

Phylogenetic estimates of species-level phenology improve ecological forecasting

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Abstract

Our ability to adapt to climate change requires accurate ecological forecasting to predict shifts in key ecosystem services, such as carbon storage and biodiversity maintenance. Current forecasts, however, have generally failed to capture the important variability in biological responses, especially observed across species. Here, using novel Bayesian hierarchical phylogenetic models, we overcome this challenge to estimate species-level responses. We illustrate our method with an dataset on phenological responses to temperature and daylength cues, determined experimentally for 191 woody species. We find that variation is greater across species than across cues, suggesting the current focus on identifying which cue is the strongest may be less important than predicting how each species responds to a combination of cues. Further, our model provides insights on how evolutionary history has shaped responses to cues, suggesting stronger evolutionary dynamics in responses to temperature—especially cool winter temperatures—than daylength. Our approach provides a major advance in ecological forecasting, with implications for predicting the impacts of climate change and other anthropogenic forces on species interactions and ecosystem functioning.

¹ Introduction

² The biological impacts of climate change will have major implications for ecosystem functioning
³ and stability. With rising global temperatures many species have shifted their geographic distri-
⁴ butions northward in space and recurring life-history events—their phenology—earlier in time
⁵ (IPCC, 2014; Parmesan and Yohe, 2003), against a background of high variability. These shifts
⁶ have cascading consequences on many ecosystem services including carbon storage, making both
⁷ mitigation and human adaptation to future warming dependent on accurate ecological forecasts
⁸ (Richardson et al., 2013).

⁹ While ecological forecasting has improved over recent years (Dietze, 2017; Lewis et al., 2022),
¹⁰ it remains a challenge to reproduce the high variability observed in biological responses such
¹¹ as phenology, physiology or demography to environmental cues (IPCC, 2014). Some of this
¹² variability results from the complexity of climate change itself, including regional and seasonal
¹³ variation in warming that underlies average trends alongside shifts in other climate axes (e.g.,
¹⁴ precipitation). Much of it, however, could be driven by species-specific variation, reflecting
¹⁵ evolved differences in species sensitivities to underlying environmental cues and their interac-
¹⁶ tions. Unfortunately, we can only estimate the sensitivities to cues for a few well-studied species
¹⁷ (Chuine and Regniere, 2017; Ettinger et al., 2020). In the absence of detailed data on individual
¹⁸ species, species groupings (e.g., functional groups) have improved ecosystem models (Moorcroft
¹⁹ et al., 2001; Griffith et al., 2020), but still capture only a fraction of the important variability
²⁰ (Fuccillo Battle et al., 2022).

²¹ Recent efforts that have attempted to model species-specific responses to the environment (Diez
²² et al., 2012) are often restricted by data availability—especially the common problem that data
²³ are often prevalent for some species and sparse across others. The rise of Bayesian hierarchical
²⁴ models can allow inference across species in such cases. However, underlying most hierarchical
²⁵ models is an implicit assumption that species are exchangeable (all species represent samples
²⁶ drawn from the same underlying distribution, Gelman and Hill, 2006), and they thus partially
²⁷ pool ('shrink') towards estimates for species with the most data and least variable responses,
²⁸ making inference at the species-level unreliable (Ettinger et al., 2020). More reliable estimates
²⁹ of species-level responses would allow us to better incorporate species differences into models of
³⁰ ecosystem change.

³¹ Including the evolutionary history of species relationships in models of species responses could
³² provide more robust species-level estimates than current approaches and a better understanding
³³ of the evolutionary constraints that might limit adaptation to change. For example, strong
³⁴ phylogenetic niche conservatism (Wiens et al., 2010) could potentially inhibit adaptive responses
³⁵ by drawing species back to an evolutionary conserved optimum, which is sub-optimal under new
³⁶ conditions. While incorporating such evolutionary history is traditionally seen as necessary,
³⁷ either as a statistical correction or to better understand species evolutionary history, the use of
³⁸ such phylogenetic information should also improve model fitting and forecasts (Freckleton et al.,

39 2002).

40 Research using long-term observational data has highlighted the role that evolutionary history
41 may play in structuring plant phenological responses—which are critical to accurate forecasts
42 of carbon storage. Phylogenetic signal in plant phenology, including dates of budburst, leafout
43 and first flowering (Kochmer and Handel, 1986; Willis et al., 2008; Davies et al., 2013), suggests
44 that more closely related species share more similar phenologies, likely reflecting evolutionary
45 conservatism in responses to common cues. There are two broad explanations for why we might
46 expect phylogenetic conservatism in phenological traits. First, close relatives will tend to share
47 similar ecologies and physiologies, and thus be sensitive to similar environmental pressures.
48 Second, close relatives derive from common geographic centers of origin, and thus their an-
49 cestors will have been exposed to—and have adapted to—similar environmental cues (Davies
50 et al., 2013). However, approaches using traditional phylogenetic comparative methods, have
51 produced conflicting results, with some studies reporting evidence of phylogenetic structure in
52 phenology-linked species declines (e.g., Willis et al., 2008) and in some phenophases, such as first
53 flowering, but not others, such as peak or last flowering (e.g., CaraDonna and Inouye, 2014),
54 and in responses to some cues but not others (e.g., Yang et al., 2021). In addition, evidence for
55 phylogenetic conservatism of phenological responses appears to depend on method and species,
56 even varying between sites with overlapping species sets (e.g., Rafferty and Nabity, 2017), which
57 violates the fundamental idea of shared evolutionary history (the common ancestor of two sets of
58 species cannot possess two separate evolutionary histories for the same trait). A first challenge is
59 thus how to better integrate evolutionary history into multi-species models of plant phenological
60 responses.

61 Generating robust ecological forecasts requires addressing a second major hurdle—underlying
62 environmental cues that are complex and interacting. Decades of research have informed our
63 understanding of how species use environmental cues to time their phenotypic responses with
64 the temporal distribution of key resources and to avoid periods of high abiotic or biotic stress
65 (Larcher, 1980; Bonamour et al., 2019). Commonly, however, responses to environmental cues,
66 and their evolution, are studied individually, linking a given phenotypic response to a single cue,
67 for example, time of leafout responding to summed heat during early spring (Davies et al., 2013).
68 These efforts fail to capture the more likely scenario for most phenotypic traits in which multiple
69 cues interacting along evolutionary history have shaped species responses (Ackerly, 2009). For
70 many plant species, phenological events are determined by a combination of temperature and
71 light (Chuine and Regnieri, 2017), with additional factors (e.g., other cues—like humidity, or
72 species physiology—vasculature or leaf structure) likely further mediating species responses.
73 Although these mediating factors are not well understood (Chuine and Regnieri, 2017), they
74 can be accounted for in models either as latent processes or by allowing non-stationarity in
75 responses across species (Davies et al., 2019).

76 Spring plant phenology may represent our best opportunity to improve forecasts of species
77 responses to interacting environmental cues. Beyond being the most studied biological impact

of climate change, the primary cue system is well established (Chuine and Regniere, 2017), especially for temperate woody species where phenology is generally thought to be determined by two components of temperature—chilling (cool temperatures during dormancy period over winter) and forcing (warm temperatures, generally in the spring)—and photoperiod (Ettinger et al., 2021). Plant phenology is also one of few phenotypic traits with extensive experimental data on responses to multiple environmental cues across species. Recent multi-species analyses considering forcing, chilling and photoperiod have shown that chilling and forcing together often determine complex non-linear responses to warming, but cannot forecast beyond several well-studied species (Ettinger et al., 2020).

