

Exploring grapevine phenology and high temperatures response under controlled conditions

N K. Merrill^{1, 2, 3*}, I García de Cortázar-Atauri⁴, A K. Parker⁵, M A. Walker⁶, E M. Wolkovich^{1, 2, 7}

¹Arnold Arboretum, Harvard University, United States, ²Department of Organismic and Evolutionary Biology, Faculty of Arts and Sciences, Harvard University, United States, ³Department of Geography, College of Arts and Sciences, University of Oregon, United States, ⁴Institut National de la Recherche Agronomique (INRA), France, ⁵Department of Wine, Food and Molecular Biosciences, Faculty of Agriculture and Life Sciences, Lincoln University, New Zealand, ⁶Department of Viticulture and Enology, University of California, Davis, United States, ⁷Department of Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, Canada

Submitted to Journal:
Frontiers in Environmental Science

Specialty Section:
Biogeochemical Dynamics

Article type:
Brief Research Report Article

Manuscript ID:
516527

Received on:
16 Dec 2019

Revised on:
23 Mar 2020

Frontiers website link:
www.frontiersin.org

Conflict of interest statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

Author contribution statement

EMW and NKM designed the methods for the experiment with input from IGdCA, AKP, and MAW. NKM collected data, and NKM and EMW performed analysis. NKM wrote the manuscript, and EMW, IGdCA, AKP, and MAW edited it.

Keywords

Phenology, Climate Change, Heat stress, flowering, *Vitis vinifera* subsp. *vinifera*, lab conditions

Abstract

Word count: 200

Climate change has challenged growers and researchers alike to better understand how warm temperatures may impact winegrape plant development across varieties. Yet multi-variety studies present challenges. Here we review studies of controlled warming on winegrape varieties alongside a new study of the budburst and flowering phenology of 50 varieties of *Vitis vinifera* subsp. *vinifera* in the lab, with a small set of plants exposed to higher temperatures (20, 26, 30, 34, 37°C mean temperatures in growth chambers) during flowering. We found few studies have examined more than one variety, which may be due to the challenge of growing diverse varieties together. Indeed, we found high variability in flowering success across varieties in the lab (28 out of 50 varieties had no flowering), which made it impossible to study variety-specific response to temperature. Across varieties, however, we found results in line with our literature review: higher temperatures did not have a significant effect on the rate at which vines progressed through the flowering stage, but higher temperatures did correlate with flower abortion. These results suggest a potential decrease in winegrape yields in a warmer climate due to flower abortion, but also highlight the challenges of understanding heat responses across many varieties.

Contribution to the field

The viticulture industry is currently faced with the task of adapting to a changing climate. Phenological timing, largely controlled by temperature, varies by winegrape variety, such that certain varieties may be better suited to produce high quality wine under a warmer climate. However, there is relatively little phenological data on most varieties of *Vitis vinifera* subsp. *vinifera*. We studied budburst and flowering phenology of 50 varieties of winegrape in a greenhouse. We found that the budburst and leafout phenology of greenhouse-grown vines significantly correlates to field observations, but many (28/50) varieties did not flower in the greenhouse. Of the varieties that formed inflorescences, ten were studied in growth chambers set at different temperatures (20 to 37 °C) to assess the rate the vines progressed from inflorescence development through flowering. While phenological timing was not significantly affected, plants in higher temperature chambers were more likely to abort their flower buds before flowering. This suggests that one potential outcome of climate change could be lower yields due to abortion of flower buds. Yet our research also highlights the challenges of growing diverse varieties in the greenhouse and suggests more studies on larger vines will be critical for understanding and exploiting variety diversity.

Funding statement

This research was funded by Harvard University.

Ethics statements

Studies involving animal subjects

Generated Statement: No animal studies are presented in this manuscript.

Studies involving human subjects

Generated Statement: No human studies are presented in this manuscript.

Inclusion of identifiable human data

Generated Statement: No potentially identifiable human images or data is presented in this study.

In review

Data availability statement

Generated Statement: All datasets generated for this study are included in the manuscript/supplementary files.

In review

Submitted as a Brief Research Report to Research Topic: Biogeosciences and Wine: the Management and Environmental Processes that Regulate the Terroir Effect in Space and Time

Main body text word count: 3,725

Figures: 3

Table: 1

Exploring grapevine phenology and high temperatures response under controlled conditions

N. K. Merrill^{*1-3}, I. García de Cortázar-Atauri⁴, A. Parker⁵, M. A. Walker⁶ & E. M. Wolkovich^{1-2,7}

Author affiliations:

¹Arnold Arboretum of Harvard University, Boston, MA, USA

²Organismic & Evolutionary Biology, Harvard University, Cambridge, MA, USA

³Department of Geography, University of Oregon, Eugene, OR 97403

⁴INRAE, US AGROCLIM, Avignon, France

⁵Department of Wine, Food and Molecular Biosciences, Faculty of Agriculture and Life Sciences, Lincoln University, PO Box 85084, Lincoln 7647, Christchurch, New Zealand

⁶Department of Viticulture and Enology, University of California, Davis, CA 95616, USA

⁷Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, Vancouver, BC, V6T 1Z4, Canada

Correspondence: N. K. Merrill (nmerrill@uoregon.edu)

Keywords

Phenology, climate change, heat stress, flowering, lab conditions, *Vitis vinifera* subsp. *vinifera*

Abstract

Climate change has challenged growers and researchers alike to better understand how warm temperatures may impact winegrape plant development across varieties. Yet multi-variety studies present challenges. Here we review studies of controlled warming on winegrape varieties alongside a new study of the budburst and flowering phenology of 50 varieties of *Vitis vinifera* subsp. *vinifera* in the lab, with a small set of plants exposed to higher temperatures (20, 26, 30, 34, 37°C mean temperatures in growth chambers) during flowering. We found few studies have examined more than one variety, which may be due to the challenge of growing diverse varieties together. Indeed, we found high variability in flowering success across varieties in the lab (28 out of 50 varieties had no flowering), which made it impossible to study variety-specific response to temperature. Across varieties, however, we found results in line with our literature review: higher temperatures did not have a significant effect on the rate at which vines progressed through the flowering stage, but higher temperatures did correlate with flower abortion. These results suggest a potential decrease in winegrape yields in a

- 43 warmer climate due to flower abortion, but also highlight the challenges of understanding
- 44 heat responses across many varieties.

In review

Introduction

As the climate changes, the viticulture industry needs to adapt to shifting terroir. Terroir—the critical link between the flavor and style of a wine and the characteristics of the environment in which it is grown—is shaped strongly by climate, and the matching of climates to varieties (Van Leeuwen et al., 2019). Thus, as climate change continues to raise temperatures in winegrowing regions across the world, the viticulture industry will be continually challenged to adapt to new terroirs over future decades. Already, the industry has shifted growing areas towards the poles and higher elevations to maintain ideal growing temperatures for winegrapes. This trend is predicted to continue (Schultz and Jones, 2010; Hannah et al., 2013), raising concerns that vineyards could move to land that is currently conserved for biodiversity and ecosystem services (Hannah et al., 2013).

Alternatively, vineyards could take advantage of the high geno- and phenotypic diversity that already exists by planting varieties better suited to the new climate (Ollat et al., 2015, 2016; Wolkovich et al., 2017; Morales-Castilla et al., 2020) or breeding new varieties (Myles, 2013; Duchêne, 2016). *Vitis vinifera* subsp. *vinifera* (winegrape) has at least 6000 genetically distinct varieties grown for many purposes, but only ~1100 are grown currently by the viticulture industry, and an even smaller number dominate the global market (Lacombe, 2012; Anderson, 2013). However, for this adaptation to be effective, growers need better information on how different varieties fare in warmer climate regimes, with phenology being one important component (Ollat et al., 2016).

