Supplementary Material: Budburst timing within a functional trait framework

4 Figures & Tables

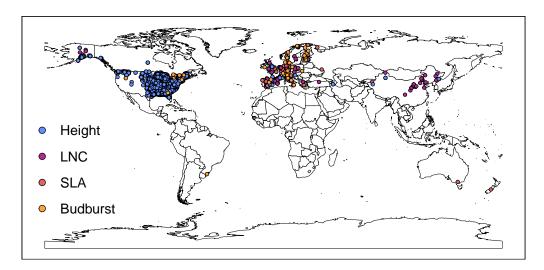


Figure S1: We obtained data from the TRY and BIEN plant trait databases and the OSPREE database of plant phenology experiments for temperate tree species. Following our cleaning of the data and selection of uncorrelated traits, our final dataset included 37 species from 24 unique datasources for the trait data and 34 unique studies for the budburst data. Our data is focused on temperate ecosystems globally with most data originating from North America and Europe.

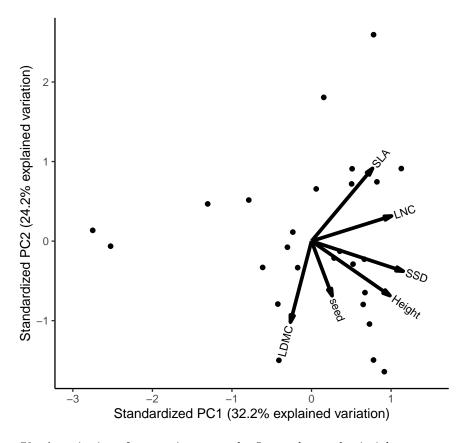


Figure S2: A projection of tree traits across the first and second principle component axis. Arrows represent the direction of vectors for the six functional traits with complete trait data. Points represent the 26 species for which complete trait data was available.

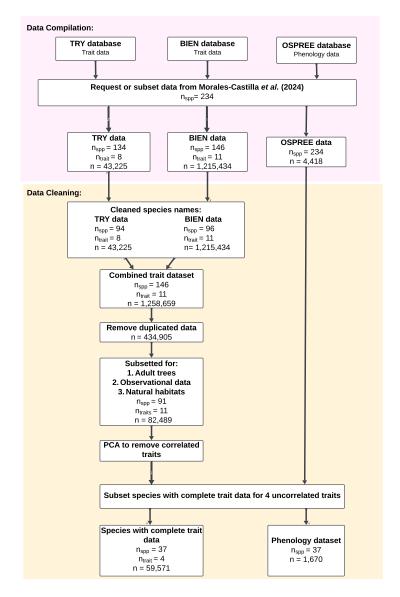


Figure S3: Our initial aim was to include all species from (analyzed by Morales-Castilla *et al.*, 2024), however trait data was only available for a subset of these species. Data was cleaned extensively, with each step depicted by a box, and subset to only include traits for adult trees growing under natural conditions for which we had a complete suite of trait values.

