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How environmental tracking shapes communities in stationary & non-stationary systems

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Abstract

Climate change is reshaping the environments of all species. Predicting how communities will shift in response requires understanding the mechanisms that govern how communities assemble, and how these mechanisms will shift with warming. Growing empirical evidence suggests that environmental tracking—how much an organism can shift the timing of key life history events in response to the environment—is linked to species performance and is a structuring force in communities today. Here, we review current knowledge on temporal environmental tracking both in empirical data and through the lens of community ecology theory. We focus on how climate change has altered the start of the growing season, examine the available evidence that tracking may trade-off with other traits, and provide an initial test of how well basic theory supports the paradigm that climate change should favor environmental tracking. We show how trade-offs that promote coexistence in stationary environments break down in non-stationary environments and may shift the fundamental mechanisms that structure ecological communities. Finally, we consider how the reality that climate change has widespread effects beyond mean temperature, including shifts in growing season length, variability, and in extreme events, may complicate simple predictions of winners and losers.

1 Main text

Anthropogenic climate change is causing widespread changes in species distributions, with many species shifting in both time and space (IPCC, 2014). Reports often focus on species shifting to higher elevations and poleward (Chen et al., 2011) and/or shifting their recurring life history events (phenology) earlier as climate warms (Wolkovich et al., 2012; Cohen et al., 2018). Across species, however, there is high variability. A large proportion of species are not shifting at all (Cook et al., 2012), which has raised concerns about whether these species may be more vulnerable to population declines with continued warming. Such concerns come in part from increasing research that links how well species track climate change—especially through temporal shifts—to shifts in biomass, growth and other metrics related to performance (Cleland et al., 2012). Tracking may then be a major component to understanding and predicting the indirect effects of climate change, including population declines, with cascading effects on community and ecosystem structure.

How well a species tracks the environment through its phenology has repeatedly been linked to other species' responses to climate change. Species that phenologically track warming perform better in field warming experiments (Cleland et al., 2012), and exotic plant species appear to gain a foothold in warming environments by phenologically tracking climate change (Willis et al., 2010). Simple community ecology theory supports these findings, suggesting that a warming climate should open up new temporal niche space and favor species that can exploit that space (Gotelli & Graves, 1996; Wolkovich & Cleland, 2011; Zettlemoyer et al., 2019). Thus, a shift toward earlier spring should favor earlier species, especially those that can environmentally track ever-earlier seasons. This hypothesis has gained significant traction in the ecological literature focused on global change (e.g., Cleland et al., 2012); however, there has been comparatively little work examining whether it is supported through coexistence theory and models.

Current or 'modern' coexistence theory is based strongly on understanding how variable environments may promote coexistence—providing one way to study how communities may be shaped by a temporally varying environment and how tracking may allow a species to take advantage of that variability. Most theory, however, is based on the assumption of stationarity: though the environment is variable, its underlying distribution is unchanged across time (i.e., constant mean and variance, Barabas *et al.*, 2018). This assumption is common not just to coexistence theory, but to much of the theory that underlies ecology, evolution, and myriad other research fields (e.g., Milly *et al.*, 2008; Nosenko *et al.*, 2013).

Climate change upends the assumption of stationarity. By causing increases in temperature, larger pulses of precipitation, increased drought, and more storms (Stocker et al., 2013), climate change has fundamentally shifted major attributes of the environment from stationary to non-stationary regimes. This transition is reshaping ecological systems, and, while new work has aimed to adapt coexistence theory to non-stationary environments (Chesson, 2017), little work has examined what such a transition may mean for communities and the species within them.

Here, we review current knowledge on temporal environmental tracking both in empirical data and through the lens of basic community ecology theory. We begin with a review of environmental variability in stationary and non-stationary environments as well as current coexistence theory for variable environments, then provide an initial test of how well basic theory supports the current paradigm that climate change should favor species with environmental tracking. Finally, we provide a framework using existing ecological theory to understand how tracking in stationary and non-stationary systems may shape communities, and thus help predict the community consequences of climate change.

1.1 Environmental variability & change

Decades of ecological research highlight how temporally variable environments shape species and their communities at multiple scales (Sale, 1977; Chesson & Huntly, 1997). In seasonal land-scapes, the environment limits periods for growth each year (e.g., by temperature or drought); within-year variability in the environment (e.g., daily, hourly or finer resolution temperatures or rainfall amounts) compounds into inter-annual variability that shapes the distribution of the start and end of growing seasons. For long stretches of history this variability has been stationary; that is, the underlying probability distribution that describes the start (or end) of the season (e.g., the date of the last major frost) does not change, even though the date may be dramatically different from one year to the next.

In other time periods, variability has been non-stationary in one or multiple dimensions. For example, climate in the northern hemisphere includes long warming and then cooling periods (i.e., increasing then decreasing means of the probability distribution) at the start of the Holocene, when the earth was coming out of the last glacial maximum. Anthropogenic climate change (henceforth, referred to simply as 'climate change') is a similar non-stationary process, with warming evident around the globe and knock-on effects for other climate metrics, such as heat extremes and the size of precipitation events. While only several decades ago, ecology was focused strongly on stochasticity in stationary systems (e.g., Ripa & Lundberg, 1996; Kaitala et al., 1997), climate change has shifted the focus to understanding stochasticity in a non-stationary framework (e.g., Cazelles et al., 2008; Ehrlen et al., 2016; Legault & Melbourne, 2019).

Understanding non-stationarity in ecological systems requires first identifying which aspects of the environment have shifted—and how they have shifted with respect to one another—as the underlying distributions transition from stationary to non-stationary (Fig. 1). For example, with climate change, warming has increased mean temperatures over time, with minimum temperatures generally increasing more than maximum—this results in an underlying distribution for daily temperature where the mean is increasing through time while the within-day variance is decreasing (Stocker et al., 2013; Screen, 2014).

1.2 Environmental tracking in time

Most species track their environments through time by adjusting their phenologies, but measuring this tracking depends on many factors (see Box 'What underlies variability in species tracking?'). Many (or potentially all) species use abiotic cues to trigger major phenological

events. These cues in turn result in different rates of tracking. At one extreme, some cues yield a fixed timing, resulting in no tracking over time. A common example of a fixed cue is photoperiod, which results in event timing that is constant across years (but variable across space, allowing—for example, later timings poleward for spring events) and appears widespread for some insect emergence and for fall senescence of many trees (Denlinger et al., 2017; Lechowicz, 2002). Fixed timings are perhaps the simplest option and may be efficient for events where there is low predictability, low variability, or low costs to being too late or early. In cases where there is a high cost to mis-timing an event across a variable environment, cues that yield more variability in timing are far more prevalent and usually rely on climate. Temperature is a widespread cue for start of season events with many organisms needing a certain thermal sum to start visible growth. Such a cue has the benefit of shifting the date of an event early or late, depending on climatic conditions, each year, but may be a poor cue in years with aberrant events (e.g., a late frost). In most systems, species must use environmental cues such as temperature to forecast the ideal date for an event—a date which is only obvious in retrospect.

Environmental cues represent the proximate cause of tracking; at the ultimate level, tracking is shaped by resources that species need to grow and reproduce. This is perhaps best recognized in the literature on trophic synchrony where focus is often on how well consumers track their prey resources (Deacy et al., 2018; Kharouba et al., 2018). For example, decades of work has studied how birds (e.g., Parus major) time their peak food demands—during their nesting season—to maximum prey (caterpillar) abundance (e.g., Charmantier et al., 2008). Failure to track prey year-to-year or over time with long-term warming has been well tied to individual-level fitness consequences in some systems (Charmantier et al., 2008), but not all (Visser et al., 2006). Tracking of plants and other lower trophic levels is also about resources. Alpine plant species that emerge in step with snowmelt are likely responding, at least in part, to light resources for photosynthesis. Light equally appears critical to the sequence of phenology in many temperate forests: with lower-canopy species, and younger (shorter) individuals of higher-canopy species, routinely risking frost damage to leaf out before the canopy closes and access to light becomes severely reduced (Vitasse, 2013; Heberling et al., 2019). In both temperate as well as alpine systems, however, access to critical belowground resources also occurs in the spring—both for available water and for nutrients released with the turnover of seasonal microbial communities (Zak et al., 1990; Edwards & Jefferies, 2010). Thus, plants' spring phenology in many systems is about careful tracking to optimally compete for nitrogen and other soil resources. As in higher trophic level systems, research has linked how well plants track to performance, with species that track warming tending to grow larger and/or produce more offspring (Cleland et al., 2012).