Studying the phenology of different varieties of winegrapes would help viticulturists better adapt to climate change, because winegrape phenology is extremely sensitive to temperature (Parker et al., 2011, 2013; Jones, 2013; García de Cortázar-Atauri et al., 2017). Timing for leafout and flowering of diverse plant species has advanced six to 20 days in the last 30-40 years of warming (Root et al., 2003; Menzel et al., 2006), equivalent to four to six days per °C. A similar advance is seen for winegrape harvest dates, which can change about 6 days per °C (Cook and Wolkovich, 2016; Labbé, 2019). The time between flowering and veraison also decreased by a little more than 1 day per °C. (Duchêne and Schneider, 2005). In winegrapes, phenological timing varies across varieties, and this variability could be used to better adapt to future climates. Generally, timing of phenology can vary from three to six weeks across varieties (Boursiquot et al., 1995; Wolkovich et al., 2017).

However, most varieties still have little phenological data and far fewer varieties have data from many different environments. In this context, it is difficult to describe where many varieties could best be grown and how they respond to higher temperatures during critical phenological phases, such as flowering. While recent efforts have greatly expanded our resources for understanding phenological responses to climate in the field across varieties—yielding information on approximately 100 varieties (Parker et al., 2011, 2013) this is still less than 10% of currently planted varieties. For growers to select varieties for adapting to shifting terroirs, they will need information on more varieties and across diverse temperature regimes.

A first step towards this goal is research on an increased number of varieties and an understanding of whether phenology in semi-artificial conditions (i.e., greenhouses, labs, and growth chambers), where temperatures can be controlled more easily, matches field-based phenology. To date, much research has focused on a limited number of varieties (Sepúlveda et al., 1986; Mullins, 1992), making it difficult to know how much results for one variety can be extrapolated to another. Yet, if a greater diversity of varieties can be grown in lab conditions, lab studies could quickly increase our understanding across varieties. Further, if lab phenology appears similar to field phenology, it would suggest such results could be relevant to field conditions. Beyond this first step then, researchers will want to examine how varying temperature regimes affect particular phenological stages.

Understanding how climate change will affect winegrape flowering may be a particularly important aspect of the overall effect on phenology and the impact of temperature on the flowering process will ultimately influence harvest yields. Studies of vegetative growth and photosynthesis in other perennial crops exposed to a range of temperatures exhibited that extreme temperatures tend to slow or inhibit certain processes in the plants (Zaka et al., 2016, 2017). In this context, we would expect that grapevine flowering development may similarly slow down at higher temperatures.

Here we address these issues through first, a literature review of warming studies on winegrape phenology to examine how many varieties have been studied, over which temperatures and their findings, second, our efforts to examine experimentally how temperature affects flowering in a variety-rich study. To explore this second issue, we had two major aims: (1) to test whether the phenological stages of budburst and leafout in lab conditions correlated with field phenology for 50 varieties in the lab, and (2) to examine the effect of higher temperatures on flowering development, by following the flowering response of a small subset of these varieties across mean temperatures of 20 °C to 34 °C in growth chambers. Overall, we aim to provide both an overview of experiments to date, and to outline how our findings and challenges may guide future efforts to conduct variety-rich lab experiments.

Materials and Methods

Literature review:

We conducted a literature review by searching Google Scholar, ISI Web of Science and ScholarOneSearch for searches including 'Vitis vinifera' with combinations of 'heat toleran*', 'growth chamber', 'phenolog*', 'temperature manipulation'. Then we reviewed papers that experimentally manipulated temperatures of growing grapevines and reported phenological responses (excluding all studies without experimental warming or of warming applied to dormant cuttings or plants or focused only on berry ripening). We additionally included any relevant papers of which we were aware that we did not find in these searches. While some studies included additional treatments (e.g., drought, CO₂ manipulation) we focus on results relating to warming and phenology.

Variety-rich study:

Observations of field-grown winegrapes in the UC Davis Robert Mondavi Institute (RMI) Vineyard (Davis, California, USA) using the modified Eichorn-Lorenz (EL) scale (Coombe, 1995) began 6 March 2015 and continued generally every 3-4 days until 2 April 2015, when almost all plants had reached EL stage 11 or higher (data and full methods available at: <https://knb.ecoinformatics.org/view/doi:10.5063/F18G8J29>). Dormant winegrape cuttings were then taken in December of 2015.

Following collection, cuttings were chilled for 21 days (4° C) at the Arnold Arboretum (Boston, Massachusetts, USA), then forced in greenhouses in 26 cm diameter (9.6L) pots in January 2016 (9.6L). After several months of growth, on 27 May they were placed in growth chambers with day/night temperatures of 6/4 °C and an 8-hour photoperiod to induce dormancy, though the plants did not appear visibly dormant until 20 June 2016.

On 15 August 2016, the 351 potted cuttings were moved out of the chambers and into a greenhouse where the initial day temperature was 18.5 ± 1.5 °C and night temperature was 16.75 ± 1.25 °C. After the first week, the temperatures were slowly raised to 25.5 ± 2.5 °C during the day and lowered to 10 °C at night. The cuttings were pruned the day they were removed from the chambers so that each cutting had two spurs and each spur had two nodes. Then, the diameter of each spur and node and the distance between the two nodes on each spur were measured with calipers. About every two days, the plants' soil was checked for moisture, and they were watered as needed to keep soils moist. Starting 1 October, plants were also fertilized once a week with a 50% dilution.

Twice a week, beginning 22 August, each plant's development was recorded using the modified Eichorn-Lorenz scale (Coombe, 1995) and soil moisture was measured with a probe in three locations in each pot. Each spur was kept at two shoots, but only the dominant shoot on each spur had observations recorded. Each shoot was trained up a stake for support. When an inflorescence had developed (EL stage 12), the plant was randomly assigned to one of five growth chambers if it was a part of the heat tolerance experiment. Otherwise, observations on each plant continued in the greenhouse. Varieties were chosen for inclusion in the experiment to include a diversity of phenology from those varieties for which there were five or more replicates growing.

The five chambers all had a 12-hour photoperiod with $800 \text{ m}^{-2}\text{s}^{-1}$ of fluorescent light, but varied in their temperature: Chamber 1 was set at 17/23 °C Chamber 2 was set at 23/29 °C, Chamber 3 was set at 27/33 °C, Chamber 4 was set at 31/37 °C, and Chamber 5 was set at 34/40 °C (all temperatures given as night/day). Initially, CO₂ levels were set at 400 ppm during the day and 600 ppm at night, because plants respire at night, increasing CO₂ levels. Each inflorescence was contained in a paper bag to collect the flower caps as they fell. Every 10 days, the plants and their assigned temperatures were rotated to a new chamber to minimize individual chamber effects on the experiment.

Observations of the percent of flower buds that flowered on each inflorescence (% flowering), leaf number, stem length, and number of fallen flower caps were made three times a week, along with soil moisture. On 19 September, it was noted that some inflorescence bags also contained aborted buds that had yet to flower, and thereafter observations of aborted buds were also recorded. Once a plant had reached 100% flowering, or, in the case of plants where the entire inflorescence had abscised, each plant had spent a minimum 14 days in the chamber, it was returned to the greenhouse. No further observations were made once no more plants were developing inflorescences and all plants in the chambers had finished flowering.

To determine if there was any correlation between the chamber temperatures and the other variables, we used ANOVA. Linear regression was used to compare the development of the plants in the greenhouse with the data collected in the RMI Vineyard growing season. All analyses were performed in R version 3.3.3 (R Core team, 2013). Given limited replicates per variety all analyses of the growth chamber study were done across varieties.