Here we present a novel Bayesian framework that extends upon phylogenetic mixed models (Housworth et al., 2004) to examine how chilling, forcing (both metrics of temperature) and photoperiod together determine spring plant phenology. By allowing non-stationarity in species responses across the phylogeny (Davies et al., 2019), our model departs from previous work and assumptions of traditional phylogenetic comparative methods concerned with phylogenetic correction (e.g., Freckleton et al., 2002), and moves towards integrating evolutionary history in models of phenological responses to environmental change. To understand how evolution has shaped the cues underlying shifting phenology with climate change (Uyeda et al., 2017), we explicitly incorporate phylogenetic structure across model intercepts and slopes (that is, allowing a separate model of evolutionary history for chilling, forcing and photoperiod, see Methods & Materials for a complete description).

We illustrate our method with an unprecedented dataset on phenological responses to environmental cues (chilling, forcing and photoperiod) determined experimentally for 191 deciduous woody species (by far the most studied group of species in phenology experiments, see Ettinger et al., 2020), in an updated version of the Observed Spring Phenology Responses in Experimental Environments (OSPRE) database (Wolkovich et al., 2019). These data combined with the Smith and Brown (2018) megatree adjusted to our species, and modeling approach allows us to address the common question of which cue has the largest effect on budburst and, at the same time, provide robust estimates of how cues vary across species. Using spring phenology, we identify historical regime shifts (Uyeda et al., 2017) in phenological responses, and highlight how our approach could advance forecasting of other critical responses to ongoing global change.

Results & Discussion

Most species respond to all three primary cues—forcing, chilling, and photoperiod (Fig. 1, Supporting Table S2)—with responses to chilling approximately five-fold greater than to photoperiod (phenological advances of 6.9 days per standardized unit vs 1.2 days, for chilling and photoperiod, respectively; see Table S2). We estimated lower average responses to temperature compared to a model without phylogeny (model slopes for forcing and chilling decreased by 18% and 22%, respectively); responses to chilling and forcing were also more similar when including

115 phylogeny (though chilling was still greater: 6.9 vs. 6.1 per standard unit), which contrasts with
116 previous results suggesting chilling responses are much greater than forcing (Laube et al., 2014;
117 Ettinger et al., 2020).

118 These average estimates, however, fail to capture the large differences in species responses to
119 both chilling and forcing (Fig. 1, Supporting Table S6). By allowing species responses to
120 vary, based on a model including their shared evolutionary history, we found species differences
121 dwarfed the mean differences between cues, especially temperature cues (Fig. 1). The largest cue
122 in magnitude—chilling—varied 24-fold between species, while variation to forcing varied 7-fold.
123 This variation indicates large differences between chilling and forcing occur at the species-level
124 and not across species (as previously suggested, Laube et al., 2014; Ettinger et al., 2020). These
125 results highlight why robust phenological forecasts must account for both the complexity of
126 multiple cues and species-level variation in responses to them.

127 *Differences across clades & cues*

128 The large differences across species produced striking differences between clades. For example,
129 several groups—oaks and beeches (Fagaceae), elms (Ulmaceae) and buckthorns (Rhamnaceae)—
130 are highly sensitive to chilling while others—rhododendrons (Ericaceae), butterfly bushes (Scro-
131 phulariaceae) and spindles (Celastraceae)—show little to no response to chilling (Fig. 1a).
132 Similar clade-level variation was observed for forcing, where some of these clades—e.g., Eri-
133 caceae, Rhamnaceae, Ulmaceae, or Fagaceae—were particularly sensitive (advancing their bud-
134 burst more than 10 days per standardized unit of forcing) and others, such as the Sapindaceae,
135 Cornaceae or Juglandaceae, show little response (Fig. 1b).

136 Some species responded strongly to both temperature cues, which could suggest the existence
137 of syndromes where the genetic basis for responses to one cue—e.g., forcing—has been selected for alongside responses to another cue—e.g., chilling. This could occur if selection
138 operates jointly on responses to both cues; for example, if sensitivity to multiple cues provides
139 greater insurance against leafing out before the last frost (Bonamour et al., 2019; Wolkovich and
140 Donahue, 2021). Additionally, linkage or pleiotropism among loci associated with different cues
141 (Nakagawa et al., 2005) could induce across-cue correlations. However, the correlation in species
142 responses across cues was generally weak ($r = 0.31$; between forcing and chilling) and some
143 genera, such as *Tilia* and *Rhododendron* (Ericaceae), displayed strong responses to forcing but
144 weak responses to chilling, while others, such as *Acer* (Sapindaceae), show moderately strong
145 responses to chilling but weak responses to forcing (Fig. 1). Species sensitivity to one cue, thus,
146 does not constrain sensitivity to another cue, and it seems selection can operate independently
147 on responses to different cues (Bonamour et al., 2019).

149 In contrast to temperature cues (chilling and forcing), species-level responses to photoperiod were
150 almost uniform across species. This consistency provides novel insight on a large debate over
151 the prevalence of photoperiod cues in temperate trees, where previous experiments (Basler and
152 Körner, 2012; Zohner et al., 2016) and models (e.g., Hunter and Lechowicz, 1992; Schaber and

153 Badeck, 2003) suggested important variability across species that may constrain the responses
154 of certain species to warming (Way and Montgomery, 2015). Our results indicate variability is
155 limited to a handful of species in Fagaceae, which have been particularly well studied, especially
156 *Fagus sylvatica* (e.g., Basler and Körner, 2012; Zohner et al., 2016; Kramer et al., 2017). As
157 *Fagus sylvatica* is nearly five times more sensitive to photoperiod than most other measured tree
158 species, our results caution against using it to draw inferences of photoperiod responses more
159 widely. These same few species are also where most evidence of local adaptation in photoperiod
160 cues for spring phenology comes from (e.g., Kramer et al., 2017), in contrast with common
161 garden studies of other species, which find little evidence of local adaptation in spring (but not
162 fall) phenology (Aitken and Bemmels, 2016). The uniformity of response to photoperiod in our
163 results supports this latter view of generally low local adaptation in photoperiod cues for spring
164 phenology (i.e., if local adaptation were high in photoperiod cues we would have expected more
165 variability across species).

166 *Phylogenetic structure of phenological cues*

167 Variation—or lack thereof—in cues across species and clades provides possible insights into
168 the evolution of cues across the phylogeny. While responses to each cue were phylogenetically
169 structured, with closely related species exhibiting more similar sensitivities than distantly related
170 species, the strength of phylogenetic conservatism in response differed between cues (Fig. 2).
171 Responses to temperature (forcing and chilling) were moderately structured ($\lambda = 0.65$ and
172 $\lambda = 0.54$, for forcing and chilling, respectively). Phylogenetic structure in species responses to
173 photoperiod was comparatively weak ($\lambda = 0.4$; see Fig. 2, Table S2).

