal creciente

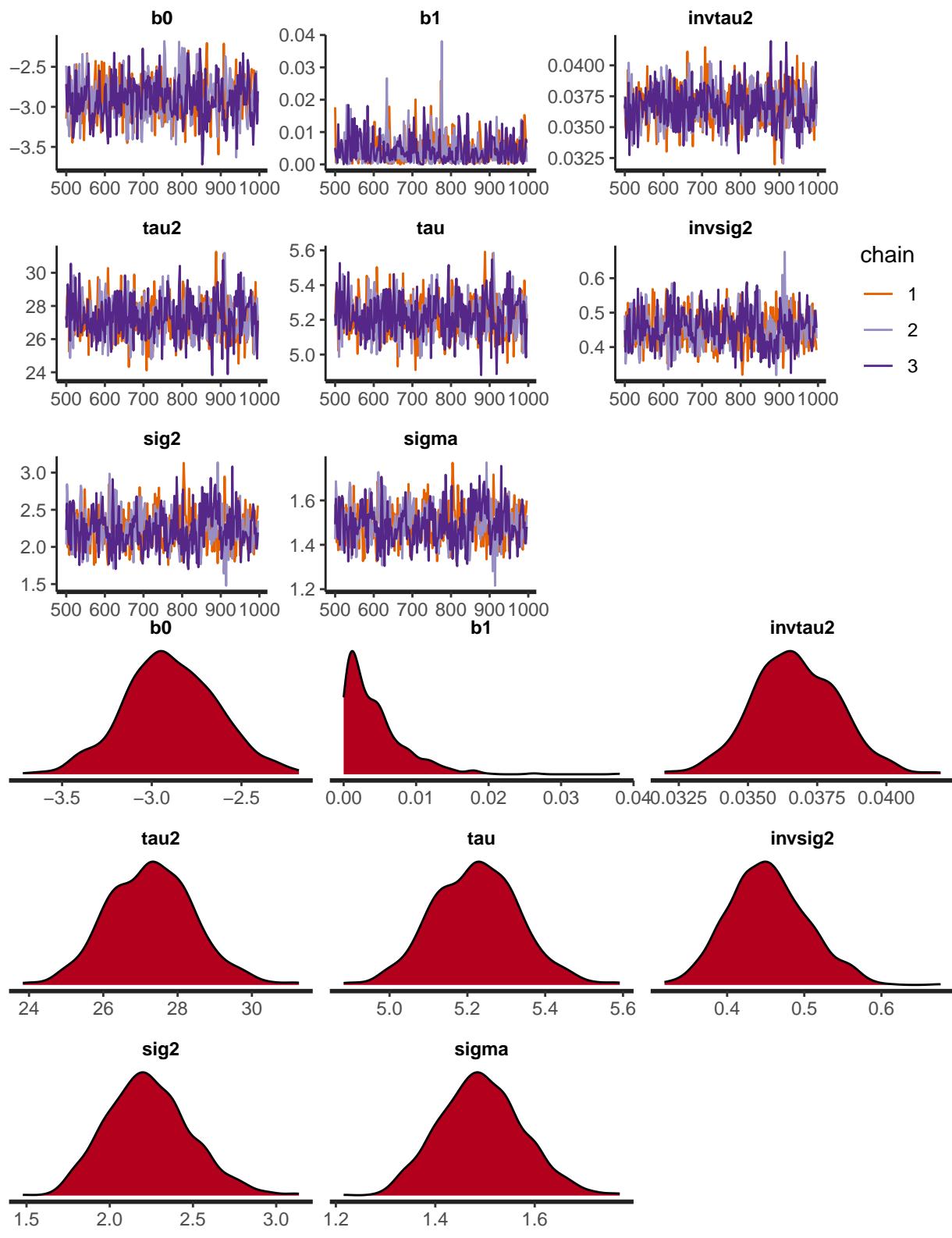
6.2. LME: Lineal creciente

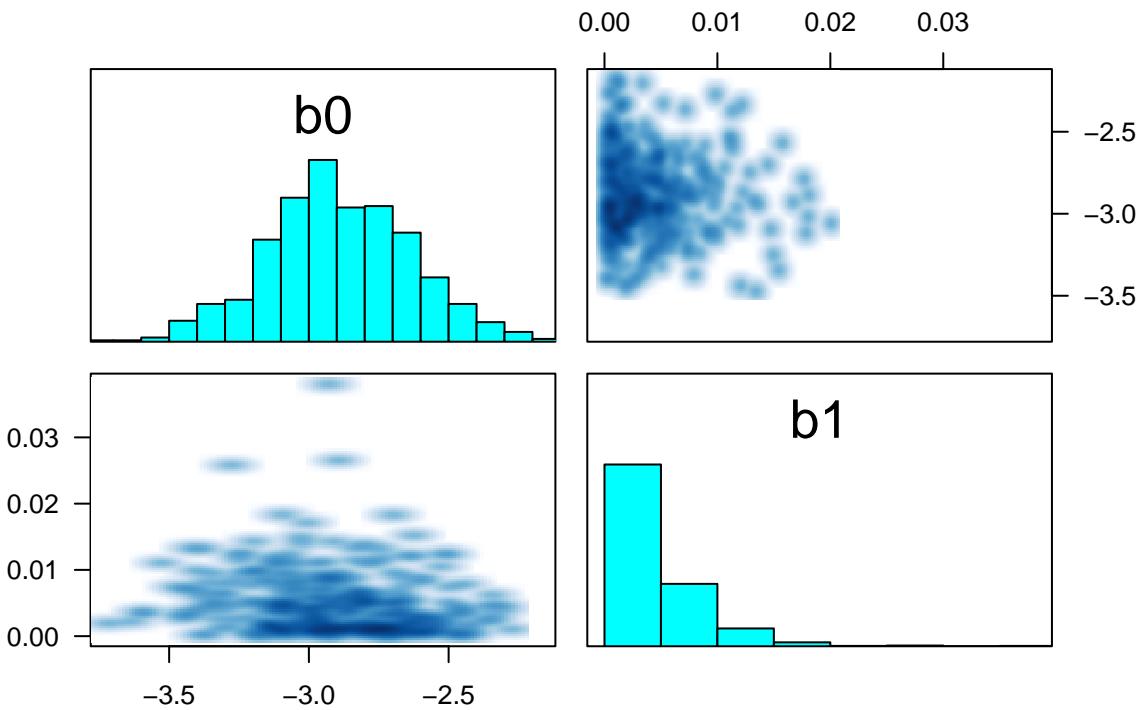
```
fit.lme.incr.reslope <- stan("jagam_9_aneur_lme_incr_reslope.stan",
  data=datos.lme,
  chains=3,warmup=500,iter=1000,thin=2,cores=6 )

print(fit.lme.incr.reslope, pars=param.lme, digits=5)

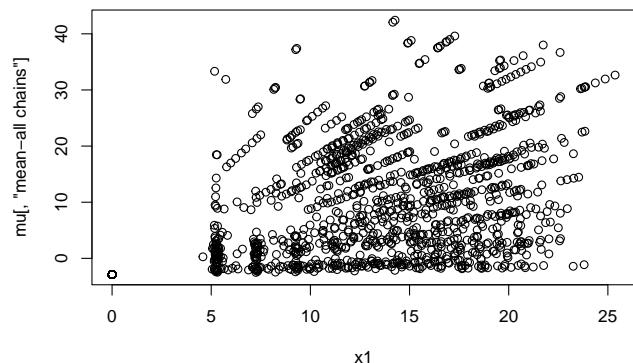
## Inference for Stan model: jagam_9_aneur_lme_incr_reslope.
## 3 chains, each with iter=1000; warmup=500; thin=2;
## post-warmup draws per chain=250, total post-warmup draws=750.
##
##          mean se_mean     sd    2.5%    25%    50%    75%   97.5%
## b0      -2.89304 0.01114 0.25649 -3.40123 -3.06703 -2.90904 -2.71514 -2.35359
## b1       0.00428 0.00016 0.00414  0.00017  0.00124  0.00315  0.00593  0.01437
## invtau2  0.03671 0.00006 0.00156  0.03365  0.03564  0.03663  0.03779  0.03989
## tau2     27.28711 0.04603 1.16124 25.06678 26.46436 27.29714 28.05491 29.71447
## tau      5.22253 0.00439 0.11104  5.00667  5.14435  5.22467  5.29669  5.45110
## invsig2  0.45313 0.00222 0.05075  0.36050  0.41899  0.45030  0.48579  0.55917
## sig2     2.23464 0.01118 0.25145  1.78837  2.05851  2.22076  2.38670  2.77393
## sigma    1.49253 0.00371 0.08368  1.33730  1.43475  1.49022  1.54490  1.66551
##          n_eff    Rhat
## b0        531 1.00189
## b1        661 0.99830
## invtau2  649 0.99775
## tau2     636 0.99787
## tau       639 0.99784
## invsig2  525 1.00138
## sig2     505 1.00136
## sigma    510 1.00136
##
## Samples were drawn using NUTS(diag_e) at Tue Jan  9 14:22:20 2024.
## For each parameter, n_eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).

stan_trace(fit.lme.incr.reslope,pars=param.lme)
stan_dens(fit.lme.incr.reslope,pars=param.lme)
```





```
mu=get_posterior_mean(fit.lme.incr.reslope,"mu")
plot(x1,mu[, "mean-all chains"])
```



