

Running title: Environmental tracking

How temporal tracking shapes species and communities in stationary and non-stationary environments

E. M. Wolkovich¹ & M. J. Donahue²

¹ *Forest & Conservation Sciences, Faculty of Forestry, University of British Columbia, 2424 Main Mall, Vancouver, BC V6T 1Z4 (e.wolkovich@ubc.ca)*

² *Hawai'i Institute of Marine Biology, University of Hawai'i at Mānoa, Kānāeohe, HI 96744 (donahuem@hawaii.edu)*

Corresponding author: see¹ above; Ph: 604.827.5246 (no fax).

Authorship statement: EMW and MJD both conceived of the paper, performed modeling work and edited the paper, EMW additionally wrote the paper and did the literature review, while MJD additionally wrote the supplementary information on the model.

Data statement: Review, so no new primary data, but data from a comprehensive literature review will be archived in an appropriate public repository and the data DOI will be included at the end of the article.

Keywords: community assembly, global change, climate change, phenology, environmental variability

Article type: Reviews and Syntheses

Article information: Abstract: 194 words; Main text: 6,538; Figures: 4; Boxes: 4 (text in Box 1: 343; Box 2: 500; Box 3: 264, Box 4: 738); 115 references

Abstract

Climate change is reshaping the environments of all species. Predicting species responses requires understanding how well species can track environmental change, and how such tracking shapes communities. Growing empirical evidence suggests that environmental temporal tracking—how an organism shifts the timing of major biological events in response to proximate environmental cues—is linked to species performance and is a structuring force of species and communities today. Here, we review the concept of temporal tracking both in empirical climate change impacts studies and through the lens of community ecology theory. After reviewing how life history theory makes predictions for variation in tracking, and trade-offs with other traits, we examine how well basic community ecology theory can be extended to test the current paradigm that climate change should favor species with environmental tracking. We find that existing community assembly theory can be leveraged to understand tracking in stationary and non-stationary systems and, thus, help predict the species- and community-level consequences of climate change. But it will require greater efforts to integrate priority effects into modern coexistence theory and much improved empirical estimates of both fundamental tracking and the underlying cues that shape measures of environmental tracking.

1 Main text

Anthropogenic climate change is causing widespread changes in species distributions, with many species shifting in both time and space (IPCC, 2014; Hoegh-Guldberg *et al.*, 2018). Reports focus on species shifting to higher elevations and poleward (Chen *et al.*, 2011) shifting their recurring life history events (phenology) earlier as climate warms (Wolkovich *et al.*, 2012; Cohen *et al.*, 2018), or both (Amano *et al.*, 2014; ?). These general trends, however, hide high variability across species. A large proportion of species are not shifting at all (Cook *et al.*, 2012; Amano *et al.*, 2014), which has raised concerns about whether these species may be more vulnerable to population declines with continued warming. Such concerns come in part from increasing research that links how well species track climate change—especially through temporal shifts—to shifts in biomass, growth and other metrics related to performance (Cleland *et al.*, 2012). Tracking climate change may then be a major component to understanding and predicting the fitness consequences of climate change, including population declines, with cascading effects on community and ecosystem structure (Menzel *et al.*, 2006; Parmesan, 2006).

The hypothesis that tracking predicts fitness outcomes with climate change has gained significant traction in the ecological literature focused on global change (e.g., Cleland *et al.*, 2012) and several areas of theory support it. Considering tracking as a form of phenotypic flexibility (Piersma & Drent, 2003), evolutionary models predict species that track will be favored in novel environmental conditions (Chevin *et al.*, 2010). Niche models of community assembly suggest that a warming climate should open up new temporal niche space and favor species that can exploit that space (Gotelli & Graves, 1996; Wolkovich & Cleland, 2011; Zettlemoyer *et al.*, 2019). Yet not all studies find the purported link (e.g., Block *et al.*, 2019), and there has been comparatively little work to improve predictions by formally connecting tracking to community assembly theory.

This disconnect could be because most ecological theory today is for stationary systems (e.g., Chesson & Huntly, 1997). While major arenas of research such as ‘modern coexistence theory’ or population ecology are now built on assumptions of a stochastic environment, they generally still assume stationarity, where the underlying distribution of the environment is unchanged across time (i.e., constant mean and variance, Barabas *et al.*, 2018). This assumption is common to much of the theory that underlies ecology, evolution, and myriad other research fields (e.g., Milly *et al.*, 2008; Nosenko *et al.*, 2013).

Climate change upends the assumption of stationarity. By causing increases in temperature, larger pulses of precipitation, shifts in snowpack, increased drought, and more storms (Stocker *et al.*, 2013), climate change has fundamentally shifted major attributes of the environment from stationary to non-stationary regimes (see Fig. 1 and Box: Environmental variability & change). This transition is reshaping ecological systems. While new work has aimed to adapt coexistence theory to non-stationary environments (Chesson, 2017; Rudolf, 2019), there is still little theoretical work on what such a transition may mean for communities and the species within them, despite growing empirical studies.

Here, we review the concept of tracking used in the empirical climate change impacts literature and in related ecological theory. We provide definitions that distinguish between fundamental

and environmental tracking, highlighting the distinction between measuring tracking and its fitness outcomes in empirical systems. Then, after briefly reviewing evolutionary theory that predicts variation in tracking across species and environments, we focus on ecological theory that could advance our understanding of tracking on ecological timescales. Specifically, we examine how well community assembly theory—especially priority effects and modern coexistence theory—can be extended to test the current paradigm that climate change should favor species with environmental tracking.

1.1 Defining & measuring tracking

Defining tracking

Tracking is a commonly used word in the climate change literature, especially relating to phenology (e.g., Menzel *et al.*, 2006; Parmesan, 2006; Cleland *et al.*, 2012; Deacy *et al.*, 2018). Yet there are few, if any, definitions of it. Conceptual and theoretical treatments of tracking often relate how well an organism matches the timing of a life history event to the ideal timing for that event, what we refer to as ‘fundamental tracking’. In contrast, empirical studies of tracking often focus on estimating a change in the timing of an event per unit change in an environmental variable, something closer to what we refer to as ‘environmental tracking’—the change in timing of a major biological event due to an organism’s cue system given change in the environment. Both these definitions are readily applied to phenology—the timing of recurring life history events—though they can also apply to non-recurring life history events (e.g., seed germination), or events not normally defined as part of life history (e.g., spring algal blooms).

Fundamental tracking rests on an assumption that there is a timing (an ‘ideal timing’) that yields maximum fitness, and event timings moving away from this ideal result in reduced fitness. This is a foundational concept of the trophic mismatch literature (Visser & Gienapp, 2019), which often assumes the peak timing of a resource defines the ideal timing for phenological events dependent on that resource (e.g. egg laying dates dependent on caterpillar abundance, Visser & Both, 2005). For most phenological events, however, fitness outcomes are likely dependent on a suite of interacting forces (e.g., Reed *et al.*, 2013)—for example, egg laying dates for migratory birds may depend both on the timing of peak prey abundance and the need to leave nesting grounds before winter. Whatever their full underlying causes, fitness consequences of suboptimal timing should drive the evolution of phenological cues to predict and best match the timing of events to the ideal (maximum fitness) timing (Fig. 2). These cues combined with environmental variation define what we refer to as temporal environmental tracking (henceforth, ‘environmental tracking’).

Environmental tracking depends on the intersection of the environment’s variability—which aspects of the environment vary, how (e.g., temporally each year, spatially at x scale) and how much—and an organism’s response to the environment via its proximate cues. If the varying components of the environment are not in the organism’s set of cues, then the organism does not ‘track’ per this definition. Environmental tracking at the individual-level is a purely plastic response to environmental variation (with the plasticity itself an outcome of selection, Chevin *et al.*, 2010). At the population-level, tracking may also incorporate evolutionary change in the

cue system, depending on both the timescales of study and the species' generation time. This evolutionary response can be predicted as the difference between the environmental sensitivity of phenotypic selection and an organism's plasticity ($|B - b|$ in Chevin *et al.*, 2010). Given our focus on current responses to climate change, we focus on environmental tracking here as a mainly plastic response (Bonamour *et al.*, 2019), though over longer timescales or in certain systems it should be shaped by selection (Franks & Hoffmann, 2012).

Measuring tracking

Measuring 'tracking' and comparing variation in it across species, space and time is a rapidly growing area of ecological research (e.g., Cook *et al.*, 2012; Fu *et al.*, 2015; Thackeray *et al.*, 2016; Cohen *et al.*, 2018). Studies estimating fundamental tracking are uncommon (but see Visser *et al.*, 2006; Charmantier *et al.*, 2008), given in part the difficulty of measuring fitness, though many studies in the synchrony literature attempt to link consumer change to resource change, with an assumption that the resource is the dominant determinant of ideal timing for the consumer, though this may rarely be true (Singer & Parmesan, 2010; Johansson & Jonzen, 2012; Reed *et al.*, 2013). Instead, most studies focus on estimating something akin to environmental tracking. Some studies estimate simply change in days over time (e.g., Parmesan, 2007; Kharouba *et al.*, 2018), though most studies now estimate shifts in response to temperature (for example, multiple meta-analyses show plants' spring phenology shifts with spring or annual temperatures 4-6 days/°C on average across species, Richardson *et al.*, 2006; Wolkovich *et al.*, 2012; Thackeray *et al.*, 2016) or precipitation (??).

All species-rich studies of phenology-climate relationships find high variation (Cook *et al.*, 2012; Thackeray *et al.*, 2016), including some species that do not track or track poorly (i.e., high noise surrounding observed statistical relationships). Researchers have worked to link such variation to the underlying cues (e.g., Cook *et al.*, 2012), species traits (e.g., Cohen *et al.*, 2018) and trophic level (e.g., Thackeray *et al.*, 2016). These approaches hint at the three majors classes of reasons that underlie species that do not appear to track climate or appear to track poorly: (1) species do not track, as environmental tracking may either not be possible or optimal for all species (Simons, 2011), (2) lack of firm biological understanding of the cues that underlie tracking (Chmura *et al.*, 2019), and (3) statistical artifacts that make it difficult to measure tracking robustly (see Box 'Challenges in measuring tracking').

Limited understanding of organisms' phenological cues combined with statistical issues may make many current estimates of variation in tracking less reliable than they appear. This in turn makes robust quantitative analyses across species difficult (Brown *et al.*, 2016; Kharouba *et al.*, 2018), yeilding only muddy picture of which species, when, and where, do and do not track. Given this current difficulty, we believe clear testable predictions from ecological theory are especially critical to guide research (Smaldino & McElreath, 2016).