1.3 Interspecific variation in tracking

Despite the clear importance of tracking for resource access, not all species appear to track their environments equally well (Thackeray et al., 2016). Many plant species track spring temperatures strongly (multiple meta-analyses now show plants' spring phenology on average track spring or annual temperatures 4-6 days/°C Richardson et al., 2006; Wolkovich et al., 2012; Thackeray et al., 2016, and simple temperature models can often explain over 90% of interannual variation in phenology), but other species do not (Cook et al., 2012) and do not

appear linked to other major climate variables (Thackeray et al., 2016). Variability equally exists when examining consumers tracking their prey (across diverse species tracking over time is 6.1 days/decade but ranges from zero to 15 days/decade, see Kharouba et al., 2018). Such variation in tracking across taxa is driven in part by difficulties in measuring tracking (see Box 'Statistical challenges in measuring tracking'). Yet other variation may be real and suggests perfect environmental tracking may either not be possible or optimal for all species.

Within populations, life-history can help predict how much individuals should track while also balancing trade-offs within and across seasons and years. Tracking has been repeatedly linked to fitness benefits (e.g., Farzan & Yang, 2018; Deacy et al., 2018). Such benefits usually break down into avoiding tissue loss or maximizing growth and, relatedly, maximizing reproduction. For species with bounded growing seasons, much literature has reviewed how tracking is a multivariate equation balancing early-season access to resources and its associated risks of tissue loss, with later season tracking of resources for reproduction and time for offspring to mature (Donohue, 2002; Morin & Chuine, 2005; Burghardt et al., 2015). These trade-offs should also scale up to predictions of variation in tracking across species.

Across species, community ecology theory makes predictions for suites of traits that may tradeoff with tracking. As tracking often relates to the timing of a resource pulse, traits related to resource acquisition are likely contenders for a trade-off. Species with traits that make them poor resource competitors may need to track the environment closely to take advantage of transient periods of available resources, but will risk tissue loss to harsh environmental conditions more prevalent early in the season (e.g., frost or snow). In contrast, species with traits that make them superior resource competitors may perform well even if they track environments less closely, because their resource acquisition is not strongly constrained by competitors. Examples include under-canopy species leafing out earlier to gain access to light (Heberling et al., 2019) or species with shallow roots starting growth sooner in an alpine meadow system, while species with deeper roots begin growth later (Zhu et al., 2016). In such cases, tracking is akin to a competition-colonization trade-off (Amarasekare, 2003), where species that track well gain priority access to resources and, thus, may co-exist with superior competitors. Research to date supports this, with several studies linking higher tracking to traits associated with being poor competitors (Dorji et al., 2013; Lasky et al., 2016; Zhu et al., 2016). Further, many studies have found a correlation between higher tracking and 'earlyness' each season, which has been linked to resource acquisition traits associated with lower competitive abilities (Wolkovich & Cleland, 2014, see Box 'Trait trade-offs with tracking').

These trade-offs with tracking, predicted by basic ecological theory and tentatively supported by growing empirical work, would have fundamental consequences for community assembly, especially with climate change. Applying ecological theory to current environments, however, is difficult because most theory has been developed for stationary systems, which are mathematically more tractable, but can sometimes be extended to non-stationary systems (Chesson, 2017). Almost no community assembly research, however, has examined the consequences of shifting from a stationary to non-stationary environment.

1.4 The role of the environment in coexistence

Recent advances in coexistence models, sometimes called 'modern coexistence theory,' recognize that both mechanisms independent of fluctuations in the environment (e.g., R* and other classical niche differences) and mechanisms dependent on fluctuations in the environment (relative non-linearity and storage effect) can drive coexistence (Chesson & Huntly, 1997; Chesson, 2000). Models under this paradigm are thus often composed of parameters that the describe the environment and the species within it. Parameters related to species must always include mechanisms for growth, death, interactions with other species, and generally a bet-hedging strategy for survival across years (e.g., a seedbank or other long-lived lifestage)—though exactly how these are defined varies across models.

How the environment is defined in most coexistence models falls into two broad categories. In some models the environment is expressed as variation in parameters related to species (e.g., in some lottery models the environment appears, effectively, as variation in birth and death rates). In other models, the environment is more specifically defined. For example, many seed germination models define an environment that begins with a resource pulse each year. Building a changing environment into models thus may require knowing how environmental shifts filter through to species-level parameters (Tuljapurkar et al., 2009) or—perhaps more simply—how the environment is changing.

These models, which underlie much of current community ecology research (Mayfield & Levine, 2010; Barabas et al., 2018; Ellner et al., 2019), allow tests of basic predictions of how tracking may shape communities. In particular, growing empirical research supports that tracking is an important trait—especially in a changing environment. Below we provide an initial test of this prediction and, more generally, how tracking may shape communities. We first consider stationary environments to examine how tracking alters basic models of coexistence; we then test the current paradigm that climate change may favor tracking by examining how communities shift as previously stationary environments become non-stationary.

1.4.1 Model description & simulations

We use a simple model that includes dynamics at both the intra- and inter-annual scales. As the model is akin to many commonly used seed germination models (Chesson *et al.*, 2004), we follow a similar terminology for ease; however the basic structure of our model could apply to other systems with one dominant pulse of a limiting resource each season (e.g., water from rain or snowpack), or a multivariate resource pulse that acts effectively as one resource (e.g., nitrogen and light drawn down together over the season). This model allows within- and between-year dynamics to contribute to coexistence. Between-years the environment is included via variable germination, and within-years the environment is explicitly included as a resource pulse at the start of the season.

The model includes a suite of species traits, including (and particularly relevant for our aims) traits controlling species response to the environment via germination each year and traits related to how species may bet-hedge across years (via a seedbank), as well as traits relating to

resource competition each year. Within-season dynamics are controlled by resource competition resulting in fitness differences, while interannual variation in the environment provides opportunities for coexistence via fluctuation-dependent mechanisms (i.e., niche differences resulting from different germination functions).

Across years, for a community of n species, the seedbank (N) of species i at time t+1 is determined by the survival (s) of seeds that did not germinate in last season $(1 - g_i(t))$ plus new biomass (B_i) produced during the length of the growing season (δ) converted to seeds at rate ϕ :

$$N_i(t+1) = sN_i(t)(1 - g_i(t)) + \phi B_i(t+\delta)$$
(1)

The production of new biomass each season follows a basic R* competition model: new biomass production depends on its resource uptake (f(R)) converted into biomass at rate c_i) less maintenance costs (m), with uptake controlled by a, u, and θ :

$$\frac{\mathrm{d}B_i}{\mathrm{d}t} = c_i f(R) - mB_i \tag{2}$$

$$\frac{\mathrm{d}B_i}{\mathrm{d}t} = c_i f(R) - mB_i \tag{2}$$

$$f(R) = \frac{aR^{\theta}}{1 + auR^{\theta}} \tag{3}$$

With the initial condition:

$$B_i(t+0) = N_i(t)g_i(t)b_0 \tag{4}$$

where b_0 is the initial biomass per seed.

The resource (R) itself declines across a growing season due to uptake by all species and abiotic loss (ϵ) :

$$\frac{\mathrm{d}R}{\mathrm{d}t} = -\sum_{i=1}^{n} f_i(R)B_i - \epsilon R \tag{5}$$

Germination depends both on the traits of the species and on the environment that year. The fraction of seeds germinating for a species each year is determined by the distance between τ_i , a species characteristic, and $\tau_p(t)$, an attribute of the environment, which varies year-to-year. Germination fraction declines according to a Gaussian as the distance between τ_i and $\tau_p(t)$ grows (we refer to this distribution as the 'germination curve').

$$g_i(t) = g_{max}e^{-h(\tau_p(t) - \tau_i)^2}$$
(6)

The model is designed for multiple conceptualizations (Chesson et al., 2004); given our focus here, we consider $\tau_p(t)$ to represent the environmental (abiotic) start of the growing season that varies from year-to-year and refer to it as the 'environmental start time.' τ_i represents the 'intrinsic biological start time' for species i. How well matched a species is to its environment each year can be measured as τ_i - $\tau_p(t)$, or the distance between the intrinsic (biological) start time and the environmental start time.