Results

Literature review:

Most studies (7/10) examined only one variety, while at most one study examined 5 varieties. Further, as certain varieties were studied often (e.g., Semillon, Shiraz and Cabernet-Sauvignon) all 10 studies yield information on only a total of 10 varieties. Experimental warming was split between being applied in the vineyard (through passive and active warming) or in the lab (growth chambers or greenhouses) with temperatures generally ranging from 20-40°C, while some field conditions exceeded 40°C. Warming generally advanced phenology, save for one field study that showed temperatures above 40°C delayed veraison (Greer and Weedon, 2013). Studies focused on flowering found decreased flowering at higher temperatures applied near budburst (Petrie and Clingeleffer, 2005) and flower abscission at higher temperature applied during flowering (Greer and Weston, 2010).

Variety-rich study:

The plants underwent budbreak (EL 4) between 17 August and 6 September (mean = 29 August) and leafout (EL 7) between 22 August and 22 September (mean = 4 September). Budbreak and leafout timing among the varieties were similar in the lab and field (Figure 1, budburst: $F(1,47)=14.55$, $p<0.001$; leafout: $F(1,47)=18.51$, $p<0.001$). The first inflorescence formed on 5 September, and 51 plants reached this stage (EL 12) later, with substantial variation in terms of the number of plants of each variety that flowered at all (Table 1). Most varieties (28/50 total) did not form inflorescences, while for a few varieties nearly half of the plants underwent flowering (e.g., Sauvignon blanc, Tempranillo, Verdelho). Due to this high variation in inflorescence appearance, only 26 of the flowering plants were used in the experiment corresponding to 10 varieties.

Given the low number of plants that formed inflorescence, most varieties could be placed in only one or two temperature treatments (with very low or no replication per variety: chamber 1 (20°C) had one plant each of Cabernet Sauvignon, Durif, Sauvignon Blanc, and Verdelho. Chamber 2 (26°C) had one plant each of Durif, Pinot Gris, Sauvignon Blanc, and Verdelho. Chamber 3 (30°C) had three Durif plants, then one plant each of Gewürztraminer, Tempranillo, and Verdelho. Chamber 4 (34°C) had two Tempranillo plants, then one each of Dolcetto, Pinot Gris, Sauvignon Blanc, Syrah, and Verdelho. Chamber 5 (mean of 37°C) had two Tempranillo plants, and one each of Sauvignon Blanc, Verdelho, and Vinhão). Because of the limited number of replicated per variety, we do not report variety-specific estimates and all statistics are done across varieties. Plants that had thicker spurs were more likely to develop inflorescence ($Z(340)=2.21$, $p=0.03$), and more likely to reach 50% flowering (Figure 2, $Z(340)=2.85$, $p=0.004$).

Soil moisture in the chambers varied by chamber temperature ($F(1,24)=8.05$, $p=0.01$), ranging from 69% to 76% over time. There was no directional relationship between the moisture levels and the chamber temperature (i.e., the warmest chambers were not the driest) and means were similar across treatments, ranging from 71% to 74%.

There was also no directional relationship between chamber temperature and either change in stem length or leaf appearance rate (stem length: $F(1,24)=0.53$, $p=0.47$; leaf appearance: $F(1,24)=0.05$, $p=0.83$).

Chamber temperatures did not affect the time it took for the plants to reach 10% and 50% flowering and there was no trend in the duration of flowering (Figure 3, 10%: $F(1,20)=0.43$, $p=0.52$; 50%: $F(1,15)=0.50$, $p=0.49$). Within treatments, the number of days after forcing it took plants to reach 10% flowering ranged from 34 to 51 days (mean = 42.6 ± 0.9).

The number of flower buds aborted per plant was significantly affected by the chamber temperature (Figure 3, $F(1,24)=7.43$, $p=0.01$). The two warmest chambers saw the greatest number of flower buds lost during the time in the chamber, with the greatest average number of flower buds aborted seen in 37 °C treatment (mean number of flower buds aborted at 20 °C: 4.5, 26 °C: 2.8, 30 °C: 5.8, 34 °C: 27.6, 37 °C: 57.3).

Discussion

Increasingly, winegrape diversity is suggested as a way for growers to cope with warming, but we know little about how experimental warming temperature differentially affects most varieties (Ollat et al., 2015, 2016; Wolkovich et al., 2017). Research to date has focused on very few (only 10 according to our literature review) varieties, but suggests responses vary depending on variety. For example, Greer & Weedon found a curvilinear ripening response to temperature increased across three varieties—but the temperature yielding the highest ripening varied for each variety (25°C, 35°C and 40°C for Chardonnay, Semillion and Merlot, respectively; 2014). Such variation is critical for growers who want to adapt to warming by shifting varieties, but to make useful variety recommendations we need more information on how temperature affects development across varieties and developmental stages. Our lab work

on 50 varieties, however, highlights the challenges of growing diverse varieties for experimental research.

Effects of high temperatures on winegrape flowering

Our lab work to examine how temperature affects flowering across diverse varieties failed to produce enough grapevines to study variety-specific effects. Yet, in pooling results across varieties, we found trends in line with previous studies.

Overall, we studied the effects of temperatures between a minimum of 17°C and maximum of 37 °C (means of 20 °C to 34 °C) on flowering for 26 winegrape plants. We found no directional relationship between temperature and soil moisture, stem length, leaf number, or the number of days it took to reach 10% or 50% flowering. Contrary to expectations of most phenological models (García de Cortázar-Atauri et al., 2010; Cuccia et al., 2014) and one previous growth chamber studies (Buttrose and Hale, 1973), we found that flowering phenology was not significantly delayed in either the coldest or warmest chambers. This result, however, is in line with much of our literature review which found growth and phenology generally advanced up to 40°C (Table 1). Further, our results support previous work, which found that plants in the hotter treatments aborted a higher number of flowers than those in the cooler treatments (Greer and Weston, 2010). This abortion, because it translated to fewer observations of higher percentages of flowering (i.e., 50%), may have limited detection of slowed phenology at higher temperatures. Furthermore, our plants were only exposed to the higher temperatures during flowering, not before, which could have diminished potential differences in timing of phenology during that developmental phase.

The majority of literature on winegrape heat tolerance focuses on the effects of heat on berry ripening. In their 2010 study of Semillon winegrapes, Greer and Weston noted that plants treated with elevated temperatures at fruit set were much less vulnerable and suffered few ill-effects when compared with those treated at flowering, veraison, and mid-ripening. When heat-treated at fruit set, berry growth was unimpeded and sugar content increased normally. This could mean that winegrapes are more vulnerable to high temperatures during certain periods of development, i.e. flowering. If winegrapes are especially susceptible to heat during flowering, viticulturists will have to take extra precautions during this period to ensure the survival of the flowers through to fruit set.

Although we did not measure fruit-set, future studies may want to investigate how it could be affected by elevated temperatures during the flowering period. There could be a delay in response between the period of warming and the effects of high temperatures that was not seen in our experiment because the plants were heated during the developmental phase in which we were interested. Continuing observations through fruit set could be an important next step to help understand more exactly how harvest yields might be impacted in a warming climate.

Utility of lab-grown winegrape plants for future research

Because the majority of the plants' development did not progress to the flowering stage (EL stage 11), sample sizes for our heat experiment were smaller than planned (each chamber had four to six plants). This meant there were not enough plants of each variety in each chamber to test for a difference in varietal response to the heat treatments, and instead we analysed our findings across varieties (as most varieties were only represented in a single treatment). Still, it is important to note that we studied ten different varieties in the chambers, which greatly increased the genetic diversity of the experiment. It has been shown that controlled ecological experiments in labs that include greater genetic diversity are more easily replicated (Milcu et al., 2018).

Further, we found high variation in flowering success—plants with larger spurs were more likely to form inflorescence and flower and some varieties were far more successful in flowering than others. This suggests plants with greater carbohydrate reserves were more likely to develop inflorescence and flower, similar to the results of Eltom's study of the effects of girdling and leaf removal on inflorescence development (2013), but with additional variation across varieties, as other studies have found (Lebon et al., 2005). Thus, future experiments may want to (at least initially) focus lab efforts on these more successful varieties and tease out high and low temperature limits to help guide further research.