7. Spline NO restricciones

7.1 For a spline-based fit without constraints:

7.2 LME: For a spline-based fit without constraints:

```
set.seed(123)
mod.lme.s <- lme( y ~ XI1 , random=list(id=~1) )
summary(mod.lme.s)

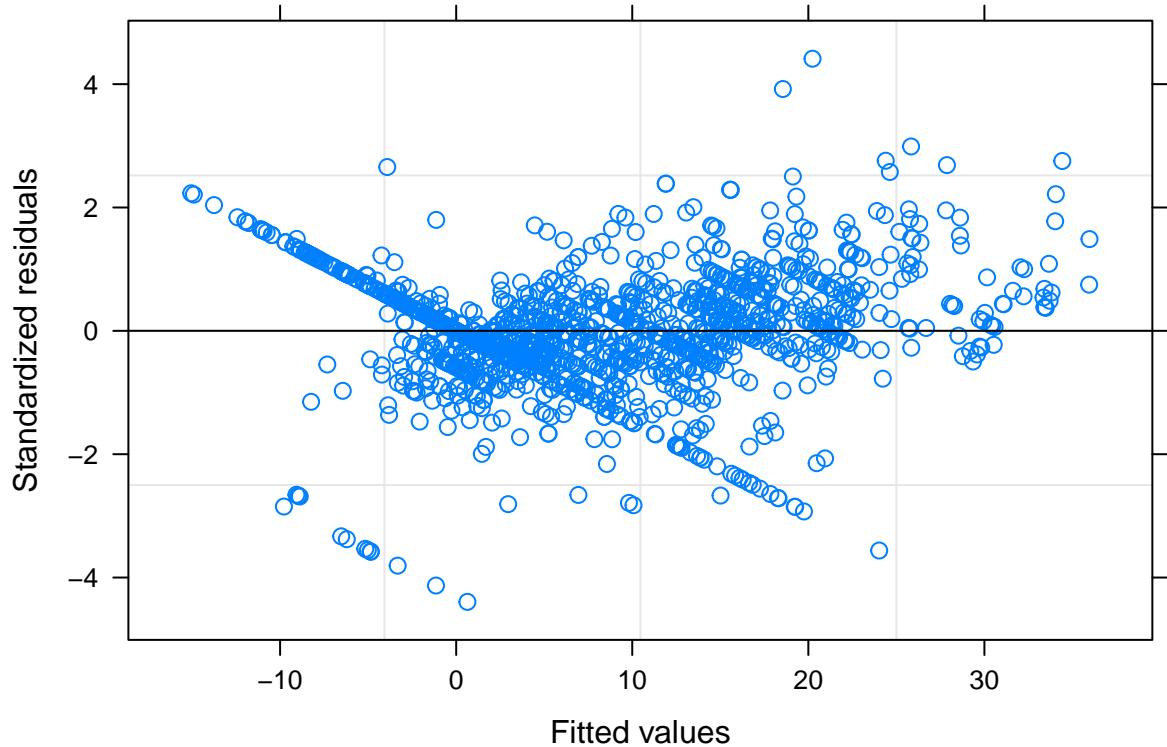
## Linear mixed-effects model fit by REML
## Data: NULL
##      AIC      BIC      logLik
## 9753.189 9789.808 -4869.595
##
## Random effects:
##   Formula: ~1 | id
##             (Intercept) Residual
## StdDev:     8.51601 6.744787
##
## Fixed effects: y ~ XI1
##                  Value Std.Error DF t-value p-value
## (Intercept) 0.045604 0.7173634 1154 0.063571 0.9493
## XI11        0.147201 0.2661071 1154 0.553166 0.5803
## XI12        0.970901 0.2158582 1154 4.497864 0.0000
## XI13        0.253511 0.2281802 1154 1.111010 0.2668
## XI14        3.889209 0.6769512 1154 5.745184 0.0000
##
## Correlation:
##   (Intr) XI11  XI12  XI13
## XI11 -0.264
## XI12  0.039 -0.860
## XI13 -0.013  0.300 -0.605
## XI14  0.007 -0.160  0.290 -0.662
##
## Standardized Within-Group Residuals:
##      Min       Q1       Med       Q3       Max
## -4.394201884 -0.453908702 -0.003561512  0.499733599  4.412553978
##
## Number of Observations: 1387
## Number of Groups: 229

sigma(mod.lme.s)

## [1] 6.744787

plot(mod.lme.s)
anova(mod.lme,mod.lme.s)

##          Model df      AIC      BIC      logLik   Test  L.Ratio p-value
## mod.lme       1  4 9776.121 9797.055 -4884.060
## mod.lme.s     2  7 9753.189 9789.808 -4869.595 1 vs 2 28.93154 <.0001
```



```
set.seed(123)
mod.lme.s1 <- lme( y ~ XI1 , random=list(id=~0+x1) )
summary(mod.lme.s1)
```

```
## Linear mixed-effects model fit by REML
##   Data: NULL
##   AIC      BIC    logLik
## 8950.966 8987.585 -4468.483
##
## Random effects:
##   Formula: ~0 + x1 | id
##             x1 Residual
## StdDev: 1.401535 4.375082
##
## Fixed effects: y ~ XI1
##                  Value Std.Error DF t-value p-value
## (Intercept) 0.021674 0.2886553 1154 0.075084 0.9402
## XI11       -0.217886 0.2073866 1154 -1.050629 0.2936
## XI12        1.053298 0.1960705 1154  5.372035 0.0000
## XI13        1.768322 0.2216114 1154  7.979381 0.0000
## XI14        4.374687 0.5119063 1154  8.545874 0.0000
##
## Correlation:
##   (Intr) XI11  XI12  XI13
## XI11 -0.353
## XI12  0.041 -0.466
## XI13 -0.015  0.297 -0.121
## XI14  0.007 -0.067  0.310 -0.346
##
## Standardized Within-Group Residuals:
```

```

##          Min           Q1           Med           Q3           Max
## -5.827242762 -0.328706726 -0.004953857  0.373749636  4.595933776
##
## Number of Observations: 1387
## Number of Groups: 229

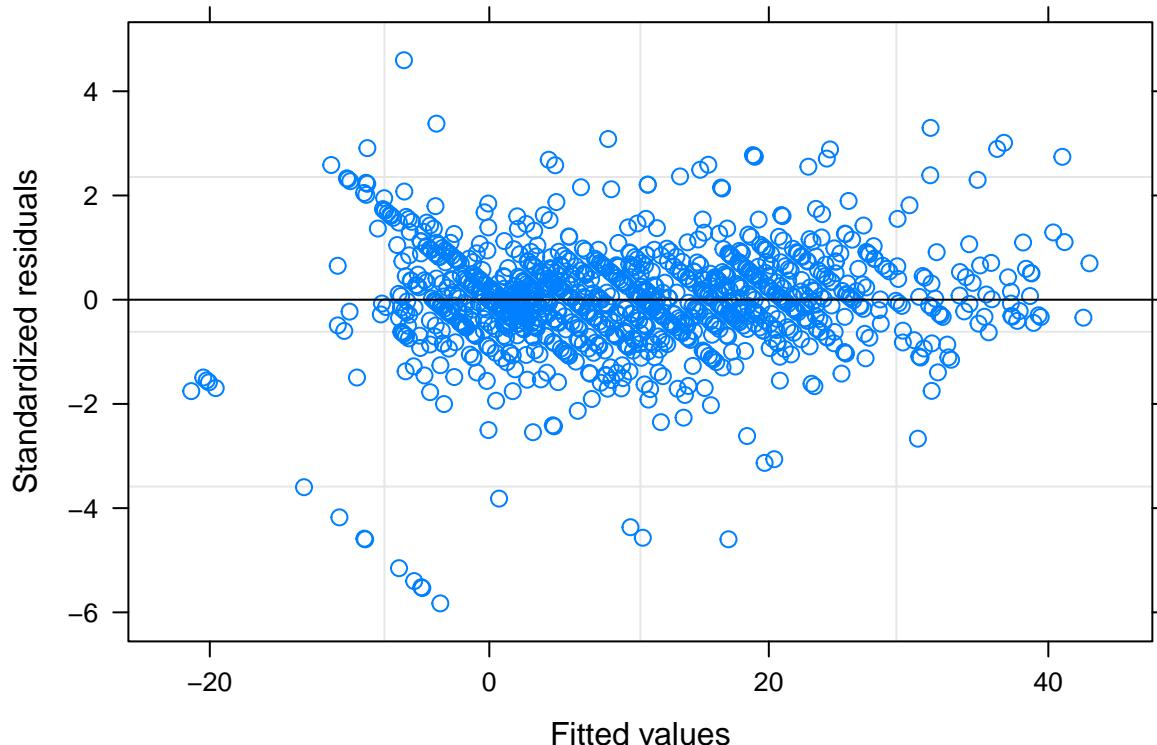
sigma(mod.lme.s1)

## [1] 4.375082

plot(mod.lme.s1)
anova(mod.lme,mod.lme.s)

```

##	Model	df	AIC	BIC	logLik	Test	L.Ratio	p-value
##	mod.lme	1	4	9776.121	9797.055	-4884.060		
##	mod.lme.s	2	7	9753.189	9789.808	-4869.595	1 vs 2	28.93154 <.0001



```

mod.gamm <- gamm( y ~ s(x1, k=k1) , random=list(id=~1) )
summary(mod.gamm$gam)

```

```

##
## Family: gaussian
## Link function: identity
##
## Formula:
## y ~ s(x1, k = k1)
##

```

```

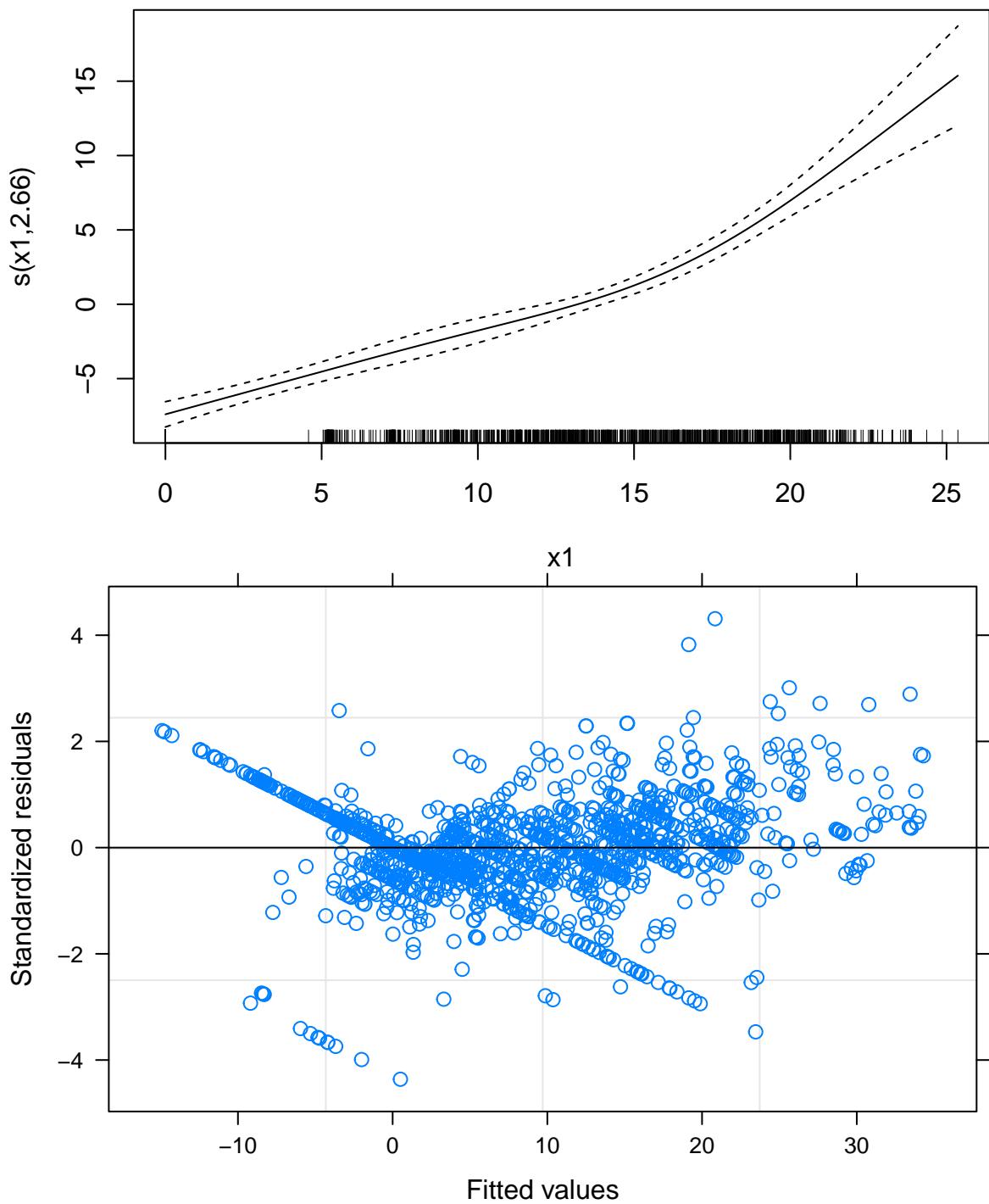
## Parametric coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 7.2836    0.6059   12.02 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Approximate significance of smooth terms:
##          edf Ref.df F p-value
## s(x1) 2.659  2.659 147 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## R-sq.(adj) =  0.16
## Scale est. = 45.747    n = 1387

summary(mod.gamm$lme)

## Linear mixed-effects model fit by maximum likelihood
## Data: strip.offset(mf)
##      AIC      BIC logLik
## 9765.232 9791.406 -4877.616
##
## Random effects:
## Formula: ~Xr - 1 | g
## Structure: pdIdnot
##           Xr1     Xr2
## StdDev: 11.6555 11.6555
##
## Formula: ~1 | id %in% g
## (Intercept) Residual
## StdDev: 8.542761 6.763645
##
## Fixed effects: y.0 ~ X - 1
##                 Value Std.Error DF t-value p-value
## X(Intercept) 7.283611 0.6060915 1157 12.017345     0
## Xs(x1)Fx1    7.373808 1.0052097 1157  7.335591     0
## Correlation:
##           X(Int)
## Xs(x1)Fx1  0
##
## Standardized Within-Group Residuals:
##      Min        Q1        Med        Q3        Max
## -4.361985592 -0.455510067 -0.004707745  0.493794967  4.310914825
##
## Number of Observations: 1387
## Number of Groups:
##      g id %in% g
##      1      229

plot(mod.gamm$gam)
plot(mod.gamm$lme)