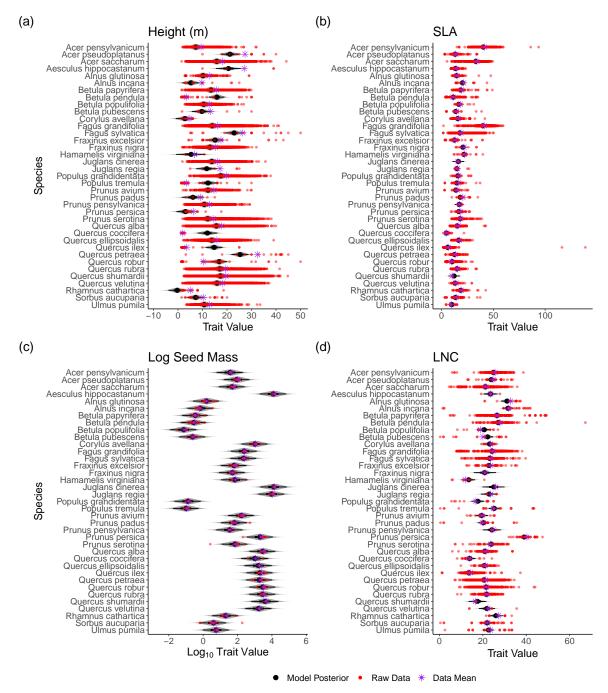


Figure S4: Comparisons of estimated model fits and raw data from joint models of trait effects on budburst phenological cues for 37 species of woody deciduous plants. Four functional traits—(a) height, (b) SLA, (c) seed mass, and (d) LNC— were modeled individually, with the calculated trait value being used to jointly model species' responses to standardized chilling, forcing, and photoperiod cues. Model posteriors are shown in black, with the thicker line depicting the 50% uncertainty interval and the thinner black line the 95% uncertainty interval. Overall species-level model posterior distributions were well-aligned with the raw data, shown in red, and the species-level means from the raw data, denoted as a purple stars.

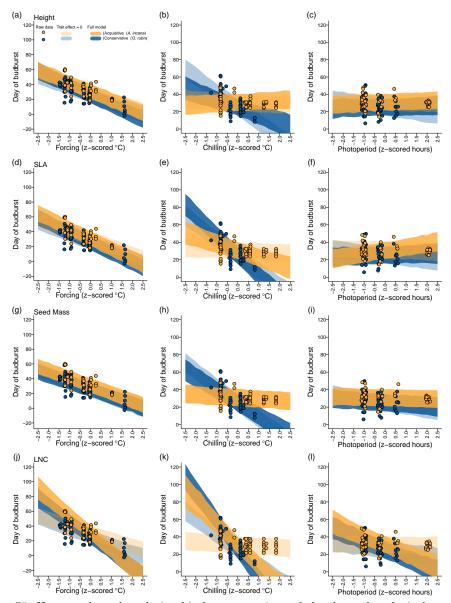


Figure S5: Here we show the relationship between traits and the three phenological cues using as an example an acquisitive species, $Alnus\ incana$ shown in blue, and a conservative species, $Quercus\ rubra$ shown in yellow. The posterior estimates from our height (a-c), SLA (d-f), seed mass (g-i), and LNC (j-l) models were used to estimate the day of budburst for the two species. The effect of height on budburst timing was smallest in response to forcing (a), while the greatest effect of height on budburst was in response to chilling (b), followed by photoperiod (c). The greatest effect of SLA on budburst was only in relation to photoperiod (f). Seed mass had a negligible effect on our estimates budburst (g-i). But LNC had a considerable effect on budburst timing in relation to each cue (j-l). The strength of the relationships can be assessed by comparing the colored bands, with the lighter bands depicting the estimated 50% uncertainty interval without the effect of traits ($\mu_{k,g}$ equal to zero) and darker bands the estimated budburst with the effects of traits included. Points represent the raw budburst data.

Table S1: Bibliographic information for trait data sources from both BIEN and TRY trait databases. Datasets without references or incomplete references are denoted below as 'unreferenced'.

