

# The problem and promise of modeling plant leafout under warmer winters

E.M. Wolkovich<sup>1</sup>, Justin Ngo<sup>1</sup>, Victor van der Meersch<sup>1</sup>, Jonathan Auerbach<sup>2</sup>

November 30, 2025

<sup>1</sup> Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, 2424 Main Mall, Vancouver, BC V6T 1Z4, Canada

<sup>2</sup> Department of Statistics, George Mason University, 4511 Patriot Circle, Fairfax, VA 22030, United States

## Abstract

How warmer winters affect plant leafout has major implications for carbon storage and ecosystem change, but is poorly understood. Recent evidence of a slow-down in the advance of spring leafout with increasing anthropogenic climate change has reinvigorated a decades-old debate over winter ‘chilling’—the concept that woody temperate plants need sufficient winter cool temperatures to budburst each spring. Decades of research has argued that chilling is a critical plant process—one that is never observed but often assumed.

We review how current work, which routinely models ‘chilling’ with highly limited data has likely led to the proliferation of models—and thus forecasts—that yield little additional insight about observable plant behavior. The result is that chilling, like other theoretical constructs such as the “ether”, has become a placeholder that does not lead to unique predictions and thus has stalled scientific progress. We argue that, until we overcome this, new insights and approaches from molecular biology that could better define the mechanism of chilling will likely lead to only limited progress. By understanding the limits of chilling as a theory, we can both 1. build better models by looking for the necessary insight from molecular biology to fill in the details, and 2. design better experiments to test those theories.

Plant leafout in the spring has shifted weeks earlier in many regions due to warming from anthropogenic climate change with consequences for a suite of ecosystem services, including carbon sequestration (Keenan *et al.*, 2014; Pörtner *et al.*, 2022). Which underlying processes drive this trend, however, is debated, as recent research suggests winter warming may slow or stall this advance (Fu *et al.*, 2015; Piao *et al.*, 2017). Such reports focus on a two-step model of leafout where plants first require cool winter temperatures, often called ‘chilling,’ before they can accumulate enough warm temperatures—‘forcing’—to leafout each year. This model of chilling is only one of many models proposed since the concept was first introduced (Basler, 2016; Hufkens *et al.*, 2018). But, while forcing ends in an observable event such as leafout, the day plants achieve their chilling requirements coincides with no known measurable event.

Current models of chilling are consistent with a diverse suite of possible future leafout. Chilling in the same location under different models can easily forecast significantly increased or greatly reduced chilling and thus greatly advanced or slowed leafout (Guy, 2014; Chuine *et al.*, 2016). This variability often occurs within any one model of chilling, given different parameter values. Such extreme variability of spring phenology predicted from current models suggests fundamental gaps in our understanding. While decades of experimental evidence suggest that winter temperatures impact leafout (Baumgarten *et al.*, 2021) new research has highlighted major flaws in these models as currently applied, including multiple papers now suggesting estimates of ‘chilling’ could easily be artifacts of poor models or correlated observational climate data (Wolkovich *et al.*, 2021; Gao *et al.*, 2024).

Now appears an opportune time to address the problems in our concept of chilling in models of leafout. Because accurate forecasts of spring phenology are critical for carbon storage, many crops and a number of other services important to humans, there is widespread interest in improving chilling models (Luedeling *et al.*, 2015; Chuine *et al.*, 2016). At the same time, new results from molecular and cellular studies of dormancy are providing new insights into when and how chilling works (Pan *et al.*, 2023; Zhu *et al.*, 2021), which could rule in—or out—some of the myriad current models. Here, we review the concept of chilling, its origins and potential problems, as well as new opportunities for major advances and how shifting current practices could accelerate progress. We argue that revisiting the chilling mechanism has two benefits: 1. building better models, and 2. designing better experiments.

## What is chilling?

How plants in temperature-limited systems avoid leafout during warm spells in the winter has long been debated by plant biologists (e.g., Lamb, 1948; Weinberger, 1950). Most work to date has focused on the idea that plants enter some form of dormancy in the fall, which is then released before warm temperatures in the spring begin. This idea hypothesizes that the slow accumulation of cool temperatures—or chilling—over the winter extends through periods of short warm spells in the winter and thus prevents leafout before spring.