174 Differences among species in their temperature responses represent shifts in the slope of the
175 relationship between the observed phenology and the cue. The observed phylogenetic structure
176 in temperature responses (forcing and chilling) would be consistent with an interaction with a
177 latent trait that moderates responses, and which also covaries with phylogeny (Davies et al.,
178 2019). This fits fundamentally with the idea that early-season phenology plays a critical role in
179 shaping species temporal niches (Gotelli and Graves, 1996) and thus should covary with a suite
180 of life-history traits, including whether species are early-active with rapid return on investment
181 traits, or start later in the season and have traits associated with higher competitive abilities
182 (e.g., Grime, 1977; Wolkovich and Donahue, 2021).

183 Weak phylogenetic signal in photoperiod sensitivity (Fig. 2) might seem at odds with the unifor-
184 mity of species response—i.e., there is very little variation in the responses to photoperiod across
185 species. However, somewhat counterintuitively, both uniform and random responses can man-
186 ifest as low phylogenetic signal when indexed by Brownian motion expectations (Wiens et al.,
187 2010). While rapid local adaptation within species might erase the phylogenetic structure in
188 photoperiod responses, it does not agree with the uniformity we find in species responses. How-
189 ever, if responses to photoperiod evolved early in plants, as seems likely (Serrano-Bueno et al.,
190 2017), and subsequent selection on photoperiod sensitivity was constrained by stabilizing selec-

tion operating on other life-history attributes sensitive to photoperiod (e.g., Rinne et al., 1994; Wilczek et al., 2014; Azeez and Sane, 2015), we would predict both low interspecific variation and weak phylogenetic signal in responses, matching observations. This latter interpretation is also consistent with our estimates of lower σ for photoperiod responses (Fig. 2). Here, as in more traditional phylogenetic comparative methods, σ represents the rate of evolution, and thus our results suggest photoperiod responses are also evolving slower than temperature responses (see Fig. S12).

Phylogenetic conservatism (high λ) and slow evolutionary rates (low σ) in traits has sometimes been interpreted as indicative of evolutionary constraints to adaptive change (Wiens et al., 2010; Bennett et al., 2021). If this were the case, we might then conclude that species where responses are dominated by forcing cues might be more vulnerable to future warming because phylogenetic conservatism (λ) in forcing is higher compared to other cues and its evolutionary rate (σ) is lower than that estimated for chilling. This is misleading, however, as estimates of λ are independent from the rate of evolution, and macroevolutionary rates are estimated on phylogenetic trees that integrate across millions of years of evolutionary history, and thus do not necessarily inform us of maximum possible rates of evolution over much shorter timescales. Indeed, there is accumulating evidence for rapid evolution to shifting climates (Bradshaw and Holzapfel, 2006; Franks et al., 2014). Our estimates are thus more useful in providing unique insights into the evolutionary history of phenological cues, and emphasize the critical importance of incorporating species-level differences in ecological forecasts.

211 *Forecasting species-level responses*

Our results highlight that species-level variability can be extremely high—when properly estimated. Our approach, which partially pooled species responses based on their shared evolutionary history, estimated substantially higher variation across species compared with more widely used hierarchical models. This was especially noticeable in temperature responses (for chilling variance across species means, $var(\beta_{chill,j}$ from eqn. 2), was estimated as 23.55 in the phylogenetic model, versus 17.47 in the non-phylogenetic model; variance across means, $var(\beta_{force,j})$: 8.75 compared to 5.01) while photoperiod, which had low phylogenetic structure, was more similar across approaches (variance across means, $var(\beta_{photo,j})$: 0.83 compared to 0.64).

The consequences of including shared evolutionary history in forecasting are most apparent for poorly sampled species nested within more well-sampled clades. For example, forecasts for *Acer campestre*, which has only 6 observations, shift by up to 35% in the number of days until budburst after forcing starts, when comparing our phylogenetically informed model to one without phylogeny (see Fig. 4 for details). In contrast, forecasts for *Betula pendula*, which is one of the most sampled species, are nearly identical across models (Fig. 4). This occurs because cue estimates for *Acer campestre* in the phylogenetically informed model are strongly influenced by other *Acer* species, which diverge from other clades. In the non-phylogenetically informed model all species are equally exchangeable and thus *Acer campestre* is pulled strongly towards

229 well-sampled species, such as *Betula pendula* ($n = 311$), leading to forecasted shifts that are
230 more similar across all species (Fig. 4).

231 The increase in variability across species in our model with phylogenetic structure also decreased
232 the uncertainty in estimates for each individual species temperature responses (Fig. S7). Thus,
233 traditional (non-phylogenetically informed) approaches that partially pool across species (most
234 hierarchical models in ecology, e.g., Flynn and Wolkovich, 2018; Ettinger et al., 2020) may also
235 lead to less precise predictions and forecasts of phenology for individual species, although overall
236 model accuracy might still appear reasonable (see Fig. S8). Another advantage of our Bayesian
237 approach is that we are also better able to accommodate imprecision in the data that informs
238 our model, which might arise from multiple sources, including measurement or experimental
239 error, and the general stochasticity associated with limited sample sizes and unbalanced species
240 representation. Critically, by partially pooling across species and weighting by phylogeny, we
241 gain strength from species estimates that are informed by more data, such as within *Betula* and
242 *Fagaceae*, but avoid skewing estimates for phylogenetically distant clade that may have been
243 exposed to different selective regimes. Robustness of species estimates was confirmed by cross-
244 validation, showing the phylogenetic model better predicted observed values for held-out data,
245 and yielded more stable species coefficients compared to a hierarchical model (Figs. S2-S1; see
246 “Leave-One-Clade-Out model cross validation”).

247 The contrasts between temperature and photoperiod responses—in both their variability across
248 species and phylogenetic structure—have important implications for generating multi-species
249 forecasts. Notably, responses to photoperiod appear weaker, more uniform across species, and
250 less phylogenetically constrained compared to temperature. For temperature responses, the
251 large variability among species makes predicting species-level responses challenging, but the
252 phylogenetic structure in responses lets us borrow information from close relatives to improve
253 our predictions. However, given that Brownian motion (our assumed model of evolution) is an
254 extremely noisy process, we recommend imputation only for missing taxa that are closely related
255 to other well sampled species or clades (Molina-Venegas et al., 2018, 2023).

256 While we focused on spring phenology here, our approach suggests a path forward for more
257 general forecasting of species-level climate change responses. Our results show how including
258 the phylogenetic relationship of species in a mechanistic model of underlying cues can overcome
259 major limitations of most current hierarchical models—correcting biased model estimates, es-
260 timating the full variability across species and reducing uncertainty around individual species
261 estimates—while at once providing insight into the evolutionary history of biological responses.
262 Using this approach improved forecasts of phenological responses to climate change and could
263 help anticipate impacts on critical ecosystem services from species-level shifts and thus aid mit-
264 igation and human adaption to warming.