Database	Reference	Trait name	Unit	No. observations	No. Species
BIEN	Mchugh <i>et al.</i> (2015)	Height	m	26	8
BIEN	Marx et al. (2016)	Height	\mathbf{m}	2	2
BIEN	Price <i>et al.</i> (2014)	Height	\mathbf{m}	27	19
BIEN	Unreferenced	Height	m	18	16
BIEN	Kleyer <i>et al.</i> (2008)	Height	m	90	19
BIEN	Unreferenced	Height	m	10	10
BIEN	Moles, Angela; unreferenced	Height	m	21	14
BIEN	Reams, Greg; unreferenced	Height	m	47036	19
BIEN	Grime, Hodgson, & Hunt; unreferenced	Height	m	5	5
BIEN	Unreferenced	Height	m	8	5
BIEN	Pérez-de Lis et al. (2017)	Height	m	18	1
BIEN	Robinson et al. (2015)	Height	m	120	1
BIEN	Anderson-teixeira et al. (2015)	Height	m	20	1
TRY	Bond-Lamberty et al. (2002)	Height	m	2	1
TRY	Unpublished	Height	m	275	3
TRY	Wright <i>et al.</i> (2004)	Height	m	28	19
TRY	Prentice et al. (2011)	Height	\mathbf{m}	2	2
TRY	Schweingruber & Landolt (2010)	Height	m	21	21
TRY	Unpublished	Height	m	35	2
TRY	Moles <i>et al.</i> (2004)	Height	m	5	5
TRY	Cavender-Bares et al. (2006)	Height	m	1	1
TRY	Diaz et al. (2004)	Height	m	11	10
TRY	Craine <i>et al.</i> (2009)	$\widetilde{\mathrm{LNC}}$	mg/g	287	12
TRY	Wilson et al. (2000)	LNC	mg/g	44	2
TRY	Wenxuan et al. (2012)	LNC	mg/g	7	4
TRY	Yahan et al. (2013)	LNC	mg/g	7	3
TRY	Wright <i>et al.</i> (2004)	LNC	m mg/g	65	32
TRY	Prentice et al. (2011)	LNC	mg/g	3	2
TRY	Vergutz et al. (2012)	LNC	mg/g	120	20
TRY	Atkin et al. (2015)	LNC	m mg/g	24	8
TRY	Marie <i>et al.</i> (2015)	LNC	m mg/g	72	22
TRY	Cornelissen et al. (2003)	LNC	m mg/g	2	1
TRY	Unpublished	LNC	m mg/g	3216	37
TRY	Wang et al. (2017)	LNC	m mg/g	6	2
BIEN	Marx et al. (2016)	Seed mass	mg	3	3
BIEN	Unreferenced	Seed mass	$^{ m mg}$	4	2
BIEN	Liu et al. (2018)	Seed mass	$^{ m mg}$	250	37
BIEN	Ameztegui et al. (2017)	Seed mass	$^{ m mg}$	12	12
BIEN	Paine <i>et al.</i> (2015)	Seed mass	$\overline{\mathrm{mg}}$	12	7
TRY	Wilson et al. (2000)	SLA	mm2 mg-1	44	2
TRY	Unpublished	SLA	mm2 mg-1	204	3
TRY	Wright <i>et al.</i> (2004)	SLA	mm2 mg-1	93	33
TRY	Prentice et al. (2011)	SLA	mm2 mg-1	2	2
TRY	Kleyer et al. (2008)	SLA	mm2 mg-1	102	18
TRY	Unpublished	SLA	mm2 mg-1	83	2
TRY	Atkin et al. (2015)	SLA	mm2 mg-1	40	11
TRY	Marie <i>et al.</i> (2015)	SLA	mm2 mg-1	86	23
TRY	Cornelissen et al. (2003)	SLA	mm2 mg-1	615	14
TRY	Unpublished	SLA	mm2 mg-1	6307	37
TRY	Wang et al. (2017) 6	SLA	mm2 mg-1	6	2
TRY	Shipley & Vu (2002)	SLA	mm2 mg-1	20	2
TRY	Cavender-Bares et al. (2006)	SLA	mm2 mg-1	42	2
TRY	Unpublished	SLA	mm2 mg-1	1	1
TRY	Diaz <i>et al.</i> (2004)	SLA	mm2 mg-1	11	10
	·		=		

Table S2: Summary of estimates from our model of height (n=42781) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with parameters with intervals that cross zero depicting weak relationships

	mean	5%	25%	75%	95%
$\mu_{grand.trait}$	12.7	9.4	11.4	14.0	15.8
$\mu_{k,g}$	32.1	27.8	30.3	33.9	36.5
μ_{force}	-10.7	-15.5	-12.6	-8.8	-6.2
μ_{chill}	-4.1	-10.8	-6.8	-1.4	2.6
μ_{photo}	1.1	-2.6	-0.3	2.5	4.6
$\beta_{trait.force}$	0.2	-0.2	0.0	0.3	0.5
$\beta_{trait.chill}$	-0.5	-1.0	-0.7	-0.3	-0.1
$\beta_{trait.photo}$	-0.2	-0.5	-0.4	-0.1	0.0
$\sigma_{species}$	5.9	4.8	5.4	6.4	7.3
σ_{study}	7.5	5.8	6.7	8.3	9.7
σ_{trait}	5.4	5.4	5.4	5.4	5.4
σ_{pheno}	15.1	11.8	13.8	16.4	18.6
σ_{force}	5.0	3.3	4.1	5.7	7.1
σ_{chill}	8.5	5.6	7.0	9.8	12.4
σ_{photo}	3.2	2.0	2.6	3.8	4.8
σ_d	14.2	13.8	14.0	14.3	14.6