```



```
mod.gamm1 <- gamm( y ~ s(x1, k=k1) , random=list(id=~0+x1) )
summary(mod.gamm1$gam)
```

```
##
## Family: gaussian
## Link function: identity
##
## Formula:
```

```

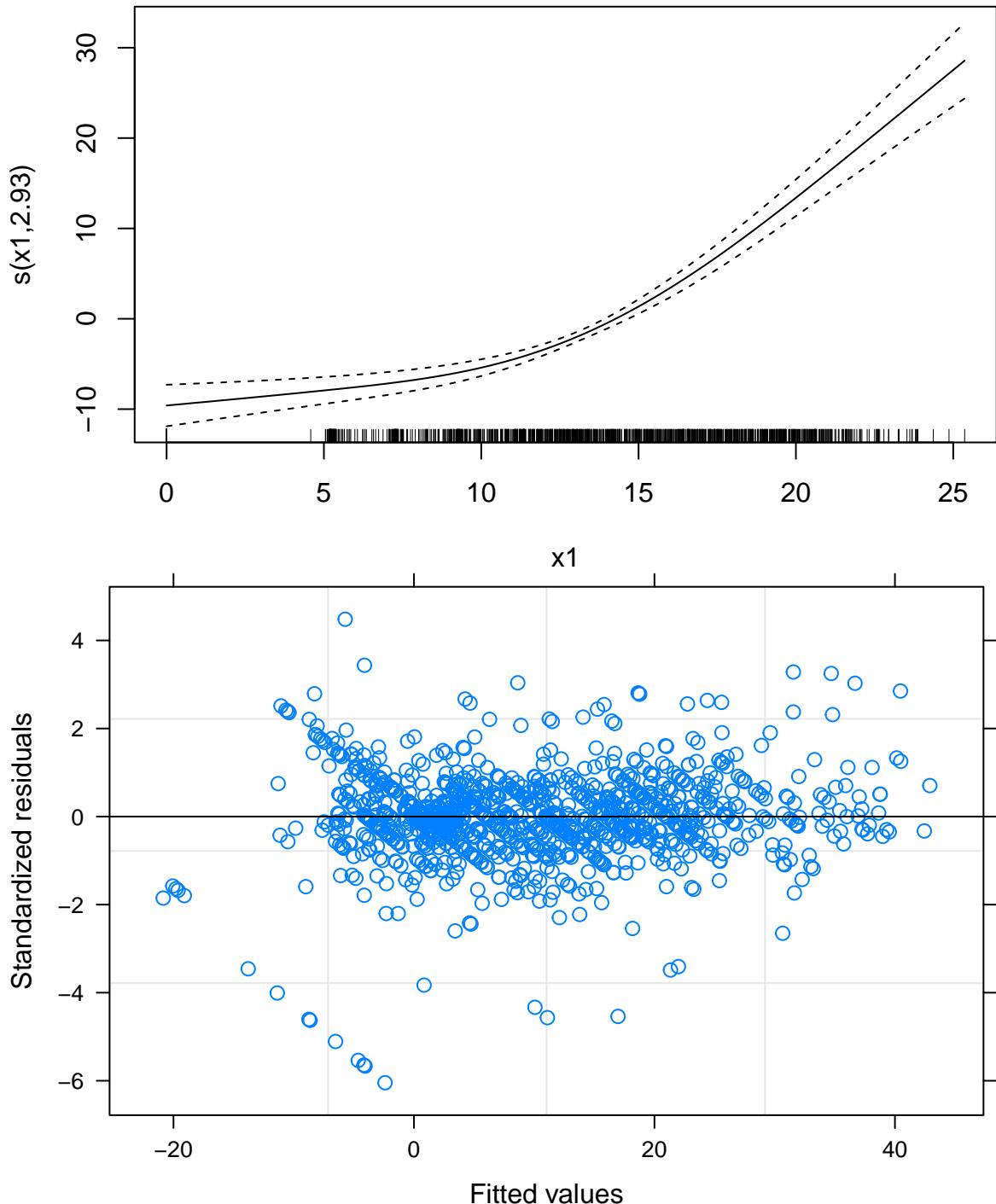
## y ~ s(x1, k = k1)
##
## Parametric coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 9.485      1.118    8.485 <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Approximate significance of smooth terms:
##          edf Ref.df   F p-value
## s(x1) 2.928 2.928 69.47 <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## R-sq.(adj) = 0.0128
## Scale est. = 19.36     n = 1387

summary(mod.gamm1$lme)

## Linear mixed-effects model fit by maximum likelihood
## Data: strip.offset(mf)
##       AIC     BIC   logLik
## 8964.745 8990.919 -4477.372
##
## Random effects:
## Formula: ~Xr - 1 | g
## Structure: pdIdnot
##           Xr1     Xr2
## StdDev: 22.62108 22.62108
##
## Formula: ~0 + x1 | id %in% g
##           x1 Residual
## StdDev: 1.380861 4.40003
##
## Fixed effects: y.0 ~ X - 1
##                 Value Std.Error DF t-value p-value
## X(Intercept) 9.485195 1.118241 1157 8.482244     0
## Xs(x1)Fx1   10.777078 1.074914 1157 10.025993     0
## Correlation:
##           X(Int)
## Xs(x1)Fx1 0.573
##
## Standardized Within-Group Residuals:
##      Min        Q1        Med        Q3        Max
## -6.04472640 -0.32800007  0.02577703  0.36868738  4.48138337
##
## Number of Observations: 1387
## Number of Groups:
##           g id %in% g
##           1      229

plot(mod.gamm1$gam)
plot(mod.gamm1$lme)

```



```
mod.gamm.ar <- gamm( y ~ s(x1, k=k1) , random=list(id=~1) ,
correlation=corAR1() )
summary(mod.gamm.ar$gam)
```

```
##
## Family: gaussian
## Link function: identity
##
```

```

## Formula:
## y ~ s(x1, k = k1)
##
## Parametric coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 7.7723     0.5601   13.88  <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Approximate significance of smooth terms:
##          edf Ref.df    F p-value
## s(x1)     1     1 263.2  <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## R-sq.(adj) =  0.16
## Scale est. = 90.909    n = 1387

```

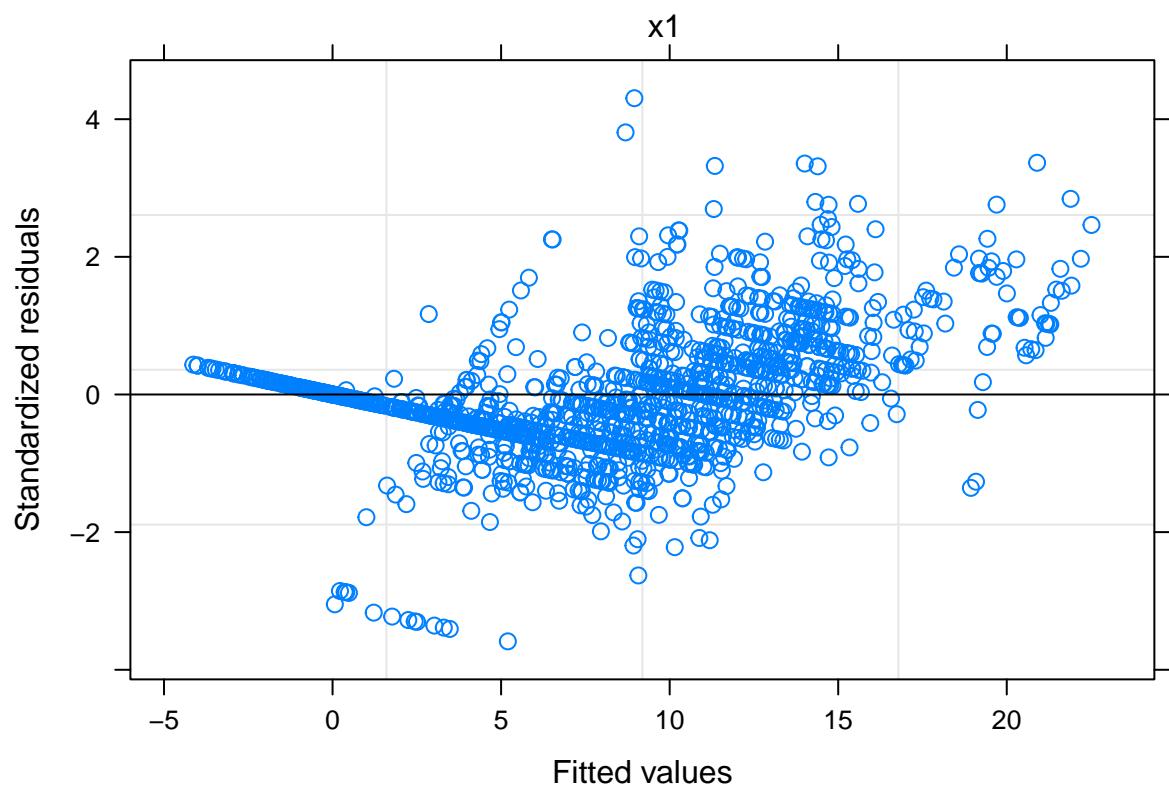
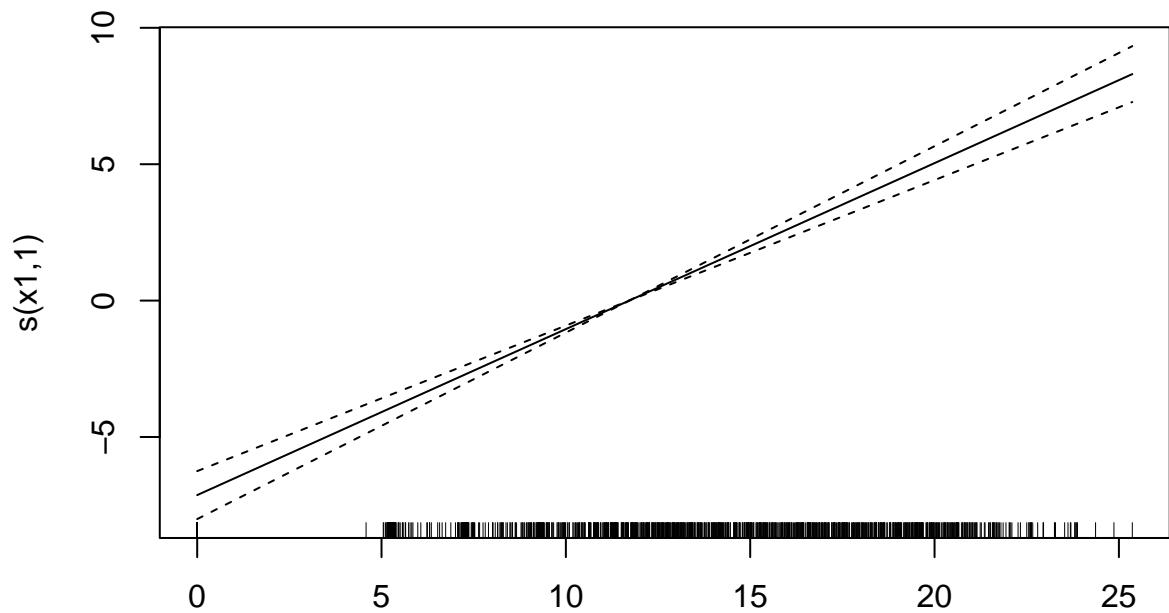
```
summary(mod.gamm.ar$lme)
```

```

## Linear mixed-effects model fit by maximum likelihood
## Data: strip.offset(mf)
##      AIC      BIC      logLik
## 9587.898 9619.308 -4787.949
##
## Random effects:
## Formula: ~Xr - 1 | g
## Structure: pdIdnot
##           Xr1         Xr2
## StdDev: 0.001703091 0.001703091
##
## Formula: ~1 | id %in% g
##          (Intercept) Residual
## StdDev: 4.724431 9.534615
##
## Correlation Structure: AR(1)
## Formula: ~1 | g/id
## Parameter estimate(s):
##       Phi
## 0.6751387
## Fixed effects: y.0 ~ X - 1
##                 Value Std.Error DF t-value p-value
## X(Intercept) 7.772282 0.5603182 1157 13.87119     0
## Xs(x1)Fx1    4.124876 0.2543414 1157 16.21787     0
## Correlation:
##       X(Int)
## Xs(x1)Fx1 0.153
##
## Standardized Within-Group Residuals:
##       Min        Q1        Med        Q3        Max
## -3.58692289 -0.53267218 -0.08303584  0.40754736  4.30503270
##
## Number of Observations: 1387
## Number of Groups:
```

```
##          g id %in% g
## 1           229
```

```
plot(mod.gamm.ar$gam)
plot(mod.gamm.ar$lme)
```



```

mod.gamm.ar1 <- gamm( y ~ s(x1, k=k1) , random=list(id=~0+x1) ,
                      correlation=corAR1() )
summary(mod.gamm.ar1$gam)