265 **Methods & Materials**

266 **Phenological and Phylogenetic Data**

267 *Phenological data:* To estimate phenological responses to chilling, forcing and photoperiod we
268 used data from phenological experiments in controlled environments of temperate woody species,
269 brought together in the Observed Spring Phenology Responses in Experimental Environments
270 (OSPREE) database. In July 2019, we updated an earlier version of this database (Wolkovich
271 et al., 2019) by reviewing all papers found through searching ISI Web of Science and Google
272 Scholar with the following terms:

- 273 1. TOPIC = (budburst OR leaf-out) AND (photoperiod OR daylength) AND temperature*,
274 which yielded 623 publications
- 275 2. TOPIC = (budburst OR leaf-out) AND dorman*, which yielded 270 publications

276 We scraped data from all papers of woody species that tested for photoperiod and/or tempera-
277 ture effects on budburst, leafout, or flowering, resulting in data from 155 experiments across 97
278 papers in the updated database. Ettinger et al. (2020) used a portion (72 experiments across
279 49 papers) of the earlier OSPREE database and provides extensive methods on the database
280 creation and cleaning.

281 We focused on angiosperms (as gymnosperms are very poorly represented in spring phenology
282 experiments), and included all budburst experiments where we could quantify chilling, forcing
283 and photoperiod levels, resulting in 44 studies from 33 papers, resulting in 2940 data points. In
284 our dataset most studies come from Europe ($n=37$) and a few from North America ($n=7$). The
285 same bias towards Europe is found across the full OSPREE dataset with less North American
286 ($n=19$) than European ($n=60$) studies and only 3 studies located in the Southern Hemisphere.
287 Given our need of daily gridded data for estimating chilling we only include studies from Eu-
288 rope and North America, with most of these sites in temperate areas and a few in European
289 Mediterranean areas (see Fig. S3). Our final dataset is both geographically and taxonomically
290 limited, mirroring the existing literature on phenological experiments. Enlarging this body of
291 literature will be critical to address important research questions, such as whether variability in
292 cue sensitivity across species is larger in temperate than in tropical latitudes.

293 Across experiments chilling treatments were often fully or partially applied in the field, thus we
294 estimated chilling—both in the field and applied in controlled environments—using Utah units
295 with the **chillR** package. We estimated field chilling from 1 September to the date given for when
296 samples were taken from the field using daily temperature data (converted to hourly) from both
297 European (E-OBS, version 16, calculating the average of minimum and maximum daily temps,
298 Cornes et al., 2018) and North American (v3, Sheffield et al., 2006) gridded climate datasets.
299 We also converted experimental chilling into Utah chill units, based on reported treatments
300 (for studies with a mix of field and experimental treatments, we added field and experimentally

301 applied Utah units). To avoid numerical instability in our models (from having predictor values
302 on very different scales), we divided Utah units by 240 (roughly equivalent to 10 days of average
303 chilling).

304 We report the Utah model because a small number of studies reported chilling only in Utah
305 units, thus using this common metric allowed us to include the most data. The Utah model
306 relies on the assumption that temperatures between 1.4 and 15.9C affect endodormancy release
307 differently, though recent findings show possibly similar effects for a wide range of temperatures
308 (-2 to 10C, see Baumgarten et al., 2021). Because chilling is a latent process an accurate model
309 of it, especially for the 191 species in our dataset, is not currently possible (Ettinger et al., 2020).
310 We found consistent results, however, using another common model of chilling—chill portions,
311 suggesting our results are robust to the exact chilling metric used (see Tables S4 and S5 in
312 Supporting Information).

313 Forcing and photoperiod treatments occurred after chilling treatments; we report photoperiod as
314 the length of light and weighted these treatments by the reported photo- and thermo-periodicity
315 (Buonaiuto et al., 2023). Most studies reported two temperatures per day across the whole
316 experiment, one for day and night, but some had ramped temperatures and/or photoperiods (or
317 other complexities). In these cases we built an hourly model of the full treatment period until
318 budburst and took the mean value.

319 For a phylogenetic tree, we pruned the megatree for seed plants (Smith and Brown, 2018)
320 to extract the sub-tree containing only the species present in the OSPREE dataset, species
321 not included in the megatree were added to the congeneric basal node age (using the function
322 ‘congeneric.merge’ in Pearse et al., 2015), and assigned branch lengths to maintain tree ultra-
323 metricity. This addition of species to the tree can introduce polytomies (multifurcations) when
324 many species are added to the same ancestral node, as was the case for *Acer*, for which several
325 species where included in the OSPREE dataset but the megatree lacked species-level resolution
326 within the genus. In total, our pruned tree had 8 polytomies affecting 46 out of 191 species.
327 Our Bayesian hierarchical model, described below, is informed by the phylogenetic structure de-
328 scribing species evolutionary relationships. Errors in phylogenetic topology and branching times
329 could thus impact model estimates, although if errors were large the contribution of phylogeny
330 would simply be scaled to zero. To assess whether the inclusion of polytomies in our data biased
331 model estimates, we ran sensitivity analyses excluding these species from models (see Table S8
332 in Supporting Information). Our approach assumes a tree where branch lengths represent time,
333 but it could be possible to change this assumption. For example, if the genes underlying plant
334 responses to particular cues were known, branch lengths would directly represent mutational
335 changes along gene sequences. In the absence of such detailed gene specific data, evolutionary
336 time provides a useful proxy for species differences.

337

338 **Bayesian hierarchical phylogenetic model**

339 Commonly used phylogenetic regression methods today (e.g., phylogenetic generalized least
340 squares models, PGLS, Freckleton et al. (2002); phylogenetic mixed models, PMM, Hous-
341 worth et al. (2004)) were originally conceived as statistical corrections for phylogenetic non-
342 independence across observations—generally species—thus allowing multi-species studies to meet
343 the assumptions of linear regression (Freckleton et al., 2002). These corrections incorporated
344 phylogenetic structure by estimating the magnitude of a transformation of a variance-covariance
345 (VCV) matrix whose elements were derived from the amount of evolutionary history (branch
346 lengths) shared between species on a phylogeny. The most commonly used transformation was
347 Pagel’s λ —a multiplier of the off-diagonal elements—where estimates of $\lambda = 1$ essentially left the
348 VCV untransformed and suggested that the residuals of the regression had phylogenetic signal
349 consistent with Brownian motion; estimates of $\lambda = 0$ suggested no phylogenetic signal. Be-
350 cause the original aim of these methods was to correct for statistical bias introduced by shared
351 evolutionary history among species, the underlying assumption of phylogenetic regressions is
352 that phylogenetic relatedness would only affect either model residuals (in PGLS approaches,
353 Freckleton et al., 2002) or model intercepts (e.g., in many PMM approaches, Housworth et al.,
354 2004).

355 Because our aim is to understand how evolution may have imprinted biological responses to
356 multiple interactive cues, our approach expands the above methods by explicitly incorporating
357 phylogenetic structure across model intercepts and slopes. Doing so allows explicitly estimating
358 the amount of phylogenetic relatedness in species sensitivities to each cue, when these sensitivities
359 are modelled in a multi-predictor regression setting.

360 For each observation i of species j , we assumed that the timing of phenological events were
361 generated from the following sampling distribution:

$$y_{i,j} \sim \mathcal{N}(\mu_j, \sigma_e^2) \quad (1)$$

where

$$\mu_j = \alpha_j + \beta_{chill,j} X_{chill} + \beta_{force,j} X_{force} + \beta_{photo,j} X_{photo} \quad (2)$$

362 and σ_e^2 represents random error unrelated to the phylogeny.