1.2 Tracking in single-species environments

Predicting variation in environmental tracking in stationary systems

Considering phenology as a trait (as we and others do, e.g., Charmantier *et al.*, 2008; Nicotra *et al.*, 2010; Forsman, 2015; Inouye *et al.*, 2019), environmental tracking is one type of

plasticity, and thus should be predictable based on evolutionary theory of when, and to what degree, organismal traits should be plastic. Generally there should be selection for plasticity in heterogeneous but predictable environments where there is the underlying architecture for plasticity and strong selection on that architecture (Piersma & Drent, 2003; Reed *et al.*, 2010). Strong inheritance of the underlying architecture of tracking seems likely as many phenological cues are strongly heritable (CITES). Selection, however, can be lower than expected from reaction norms predicted by simple models of plasticity (CITES) for many reasons, including unavoidable trade-offs with tracking (Singer & Parmesan, 2010; Johansson & Jonzen, 2012), gene flow from other environments that may continually push a population away from its local optimum (Lenormand, 2002), limits due to standing genetic variation (Franks *et al.*, 2007; Ghalambor *et al.*, 2015), or deeper evolutionary history that may produce co-evolved traits making it difficult for selection to act solely on tracking (Ackerly, 2009). The extent to which these constraints will limit selection depends in large part on the underlying machinery of tracking—an organisms’ cues for the event.

While plasticity theory predicts the conditions under which plastic traits should evolve, optimal control theory provides a conceptual framework for understanding the cues evolved for tracking, considering the costs, benefits, and constraints of an organism in its environment. (Donahue *et al.*, 2015). In this context, the cost of cues includes the machinery an organism uses to monitor its environment (e.g., accumulated temperature or daylength), while the fitness benefits are the increased reproductive success that arises from better timing (e.g., how much tissue is saved by avoiding a coldsnap). Constraints can arise from the organisms itself or from its environment. Organismal constraints include the availability of machinery to track the environment as well as other fundamental differences in life history—for example, the type and amount of loss an organism can sustain each season is limited by its generation time and other attributes related to long-lived lifestages that yield buffered population growth (Chesson & Huntly, 1997). Environmental constraints include the fundamental predictability of the environment: are there early season environmental variables that can predict later season phenomena? Apparently poor cues may occur for organisms in environments where there is both a low cost and low benefit to the cue(s). Similarly, expensive cues, such as complex multivariate ones, are possible given a high pay-off. Most in-depth studies of species’ phenological cues find evidence for complex multivariate ones (Chuine & Regniere, 2017). These cues almost always appear adapted to handle unusual—though not completely uncommon—years when the simple cue alone would fail (that is, would trigger growth, reproduction or another life history event at a suboptimal time), suggesting that multivariate cues may provide a large benefit in best coupling the timing of cues to the ultimate controls (i.e., cues that couple environmental tracking strongly to fundamental tracking).

Tracking should generally not be favored in unpredictable environments, or environments where species otherwise face high uncertainty in the timing of investment decisions; instead theory suggests the optimal strategy may often be to bet-hedge (Venable, 2007; Donaldson-Matasci *et al.*, 2012; de Casas *et al.*, 2015) via a high diversity of timings or one conservative timing. Because bet-hedging, by definition, maximizes geometric-mean fitness in the long-run, its short-term outcomes can appear maladaptive. How often observed ‘maladaptations,’ which may easily include species that do not track or appear to track poorly, are actually the outcome of bet-

hedging is difficult to estimate, as robustly assessing bet-hedging requires studies of fitness over longer timescales than many current field experiments (Simons, 2011).

Environmental variation often includes both predictable and less predictable aspects. In such cases theory predicts organisms may evolve tracking that is a mixed strategy between bet-hedging and plasticity (Wong & Ackerly, 2005). Taken together, life history theory provides multiple reasons species may not track or track weakly, suggesting that—at least in stationary systems—we should expect a number of species that do not track.

Predicting variation in environmental tracking in non-stationary systems

Expectations from life history theory of which species should track are generally based on assumptions of stationarity, thus a major open area of research is adapting life history theory to non-stationary environments. Multivariate cues may be especially robust to a non-stationary environment if they provide a tight coupling of cues to fundamental tracking, and that coupling is maintained in the non-stationary environment (Dore *et al.*, 2018). But multivariate cues may equally be most vulnerable to failure if non-stationarity decouples the cues from fundamental tracking (Bonamour *et al.*, 2019). For example, consider an organism’s whose cues evolved based on a correlation between peak prey abundance and daylength: the daylength cue that could be reliable in a stationary environment (generally predicting peak prey abundance based on daylength, with some interannual variation), but would become unreliable if warming advances peak prey abundance. Predicting the outcome of non-stationarity would be possible from the stationary environment in this case given researchers know (1) the full cue system of an organism, (2) how it relates to fundamental tracking, and (3) how both that cue system and the underlying fundamental model shift with non-stationarity.

In recent years, plasticity theory has developed to provide insights on non-stationarity (or ‘sustained environmental change,’ see Chevin *et al.*, 2010). Models of the role of plasticity in novel environments provide an important bridge to understanding the outcomes of non-stationarity, generally predicting non-stationarity should favor highly plastic species. This outcome, however, assumes there are no costs related to plasticity (Ghalambor *et al.*, 2007; Tufto, 2015). If there are costs associated with plasticity (here, directly akin to costs associated with tracking), then species may evolve lower tracking, because it should trade-off with other traits (Auld *et al.*, 2010).

1.3 Tracking in multi-species environments

Life history theory that may help predict tracking often ignores other (non-focal) species or abstracts them as an aspect of the environment. However, a multi-species perspective may be critical to understanding environmental tracking (Metcalf *et al.*, 2015). Considering selection in multi-species environments highlights that tracking cannot be considered as a singular trait, but must be evaluated as part of a trait syndrome (or mosaic of traits, Ghalambor *et al.*, 2007) and should ultimately produce communities of species where tracking trades-off with other traits.

As tracking often relates to the timing of a resource pulse, traits related to resource acquisition are likely contenders for a trade-off. Species with traits that make them poor resource com-

petitors may need to track the environment closely to take advantage of transient periods of available resources, but will risk tissue loss to harsh environmental conditions more prevalent early in the season (e.g., frost or snow) and thus may need to be more stress tolerant. In contrast, species with traits that make them superior resource competitors may perform well even if they track environments less closely, because their resource acquisition is not strongly constrained by competitors. Examples include under-canopy species leafing out earlier to gain access to light (Heberling *et al.*, 2019) or species with shallow roots starting growth sooner in an alpine meadow system, while species with deeper roots begin growth later (Zhu *et al.*, 2016). In such cases, tracking is akin to a competition-colonization trade-off (Amarasekare, 2003), where species that track well gain priority access to resources and, thus, may co-exist with superior competitors. Research to date supports this, with several studies linking higher tracking to traits associated with being poor competitors (Dorji *et al.*, 2013; Lasky *et al.*, 2016; Zhu *et al.*, 2016). Further, many studies have found a correlation between higher tracking and ‘early-ness’ each season, which has been linked to resource acquisition traits associated with lower competitive abilities (Wolkovich & Cleland, 2014, see Box ‘Trait trade-offs with tracking’).

Understanding these trade-offs is clearly critical, but understanding the short-term dynamics of a changing environment with plastic species is additionally important and highlights how little ecological theory we have for tracking. While evolutionary theory sometimes predicts the outcome of a new environment, non-stationarity in the climate today means understanding the trajectory to that outcome may be most relevant—and bridges across evolutionary and ecological timescales. Evolutionary models show how plasticity may limit standing variation and thus reduce fitness in novel environments (Ghalambor *et al.*, 2007; Fox *et al.*, 2019). But whether such findings extend to systems transitioning from stationary to non-stationary will likely depend on how non-stationarity affects the rate of adaptation (Chevin *et al.*, 2010), and how ecological shifts reshape the environment. Efforts to model expected outcomes given climate projections and current understanding of plasticity and genetic variation underlying event timing in some organisms provide the empirical start to this (e.g., Fournier-Level *et al.*, 2016), but more eco-evolutionary models that bridge this gap may prove especially useful.

Including tracking in multi-species community assembly models

Predicting how tracking may determine which species are effectively winners and losers with climate change requires integrating non-stationary environments into models of community assembly. Recent advances in coexistence models, sometimes called ‘modern coexistence theory,’ recognize that both mechanisms independent of fluctuations in the environment (e.g., R^* and other classical niche differences) and mechanisms dependent on fluctuations in the environment (relative non-linearity and storage effect) can lead to coexistence (Chesson & Huntly, 1997; Chesson, 2000). These models, which underlie much of current community ecology research (Mayfield & Levine, 2010; Barabas *et al.*, 2018; Ellner *et al.*, 2019), provide a framework to begin to model environmental tracking and non-stationarity.

How the environment is defined in most community models falls into two broad categories. In some models the environment is expressed as variation in parameters related to species. For example, in some lottery models the environment appears, effectively, as variation in birth and death rates. Building a changing environment into such models thus requires knowing how

environmental shifts filter through to species-level parameters (Tuljapurkar *et al.*, 2009). For example, Rudolf (2019) added the temporal environment to competition models by defining interaction strength as dependent on the temporal distance between species. This is somewhat similar to models that include the environment effectively through different levels of asynchrony (e.g., Nakazawa & Doi, 2012; Revilla *et al.*, 2014). In other models, the environment is more specifically defined. Many of these models define the environment as a resource (e.g., many seed germination models that begin with a resource pulse each year), and thus generally model something close to fundamental tracking. Building a changing environment into these models requires knowing how the environment is changing.

Models that explicitly include the environment provide a major opportunity to predict how environmental tracking and non-stationarity determine future communities (see Fig. 3 and Box: ‘Adding tracking and non-stationarity to a common coexistence model’). Yet most current models generally examine the environment from only one of two relevant angles: they represent the environment as used for species’ cues (e.g., many models of plasticity) or they represent the environment as directly affecting fitness (e.g., the storage effect model). Combining these two angles may be especially critical to understanding the costs and benefits of tracking in non-stationary environments.

Layered onto the different angles that different models take on the environment is how species responses to the environment are defined. In general, species responses to the (resource) environment can be broadly grouped into models that explicitly define when species start an event (e.g., spawning or germination) in response to the environment versus those that model the magnitude (e.g., the number of propagules or seeds) of response to the environment. Models that explicitly model when a species starts an event are often focused on situations where order of arrival is critical to predicting coexistence outcomes. For example, models of priority effects through niche pre-emption highlight the advantage tracking may provide when it allows species to be early (and when there is no cost to being too early): early arrivals receive a head-start advantage, by gaining priority access to resources (the environment) they can draw down the resources available to later arrivals (Fukami, 2015). Such models predict the early-arriving species to out-compete other species, unless the order of arrival varies by year or there are trade-offs with other species’ traits (see Fig. 4).

Other models canalize species’ responses to the environment into production and investment. For example, most models of inter-annual competition (much of ‘modern coexistence theory’) fall into this camp. Species produce (via offspring, tissue etc.) differentially depending on the environment each year and outcomes are mediated through density. While these models superficially may seem disconnected from timing, they critically highlight how phenology relates to production and, thus, investment across years. Further, they almost always model the environment as a distribution (see Fig. 4), which provides the opportunity for the environment to alter the competitive environment each year and, thus, structure coexistence.