```

```

##
## Family: gaussian
## Link function: identity
##
## Formula:
## y ~ s(x1, k = k1)
##
## Parametric coefficients:
##             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 9.379     1.112    8.432  <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Approximate significance of smooth terms:
##          edf Ref.df   F p-value
## s(x1) 2.907 2.907 56.46  <2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## R-sq.(adj) = 0.0301
## Scale est. = 20.205 n = 1387

```

```
summary(mod.gamm.ar1$lme)
```

```

## Linear mixed-effects model fit by maximum likelihood
## Data: strip.offset(mf)
##      AIC      BIC logLik
## 8944.268 8975.678 -4466.134
##
## Random effects:
## Formula: ~Xr - 1 | g
## Structure: pdIdnot
##           Xr1      Xr2
## StdDev: 21.58851 21.58851
##
## Formula: ~0 + x1 | id %in% g
##           x1 Residual
## StdDev: 1.366815 4.495042
##
## Correlation Structure: AR(1)
## Formula: ~1 | g/id
## Parameter estimate(s):
##           Phi
## 0.1527438
## Fixed effects: y.0 ~ X - 1
##                 Value Std.Error DF t-value p-value
## X(Intercept) 9.379081 1.112715 1157 8.429006 0
## Xs(x1)Fx1   10.584722 1.134490 1157 9.329937 0

```

```

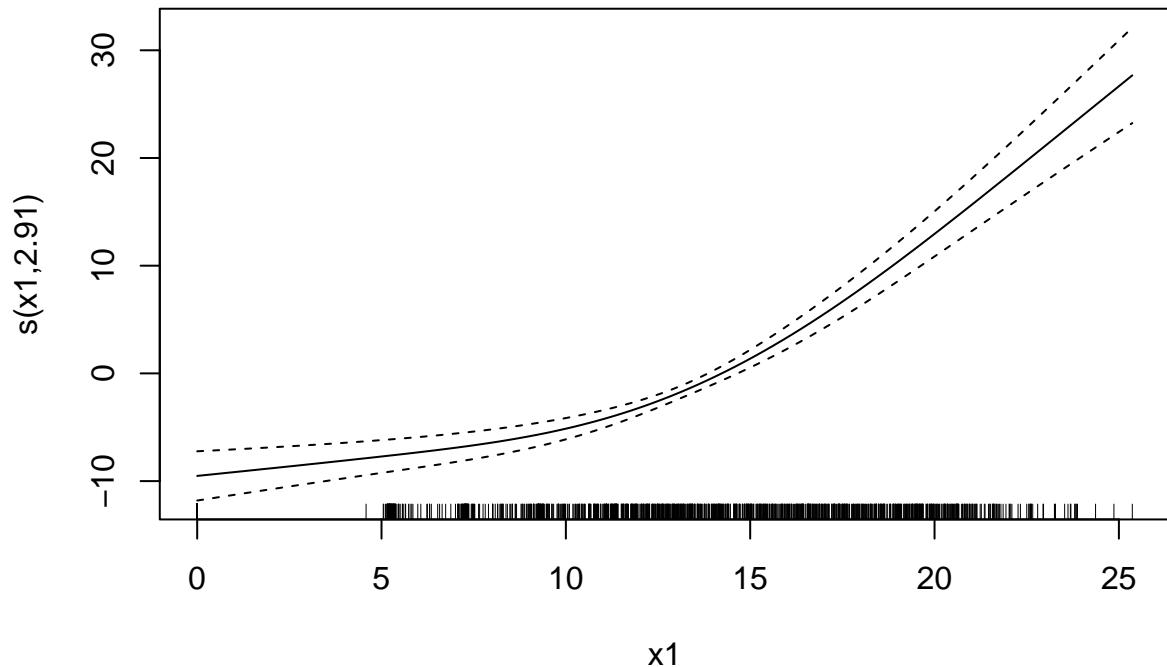
## Correlation:
##          X(Int)
## Xs(x1)Fx1 0.535
##
## Standardized Within-Group Residuals:
##      Min       Q1       Med       Q3      Max
## -5.91253795 -0.33640243  0.03088513  0.35053651  4.28869217
##
## Number of Observations: 1387
## Number of Groups:
##      g id %in% g
##      1     229

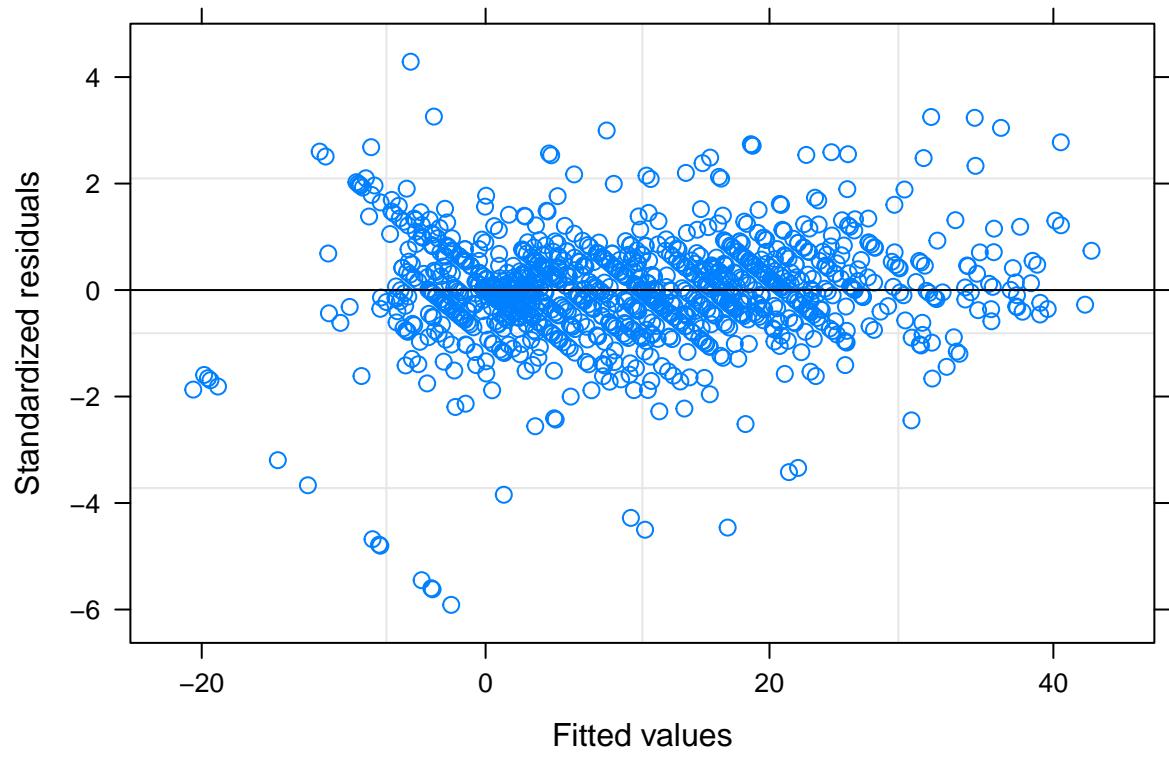
```

```

plot(mod.gamm.ar1$gam)
plot(mod.gamm.ar1$lme)

```





```

datos.lme.add.non <- list( y = y ,
                           id = id ,
                           n = length(y) ,
                           N = N , Ni = Ni ,
                           k1=k1 ,
                           XI1 = XI1 ,
                           x1 = x1 ,
                           zero = rep(0,1+k1) ,
                           S1=S1 )
inits.lme.add.non <- function(){   list(
  "b0" = rnorm(1,0,0.1) ,
  "b1" = rnorm(k1,0,0.1) ,
  "invtau2" = rgamma(1,1,1) ,
  "lambda" = rgamma(1,1,1) ,
  "invsig2" = rgamma(1,1,1) ,
  "bre0" = rnorm(N,0,0.1)
) }
param.lme.add = c("b0","b1", "invtau2","tau2","tau", "lambda","rho", "invsig2","sig2","sigma")

fit.lme.add.non.reslope <- stan("jagam_9_aneur_lme_add_non_reslope.stan",
                                 data=datos.lme.add.non,
                                 chains=3,warmup=500,iter=1000,thin=2,cores=6,
                                 init= inits.lme.add.non)

print(fit.lme.add.non.reslope, pars=param.lme.add, digits=5)

## Inference for Stan model: jagam_9_aneur_lme_add_non_reslope.
## 3 chains, each with iter=1000; warmup=500; thin=2;
## post-warmup draws per chain=250, total post-warmup draws=750.
##
##           mean    se_mean      sd     2.5%     25%     50%     75%
## b0       0.00645  0.01116  0.28193 -0.55818 -0.18017 -0.00824  0.20660
## b1[1]   -0.17681  0.01082  0.19754 -0.57032 -0.31189 -0.17659 -0.03990
## b1[2]    0.93244  0.01723  0.18581  0.56522  0.79819  0.94173  1.06738
## b1[3]    1.80348  0.01905  0.20723  1.36371  1.67998  1.80463  1.94244
## b1[4]    3.62571  0.02522  0.52394  2.63619  3.25662  3.61182  3.96627
## invtau2  0.05191  0.00010  0.00224  0.04776  0.05035  0.05184  0.05328
## tau2    19.30049  0.03565  0.82697  17.70262 18.76798 19.28854 19.86125
## tau     4.39222  0.00406  0.09420  4.20745  4.33220  4.39187  4.45660
## lambda  1182.71253 16.54124 353.24060 646.72960 931.38378 1137.21453 1368.79723
## rho     7.03351  0.01338  0.28898  6.47193  6.83667  7.03633  7.22169
## invsig2  0.51525  0.00229  0.05767  0.41268  0.47521  0.51246  0.54996
## sig2    1.96515  0.00885  0.22103  1.55847  1.81831  1.95136  2.10433
## sigma   1.39965  0.00313  0.07835  1.24839  1.34845  1.39691  1.45063
##             97.5% n_eff    Rhat
## b0       0.57039  638  0.99819
## b1[1]   0.20983  333  1.00530
## b1[2]   1.26549  116  1.01307
## b1[3]   2.18503  118  1.02319
## b1[4]   4.70298  432  0.99964
## invtau2  0.05649  537  1.00186
## tau2    20.93845 538  1.00134
## tau     4.57585  538  1.00147