363 Predictors X_{chill} , X_{force} , X_{photo} are standardized chilling, forcing, and photoperiod, and their
364 effects on the phenology of species j are determined by parameters $\beta_{chill,j}$, $\beta_{force,j}$, $\beta_{photo,j}$,
365 representing species responses (or sensitivities) to each of the cues. These responses, including
366 the species-specific intercept α_j , are elements of the following normal random vectors:

$$\boldsymbol{\alpha} = [\alpha_1, \dots, \alpha_n]^T \text{ such that } \boldsymbol{\alpha} \sim \mathcal{N}(\mu_{\alpha}, \boldsymbol{\Sigma}_{\alpha}) \quad (3)$$

$$\boldsymbol{\beta}_{chill} = [\beta_{chill,1}, \dots, \beta_{chill,n}]^T \text{ such that } \boldsymbol{\beta}_{chill} \sim \mathcal{N}(\mu_{\beta_{chill}}, \boldsymbol{\Sigma}_{\beta_{chill}})$$

$$\boldsymbol{\beta}_{force} = [\beta_{force,1}, \dots, \beta_{force,n}]^T \text{ such that } \boldsymbol{\beta}_{force} \sim \mathcal{N}(\mu_{\beta_{force}}, \boldsymbol{\Sigma}_{\beta_{force}})$$

$$\boldsymbol{\beta}_{photo} = [\beta_{photo,1}, \dots, \beta_{photo,n}]^T \text{ such that } \boldsymbol{\beta}_{photo} \sim \mathcal{N}(\mu_{\beta_{photo}}, \boldsymbol{\Sigma}_{\beta_{photo}})$$

where the means of the multivariate normal distributions are root trait values (i.e., values of cue responses prior to evolving across a phylogenetic tree) and $\boldsymbol{\Sigma}_i$ are $n \times n$ phylogenetic variance-covariance matrices of the form:

$$\begin{bmatrix} \sigma_i^2 & \lambda_i \times \sigma_i \times \rho_{12} & \dots & \lambda_i \times \sigma_i \times \rho_{1n} \\ \lambda_i \times \sigma_i \times \rho_{21} & \sigma_i^2 & \dots & \lambda_i \times \sigma_i \times \rho_{2n} \\ \vdots & \vdots & \ddots & \vdots \\ \lambda_i \times \sigma_i \times \rho_{n1} & \lambda_i \times \sigma_i \times \rho_{n2} & \dots & \sigma_i^2 \end{bmatrix} \quad (4)$$

where σ_i^2 is the rate of evolution across a tree for a given trait or predictor (here assumed to be constant along all branches), λ_i scales branch lengths and therefore is a measure of the phylogenetic signal or extent of phylogenetic relatedness on each model parameter (i.e., α_j , $\beta_{force,j}$, $\beta_{photo,j}$), and ρ_{xy} is the phylogenetic correlation between species x and y , or the fraction of the tree shared by the two species.

The above specification is equivalent to writing equation 2 in terms of root trait values and residuals, such that:

$$\mu_j = \mu_{\alpha} + \mu_{\beta_{chill}} X_{chill} + \mu_{\beta_{force}} X_{force} + \mu_{\beta_{photo}} X_{photo} + e_{\alpha_j} + e_{\beta_{force,j}} + e_{\beta_{chill,j}} + e_{\beta_{photo,j}} \quad (5)$$

where the residual phylogenetic error terms (e.g., e_{α_j}) are elements of normal random vectors from multivariate normal distributions centered on 0 with the same phylogenetic variance-covariance matrices as in equation 4. Model code, including priors used here, are given in the supplement. We fit all models to our data using RStan using 4 chains of 4,000 iterations with a warmup of 2,000 each (resulting 8,000 posterior samples), and assessed fit via \hat{R} near 1 and adequate effective sample sizes (see Tables S2-S3).

To assess if the phylogenetic mixed model (PMM) presents any advantages with respect commonly used hierarchical mixed models (HMM; see e.g., Ettinger et al., 2020), beyond fitting evolutionary parameters to model predictors, we compare results of PMM and HMM. HMM is a simplified version of PMM where off-diagonal elements of the variance-covariance phylogenetic matrices are multiplied by zero ($\lambda = 0$). Both models account for differences in sample sizes and variances for each species, by partially pooling across all data while at the same time providing species-level estimates; however the PMM will pool more strongly to closely-related species

when λ is high. Additional grouping factors beyond species could be added to these models. For example, similar approaches can be used to estimate study or location effects; however, many species occurred in only one study and location in our dataset, making it difficult to separate these effects, thus we do not include study as a separate parameter here (and thus average over it in our model estimates).

References

- Ackerly, D. 2009. Conservatism and diversification of plant functional traits: Evolutionary rates versus phylogenetic signal. *Proceedings of the National Academy of Sciences of the United States of America* 106:19699–19706.
- Aitken, S. N., and J. B. Bemmels. 2016. Time to get moving: assisted gene flow of forest trees. *Evol Appl* 9:271–90.
- Azeez, A., and A. P. Sane. 2015. Photoperiodic growth control in perennial trees. *Plant signaling & behavior* 10:e1087631.
- Basler, D., and C. Körner. 2012. Photoperiod sensitivity of bud burst in 14 temperate forest tree species. *Agricultural and Forest Meteorology* 165:73–81.
- Baumgarten, F., C. M. Zohner, A. Gessler, and Y. Vitasse. 2021. Chilled to be forced: the best dose to wake up buds from winter dormancy. *New Phytologist* 230:1366–1377.
- Bennett, J. M., J. Sunday, P. Calosi, F. Villalobos, B. Martínez, R. Molina-Venegas, M. B. Araújo, A. C. Algar, S. Clusella-Trullas, B. A. Hawkins, et al. 2021. The evolution of critical thermal limits of life on earth. *Nature communications* 12:1–9.
- Bonamour, S., L. M. Chevin, A. Charmantier, and C. Teplitsky. 2019. Phenotypic plasticity in response to climate change: the importance of cue variation. *Philosophical Transactions of the Royal Society B-Biological Sciences* 374.
- Bradshaw, W. E., and C. M. Holzapfel. 2006. Evolutionary response to rapid climate change. *Science* 312:1477–1478.
- Buonaiuto, D. M., M. Donahue, and E. M. Wolkovich. 2023. Experimental designs for testing the interactive effects of temperature and light in ecology: the problem of periodicity. *Functional Ecology* .
- CaraDonna, P. J., and D. W. Inouye. 2014. Phenological responses to climate change do not exhibit phylogenetic signal in a subalpine plant community. *Ecology* 96:355–361.
- Chuine, I., and J. Regniere. 2017. Process-based models of phenology for plants and animals. *Annual Review of Ecology, Evolution, and Systematics* 48:159–182.