Much of our fundamental understanding of chilling comes from studies on temperate woody fruit crops where chilling can be critical to yield. Peach trees planted into warmer climates well outside their range often have extremely low fruitset because most flower buds do not burst (Weinberger, 1950; Overcash & Campbell, 1955; Erez *et al.*, 1971). Initial studies of this phenomenon with related experiments—where cut ends of dormant branches (cuttings) exposed to cooler temperatures in chambers burst more fully and more quickly—underlies most of the models of chilling used today for crops and wild tree species (Weinberger, 1950; Ettinger *et al.*, 2020).

The term ‘chilling’ is now used across numerous fields in plant biology to refer to a process where dormant buds exposed to cool temperatures accelerate a phenological event that later occurs after warm temperatures. Focused on how chilling can accelerate events, researchers have calculated ‘chilling’ required for leafout of forest trees from cutting experiments similar to those used for peaches (reviewed in Ettinger *et al.*, 2020), ground observations of budburst (Luedeling *et al.*, 2009b), and satellite measures of greenup (Kaduk & Los, 2011). These estimates rely on phenolog-

ical models that have become critical across a suite of fields, from climatology where modeling how vegetation on the land surface responds to anthropogenic climate change affects carbon storage, to crop biology, where estimated chill units guide growers in which specific cultivar to plant, and has led to the cross-disciplinary field of phenology (Schwartz, 1994; Cleland *et al.*, 2007; Chuine *et al.*, 2013).

Alongside these more macro-scale studies of chilling, molecular approaches have also examined chilling. Many studies have focused on vernalization—cool temperatures required for flowering (Kim *et al.*, 2009)—in *Arabidopsis thaliana*, with studies in woody species, especially *Populus* examining chilling before budbreak (Azeez *et al.*, 2021; Cai *et al.*, 2024). These studies generally use controlled temperatures to vary the hypothesized amount of chilling then examine molecular and cellular responses (e.g., Pan *et al.*, 2021; Azeez *et al.*, 2021; Cai *et al.*, 2024).

Today these studies have led to over 30 basic models where accumulated chilling releases plants from dormancy and hundreds more when considering different species and cultivars (Basler, 2016; Hufkens *et al.*, 2018). Though early debates considered whether plants were truly dormant or only growing slow (‘dormancy’ or ‘rest’ versus ‘quiescent’; Considine & Considine, 2016), today most research assumes a model with two phases of dormancy: endodormancy—where chilling occurs—and a period after but before the observed event, called ecodormancy (Fig. ??). In most models, chilling can only be accumulated under certain temperatures—traditionally above zero but below 10°C—with certain temperatures being optimal for the most rapid accumulation of ‘chill units,’ where some unknown sum of chill units breaks endo-dormancy. Which temperatures are most effective at providing chilling is a common question addressed in experiments, with different experiments providing different answers (Vitasse *et al.*, 2013; Baumgarten *et al.*, 2021).

Further complexity comes from the hypothesized diversity of these temperature thresholds and sums across species and populations. Most assume different species require different sums of chill units, and may have different lower, upper and optimal chill temperatures. Within species, populations may require different sums of chill units, with populations in more mild climates—where warm interruptions in the winter are more common—requiring more chill units than those in areas with cold winters, where temperatures rarely rise above zero before spring (Campbell & Sugano, 1979; Leinonen, 1996).

## The problem with chilling

Chilling is a latent, unobserved process. Typically, ‘chilling’ describes the physiological phase (endodormancy) in which a plant experiences environmental temperatures that induce progress towards the next physiological phase (eco-dormancy) that ends in budburst (Fig. ??). The problem is that the transition from endo- to eco-dormancy corresponds with no clearly measurable phenomenon, and thus the properties of each period cannot be determined from the observed leafout, even under experimental conditions in which the temperature can be manipulated. The parameters of a model governing this system are said to be underdetermined or unidentified.

For example, consider a simple model in which one would need to estimate the minimum and maximum temperatures that allow chilling to accumulate (two parameters) and the total sum of

those temperature units needed to trigger a shift into the next physiological phase (often called, ‘endo-dormancy break,’ for one additional parameter for three total). Models then need to estimate when plants start and stop accumulating (two more parameters). An experiment that raises temperatures and observes an earlier leafout could be explained by more rapid chilling accumulation leading to earlier endo-dormancy break, more rapid ecodormancy break, or an almost limitless mix of the two.