The rate of development seen in the plants grown in the greenhouse was significantly correlated with that seen in the winegrapes grown in the Robert Mondavi Institute Vineyard, from which the cuttings in this experiment were taken (Figure 1). This suggests that the overall progression and timing of phenological development was not dramatically altered by the lab setting and supports the use of potted plants in the lab used alongside field data to better understand and predict winegrape responses to climate change. Our finding that plants with larger spurs were more likely to flower, however, suggests that our results regarding flower development in the greenhouse and flowering (and flower abortion) in the growth chambers should be interpreted cautiously.

Our vines, taken from field cuttings, were in only their first growing season, and we expect flowering success across varieties would be greater for older, larger vines. In the literature studies vary in using <1 year-old potted cutting, to 3-to 5-year-old potted vines, to established vineyard plants. This diversity of vine age across studies that also vary treatments makes it difficult to attribute variation in findings to age, but our results suggest older vines may be most relevant and useful for studies on heat tolerance and warming effects.

While this study was unable to adequately address varietal differences in response to warming as a result of climate change, it provided valuable insight into challenges of variety-rich winegrape studies. Based on the outcome of our study, we recommend the following strategies to improve the success of similar future studies: (1) Use older vines or those with thicker diameters (which indirectly corresponds to greater carbohydrate reserves) to ensure a higher number of plants form inflorescences and undergo flowering. (2) Consider mesh bags to trap flowers; because we contained inflorescences in paper bags, we may have restricted air flow during a critical period of development, limiting photosynthesis. (3) Examine effects of

gradual versus sudden temperature increases. Providing a transitional period for plants when they are moved into chambers and raising temperatures gradually could prevent shock or stress on the plants that could exacerbate flower abscission, but a more sudden temperature changes may be relevant for weather changes with climate change (Gouot et al., 2019).

Conclusions

Helping growers adapt to shifting terroirs requires research on a greater diversity of *Vitis vinifera* varieties across diverse temperature regimes. Here we showed that budburst and leafout phenology of 50 varieties grown in the field correlated with field-based phenology and that higher temperatures can negatively impact flowering. While heat treatments during flowering did not affect the phenology of the grapes we studied, we found a significant impact from the elevated temperatures on flower abortions, in line with previous studies, which could lead to substantial negative impacts on yield. Despite the difficulties we faced implementing a variety-rich experiment, lessons we learned can inform future studies to increase success and provide further guidance for academics and professionals alike. Our findings underscore the importance of modeling more than the plants' phenology to fully understand the impacts climate change will have on the viticulture industry. As data across more diverse varieties and temperature regimes increases, it can help support mapping when and where different varieties may perform best as warming continues.

Acknowledgements

We thank R. Antolick for assistance with the literature review, E. Forrestel for taking cuttings and starting plants in the greenhouse, and S. Sutphin for data collection in the field in Davis, and two anonymous reviewers for comments that improved the manuscript.

Figures (below)

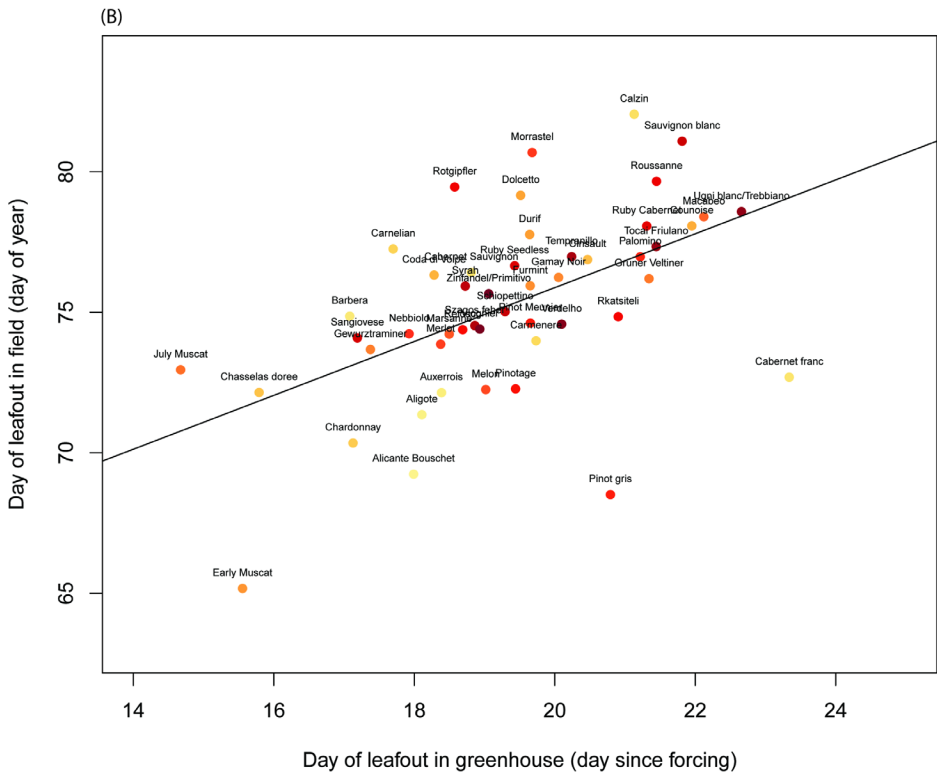
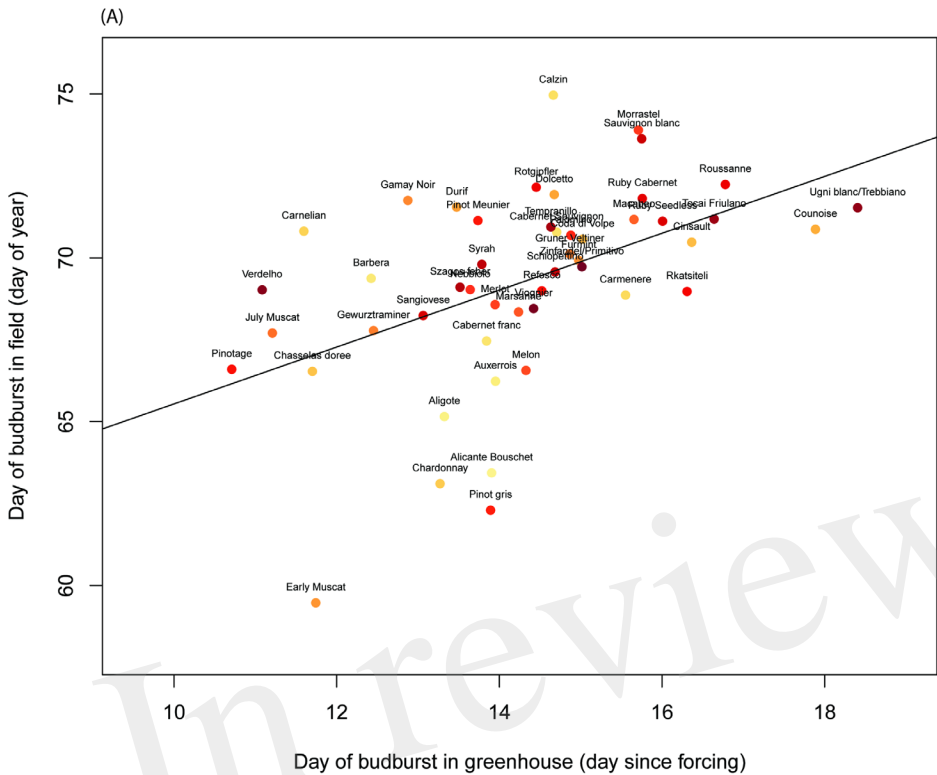
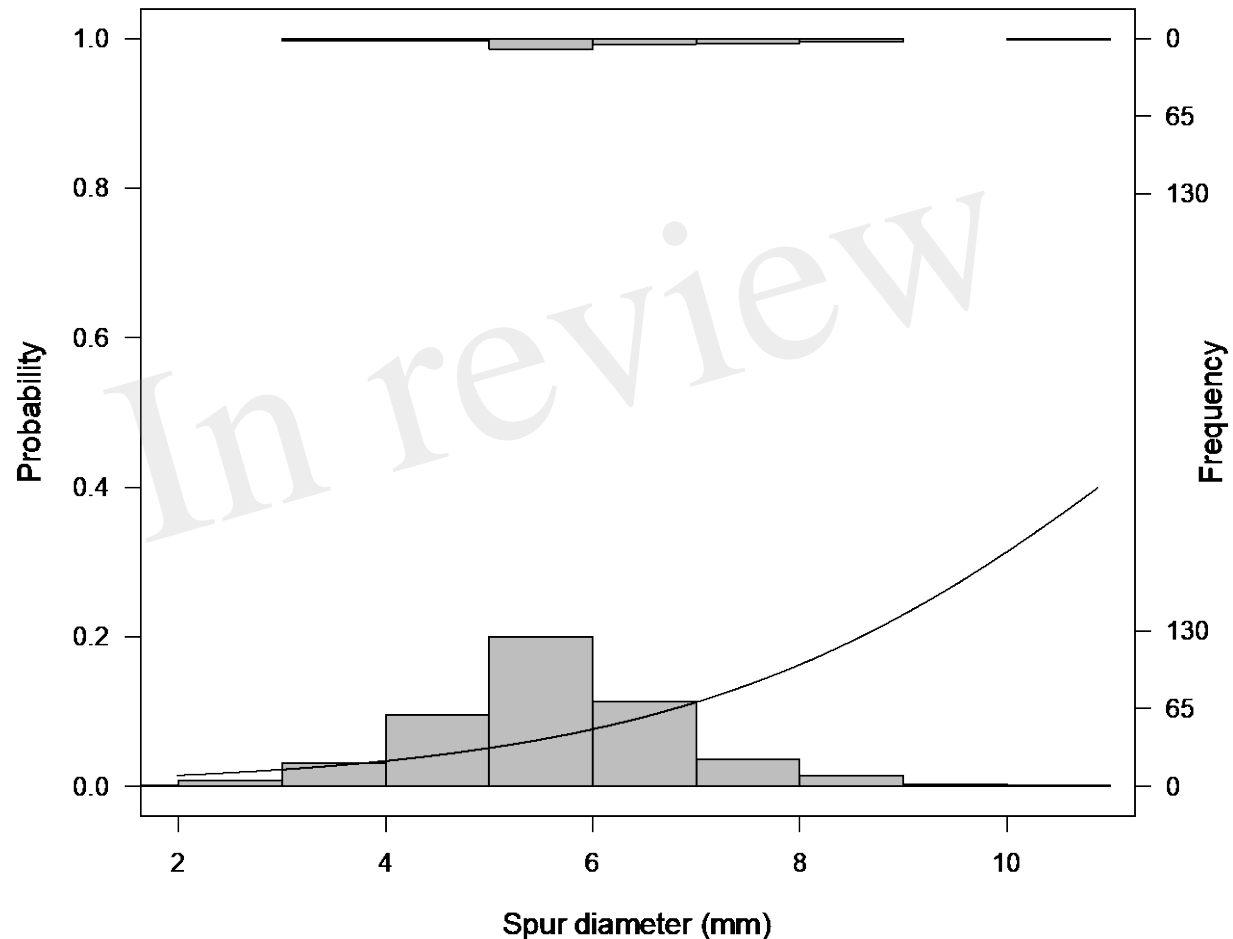


