Supplementary Material: Budburst timing within a functional trait framework

4 Figures & Tables

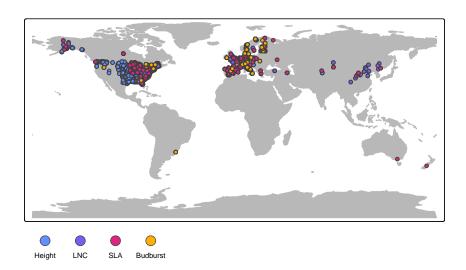


Figure S1: We obtained data from the TRY and BIEN plant trait databases and the OSPREE database of plant phenology experiments for temperate tree species. Following our cleaning of the data and selection of uncorrelated traits, our final dataset included 37 species from 24 unique datasources for the trait data and 34 unique studies for the budburst data. Our data is focused on temperate ecosystems globally with most data originating from North America and Europe.

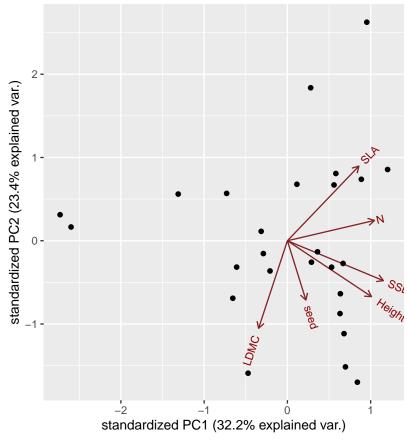


Figure S2: A projection of tree traits across the first and second principle component axis. Arrows represent the direction of vectors for the six functional traits with complete trait data. Points represent the 26 species for which complete trait data was available.

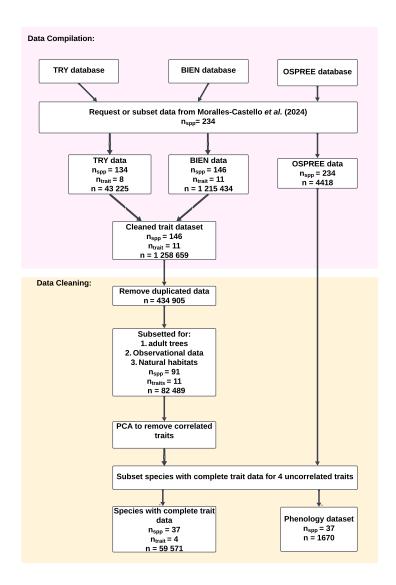


Figure S3: Our initial aim was to include all species from (analyzed by Morales-Castilla *et al.*, 2024), however trait data was only available for a subset of these species. Data was cleaned extensively, with each step depicted by a box, and subset to only include traits for adult trees growing under natural conditions for which we had a complete suite of trait values.