Adding phenological tracking to model:

Biological start time, τ_i , can be considered a fixed characteristic of a species, but it may also respond to the environment dynamically through what we refer to as environmental tracking. Tracking (α , which can vary between 0 to 1) decreases the distance between τ_i and $\tau_p(t)$, i.e., moving the intrinsic start time closer to the environmental start time in that year, resulting in a higher germination fraction (e.g., species B in Fig. 2b-c).

$$\alpha_i \in [0, 1]$$

$$\hat{\tau}_i = \alpha_i \tau_p(t) + (1 - \alpha_i) \tau_i \tag{7}$$

Thus,

when
$$\alpha_i = 0, \hat{\tau}_i = \tau_i$$

when $\alpha_i = 1, \hat{\tau}_i = \tau_p(t)$

We refer to $\hat{\tau}_i$ as 'effective biological start time' for species i (or 'effective τ_i ').

Simulations:

Using this model framework, we simulated a suite of two-species communities in stationary and non-stationary environments and examined persistence. As our interest is primarily in the role of environmental tracking, we focus on situations where species vary in their match to the environment through both a fixed response (τ_i) or tracking (α) and, thus, examined simulations with only non-trackers and simulations where all species had some level of tracking. We also varied species' resource use efficiency (via c_i), yielding species with different R^* (where a lower R^* means a species can draw the resource down to a lower level and is thus considered the superior competitor). We focused on resource competition as it has been often linked to environmental tracking (see Box 'Trait trade-offs with tracking'). Each simulation was composed of two sequential parts: first, a 500-year stationary period where the underlying distribution of the environment does not change (but is stochastic, yielding year-to-year variation) followed by a 500-year non-stationary period where the underlying distribution of the environment shifts to an earlier start of season. Thus, only species that persisted through the first stationary period continued into the non-stationary period (see Supporting Information, SI, for more details).

1.4.2 Tracking in stationary environments

Species must be sufficiently matched to their environment across years to persist for any long period of time. In our modeling framework, this means species must have a germination curve such that their effective biological start time $(\hat{\tau}_i)$ is sufficiently close to the environmental start time (τ_p) to allow germination of new seeds before the species' seedbank is exhausted. This can happen in effectively two ways: (1) species have fixed intrinsic biological start time values (τ_i) close enough to the environmental start time $(\tau_p; \text{ e.g.}, \text{ species A in Fig. 2b})$ to persist, or

(2) species have a combination of an intrinsic biological start time (τ_i) and tracking (α) that brings the species' effective biological start time $(\hat{\tau}_i)$ close enough to the environmental start time (e.g., see species B in Fig. 2b) to persist.

A simple outcome of this model is that in temporally variable environments where all other species characteristics are identical, the species with the effective biological start time closest to the average environmental start time will always win—regardless of whether this effective biological start is due to a fixed intrinsic start time or due to tracking (or some combination of the two). Put another way, in a stationary environment both tracking and a fixed intrinsic start time are equally useful ways to match to the environment—all that matters is the effective distance between the biological and environmental start of the season. This is because both represent the same niche axis—the temporal niche.

As both a fixed intrinsic start time and tracking represent the same major niche axis, species cannot coexist given only variation in these traits—coexistence requires variation in another trait axis. As discussed above, theory and empirical work suggest this trade-off may involve traits related closely to resource competition. With this added variation—here we varied species' R^* (via c_i)—species can persist together as long as those species with a temporal niche advantage are also the inferior competitors (Fig. 3-4). That is, species that can draw resources down to a lower level and are thus the superior within-season resource competitors (lower R^*) can persist with species with that are inferior competitors but have realized biological start times closer to the environmental start time (regardless of whether that realized biological start time is a result of a fixed trait or tracking)—a finding inline with currently observed empirical trade-offs (see Box 'Trait trade-offs with tracking'). These trade-offs, however, are all environmentally dependent. They hold only so long as the environment is stationary.

1.4.3 Tracking in non-stationary environments

We examined how trade-offs may be transformed by a non-stationary environment, by transitioning a stationary environment—in which two-species communities had persisted for 500 years—to non-stationary, via an earlier start of season (Fig. 2a; see SI for more details). By changing a fundamental niche axis (the distribution of the environment, an axis along which these communities were structured), we shifted one major part of the trade-off: the new non-stationary environment favored an earlier effective biological start time than the previous stationary environment. This, in turn, reshaped our two-species communities, which depended on this trade-off for persistence.

In communities where species traded-off competitive traits (R^*) with non-trackers—species with fixed intrinsic biological start times (τ_i) —species with earlier start times were clearly favored in the non-stationary environment, generally driving the other species (with a lower R^* and later start time) extinct before the end of our 500 year time-period (Fig. 3). Very few two-species communities persisted through the end of the non-stationary period (2 out of 547 two-species communities persisting after end of stationary, or 0.04%); those that persisted did so because the strong similarity between the two species slowed competitive exclusion (i.e., the two species were nearly identical in both R^* and intrinsic biological start time, τ_i). These

species were thus persisting mainly through equalizing mechanisms. In the previous stationary environment species coexisted through both equalizing and stabilizing mechanisms, but the stabilizing mechanisms were lost in the non-stationary environment, as the system shifted away from the region of the temporal niche axis that the communities formed in.

Persistence of two-species communities via both stabilizing and equalizing mechanisms occurred more often in communities where species traded off competitive traits (R^*) with tracking (α) . While the non-stationary environment favored higher trackers (who in turn drove the extinction of species with lower tracking values from many two-species communities) some two-species communities persisted (257 out of 1698 two-species communities persisting after end of stationary, or 15.1%, Fig. 4). These two-species communities persisted because the same fundamental trade-off between biological start time and within-season competitive ability, while narrowed, was not fully lost (Fig. 4). Tracking, in contrast to fixed biological start times, allowed the species with the competitive advantage in effective start time to shift along the temporal niche axis as the environment shifted.

1.4.4 Model conclusions

Taken together, these simple simulations show how non-stationarity can drive local species extinction and reshape the underlying assembly mechanisms of communities. While in stationary systems both tracking and a fixed intrinsic start time can allow a good match to the environment, tracking is superior as environments shift to non-stationarity, confirming the current paradigm that climate change favors species that track the environment. Our simulations, however, also support growing work that tracking should not be considered alone (Diamond et al., 2011; Dorji et al., 2013; Ishioka et al., 2013; Kharouba et al., 2014; Du et al., 2017), but may be part of a larger trait syndrome. Indeed, our models trivially show that multi-species communities cannot form given only variation in fixed intrinsic start times and/or tracking—a trade-off is required. Our results thus also support empirical work showing a trade-off where trackers are also inferior resource competitors (Lasky et al., 2016; Zhu et al., 2016)—we show this must be the case for multi-species persistence; otherwise, the species best matched to the environment would drive the other extinct. Finally, our results highlight that non-stationarity may reshape the balance of equalizing versus stabilizing mechanisms (discussed further below).

Our model, however, makes a number of assumptions about how species respond to non-stationarity in the environment. We imposed environmental non-stationarity on an axis fundamental to coexistence. Yet, non-stationarity in the environment can take on many forms—both in what variable it affects and how it reshapes the underlying distribution of that variable. Communities that assemble via other axes of the environment than start of season timing may be far less impacted than our simulations suggest. Further, we examined a common trend with climate change—shifts in the mean of the environment. Changes can also occur in the variance or the fundamental shape of the distribution (e.g., shifting from a normal distribution to one that is more similar to a Gamma). Additionally, we applied a shift to only one aspect of the environment. In reality, climate change may impose multivariate shifts.

Human modification of the globe imposes complex shifts in the environments of most species.

If the environment shifts along multiple niche axes involved in community assembly, it may allow trade-offs that structure communities to persist through non-stationary periods. We examined this possibility by again shifting the mean start of season earlier (i.e., changing the temporal niche) and, at the same time, decreasing the mean size of the resource pulse by half (i.e., changing the resource niche). Thus, our environment simultaneously favored species with earlier start times and superior within-season competitive abilities (lower R*). We found little evidence, however, of communities persisting via a maintained trade-off—instead the inherent variability of a system shifting in two dimensions drove species extirpations higher (14.8% of two-species communities remaining after non-stationary). Thus, while a multivariate nonstationary environments may, in principle, maintain trade-offs, the shifts in the joint distribution would need to be so balanced that it seems unlikely. More likely appears the possibility that multivariate shifts in the environment make species more vulnerable to local extirpation (Barnosky et al., 2011; IPCC, 2014).