Figure 1

Day of budburst (A) and leafout (B) in the Robert Mondavi Institute Vineyard (Davis, California, USA) from the 2015 growing season correlates to the day of budburst ($F(1,47)=14.55$, $p<0.001$) and leafout ($F(1,47)=18.51$, $p<0.001$) in greenhouse conditions across 50 varieties (each point represents a different variety that was grown both in the vineyard and in the greenhouse).

**Figure 2**

Spur diameter in greenhouse-grown vines (measured when plants were removed from dormancy) related to the probability that a plant would reach 50% flowering ($Z(340)=2.85$, $p=0.004$), with larger spur vines more often reaching 50% flowering. Histograms show the vines that did not reach 50% flowering (recorded in this analysis as 0 values, bottom) and those that did reach 50% flowering (recorded in this analysis as 1 values, top).

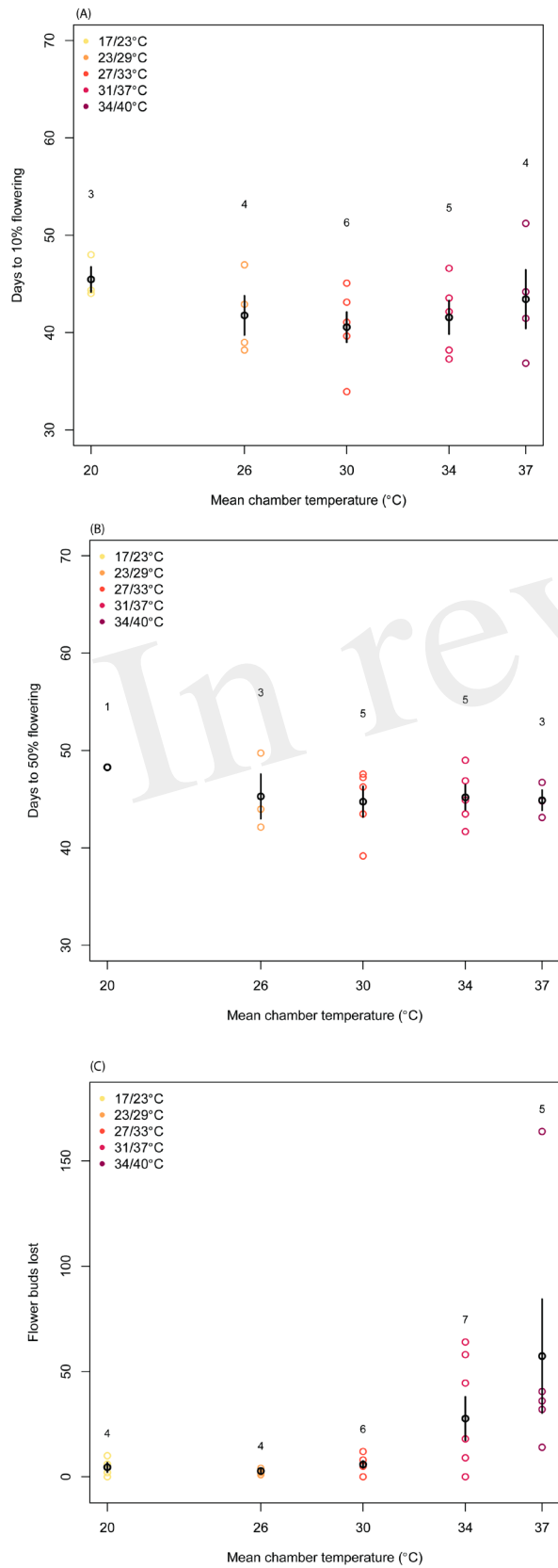


Figure 3

These figures illustrate the relationship between mean chamber temperature and (A) the days it took the plants to reach 10% flowering ($F(1,20)=0.43$, $p=0.52$), (B) the days it took the plants to reach 50% flowering (50%: $F(1,15)=0.50$, $p=0.49$), or (C) the number of flower buds lost while in the chamber ($F(1,24)=7.43$, $p=0.01$). The black points and bars show the average and error in each chamber. The number above each chamber's data is the sample size. The colored points represent individual plants. The legend in the top left corner gives the night/day temperature for each chamber.