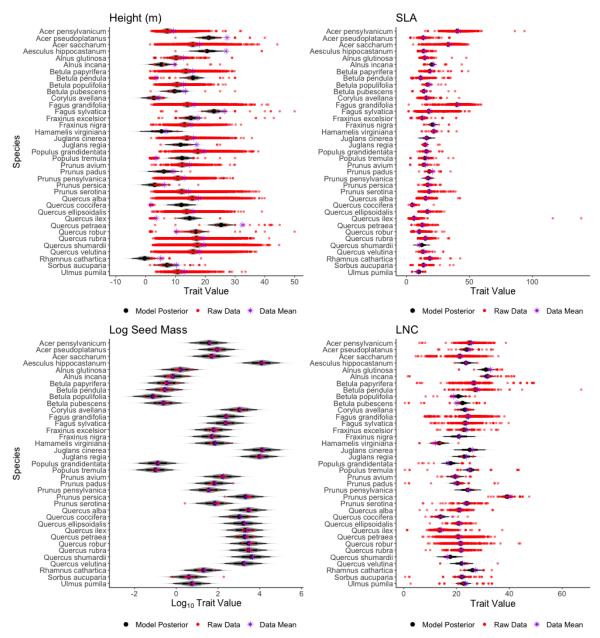


Figure S4: Comparisons of estimated model fits and raw data from joint models of trait effects on budburst phenological cues for 37 species of woody deciduous plants. Four functional traits—a. height, b. SLA, c. seed mass, and d. LNC—were modeled individually, with the calculated trait value being used to jointly model species responses to standardized chilling, forcing, and photoperiod cues. Model posteriors are shown in black, with the thicker line depicting the 66% interval and the thinner black line the 97% interval. Overall species level model posterior distributions were well aligned with the raw data, shown in red, and the species level means from the raw data, denoted as a purple stars.

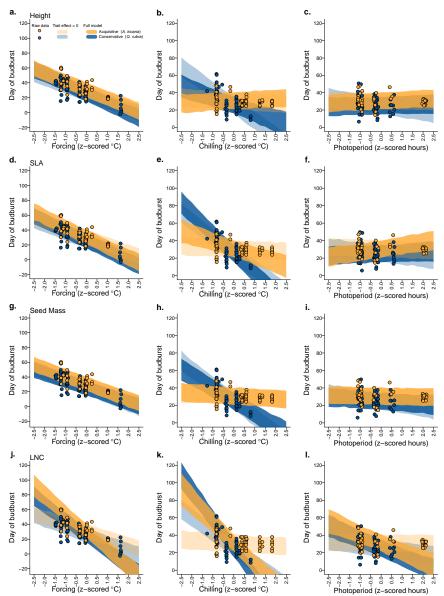


Figure S5: Here we show an example with an acquisitive species, Alnus incana shown in blue, and a conservative species, Quercus rubra shown in yellow, using the posterior estimates from our height (a-c), SLA (d-f), seed mass (g-i), and LNC (j-l) models to estimate the day of budburst. a,d, The effect of height on budburst timing was smallest in response to forcing, b while the greatest effect of height on budburst was in response to chilling, c, followed by photoperiod. e, The greatest effect of SLA on budburst was only in relation to photoperiod. g,h,i, Seed mass had a negligible effect on our estimates budburst. j,k,l, But LNC had a considerable effect on budburst timing in relation to each cue. The strength of the relationships can be assessed by comparing the colored bands, with the lighter bands depicting the estimated 50% uncertainty interval without the effect of traits ($\mu_{k,g}$ equal to zero) and darker bands the estimated budburst with the effects of traits included. Points represent the raw budburst data.

Table S1: Bibliographic information for trait data sources from both BIEN and Try trait databases. Datasets without references or incomplete references are denoted below as 'unreferenced'.