1.4.5 Future research in environmental tracking & non-stationary systems

As we have reviewed, growing empirical research highlights that environmental tracking is linked to species performance and, thus, may be critical to understanding the forces that assemble communities and determine species persistence—especially as anthropogenic climate change is reshaping the environment of all species. Current models of coexistence are clearly primed for understanding how a variable environment can shape the formation and persistence of communities. Moving forward, we need more focus on understanding the attributes of an environment shaped strongly by humans, and, thus, what advances in theory may be most useful for making predictions in the Anthopocene. To this aim, we review several major questions that we believe could most rapidly unite empirical and theoretical research in environmental tracking to advance the field.

How is the environment changing?

Climate change has shifted the environment of all species, often in multivariate ways (Fig. 1). Most systems are seeing increases in mean temperatures, which can rapidly impact the metabolism and activity periods of many species (Monson *et al.*, 2006; IPCC, 2014). This warming is also altering many other attributes of the climate system, including precipitation regimes (Diffenbaugh *et al.*, 2015), and cloud cover (Hofer *et al.*, 2017), which can all further influence species via altering environmental cues.

While we focused on one major shift in the climate system (earlier growing seasons), much more research is needed to understand how multivariate environmental shifts may alter these predictions. Ecologists can guide these efforts by identifying environmental shifts that are often linked (e.g., warming temperatures may drive earlier seasons and higher evaporative loss of some resources such as water). Researchers can also aim to more consistently and fully characterize the environmental distributions of their systems that appear to most drive species performance and interactions: the environment of the years of study should be clearly reported

and compared against long-term and recent climate for each system.

What major traits trade-off with tracking?

Basic theory requires that environmental tracking must trade-off with other traits to allow multi-species communities. Yet to date empirical work has mainly documented tracking, linked it to performance, or focused on how it varies between native and non-native species (Willis et al., 2010; Wolkovich et al., 2013; Zettlemoyer et al., 2019). Such work lays the groundwork that environmental tracking is important, but future empirical research should address how this trait co-occurs with other traits. Research has highlighted some traits that co-vary with tracking (e.g., Kharouba et al., 2014; Lasky et al., 2016; Zhu et al., 2016), but to tie this empirical work to models requires more research on traits that link clearly to theory, and a fuller understanding of how tracking and other traits jointly contribute to performance under varying environments. Traits that link to resource competition, as we focus on here (as others have as well, see Rudolf, 2019), may be especially fruitful for greater research, but should not be the only ones considered. For example, traits related to predator tolerance or avoidance may also play a role, but have been effectively unstudied. As empirical research in this area grows, models can aid progress in understanding the outcomes of these trade-offs for community assembly.

How do shifts to non-stationary environments re-shape the relative influence of stabilizing versus equalizing mechanisms?

Our simple models showed that as environments shift from stationarity to non-stationarity, species that co-occur via equalizing mechanisms can persist longer. While this is a rather obvious outcome—as equalized species will be more similarly affected by environmental shifts—it has several important implications. First, it may make identifying which traits climate change promotes through stabilizing mechanisms more difficult. Second, it suggests climate change—or other factors that cause an environment to shift from stationary to non-stationary—may cause a fundamental shift away from assembly via stabilizing mechanisms. Thus understanding the prevalence of stabilizing versus equalizing mechanisms (which ecology has worked on for many decades, Caswell, 1976; Chesson, 2000) becomes critical for understanding the implications of transitions to non-stationary environments.

If equalizing mechanisms are rare in natural communities then climate change could promote species loss by fundamentally re-shaping stabilizing mechanisms. This finding, however, comes from our modeling approach here, which assumed a closed community without dispersal or evolution. In practice, dispersal of species or individuals with traits that make them better matched to the non-stationary environment (e.g., fixed intrinsic start times that are earlier or with a suite of traits that match to the transformed trade-off axis) would lead to new communities that may persist longer or be continually re-assembled as long as the environment remains non-stationary. Indeed, this logic underlies the argument that invasive species may be superior trackers benefiting from how climate change has altered growing seasons (Willis et al., 2010; Wolkovich & Cleland, 2011). Evolution equally could alter our findings by allowing

species traits to evolve in step with environmental change. Long-term population (e.g., Colautti et al., 2017) and resurrection studies (Wilczek et al., 2014; Yousey et al., 2018), as well as field experiments (Colautti et al., 2017; Exposito-Alonso et al., 2019), have repeatedly shown species can shift to earlier flowering times, higher thermal tolerances or related genetically-controlled traits that confer higher fitness in warmer climates. Yet these studies also highlight that responses can be lagged (e.g., Wilczek et al., 2014), associated with reduced population viability (e.g., Colautti et al., 2017), or other factors that may constrain adaptive responses

1.5 Stationarity in the future

While most environments today are climatically non-stationary and have been for decades, the climate will return to stationarity in the future. There are many possible pathways to climatic stabilization, but almost all require first the stabilization of greenhouse gases—the subject of much policy and political debate. Once greenhouse gas emissions stabilize climate will not quickly snap back to a new stationary phase. Instead systems will slowly approach a new climatic stationarity depending on how they are effected by the earth's multiple thermal reservoirs, and, in turn, how quickly those reservoirs stabilize. The timescale of this approach is generally expected to be on the scale of centuries, but could be much longer in certain oceanic systems (Collins et al., 2013). Thus, ecologists are—and will remain for the forseeable furture—in a research area structured by climatic non-stationarity.

As paleobiologists and evolutionary biologists often point out, climatic nonstationarity is a common part of the earth's history (Jansson & Dynesius, 2002)—even if stationary periods—be they cold or warm (glacial and interglacial periods)—are more common. Indeed, while much of this work has examined how species survive for millions of years given large oscillations in climate (Provan & Bennett, 2008), the periods that provide the most dramatic community reshuffling are periods shifting from stationary to non-stationary climate regimes (Vrba, 1980, 1985). Such stories of the past are now fundamentally happening today, and ecology is challenged to understand how transitions between stationary and non-stationary environments are reshaping the species and communities we have today and will in our warmer future.

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3 Box: What underlies variability in species tracking?

Much recent research in phenological tracking has focused on variability across species (e.g., Willis et al., 2008; Cook et al., 2012; Bolmgren et al., 2013; CaraDonna et al., 2014), with

growing work highlighting that some species do not appear to track climate closely. Indeed, theory predicts some species, for some events, should not track the environment, but identifying non-trackers is difficult in most systems.

We argue three majors classes of reasons underlie species that do not appear to track climate: (1) species do not track, (2) lack of firm biological understanding of the cues that underlie tracking, and (3) statistical artifacts that make it difficult to measure tracking robustly (see Box 'Statistical challenges in measuring tracking'). In some cases, species may be best served to not track; this includes species in highly variable environments or which otherwise face high uncertainty in when to time investment decisions. In such cases, species should gain a substantial benefit from bet-hedging or employing other approaches that spread out risk given uncertainty (Venable, 2007; Donaldson-Matasci et al., 2012). Additionally, evolutionary limitations may prevent tracking: species may not be able to closely measure relevant environmental cues (Arnold, 1992; Singer & Parmesan, 2010), gene flow from other environments may continually push a population away from its local optimum (Lenormand, 2002), or there may be unavoidable trade-offs (Levins, 1968) with tracking. Growing evidence suggests a potential fundamental trade-off where early species track and possess a suite of traits to related to faster growth and shorter lifespans, while later species track less and possess traits related to slower growth and longer lifespans—these later species may bet-hedge more given their longer investment window. This, however, could equally be an artifact where early species use simpler cues, and, thus, their tracking is measured more accurately given current methods.

Accurately measuring environmental tracking depends on the temporal scale of the question (e.g., intra-annual versus inter-annual versus decadal), and how well researchers understand a species' underlying physiology and ecology. Tracking is often measured simply by the relationship between the dates of the phenological event and a simple abiotic metric, such as mean monthly temperature (with variation in temperature derived from multiple periods of observation or induced through experiments). Simple environmental metrics, however, are almost always proxies for a more complicated underlying physiology where simple cues—such as warm temperatures—can be modified by other cues, such as photoperiod, drought or light spectra (Bagnall, 1993; Stinchcombe et al., 2004). Indeed, multiple studies have shown how simple correlations between phenological events and environmental variables can mask complicated relationships (Cook et al., 2012; Tansey et al., 2017). Most well-studied species have multiple cues to time critical biological events (Chuine & Regniere, 2017). These additional cues almost always appear adapted to handle unusual—though not completely uncommon—years when the simple cue alone would fail (that is, would trigger growth, reproduction or another life history event at a suboptimal time).