```

```

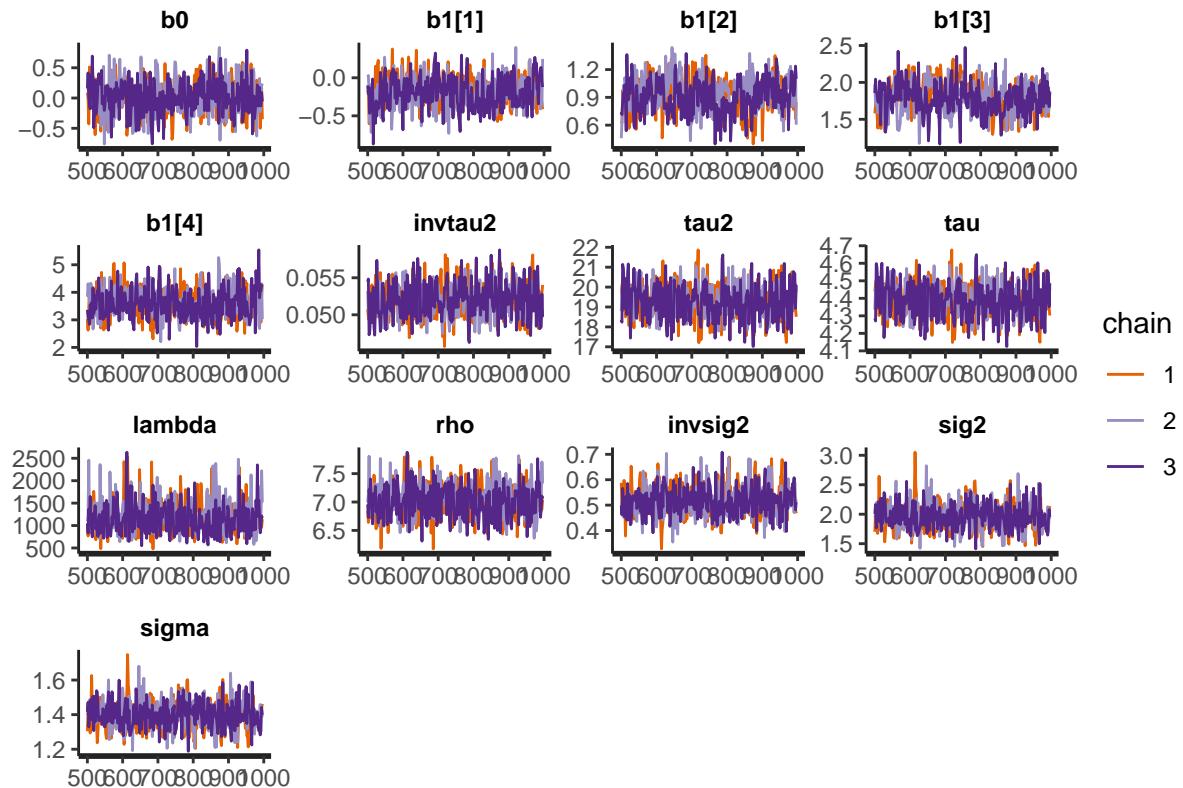
## lambda 2064.98819 456 1.00403
## rho      7.63287 466 1.00500
## invsig2   0.64166 634 0.99920
## sig2     2.42320 623 0.99921
## sigma    1.55666 627 0.99923
##
## Samples were drawn using NUTS(diag_e) at Tue Jan  9 14:24:04 2024.
## For each parameter, n_eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).

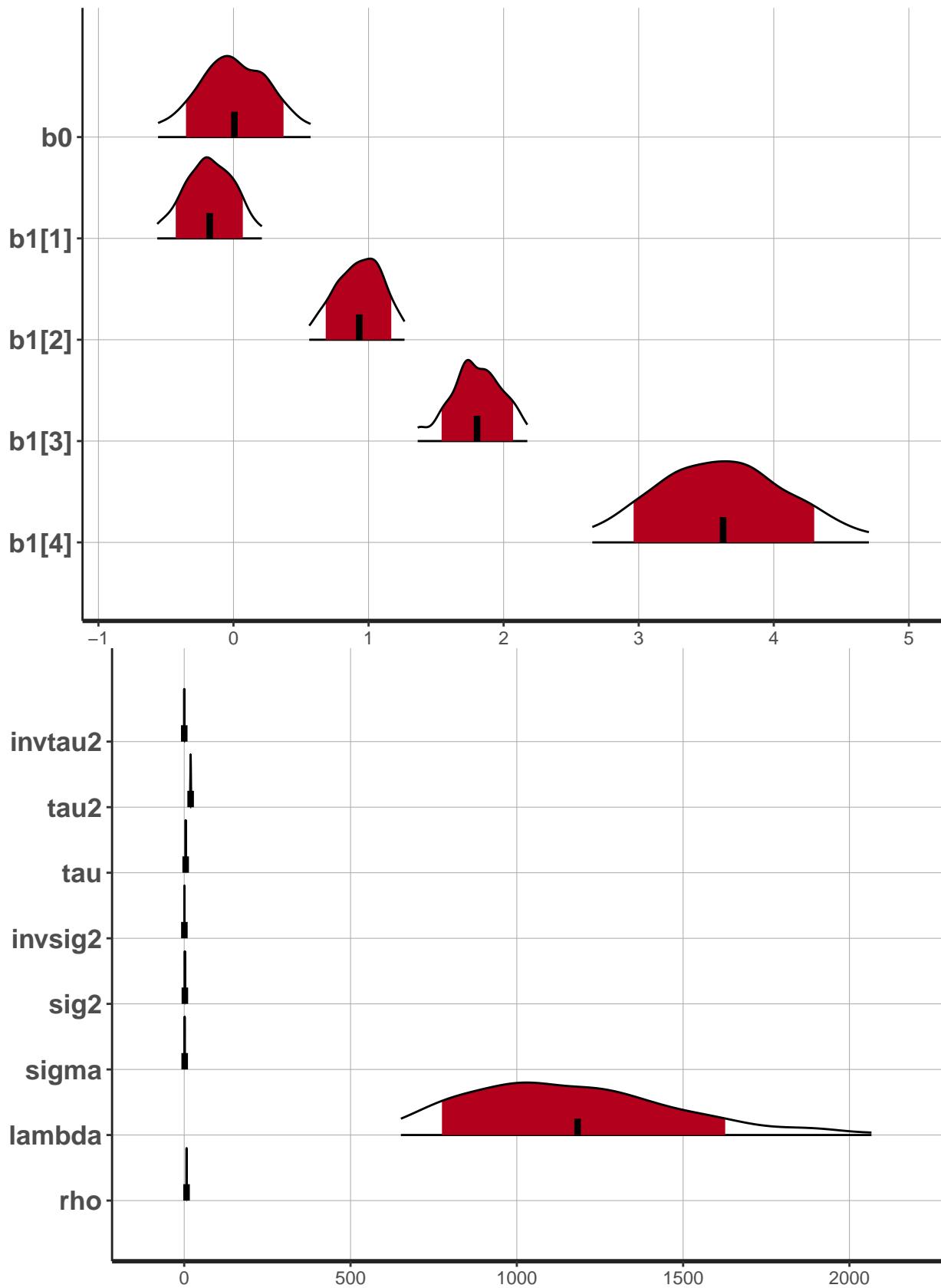
```

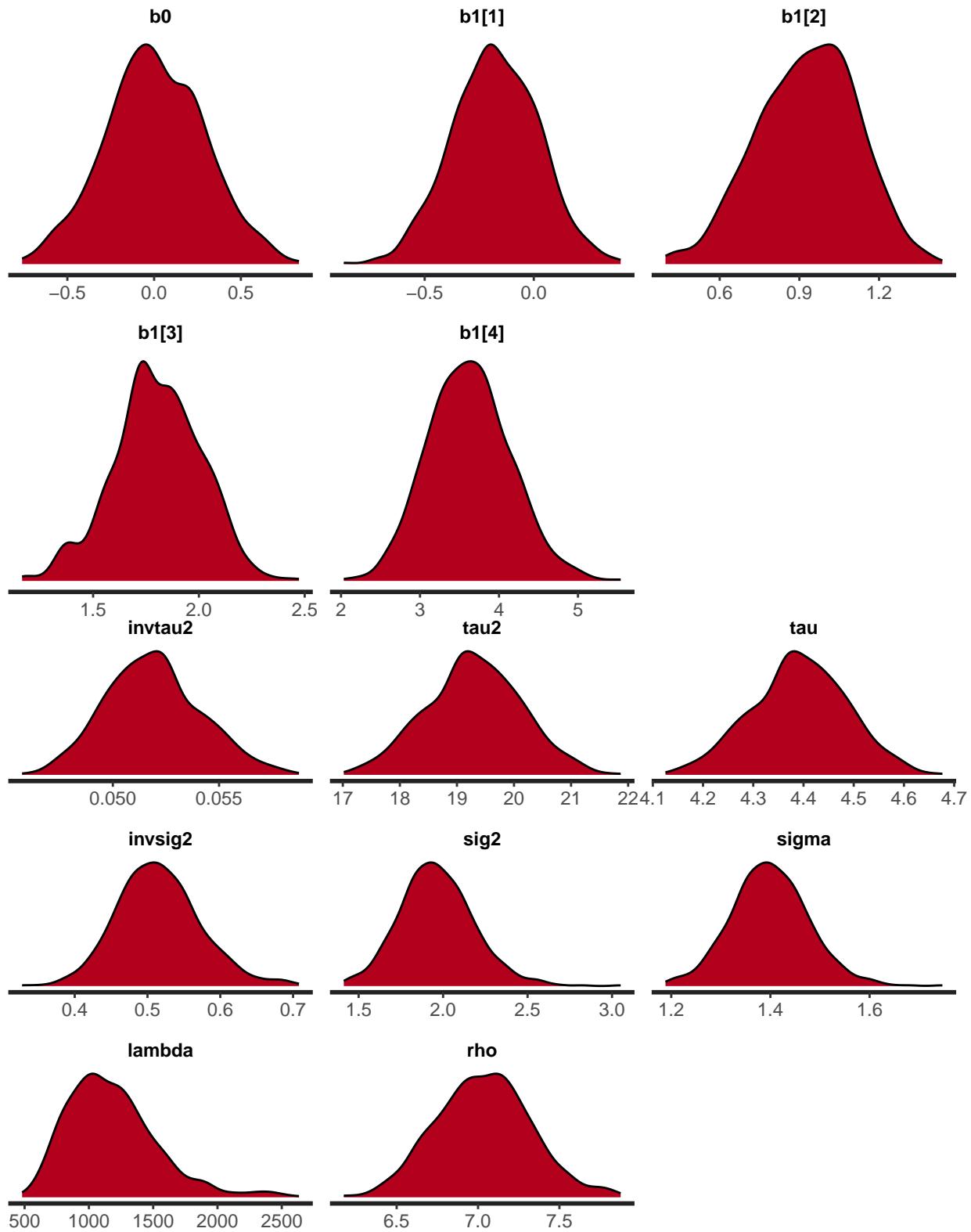
```

stan_trace(fit.lme.add.non.reslope, pars=param.lme.add)
stan_plot(fit.lme.add.non.reslope, pars=c("b0", "b1"), point_est = "mean", show_density = TRUE)
stan_plot(fit.lme.add.non.reslope, pars=c("invtau2", "tau2", "tau", "invsig2", "sig2", "sigma", "lambda", "rho"))
stan_dens(fit.lme.add.non.reslope, pars=c("b0", "b1"))
stan_dens(fit.lme.add.non.reslope, pars=c("invtau2", "tau2", "tau", "invsig2", "sig2", "sigma", "lambda", "rho"))

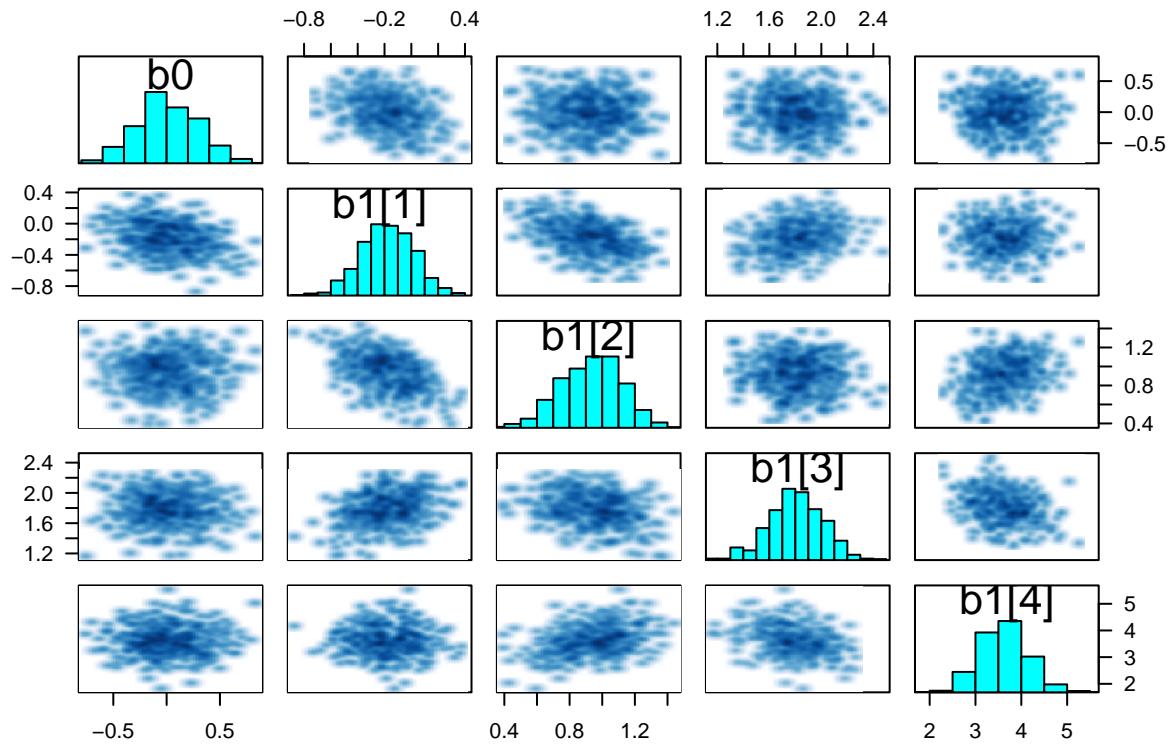
```