Database	Reference	Trait name	Unit	No. observations	No. Species
bien	Mchugh et al. (2015)	Height	m	26	3
bien	Marx et al. (2016)	Height	m	2	2
oien	Price et al. (2014)	Height	m	27	19
bien	Unreferenced	Height	m	18	16
oien	Kleyer et al. (2008)	Height	m	90	19
oien	Unreferenced	Height	m	10	10
bien	Moles, Angela; unreferenced	Height	m	21	1
oien	Reams, Greg; unreferenced	Height	m	47036	19
oien	Grime, Hodgson, & Hunt; unreferenced	Height	m	5	į.
oien	Unreferenced	Height	m	8	
oien	Pérez-de Lis et al. (2017)	Height	m	18	-
oien	Robinson et al. (2015)	Height	m	120	
oien	Anderson-teixeira et al. (2015)	Height	m	20	
ry	Bond-Lamberty et al. (2002)	Height	m	2	
ry	Unpublished	Height	\mathbf{m}	275	;
ry	Wright et al. (2004)	Height	\mathbf{m}	28	19
ry	Prentice et al. (2011)	Height	\mathbf{m}	2	
ry	Schweingruber and Landolt (2010)	Height	\mathbf{m}	21	2
ry	Unpublished	Height	\mathbf{m}	35	
ry	Moles et al. (2004)	Height	\mathbf{m}	5	
ry	Cavender-Bares et al. (2006)	Height	\mathbf{m}	1	
ry	Diaz et al. (2004)	Height	\mathbf{m}	11	1
ry	Craine et al. (2009)	LNC	$\mathrm{mg/g}$	287	1
ry	Wilson et al. (2000)	LNC	$\mathrm{mg/g}$	44	
ry	Wenxuan et al. (2012)	LNC	$\mathrm{mg/g}$	7	
ry	Yahan et al. (2013)	LNC	$\mathrm{mg/g}$	7	
ry	Wright et al. (2004)	LNC	$\mathrm{mg/g}$	65	3
ry	Prentice et al. (2011)	LNC	$\mathrm{mg/g}$	3	
ry	Vergutz et al. (2012)	LNC	$\mathrm{mg/g}$	120	2
ry	Atkin et al. (2015)	LNC	$\mathrm{mg/g}$	24	
ry	Marie et al. (2015)	LNC	$\mathrm{mg/g}$	72	2
ry	Cornelissen et al. (2003)	LNC	$\mathrm{mg/g}$	2	
ry	Unpublished	LNC	mg/g	3216	3
ry	Wang et al. (2017)	LNC	mg/g	6	
oien	Marx et al. (2016)	Seed mass	$_{ m mg}$	3	
oien	Unreferenced	Seed mass	$\overline{\mathrm{mg}}$	4	
oien	Liu et al. (2018)	Seed mass	$_{ m mg}$	250	3
oien	Ameztegui et al. (2017)	Seed mass	mg	12	1
oien	Paine et al. (2015)	Seed mass	$_{ m mg}$	12	
ry	Wilson et al. (2000)	SLA	mm2 mg-1	44	
ry	Unpublished	SLA	mm2 mg-1	204	
ry	Wright et al. (2004)	SLA	mm2 mg-1	93	3
ry	Prentice et al. (2011)	SLA	mm2 mg-1	2	
ry	Kleyer et al. (2008)	SLA	mm2 mg-1	102	1
ry	Unpublished	SLA	mm2 mg-1	83	
ry	Atkin et al. (2015)	SLA	mm2 mg-1	40	1
ry	Marie et al. (2015)	SLA	mm2 mg-1	86	2
ry	Cornelissen et al. (2003)	SLA	mm2 mg-1	615	1
ry	Unpublished	SLA	mm2 mg-1	6307	3
ry	Wang et al. (2017) 6	SLA	mm2 mg-1	6	· ·
ry	Shipley and Vu (2002)	SLA	mm2 mg 1	20	
ry	Cavender-Bares et al. (2006)	SLA	mm2 mg-1	42	
				42	
ry	Unpublished	SLA	mm2 mg-1	1	

Table S2: Summary of estimates from our model of height (n=42781) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with paremeters with intervals that cross zero depicting weak relationships

	mean	X5.	X25.	X75.	X95.
$\mu_{grand.trait}$	12.7	9.4	11.4	14.0	15.8
$\mu_{k,g}$	32.1	27.8	30.3	33.9	36.5
μ_{force}	-10.7	-15.5	-12.6	-8.8	-6.2
μ_{chill}	-4.1	-10.8	-6.8	-1.4	2.6
μ_{photo}	1.1	-2.6	-0.3	2.5	4.6
$\beta_{trait.force}$	0.2	-0.2	0.0	0.3	0.5
$\beta_{trait.chill}$	-0.5	-1.0	-0.7	-0.3	-0.1
$\beta_{trait.photo}$	-0.2	-0.5	-0.4	-0.1	0.0
$\sigma_{species}$	5.9	4.8	5.4	6.4	7.3
σ_{study}	7.5	5.8	6.7	8.3	9.7
σ_{trait}	5.4	5.4	5.4	5.4	5.4
σ_{pheno}	15.1	11.8	13.8	16.4	18.6
σ_{force}	5.0	3.3	4.1	5.7	7.1
σ_{chill}	8.5	5.6	7.0	9.8	12.4
σ_{photo}	3.2	2.0	2.6	3.8	4.8
σ_d	14.2	13.8	14.0	14.3	14.6