A model where species vary both when they start an event and how much they produce dependent on the environment would capture the important attributes of tracking—combining head-start advantages from being early with production variation based on the fitness of the environment. To our knowledge, however, most models approach these questions separately, though

models of bet-hedging come closest (Gourbiere & Menu, 2009; Tufto, 2015). A model that explicitly includes the linked decisions of when to time an event and how much offspring/tissue to produce during the event could provide fundamental insights on the relative importance of each aspect of this process. Such a model could be adapted to address multiple questions of environmental tracking, including how these decisions (‘when’ and ‘how much’) may trade-off and which other traits may be most strongly linked to tracking, as well as explicitly modeling the costs and benefits of tracking in stationary systems—a critical precursor to extending it to non-stationary systems.

Extending models to non-stationary systems is crucial to testing how environmental tracking relates to species persistence with climate change, and research has already begun to tackle this non-trivial challenge (Chesson, 2017; Legault & Melbourne, 2019; Rudolf, 2019). Most work to date, however, focuses on conclusions from stationary or non-stationary systems. Thus, the transition between stationary and non-stationary is often ignored, yet we expect it may be most critical. Communities formed in stationary environment periods (or periods with environments lower non-stationarity) are effectively filtered and assembled by that environmental regime and thus produce the baseline of variation and assembly dynamics for a shifting environment. While analytical solutions for systems transitioning from stationary to non-stationary may take time to develop (Chesson, 2017), simulation work can provide an immediate intuition and framework to address this challenge (see Box: Adding tracking and non-stationarity to a common coexistence model).

1.4 Future directions

Growing empirical research highlights that environmental tracking is linked to species performance and, thus, may be critical to understanding the forces that assemble communities and determine species persistence—especially as anthropogenic climate change is reshaping the environment of all species. Ecological theory is clearly primed for understanding how a variable environment can shape the formation and persistence of species and communities, but to understand what advances in theory may be most useful for making predictions in the Anthropocene, we need more focus on understanding the attributes of an environment shaped strongly by humans. In turn, to test theory we need more robust estimates of environmental tracking and how it fits within a mosaic of other correlated traits. To this aim, we review several major areas of research that we believe could most rapidly unite empirical and theoretical research in environmental tracking to advance the field.

How is an organism’s environment changing?

Currently, much research has focused on one major shift in the climate system (earlier growing seasons), but research on multivariate environmental shifts is growing and will be critical to understanding how climate change affects an organism’s whole environment. Ecologists can guide these efforts by identifying environmental shifts that are often linked. For example, warming temperatures may drive earlier seasons and higher evaporative loss of some resources such as water. Researchers can also aim to more consistently and fully characterize the environmental

distributions of their systems that appear to drive species performance and interactions: the environment of the years of study should be clearly reported and compared against long-term and recent climate for each system.

More interdisciplinary research with climate science could also speed a fuller understanding of what shifts are and are not expected with climate change, and what climate variables are inherently correlated. Such correlations make estimating cues and other biological parameters from long-term data especially precarious (Tansey *et al.*, 2017). But these correlations are equally critical in considering how species may view their environment and whether environmental change will couple or uncouple links between proximate cues and fundamental tracking (Bonamour *et al.*, 2019).

Understanding and measuring ‘tracking’

Understanding how the environment is changing represents just one step towards robust measures of environmental tracking. Shifting environmental regimes must then be filtered through species cues. As robust estimates of these cues are currently available for few species, we suggest several major improvements on current methods to help interpret current trends and make comparisons more feasible.

Studies should clarify their definition of tracking, how the environment is defined and how well, or not, the underlying cue system is understood for study species. Currently, many studies examine fundamental and environmental tracking at once (e.g., Visser *et al.*, 2006; Charmantier *et al.*, 2008; Cleland *et al.*, 2012; Yang & Censer, 2020), which is clearly helpful in advancing the field. However, the more researchers can clarify when and how they are addressing fundamental tracking versus environmental tracking, the more easily we can compare results across studies. Next, and relatedly, studies should define their environment: are they considering primarily the abiotic environment or measuring an environment fundamentally shaped by other species? Finally, all researchers working on environmental tracking need to embrace their inner-physiologist, or collaborate with one. For many species, there is often a related species (albeit, sometimes distantly) whose cue system has been studied (e.g., researchers of *Salix* could examine *Populus*), and thus could provide some insight into possible cues. We expect progress will come from a balance between measures of fundamental tracking (measuring both event date variation and fitness), estimating an organism’s system of cues (generally through controlled experiments followed by tests in the field), and measuring environmental tracking—that is the change in an event date from environmental variation that is due to cues. Clear statements of what is and is not known and measured will help.

Embrace the sometimes contradictory pulls of conducting experiments to identify mechanistic cues and the multivariate climate of the real world. Clearly, we need more experiments to identify which specific aspects of the environment different species cue to and how these cues are filtered by their actual environmental regime (as outlined above and see Chmura *et al.*, 2019). Suites of experiments, which build from identifying cues to understanding how they act when correlated, are a major gap for most organisms.

Build a model of your species’ cues and interrogate it. As research progresses in trying to estimate environmental tracking, greater progress will come from fuller and more diverse inter-

rogations of current (and future) models. Define the framework under which you expect your cue system developed or works (e.g., bet-hedging) then test how well it works, and where it fails (see Johansson & Bolmgren, 2019, for an example). Model interrogation can also use realistic environmental regimes to provide field predictions (Wilczek *et al.*, 2010, 2009) or predict future species and communities. One example of this comes from in silica resurrection experiments of model organisms where future environmental regimes included a mix of regular climate projections and projections modified to test and advance understanding of environmental tracking for the study species (e.g., warmer winter and altered photoperiod scenarios in Fournier-Level *et al.*, 2016).

What major traits trade-off with tracking?

Basic theory of plasticity and competition suggest that environmental tracking must trade-off with other traits to allow multi-species communities. Yet to date empirical work has mainly documented tracking, linked it to performance, or focused on how it varies between native and non-native species (Willis *et al.*, 2010; Wolkovich *et al.*, 2013; Zettlemoyer *et al.*, 2019). Such work lays the groundwork that environmental tracking is important, but future empirical research should address how this trait co-occurs with other traits. Research has highlighted some traits that co-vary with tracking (e.g., Kharouba *et al.*, 2014; Lasky *et al.*, 2016; Zhu *et al.*, 2016), but to tie this empirical work to models requires more research on traits that link clearly to theory, and a fuller understanding of how tracking and other traits jointly contribute to performance under varying environments.

Traits that link to resource competition, as we focus on here (as others have as well, see Rudolf, 2019), may be especially fruitful for greater research, but should not be the only ones considered. For example, traits related to predator tolerance or avoidance may also play a role, but have been effectively unstudied. As empirical research in this area grows, models can aid progress in understanding the outcomes of these trade-offs for community assembly.

Developing new models for tracking in stationary to non-stationary systems

As outlined above many areas of ecological and evolutionary modeling contribute to our understanding of environmental tracking. But most are limited in various ways. Community ecology models generally bifurcate in modeling differences in timing versus production amounts across species, thus studies of whether these models lead to similar or different conclusions would help predict community outcomes and advance our understanding of trade-offs. As outlined above, understanding tracking likely requires models that combine effects. This includes models that combine effects of variation in timing and production amounts and models that include environment as impacting species' cues, as well as species' fitness. Such models would explicitly allow the potential costs and benefits of tracking depending on how closely environmental tracking matches fundamental tracking.

New models will also need to examine how relaxing assumptions of closed communities (i.e., without dispersal or evolution) alter predictions. In practice, dispersal of species or individuals with traits that make them better matched to the non-stationary environment would lead to new communities that may persist or be continually re-assembled as long as the environment remains

non-stationary. Indeed, this logic underlies the argument that invasive species may be superior trackers benefiting from how climate change has altered growing seasons (Willis *et al.*, 2010; Wolkovich & Cleland, 2011). Evolution equally could alter predictions by allowing species traits to evolve in step with environmental change. Long-term population (e.g., Colautti *et al.*, 2017) and resurrection studies (Wilczek *et al.*, 2014; Yousey *et al.*, 2018), as well as field experiments (Colautti *et al.*, 2017; Exposito-Alonso *et al.*, 2019), have repeatedly shown species can shift to earlier flowering times, higher thermal tolerances or related genetically-controlled traits that confer higher fitness in warmer climates. Yet these studies also highlight that responses can be lagged (e.g., Wilczek *et al.*, 2014), associated with reduced population viability (e.g., Colautti *et al.*, 2017), or other factors that may constrain adaptive responses.

1.5 Stationarity in the future

While most environments today are climatically non-stationary and have been for decades, the climate will return to a more stationarity form in the future. There are many possible pathways to climatic stabilization, but almost all require first the stabilization of greenhouse gases—the subject of much policy and political debate. Once greenhouse gas emissions stabilize climate will not quickly snap back to a new stationary phase. Instead systems will slowly approach a new climatic stationarity depending on how they are effected by the earth’s multiple thermal reservoirs, and, in turn, how quickly those reservoirs stabilize. The timescale of this approach is generally expected to be on the scale of centuries, but could be much longer in certain oceanic systems (Collins *et al.*, 2013). Thus, ecologists are—and will remain for the foreseeable future—in a research area structured by climatic non-stationarity.

As paleobiologists and evolutionary biologists often point out, climatic nonstationarity is a common part of the earth’s history (Jansson & Dynesius, 2002)—even if stationary periods—be they cold or warm (glacial and interglacial periods), or dry or wet (e.g., megadroughts or pluvials)—are more common. Indeed, while much of this work has examined how species survive for millions of years given large oscillations in climate (Provan & Bennett, 2008), the periods that provide the most dramatic community reshuffling are periods shifting from stationary to non-stationary climate regimes (Vrba, 1980, 1985). Such stories of the past are now fundamentally happening today, and ecology is challenged to understand how transitions between stationary and non-stationary environments are reshaping the species and communities we have today and will in the altered climates of our future.

2 Acknowledgments

We thank I. Breckheimer, D. Buonaiuto, E. Cleland, J. Davies, G. Legault and two anonymous reviewers for helpful comments that improved the manuscript, K. Slimon for help with trade-offs review, and the NSERC Discovery Award (RGPIN-05038 to EMW) and Canada Research Chair in Temporal Ecology (EMW) programs that provided funding.

3 Boxes

3.1 Box: Environmental variability & change

Decades of ecological research highlight how temporally variable environments shape species and their communities at multiple scales (Sale, 1977; Chesson & Huntly, 1997). In seasonal landscapes, the environment limits periods for growth each year (e.g., by temperature or drought); within-year variability in the environment (e.g., daily, hourly or finer resolution temperatures or rainfall amounts) compounds into inter-annual variability that shapes the distribution of the start and end of growing seasons. For long stretches of history this variability has been effectively stationary; that is, the underlying probability distribution that describes the start (or end) of the season (e.g., the date of the last major frost) does not change, even though the date may be dramatically different from one year to the next.

