



WISER: WhItening and successive least Squares Estimation Refinement for phenotype estimation

Licence, status and metrics

License **GPLv3**

Lifecycle **Stable**

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languages **1**

r **100.0%**

code size **41.1 KiB**

downloads **0**

Stars **2**

Languages and technologies



Overview

The **wiser** package offers *user-friendly* advanced tools for more accurate phenotypic estimation, by adjusting fixed effects (e.g. environmental effects) for the experimental design population structure—or simply called population structure—to eliminate confounding factors between fixed and genetic effects. By employing a whitening transformation followed by successive ordinary least squares (OLS) estimation, **wiser** improves genetic values estimation, called WISER-estimated phenotypes. This approach is particularly beneficial in complex experimental designs where genetic and environmental factors are intricately linked. **wiser** includes methods for computing whitening matrices, fixed effects, residuals, and estimating phenotypes. Additionally, the package provides fast and stable variance component estimation using a parallelized approximate Bayesian computation (ABC) algorithm, specifically designed for large datasets associated with complex experimental designs.

For an experimental design, **wiser** solves the following model, in order to estimate a vector $v = (v_1, \dots, v_q)'$ of q genetic values, which are treated as fixed effects and referred to as the WISER-estimated phenotypes:

$$Y = WX\beta + Zv + \varepsilon = \tilde{X}\beta + \xi$$

where :

- Y ($n \times 1$) is the vector of raw phenotypic measurements, with values possibly repeated for each genotype across one or more environments.

- W ($n \times n$) is a whitening matrix built using omic data (e.g., SNP markers, metabolites, or wavelength reflectances), which adjust the fixed effects variables for the experimental design population structure.
- X ($n \times l$) is the design matrix linking fixed effects to raw phenotypic measurements.
- β ($l \times 1$) is the vector of fixed effects.
- v ($q \times 1$) corresponds to the vector of q WISER-estimated phenotypes.
- Z ($n \times q$) corresponds to the design matrix linking genetic values to raw phenotypic values in the experimental design.
- ε ($n \times 1$) is the vector of residuals.
- $\tilde{X} = XW$ and $\xi = Zv + \varepsilon$.

In the **wiser** framework, the whitening matrix W can be constructed using three different procedures: zero-phase component correlation analysis (**ZCA-cor**), principal component correlation analysis (**PCA-cor**) and cholesky (**Cholesky**). Let $\Sigma_u = \sigma_u^2 ZKZ'$ represent the genetic covariance matrix for the vector $U = Zu = (U_1, U_2, \dots, U_n)$, which corresponds to the vector of individual genetic effects in the experimental design. In **wiser**, the vector $u = (u_1, \dots, u_q)'$, representing the genetic values associated with q genotypes, is assumed to follow the distribution $u \sim \mathcal{N}_q(0, \sigma_u^2 K)$, where K is the genetic covariance matrix between genotypes, estimated from omic data. Define $R_u = V^{-\frac{1}{2}} \Sigma_u V^{-\frac{1}{2}}$ as the correlation matrix associated with Σ_u , where $V = \text{diag}(\Sigma_u)$ is a diagonal matrix with the variances of Σ_u on its diagonal. The matrices Σ_u and R_u , which are symmetric and positive semi-definite, have the following spectral decompositions: $\Sigma_u = U\Lambda U'$ and $R_u = G\Theta G'$, where U and G are orthogonal eigenvectors matrices for Σ_u and R_u , respectively, while Λ and Θ are diagonal matrices of positive eigenvalues for Σ_u and R_u , respectively. The inverse square root matrices of Σ_u and R_u are given by $\Sigma_u^{-\frac{1}{2}} = U\Lambda^{-\frac{1}{2}}U'$ and $R_u^{-\frac{1}{2}} = G\Theta^{-\frac{1}{2}}G'$. These satisfy $\Sigma_u^{-1} = \Sigma_u^{-\frac{1}{2}}\Sigma_u^{-\frac{1}{2}}$ and $R_u^{-1} = R_u^{-\frac{1}{2}}R_u^{-\frac{1}{2}}$. According to Kessy *et al.* (2015), the whitening matrices associated to **ZCA-cor**, **PCA-cor** and **Cholesky** are given by :