Modeling this multi-cue complexity well is inherently difficult (Chuine et al., 2016), especially since one cue may dominate in many conditions. For example, woody plant leafout responds strongly to warm spring temperatures, but also to cool winter temperatures to prevent leafout in mid-winter warm snaps that occur long before the last frost. Often this cool-temperature effect may be masked by sufficiently cold conditions. With warming, however, this additional trigger—which appears to vary by site, species and even inter-annual conditions (Dennis, 2003)—may become critical. In some semi-arid systems, species time growth to pulses of rain, but only when

those rain events occur with cooler temperatures that indicate the start of the rainy season, and not a rare summer rainfall event in the middle of months of drought (Wainwright et al., 2012; Wainwright & Cleland, 2013). Thus, despite the apparent efficacy of many current phenological models, many models may fail spectacularly in the future as additional cues come into play (Chuine et al., 2016). Tracking in species with longer generation times may be especially complicated, as species may track low frequency climate signals and make investment choices on far longer timescales than species with shorter lifespans (Morris et al., 2008).

4 Box: Statistical challenges in measuring tracking

Perhaps the most widespread reason for observations of species that do not track is statistical artifacts, including non-stationarity in units and unrecognized low power. All of these can be addressed given improved statistical approaches, though such approaches may (uncomfortably) highlight how uncertain many current estimates are (Brown et al., 2016). Non-stationarity in units comes in many forms—estimates of mean days shifted per decade (i.e., days/decade, a common metric summarizing observed shifts in phenology over time in long-term datasets) depend strongly on the climate of the decade(s) studied, which is not consistent in many systems (Ault et al., 2011; McCabe et al., 2012). Estimates based on a relevant climate variable can sometimes ameliorate this problem, but may be equally vulnerable to non-stationarity in units. For example, processes that depend on thermal sums reported as days/°C will generally appear to decline with warming, as the thermal sum of an average day has increased in most regions with climate change. Relatedly, estimates of long-term change using simple linear regression are influenced by the climate at the start of the time-series (with greater changes seen from timeseries that started in unusually cold decades, such as the 1950s for much of North America). Impacts of start-years for long-term time-series can be muted by applying change-point or hinge models (e.g., Kharouba et al., 2018).

Low power is widespread in ecology, where even 'long' time-series may be far too short for robust analyses (Bolmgren et al., 2013; Kharouba et al., 2018). Authors should be especially cautious if they find only large effects appear significant (e.g., CaraDonna et al., 2014), which is a well-known statistical bias associated with p-values (Loken & Gelman, 2017). Additionally, effect sizes that are higher when climate variability is higher (for example, in temperate habitats temperature is highly variable in the spring and autumn compared to summer) may be more related to variation in statistical power than to biology (periods with higher variation yield greater variation in the predictor variable, and thus higher power). Mixed models can help better leverage understanding by pooling information across species, and often better capture uncertainty (Pearse et al., 2017). We suggest mixed models should be used more widely along-side randomization and/or data-simulation approaches (e.g., Bolmgren et al., 2013; Kharouba et al., 2018) to better estimate and communicate uncertainty in studies.

5 Box: Trait trade-offs with tracking

Research on phenological tracking and traits has increased greatly in recent years, with a major uptick in studies after 2010 (see SI Fig. S1). Most papers examining tracking and other traits across species have focused on plants (20/30), followed by birds and Lepidoptera (both 4/30), plankton and aphids (both 1/30). The most studied trait was how early or late a phenophase occurred (e.g., date of flowering of start of migration for a species, termed 'earlyness' by some authors), with earlier species tending to track more (studies included both birds and Lepidotera, Diamond et al., 2011; Ishioka et al., 2013; Kharouba et al., 2014; Jing et al., 2016; Du et al., 2017). While this is an important link, it is vulnerable to statistical challenges (see Box 'Statistical challenges in measuring tracking'). Few studies examined whether tracking correlates with resource acquisition traits; those that did generally found species with higher tracking also had traits associated with lower competitive abilities under low resources (e.g., being shallower or lacking a taproot rooted Dorji et al., 2013; Lasky et al., 2016; Zhu et al., 2016). These species were often also early (e.g., Dorji et al., 2013; Zhu et al., 2016), suggesting tracking may relate to a syndrome of traits that allow species to be rapid colonizers each season, but poor competitors for resources. Indeed, previous work has documented that species with earlier phenophases tend to have resource acquisition traits associated with lower competitive abilities (e.g., they tend to be of lower height, have shallower roots, narrower diameter vessels, thinner leaves, and grow faster, reviewed in Wolkovich & Cleland, 2014).

References

- Amarasekare, P. (2003). Competitive coexistence in spatially structured environments: a synthesis. *Ecology Letters*, 6.
- Arnold, S.J. (1992). Constraints on phenotypic evolution. American Naturalist, 140, S85–S107.
- Ault, T.R., Macalady, A.K., Pederson, G.T., Betancourt, J.L. & Schwartz, M.D. (2011). Northern hemisphere modes of variability and the timing of spring in western north america. *Journal of Climate*, 24, 4003–4014.
- Bagnall, D.J. (1993). Light quality and vernalization interact in controlling late flowering *Arabidopsis* ecotypes and mutants. *Annals of Botany*, 71, 75–83.
- Barabas, G., D'Andrea, R. & Stump, S.M. (2018). Chesson's coexistence theory. *Ecological Monographs*, 88, 277–303.
- Barnosky, A.D., Matzke, N., Tomiya, S., Wogan, G.O.U., Swartz, B., Quental, T.B., Marshall, C., McGuire, J.L., Lindsey, E.L., Maguire, K.C., Mersey, B. & Ferrer, E.A. (2011). Has the earth's sixth mass extinction already arrived? *Nature*, 471, 51–57.
- Bolmgren, K., Vanhoenacker, D. & Miller-Rushing, A.J. (2013). One man, 73 years, and 25 species. evaluating phenological responses using a lifelong study of first flowering dates. *International Journal of Biometeorology*, 57, 367–375.
- Brown, C.J., O'Connor, M.I., Poloczanska, E.S., Schoeman, D.S., Buckley, L.B., Burrows, M.T., Duarte, C.M., Halpern, B.S., Pandolfi, J.M., Parmesan, C. & Richardson, A.J. (2016). Ecological and methodological drivers of species' distribution and phenology responses to climate change. *Global Change Biology*, 22, 1548–1560.
- Burghardt, L.T., Metcalf, C.J.E., Wilczek, A.M., Schmitt, J. & Donohue, K. (2015). Modeling the influence of genetic and environmental variation on the expression of plant life cycles across landscapes. *American Naturalist*, 185, 212–227.
- CaraDonna, P.J., Iler, A.M. & Inouye, D.W. (2014). Shifts in flowering phenology reshape a subalpine plant community. *Proceedings of the National Academy of Sciences*, 111, 4916–4921.
- Caswell, H. (1976). Community structure neutral model analysis. *Ecological Monographs*, 46, 327–354.
- Cazelles, B., Chavez, M., Berteaux, D., Menard, F., Vik, J.O., Jenouvrier, S. & Stenseth, N.C. (2008). Wavelet analysis of ecological time series. *Oecologia*, 156, 287–304.
- Charmantier, A., McCleery, R.H., Cole, L.R., Perrins, C., Kruuk, L.E.B. & Sheldon, B.C. (2008). Adaptive phenotypic plasticity in response to climate change in a wild bird population. *Science*, 320, 800–803.
- Chen, I.C., Hill, J.K., Ohlemuller, R., Roy, D.B. & Thomas, C.D. (2011). Rapid range shifts of species associated with high levels of climate warming. *Science*, 333, 1024–1026.