Table S3: Summary of estimates from our model of specific leaf area (SLA, n=7656) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with parameters with intervals that cross zero depicting weak relationships

	mean	5%	25%	75%	95%
$\mu_{grand.trait}$	16.8	14.5	15.8	17.8	19.3
$\mu_{k,g}$	31.3	27.2	29.6	33.0	35.6
μ_{force}	-11.4	-16.2	-13.1	-9.6	-7.2
μ_{chill}	-16.7	-24.6	-19.8	-13.3	-9.1
μ_{photo}	1.9	-2.4	0.2	3.6	5.8
$\beta_{trait.force}$	0.2	-0.1	0.1	0.3	0.4
$\beta_{trait.chill}$	0.3	-0.1	0.2	0.5	0.7
$\beta_{trait.photo}$	-0.2	-0.5	-0.3	-0.1	0.0
$\sigma_{species}$	7.8	6.4	7.1	8.4	9.4
σ_{study}	3.3	2.0	2.6	3.8	5.1
σ_{trait}	6.2	6.1	6.1	6.2	6.3
σ_{pheno}	13.9	10.7	12.4	15.3	17.6
σ_{force}	5.0	3.3	4.2	5.6	7.0
σ_{chill}	10.6	7.2	8.9	12.0	14.8
σ_{photo}	3.5	2.3	2.9	3.9	5.0
σ_d	14.2	13.8	14.0	14.3	14.6

Table S4: Summary of estimates from our model of seed mass (n=281) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with parameters with intervals that cross zero depicting weak relationships

	mean	5%	25%	75%	95%
$\mu_{grand.trait}$	1.9	1.0	1.5	2.2	2.7
$\mu_{k,g}$	31.4	27.1	29.6	33.1	35.8
μ_{force}	-8.2	-10.8	-9.2	-7.1	-5.6
μ_{chill}	-9.4	-14.0	-11.2	-7.5	-4.8
μ_{photo}	-1.3	-3.4	-2.1	-0.4	0.8
$\beta_{trait.force}$	-0.3	-1.4	-0.8	0.1	0.9
$\beta_{trait.chill}$	-1.1	-2.9	-1.8	-0.4	0.7
$\beta_{trait.photo}$	-0.6	-1.5	-0.9	-0.2	0.4
$\sigma_{species}$	1.6	1.3	1.5	1.7	2.0
σ_{study}	1.0	0.8	0.9	1.0	1.1
σ_{trait}	0.2	0.2	0.2	0.3	0.3
σ_{pheno}	14.8	11.2	13.3	16.3	18.6
σ_{force}	4.9	3.4	4.2	5.5	6.7
σ_{chill}	10.7	7.1	8.8	12.1	15.4
σ_{photo}	3.6	2.3	3.0	4.1	5.1
σ_d	14.1	13.7	14.0	14.3	14.5

Table S5: Summary of estimates from our model of leaf nitrogen content (LNC, n=3853) using data from 37 focal species. Values represent the mean estimate for model parameters as well as the 50% and 90% uncertainty interval (UI). The strength of the relationship can be assessed by comparing across UI, with parameters with intervals that cross zero depicting weak relationships

	mean	5%	25%	75%	95%
$\mu_{grand.trait}$	22.6	20.4	21.7	23.5	24.9
$\mu_{k,g}$	31.1	27.1	29.4	32.8	35.3
μ_{force}	-19.3	-27.9	-22.9	-15.8	-10.5
μ_{chill}	-27.1	-38.5	-31.7	-22.4	-15.4
μ_{photo}	-9.4	-17.0	-12.5	-6.2	-1.9
$\beta_{trait.force}$	0.5	0.1	0.3	0.6	0.8
$\beta_{trait.chill}$	0.7	0.2	0.5	0.9	1.2
$\beta_{trait.photo}$	0.3	0.0	0.2	0.4	0.6
$\sigma_{species}$	5.1	4.2	4.7	5.5	6.2
σ_{study}	3.6	2.2	2.9	4.1	5.3
σ_{trait}	5.1	5.0	5.1	5.2	5.2
σ_{pheno}	14.0	10.9	12.7	15.3	17.4
σ_{force}	4.6	3.0	3.8	5.2	6.6
σ_{chill}	8.9	6.1	7.5	10.1	12.5
σ_{photo}	3.6	2.4	3.0	4.1	5.0
σ_d	14.2	13.8	14.0	14.3	14.6