$$\begin{cases} W^{\text{ZCA-cor}} = R_u^{-\frac{1}{2}}V^{-\frac{1}{2}} = G\Theta^{-\frac{1}{2}}G'V^{-\frac{1}{2}} = G[\Theta^{-\frac{1}{2}}G'V^{-\frac{1}{2}}] \\ W^{\text{PCA-cor}} = \Theta^{-\frac{1}{2}}G'V^{-\frac{1}{2}} \\ W^{\text{Cholesky}} = L^{-1} \end{cases}$$

where L is derived from the Cholesky decomposition of $\Sigma_u = LL'$. The **PCA-cor** whitening procedure can be seen as standardizing variables using $V^{-\frac{1}{2}}$, followed by a rotation using the transposed correlation eigenmatrix G' , and then scaling using the inverted correlation singular values matrix $\Theta^{-\frac{1}{2}}$. **ZCA-cor** whitening extends this by applying an additional rotation G to revert to the original basis of the standardized variables. Each whitening method is optimal according to specific criteria. For instance, **ZCA-cor** is unique in ensuring that the whitened variables retain the maximum correlation with the original variables. Details of these criteria and the optimality of each method are discussed in Kessy *et al.* (2015).

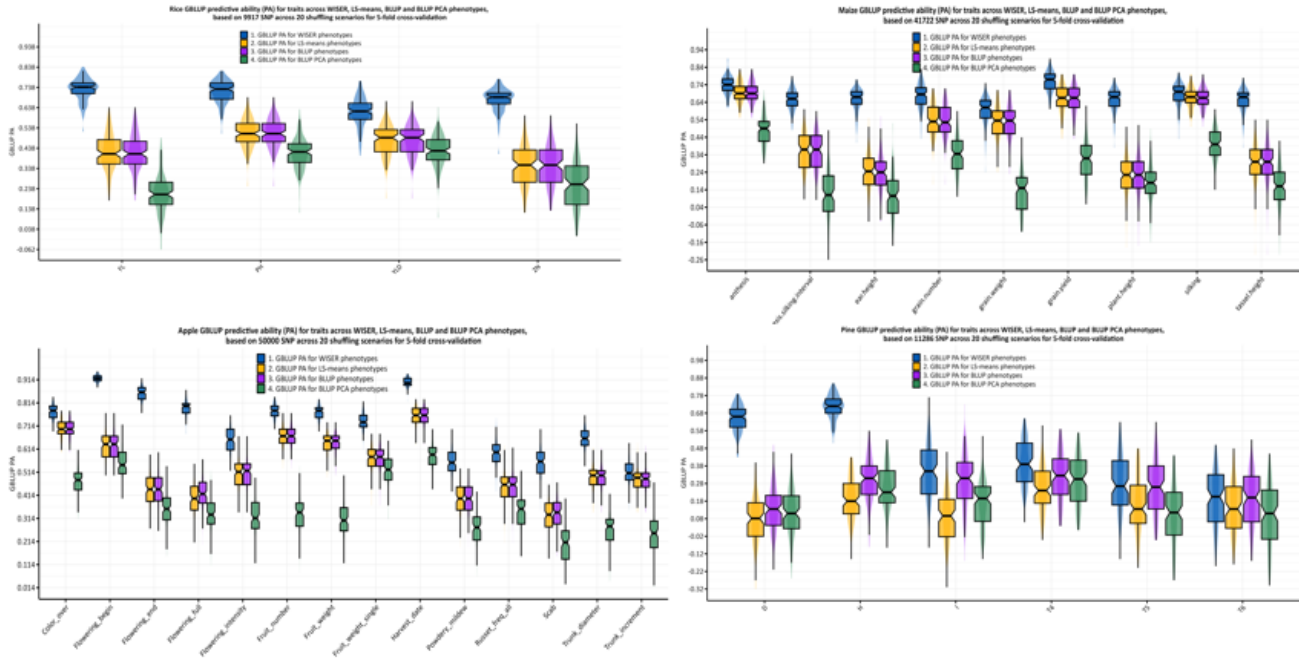
The rationale for transforming X into \tilde{X} through whitening, to adjust fixed effect variables for the experimental design population structure, is comprehensively addressed in Jacquin *et al.* (2025). In contrast, the chosen approach of successive OLS estimation in **wiser** avoids making assumptions about the properties of β and v . Specifically, **wiser** does not assume that v is a random vector drawn from a distribution with a specified covariance matrix. This approach prevents enforcing an unnecessary covariance structure during the estimation of v , which could be detrimental. For example, assuming $v \sim \mathcal{N}_q(0, \sigma_v^2 I_q)$ is often unrealistic and would lead to using a decorrelated covariance structure in the best linear unbiased predictor (BLUP) of v , which can be highly undesirable—particularly in the context of imbalanced genotype frequencies (Holland and Phiepho, 2024). This issue is nearly inevitable, even in carefully balanced experimental designs, due to missing data or incomplete block designs.