In review

Table 1

Literature review of studies applying experimental warming to winegrapes during development and following phenological responses. We provide the rate at which temperature treatments were applied parenthetically.

Paper	Varieties	Type	Temperature	Effects	Vine age
Edwards <i>et al.</i> 2017	Shiraz	Field experimental warming (passive chambers)	2°C warming from average temperature (passive heating)	All aspects of vine phenology advanced	"mature"
Gouot <i>et al.</i> 2019	Shiraz	System to heat only aboveground parts of plants	+6°C at end of fruit set and again prior to veraison (immediate)	Photosynthesis decreased when heating led to 45°C temperatures, but not when only 40°C	7 years
Greer & Weedon 2013	Semillon	Field experimental cooling	Some vines protected from 40+°C ambient temperatures (passive heating)	Heat delayed ripening	6 years
Greer & Weedon 2014	Merlot, Chardonnay, Semillon	Growth chambers	20-40°C range, 4 treatments, post-veraison (temperatures raised in chamber 10 days after plants were allocated to chambers)	Growth effects varied by variety: Merlot: no effect on berry, Chardonnay: rapid expansion at 20 and 25°C but decline in size at 40°C, Semillon: expansion at 20 and 25°C but not at higher temps	5 years
Greer & Weston 2010	Semillon	Growth chambers	40/25°C at flowering, fruit set, veraison and mid-ripening stages (immediate)	Heat did not affect leaf growth or stem extension, but flowers completely abscised. Berries treated at fruit set developed normally and those treated at veraison and mid-ripening stopped expanding and sugar content stopped increasing	3 years
Kadir <i>et al.</i> 2006	Semillon, Pinot Noir, Chardonnay, Cabernet Sauvignon, Cynthiana (<i>V. aestivalus</i>)	Growth chambers	20-40°C range, 3 treatments (immediate)	Growth effects increase in vegetative growth for <i>V. vinifera</i> from 20 to 30°C, but most growth stunted at 40°C -- <i>V. vinifera</i> affected less by high temperatures	1 year

Winegrape phenology and heat stress

Kliewer 1977	Cabernet Sauvignon, Tokay, Pinot Noir, Carignane	Growth chambers	35, 40°C warming during 2-8 days before to 12-18 days after bloom 25/20°C controls (immediate)	Variable effects on berry set and weight, depending on variety; no effect on rate of shoot growth	3-4 years
Petrie & Clingeleffer 2005	Chardonnay	Field experimental warming (passive chambers)	Range of 3°C across treatments (passive heating)	Resulted in decrease in flowering of 15 to 25% due to temperature	"established vineyard"
Salazar-Parra et al. 2010	Tempranillo	Greenhouse	28/18°C vs. 24/14°C, day/night at veraison (immediate)	Warming shortened the time between grape veraison and full maturity	<1 year
Soar <i>et al.</i> 2009	Shiraz	Field experimental warming (chambers with fans)	6.5-7.3°C above ambient for 3 days (passive heating)	No effect on berry growth or sugar accumulation	10 years

410
411
412

REFERENCES

- Anderson, K. (2013). *Which winegrape varieties are grown where? : a global picture*. Adelaide, South Australia: University of Adelaide Press.
- Boursiquot, J. M., Dessup, M., and Rennes, C. (1995). Distribution des principaux caracteres phenologiques, agronomiques et technologiques chez *Vitis vinifera* L. *Vitis* 34, 31–35.
- Buttrose, M. S., and Hale, C. R. (1973). Effect of Temperature on Development of the Grapevine Inflorescence after Bud Burst. *Am. J. Enol. Vitic.* 24, 14.
- Cook, B. I., and Wolkovich, E. M. (2016). Climate change decouples drought from early wine grape harvests in France. *Nat. Clim. Change* 6. doi:10.1038/nclimate2960.
- Coombe, B. G. (1995). Growth Stages of the Grapevine: Adoption of a system for identifying grapevine growth stages. *Aust. J. Grape Wine Res.* 1, 104–110. doi:10.1111/j.1755-0238.1995.tb00086.x.
- Cuccia, C., Bois, B., Richard, Y., Parker, A. K., García de Cortázar-Atauri, I., Van Leeuwen, C., et al. (2014). Phenological model performance to warmer conditions: application to Pinot Noir in Burgundy. *J. Int. Sci. Vigne Vin*, 169–178.
- Duchêne, E. (2016). How can grapevine genetics contribute to the adaptation to climate change? *OENO One* 50. doi:10.20870/oeno-one.2016.50.3.98.
- Duchêne, E., and Schneider, C. (2005). Grapevine and climatic changes: a glance at the situation in Alsace. *Agron. Sustain. Dev.* 25, 93–99. doi:10.1051/agro:2004057.
- Edwards, E. J., Unwin, D., Kilmister, R., and Treeby, M. (2017). Multi-seasonal effects of warming and elevated CO₂ on the physiology, growth and production of mature, field grown, Shiraz grapevines. *OENO One* 51, 127. doi:10.20870/oeno-one.2016.0.0.1586.
- Eltom, M. (2013). Influence of temperature and carbohydrate availability on the bunch architecture of *Vitis vinifera* L. sauvignon blanc.
- García de Cortázar-Atauri, I., Chuine, I., Donatelli, M., Parker, A. K., and van Leeuwen, C. (2010). *A curvilinear process-based phenological model to study impacts of climatic change on grapevine (Vitis vinifera L.)*.
- García de Cortázar-Atauri, I., Duchêne, E., Destrac-Irvine, A., Barbeau, G., Rességuier, L. de, Lacombe, T., et al. (2017). Grapevine phenology in France: from past observations to future evolutions in the context of climate change. *OENO One* 51. doi:10.20870/oeno-one.2017.51.2.1622.

- 445 Gouot, J. C., Smith, J., Holzapfel, B., and Barril, C. (2019). Single and cumulative effects of whole-
446 vine heat events on Shiraz berry composition. *OENO One* 53. doi:10.20870/oen-
447 one.2019.53.2.2392.
- 448 Greer, D. H., and Weedon, M. M. (2013). The impact of high temperatures on *Vitis vinifera* cv.
449 Semillon grapevine performance and berry ripening. *Front. Plant Sci.* 4.
450 doi:10.3389/fpls.2013.00491.
- 451 Greer, D. H., and Weston, C. (2010). Heat stress affects flowering, berry growth, sugar
452 accumulation and photosynthesis of *Vitis vinifera* cv. Semillon grapevines grown in a
453 controlled environment. *Funct Plant Biol* 37, 206–214. doi:10.1071/FP09209.
- 454 Greer, D., and Weedon, M. (2014). Temperature-dependent responses of the berry
455 developmental processes of three grapevine (*Vitis vinifera*) cultivars. *N. Z. J. Crop*
456 *Hortic. Sci.* 42, 233–246. doi:10.1080/01140671.2014.894921.
- 457 Hannah, L., Roehrdanz, P. R., Ikegami, M., Shepard, A. V., Shaw, M. R., Tabor, G., et al. (2013).
458 Climate change, wine, and conservation. *Proc. Natl. Acad. Sci.* 110, 6907–6912.
459 doi:10.1073/pnas.1210127110.
- 460 Jones, G. V. (2013). “Winegrape phenology,” in *Phenology: An Integrative Environmental*
461 *Science*, ed. M. D. Schwartz (Springer), 563–584.
- 462 Kadir, S. (2006). Thermostability of Photosynthesis of *Vitis aestivalis* and *V. vinifera*. *J. Am. Soc.*
463 *Hortic. Sci.* 131, 476–483. doi:10.21273/JASHS.131.4.476.
- 464 Kliewar, W. M. (1977). Effect of high temperatures during the bloom-set period on fruit-set,
465 ovule fertility, and berry growth of several grape cultivars. *Am. J. Enol. Vitic.* 28.
- 466 Labbé, T. P. (2019). The longest homogenous series of grape harvest dates, Beaune 1354-2018,
467 and its significance for the understanding of past and present climate. *Clim. Past*, 1485–
468 1501. doi:https://doi.org/10.5194/cp-15-1485-2019.
- 469 Lacombe, T. (2012). Contribution à l’étude de l’histoire évolutive de la vigne cultivée (*Vitis*
470 *vinifera* L.) par l’analyse de la diversité génétique neutre et de gènes d’intérêt. in.
- 471 Lebon, G., Duchêne, E., Brun, O., and Clément, C. (2005). Phenology of Flowering and Starch
472 Accumulation in Grape (*Vitis vinifera* L.) Cuttings and Vines. *Ann. Bot.* 95, 943–948.
473 doi:10.1093/aob/mci108.
- 474 Menzel, A., Sparks, T. H., Estrella, N., Koch, E., Aasa, A., Ahas, R., et al. (2006). European
475 phenological response to climate change matches the warming pattern. *Glob. Change*
476 *Biol.* 12, 1969–1976. doi:10.1111/j.1365-2486.2006.01193.x.