Stan model code

```
6
  data {
     int<lower = 1> n_spec;
     int<lower = 1> N;
     int<lower = 1, upper = n_spec> trait_species[N];
10
     int<lower = 1> n_study;
11
     int<lower = 1, upper = n_study> study[N];
12
     vector[N] yTraiti;
13
14
     int<lower = 1> Nph;
15
     int<lower = 1, upper = n_spec> phenology_species[Nph];
16
     vector[Nph] yPhenoi;
17
     vector[Nph] forcei;
18
     vector[Nph] chilli;
19
     vector[Nph] photoi;
20
21
22
   parameters{
23
     real mu_grand;
25
     vector[n_spec] muSp;
     vector[n_study] muStudy;
27
     real<lower = 0> sigma_traity;
28
     real<lower = 0> sigma_sp;
29
     real<lower = 0> sigma_study;
30
31
     real alphaForceSp[n_spec];
32
     real muForceSp;
33
     real<lower = 0> sigmaForceSp;
     real alphaChillSp[n_spec];
35
     real muChillSp;
     real<lower = 0> sigmaChillSp;
37
     real alphaPhotoSp[n_spec];
38
     real muPhotoSp;
     real<lower = 0> sigmaPhotoSp;
40
     real alphaPhenoSp[n_spec];
41
     real muPhenoSp;
42
     real<lower = 0> sigmaPhenoSp;
     real betaTraitxForce;
44
     real betaTraitxChill;
     real betaTraitxPhoto;
46
     real<lower = 0> sigmapheno_y;
47
48
49
   transformed parameters{
50
51
     vector[N] y_hat;
52
     vector[n_spec] mu_grand_sp;
53
     real betaForceSp[n_spec];
```

```
real betaPhotoSp[n_spec];
56
     real betaChillSp[n_spec];
57
58
     for(i in 1:n_spec){
59
       mu_grand_sp[i] = mu_grand + muSp[i];
60
61
     for (i in 1:N){
62
       y_hat[i] = mu_grand + muSp[trait_species[i]] + muStudy[study[i]];
63
64
65
     for (isp in 1:n_spec){
66
       betaForceSp[isp] = alphaForceSp[isp] + betaTraitxForce * (mu_grand_sp[isp]);
68
     for (isp in 1:n_spec){
       betaPhotoSp[isp] = alphaPhotoSp[isp] + betaTraitxPhoto * (mu_grand_sp[isp]);
70
71
     for (isp in 1:n_spec){
72
       betaChillSp[isp] = alphaChillSp[isp] + betaTraitxChill * (mu_grand_sp[isp]);
73
     }
74
   }
75
76
   model{
77
     yTraiti ~ normal(y_hat, sigma_traity);
     muSp ~ normal(0, sigma_sp);
79
     muStudy ~ normal(0, sigma_study);
     mu_grand ~ normal(20,10);
81
     sigma_sp ~ normal(4,5);
82
     sigma_study ~ normal(2,5);
83
     sigma_traity ~ normal(3,5);
85
     for (i in 1:Nph){
       yPhenoi[i] ~ normal(alphaPhenoSp[phenology_species[i]] +
87
       betaForceSp[phenology_species[i]] * forcei[i] +
       betaPhotoSp[phenology_species[i]] * photoi[i] +
89
       betaChillSp[phenology_species[i]] * chilli[i],
       sigmapheno_y);
91
     }
92
93
     alphaPhenoSp ~ normal(muPhenoSp, sigmaPhenoSp);
     alphaForceSp ~ normal(muForceSp, sigmaForceSp);
     alphaChillSp ~ normal(muChillSp, sigmaChillSp);
     alphaPhotoSp ~ normal(muPhotoSp, sigmaPhotoSp);
     muPhenoSp ~ normal(40,10);
99
     sigmaPhenoSp ~ normal(5,5);
100
     sigmapheno_y ~ normal(10,5);
102
103
     muForceSp ~ normal(-15,10);
104
     sigmaForceSp ~ normal(5,5);
105
106
     muChillSp ~ normal(-15,10);
107
```

```
sigmaChillSp ~ normal(5,5);
108
109
     muPhotoSp ~ normal(-15,10);
110
     sigmaPhotoSp ~ normal(5,5);
111
112
     betaTraitxForce ~ normal(0,1);
113
     betaTraitxPhoto ~ normal(0,1);
114
     betaTraitxChill ~ normal(0,1);
116
117 }
118
```