In other time periods, variability has been non-stationary in one or multiple dimensions. For example, climate in the northern hemisphere includes long warming and then cooling periods (i.e., increasing then decreasing means of the probability distribution) at the start of the Holocene, when the earth was coming out of the last glacial maximum. Anthropogenic climate change is a similar non-stationary process, with warming evident around the globe and knock-on effects for other climate metrics, such as heat extremes and the size of precipitation events.

Understanding non-stationarity in ecological systems requires first identifying which aspects of the environment have shifted—and how they have shifted with respect to one another—as the underlying distributions transition from stationary to non-stationary (Fig. 1). For example, with climate change, warming has increased mean temperatures over time, with minimum temperatures generally increasing more than maximum—this results in an underlying distribution for daily temperature where the mean is increasing through time while the within-day variance is decreasing (Stocker *et al.*, 2013; Screen, 2014). Understanding the impacts of climate change further requires recognizing that most systems can be considered stationary or non-stationary depending on the timescale and period of study. Thus, predicting the consequences of current non-stationarity in ecological systems benefits from identifying the type and scale of non-stationarity, relative to long-term trends.

3.2 Box: Challenges in measuring tracking

Increasingly, research has outlined statistical difficulties in measuring tracking, which may underlie many observations of species that do not track. These issues mostly relate to the challenges of observing phenological distributions (Steer *et al.*, 2019; Carter *et al.*, 2018) through anthropogenically defined temporal units (e.g., day or month) in non-stationary systems, low power, and the complexity of climate data. Most of these can be addressed given improved statistical approaches (e.g., Gienapp *et al.*, 2005; Pearse *et al.*, 2017), though such approaches may uncomfortably highlight how uncertain many current estimates are (Brown *et al.*, 2016).

Non-stationarity in units comes in many forms—estimates of mean days shifted per decade (i.e., days/decade, a common metric summarizing observed shifts in phenology over time in long-

term datasets) depend strongly on the climate of the decade(s) studied, which is not consistent in many systems (Ault *et al.*, 2011; McCabe *et al.*, 2012). Estimates based on a relevant climate variable can sometimes ameliorate this problem, but may be equally vulnerable to non-stationarity in units (e.g., Sagarin, 2001). For example, processes that depend on thermal sums reported as days/°C will generally appear to decline with warming, as the thermal sum of an average day has increased in most regions with climate change. Relatedly, estimates of long-term change using simple linear regression are influenced by the climate at the start of the time-series (with greater changes seen from time-series that started in unusually cold decades, such as the 1950s for much of North America). Impacts of start-years for long-term time-series can be muted by applying change-point or hinge models (e.g., Kharouba *et al.*, 2018), while metrics that move away from calendar units (e.g., day) can help address non-stationarity in units.

Low power is widespread in ecology, where even ‘long’ time-series may be far too short for robust analyses (Bolmgren *et al.*, 2013; Kharouba *et al.*, 2018). Authors should be especially cautious if they find only large effects appear significant (e.g., CaraDonna *et al.*, 2014), which is a well-known statistical bias associated with p-values (Loken & Gelman, 2017). Additionally, effect sizes that are higher when climate variability is higher (for example, in temperate habitats temperature is highly variable in the spring and autumn compared to summer) may be more related to variation in statistical power than to biology (periods with higher variation yield greater variation in the predictor variable, and thus higher power). Mixed models can help better leverage understanding by pooling information across species, and often better capture uncertainty (Pearse *et al.*, 2017). We suggest mixed models should be used more widely alongside randomization and/or data-simulation approaches (e.g., Bolmgren *et al.*, 2013; Kharouba *et al.*, 2018) to better estimate and communicate uncertainty in studies. And researchers should identify what results bias may produce. For example, growing evidence suggests a potential fundamental trade-off where early species track and possess a suite of traits related to faster growth and shorter lifespans, while later species track less and possess traits related to slower growth and longer lifespans—these later species may bet-hedge more given their longer investment window. This, however, could equally be an artifact where early species use simpler cues, and, thus, their tracking is measured more accurately given current methods.

Even without statistical issues, translating phenological and climate data into estimates of tracking requires a firm biological understanding of an organism’s cues, critical knowledge that researchers rarely have (Chmura *et al.*, 2019). Currently, ‘tracking’ is often measured simply as the relationship between the dates of the phenological event and a simple abiotic metric, such as mean monthly temperature (with variation in temperature derived from multiple periods of observation or induced through experiments). Simple environmental metrics, however, are almost always proxies for a more complicated underlying physiology where simple cues—such as warm temperatures—can be modified by other cues, such as photoperiod, drought or light spectra (Bagnall, 1993; Stinchcombe *et al.*, 2004). Teasing out these other cues, however, generally requires nuanced approaches to observational data with explicit assumptions (Tansey *et al.*, 2017) or controlled experiments (Wilczek *et al.*, 2009; Caffarra *et al.*, 2011).

Modeling multivariate cues well is inherently difficult (Chuine *et al.*, 2016), especially since one

cue may dominate in many conditions. For example, woody plant leafout responds strongly to warm spring temperatures, but also to cool winter temperatures to prevent leafout in mid-winter warm snaps that occur long before the last frost. Often this cool-temperature effect may be masked by sufficiently cold conditions. With warming from climate change, however, this additional trigger—which appears to vary by site, species and even inter-annual conditions (Dennis, 2003)—may become critical (and potentially lead many phenological models to fail spectacularly in the future as additional cues come into play, see Chuine *et al.*, 2016). Tracking in species with longer generation times may be especially complicated, as species may track low frequency climate signals and make investment choices on far longer timescales than species with shorter lifespans (Morris *et al.*, 2008). In some semi-arid systems, species time growth to pulses of rain, but only when those rain events occur with cooler temperatures that indicate the start of the rainy season, and not a rare summer rainfall event in the middle of months of drought (Wainwright *et al.*, 2012; Wainwright & Cleland, 2013).

3.3 Box: Trait trade-offs with tracking

Research on phenological tracking and traits has increased greatly in recent years, with a major uptick in studies after 2010 (see SI Fig. ??). Most papers examining tracking and other traits across species have focused on plants (20/30), followed by birds and Lepidoptera (both 4/30), plankton and aphids (both 1/30). The most studied trait was how early or late a phenophase occurred (e.g., date of flowering or start of migration for a species, termed ‘earlyness’ by some authors), with earlier species tending to track more (studies included both birds and Lepidoptera, Diamond *et al.*, 2011; Ishioka *et al.*, 2013; Kharouba *et al.*, 2014; Jing *et al.*, 2016; Du *et al.*, 2017). While this is an important link, it is vulnerable to statistical challenges (see Box ‘Statistical challenges in measuring tracking’). Few studies examined whether tracking correlates with resource acquisition traits; those that did generally found species with higher tracking also had traits associated with lower competitive abilities under low resources (e.g., being shallower or lacking a taproot rooted Dorji *et al.*, 2013; Lasky *et al.*, 2016; Zhu *et al.*, 2016). These species were often also early (e.g., Dorji *et al.*, 2013; Zhu *et al.*, 2016), suggesting tracking may relate to a syndrome of traits that allow species to be rapid colonizers each season, but poor competitors for resources. Indeed, previous work has documented that species with earlier phenophases tend to have resource acquisition traits associated with lower competitive abilities (e.g., they tend to be of lower height, have shallower roots, narrower diameter vessels, thinner leaves, and grow faster, reviewed in Wolkovich & Cleland, 2014).

3.4 Box: Adding tracking and non-stationarity to a common coexistence model

To understand the role of environmental tracking by species in variable environments we use a simple model that allows within- and between-year dynamics to contribute to coexistence. As the model is akin to many commonly used seed germination models (Chesson *et al.*, 2004), we follow a similar terminology for ease; however the basic structure of our model could apply to other systems with one dominant (non-renewing) pulse of a limiting resource each season

(e.g., water from rain or snowpack), or a multivariate resource pulse that acts effectively as one resource (e.g., nitrogen and light drawn down together over the season). In this model the environment is included between-years via variable germination, and within-years the environment is explicitly included as a resource pulse at the start of the season. We adjust the biological start time of species (τ_i for species i) to also allow species to respond to the environment dynamically through what we refer to as tracking. Here, tracking effectively moves a species intrinsic start time closer to the environmental start time in that year, resulting in a higher germination fraction (see SI for complete description and equations).

Species can co-occur via equalizing mechanisms, but they require stabilizing mechanisms to coexist. Species that is, cannot coexist given only variation in tracking—coexistence requires variation in another trait axis. As theory and empirical work suggest this trade-off may involve traits related closely to resource competition, we varied species' R^* . With variation in tracking and in R^* species can persist together as long as those species with a temporal niche advantage are also the inferior competitors (Fig. 3). That is, species that can draw resources down to a lower level and are thus the superior within-season resource competitors (lower R^*) can persist with species with that are inferior competitors but have realized biological start times closer to the environmental start time—a finding inline with currently observed empirical trade-offs (see Box ‘Trait trade-offs with tracking’). These trade-offs, however, are all environmentally dependent. They hold only so long as the environment is stationary.

We examined how trade-offs may be transformed by a non-stationary environment, by transitioning a stationary environment—in which two-species communities had persisted for 500 years—to non-stationary, via an earlier start of season (earlier timing of the resource pulse, τ_p , Fig. 3; see SI for more details). By changing a fundamental niche axis (the distribution of the environment, an axis along which these communities were structured), we shifted one major part of the trade-off: the new non-stationary environment favored an earlier start time than the previous stationary environment. This, in turn, reshaped our two-species communities, which depended on this trade-off for persistence.

While the non-stationary environment favored higher trackers (who in turn drove the extinction of species with lower tracking values from many two-species communities) some two-species communities persisted (257 out of 1698 two-species communities persisting after end of stationary, or 15.1%, Fig. 3). These two-species communities persisted because the same fundamental trade-off between biological start time and within-season competitive ability, while narrowed, was not fully lost. Taken together, these simple simulations show how non-stationarity can drive local species extinction and reshape the underlying assembly mechanisms of communities.

Our simulations support growing work that tracking should not be considered alone (Diamond *et al.*, 2011; Dorji *et al.*, 2013; Ishioka *et al.*, 2013; Kharouba *et al.*, 2014; Du *et al.*, 2017), but may be part of a larger trait syndrome. Indeed, this model trivially shows that multi-species communities cannot form given only variation in the temporal niche—a trade-off is required. Our results thus support empirical work showing a trade-off where trackers are also inferior resource competitors (Lasky *et al.*, 2016; Zhu *et al.*, 2016)—we show this must be the case for multi-species persistence; otherwise, the species best matched to the environment would drive the other extinct.