- Chesson, P. (2000). Mechanisms of maintenance of species diversity. *Annual Review of Ecology and Systematics*, 31, 343–366.
- Chesson, P. (2017). Aedt: A new concept for ecological dynamics in the ever-changing world. *Plos Biology*, 15.
- Chesson, P., Gebauer, R.L.E., Schwinning, S., Huntly, N., Wiegand, K., Ernest, M.S.K., Sher, A., Novoplansky, A. & Weltzin, J.F. (2004). Resource pulses, species interactions, and diversity maintenance in arid and semi-arid environments. *Oecologia*, 141, 236–253.
- Chesson, P. & Huntly, N. (1997). The roles of harsh and fluctuating conditions in the dynamics of ecological communities. *American Naturalist*, 150, 519–553.
- Chuine, I., Bonhomme, M., Legave, J.M., de Cortazar-Atauri, I.G., Charrier, G., Lacointe, A. & Ameglio, T. (2016). Can phenological models predict tree phenology accurately in the future? the unrevealed hurdle of endodormancy break. *Global Change Biology*, 22, 3444–3460.
- Chuine, I. & Regniere, J. (2017). Process-based models of phenology for plants and animals. *Annual Review of Ecology, Evolution, and Systematics*, 48, 159–182.
- Cleland, E.E., Allen, J.M., Crimmins, T.M., Dunne, J.A., Pau, S., Travers, S.E., Zavaleta, E.S. & Wolkovich, E.M. (2012). Phenological tracking enables positive species responses to climate change. *Ecology*, 93, 1765–1771.
- Cohen, J.M., Lajeunesse, M.J. & Rohr, J.R. (2018). A global synthesis of animal phenological responses to climate change. *Nature Climate Change*, 8, 224–+.
- Colautti, R.I., Aring; gren, J. & Anderson, J.T. (2017). Phenological shifts of native and invasive species under climate change: insights from the boechera lythrum model. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 372.
- Collins, M., Knutti, R., Arblaster, J.L., Dufresne, T., Fichefet, P., Friedlingstein, X., Gao, W., Gutowski, T., Johns, G., Krinner, M., Shongwe, C., Tebaldi, A., Weaver & Wehner, M. (2013). Climate Change 2013: The Physical Science Basis Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge University Press, chap. Long-term Climate Change: Projections, Commitments and Irreversibility, pp. 1029–1119.
- Cook, B.I., Wolkovich, E.M. & Parmesan, C. (2012). Divergent responses to spring and winter warming drive community level flowering trends. *Proceedings of the National Academy of Sciences of the United States of America*, 109, 9000–9005.
- Deacy, W.W., Erlenbach, J.A., Leacock, W.B., Stanford, J.A., Robbins, C.T. & Armstrong, J.B. (2018). Phenological tracking associated with increased salmon consumption by brown bears. *Scientific Reports*, 8.
- Denlinger, D.L., Hahn, D.A., Merlin, C., Holzapfel, C.M. & Bradshaw, W.E. (2017). Keeping time without a spine: what can the insect clock teach us about seasonal adaptation? *Philosophical Transactions of the Royal Society B-Biological Sciences*, 372.

- Dennis, F. (2003). Problems in standardizing methods for evaluating the chilling requirements for the breaking of dormancy in buds of woody plants. *HortScience*, 38, 347–350.
- Diamond, S.E., Frame, A.M., Martin, R.A. & Buckley, L.B. (2011). Species' traits predict phenological responses to climate change in butterflies. *Ecology*, 92, 1005–1012.
- Diffenbaugh, N.S., Swain, D.L. & Touma, D. (2015). Anthropogenic warming has increased drought risk in california. *Proceedings of the National Academy of Sciences of the United States of America*, 112, 3931–3936.
- Donaldson-Matasci, M.C., Bergstrom, C.T. & Lachmann, M. (2012). When unreliable cues are good enough. *American Naturalist*, 182, 313–327.
- Donohue, K. (2002). Germination timing influences natural selection on life-history characters in *Arabidopsis thaliana*. *Ecology*, 83, 1006–1016.
- Dorji, T., Totland, O., Moe, S.R., Hopping, K.A., Pan, J. & Klein, J.A. (2013). Plant functional traits mediate reproductive phenology and success in response to experimental warming and snow addition in Tibet. *Global Change Biology*, 19, 459–472.
- Du, Y.J., Chen, J.R., Willis, C.G., Zhou, Z.Q., Liu, T., Dai, W.J., Zhao, Y. & Ma, K.P. (2017). Phylogenetic conservatism and trait correlates of spring phenological responses to climate change in northeast china. *Ecology and Evolution*, 7, 6747–6757.
- Edwards, K.A. & Jefferies, R.L. (2010). Nitrogen uptake by *Carex aquatilis* during the winterspring transition in a low Arctic wet meadow. *Journal of Ecology*, 98, 737–744.
- Ehrlen, J., Morris, W.F., von Euler, T. & Dahlgren, J.P. (2016). Advancing environmentally explicit structured population models of plants. *Journal of Ecology*, 104, 292–305.
- Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexistence theory for empirical applications. *Ecology Letters*, 22, 3–18.
- Exposito-Alonso, M., Burbano, H.A., Bossdorf, O., Nielsen, R., Weigel, D., Exposito-Alonso, M., Rodriguez, R.G., Barragan, C., Capovilla, G., Chae, E., Devos, J., Dogan, E.S., Friedemann, C., Gross, C., Lang, P., Lundberg, D., Middendorf, V., Kageyama, J., Karasov, T., Kersten, S., Petersen, S., Rabbani, L., Regalado, J., Reinelt, L., Rowan, B., Seymour, D.K., Symeonidi, E., Schwab, R., Tran, D.T.N., Venkataramani, K., Van de Weyer, A.L., Vasseur, F., Wang, G., Wedegartner, R., Weiss, F., Wu, R., Xi, W.Y., Zaidem, M., Zhu, W.S., Garcia-Arenal, F., Burbano, H.A., Bossdorf, O., Weigel, D. & Genomes Field Expt, T. (2019). Natural selection on the arabidopsis thaliana genome in present and future climates. Nature, 573, 126-+.
- Farzan, S. & Yang, L.H. (2018). Experimental shifts in phenology affect fitness, foraging, and parasitism in a native solitary bee. *Ecology*, 99, 2187–2195.
- Gotelli, N.J. & Graves, G.R. (1996). The temporal niche. In: *Null Models in Ecology* (eds. Gotelli, N.J. & Graves, G.R.). Smithsonian Institution, Washington, DC, pp. 95–111.

- Heberling, J.M., Cassidy, S.T., Fridley, J.D. & Kalisz, S. (2019). Carbon gain phenologies of spring-flowering perennials in a deciduous forest indicate a novel niche for a widespread invader. *New Phytologist*, 221, 778–788.
- Hofer, S., Tedstone, A.J., Fettweis, X. & Bamber, J.L. (2017). Decreasing cloud cover drives the recent mass loss on the greenland ice sheet. *Science Advances*, 3.
- IPCC (2014). Climate Change 2014: Impacts, Adaptation, and Vulnerability. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Ishioka, R., Muller, O., Hiura, T. & Kudo, G. (2013). Responses of leafing phenology and photosynthesis to soil warming in forest-floor plants. *Acta Oecologica-International Journal of Ecology*, 51, 34–41.
- Jansson, R. & Dynesius, M. (2002). The fate of clades in a world of recurrent climatic change: Milankovitch oscillations and evolution. Annual Review of Ecology and Systematics, 33, 741–777.
- Jing, J., Li, K. & Liu, Z.G. (2016). Effects of varying temperature on leaf phenology and herbivory of dominant tree species in subtropical evergreen broad-leaves forest in eastern china. *Polish Journal of Ecology*, 64, 53–69.
- Kaitala, V., Ylikarjula, J., Ranta, E. & Lundberg, P. (1997). Population dynamics and the colour of environmental noise. *Proceedings of the Royal Society B-Biological Sciences*, 264, 943–948.
- Kharouba, H.M., Ehrlen, J., Gelman, A., Bolmgren, K., Allen, J.M., Travers, S.E. & Wolkovich, E.M. (2018). Global shifts in the phenological synchrony of species interactions over recent decades. *Proceedings of the National Academy of Sciences of the United States of America*, 115, 5211–5216.
- Kharouba, H.M., Paquette, S.R., Kerr, J.T. & Vellend, M. (2014). Predicting the sensitivity of butterfly phenology to temperature over the past century. *Global Change Biology*, 20, 504–514.
- Lasky, J.R., Uriarte, M. & Muscarella, R. (2016). Synchrony, compensatory dynamics, and the functional trait basis of phenological diversity in a tropical dry forest tree community: effects of rainfall seasonality. *Environmental Research Letters*, 11.
- Lechowicz, M.J. (2002). Phenology, John Wiley Sons, Chichester, vol. 2, pp. 461–465.
- Legault, G. & Melbourne, B.A. (2019). Accounting for environmental change in continuous-time stochastic population models. *Theoretical Ecology*, 12, 31–48.
- Lenormand, T. (2002). Gene flow and the limits to natural selection. *Trends in Ecology Evolution*, 17, 183–189.
- Levins, R. (1968). Evolution in changing environments: some theoretical explorations. Princeton University Press, Princeton, NJ.