Crucially, the successive OLS estimation procedure implemented in **wiser** operates without such assumptions, ensuring that the estimation of v remains independent of omic information or any imposed covariance structure. The only assumptions in the **wiser** framework are $u \sim \mathcal{N}_q(0, \sigma_u^2 K)$ and $\varepsilon \sim \mathcal{N}_n(0, \sigma_e^2 I_n)$, which are necessary to estimate σ_u^2 for constructing the whitening matrix W . In this framework, the estimation of σ_u^2 and σ_e^2 is performed using an ABC algorithm.

In **wiser**, two kernel functions are also provided to build K : **linear** and **identity**. The **identity** kernel, which is **not estimated from omic data**, is generally discouraged due to its poor associated phenotypic predictive ability. The **identity** kernel can be used solely to assess the impact of including or excluding the genetic covariance structure. The **linear** kernel is used by default in **wiser**.

Expected results

The blue, yellow, violet, and green box and violin plots in the figures below represent the distributions of predictive abilities (PA) for phenotypes estimated using WISER, least-squares means (LS-means), BLUP, and BLUP-PCA respectively, as reported by Jacquin *et al.* (2025). These distributions of PA values were computed for 33 traits across four species: rice, maize, apple, and pine. For each trait, the distributions of PA for phenotypes estimated using WISER, LS-means, BLUP, and BLUP-PCA were estimated using a 5-fold cross-validation (CV) scheme with 20 random shufflings of the datasets. As described by Jacquin *et al.* (2025), the average increase in median PA across all species and traits—when using WISER compared to LS-means and BLUP—was **0.21**, highlighting WISER's superior accuracy in phenotypic estimation. Additionally, Jacquin *et al.* (2025) reported a general improvement in genomic heritability estimation when using WISER phenotypes, compared to those estimated with LS-means and BLUP.



Installation

You can install the latest version of the **wiser** package with :

```
install.packages("devtools")
library(devtools)
install_github("ljacquin/wiser")
```

PDF documentation

The PDF version of WISER's documentation can be downloaded [here](#)

Key Features

Phenotype estimation: estimate genetic values which are referred as WISER-estimated phenotypes, by applying whitening methods to adjust for the genetic covariance structure in experimental designs (i.e., population structure correction).

Whitening methods: implement various whitening techniques, such as ZCA-cor, PCA-cor, and Cholesky, to effectively adjust for population structure.

Genetic covariance matrix regularization: ensure the stability of genetic covariance matrices, by applying regularization and shrinkage techniques which ensure positive definiteness.

Optimal whitening and regularization: automatically determine the best whitening method and regularization parameter minimizing mean squared error (MSE), through k-fold cross-validation using a subset of the data.

Fast and stable variance component estimation: use a parallelized ABC algorithm to achieve fast and stable variance component estimation in large datasets associated to complex experimental designs.

Main Functions

`estimate_wiser_phenotype`: computes WISER-estimated phenotypes.
`optimize_whitening_and_regularization`: finds the optimal combination of whitening method and regularization parameter through cross-validation minimizing MSE.

Examples of phenotypic estimation using WISER

Below are straightforward examples demonstrating the use of the `estimate_wiser_phenotype` function to compute WISER-estimated phenotypes, across the following four species: apple, pine, maize and rice. The datasets used are small subsets derived from the original datasets featured in Jacquin *et al.* (2025). These subsets are provided for illustrative purposes within the WISER package and are not intended to serve as reference populations for genomic prediction or GWAS. They include genomic data and raw phenotypic measurements for 30 randomly selected genotypes associated with an experimental design specific to each one of the four species.