Table S3: Summary of estimates from our model of specific leaf area (SLA, n=7656) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with paremeters with intervals that cross zero depicting weak relationships

	mean	X5.	X25.	X75.	X95.
$\mu_{grand.trait}$	16.8	14.5	15.8	17.8	19.3
$\mu_{k,g}$	31.3	27.2	29.6	33.0	35.6
μ_{force}	-11.4	-16.2	-13.1	-9.6	-7.2
μ_{chill}	-16.7	-24.6	-19.8	-13.3	-9.1
μ_{photo}	1.9	-2.4	0.2	3.6	5.8
$\beta_{trait.force}$	0.2	-0.1	0.1	0.3	0.4
$\beta_{trait.chill}$	0.3	-0.1	0.2	0.5	0.7
$\beta_{trait.photo}$	-0.2	-0.5	-0.3	-0.1	0.0
$\sigma_{species}$	7.8	6.4	7.1	8.4	9.4
σ_{study}	3.3	2.0	2.6	3.8	5.1
σ_{trait}	6.2	6.1	6.1	6.2	6.3
σ_{pheno}	13.9	10.7	12.4	15.3	17.6
σ_{force}	5.0	3.3	4.2	5.6	7.0
σ_{chill}	10.6	7.2	8.9	12.0	14.8
σ_{photo}	3.5	2.3	2.9	3.9	5.0
σ_d	14.2	13.8	14.0	14.3	14.6

Table S4: Summary of estimates from our model of seed mass (n=281) using data from 37 focal species. Values represent the mean estimat for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with paremeters with intervals that cross zero depicting weak relationships

	mean	X5.	X25.	X75.	X95.
$\mu_{grand.trait}$	1.9	1.0	1.5	2.2	2.7
$\mu_{k,g}$	31.4	27.1	29.6	33.1	35.8
μ_{force}	-8.2	-10.8	-9.2	-7.1	-5.6
μ_{chill}	-9.4	-14.0	-11.2	-7.5	-4.8
μ_{photo}	-1.3	-3.4	-2.1	-0.4	0.8
$\beta_{trait.force}$	-0.3	-1.4	-0.8	0.1	0.9
$\beta_{trait.chill}$	-1.1	-2.9	-1.8	-0.4	0.7
$\beta_{trait.photo}$	-0.6	-1.5	-0.9	-0.2	0.4
$\sigma_{species}$	1.6	1.3	1.5	1.7	2.0
σ_{study}	1.0	0.8	0.9	1.0	1.1
σ_{trait}	0.2	0.2	0.2	0.3	0.3
σ_{pheno}	14.8	11.2	13.3	16.3	18.6
σ_{force}	4.9	3.4	4.2	5.5	6.7
σ_{chill}	10.7	7.1	8.8	12.1	15.4
σ_{photo}	3.6	2.3	3.0	4.1	5.1
σ_d	14.1	13.7	14.0	14.3	14.5

Table S5: Summary of estimates from our model of leaf nitrogen content (LNC, n=3853) using data from 37 focal species. Values represent the mean estimat for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with paremeters with intervals that cross zero depicting weak relationships

	mean	X5.	X25.	X75.	X95.
$\mu_{grand.trait}$	22.6	20.4	21.7	23.5	24.9
$\mu_{k,g}$	31.1	27.1	29.4	32.8	35.3
μ_{force}	-19.3	-27.9	-22.9	-15.8	-10.5
μ_{chill}	-27.1	-38.5	-31.7	-22.4	-15.4
μ_{photo}	-9.4	-17.0	-12.5	-6.2	-1.9
$\beta_{trait.force}$	0.5	0.1	0.3	0.6	0.8
$\beta_{trait.chill}$	0.7	0.2	0.5	0.9	1.2
$\beta_{trait.photo}$	0.3	0.0	0.2	0.4	0.6
$\sigma_{species}$	5.1	4.2	4.7	5.5	6.2
σ_{study}	3.6	2.2	2.9	4.1	5.3
σ_{trait}	5.1	5.0	5.1	5.2	5.2
σ_{pheno}	14.0	10.9	12.7	15.3	17.4
σ_{force}	4.6	3.0	3.8	5.2	6.6
σ_{chill}	8.9	6.1	7.5	10.1	12.5
σ_{photo}	3.6	2.4	3.0	4.1	5.0
σ_d	14.2	13.8	14.0	14.3	14.6