- 477 Milcu, A., Puga-Freitas, R., Ellison, A. M., Blouin, M., Scheu, S., Freschet, G. T., et al. (2018).
478 Genotypic variability enhances the reproducibility of an ecological study. *Nat. Ecol. Evol.*
479 2, 279–287. doi:10.1038/s41559-017-0434-x.
- 480 Morales-Castilla, I., García de Cortázar-Atauri, I., Cook, B. I., Lacombe, T., Parker, A., van
481 Leeuwen, C., et al. (2020). Diversity buffers winegrowing regions from climate change
482 losses. *Proc. Natl. Acad. Sci.* 117, 2864–2869. doi:10.1073/pnas.1906731117.
- 483 Mullins, M. G. (1992). *Biology of the grapevine*. Cambridge: Cambridge.
- 484 Myles, S. (2013). Improving fruit and wine: what does genomics have to offer? *Trends Genet.*
485 29, 190–196. doi:http://dx.doi.org/10.1016/j.tig.2013.01.006.
- 486 Ollat, N., Quénel, H., Barbeau, G., van Leeuwen, C., Darriet, P., García de Cortázar-Atauri, I., et
487 al. (2015). Adaptation to Climate Change: strategic issues to face for the French wine
488 industry.
- 489 Ollat, N., Touzard, J.-M., and van Leeuwen, C. (2016). Climate Change Impacts and Adaptations:
490 New Challenges for the Wine Industry.
- 491 Parker, A., García de Cortázar-Atauri, I., Chuine, I., Barbeau, G., Bois, B., Boursiquot, J.-M., et al.
492 (2013). Classification of varieties for their timing of flowering and veraison using a
493 modelling approach: A case study for the grapevine species *Vitis vinifera* L. *Agric. For.*
494 *Meteorol.* 180, 249–264. doi:https://doi.org/10.1016/j.agrformet.2013.06.005.
- 495 Parker, A. K., De Cortazar-Atauri, I. G., Van Leeuwen, C., and Chuine, I. (2011). General
496 phenological model to characterise the timing of flowering and veraison of *Vitis vinifera*
497 L. *Aust. J. Grape Wine Res.* 17, 206–216. doi:10.1111/j.1755-0238.2011.00140.x.
- 498 Petrie, P. R., and Clingeleffer, P. R. (2005). Effects of temperature and light (before and after
499 budburst) on inflorescence morphology and flower number of Chardonnay grapevines
500 (*Vitis vinifera* L.). *Aust. J. Grape Wine Res.* 11, 59–65. doi:10.1111/j.1755-
501 0238.2005.tb00279.x.
- 502 R Core team (2013). *R: A language and environment for statistical computing*.
- 503 Root, T. L., Price, J. T., Hall, K. R., Schneider, S. H., Rosenzweig, C., and Pounds, J. A. (2003).
504 Fingerprints of global warming on wild animals and plants. *Nature* 421, 57.
505 doi:10.1038/nature01333
506 https://www.nature.com/articles/nature01333#supplementary-information.
- 507 Salazar-Parra, C., Aguirreolea, J., Sánchez-Díaz, M., Irigoyen, J., and Morales, F. (2010). Effects of
508 climate change scenarios on Tempranillo grapevine (*Vitis vinifera* L.) ripening: Response
509 to a combination of elevated CO₂ and temperature, and moderate drought. *Plant Soil -*
510 *PLANT SOIL* 337, 179–191. doi:10.1007/s11104-010-0514-z.

- 511 Schultz, H. R., and Jones, G. V. (2010). Climate Induced Historic and Future Changes in
512 Viticulture. *J. Wine Res.* 21, 137–145. doi:10.1080/09571264.2010.530098.
- 513 Sepúlveda, G., Kliewer, W. M., and Ryugo, K. (1986). Effect of High Temperature on Grapevines
514 (*Vitis vinifera* L.). I. Translocation of ¹⁴C-Photosynthates. *Am. J. Enol. Vitic.* 37, 13–19.
- 515 Soar, C., Collins, M., and Sadras, V. (2009). Irrigated Shiraz vines (*Vitis vinifera*) upregulate gas
516 exchange and maintain berry growth in response to short spells of high maximum
517 temperature in the field. *Funct. Plant Biol. - FUNCT PLANT BIOL* 36.
518 doi:10.1071/FP09101.
- 519 Van Leeuwen, C., Destrac-Irvine, A., Dubernet, M., Duchêne, E., Gowdy, M., Marguerit, E., et al.
520 (2019). An Update on the Impact of Climate Change in Viticulture and Potential
521 Adaptations. *Agronomy* 9, 514. doi:10.3390/agronomy9090514.
- 522 Wolkovich, E. M., Burge, D. O., Walker, M. A., and Nicholas, K. A. (2017). Phenological diversity
523 provides opportunities for climate change adaptation in winegrapes.
- 524 Zaka, S., Ahmed, L. Q., Escobar-Gutiérrez, A. J., Gastal, F., Julier, B., and Louarn, G. (2017). How
525 variable are non-linear developmental responses to temperature in two perennial
526 forage species? *Agric. For. Meteorol.* 232, 433–442.
527 doi:10.1016/j.agrformet.2016.10.004.
- 528 Zaka, S., Frak, E., Julier, B., Gastal, F., and Louarn, G. (2016). Intraspecific variation in thermal
529 acclimation of photosynthesis across a range of temperatures in a perennial crop. *AoB*
530 *PLANTS* 8. doi:10.1093/aobpla/plw035.
- 531

Day of budburst in greenhouse (day since forcing)