To address this, models often assign the start date of the endo-dormancy as known (e.g., starting 1 September in the northern hemisphere) and rely on assumptions to set the end date (see Box: Why has progress on modeling stalled for decades?). A common assumption, developed by early work on peaches, is that high and rapid budburst (leaf or flower) is evidence that chilling has been met (i.e. endo-dormancy has ended Erez & Lavee, 1971). While this assumption is widely used, it is rarely if ever tested beyond the early work on peaches. This approach of assigning some unknown parameters as known has the benefit of avoiding adding more unknown model parameters, but it also has led researchers to be overly confident in a model of chilling where more is actually unobserved and unknown than acknowledged.

Hidden assumptions and numerous parameters can easily drive diverging models. Even if we assume high and rapid percent budburst signals sufficient chilling, most models today include parameters that cannot be uniquely identified with current data. Given experiments and models have suggested many variants on a more complicated model of chilling—for example minimum, maximum and optimal temperatures, or high temperatures that reduce previous accumulation (Fig. ??, and see Luedeling *et al.*, 2011, 2012; Chuine *et al.*, 2016)—current data are relatively uninformative to try to estimate all the parameters the models include. Further, recent models have often relied on even less data (Hänninen *et al.*, 2019). Many current methods use only observational data of the timing of leafout (or flowering) to attempt to estimate a model of chilling for different species or locations and project it forward to understand effects of anthropogenic climate change (Luedeling *et al.*, 2011, 2012; Gao *et al.*, 2024). Perhaps not surprisingly then, which model is deemed best varies strongly by method and approach (Caffarra *et al.*, 2011; Basler, 2016; Hufkens *et al.*, 2018), with no clear pattern.

## New molecular insights could reshape the field and its models

New molecular research provides hope that the transition from endo- to eco-dormancy is measurable. Molecular insights have long contributed to crop and forest tree models of chilling (Chuine & Regniere, 2017). Decades of work on vernalization have outlined the pathways—and genes—that lead to flowering only after winter’s cool temperatures in biennial (herbaceous) populations of *Arabidopsis thaliana* (Fig. ??, Wilczek *et al.*, 2009; Kim *et al.*, 2009). Research has linked some of these pathways to similar ones in woody species, and have also highlighted the sugar callose (1,3- $\beta$ -D-glucan) as potentially pivotal for chilling (van der Schoot *et al.*, 2014; Pan *et al.*, 2021). Multiple studies across multiple species have now shown that (1) lower temperatures appear to degrade callose and (2) the release/loss of callose appears to re-start cell-to-cell communication before budburst (van der Schoot *et al.*, 2014). Taken together, these results suggest the loss of callose—generally degraded through 1,3- $\beta$ -glucanases (a group of enzymes)—may be an indicator of endo-dormancy

release, though other factors, such as ABA, also often change at the same time (Tylewicz *et al.*, 2018; Pan *et al.*, 2021), and may provide a similar observable signal of endo-dormancy release (Rinne *et al.*, 2018; André *et al.*, 2022).

If callose is functionally a major controller of endo-dormancy and its release, then chilling models could be limited to those that match the idea of glucanase degrading callose—meaning models that include a temperature range over which the enzyme is active (Fig. ??). In contrast, models using simple temperature thresholds (e.g., all hours below  $-5^{\circ}\text{C}$  equally allow chilling) would appear less biologically accurate, as enzymes generally do not work over such a wide range of temperatures.

Other new molecular insights similarly suggest that such simplified temperature metrics used in many chilling models may not map to molecular realities. For example, new work on how slow growth itself may act a ‘long-term thermosensor’ (Zhao *et al.*, 2020) adds to an increasing number of molecular studies that suggest plants integrate long-term thermo-sensing in the winter alongside responses to short-term temperatures (Antoniou-Kourounioti *et al.*, 2021; Satake *et al.*, 2022). The best models of leafout may thus need to integrate across multiple timescales.

New molecular results could easily add complexity to models of spring phenology that are already challenged by too much complexity. But they could also begin to rule out models by focusing on new experiments and modeling approaches that target the major problems facing models of chilling—if the field is more open about those issues and open to new approaches.

## Overcoming the chilling problem

Richer, more informative data from molecular biology studies and other approaches to identify chilling (Fouché *et al.*, 2023; Walde *et al.*, 2024) hold the promise to shift chilling from an unobserved complex process to something we understand and can robustly forecast. Taking full advantage of this opportunity, however, would benefit from re-examining our concept of chilling by working across different disciplines to combine observational and experimental evidence, while also leaving opportunity to question the future utility of ‘chilling.’