627 Finally, our results highlight that non-stationarity may reshape the balance of equalizing versus
628 stabilizing mechanisms. As environments shifted from stationarity to non-stationarity, species
629 that co-occurred via equalizing mechanisms persisted longer. While the outcome that equalized
630 species will be more similarly affected by environmental shifts is rather obvious, it has several
631 important implications. First, it may make identifying which traits climate change promotes
632 through stabilizing mechanisms more difficult. Second, it suggests climate change—or other
633 factors that cause an environment to shift from stationary to non-stationary—may cause a
634 fundamental shift away from assembly via stabilizing mechanisms.

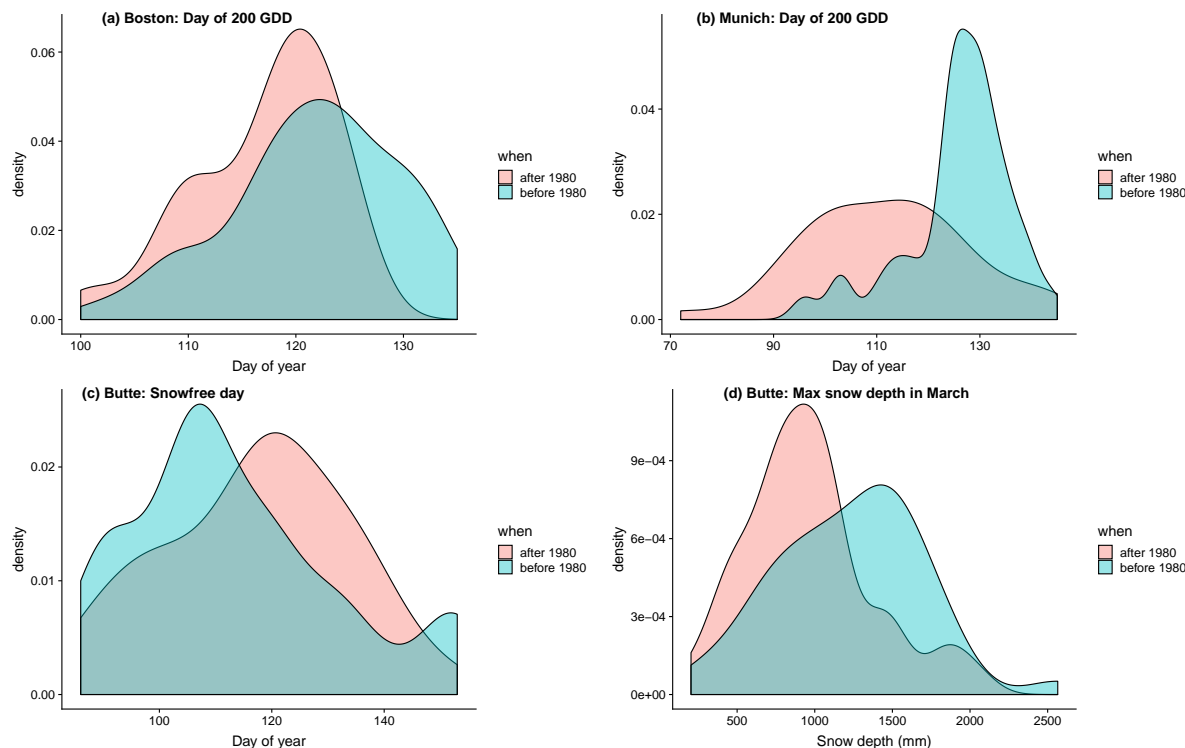


Figure 1: Examples of non-stationarity in climate variables linked to environmental tracking: shifts before and after 1980 (a major change-point in climate for many regions) in several metrics related to the start of growing seasons (a-c) or resource pulse connected to growing season length (d). Density plots of day of 200 growing degree day units (a metric of thermal sum, here based on 0 degree base temperature using daily minima in $^{\circ}\text{C}$) in Boston, MA, USA (a), and Munich, Germany (b), first snowfree day (followed by at least 9 snowfree days) in Crested Butte, CO, USA (c) and maximum snowdepth (mm) in March (often the month before the first snowfree day) in Crested Butte, CO, USA (d). Note that (c) and (d) are likely related, with lower snowpacks leading to an earlier first snowfree day. We selected sites that have been studied for plant phenological data and included at least 80 years of daily climate data from a Global Historical Climatology Network site; we subsetting data so that there were 40 years before and after 1980 for all sites.

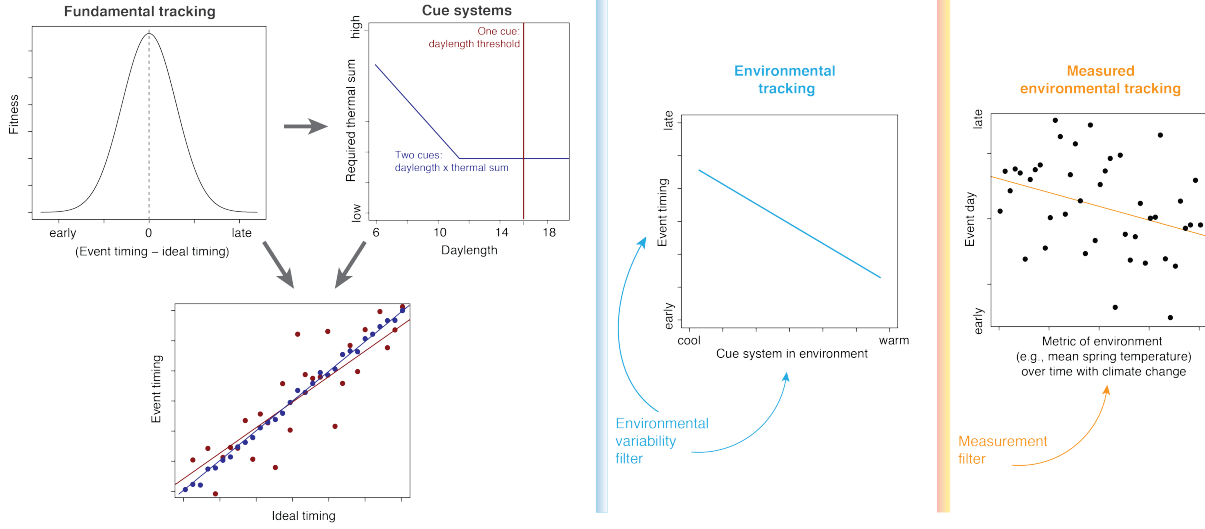


Figure 2: Defining tracking: Fundamental tracking (top left) represents how an organism matches the actual timing of a life history event to the timing that maximizes its fitness (i.e., ideal timing). Here, we show one conceptualization, where fitness declines as event timing moves away from the ideal timing, though realizations in nature may take diverse forms. The cue system is the physiological machinery that an organism uses to predict the ideal timing; we show one dependent only on daylength (red line: the event occurs when the organism’s environment exceeds a certain daylength) and one that depends on a combination of daylength and thermal sums (navy line: the event occurs when the organism accumulates enough temperature for the current daylength). The skill of a cue system in predicting the ideal timing represents cue quality (bottom left). The combination of environmental variability and an organism’s cue system results in different event timings depending on the environment, what we define as environmental tracking (middle). Measured environmental tracking (right) depends both on environmental tracking and the metric(s) of the environment that researchers measure.

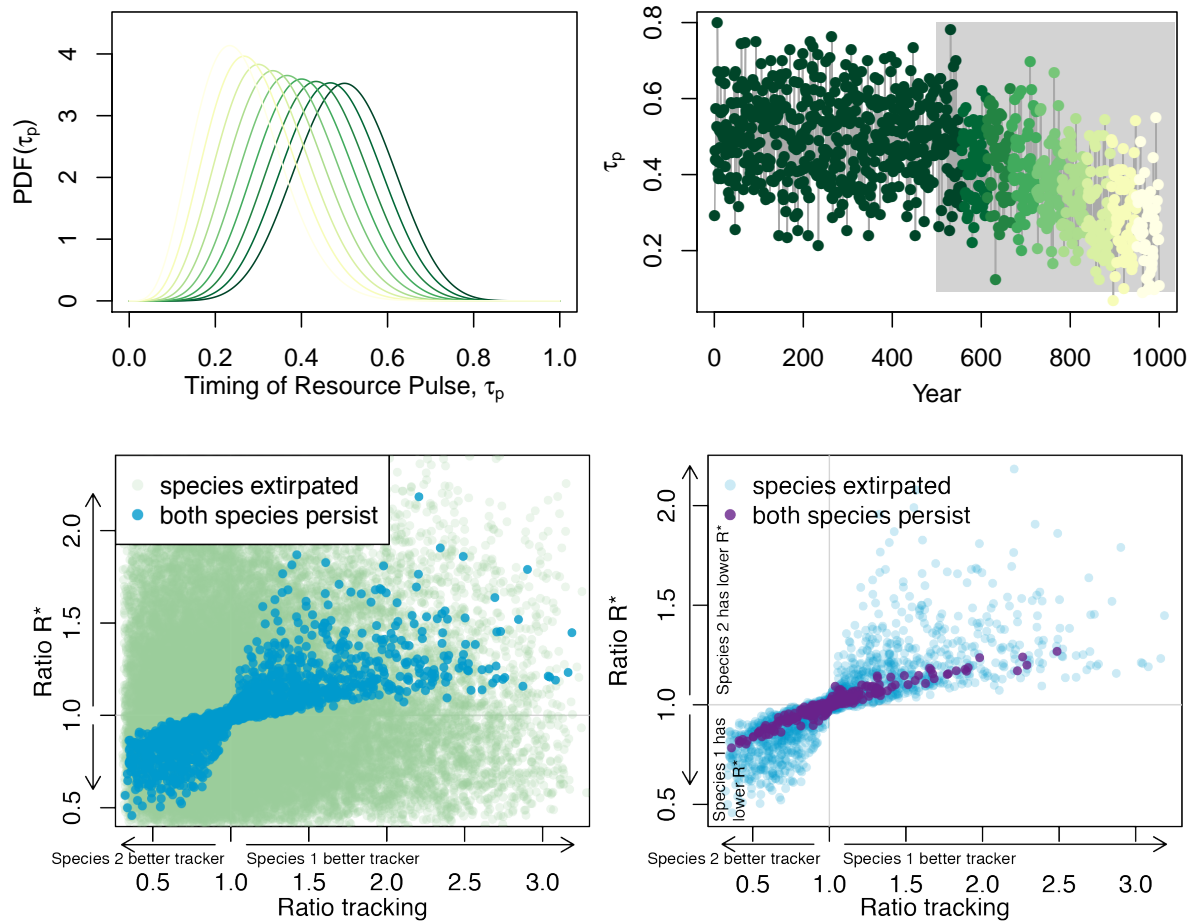


Figure 3: Example of how non-stationarity can reshape communities in a simple model. We shifted the environment (top panels) by changing the timing of the resource pulse from a stationary period ($\tau_p \sim \beta(10, 10)$ for the 500 years) to a nonstationary period ($\tau_p \sim \beta(5, 15)$ over the 500 years), then examined outcomes for two-species communities (bottom panels) where tracking (X axis: species 1/species 2) trades off with R^* (Y axis: species 1/species 2): each point represents one two-species community color-coded by whether both species persisted or one or more species was extirpated through 500 years of a stationary environment (bottom-left), followed by an additional 500 years of non-stationary environment (bottom-right, only two-species communities that persisted through the stationary period are shown).