- 419 Cornes, R. C., G. van der Schrier, E. J. van den Besselaar, and P. D. Jones. 2018. An ensemble
420 version of the E-OBS temperature and precipitation data sets. *Journal of Geophysical*
421 *Research: Atmospheres* 123:9391–9409.
- 422 Davies, T., E. Wolkovich, N. Kraft, N. Salamin, and S. E. Travers. 2013. Phylogenetic conservatism
423 in plant phenology. *Journal of Ecology* 101:1520–1530.
- 424 Davies, T. J., J. Regetz, E. M. Wolkovich, and B. J. McGill. 2019. Phylogenetically weighted
425 regression: A method for modelling non-stationarity on evolutionary trees. *Global ecology*
426 and biogeography 28:275–285.
- 427 Dietze, M. 2017. Ecological forecasting. In *Ecological Forecasting*. Princeton University Press.
- 428 Diez, J. M., I. Ibáñez, A. J. Miller-Rushing, S. J. Mazer, T. M. Crimmins, M. A. Crimmins,
429 C. D. Bertelsen, and D. W. Inouye. 2012. Forecasting phenology: from species variability to
430 community patterns. *Ecology letters* 15:545–553.
- 431 Ettinger, A., C. Chamberlain, I. Morales-Castilla, D. Buonaiuto, D. Flynn, T. Savas, J. Samaha,
432 and E. Wolkovich. 2020. Winter temperatures predominate in spring phenological responses
433 to warming. *Nature Climate Change* pages 1–6.
- 434 Ettinger, A. K., D. M. Buonaiuto, C. J. Chamberlain, I. Morales-Castilla, and E. M. Wolkovich.
435 2021. Spatial and temporal shifts in photoperiod with climate change. *New Phytologist*
436 230:462–474.
- 437 Flynn, D. F. B., and E. M. Wolkovich. 2018. Temperature and photoperiod drive spring phe-
438 nology across all species in a temperate forest community. *New Phytologist* 219:1353–1362.
- 439 Franks, S. J., J. J. Weber, and S. N. Aitken. 2014. Evolutionary and plastic responses to climate
440 change in terrestrial plant populations. *Evolutionary Applications* 7:123–139.
- 441 Freckleton, R. P., P. H. Harvey, and M. Pagel. 2002. Phylogenetic analysis and comparative
442 data: a test and review of evidence. *The American Naturalist* 160:712–726.
- 443 Fuccillo Battle, K., A. Duhon, C. R. Vispo, T. M. Crimmins, T. N. Rosenstiel, L. L. Armstrong-
444 Davies, and C. E. de Rivera. 2022. Citizen science across two centuries reveals phenological
445 change among plant species and functional groups in the northeastern us. *Journal of Ecology*
446 110:1757–1774.
- 447 Gelman, A., and J. Hill. 2006. Data analysis using regression and multilevel/hierarchical models.
448 Cambridge University Press.
- 449 Gotelli, N. J., and G. R. Graves. 1996. The temporal niche. Pages 95–111 in N. J. Gotelli and
450 G. R. Graves, eds. *Null Models in Ecology*. Smithsonian Institution, Washington, DC.

- 451 Griffith, D. M., C. P. Osborne, E. J. Edwards, S. Bachle, D. J. Beerling, W. J. Bond, T. J. Gal-
452 laher, B. R. Helliker, C. E. R. Lehmann, L. Leatherman, J. B. Nippert, S. Pau, F. Qiu, W. J.
453 Riley, M. D. Smith, C. A. E. Stromberg, L. Taylor, M. Ungerer, and C. J. Still. 2020. Lineage-
454 based functional types: characterising functional diversity to enhance the representation of
455 ecological behaviour in land surface models. *New Phytologist* 228:15–23.
- 456 Grime, J. P. 1977. Evidence for existence of 3 primary strategies in plants and its relevance to
457 ecological and evolutionary theory. *American Naturalist* 111:1169–1194.
- 458 Housworth, E. A., E. P. Martins, and M. Lynch. 2004. The phylogenetic mixed model. *The
459 American Naturalist* 163:84–96.
- 460 Hunter, A. F., and M. J. Lechowicz. 1992. Predicting the timing of budburst in temperate trees.
461 *Journal of Applied Ecology* 29:597–604.
- 462 IPCC. 2014. Climate Change 2014: Impacts, Adaptation, and Vulnerability. Cambridge Uni-
463 versity Press, Cambridge, United Kingdom and New York, NY, USA.
- 464 Kochmer, J. P., and S. N. Handel. 1986. Constraints and competition in the evolution of flowering
465 phenology. *Ecological monographs* 56:303–325.
- 466 Kramer, K., A. Ducousoo, D. Gomory, J. K. Hansen, L. Ionita, M. Liesebach, A. Lorent,
467 S. Schueler, M. Sulkowska, S. de Vries, and G. von Wuehlisch. 2017. Chilling and forc-
468 ing requirements for foliage bud burst of European beech (*Fagus sylvatica* L.) differ between
469 provenances and are phenotypically plastic. *Agricultural and Forest Meteorology* 234:172–181.
- 470 Larcher, W. 1980. Plant Physiological Ecology. Springer-Verlag.
- 471 Laube, J., T. H. Sparks, N. Estrella, J. Höfler, D. P. Ankerst, and A. Menzel. 2014. Chilling
472 outweighs photoperiod in preventing precocious spring development. *Global Change Biology*
473 20:170–182.
- 474 Lewis, A. S., C. R. Rollinson, A. J. Allyn, J. Ashander, S. Brodie, C. B. Brookson, E. Collins,
475 M. C. Dietze, A. S. Gallinat, N. Juvigny-Khenafou, et al. 2022. The power of forecasts to
476 advance ecological theory. *Methods in Ecology and Evolution* .
- 477 Molina-Venegas, R., I. Morales-Castilla, and M. Á. Rodríguez. 2023. Unreliable prediction of
478 b-vitamin source species. *Nature Plants* 9:31–33.
- 479 Molina-Venegas, R., J. C. Moreno-Saiz, I. Castro Parga, T. J. Davies, P. R. Peres-Neto, and
480 M. Á. Rodríguez. 2018. Assessing among-lineage variability in phylogenetic imputation of
481 functional trait datasets. *Ecography* 41:1740–1749.
- 482 Moorcroft, P., G. Hurtt, and S. Pacala. 2001. A method for scaling vegetation dynamics: The
483 ecosystem demography model (ed). *Ecological Monographs* 71:557–585.