### Model experimental and observational data together

Research on chilling could accelerate by working across what today are three fairly separated silos of model building for crops and other forecasting (usually called process-based models), experiments at the branch or whole-plant level, and molecular research. Currently, crop biologists, phenological process-based modelers of forest trees, molecular biologists and hardiness modelers all develop unique and rarely compared models of dormancy and budburst (but see Kovaleski, 2022), highlighting a major problem, but also a major opportunity. Synthesizing models—and their underlying biological understanding of chilling—across the many research fields developing chilling models today would help identify models that are equivalent.

Synthesizing would hopefully driver new approaches that estimate chilling by fitting experimental and observational data together in one model. This is rarely (if ever) done, in part because of how differently they may be observed, including the challenging diversity of environmental conditions across these two data types (for example, many experiments apply cold temperatures in the dark and

include extreme temperature differences, while photoperiod shifts each day in observational data and temperatures are more similar) but also because of separate modeling approaches. Researchers rarely if ever fit process-based models to new empirical data; instead they use so-called ‘statistical models’ that often follow canonical treatment designs (e.g., ANOVA). Statistical models are usually far simpler, and make a suite of unstated assumptions that contradict the current understanding of chilling (see Fig. ??). Many of the original studies that led to the concept of chilling, however, were developed from datasets that created greater extremes in observational data—focusing on crops planted well outside their natural range (e.g., peaches in Florida and Israel) and bridged across observational and experimental studies more often (Erez & Lavee, 1971; Richardson, 1974).

Experiments bridging across methods may have the greatest opportunity to provide data that would truly challenge current models of chilling. Testing models across large environmental gradients in the field is one of the best ways to find out where models work—and where they fail. For example, molecular biologists tested vernalization models by through comparing predicted to observed flowering times in a common garden study across Europe (Wilczek *et al.*, 2009), and supported the temperature-dependent growth model by testing its predictions of what happens when growth is altered but temperature is held constant (Zhao *et al.*, 2020). Similar examples for challenging other models of chilling date back over 40 years to when many of the models used today were developed (Richardson, 1974; Chuine *et al.*, 2016; Ettinger *et al.*, 2020), but could take place now. Models of chilling can make predictions under lower field chilling then test them using individuals planted beyond the range (either planting those individuals now or identifying such cases in forestry provenance trials or similar). Process-based modelers could also challenge their models more through more dramatic variation in biology, via experiments that include mutants or similar variants.

## Is ‘chilling’ necessary for scientific progress?

One important way to leverage new molecular insights for modeling is through new experiments designed to identify novel ways to more directly observe and measure chilling. Experiments testing for evidence of callose loss using the temperature treatments commonly applied in past studies (Ettinger *et al.*, 2020) could be complemented by studies with other cellular and molecular markers (Yu *et al.*, 2024). Testing these methods together alongside previous-used markers of dormancy shifts—including the often-used bioassay of high and rapid budburst at higher temperatures (indicating endo-dormancy release), and additional methods, such as weighing flower buds (Chuine *et al.*, 2016) or tracing water reactivation into cells (Faust *et al.*, 1991; Kalcsits *et al.*, 2009; Walde *et al.*, 2024)—could help align both new and old methods.

Because chilling is an unobserved process we argue that comparing methods to measure chilling should be a major priority for the field. This comparison will need to allow for the reality that different methods may measure different processes and, thus, terminology may need to adapt as well. As a first step, research could stop referring to treatments in experiments as ‘chilling’ or ‘forcing,’ or other terms that assume an underlying physiological state, and instead focus on the actual treatments (e.g., ‘cool temperatures before warm’). Currently, many experiments use the term ‘chilling’ to refer to a treatment where researchers do not know the physiological phase (Flynn & Wolkovich, 2018; Ettinger *et al.*, 2020); for example, cuttings or buds from woody plants are often chilled at 5°C for 6 weeks in the dark in a ‘chilling’ treatment, then transferred to warming

‘forcing’ conditions.

We argue that one of the main reasons for stalled progress on modeling chilling is that most models—from the old to new ones—cannot actually estimate the parameters in them. Taking a simple example of a chilling model with three parameters—the minimum temperature for chilling, the maximum temperature, and the accumulation needed—shows that there are multiple solutions. Considering just two of these possible solutions highlights how the temperature range trade-offs with the accumulation: if the temperature range is wide (lower minimum, higher maximum) then the accumulation required will be higher, while if the range is smaller, then the accumulation needed is lower. The full suite of possible solutions is effectively endless (and the trade-off between range and accumulation is not linear, as it depends on the full width of the range, but also its placement relative to 0). Further, this model is not actually one of only three parameters as two additional parameters were set as known (start day of accumulation was set at 1 September, and the endodormancy break date at 30 January) so that the model could even be fit using common algorithms. This reality is present in every model of chilling, but it is rarely presented clearly.