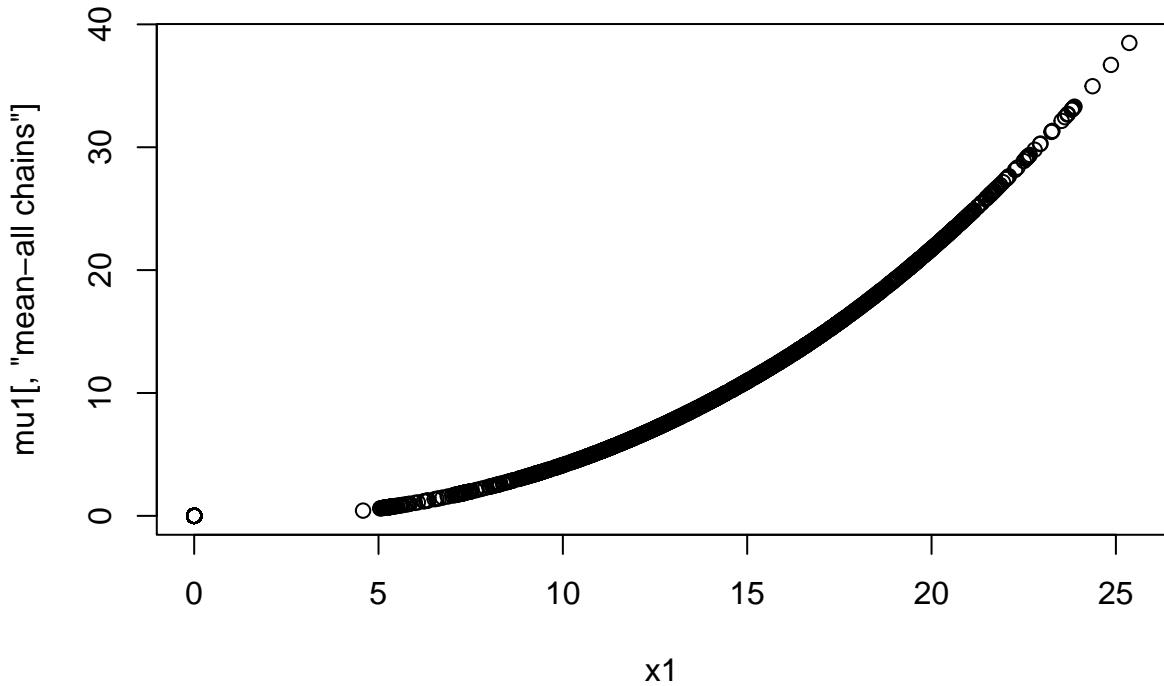




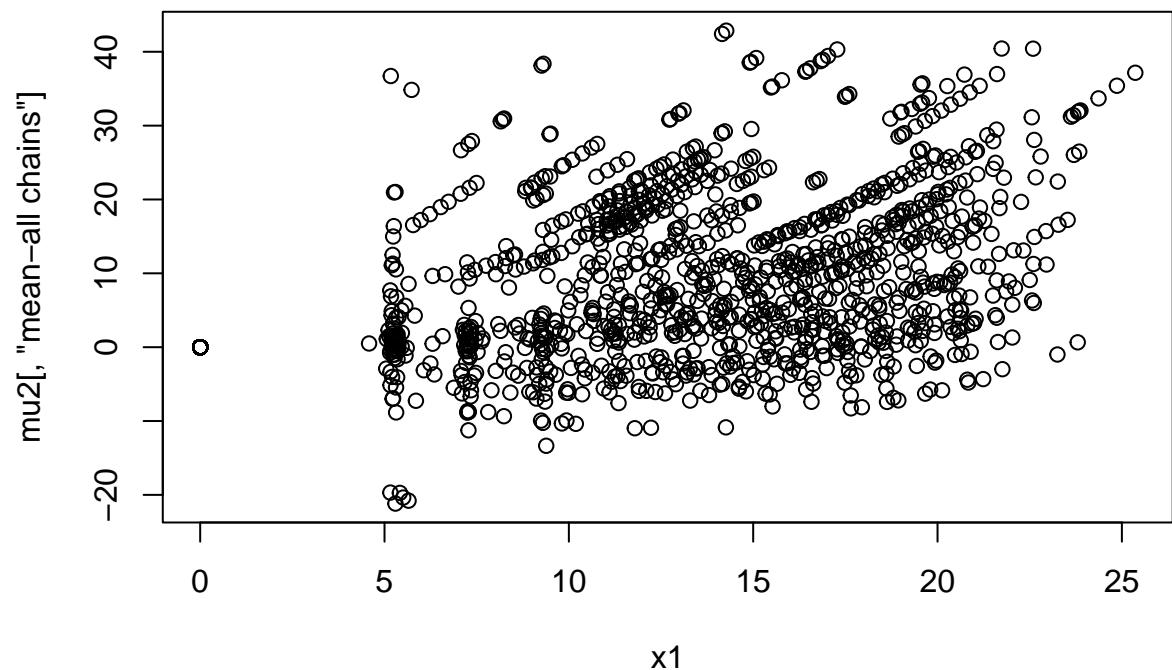
```
pairs(fit.lme.add.non.reslope, pars = c("b0", "b1"), las = 1)
```



```
mu1 = get_posterior_mean(fit.lme.add.non.reslope, "mu1")
plot(x1,mu1[, "mean-all chains"])
```



```
mu2 = get_posterior_mean(fit.lme.add.non.reslope, "mu2")
plot(x1,mu2[, "mean-all chains"])
```



8. Spline con restricciones creciente

8.1. LIN: Spline con restricciones creciente

8.2. LME: Spline con restricciones creciente

```

datos.lme.add.incr <- list( y = y ,
                            id = id ,
                            n = length(y) ,
                            N = N , Ni = Ni ,
                            k1=k1 ,
                            XI1 = XI1 ,
                            x1 = x1 ,
                            zero = rep(0,1+k1) ,
                            S1=S1 )
inits.lme.add.incr <- function(){   list(
  "b0" = rnorm(1,0,0.1) ,
  "b1" = abs(rnorm(k1,0,0.1)),
  "invtau2" = rgamma(1,1,1) ,
  "lambda" = rgamma(1,1,1) ,
  "invsig2" = rgamma(1,1,1)
) }

fit.lme.add.incr.reslope <- stan("jagam_9_aneur_lme_add_incr_reslope.stan",
                                   data=datos.lme.add.incr,
                                   chains=3,warmup=500,iter=1000,thin=2,cores=6,
                                   init= inits.lme.add.incr)

print(fit.lme.add.incr.reslope, pars=param.lme.add, digits=5)

```

```

## Inference for Stan model: jagam_9_aneur_lme_add_incr_reslope.
## 3 chains, each with iter=1000; warmup=500; thin=2;
## post-warmup draws per chain=250, total post-warmup draws=750.
##
##           mean se_mean     sd    2.5%    25%    50%    75%
## b0      -2.88538 0.01432 0.25326 -3.35610 -3.06045 -2.89640 -2.71761
## b1[1]    0.00050 0.00007 0.00174 0.00000 0.00000 0.00000 0.00011
## b1[2]    0.00050 0.00008 0.00192 0.00000 0.00000 0.00000 0.00010
## b1[3]    0.00104 0.00019 0.00517 0.00000 0.00000 0.00000 0.00011
## b1[4]    0.00165 0.00082 0.02240 0.00000 0.00000 0.00000 0.00010
## invtau2  0.03691 0.00006 0.00154 0.03386 0.03590 0.03694 0.03787
## tau2    27.14214 0.04397 1.14260 24.98860 26.40683 27.07197 27.85748
## tau     5.20866 0.00421 0.10943 4.99886 5.13876 5.20307 5.27802
## lambda  0.08767 0.06396 1.75998 0.00000 0.00000 0.00000 0.00002
## rho     -22.35671 1.22923 17.30482 -66.51326 -29.33107 -18.17919 -10.62172
## invsig2 0.45444 0.00242 0.05333 0.36123 0.41633 0.45104 0.48756
## sig2    2.23053 0.01180 0.26012 1.73718 2.05104 2.21712 2.40197
## sigma   1.49098 0.00395 0.08678 1.31802 1.43214 1.48900 1.54983
##             97.5% n_eff     Rhat
## b0      -2.34516 313 1.00821
## b1[1]    0.00551 630 0.99792
## b1[2]    0.00529 637 0.99812
## b1[3]    0.01003 741 0.99887
## b1[4]    0.00803 755 0.99907
## invtau2  0.04002 673 1.00064
## tau2    29.53686 675 1.00071