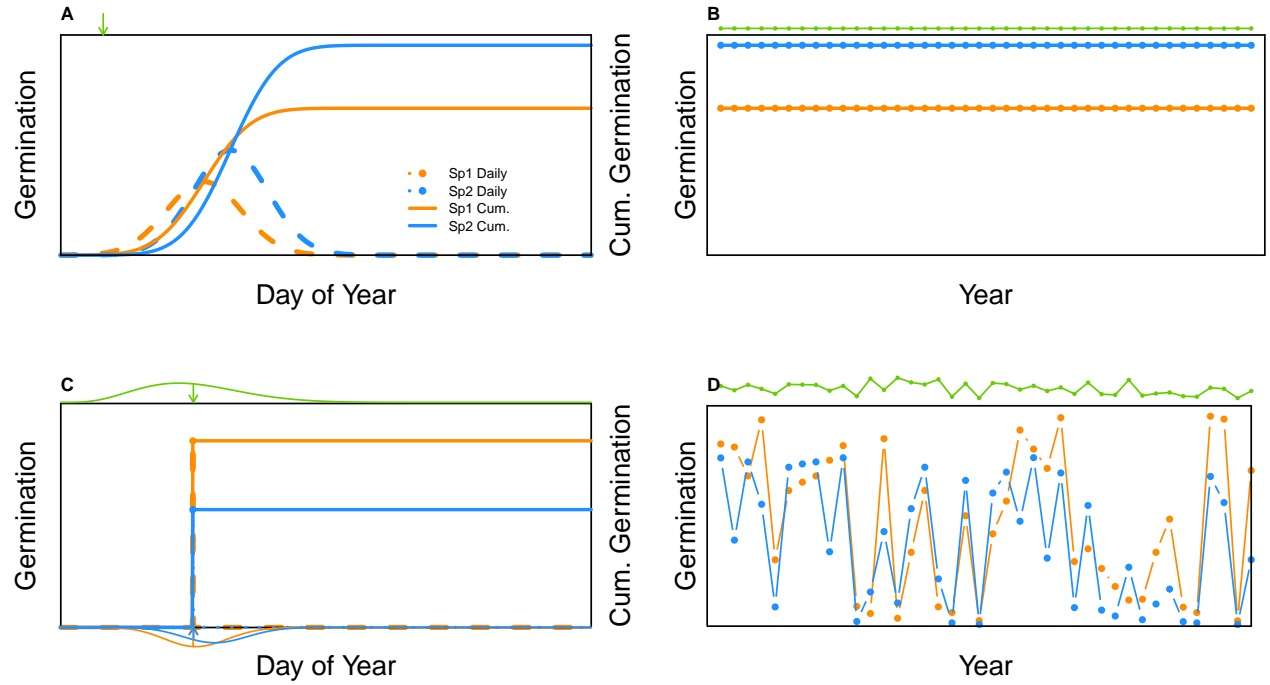


Figure 4: Tracking can be conceptualized as changes in priority effects or changes in storage effects. In a priority effect model (A,B), the coexistence mechanism is a within-year tradeoff between, an early-germinating species that pre-empts resources (sp 1) and late-germinating species that is a superior resource competitor (sp 2) (A, where green arrow indicates the start of season); no between-year variation is required to maintain coexistence. In a storage effect model (C,D), variation in the timing of the start of season (indicated by the distribution in green, top of C) results in differential species-response to the environment (illustrated by species-specific germination curves, bottom of C); this interannual variation in species-response to the environment (D)—along with a seedbank or other interannual storage mechanism—can maintain coexistence through reduced interspecific competition.

References

- Ackerly, D. (2009). Conservatism and diversification of plant functional traits: Evolutionary rates versus phylogenetic signal. *Proceedings of the National Academy of Sciences of the United States of America*, 106, 19699–19706.
- Amano, T., Freckleton, R.P., Queenborough, S.A., Doxford, S.W., Smithers, R.J., Sparks, T.H. & Sutherland, W.J. (2014). Links between plant species’ spatial and temporal responses to a warming climate. *Proceedings of the Royal Society B-Biological Sciences*, 281.
- Amarasekare, P. (2003). Competitive coexistence in spatially structured environments: a synthesis. *Ecology Letters*, 6.
- Auld, J.R., Agrawal, A.A. & Relyea, R.A. (2010). Re-evaluating the costs and limits of adaptive phenotypic plasticity. *Proceedings of the Royal Society B-Biological Sciences*, 277, 503–511.
- Ault, T.R., Macalady, A.K., Pederson, G.T., Betancourt, J.L. & Schwartz, M.D. (2011). Northern hemisphere modes of variability and the timing of spring in western north america. *Journal of Climate*, 24, 4003–4014.
- Bagnall, D.J. (1993). Light quality and vernalization interact in controlling late flowering *Arabidopsis* ecotypes and mutants. *Annals of Botany*, 71, 75–83.
- Barabas, G., D’Andrea, R. & Stump, S.M. (2018). Chesson’s coexistence theory. *Ecological Monographs*, 88, 277–303.
- Block, S., Alexander, J. & Levine, J.M. (2019). Phenological plasticity is a poor predictor of subalpine plant population performance following experimental climate change. *Oikos*, p. 10.1111/oik.06667.
- Bolmgren, K., Vanhoenacker, D. & Miller-Rushing, A.J. (2013). One man, 73 years, and 25 species. evaluating phenological responses using a lifelong study of first flowering dates. *International Journal of Biometeorology*, 57, 367–375.
- Bonamour, S., Chevin, L.M., Charmantier, A. & Teplitsky, C. (2019). Phenotypic plasticity in response to climate change: the importance of cue variation. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 374.
- Brown, C.J., O’Connor, M.I., Poloczanska, E.S., Schoeman, D.S., Buckley, L.B., Burrows, M.T., Duarte, C.M., Halpern, B.S., Pandolfi, J.M., Parmesan, C. & Richardson, A.J. (2016). Ecological and methodological drivers of species’ distribution and phenology responses to climate change. *Global Change Biology*, 22, 1548–1560.
- Caffarra, A., Donnelly, A. & Chuine, I. (2011). Modelling the timing of *Betula pubescens* budburst. II. Integrating complex effects of photoperiod into process-based models. *Climate Research*, 46, 159–170.
- CaraDonna, P.J., Iler, A.M. & Inouye, D.W. (2014). Shifts in flowering phenology reshape a subalpine plant community. *Proceedings of the National Academy of Sciences*, 111, 4916–4921.

- Carter, S.K., Saenz, D. & Rudolf, V.H.W. (2018). Shifts in phenological distributions reshape interaction potential in natural communities. *Ecology Letters*, 21, 1143–1151.
- de Casas, R.R., Donohue, K., Venable, D.L. & Cheptou, P.O. (2015). Gene-flow through space and time: dispersal, dormancy and adaptation to changing environments. *Evolutionary Ecology*, 29, 813–831.
- Charmantier, A., McCleery, R.H., Cole, L.R., Perrins, C., Kruuk, L.E.B. & Sheldon, B.C. (2008). Adaptive phenotypic plasticity in response to climate change in a wild bird population. *Science*, 320, 800–803.
- Chen, I.C., Hill, J.K., Ohlemuller, R., Roy, D.B. & Thomas, C.D. (2011). Rapid range shifts of species associated with high levels of climate warming. *Science*, 333, 1024–1026.
- Chesson, P. (2000). Mechanisms of maintenance of species diversity. *Annual Review of Ecology and Systematics*, 31, 343–366.
- Chesson, P. (2017). Aedt: A new concept for ecological dynamics in the ever-changing world. *Plos Biology*, 15.
- Chesson, P., Gebauer, R.L.E., Schwinning, S., Huntly, N., Wiegand, K., Ernest, M.S.K., Sher, A., Novoplansky, A. & Weltzin, J.F. (2004). Resource pulses, species interactions, and diversity maintenance in arid and semi-arid environments. *Oecologia*, 141, 236–253.
- Chesson, P. & Huntly, N. (1997). The roles of harsh and fluctuating conditions in the dynamics of ecological communities. *American Naturalist*, 150, 519–553.
- Chevin, L.M. & Lande, R. (2015). Evolution of environmental cues for phenotypic plasticity. *Evolution*, 69, 2767–2775.
- Chevin, L.M., Lande, R. & Mace, G.M. (2010). Adaptation, plasticity, and extinction in a changing environment: Towards a predictive theory. *Plos Biology*, 8.
- Chmura, H.E., Kharouba, H.M., Ashander, J., Ehlman, S.M., Rivest, E.B. & Yang, L.H. (2019). The mechanisms of phenology: the patterns and processes of phenological shifts. *Ecological Monographs*, 89.
- Chuine, I., Bonhomme, M., Legave, J.M., de Cortazar-Atauri, I.G., Charrier, G., Lacointe, A. & Ameglio, T. (2016). Can phenological models predict tree phenology accurately in the future? the unrevealed hurdle of endodormancy break. *Global Change Biology*, 22, 3444–3460.
- Chuine, I. & Regniere, J. (2017). Process-based models of phenology for plants and animals. *Annual Review of Ecology, Evolution, and Systematics*, 48, 159–182.
- Cleland, E.E., Allen, J.M., Crimmins, T.M., Dunne, J.A., Pau, S., Travers, S.E., Zavaleta, E.S. & Wolkovich, E.M. (2012). Phenological tracking enables positive species responses to climate change. *Ecology*, 93, 1765–1771.
- Cohen, J.M., Lajeunesse, M.J. & Rohr, J.R. (2018). A global synthesis of animal phenological responses to climate change. *Nature Climate Change*, 8, 224–+.