Researchers tacit approach to these major issues likely has contributed to the expansion of models over the last few decades without any clear advances. While various models have added complexity via the shape of the optimal temperature range for chilling, allowing accumulated chilling to be reduced, shifting the start date of chilling, and/or allowing chilling and forcing to act at once (Luedeling *et al.*, 2009a; Gusewell *et al.*, 2017; Hänninen, 1990; Kramer, 1994), none of these have swept through the field. These new models of chilling have all added parameters, but none of the parameters added to chilling models in 40 years that have been successful enough to be added to all models of chilling.

Being clear about model uncertainty, the full number of parameters and how well they fit would advance progress on chilling through multiple routes. First, it would help all researchers in the field recognize what is fundamentally unknown and thus focus more research in these areas. Second, it would highlight which parameters are most often fixed (effectively assumptions in the model) versus fit to data, and to which type of data. With this, more research could easily compare across models and datasets to give better overviews of what is known, assumed, or most often studied (i.e., what parameter model studies try to fit). Given the extensive list of proposed complexities to chilling models (Lamb, 1948; Campbell & Sugano, 1979; Leinonen, 1996; Cook *et al.*, 2005; Man & Lu, 2010; Jones *et al.*, 2012; Sønsteby & Heide, 2014), having simpler models (fewer parameters) that are routinely used to compare to more complex models, would likely help the field advance. Highlighting uncertainty in findings from experiments would also aid modeling studies to be more upfront about assumptions and limitations (Fig. ??).

# 1 References

- André, D., Zambrano, J.A., Zhang, B., Lee, K.C., Rühl, M., Marcon, A. & Nilsson, O. (2022) Populus svl acts in leaves to modulate the timing of growth cessation and bud set. *Frontiers in Plant Science* **13**, 823019.
- Antoniou-Kourounioti, R.L., Zhao, Y., Dean, C. & Howard, M. (2021) Feeling every bit of winter-distributed temperature sensitivity in vernalization. *Frontiers in Plant Science* **12**, 628726.
- Azeez, A., Zhao, Y.C., Singh, R.K., Yordanov, Y.S., Dash, M., Miskolczi, P., Stojković, K., Strauss, S.H., Bhalerao, R.P. & Busov, V.B. (2021) Early bud-break 1 and early bud-break 3 control resumption of poplar growth after winter dormancy. *Nature communications* **12**, 1123.
- Basler, D. (2016) Evaluating phenological models for the prediction of leaf-out dates in six temperate tree species across central europe. *Agricultural and Forest Meteorology* **217**, 10–21.
- Baumgarten, F., Zohner, C.M., Gessler, A. & Vitasse, Y. (2021) Chilled to be forced: the best dose to wake up buds from winter dormancy. *New Phytologist* **230**, 1366–1377.
- Caffarra, A., Donnelly, A. & Chuine, I. (2011) Modelling the timing of *Betula pubescens* budburst. II. Integrating complex effects of photoperiod into process-based models. *Climate Research* **46**, 159–170.
- Cai, F., Jin, X., Tian, Y., Huang, Z., Wang, X., Zhang, Y., Sun, Y. & Shao, C. (2024) Molecular regulation of bud dormancy in perennial plants. *Plant Growth Regulation* **102**, 1–11.
- Campbell, R.K. & Sugano, A.I. (1979) Genecology of bud-burst phenology in Douglas-fir - response to flushing temperature and chilling. *Botanical Gazette* **140**, 223–231.
- Chuine, I., Bonhomme, M., Legave, J.M., de Cortazar-Atauri, I.G., Charrier, G., Lacointe, A. & Ameglio, T. (2016) Can phenological models predict tree phenology accurately in the future? the unrevealed hurdle of endodormancy break. *Global Change Biology* **22**, 3444–3460.
- Chuine, I., Garcia de Cortazar Atauri, I., Hanninen, H. & Kramer, K. (2013) *Plant development models*, pp. 275–293. Kluwer, Dordrecht, the Netherlands.
- Chuine, I. & Regniere, J. (2017) Process-based models of phenology for plants and animals. *Annual Review of Ecology, Evolution, and Systematics* **48**, 159–182.
- Cleland, E.E., Chuine, I., Menzel, A., Mooney, H.A. & Schwartz, M.D. (2007) Shifting plant phenology in response to global change. *Trends in Ecology & Evolution* **22**, 357–365.
- Considine, M.J. & Considine, J.A. (2016) On the language and physiology of dormancy and quiescence in plants. *Journal of Experimental Botany* **67**, 3189–3203.
- Cook, N.C., Bellen, A., Cronjé, P.J., De Wit, I., Keulemans, W., Van den Putte, A. & Steyn, W. (2005) Freezing temperature treatment induces bud dormancy in ‘granny smith’apple shoots. *Scientia horticultrae* **106**, 170–176.