```

```

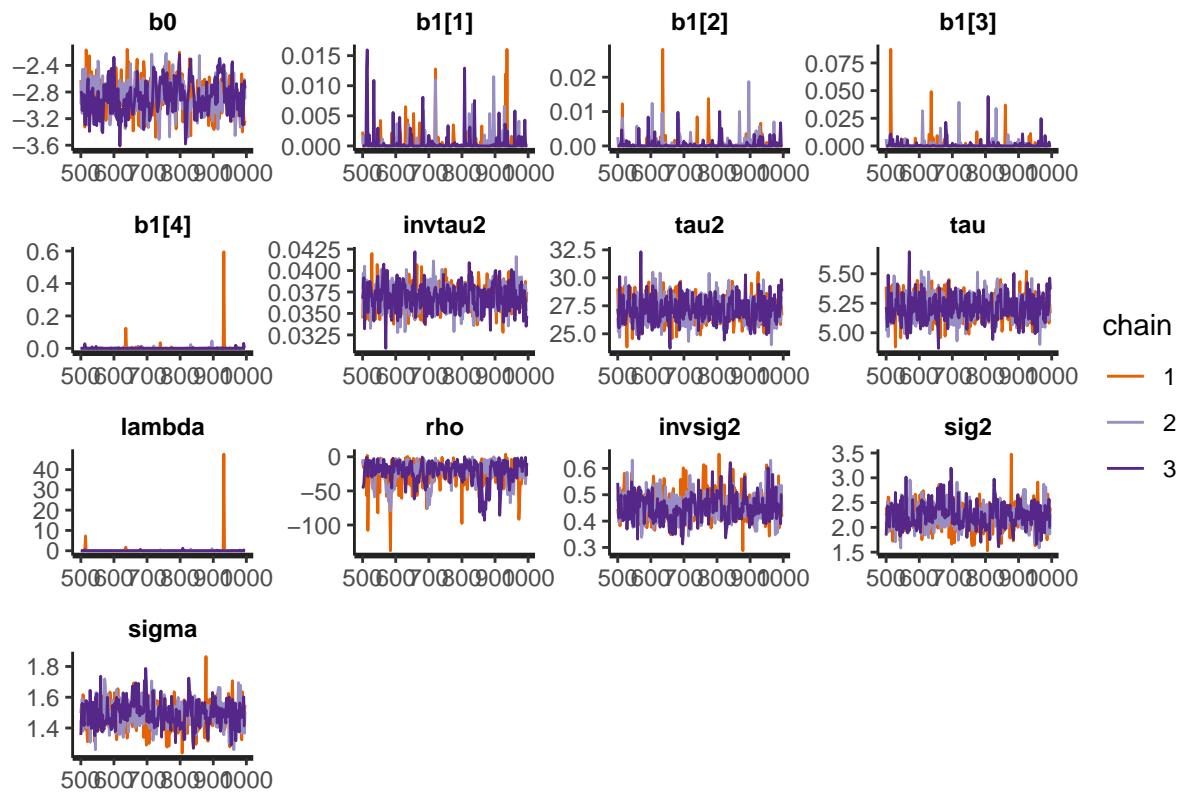
## tau      5.43478   675 1.00070
## lambda   0.12528   757 0.99956
## rho     -2.07947   198 1.00598
## invsig2  0.57565   485 1.00374
## sig2     2.76834   486 1.00353
## sigma    1.66383   482 1.00358
##
## Samples were drawn using NUTS(diag_e) at Tue Jan  9 14:28:41 2024.
## For each parameter, n_eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).

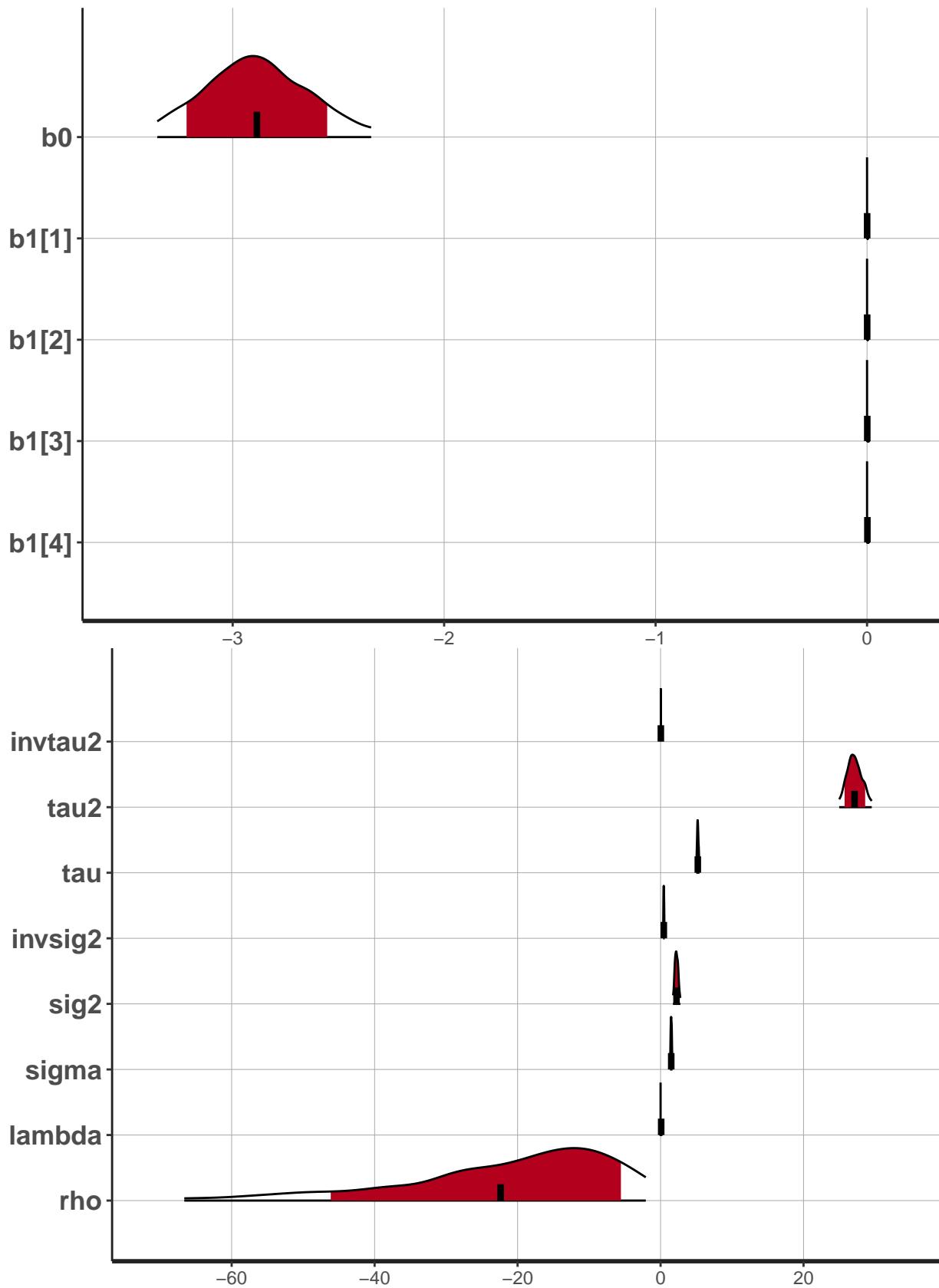
```

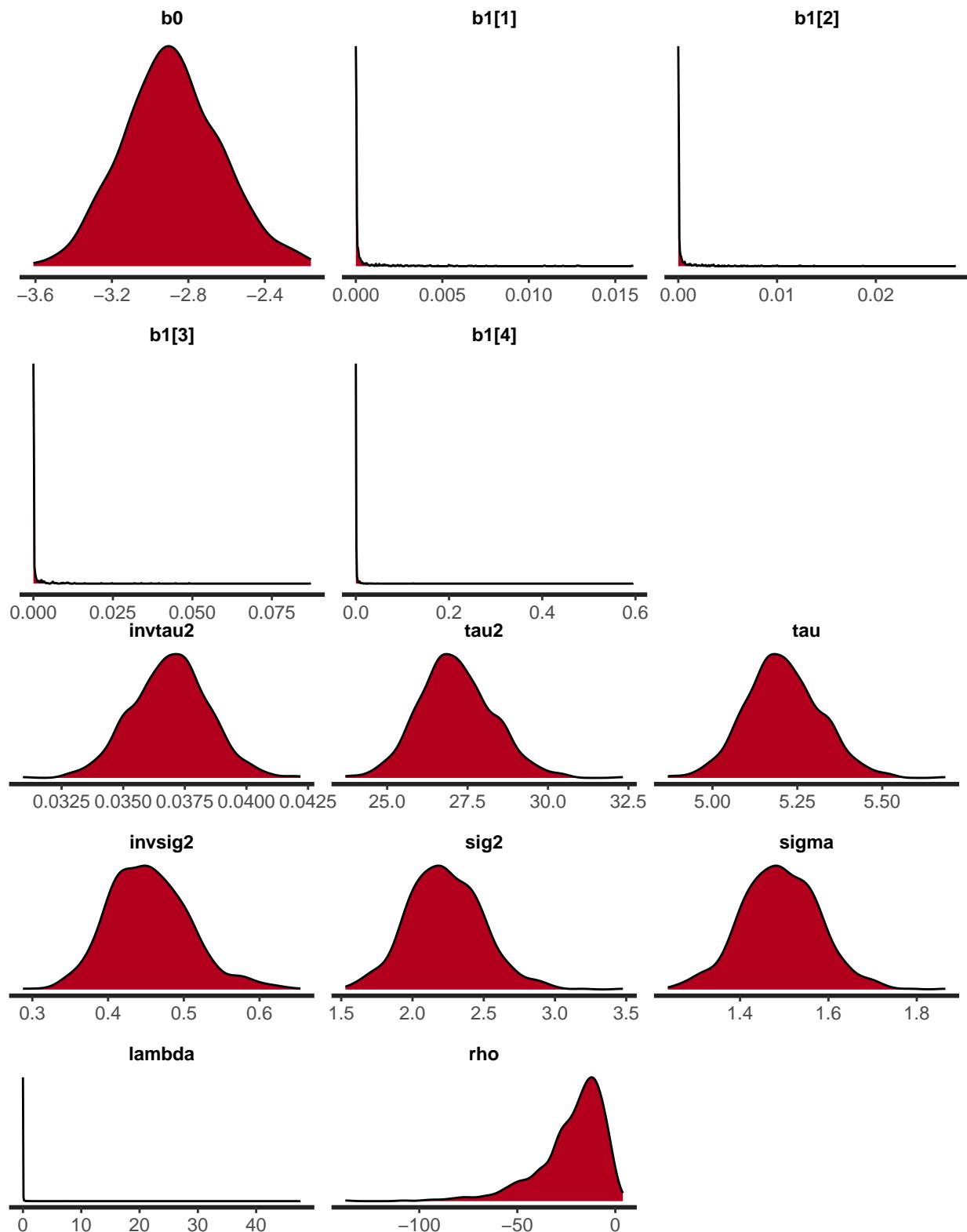
```

stan_trace(fit.lme.add.incr.reslope, pars=param.lme.add)
stan_plot(fit.lme.add.incr.reslope, pars=c("b0","b1"), point_est = "mean", show_density = TRUE)
stan_plot(fit.lme.add.incr.reslope, pars=c("invtau2","tau2","tau", "invsig2","sig2","sigma", "lambda", "rho"))
stan_dens(fit.lme.add.incr.reslope, pars=c("b0","b1"))
stan_dens(fit.lme.add.incr.reslope, pars=c("invtau2","tau2","tau", "invsig2","sig2","sigma", "lambda", "rho"))

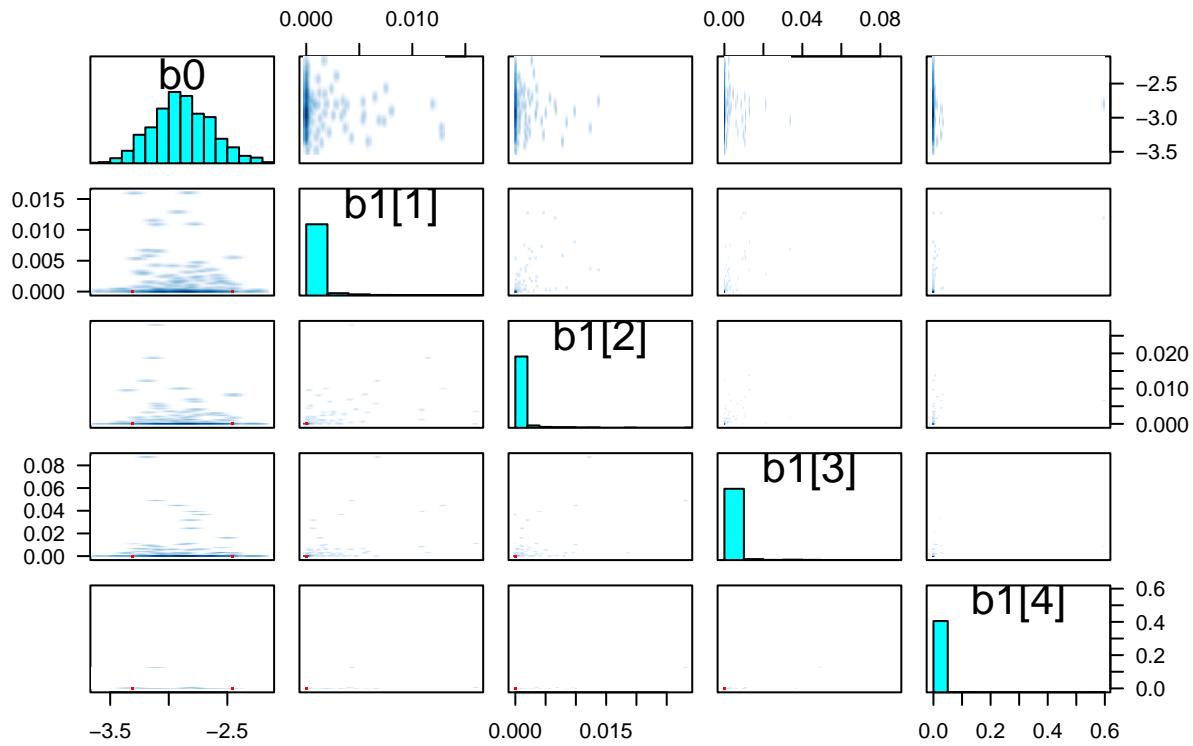
```