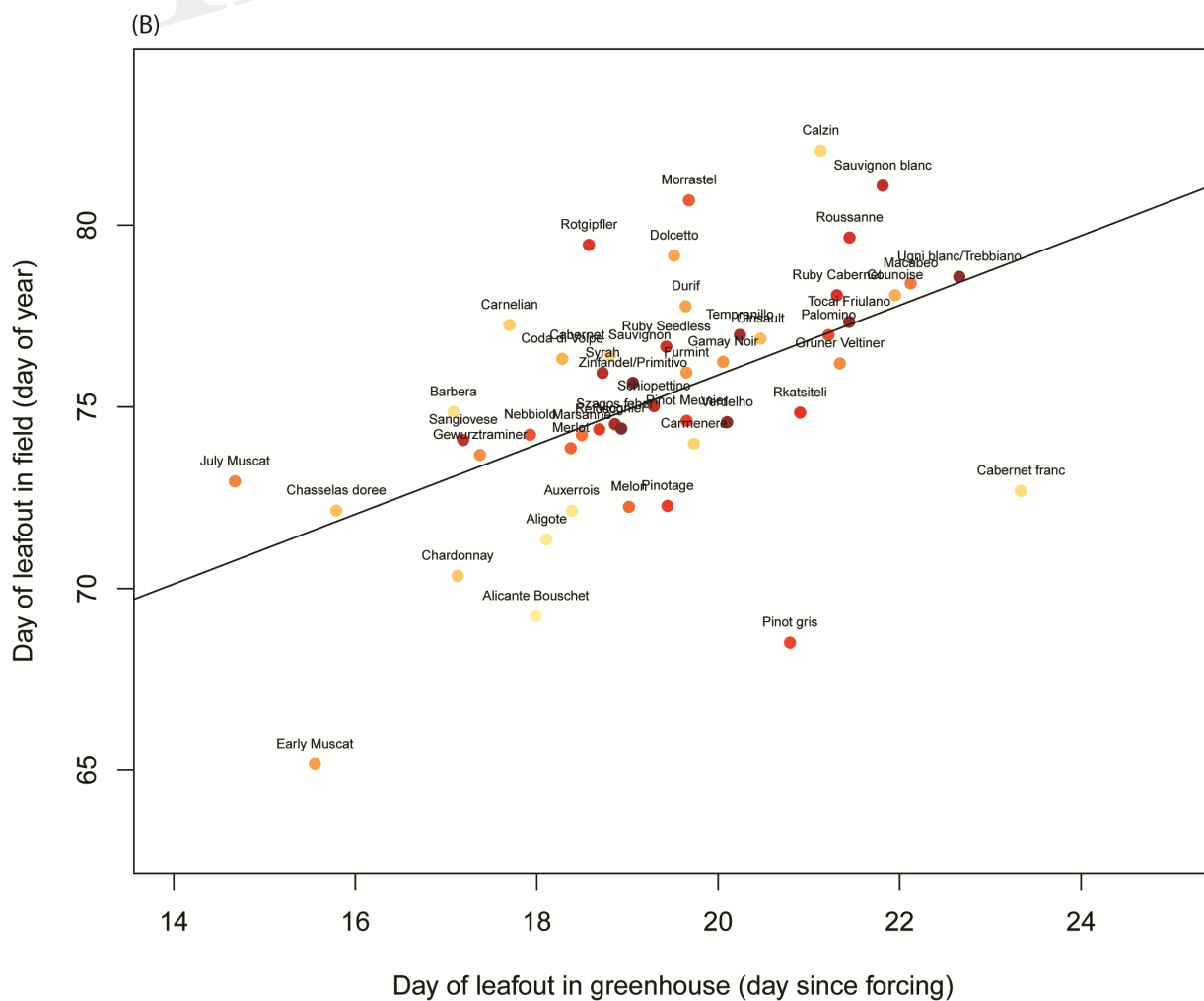
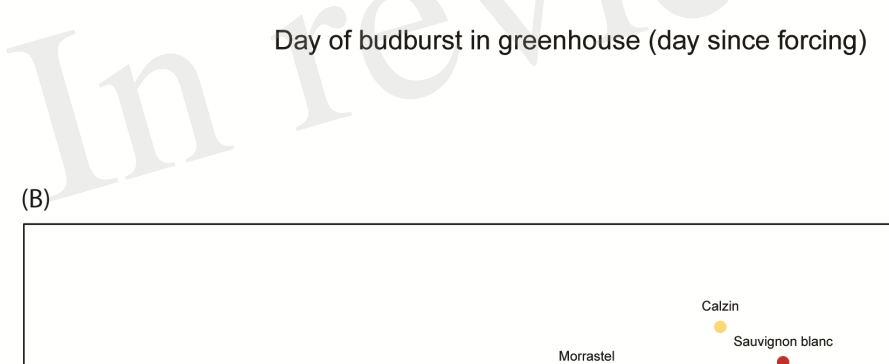


Figure 2.TIF

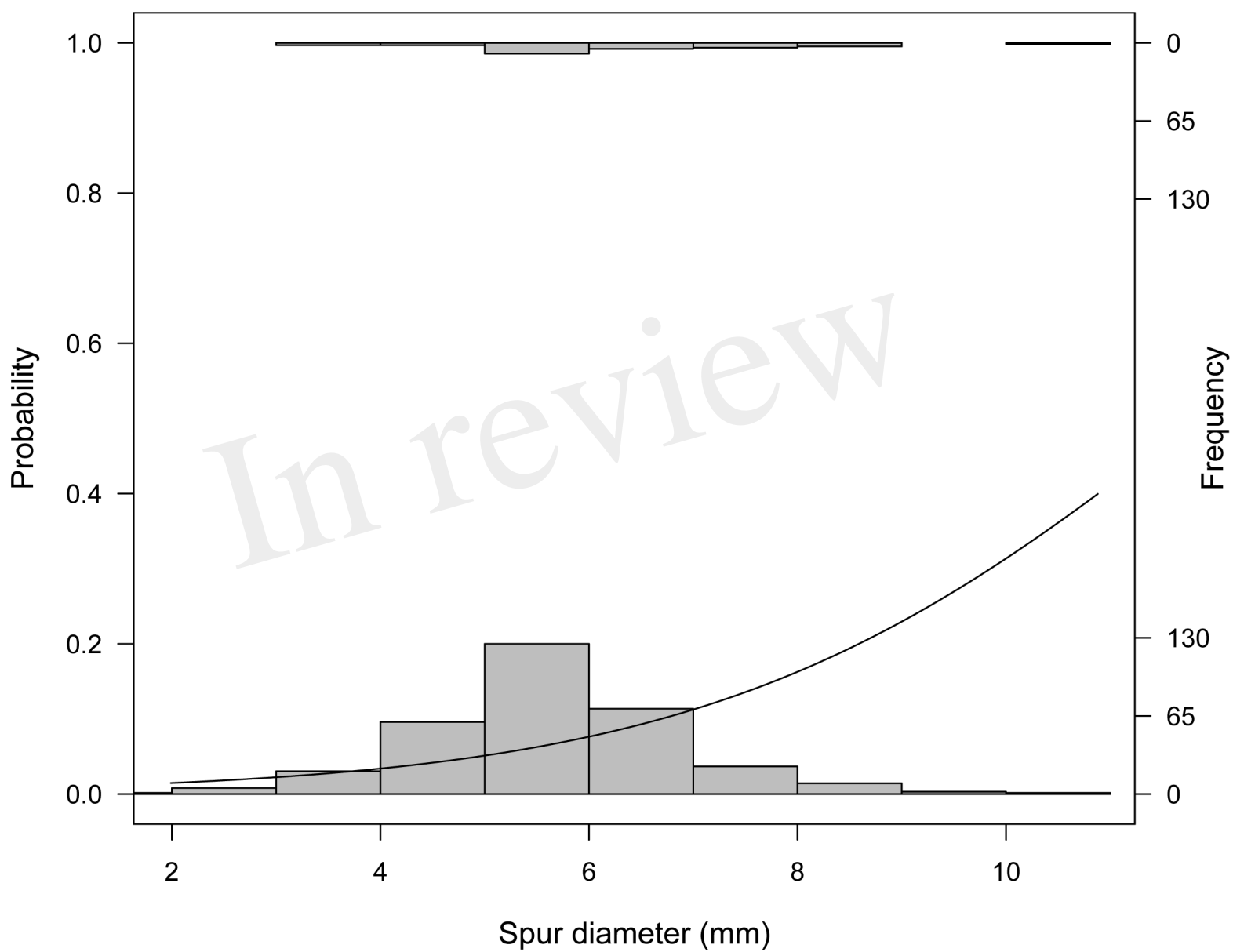


Figure 3.TIF