Stan model code

```
6
  data {
     int<lower = 1> n_spec;
     int<lower = 1> N;
     int<lower = 1, upper = n_spec> trait_species[N];
10
     int<lower = 1> n_study;
11
     int<lower = 1, upper = n_study> study[N];
12
     vector[N] yTraiti;
13
14
     int<lower = 1> Nph;
15
     int<lower = 1, upper = n_spec> phenology_species[Nph];
16
     vector[Nph] yPhenoi;
17
     vector[Nph] forcei;
18
     vector[Nph] chilli;
19
     vector[Nph] photoi;
20
21
22
   parameters{
23
     real mu_grand;
25
     vector[n_spec] muSp;
     vector[n_study] muStudy;
27
     real<lower = 0> sigma_traity;
28
     real<lower = 0> sigma_sp;
29
     real<lower = 0> sigma_study;
30
31
     real alphaForceSp[n_spec];
32
     real muForceSp;
33
     real<lower = 0> sigmaForceSp;
     real alphaChillSp[n_spec];
35
     real muChillSp;
     real<lower = 0> sigmaChillSp;
37
     real alphaPhotoSp[n_spec];
38
     real muPhotoSp;
     real<lower = 0> sigmaPhotoSp;
40
     real alphaPhenoSp[n_spec];
41
     real muPhenoSp;
42
     real<lower = 0> sigmaPhenoSp;
     real betaTraitxForce;
44
     real betaTraitxChill;
     real betaTraitxPhoto;
46
     real<lower = 0> sigmapheno_y;
47
48
49
   transformed parameters{
50
51
     vector[N] y_hat;
52
     vector[n_spec] mu_grand_sp;
53
     real betaForceSp[n_spec];
```

```
real betaPhotoSp[n_spec];
56
     real betaChillSp[n_spec];
57
58
     for(i in 1:n_spec){
59
       mu_grand_sp[i] = mu_grand + muSp[i];
60
61
     for (i in 1:N){
62
       y_hat[i] = mu_grand + muSp[trait_species[i]] + muStudy[study[i]];
63
64
65
     for (isp in 1:n_spec){
66
       betaForceSp[isp] = alphaForceSp[isp] + betaTraitxForce * (mu_grand_sp[isp]);
68
     for (isp in 1:n_spec){
       betaPhotoSp[isp] = alphaPhotoSp[isp] + betaTraitxPhoto * (mu_grand_sp[isp]);
70
71
     for (isp in 1:n_spec){
72
       betaChillSp[isp] = alphaChillSp[isp] + betaTraitxChill * (mu_grand_sp[isp]);
73
     }
74
   }
75
76
   model{
77
     yTraiti ~ normal(y_hat, sigma_traity);
     muSp ~ normal(0, sigma_sp);
79
     muStudy ~ normal(0, sigma_study);
     mu_grand ~ normal(20,10);
81
     sigma_sp ~ normal(4,5);
82
     sigma_study ~ normal(2,5);
83
     sigma_traity ~ normal(3,5);
85
     for (i in 1:Nph){
       yPhenoi[i] ~ normal(alphaPhenoSp[phenology_species[i]] +
87
       betaForceSp[phenology_species[i]] * forcei[i] +
       betaPhotoSp[phenology_species[i]] * photoi[i] +
89
       betaChillSp[phenology_species[i]] * chilli[i],
       sigmapheno_y);
91
     }
92
93
     alphaPhenoSp ~ normal(muPhenoSp, sigmaPhenoSp);
     alphaForceSp ~ normal(muForceSp, sigmaForceSp);
     alphaChillSp ~ normal(muChillSp, sigmaChillSp);
     alphaPhotoSp ~ normal(muPhotoSp, sigmaPhotoSp);
     muPhenoSp ~ normal(40,10);
99
     sigmaPhenoSp ~ normal(5,5);
100
     sigmapheno_y ~ normal(10,5);
102
103
     muForceSp ~ normal(-15,10);
104
     sigmaForceSp ~ normal(5,5);
105
106
     muChillSp ~ normal(-15,10);
107
```

```
sigmaChillSp ~ normal(5,5);
108
109
     muPhotoSp ~ normal(-15,10);
110
     sigmaPhotoSp ~ normal(5,5);
111
112
     betaTraitxForce ~ normal(0,1);
113
     betaTraitxPhoto ~ normal(0,1);
114
     betaTraitxChill ~ normal(0,1);
116
117 }
118
```