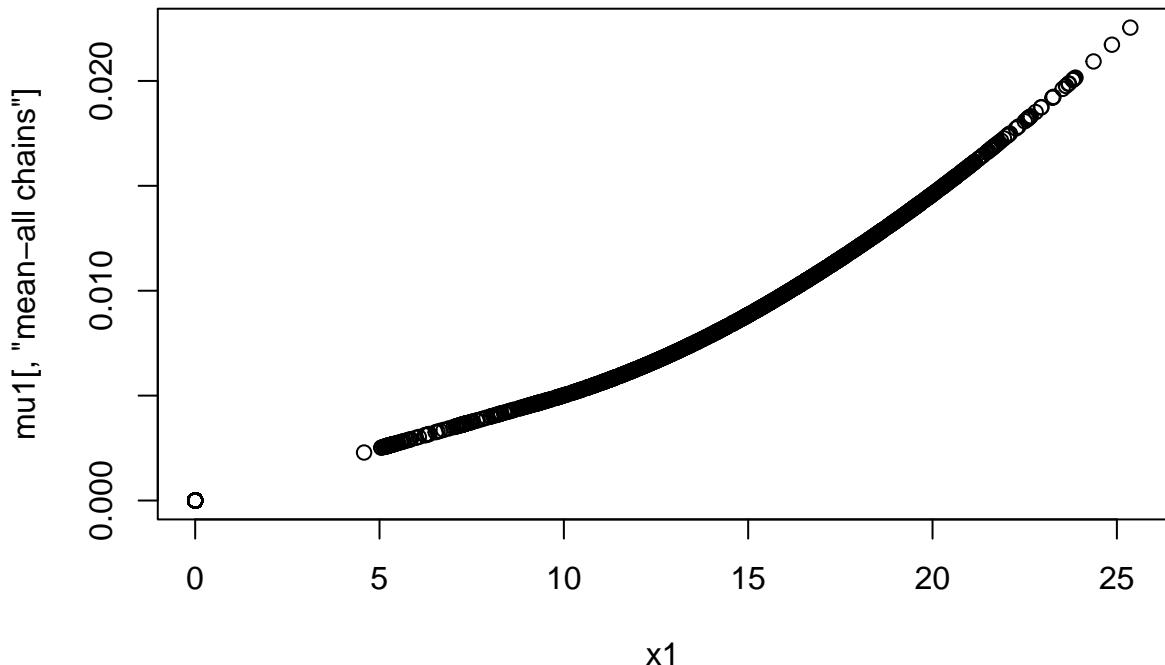




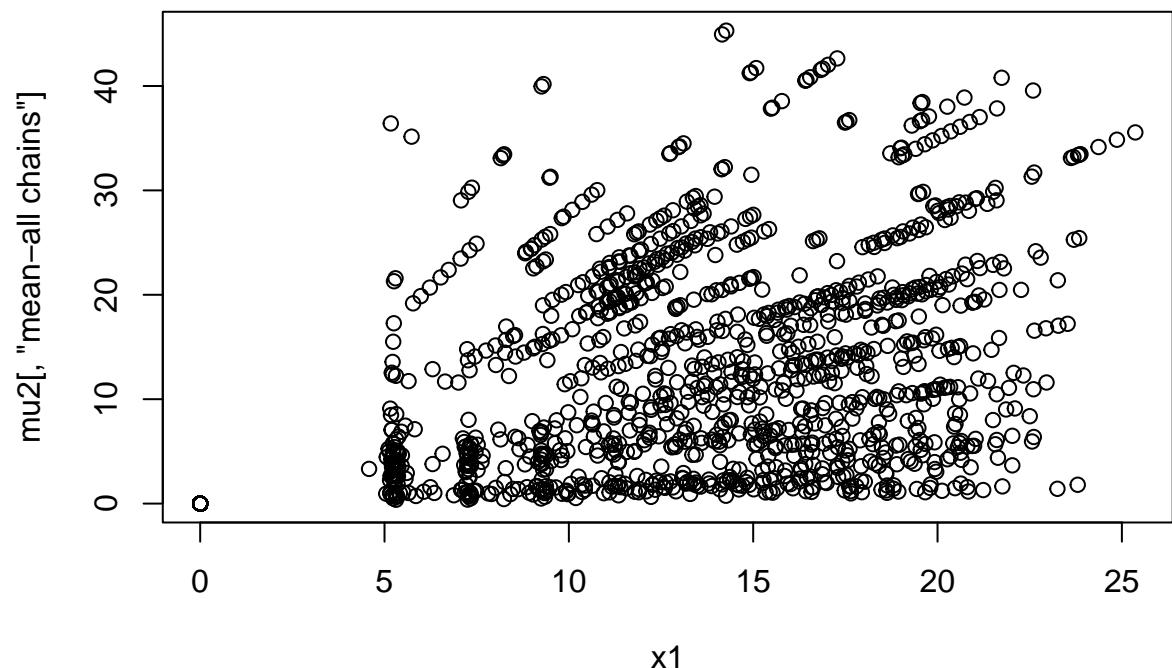
```
pairs(fit.lme.add.incr.reslope, pars = c("b0", "b1"), las = 1)
```



```
mu1 = get_posterior_mean(fit.lme.add.incr.reslope, "mu1")
plot(x1,mu1[, "mean-all chains"])
```



```
mu2 = get_posterior_mean(fit.lme.add.incr.reslope, "mu2")
plot(x1,mu2[, "mean-all chains"])
```



Comparar

```
### http://ritsokiguess.site/docs/2019/06/25/going-to-the-loo-using-stan-for-model-comparison/
library(loo)

## This is loo version 2.4.1

## - Online documentation and vignettes at mc-stan.org/loo

## - As of v2.0.0 loo defaults to 1 core but we recommend using as many as possible. Use the 'cores' arg

##
## Attaching package: 'loo'

## The following object is masked from 'package:rstan':
## 
##     loo

loo3_sample = fit.lme.non.reslope
loo6_sample = fit.lme.incr.reslope
loo9_sample = fit.lme.add.non.reslope
loo12_sample = fit.lme.add.incr.reslope

### we have to extract those log-likelihood terms that we so carefully had Stan calculate for us:
log_lik_3 =extract_log_lik(loo3_sample, merge_chains = F)
log_lik_6 =extract_log_lik(loo6_sample, merge_chains = F)
log_lik_9 =extract_log_lik(loo9_sample, merge_chains = F)
log_lik_12 =extract_log_lik(loo12_sample, merge_chains = F)

r_eff_3 =relative_eff(log_lik_3)
r_eff_6 =relative_eff(log_lik_6)
r_eff_9 =relative_eff(log_lik_9)
r_eff_12 =relative_eff(log_lik_12)

### look at the results for each model, first the one with mu estimated:
(loo_3 <- loo(log_lik_3, r_eff=r_eff_3))

## Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

##
## Computed from 750 by 1387 log-likelihood matrix
##
##          Estimate    SE
## elpd_loo   -4290.7  57.7
## p_loo      253.3   23.0
## looic     8581.4 115.4
## -----
## Monte Carlo SE of elpd_loo is NA.
##
## Pareto k diagnostic values:
```

```

##                                     Count Pct.   Min. n_eff
## (-Inf, 0.5]    (good)      1256 90.6%   78
## (0.5, 0.7]    (ok)        86   6.2%   36
## (0.7, 1]      (bad)       29   2.1%    7
## (1, Inf)      (very bad) 16   1.2%    1
## See help('pareto-k-diagnostic') for details.

```

```
(loo_6 <- loo(log_lik_6, r_eff=r_eff_6))
```

Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

```

##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_loo  -4359.2  59.9
## p_loo      156.5  14.6
## looic     8718.4 119.7
## -----
## Monte Carlo SE of elpd_loo is NA.
##
## Pareto k diagnostic values:
##                                     Count Pct.   Min. n_eff
## (-Inf, 0.5]    (good)      1283 92.5%   60
## (0.5, 0.7]    (ok)        71   5.1%   51
## (0.7, 1]      (bad)       23   1.7%   21
## (1, Inf)      (very bad) 10   0.7%    1
## See help('pareto-k-diagnostic') for details.

```

```
(loo_9 <- loo(log_lik_9, r_eff=r_eff_9))
```

Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

```

##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_loo  -4183.5  61.5
## p_loo      263.1  22.9
## looic     8367.0 122.9
## -----
## Monte Carlo SE of elpd_loo is NA.
##
## Pareto k diagnostic values:
##                                     Count Pct.   Min. n_eff
## (-Inf, 0.5]    (good)      1246 89.8%   70
## (0.5, 0.7]    (ok)        87   6.3%   20
## (0.7, 1]      (bad)       39   2.8%    9
## (1, Inf)      (very bad) 15   1.1%    1
## See help('pareto-k-diagnostic') for details.

```

```

(loo_12 <- loo(log_lik_12, r_eff=r_eff_12))

## Warning: Some Pareto k diagnostic values are too high. See help('pareto-k-diagnostic') for details.

##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_loo   -4352.3  59.5
## p_loo      151.6   13.8
## looic     8704.5 119.1
## -----
## Monte Carlo SE of elpd_loo is NA.
##
## Pareto k diagnostic values:
##                               Count Pct.    Min. n_eff
## (-Inf, 0.5]    (good)    1293 93.2%   114
## (0.5, 0.7]    (ok)       62   4.5%   47
## (0.7, 1]      (bad)      26   1.9%   10
## (1, Inf)     (very bad)  6   0.4%   2
## See help('pareto-k-diagnostic') for details.

#compare(loo_1, loo_2)
### The second model fits better than the first one, since its looic is smaller.

### look at the results for each model, first the one with mu estimated:
(waic_3 <- waic(log_lik_3, r_eff=r_eff_3))

##
## Warning:
## 102 (7.4%) p_waic estimates greater than 0.4. We recommend trying loo instead.

##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_waic  -4276.0  57.0
## p_waic     238.6   21.5
## waic      8552.0 114.0
##
## 102 (7.4%) p_waic estimates greater than 0.4. We recommend trying loo instead.

(waic_6 <- waic(log_lik_6, r_eff=r_eff_6))

##
## Warning:
## 75 (5.4%) p_waic estimates greater than 0.4. We recommend trying loo instead.

##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE

```

```
## elpd_waic -4348.3 58.8
## p_waic      145.6 13.0
## waic        8696.5 117.7
##
## 75 (5.4%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```

```
(waic_9 <- waic(log_lik_9, r_eff=r_eff_9))
```

```
## Warning:
## 114 (8.2%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```

```
##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_waic -4172.0 61.8
## p_waic     251.6 23.0
## waic       8344.0 123.5
##
## 114 (8.2%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```

```
(waic_12 <- waic(log_lik_12, r_eff=r_eff_12))
```

```
## Warning:
## 70 (5.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```

```
##
## Computed from 750 by 1387 log-likelihood matrix
##
##           Estimate     SE
## elpd_waic -4342.1 58.6
## p_waic     141.4 12.1
## waic       8684.2 117.2
##
## 70 (5.0%) p_waic estimates greater than 0.4. We recommend trying loo instead.
```

```
#compare(waic_1, waic_2)
### The second model fits better than the first one, since its looic is smaller.
```