- Loken, E. & Gelman, A. (2017). Measurement error and the replication crisis. *Science*, 355, 584–585.
- Mayfield, M.M. & Levine, J.M. (2010). Opposing effects of competitive exclusion on the phylogenetic structure of communities. *Ecology Letters*, 13, 1085–1093.
- McCabe, G.J., Ault, T.R., Cook, B.I., Betancourt, J.L. & Schwartz, M.D. (2012). Influences of the el nino southern oscillation and the pacific decadal oscillation on the timing of the north american spring. *International Journal of Climatology*, 32, 2301–2310.
- Milly, P.C.D., Betancourt, J., Falkenmark, M., Hirsch, R.M., Kundzewicz, Z.W., Lettenmaier, D.P. & Stouffer, R.J. (2008). Climate change stationarity is dead: Whither water management? Science, 319, 573–574.
- Monson, R.K., Lipson, D.L., Burns, S.P., Turnipseed, A.A., Delany, A.C., Williams, M.W. & Schmidt, S.K. (2006). Winter forest soil respiration controlled by climate and microbial community composition. *Nature*, 439, 711–714.
- Morin, X. & Chuine, I. (2005). Sensitivity analysis of the tree distribution model phenofit to climatic input characteristics: implications for climate impact assessment. *Global Change Biology*, 11, 1493–1503.
- Morris, W.F., Pfister, C.A., Tuljapurkar, S., Haridas, C.V., Boggs, C.L., Boyce, M.S., Bruna, E.M., Church, D.R., Coulson, T., Doak, D.F., Forsyth, S., Gaillard, J.M., Horvitz, C.C., Kalisz, S., Kendall, B.E., Knight, T.M., Lee, C.T. & Menges, E.S. (2008). Longevity can buffer plant and animal populations against changing climatic variability. *Ecology*, 89, 19–25.
- Nosenko, T., Schreiber, F., Adamska, M., Adamski, M., Eitel, M., Hammel, J., Maldonado, M., Mueller, W.E.G., Nickel, M., Schierwater, B., Vacelet, J., Wiens, M. & Woerheide, G. (2013). Deep metazoan phylogeny: When different genes tell different stories. *Molecular Phylogenetics and Evolution*, 67, 223–233.
- Pearse, W.D., Davis, C.C., Inouye, D.W., Primack, R.B. & Davies, T.J. (2017). A statistical estimator for determining the limits of contemporary and historic phenology. *Nature Ecology Evolution*, 1, 1876—+.
- Provan, J. & Bennett, K.D. (2008). Phylogeographic insights into cryptic glacial refugia. *Trends in Ecology Evolution*, 23, 564–571.
- Richardson, A.D., Bailey, A.S., Denny, E.G., Martin, C.W. & O'Keefe, J. (2006). Phenology of a northern hardwood forest canopy. *Global Change Biology*, 12, 1174–1188.
- Ripa, J. & Lundberg, P. (1996). Noise colour and the risk of population extinctions. *Proceedings* of the Royal Society B-Biological Sciences, 263, 1751–1753.
- Rudolf, V.H.W. (2019). The role of seasonal timing and phenological shifts for species coexistence. *Ecology Letters*.

- Sale, P.F. (1977). Maintenance of high diversity in coral reef fish communities. *American Naturalist*, 111, 337–359.
- Screen, J.A. (2014). Arctic amplification decreases temperature variance in northern mid- to high-latitudes. *Nature Climate Change*, 4, 577–582.
- Singer, M.C. & Parmesan, C. (2010). Phenological asynchrony between herbivorous insects and their hosts: signal of climate change or pre-existing adaptive strategy? *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365, 3161–3176.
- Stinchcombe, J.R., Weinig, C., Ungerer, M., Olsen, K.M., Mays, C., Halldorsdottir, S.S., Purugganan, M.D. & Schmitt, J. (2004). A latitudinal cline in flowering time in *Arabidopsis thaliana* modulated by the flowering time gene FRIGIDA. *Proceedings of the National Academy of Sciences of the United States of America*, 101, 4712–4717.
- Stocker, T., Qin, D. & Platner, G. (2013). Climate change 2013: The physical science basis. Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Summary for Policymakers (IPCC, 2013).
- Tansey, C.J., Hadfield, J.D. & Phillimore, A.B. (2017). Estimating the ability of plants to plastically track temperature-mediated shifts in the spring phenological optimum. *Global Change Biology*, 23, 3321–3334.
- Thackeray, S.J., Henrys, P.A., Hemming, D., Bell, J.R., Botham, M.S., Burthe, S., Helaouet, P., Johns, D.G., Jones, I.D., Leech, D.I., Mackay, E.B., Massimino, D., Atkinson, S., Bacon, P.J., Brereton, T.M., Carvalho, L., Clutton-Brock, T.H., Duck, C., Edwards, M., Elliott, J.M., Hall, S.J.G., Harrington, R., Pearce-Higgins, J.W., Høye, T.T., Kruuk, L.E.B., Pemberton, J.M., Sparks, T.H., Thompson, P.M., White, I., Winfield, I.J. & Wanless, S. (2016). Phenological sensitivity to climate across taxa and trophic levels. *Nature*, 535, 241.
- Tuljapurkar, S., Gaillard, J.M. & Coulson, T. (2009). From stochastic environments to life histories and back. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 364, 1499–1509.
- Venable, D.L. (2007). Bet hedging in a guild of desert annuals. *Ecology*, 88, 1086–1090.
- Visser, M.E., Holleman, L.J.M. & Gienapp, P. (2006). Shifts in caterpillar biomass phenology due to climate change and its impact on the breeding biology of an insectivorous bird. *Oecologia*, 147, 164–172.
- Vitasse, Y. (2013). Ontogenic changes rather than difference in temperature cause understory trees to leaf out earlier. *New Phytologist*, 198, 149–155.
- Vrba, E.S. (1980). Evolution, species and fossils how does life evolve. South African Journal of Science, 76, 61–84.
- Vrba, E.S. (1985). Environment and evolution alternative causes of the temporal distribution of evolutionary events. South African Journal of Science, 81, 229–236.

- Wainwright, C.E. & Cleland, E.E. (2013). Exotic species display greater germination plasticity and higher germination rates than native species across multiple cues. *Biological Invasions*, 15, 2253–2264.
- Wainwright, C.E., Wolkovich, E.M. & Cleland, E.E. (2012). Seasonal priority effects: implications for invasion and restoration in a semi-arid system. *Journal of Applied Ecology*, 49, 234–241.
- Wilczek, A.M., Cooper, M.D., Korves, T.M. & Schmitt, J. (2014). Lagging adaptation to warming climate in arabidopsis thaliana. *Proceedings of the National Academy of Sciences of the United States of America*, 111, 7906–7913.
- Willis, C.G., Ruhfel, B., Primack, R.B., Miller-Rushing, A.J. & Davis, C.C. (2008). Phylogenetic patterns of species loss in Thoreau's woods are driven by climate change. *Proceedings of the National Academy of Sciences of the United States of America*, 105, 17029–17033.
- Willis, C.G., Ruhfel, B.R., Primack, R.B., Miller-Rushing, A.J., Losos, J.B. & Davis, C.C. (2010). Favorable climate change response explains non-native species' success in Thoreau's woods. PLoS ONE, 5, e8878.
- Wolkovich, E.M. & Cleland, E.E. (2011). The phenology of plant invasions: A community ecology perspective. Frontiers in Ecology and the Environment, 9, 287–294.
- Wolkovich, E.M. & Cleland, E.E. (2014). Phenological niches and the future of invaded ecosystems with climate change. *AoB Plants*, 6, plu013.
- Wolkovich, E.M., Cook, B.I., Allen, J.M., Crimmins, T.M., Betancourt, J.L., Travers, S.E., Pau, S., Regetz, J., Davies, T.J., Kraft, N.J.B., Ault, T.R., Bolmgren, K., Mazer, S.J., McCabe, G.J., McGill, B.J., Parmesan, C., Salamin, N., Schwartz, M.D. & Cleland, E.E. (2012). Warming experiments underpredict plant phenological responses to climate change. Nature, 485, 494–497.
- Wolkovich, E.M., Davies, T.J., Schaefer, H., Cleland, E.E., Cook, B.I., Travers, S.E., Willis, C.G. & Davis, C. (2013). Temperature-dependent shifts in phenology contribute to the success of exotic species with climate change. *American Journal of Botany*, 100, 1407–1421.
- Yousey, A.M., Chowdhury, P.R., Biddinger, N., Shaw, J.H., Jeyasingh, P.D. & Weider, L.J. (2018). Resurrected 'ancient' daphnia genotypes show reduced thermal stress tolerance compared to modern descendants. *Royal Society Open Science*, 5.
- Zak, D.R., Groffman, P.M., Pregitzer, K.S., Christensen, S. & Tiedje, J.M. (1990). The vernal dam - plant microbe competition for nitrogen in northern hardwood forests. *Ecology*, 71, 651–656.
- Zettlemoyer, M.A., Schultheis, E.H. & Lau, J.A. (2019). Phenology in a warming world: differences between native and nonnative plant species. *Ecology Letters*.