Phenotypic estimation for apple data (Jung et al., 2022)

```
# >>> Load wiser library, display package help ,and attach apple data

# Load library
library(wiser)

# Show help for library
help(package = "wiser")

# Attach apple data
data("apple_raw_pheno_data")
data("apple_genomic_data")

# Display apple data
head(apple_raw_pheno_data)
head(apple_genomic_data)[, 1:10]

# For the raw phenotypic data, create the Envir variable in apple_raw_pheno_data by combining
# Country, Year, and Management. This variable represents the environments (i.e., levels)
apple_raw_pheno_data$Envir <- paste0(
  apple_raw_pheno_data$Country, "_",
  apple_raw_pheno_data$Year, "_",
  apple_raw_pheno_data$Management
)

# For the genomic data, assign genotypes as row names (if necessary) to comply with wiser
# functions, and remove the "Genotype" column (if present)
rownames(apple_genomic_data) <- apple_genomic_data$Genotype
apple_genomic_data <- apple_genomic_data[, -match(
  "Genotype",
  colnames(apple_genomic_data)
)]

# Define a trait for phenotypic estimation using wiser (note: some data should be available for
# the trait)
trait_ <- "Trunk_increment"

# >>> Compute WISER-estimated phenotypes using estimate_wiser_phenotype() with its default values for
# whitening_method ("ZCA-cor") and alpha_ (0.01). Remark that the overall mean is always fitted within
# the wiser framework

# NOTE 1: default values for whitening_method and alpha_ typically yield satisfactory results for
# phenotypic predictive ability. Therefore, using `optimize_whitening_and_regularization()` may not
# always be necessary, especially for large datasets. Nevertheless, these parameters should be
# optimized, when possible, for better results.
```

```

# NOTE 2: rows and positions are modeled as factors within the variable Envir to account for local
# spatial heterogeneity. For each environment (i.e., each level of Envir), with multiple rows and
# positions, a row or position level (i.e., row or position number dummy) will capture only a local
# effect. Therefore, Envir is also fitted separately to capture the global environmental effects.

# Estimate "Trunk_increment" (i.e. trait_) phenotypes for genotypes using estimate_wiser_phenotype()
wiser_obj <- estimate_wiser_phenotype(
  omic_df = apple_genomic_data,
  raw_pheno_df = apple_raw_pheno_data,
  trait_ = trait_,
  fixed_effect_vars = c(
    "Envir", "Row", "Position"
  ),
  fixed_effect_vars_computed_as_factor = c(
    "Envir", "Row", "Position"
  ),
  envir_var = "Envir",
  fixed_effect_vars_computed_as_factor_by_envir = c("Row", "Position"),
  random_effects_vars = "Genotype"
)

# >>> Compute WISER-estimated phenotypes using estimate_wiser_phenotype() with optimized values for
# whitening_method and alpha_

# WARNING Highly recommended: increase memory size as specified below with options() before using
# optimize_whitening_and_regularization(). For optimal performance, it is strongly advised to use
# a high-performance computing cluster (HPC) when running this function.
run_example <- F
if (run_example) {
  options(future.globals.maxSize = 16 * 1024^3)

  opt_white_reg_par <- optimize_whitening_and_regularization(
    omic_df = apple_genomic_data,
    raw_pheno_df = apple_raw_pheno_data,
    trait_ = trait_,
    whitening_method_grid = c("ZCA-cor", "Cholesky"),
    k_folds_ = 3,
    alpha_grid = c(0.01, 0.1)
  )
  print(opt_white_reg_par)
  opt_alpha_par_ <- as.numeric(opt_white_reg_par$opt_alpha_)
  opt_white_method_ <- as.character(opt_white_reg_par$opt_whitening_method)

  wiser_obj <- estimate_wiser_phenotype(
    omic_df = apple_genomic_data,
    raw_pheno_df = apple_raw_pheno_data,
    trait_ = trait_,
    fixed_effect_vars = c("Envir", "Row", "Position"),
    fixed_effect_vars_computed_as_factor = c("Envir", "Row", "Position"),
    envir_var = "Envir",
    fixed_effect_vars_computed_as_factor_by_envir = c("Row", "Position"),
    random_effects_vars = "Genotype",
    whitening_method = opt_white_method_,
    alpha_ = opt_alpha_par_
  )
}

# >>> Plot WISER-estimated phenotypes density, display wiser fixed-effect estimates (i.e.
# Envir, etc.), and verify the whitening property

# Plot the density for the estimated phenotypes