- Erez, A. & Lavee, S. (1971) Effect of climatic conditions on dormancy development of peach buds. i. temperature. *Journal of the American Society for Horticultural Science* .
- Erez, A., Lavee, S. & Samish, R.M. (1971) Improved methods for breaking rest in the peach and other deciduous fruit species. .
- Ettinger, A.K., Chamberlain, C.J., Morales-Castilla, I., Buonaiuto, D.M., Flynn, D.F.B., Savas, T., Samaha, J.A. & Wolkovich, E.M. (2020) Winter temperatures predominate in spring phenological responses to warming. *Nature Climate Change* **10**, 1137–U119.
- Faust, M., Liu, D., Millard, M.M. & Stutte, G. (1991) Bound versus free water in dormant apple buds—a theory for endodormancy. *HortScience* **26**, 887–890.
- Flynn, D.F.B. & Wolkovich, E.M. (2018) Temperature and photoperiod drive spring phenology across all species in a temperate forest community. *New Phytologist* **29**, 1135–1492.
- Fouché, M., Bonnet, H., Bonnet, D.M. & Wenden, B. (2023) Transport capacity is uncoupled with endodormancy breaking in sweet cherry buds: physiological and molecular insights. *Frontiers in Plant Science* **14**, 1240642.
- Fu, Y.S.H., Zhao, H.F., Piao, S.L., Peaucelle, M., Peng, S.S., Zhou, G.Y., Ciais, P., Huang, M.T., Menzel, A., Uelas, J.P., Song, Y., Vitasse, Y., Zeng, Z.Z. & Janssens, I.A. (2015) Declining global warming effects on the phenology of spring leaf unfolding. *Nature* **526**, 104–107.
- Gao, X., Richardson, A.D., Friedl, M.A., Moon, M. & Gray, J.M. (2024) Thermal forcing versus chilling? misspecification of temperature controls in spring phenology models. *Global Ecology and Biogeography* **33**, e13932, e13932 GEB-2024-0136.R2.
- Giertrych, M. (ed.) (1974) *International Symposium on Dormancy in Trees Discussions: 5th to 9th September 1973*. Polish Academy of Sciences.
- Gusewell, S., Furrer, R., Gehrig, R. & Pietragalla, B. (2017) Changes in temperature sensitivity of spring phenology with recent climate warming in switzerland are related to shifts of the preseason. *Global Change Biology* **23**, 5189–5202.
- Guy, R.D. (2014) The early bud gets to warm. *New Phytologist* **202**, 7–9.
- Hänninen, H. (1990) *Modelling bud dormancy release in trees from cool and temperate regions*. 213.
- Hänninen, H., Kramer, K., Tanino, K., Zhang, R., Wu, J. & Fu, Y.H. (2019) Experiments are necessary in process-based tree phenology modelling. *Trends in Plant Science* **24**, 199–209.
- Hufkens, K., Basler, D., Milliman, T., Melaas, E.K. & Richardson, A.D. (2018) An integrated phenology modelling framework in r. *Methods in Ecology and Evolution* **9**, 1276–1285.
- Jones, H., Hillis, R., Gordon, S. & Brennan, R. (2012) An approach to the determination of winter chill requirements for different *Ribes* cultivars. *Plant Biology* **15**, 18–27.
- Kaduk, J.D. & Los, S.O. (2011) Predicting the time of green up in temperate and boreal biomes. *Climatic Change* **107**, 277–304.