Zhu, J.T., Zhang, Y.J. & Wang, W.F. (2016). Interactions between warming and soil moisture increase overlap in reproductive phenology among species in an alpine meadow. *Biology Letters*, 12, 4.

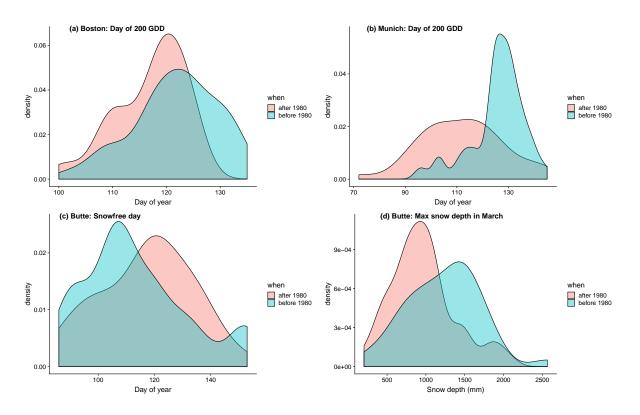
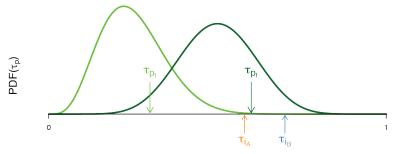
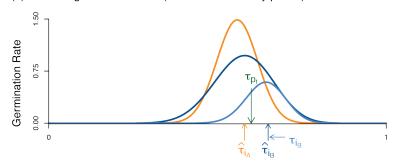


Figure 1: Examples of non-stationarity in climate variables linked to environmental tracking: shifts before and after 1980 (a major change-point in climate for many regions) in several metrics related to the start of growing seasons (a-c) or resource pulse connected to growing season length (d). Density plots of day of 200 growing degree day units (a metric of thermal sum, here based on 0 degree base temperature using daily minima in °C) in Boston, MA, USA (a), and Munich, Germany (b), first snowfree day (followed by at least 9 snowfree days) in Crested Butte, CO, USA (c) and maximum snowdepth (mm) in March (often the month before the first snowfree day) in Crested Butte, CO, USA (d). Note that (c) and (d) are likely related, with lower snowpacks leading to an earlier first snowfree day. We selected sites that have been studied for plant phenological data and included at least 80 years of daily climate data from a Global Historical Climatology Network site (downloaded from https://climexp.knmi.nl/); we subsetted data so that there were 40 years before and after 1980 for all sites.

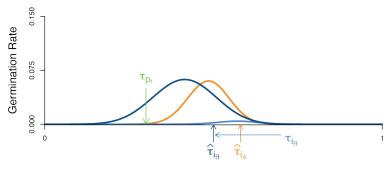
(a) Distributions of resource timing under stationary and non-stationary periods



(b) Realized germination rates (draw from stationary period)



(c) Realized germination rates (draw from end of non-stationary period)



Timing of Resource Pulse, τ_{D}

Figure 2: The distributions of the environment (a) and species' germination for two sample years (b-c) in our seed germination model. (a) The timing of the resource pulse $(\tau_p(t))$, which defines the environmental start of season, is β -distributed with parameters $\beta(10,10)$ during the stationary period (dark green) shifting to $\beta(5,15)$ through the nonstationary period (light green). (b) Realized germination rate as a function of $\tau_p(t)$ for two species during the stationary period: the orange line is a non-tracking species A with preferred germination time, τ_{iA} , that is close to the mean of the stationary period; the blue lines show the difference in realized germination rate of a tracking species with a preferred germination time, τ_{iB} , that is further from the mean of the stationary period both without (light blue) and with (dark blue) the effect of tracking; note the shift from τ_{iB} to τ_{iB} . (c) Realized germination rate of species A and species B at the end of the nonstationary period. Note the change in axes from (b) to (c) shows the decline in overall germination rate as the environment moves away from the preferred germination time of both species.

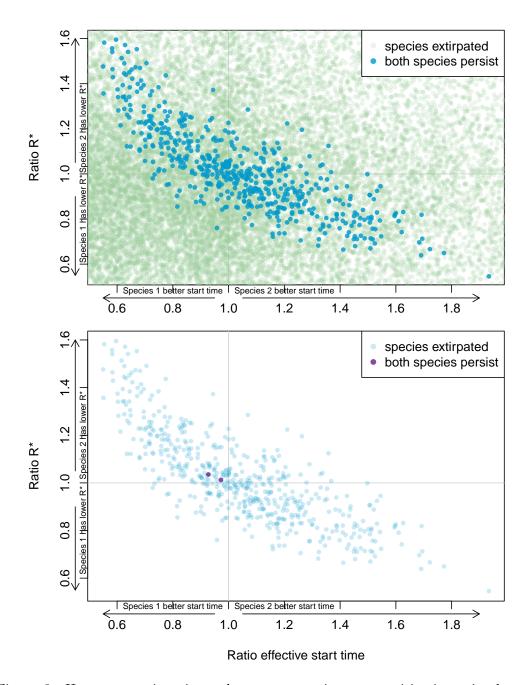


Figure 3: How non-stationarity reshapes two-species communities in a simple model where effective start time (X axis: species $1/\text{species}\ 2$) trades off with R^* (Y axis: species $1/\text{species}\ 2$): each point represents one two-species community color-coded by whether both species persisted or one or more species was extirpated through 500 years of a stationary environment (top), followed by an additional 500 years of non-stationary environment (bottom), where the abiotic start of the season shifts earlier. Only two-species communities that persisted through the stationary period are shown in the bottom panel. See Fig. S3 for an alternative version of this figure detailing one-species outcomes.

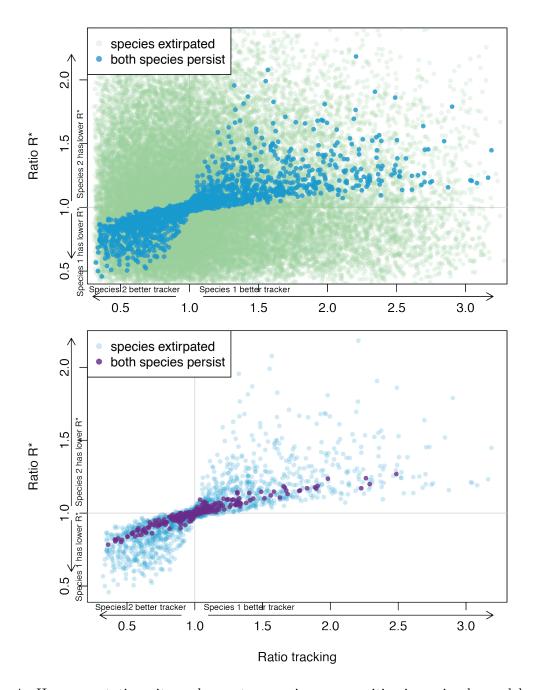


Figure 4: How non-stationarity reshapes two-species communities in a simple model where tracking (X axis: species 1/species 2) trades off with R^* (Y axis: species 1/species 2): each point represents one two-species community color-coded by whether both species persisted or one or more species was extirpated through 500 years of a stationary environment (top), followed by an additional 500 years of non-stationary environment (bottom), where the abiotic start of the season shifts earlier. Only two-species communities that persisted through the stationary period are shown in the bottom panel. See Fig. S4 for an alternative version of this figure detailing one-species outcomes.