19 References

- Ameztegui, A., Paquette, A., Shipley, B., Heym, M., Messier, C. & Gravel, D. (2017) Shade tolerance and the functional trait: demography relationship in temperate and boreal forests. *Functional Ecology* 31, 821–830.
- Anderson-teixeira, K.J., Mcgarvey, J.C., Muller-landau, H.C., Park, J.Y., Gonzalez-akre, E.B., Herrmann, V., Bennett, A.C., So, C.V., Bourg, N.A., Thompson, J.R., Mcmahon, S.M. & Mcshea, W.J. (2015) Size-related scaling of tree form and function in a mixed-age forest. *Functional Ecology* **29**, 1587–1602.
- Atkin, O., Bloomfield, K., Reich, P., Tjoelker, M., Asner, G., Bonal, D., Bönisch, G., Bradford, M., 127 Cernusak, L., Cosio, E., Creek, D., K.Y., C., Domingues, T., Dukes, J., Egerton, J., Evans, J., 128 Farquhar, G., Fyllas, N., Gauthier, P., Gloor, E., Gimeno, T., Griffin, K., Guerrieri, R., Heskel, M., 129 Huntingford, C., Ishida, F., Kattge, J., Lambers, H., Liddell, M., Lloyd, J., Lusk, C., Martin, R., 130 Maksimov, A., Maximov, T., Malhi, Y., Medlyn, B., Meir, P., Mercado, L., Mirotchnick, N., Ng, D., 131 Niinemets, Ü., O'Sullivan, O., Phillips, O., Poorter, L., Poot, P., Prentice, I., Salinas, N., Rowland, 132 L., Ryan, M., Sitch, S., Slot, M., Smith, N., Turnbull, M., VanderWel, M., Valladares, F., Veneklaas, 133 E., Weerasinghe, L., Wirth, C., Wright, I., Wythers, K., Xiang, J., Xiang, S. & Zaragoza-Castells, 134 J. (2015) Global variability in leaf respiration in relation to climate, plant functional types and leaf 135 traits. New Phytologist 206, 614-636. 136
- Bond-Lamberty, B., Wang, C. & Gower, S.T. (2002) Aboveground and belowground biomass and sapwood area allometric equations for six boreal tree species of northern Manitoba. *Canadian Journal* of Forest Research 32, 1441–1450.
- Cavender-Bares, J., Keen, A. & Miles, B. (2006) Phylogenetic structure of floridian plant communities
 depends on taxonomic and spatial scale. *Ecology* 87, 109–122.
- Cornelissen, J.H.C., Cerabolini, B., Castro-Diez, P., Villar-Salvador, P., Montserrat-Marti, G.,
 Puyravaud, J.P., Maestro, M., Werger, M.J.A. & Aerts, R. (2003) Functional traits of woody plants:
 correspondence of species rankings between field adults and laboratory-grown seedlings? *Journal of Vegetation Science* 14, 311–322.
- Craine, J.M., Elmore, A.J., Aidar, M.P.M., Bustamante, M., Dawson, T.E., Hobbie, E.A., Kahmen,
 A., Mack, M.C., Mclauchlan, K.K., Michelsen, A., Nardoto, G.B., Pardo, L.H., Penuelas, J., Reich,
 P.B., Schuur, E.A.G., Stock, W.D., Templer, P.H., Virginia, R.A., Welker, J.M. & Wright, I.J.
 (2009) Global patterns of foliar nitrogen isotopes and their relationships with climate, mycorrhizal fungi, foliar nutrient concentrations, and nitrogen availability. New Phytologist 183, 980–992.
- Diaz, S., Hodgson, J.G., Thompson, K., Cabido, M., Cornellissen, J.H.C., Jalili, A., Montserrat-Marti,
 G., Grime, J.P., Zarrinkamar, F., Asri, Y., Band, S.R., Basconcelo, S., Castro-Diez, P., Funes,
 G., Hamzehee, B., Khoshnevi, M., Pérez-Harguindeguy, N., Pérez-Rontomé, M.C., Shirvany, F.A.,
 Vendramini, F., Yazdani, S., Abbas-Azimi, R., Bogaard, A., Boustani, S., Charles, M., Dehghan,
 M., de Torres-Espuny, L., Falczuk, V., Guerrero-Campo, J., Hynd, A., Jones, G., Kowsary, E.,
 Kazemi-Saeed, F., Maestro-Martinez, M., Romo-Diez, A., Shaw, S., Siavash, B., Villar-Salvador,
 P. & Zak, M.R. (2004) The plant traits that drive ecosystems: Evidence from three continents.