- Kalcsits, L., Kendall, E., Silim, S. & Tanino, K. (2009) Magnetic resonance microimaging indicates water diffusion correlates with dormancy induction in cultured hybrid poplar (*populus* spp.) buds. *Tree Physiology* **29**, 1269–1277.
- Keenan, T.F., Gray, J., Friedl, M.A., Toomey, M., Bohrer, G., Hollinger, D.Y., Munger, J.W., O’Keefe, J., Schmid, H.P., Wing, I.S. *et al.* (2014) Net carbon uptake has increased through warming-induced changes in temperate forest phenology. *Nature Climate Change* **4**, 598–604.
- Kim, D.H., Doyle, M.R., Sung, S. & Amasino, R.M. (2009) Vernalization: winter and the timing of flowering in plants. *Annual Review of Cell and Developmental* **25**, 277–299.
- Kovaleski, A. (2022) Woody species do not differ in dormancy progression: differences in time to budbreak due to forcing and cold hardiness. *Proceedings of the National Academy of Sciences* **119**, e2112250119.
- Kramer, K. (1994) Seleecting a model to predict the onset of growth of *Fagus sylvatica*. *Journal of Applied Ecology* **31**, 172–181.
- Lamb, R.C. (1948) Effect of temperatures above and below freezing on the breaking of rest in the Latham raspberry. *Proceedings of the American Society for Horticultural Science*, vol. 51, pp. 313–315, AMER SOC HORTICULTURAL SCIENCE 113 S WEST ST, STE 200, ALEXANDRIA, VA 22314 . . . .
- Leinonen, I. (1996) Dependence of dormancy release on temperature in different origins of *pinus sylvestris* and *betula pendula* seedlings. *Scandinavian Journal of Forest Research* **11**, 122–128.
- Li, W., Xin, Q., Zhou, X., Zhang, Z. & Ruan, Y. (2021) Comparisons of numerical phenology models and machine learning methods on predicting the spring onset of natural vegetation across the northern hemisphere. *Ecological Indicators* **131**, 108126.
- Luedeling, E., Blanke, M. & Gebauer, J. (2012) Chilling challenges in a warming world. *II International Symposium on Horticulture in Europe 1099*, pp. 901–907.
- Luedeling, E., Blanke, M. & Gebauer, J. (2015) *Chilling Challenges in a Warming World*, vol. 1099 of *Acta Horticulturae*, pp. 901–907.
- Luedeling, E., Girvetz, E.H., Semenov, M.A. & Brown, P.H. (2011) Climate change affects winter chill for temperate fruit and nut trees. *Plos One* **6**.
- Luedeling, E., Zhang, M.H. & Girvetz, E.H. (2009a) Climatic changes lead to declining winter chill for fruit and nut trees in California during 1950–2009. *Plos One* **4**.
- Luedeling, E., Zhang, M.H., McGranahan, G. & Leslie, C. (2009b) Validation of winter chill models using historic records of walnut phenology. *Agricultural and Forest Meteorology* **149**, 1854–1864.
- Man, R. & Lu, P. (2010) Effects of thermal model and base temperature on estimates of thermal time to bud break in white spruce seedlings. *Canadian Journal of Forest Research* **40**, 1815–1820.
- McMaster, G.S. & Wilhelm, W. (1997) Growing degree-days: one equation, two interpretations. *Agricultural and forest meteorology* **87**, 291–300.