- 484 Nakagawa, H., J. Yamagishi, N. Miyamoto, M. Motoyama, M. Yano, and K. Nemoto. 2005.
485 Flowering response of rice to photoperiod and temperature: a qtl analysis using a phenological
486 model. *Theoretical and Applied Genetics* 110:778–786.
- 487 Parmesan, C., and G. Yohe. 2003. A globally coherent fingerprint of climate change impacts
488 across natural systems. *Nature* 421:37.
- 489 Pearse, W. D., M. W. Cadotte, J. Cavender-Bares, A. R. Ives, C. M. Tucker, S. C. Walker,
490 and M. R. Helmus. 2015. Pez: Phylogenetics for the environmental sciences. *Bioinformatics*
491 31:2888–2890.
- 492 Rafferty, N. E., and P. D. Nabity. 2017. A global test for phylogenetic signal in shifts in flowering
493 time under climate change. *Journal of Ecology* 105:627–633.
- 494 Richardson, A. D., T. F. Keenan, M. Migliavacca, Y. Ryu, O. Sonnentag, and M. Toomey. 2013.
495 Climate change, phenology, and phenological control of vegetation feedbacks to the climate
496 system. *Agricultural and Forest Meteorology* 169:156–173.
- 497 Rinne, P., A. Saarelainen, and O. Junntila. 1994. Growth cessation and bud dormancy in
498 relation to ABA level in seedlings and coppice shoots of *Betula pubescens* as affected by a
499 short photoperiod, water stress and chilling. *Physiologia Plantarum* 90:451–458.
- 500 Schaber, J., and F. Badeck. 2003. Physiology-based phenology models for forest tree species in
501 germany. *International Journal of Biometeorology* 47:193–201. International Conference on
502 the Times they are a Changin, WAGENINGEN, NETHERLANDS, DEC 05-07, 2001.
- 503 Serrano-Bueno, G., F. J. Romero-Campero, E. Lucas-Reina, J. M. Romero, and F. Valverde.
504 2017. Evolution of photoperiod sensing in plants and algae. *Current Opinion in Plant Biology*
505 37:10–17.
- 506 Sheffield, J., G. Goteti, and E. F. Wood. 2006. Development of a 50-year high-resolution global
507 dataset of meteorological forcings for land surface modeling. *Journal of Climate* 19:3088–3111.
- 508 Smith, S. A., and J. W. Brown. 2018. Constructing a broadly inclusive seed plant phylogeny.
509 *American journal of botany* 105:302–314.
- 510 Uyeda, J. C., M. W. Pennell, E. T. Miller, R. Maia, and C. R. McClain. 2017. The evolution of
511 energetic scaling across the vertebrate tree of life. *The American Naturalist* 190:185–199.
- 512 Way, D. A., and R. A. Montgomery. 2015. Photoperiod constraints on tree phenology, perfor-
513 mance and migration in a warming world. *Plant, Cell & Environment* 38:1725–1736.
- 514 Wiens, J. J., D. D. Ackerly, A. P. Allen, B. L. Anacker, L. B. Buckley, H. V. Cornell, E. I.
515 Damschen, T. Jonathan Davies, J.-A. Grytnes, S. P. Harrison, et al. 2010. Niche conservatism
516 as an emerging principle in ecology and conservation biology. *Ecology letters* 13:1310–1324.

- 517 Wilczek, A. M., M. D. Cooper, T. M. Korves, and J. Schmitt. 2014. Lagging adaptation to
518 warming climate in *Arabidopsis thaliana*. Proceedings of the National Academy of Sciences
519 111:7906–7913.
- 520 Willis, C. G., B. Ruhfel, R. B. Primack, A. J. Miller-Rushing, and C. C. Davis. 2008. Phyloge-
521 netic patterns of species loss in thoreau’s woods are driven by climate change. Proceedings of
522 the National Academy of Sciences 105:17029–17033.
- 523 Wolkovich, E. M., and M. J. Donahue. 2021. How phenological tracking shapes species and
524 communities in non-stationary environments. Biological Reviews .
- 525 Wolkovich, E. M., A. K. Ettinger, D. Flynn, T. Savas, C. Chamberlain, D. Buonaiuto, and
526 J. Samaha. 2019. Observed Spring Phenology Responses in Experimental Environments (OS-
527 PREE). doi:10.5063/F1CZ35KB.
- 528 Yang, Z., Y. Du, M. Shen, N. Jiang, E. Liang, W. Zhu, Y. Wang, and W. Zhao. 2021. Phy-
529 logenetic conservatism in heat requirement of leaf-out phenology, rather than temperature
530 sensitivity, in tibetan plateau. Agricultural and Forest Meteorology 304.
- 531 Zohner, C. M., B. M. Benito, J. C. Svenning, and S. S. Renner. 2016. Day length unlikely to
532 constrain climate-driven shifts in leaf-out times of northern woody plants. Nature Climate
533 Change 6:1120–1123.

534 **Data and code availability statement**

535 All data supporting the findings of this study are publicly available. Data analyses were per-
536 formed in R and Bayesian Hierarchical and Phylogenetic Mixed Models (HMM and PMM,
537 respectively) were performed within the Stan environment. All custom code will be made avail-
538 able through a GitHub repository for this project (see Supporting Information for details and
539 online links).

540 **Tables and Figures**

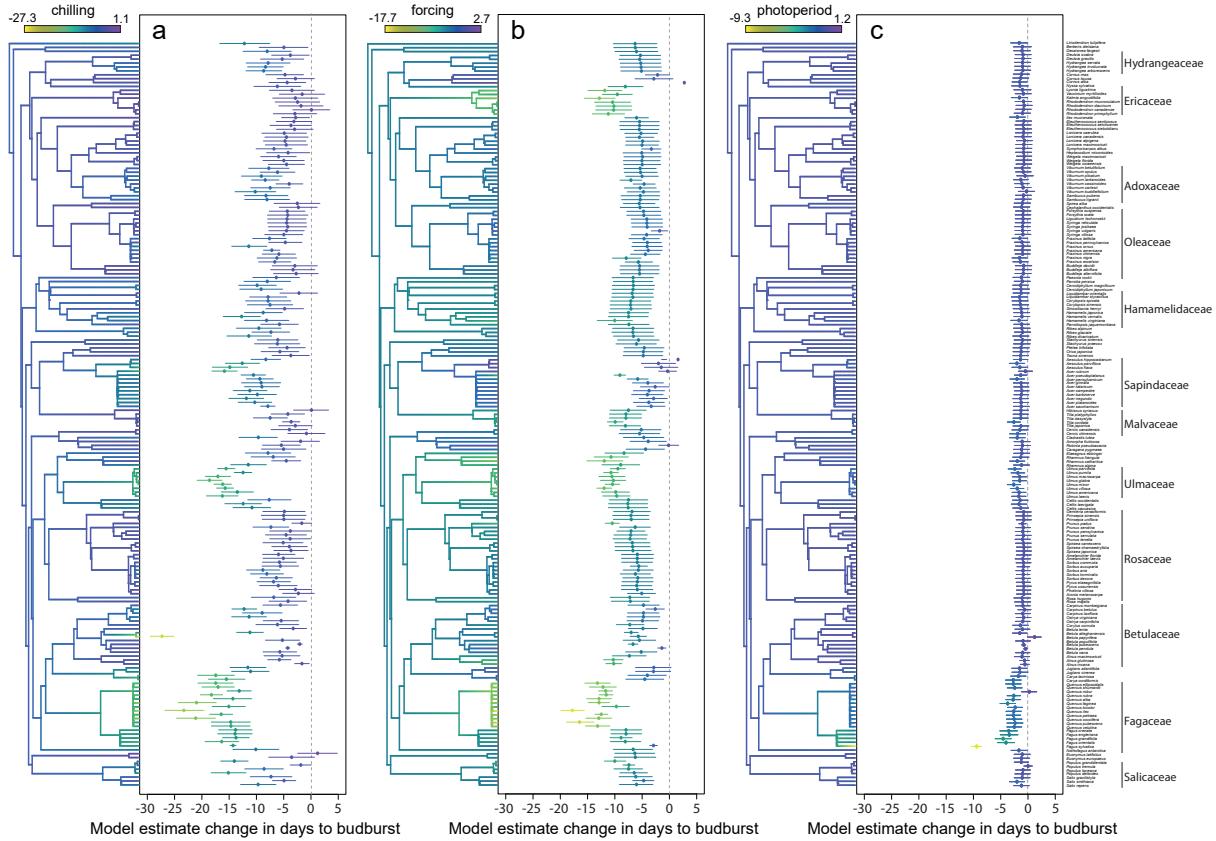


Figure 1: Phenological sensitivity to three environmental cues, chilling (a), forcing (b) and photoperiod (c) measured as change in days to budburst per standardized unit (z -transformation) of the cues across 191 woody species. The same phylogenetic tree is shown in each panel, colored according to an estimation of ancestral character states, being the states at the tips the species sensitivities to a cue, as estimated by our hierarchical phylogenetic model. Species sensitivities are shown along with 50% uncertainty intervals in the diagrams. Note that the color scale varies in each panel. Total tree depth is 81 My.