¹⁹ References

- Ameztegui, A., A. Paquette, B. Shipley, M. Heym, C. Messier, and D. Gravel. 2017. Shade tolerance and the functional trait: demography relationship in temperate and boreal forests. Functional Ecology 31:821–830.
- Anderson-teixeira, K. J., J. C. Mcgarvey, H. C. Muller-landau, J. Y. Park, E. B. Gonzalez-akre, V. Herrmann, A. C. Bennett, C. V. So, N. A. Bourg, J. R. Thompson, S. M. Mcmahon, and W. J. Mcshea. 2015. Size-related scaling of tree form and function in a mixed-age forest. Functional Ecology 29:1587–1602.
- Atkin, O., K. Bloomfield, P. Reich, M. Tjoelker, G. Asner, D. Bonal, G. Bönisch, M. Bradford, L. Cer-127 nusak, E. Cosio, D. Creek, C. K.Y., T. Domingues, J. Dukes, J. Egerton, J. Evans, G. Farquhar, 128 N. Fyllas, P. Gauthier, E. Gloor, T. Gimeno, K. Griffin, R. Guerrieri, M. Heskel, C. Huntingford, 129 F. Ishida, J. Kattge, H. Lambers, M. Liddell, J. Lloyd, C. Lusk, R. Martin, A. Maksimov, T. Maxi-130 mov, Y. Malhi, B. Medlyn, P. Meir, L. Mercado, N. Mirotchnick, D. Ng, Ü. Niinemets, O. O'Sullivan, 131 O. Phillips, L. Poorter, P. Poot, I. Prentice, N. Salinas, L. Rowland, M. Ryan, S. Sitch, M. Slot, 132 N. Smith, M. Turnbull, M. VanderWel, F. Valladares, E. Veneklaas, L. Weerasinghe, C. Wirth, 133 I. Wright, K. Wythers, J. Xiang, S. Xiang, and J. Zaragoza-Castells. 2015. Global variability in leaf 134 respiration in relation to climate, plant functional types and leaf traits. New Phytologist 206:614– 135 636. 136
- Bond-Lamberty, B., C. Wang, and S. T. Gower. 2002. Aboveground and belowground biomass and sapwood area allometric equations for six boreal tree species of northern Manitoba. Canadian Journal of Forest Research 32:1441–1450.
- Cavender-Bares, J., A. Keen, and B. Miles. 2006. Phylogenetic structure of floridian plant communities
 depends on taxonomic and spatial scale. Ecology 87:109–122.
- Cornelissen, J. H. C., B. Cerabolini, P. Castro-Diez, P. Villar-Salvador, G. Montserrat-Marti, J. P. Puyravaud, M. Maestro, M. J. A. Werger, and R. Aerts. 2003. Functional traits of woody plants: correspondence of species rankings between field adults and laboratory-grown seedlings? Journal of Vegetation Science 14:311–322.
- Craine, J. M., A. J. Elmore, M. P. M. Aidar, M. Bustamante, T. E. Dawson, E. A. Hobbie, A. Kahmen,
 M. C. Mack, K. K. Mclauchlan, A. Michelsen, G. B. Nardoto, L. H. Pardo, J. Penuelas, P. B. Reich,
 E. A. G. Schuur, W. D. Stock, P. H. Templer, R. A. Virginia, J. M. Welker, and I. J. Wright. 2009.
 Global patterns of foliar nitrogen isotopes and their relationships with climate, mycorrhizal fungi,
 foliar nutrient concentrations, and nitrogen availability. New Phytologist 183:980–992.
- Diaz, S., J. G. Hodgson, K. Thompson, M. Cabido, J. H. C. Cornellissen, A. Jalili, G. Montserrat Marti, J. P. Grime, F. Zarrinkamar, Y. Asri, S. R. Band, S. Basconcelo, P. Castro-Diez, G. Funes, B. Hamzehee, M. Khoshnevi, N. Pérez-Harguindeguy, M. C. Pérez-Rontomé, F. A. Shirvany,
 F. Vendramini, S. Yazdani, R. Abbas-Azimi, A. Bogaard, S. Boustani, M. Charles, M. Dehghan,
 L. de Torres-Espuny, V. Falczuk, J. Guerrero-Campo, A. Hynd, G. Jones, E. Kowsary, F. Kazemi Saeed, M. Maestro-Martinez, A. Romo-Diez, S. Shaw, B. Siavash, P. Villar-Salvador, and M. R.
 Zak. 2004. The plant traits that drive ecosystems: Evidence from three continents. Journal of
 Vegetation Science 15:295–304.
- Kleyer, M., R. M. Bekker, I. C. Knevel, J. P. Bakker, K. Thompson, M. Sonnenschein, P. Poschlod,
 J. M. V. Groenendael, L. Klime, J. Klimesova, S. Klotz, G. M. Rusch, M. Hermy, D. Adriaens,
 G. Boedeltje, B. Bossuyt, A. Dannemann, P. Endels, L. Götzenberger, J. G. Hodgson, A.-k. Jackel,
 I. Kühn, D. Kunzmann, W. A. Ozinga, C. Römermann, M. Stadler, J. Schlegelmilch, H. J. Steendam,
 O. Tackenberg, B. Wilmann, J. H. C. Cornelissen, O. Eriksson, E. Garnier, and B. Peco. 2008. The
 LEDA Traitbase: a database of life-history traits of the Northwest European flora. Journal of
 Ecology 96:1266–1274.