- Overcash, J. & Campbell, J. (1955) The effects of intermittent warm and cold periods on breaking the rest period of peach leaf buds. .
- Pan, W., Li, J., Du, Y., Zhao, Y., Xin, Y., Wang, S., Liu, C., Lin, Z., Fang, S., Yang, Y. *et al.* (2023) Epigenetic silencing of callose synthase by vill promotes bud-growth transition in lily bulbs. *Nature Plants* **9**, 1451–1467.
- Pan, W., Liang, J., Sui, J., Li, J., Liu, C., Xin, Y., Zhang, Y., Wang, S., Zhao, Y., Zhang, J. *et al.* (2021) Aba and bud dormancy in perennials: current knowledge and future perspective. *Genes* **12**, 1635.
- Piao, S.L., Liu, Z., Wang, T., Peng, S.S., Ciais, P., Huang, M.T., Ahlstrom, A., Burkhardt, J.F., Chevallier, F., Janssens, I.A., Jeong, S.J., Lin, X., Mao, J.F., Miller, J., Mohammad, A., Myneni, R.B., Penuelas, J., Shi, X.Y., Stohl, A., Yao, Y.T., Zhu, Z.C. & Tans, P.P. (2017) Weakening temperature control on the interannual variations of spring carbon uptake across northern lands. *Nature Climate Change* **7**, 359–+.
- Pörtner, H.O., Roberts, D.C., Tignor, M., Poloczanska, E.S., Mintenbeck, K., Alegría, A., Craig, M., Langsdorf, S., Löschke, S., Möller, V., Okem, A. & Rama, B. (2022) *Climate Change 2022: Impacts, Adaptation and Vulnerability. Contribution of Working Group II to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press.
- Richardson, E. (1974) A model for estimating the completion of rest for ‘Redhaven’ and ‘Elberta’ peach trees. *HortScience* **9**, 331–332.
- Rinne, P.L.H., Paul, L.K. & van der Schoot, C. (2018) Decoupling photo- and thermoperiod by projected climate change perturbs bud development, dormancy establishment and vernalization in the model tree populus. *BMC Plant Biology* **18**.
- Satake, A., Nagahama, A. & Sasaki, E. (2022) A cross-scale approach to unravel the molecular basis of plant phenology in temperate and tropical climates. *New Phytologist* **233**, 2340–2353.
- Schwartz, M.D. (1994) Monitoring global change with phenology – the case of the spring green wave. *International Journal of Biometeorology* **38**, 18–22.
- Sønsteby, A. & Heide, O.M. (2014) Chilling requirements of contrasting black currant (*Ribes nigrum* L.) cultivars and the induction of secondary bud dormancy. *Scientia Horticulturae* **179**, 256–265.
- Tylewicz, S., Petterle, A., Marttila, S., Miskolczi, P., Azeez, A., Singh, R.K., Immanen, J., Mähler, N., Hvidsten, T.R., Eklund, D.M. *et al.* (2018) Photoperiodic control of seasonal growth is mediated by aba acting on cell-cell communication. *Science* **360**, 212–215.
- van der Schoot, C., Paul, L.K. & Rinne, P.L.H. (2014) The embryonic shoot: a lifeline through winter. *Journal of Experimental Botany* **65**, 1699–1712.
- Vitasse, Y., Hoch, G., Randin, C.F., Lenz, A., Kollas, C., Scheepens, J.F. & Koerner, C. (2013) Elevational adaptation and plasticity in seedling phenology of temperate deciduous tree species. *Oecologia* **171**, 663–678.

- Walde, M.G., Wenden, B., Chuine, I., Gessler, A., Saurer, M. & Vitasse, Y. (2024) Stable water isotopes reveal the onset of bud dormancy in temperate trees, whereas water content is a better proxy for dormancy release. *Tree Physiology* **44**, tpae028.
- Walde, M.G., Wu, Z., Fox, T., Baumgarten, F., Fu, Y.H., Wang, S. & Vitasse, Y. (2022) Higher spring phenological sensitivity to forcing temperatures of asian compared to european tree species under low and high pre-chilling conditions. *Frontiers in Forests and Global Change* **5**, 1063127.
- Weinberger, J.H. (1950) Chilling requirements of peach varieties. *Proceedings of the American Society for Horticultural Science* **56**, 122–128.
- Wilczek, A.M., Roe, J.L., Knapp, M.C., Cooper, M.D., Lopez-Gallego, C., Martin, L.J., Muir, C.D., Sim, S., Walker, A., Anderson, J., Egan, J.F., Moyers, B.T., Petipas, R., Giakountis, A., Charbit, E., Coupland, G., Welch, S.M. & Schmitt, J. (2009) Effects of genetic perturbation on seasonal life history plasticity. *Science* **323**, 930–934.
- Wolkovich, E.M., Auerbach, J., Chamberlain, C.J., Buonaiuto, D.M., Ettinger, A.K., Morales-Castilla, I. & Gelman, A. (2021) A simple explanation for declining temperature sensitivity with warming. *Global Change Biology* **27**, 4947–4949.
- Yu, S., Wenden, B., Ferguson, L. & Tian, L. (2024) Building climate resilient deciduous tree crops by deciphering winter dormancy.
- Zhao, Y., Antoniou-Kourounioti, R.L., Calder, G., Dean, C. & Howard, M. (2020) Temperature-dependent growth contributes to long-term cold sensing. *Nature* **583**, 825–829.
- Zhu, P., Lister, C. & Dean, C. (2021) Cold-induced arabidopsis frigida nuclear condensates for flc repression. *Nature* **599**, 657–661.