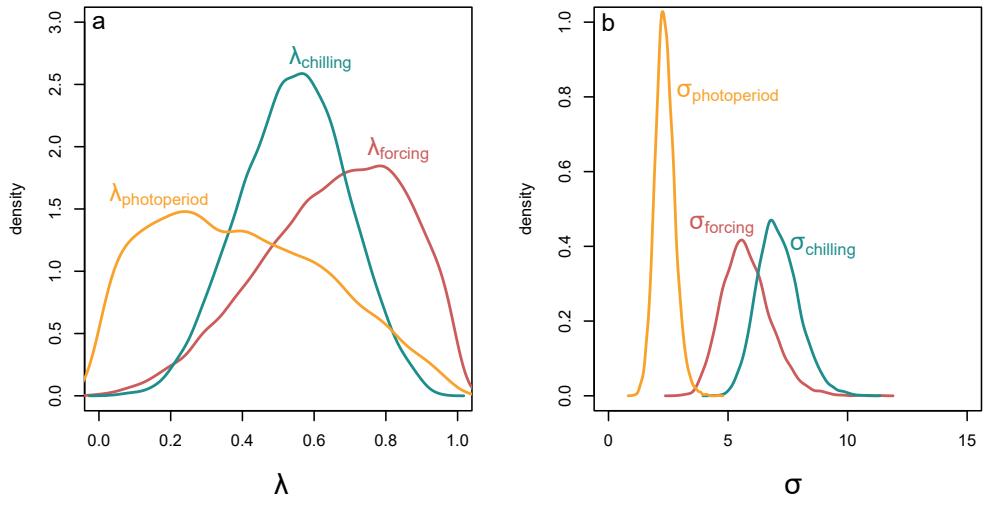


Figure 2: Density plots comparing the posterior distributions of phylogenetic parameters λ and σ estimated for each cue in the model: chilling (blue), forcing (red), and photoperiod (orange). Panels correspond to λ (a) and σ (b) from the phylogenetic model.

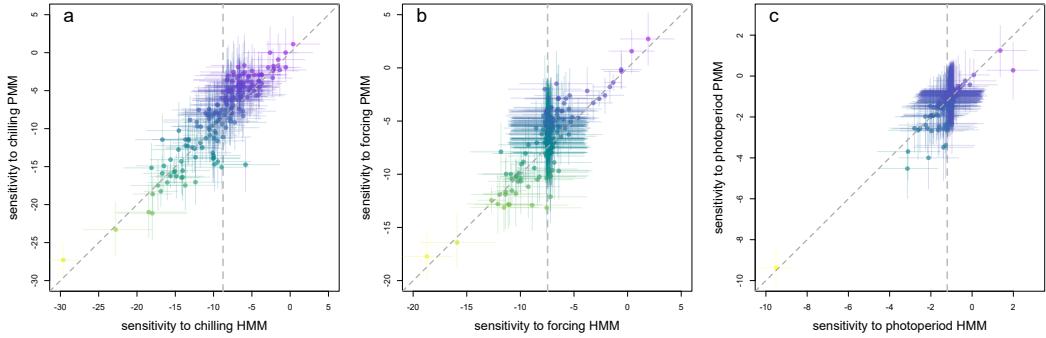


Figure 3: Correlations between model parameters as estimated by the model including phylogenetic structure on each phenological cue (phylogenetic mixed model, PMM; y -axis), and the more commonly used hierarchical mixed model (HMM) where species are exchangeable (where λ is constrained to be equal to zero, x -axis). While species with large amounts of data may be estimated similarly by both models, in the more commonly used hierarchical model (x -axis) many species are pulled towards the overall average (shown by dashed grey vertical lines). The strength and prevalence of pulling across species is particularly obvious for forcing (b). Panels correspond to sensitivity to chilling (a), forcing (b), and photoperiod (c). Dashed grey 1:1 lines also shown. Estimate colors are in the same scale as in 2, and error lines correspond to 50% uncertainty intervals.

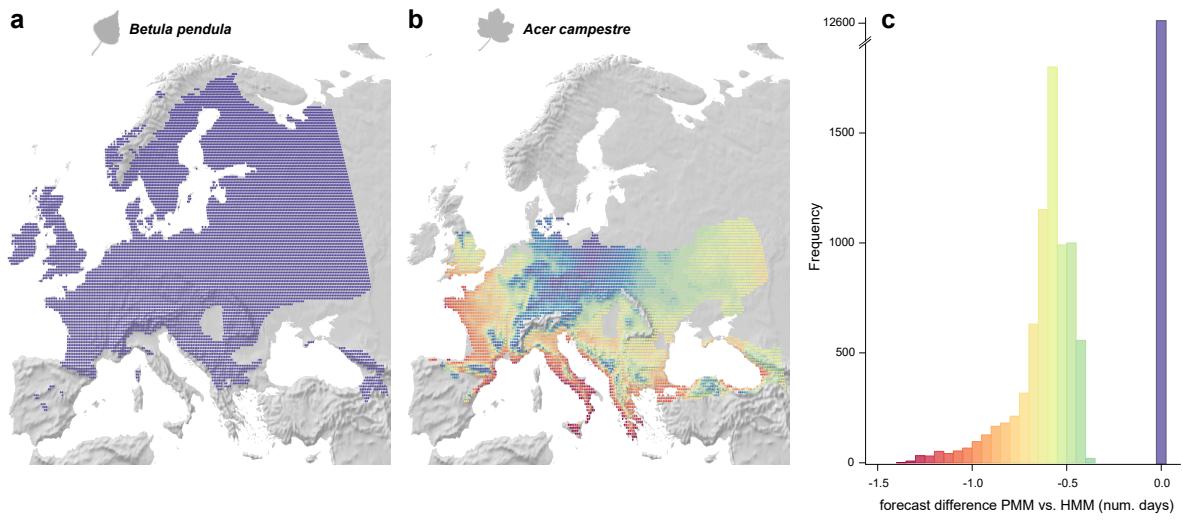


Figure 4: Comparison of forecasts of phenological shifts (i.e., computed as the difference between predictions under current climate vs. a 2°C warmer climate) resulting from a phylogenetic (PMM) and a non-phylogenetic but hierarchical (HMM) approach. Differences in forecasted shifts are negligible for well sampled species (*Betula pendula*, $n = 311$, a), but can be substantially different for poorly sampled species in well-sampled clades (*Acer campestre*, $n = 6$, b). The maps show the difference in number of days between the shifts predicted by PMM and HMM, with values colored according to histograms in panel c (days here are relative to start of forcing conditions, not calendar days). See Supplementary Material for details on forecast calculation.