 Journal of Vegetation Science 15, 295–304.
- Kleyer, M., Bekker, R.M., Knevel, I.C., Bakker, J.P., Thompson, K., Sonnenschein, M., Poschlod, P.,
 Groenendael, J.M.V., Klime, L., Klimesova, J., Klotz, S., Rusch, G.M., Hermy, M., Adriaens, D.,
 Boedeltje, G., Bossuyt, B., Dannemann, A., Endels, P., Götzenberger, L., Hodgson, J.G., Jackel,
 A.k., Kühn, I., Kunzmann, D., Ozinga, W.A., Römermann, C., Stadler, M., Schlegelmilch, J.,
 Steendam, H.J., Tackenberg, O., Wilmann, B., Cornelissen, J.H.C., Eriksson, O., Garnier, E. &
 Peco, B. (2008) The LEDA Traitbase: a database of life-history traits of the Northwest European
 flora. Journal of Ecology 96, 1266-1274.

- Liu, K., Eastwood, R.J. a d Flynn, S., Turner, R. & Stuppy, W. (2018) Kew database.
- Marie, V., Wright, I.J., Prentice, I.C., Batjes, N.H., Bhaskar, R., van Bodegom, P.M., Cornwell, W.K.,
 Ellsworth, D., Niinemets, Ü., Ordonez, A., Reich, P.B. & Santiago, L.S. (2015) Global effects of soil
 and climate on leaf photosyncthetic traits and rates. Global Ecology and Biogeography 24, 706-717.
- Marx, H.E., Giblin, D.E., Dunwiddie, P.W. & Tank, D.C. (2016) Deconstructing Darwin's Naturalization using community phylogenetics and functional traits. *Diversity and Distributions* **22**, 318–331.
- Mchugh, N., Edmondson, J.L., Gaston, K.J., Leake, J.R. & Sullivan, O.S.O. (2015) Modelling shortrotation coppice and tree planting for urban carbon management – a citywide analysis. *Journal of* Applied Ecology **52**, 1237–1245.
- Moles, A.T., Falster, D.S., Leishman, M.R. & Westoby, M. (2004) Small-seeded species produce more
 seeds per square metre of canopy per year, but not per individual per lifetime. *Journal of Ecology* 92, 384–396.
- Morales-Castilla, I., Davies, T.J., Legault, G., Buonaiuto, D.M., Chamberlain, C.J., Ettinger, A.K.,
 Garner, M., Jones, F.A.M., Loughnan, D., Pearse, W.D., Sodhi, D. & Wolkovich, E.M. (2024)
 Phylogenetic estimates of species-level phenology improve ecological forecasting. Nature Climate
 Change 14, 989–995.
- Paine, C.E.T., Amissah, L., Auge, H., Baraloto, C., Baruffol, M., Bourland, N., Bruelheide, H.,
 Dainou, K., de Gouvenain, R.C., Doucet, J.l., Doust, S., Fine, P.V.A., Fortunel, C., Haase, J., Holl,
 K.D., Jactel, H., Li, X., Kitajima, K., Koricheva, J., Martínez-Garza, C., Messier, C., Paquette, A.,
 Philipson, C., Piotto, D., Poorter, L., Posada, J.M., Potvin, C., Rainio, K., Russo, S.E., Ruiz-jaen,
 M., Scherer-lorenzen, M., Webb, C.O., Wright, S.J., Zahawi, R.A. & Hector, A. (2015) Globally,
 functional traits are weak predictors of juvenile tree growth, and we do not know why. Journal of
 Ecology 103, 978-989.
- Pérez-de Lis, G., Olano, J.M., Rozas, V., Rossi, S., Vázquez-Ruiz, R.A. & García-Gonzalez, I. (2017)
 Environmental conditions and vascular cambium regulate carbon allocation to xylem growth in
 deciduous oaks. Functional Ecology 31, 592-603.
- Prentice, I.C., Meng, T., Wang, H., Harrison, S.P., Ni, J. & Wang, G. (2011) Evidence of a universal scaling relationship for leaf CO 2 drawdown along an aridity gradient. *New Phytologist* **190**, 169–180.
- Price, C.A., Wright, I.J., Ackerly, D.D., Niinemets, Ü., Reich, P.B. & Veneklaas, E.J. (2014) Are leaf
 functional traits 'invariant' with plant size and what is 'invariance' anyway? Functional Ecology 28,
 1330–1343.
- Robinson, K.M., Hauzy, C., Loeuille, N. & Albrectsen, B.R. (2015) Relative impacts of environmental variation and evolutionary history on the nestedness and modularity of tree-herbivore networks.