- Colautti, R.I., Aringgren, J. & Anderson, J.T. (2017). Phenological shifts of native and invasive species under climate change: insights from the boechera - lythrum model. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 372.
- Collins, M., Knutti, R., Arblaster, J.L., Dufresne, T., Fichefet, P., Friedlingstein, X., Gao, W., Gutowski, T., Johns, G., Krinner, M., Shongwe, C., Tebaldi, A., Weaver & Wehner, M. (2013). *Climate Change 2013: The Physical Science Basis – Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, Cambridge University Press, chap. Long-term Climate Change: Projections, Commitments and Irreversibility, pp. 1029–1119.
- Cook, B.I., Wolkovich, E.M. & Parmesan, C. (2012). Divergent responses to spring and winter warming drive community level flowering trends. *Proceedings of the National Academy of Sciences of the United States of America*, 109, 9000–9005.
- Deacy, W.W., Erlenbach, J.A., Leacock, W.B., Stanford, J.A., Robbins, C.T. & Armstrong, J.B. (2018). Phenological tracking associated with increased salmon consumption by brown bears. *Scientific Reports*, 8.
- Dennis, F. (2003). Problems in standardizing methods for evaluating the chilling requirements for the breaking of dormancy in buds of woody plants. *HortScience*, 38, 347–350.
- Diamond, S.E., Frame, A.M., Martin, R.A. & Buckley, L.B. (2011). Species’ traits predict phenological responses to climate change in butterflies. *Ecology*, 92, 1005–1012.
- Donahue, M.J., Karnauskas, M., Toews, C. & Paris, C.B. (2015). Location isn’t everything: Timing of spawning aggregations optimizes larval replenishment. *Plos One*, 10.
- Donaldson-Matasci, M.C., Bergstrom, C.T. & Lachmann, M. (2012). When unreliable cues are good enough. *American Naturalist*, 182, 313–327.
- Dore, A.A., McDowall, L., Rouse, J., Bretman, A., Gage, M.J.G. & Chapman, T. (2018). The role of complex cues in social and reproductive plasticity. *Behavioral Ecology and Sociobiology*, 72.
- Dorji, T., Totland, O., Moe, S.R., Hopping, K.A., Pan, J. & Klein, J.A. (2013). Plant functional traits mediate reproductive phenology and success in response to experimental warming and snow addition in Tibet. *Global Change Biology*, 19, 459–472.
- Du, Y.J., Chen, J.R., Willis, C.G., Zhou, Z.Q., Liu, T., Dai, W.J., Zhao, Y. & Ma, K.P. (2017). Phylogenetic conservatism and trait correlates of spring phenological responses to climate change in northeast china. *Ecology and Evolution*, 7, 6747–6757.
- Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexistence theory for empirical applications. *Ecology Letters*, 22, 3–18.
- Exposito-Alonso, M., Burbano, H.A., Bossdorf, O., Nielsen, R., Weigel, D., Exposito-Alonso, M., Rodriguez, R.G., Barragan, C., Capovilla, G., Chae, E., Devos, J., Dogan, E.S., Friedemann, C., Gross, C., Lang, P., Lundberg, D., Middendorf, V., Kageyama, J., Karasov,

- T., Kersten, S., Petersen, S., Rabbani, L., Regalado, J., Reinelt, L., Rowan, B., Seymour, D.K., Symeonidi, E., Schwab, R., Tran, D.T.N., Venkataramani, K., Van de Weyer, A.L., Vasseur, F., Wang, G., Wedegartner, R., Weiss, F., Wu, R., Xi, W.Y., Zaidem, M., Zhu, W.S., Garcia-Arenal, F., Burbano, H.A., Bossdorf, O., Weigel, D. & Genomes Field Expt, T. (2019). Natural selection on the *Arabidopsis thaliana* genome in present and future climates. *Nature*, 573, 126–+.
- Forsman, A. (2015). Rethinking phenotypic plasticity and its consequences for individuals, populations and species. *Heredity*, 115, 276–284.
- Fournier-Level, A., Perry, E.O., Wang, J.A., Braun, P.T., Migneault, A., Cooper, M.D., Metcalf, C.J.E. & Schmitt, J. (2016). Predicting the evolutionary dynamics of seasonal adaptation to novel climates in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences of the United States of America*, 113, E2812–E2821.
- Fox, R.J., Donelson, J.M., Schunter, C., Ravasi, T. & Gaitan-Espitia, J.D. (2019). Beyond buying time: the role of plasticity in phenotypic adaptation to rapid environmental change. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 374.
- Franks, S.J. & Hoffmann, A.A. (2012). *Genetics of Climate Change Adaptation*, vol. 46 of *Annual Review of Genetics*, pp. 185–208.
- Franks, S.J., Sim, S. & Weis, A.E. (2007). Rapid evolution of flowering time by an annual plant in response to a climate fluctuation. *Proceedings of the National Academy of Sciences of the United States of America*, 104, 1278–1282.
- Fu, Y.S.H., Zhao, H.F., Piao, S.L., Peaucelle, M., Peng, S.S., Zhou, G.Y., Ciais, P., Huang, M.T., Menzel, A., Uelas, J.P., Song, Y., Vitasse, Y., Zeng, Z.Z. & Janssens, I.A. (2015). Declining global warming effects on the phenology of spring leaf unfolding. *Nature*, 526, 104–107.
- Fukami, T. (2015). *Historical Contingency in Community Assembly: Integrating Niches, Species Pools, and Priority Effects*, vol. 46 of *Annual Review of Ecology Evolution and Systematics*, pp. 1–23.
- Ghalambor, C.K., Hoke, K.L., Ruell, E.W., Fischer, E.K., Reznick, D.N. & Hughes, K.A. (2015). Non-adaptive plasticity potentiates rapid adaptive evolution of gene expression in nature. *Nature*, 525, 372–+.
- Ghalambor, C.K., McKay, J.K., Carroll, S.P. & Reznick, D.N. (2007). Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Functional Ecology*, 21, 394–407.
- Gienapp, P., Hemerik, L. & Visser, M.E. (2005). A new statistical tool to predict phenology under climate change scenarios. *Global Change Biology*, 11, 600–606.
- Gotelli, N.J. & Graves, G.R. (1996). The temporal niche. In: *Null Models in Ecology* (eds. Gotelli, N.J. & Graves, G.R.). Smithsonian Institution, Washington, DC, pp. 95–111.

- Gourbiere, S. & Menu, F. (2009). Adaptive dynamics of dormancy duration variability: evolutionary trade-off and priority effect lead to suboptimal adaptation. *Evolution*, 63, 1879–1892.
- Heberling, J.M., Cassidy, S.T., Fridley, J.D. & Kalisz, S. (2019). Carbon gain phenologies of spring-flowering perennials in a deciduous forest indicate a novel niche for a widespread invader. *New Phytologist*, 221, 778–788.
- Hoegh-Guldberg, O., Jacob, D., Taylor, M., Bindi, M., Brown, S., Camilloni, I., Diedhiou, A., Djalante, R., Ebi, K., Engelbrecht, F., Guiot, J., Hijioka, Y., Mehrotra, S., Payne, A., Seneviratne, S., Thomas, A., R. Warren, R. & Zhou, G. (2018). *Impacts of 1.5C Global Warming on Natural and Human Systems*.
- Inouye, B.D., Ehrlen, J. & Underwood, N. (2019). Phenology as a process rather than an event: from individual reaction norms to community metrics. *Ecological Monographs*, 89.
- IPCC (2014). *Climate Change 2014: Impacts, Adaptation, and Vulnerability*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Ishioka, R., Muller, O., Hiura, T. & Kudo, G. (2013). Responses of leafing phenology and photosynthesis to soil warming in forest-floor plants. *Acta Oecologica-International Journal of Ecology*, 51, 34–41.
- Jansson, R. & Dynesius, M. (2002). The fate of clades in a world of recurrent climatic change: Milankovitch oscillations and evolution. *Annual Review of Ecology and Systematics*, 33, 741–777.
- Jing, J., Li, K. & Liu, Z.G. (2016). Effects of varying temperature on leaf phenology and herbivory of dominant tree species in subtropical evergreen broad-leaves forest in eastern china. *Polish Journal of Ecology*, 64, 53–69.
- Johansson, J. & Bolmgren, K. (2019). Is timing of reproduction according to temperature sums an optimal strategy? *Ecology and Evolution*, 9, 11598–11605.
- Johansson, J. & Jonzen, N. (2012). Game theory sheds new light on ecological responses to current climate change when phenology is historically mismatched. *Ecol Lett*, 15, 881–8.
- Kharouba, H.M., Ehrlen, J., Gelman, A., Bolmgren, K., Allen, J.M., Travers, S.E. & Wolkovich, E.M. (2018). Global shifts in the phenological synchrony of species interactions over recent decades. *Proceedings of the National Academy of Sciences of the United States of America*, 115, 5211–5216.
- Kharouba, H.M., Paquette, S.R., Kerr, J.T. & Vellend, M. (2014). Predicting the sensitivity of butterfly phenology to temperature over the past century. *Global Change Biology*, 20, 504–514.
- Lasky, J.R., Uriarte, M. & Muscarella, R. (2016). Synchrony, compensatory dynamics, and the functional trait basis of phenological diversity in a tropical dry forest tree community: effects of rainfall seasonality. *Environmental Research Letters*, 11.

- 818 Legault, G. & Melbourne, B.A. (2019). Accounting for environmental change in continuous-time
819 stochastic population models. *Theoretical Ecology*, 12, 31–48.
- 820 Lenormand, T. (2002). Gene flow and the limits to natural selection. *Trends in Ecology &*
821 *Evolution*, 17, 183–189.
- 822 Loken, E. & Gelman, A. (2017). Measurement error and the replication crisis. *Science*, 355,
823 584–585.
- 824 Mayfield, M.M. & Levine, J.M. (2010). Opposing effects of competitive exclusion on the phy-
825 logenetic structure of communities. *Ecology Letters*, 13, 1085–1093.
- 826 McCabe, G.J., Ault, T.R., Cook, B.I., Betancourt, J.L. & Schwartz, M.D. (2012). Influences of
827 the el nino southern oscillation and the pacific decadal oscillation on the timing of the north
828 american spring. *International Journal of Climatology*, 32, 2301–2310.
- 829 Menzel, A., Von Vopelius, J., Estrella, N., Schleip, C. & Dose, V. (2006). Farmers’ annual
830 activities are not tracking the speed of climate change. *Climate Research*, 32, 201–207.
- 831 Metcalf, C.J.E., Burghardt, L.T. & Koons, D.N. (2015). Avoiding the crowds: the evolution
832 of plastic responses to seasonal cues in a density-dependent world. *Journal of Ecology*, 103,
833 819–828.
- 834 Milly, P.C.D., Betancourt, J., Falkenmark, M., Hirsch, R.M., Kundzewicz, Z.W., Lettenmaier,
835 D.P. & Stouffer, R.J. (2008). Climate change - stationarity is dead: Whither water manage-
836 ment? *Science*, 319, 573–574.
- 837 Morris, W.F., Pfister, C.A., Tuljapurkar, S., Haridas, C.V., Boggs, C.L., Boyce, M.S., Bruna,
838 E.M., Church, D.R., Coulson, T., Doak, D.F., Forsyth, S., Gaillard, J.M., Horvitz, C.C.,
839 Kalisz, S., Kendall, B.E., Knight, T.M., Lee, C.T. & Menges, E.S. (2008). Longevity can
840 buffer plant and animal populations against changing climatic variability. *Ecology*, 89, 19–25.
- 841 Nakazawa, T. & Doi, H. (2012). A perspective on match/mismatch of phenology in community
842 contexts. *Oikos*, 121, 489–495.
- 843 Nicotra, A.B., Atkin, O.K., Bonser, S.P., Davidson, A.M., Finnegan, E.J., Mathesius, U., Poot,
844 P., Purugganan, M.D., Richards, C.L., Valladares, F. & van Kleunen, M. (2010). Plant
845 phenotypic plasticity in a changing climate. *Trends in Plant Science*, 15, 684–692.
- 846 Nosenko, T., Schreiber, F., Adamska, M., Adamski, M., Eitel, M., Hammel, J., Maldonado,
847 M., Mueller, W.E.G., Nickel, M., Schierwater, B., Vacelet, J., Wiens, M. & Woerheide, G.
848 (2013). Deep metazoan phylogeny: When different genes tell different stories. *Molecular*
849 *Phylogenetics and Evolution*, 67, 223–233.
- 850 Parmesan, C. (2006). *Ecological and evolutionary responses to recent climate change*, vol. 37 of
851 *Annual Review of Ecology Evolution and Systematics*, pp. 637–669.
- 852 Parmesan, C. (2007). Influences of species, latitudes and methodologies on estimates of pheno-
853 logical response to global warming. *Global Change Biology*, 13, 1860–1872.