- Liu, K., S. Eastwood, R.J. a d Flynn, R. Turner, and W. Stuppy. 2018. Kew database.
- Marie, V., I. J. Wright, I. C. Prentice, N. H. Batjes, R. Bhaskar, P. M. van Bodegom, W. K. Cornwell,
 D. Ellsworth, Ü. Niinemets, A. Ordonez, P. B. Reich, and L. S. Santiago. 2015. Global effects of soil
 and climate on leaf photosyncthetic traits and rates. Global Ecology and Biogeography 24:706-717.
- Marx, H. E., D. E. Giblin, P. W. Dunwiddie, and D. C. Tank. 2016. Deconstructing Darwin's Naturalization using community phylogenetics and functional traits. Diversity and Distributions 22:318–331.
- Mchugh, N., J. L. Edmondson, K. J. Gaston, J. R. Leake, and O. S. O. Sullivan. 2015. Modelling short-rotation coppice and tree planting for urban carbon management a citywide analysis. Journal of Applied Ecology 52:1237–1245.
- Moles, A. T., D. S. Falster, M. R. Leishman, and M. Westoby. 2004. Small-seeded species produce
 more seeds per square metre of canopy per year, but not per individual per lifetime. Journal of
 Ecology 92:384–396.
- Paine, C. E. T., L. Amissah, H. Auge, C. Baraloto, M. Baruffol, N. Bourland, H. Bruelheide, K. Dainou,
 R. C. de Gouvenain, J.-l. Doucet, S. Doust, P. V. A. Fine, C. Fortunel, J. Haase, K. D. Holl, H. Jactel, X. Li, K. Kitajima, J. Koricheva, C. Martínez-Garza, C. Messier, A. Paquette, C. Philipson,
 D. Piotto, L. Poorter, J. M. Posada, C. Potvin, K. Rainio, S. E. Russo, M. Ruiz-jaen, M. Scherer-lorenzen, C. O. Webb, S. J. Wright, R. A. Zahawi, and A. Hector. 2015. Globally, functional traits are weak predictors of juvenile tree growth, and we do not know why. Journal of Ecology 103:978–989.
- Pérez-de Lis, G., J. M. Olano, V. Rozas, S. Rossi, R. A. Vázquez-Ruiz, and I. García-Gonzalez. 2017.
 Environmental conditions and vascular cambium regulate carbon allocation to xylem growth in deciduous oaks. Functional Ecology 31:592–603.
- Prentice, I. C., T. Meng, H. Wang, S. P. Harrison, J. Ni, and G. Wang. 2011. Evidence of a universal scaling relationship for leaf CO 2 drawdown along an aridity gradient. New Phytologist 190:169–180.
- Price, C. A., I. J. Wright, D. D. Ackerly, Ü. Niinemets, P. B. Reich, and E. J. Veneklaas. 2014. Are