 Ecology and Evolution 5, 2898–2915.
- ²⁰⁰ Schweingruber, F. & Landolt, W. (2010) The xylem database.
- Shipley, B. & Vu, T.T. (2002) Dry matter content as a measure of dry matter concentration in plants and their parts. New Phytologist 153, 359–364.
- Vergutz, L., Manzoni, S., Porporato, A., Novais, R. & Jackson, R. (2012) A Global Database of
 Carbon and Nutrient Concentrations of Green and Senesced Leaves. Oak Ridge National Laboratory
 Distributed Active Archive Center Oak Ridge, Tennessee, U.S.A.
- Wang, H., Harrison, S.P., Prentice, I.C., Yang, Y., Bai, F., Furstenau Togashi, H., Wang, M., Zhou,
 S. & Ni, J. (2017) The China Plant Trait Database. PANGAEA.

- Wenxuan, H., Yahan, C., Fang-Jie, Z., Tang, L., Rongfeng, J. & Fusuo, Z. (2012) Floral, climatic and
 soil pH controls on leaf ash content in China's terrestrial plants. Global Ecology and Biogeography
 21, 376–382.
- Wilson, K.B., Baldocchi, D.D. & Hanson, P.J. (2000) Spatial and seasonal variability of photosynthetic parameters and their relationship to leaf nitrogen in a deciduous forest. *Tree Physiology* **20**, 565–578.
- Wright, I.J., Westoby, M., Reich, P.B., Oleksyn, J., Ackerly, D.D., Baruch, Z., Bongers, F., Cavender-Bares, J., Chapin, T., Cornellissen, J.H.C., Diemer, M., Flexas, J., Gulias, J., Garnier, E., Navas, M.L., Roumet, C., Groom, P.K., Lamont, B.B., Hikosaka, K., Lee, T., Lee, W., Lusk, C., Midgley, J.J., Niinemets, Ü., Osada, H., Poorter, H., Pool, P., Veneklaas, E.J., Prior, L., Pyankov, V.I., Thomas, S.C., Tjoelker, M.G. & Villar, R. (2004) The worldwide leaf economics spectrum. *Nature* 428, 821–827.
- Yahan, C., Wenxuan, H., Luying, T., Zhiyao, T. & Jingyun, F. (2013) Leaf nitrogen and phosphorus concentrations of woody plants differ in responses to climate, soil and plant growth form. *Ecography* 36, 178–184.