854 Pearse, W.D., Davis, C.C., Inouye, D.W., Primack, R.B. & Davies, T.J. (2017). A statistical
855 estimator for determining the limits of contemporary and historic phenology. *Nature Ecology*
856 *& Evolution*, 1, 1876–+.

857 Piersma, T. & Drent, J. (2003). Phenotypic flexibility and the evolution of organismal design.
858 *Trends in Ecology & Evolution*, 18, 228–233.

859 Provan, J. & Bennett, K.D. (2008). Phylogeographic insights into cryptic glacial refugia. *Trends*
860 *in Ecology & Evolution*, 23, 564–571.

861 Reed, T.E., Grotan, V., Jenouvrier, S., Saether, B.E. & Visser, M.E. (2013). Population growth
862 in a wild bird is buffered against phenological mismatch. *Science*, 340, 488–491.

863 Reed, T.E., Waples, R.S., Schindler, D.E., Hard, J.J. & Kinnison, M.T. (2010). Phenotypic
864 plasticity and population viability: the importance of environmental predictability. *Proceed-*
865 *ings of the Royal Society B-Biological Sciences*, 277, 3391–3400.

866 Revilla, T.A., Encinas-Viso, F. & Loreau, M. (2014). (a bit) earlier or later is always better:
867 Phenological shifts in consumer-resource interactions. *Theoretical Ecology*, 7, 149–162.

868 Richardson, A.D., Bailey, A.S., Denny, E.G., Martin, C.W. & O’Keefe, J. (2006). Phenology
869 of a northern hardwood forest canopy. *Global Change Biology*, 12, 1174–1188.

870 Rudolf, V.H.W. (2019). The role of seasonal timing and phenological shifts for species coexis-
871 tence. *Ecology Letters*.

872 Sagarin, R. (2001). Phenology - false estimates of the advance of spring. *Nature*, 414, 600–600.

873 Sale, P.F. (1977). Maintenance of high diversity in coral reef fish communities. *American*
874 *Naturalist*, 111, 337–359.

875 Screen, J.A. (2014). Arctic amplification decreases temperature variance in northern mid- to
876 high-latitudes. *Nature Climate Change*, 4, 577–582.

877 Simons, A.M. (2011). Modes of response to environmental change and the elusive empirical
878 evidence for bet hedging. *Proceedings of the Royal Society B-Biological Sciences*, 278, 1601–
879 1609.

880 Singer, M.C. & Parmesan, C. (2010). Phenological asynchrony between herbivorous insects
881 and their hosts: signal of climate change or pre-existing adaptive strategy? *Philosophical*
882 *Transactions of the Royal Society B: Biological Sciences*, 365, 3161–3176.

883 Smaldino, P.E. & McElreath, R. (2016). The natural selection of bad science. *Royal Society*
884 *Open Science*, 3.

885 Steer, N.C., Ramsay, P.M. & Franco, M. (2019). nlstimedist: An r package for the biologi-
886 cally meaningful quantification of unimodal phenology distributions. *Methods in Ecology and*
887 *Evolution*, 10, 1934–1940.

- Stinchcombe, J.R., Weinig, C., Ungerer, M., Olsen, K.M., Mays, C., Halldorsdottir, S.S., Purugganan, M.D. & Schmitt, J. (2004). A latitudinal cline in flowering time in *Arabidopsis thaliana* modulated by the flowering time gene FRIGIDA. *Proceedings of the National Academy of Sciences of the United States of America*, 101, 4712–4717.
- Stocker, T., Qin, D. & Plattner, G. (2013). Climate change 2013: The physical science basis. *Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Summary for Policymakers (IPCC, 2013)*.
- Tansey, C.J., Hadfield, J.D. & Phillimore, A.B. (2017). Estimating the ability of plants to plastically track temperature-mediated shifts in the spring phenological optimum. *Global Change Biology*, 23, 3321–3334.
- Thackeray, S.J., Henrys, P.A., Hemming, D., Bell, J.R., Botham, M.S., Burthe, S., Helaouet, P., Johns, D.G., Jones, I.D., Leech, D.I., Mackay, E.B., Massimino, D., Atkinson, S., Bacon, P.J., Brereton, T.M., Carvalho, L., Clutton-Brock, T.H., Duck, C., Edwards, M., Elliott, J.M., Hall, S.J.G., Harrington, R., Pearce-Higgins, J.W., Høye, T.T., Kruuk, L.E.B., Pemberton, J.M., Sparks, T.H., Thompson, P.M., White, I., Winfield, I.J. & Wanless, S. (2016). Phenological sensitivity to climate across taxa and trophic levels. *Nature*, 535, 241.
- Tufto, J. (2015). Genetic evolution, plasticity, and bet-hedging as adaptive responses to temporally autocorrelated fluctuating selection: A quantitative genetic model. *Evolution*, 69, 2034–2049.
- Tuljapurkar, S., Gaillard, J.M. & Coulson, T. (2009). From stochastic environments to life histories and back. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 364, 1499–1509.
- Venable, D.L. (2007). Bet hedging in a guild of desert annuals. *Ecology*, 88, 1086–1090.
- Visser, M.E. & Both, C. (2005). Shifts in phenology due to global climate change: the need for a yardstick. *Proceedings of the Royal Society B-Biological Sciences*, 272, 2561–2569.
- Visser, M.E. & Gienapp, P. (2019). Evolutionary and demographic consequences of phenological mismatches. *Nature Ecology & Evolution*, 3, 879–885.
- Visser, M.E., Holleman, L.J.M. & Gienapp, P. (2006). Shifts in caterpillar biomass phenology due to climate change and its impact on the breeding biology of an insectivorous bird. *Oecologia*, 147, 164–172.
- Vrba, E.S. (1980). Evolution, species and fossils - how does life evolve. *South African Journal of Science*, 76, 61–84.
- Vrba, E.S. (1985). Environment and evolution - alternative causes of the temporal distribution of evolutionary events. *South African Journal of Science*, 81, 229–236.
- Wainwright, C.E. & Cleland, E.E. (2013). Exotic species display greater germination plasticity and higher germination rates than native species across multiple cues. *Biological Invasions*, 15, 2253–2264.

- Wainwright, C.E., Wolkovich, E.M. & Cleland, E.E. (2012). Seasonal priority effects: implications for invasion and restoration in a semi-arid system. *Journal of Applied Ecology*, 49, 234–241.
- Wilczek, A.M., Burghardt, L.T., Cobb, A.R., Cooper, M.D., Welch, S.M. & Schmitt, J. (2010). Genetic and physiological bases for phenological responses to current and predicted climates. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365, 3129–3147.
- Wilczek, A.M., Cooper, M.D., Korves, T.M. & Schmitt, J. (2014). Lagging adaptation to warming climate in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences of the United States of America*, 111, 7906–7913.
- Wilczek, A.M., Roe, J.L., Knapp, M.C., Cooper, M.D., Lopez-Gallego, C., Martin, L.J., Muir, C.D., Sim, S., Walker, A., Anderson, J., Egan, J.F., Moyers, B.T., Petipas, R., Giakountis, A., Charbit, E., Coupland, G., Welch, S.M. & Schmitt, J. (2009). Effects of genetic perturbation on seasonal life history plasticity. *Science*, 323, 930–934.
- Willis, C.G., Ruhfel, B.R., Primack, R.B., Miller-Rushing, A.J., Losos, J.B. & Davis, C.C. (2010). Favorable climate change response explains non-native species' success in Thoreau's woods. *PLoS ONE*, 5, e8878.
- Wolkovich, E.M. & Cleland, E.E. (2011). The phenology of plant invasions: A community ecology perspective. *Frontiers in Ecology and the Environment*, 9, 287–294.
- Wolkovich, E.M. & Cleland, E.E. (2014). Phenological niches and the future of invaded ecosystems with climate change. *AoB Plants*, 6, plu013.
- Wolkovich, E.M., Cook, B.I., Allen, J.M., Crimmins, T.M., Betancourt, J.L., Travers, S.E., Pau, S., Regetz, J., Davies, T.J., Kraft, N.J.B., Ault, T.R., Bolmgren, K., Mazer, S.J., McCabe, G.J., McGill, B.J., Parmesan, C., Salamin, N., Schwartz, M.D. & Cleland, E.E. (2012). Warming experiments underpredict plant phenological responses to climate change. *Nature*, 485, 494–497.
- Wolkovich, E.M., Davies, T.J., Schaefer, H., Cleland, E.E., Cook, B.I., Travers, S.E., Willis, C.G. & Davis, C. (2013). Temperature-dependent shifts in phenology contribute to the success of exotic species with climate change. *American Journal of Botany*, 100, 1407–1421.
- Wong, T.G. & Ackerly, D.D. (2005). Optimal reproductive allocation in annuals and an informational constraint on plasticity. *New Phytologist*, 166, 159–172.
- Yang, L.H. & Czenzer, M.L. (2020). Seasonal windows of opportunity in milkweed-monarch interactions. *Ecology*, 101.
- Yousey, A.M., Chowdhury, P.R., Biddinger, N., Shaw, J.H., Jeyasingh, P.D. & Weider, L.J. (2018). Resurrected 'ancient' daphnia genotypes show reduced thermal stress tolerance compared to modern descendants. *Royal Society Open Science*, 5.
- Zettlemoyer, M.A., Schultheis, E.H. & Lau, J.A. (2019). Phenology in a warming world: differences between native and nonnative plant species. *Ecology Letters*.

962 Zhu, J.T., Zhang, Y.J. & Wang, W.F. (2016). Interactions between warming and soil moisture
963 increase overlap in reproductive phenology among species in an alpine meadow. *Biology*
964 *Letters*, 12, 4.