 leaf functional traits 'invariant' with plant size and what is 'invariance' anyway? Functional Ecology
 28:1330–1343.
- Robinson, K. M., C. Hauzy, N. Loeuille, and B. R. Albrectsen. 2015. Relative impacts of environmental variation and evolutionary history on the nestedness and modularity of tree–herbivore networks. Ecology and Evolution 5:2898–2915.
- Schweingruber, F., and W. Landolt. 2010. The xylem database.
- Shipley, B., and T.-T. Vu. 2002. Dry matter content as a measure of dry matter concentration in plants and their parts. New Phytologist 153:359–364.
- Vergutz, L., S. Manzoni, A. Porporato, R. Novais, and R. Jackson. 2012. A Global Database of
 Carbon and Nutrient Concentrations of Green and Senesced Leaves. Oak Ridge National Laboratory
 Distributed Active Archive Center Oak Ridge, Tennessee, U.S.A.
- Wang, H., S. P. Harrison, I. C. Prentice, Y. Yang, F. Bai, H. Furstenau Togashi, M. Wang, S. Zhou,
 and J. Ni. 2017. The China Plant Trait Database. PANGAEA.
- Wenxuan, H., C. Yahan, Z. Fang-Jie, L. Tang, J. Rongfeng, and Z. Fusuo. 2012. Floral, climatic and
 soil pH controls on leaf ash content in China's terrestrial plants. Global Ecology and Biogeography
 21:376–382.

- Wilson, K. B., D. D. Baldocchi, and P. J. Hanson. 2000. Spatial and seasonal variability of photosynthetic parameters and their relationship to leaf nitrogen in a deciduous forest. Tree Physiology 20:565–578.
- Wright, I. J., M. Westoby, P. B. Reich, J. Oleksyn, D. D. Ackerly, Z. Baruch, F. Bongers, J. Cavender-Bares, T. Chapin, J. H. C. Cornellissen, M. Diemer, J. Flexas, J. Gulias, E. Garnier, M. L. Navas,
 C. Roumet, P. K. Groom, B. B. Lamont, K. Hikosaka, T. Lee, W. Lee, C. Lusk, J. J. Midgley,
 Ü. Niinemets, H. Osada, H. Poorter, P. Pool, E. J. Veneklaas, L. Prior, V. I. Pyankov, S. C.
 Thomas, M. G. Tjoelker, and R. Villar. 2004. The worldwide leaf economics spectrum. Nature
 428:821–827.
- Yahan, C., H. Wenxuan, T. Luying, T. Zhiyao, and F. Jingyun. 2013. Leaf nitrogen and phosphorus concentrations of woody plants differ in responses to climate, soil and plant growth form. Ecography 36:178–184.