```

```

dev.new()
plot(density(wiser_obj$wiser_phenotypes$v_hat), main = paste0(trait_, " v_hat"))

# Print the fixed-effect estimates computed from the whitening process and OLS
print(wiser_obj$wiser_fixed_effect_estimates)

# Print the estimated variance components (from ABC)
print(wiser_obj$wiser_abc_variance_component_estimates)

# Print the fundamental whitening property, i.e.  $In = W \Sigma_u W'$ 
id_mat <- wiser_obj$w_mat %*% wiser_obj$sig_mat_u %*% t(wiser_obj$w_mat)
print(id_mat[1:5, 1:5])

```

Phenotypic estimation for pine data (Perry et al., 2022)

```

# >>> Load wiser library, display package help ,and attach pine data

# Load library
library(wiser)

# Show help for library
help(package = "wiser")

# Attach pine data
data("pine_raw_pheno_data")
data("pine_genomic_data")

# Display pine data
head(pine_raw_pheno_data)
head(pine_genomic_data)[, 1:10]

# Generate latitude and longitude variables per environment (i.e. combination of Site, Year
# and Block in pine_raw_pheno_data)

# NOTE: the Envir variable is not created in pine_raw_pheno_data. Indeed, the two fixed-effect
# variables latitude and longitude, which are quantitative and not considered as factors with levels,
# are highly correlated to each environment for which they are fitted. Therefore, it is not necessary
# to create and fit the Envir variable due to strong redundancy and multicollinearity.
pine_raw_pheno_data <- generate_latitude_longitude_variables_by_environment(
  pine_raw_pheno_data
)

# For the genomic data, assign genotypes as row names (if necessary) to comply with wiser
# functions, and remove the "Genotype" column (if present). Note that "V1" corresponds to
# "Genotype" column here.
rownames(pine_genomic_data) <- pine_genomic_data$V1
pine_genomic_data <- pine_genomic_data[, -match(
  "V1", colnames(pine_genomic_data)
)]

# Define a trait for phenotypic estimation using wiser (note: some data should be available for
# the trait)
trait_ <- "H" # height

# Get fixed-effect variables, latitude and longitude, which are fitted with wiser as quantitative
# variables for each environment (i.e. combination of Site, Year and Block in pine_raw_pheno_data)
fixed_effect_vars_ <- grep("_latitude$|_longitude$", colnames(pine_raw_pheno_data),
  value = TRUE
)

# >>> Compute WISER-estimated phenotypes using estimate_wiser_phenotype() with its default values for

```



```

# whitening_method ("ZCA-cor") and alpha_ (0.01). Remark that the overall mean is always fitted within
# the wiser framework

# Estimate "H" (i.e. trait_) phenotypes for genotypes using estimate_wiser_phenotype()
wiser_obj <- estimate_wiser_phenotype(
  pine_genomic_data,
  pine_raw_pheno_data,
  trait_,
  fixed_effect_vars = fixed_effect_vars_,
  fixed_effect_vars_computed_as_factor = NULL,
  envir_var = NULL,
  fixed_effect_vars_computed_as_factor_by_envir = NULL,
  random_effects_vars = "Genotype"
)

# >>> Plot WISER-estimated phenotypes density, display wiser fixed-effect estimates, and verify
# the whitening property

# Plot the density for the estimated phenotypes
dev.new()
plot(density(wiser_obj$wiser_phenotypes$v_hat), main = paste0(trait_, " v_hat"))

# Print the fixed-effect estimates computed from the whitening process and OLS
print(wiser_obj$wiser_fixed_effect_estimates)

# Print the estimated variance components (from ABC)
print(wiser_obj$wiser_abc_variance_component_estimates)

# Print the fundamental whitening property, i.e.  $I_n = W \Sigma W'$ 
id_mat <- wiser_obj$w_mat %*% wiser_obj$sig_mat_u %*% t(wiser_obj$w_mat)
print(id_mat[1:5, 1:5])

```

Phenotypic estimation for maize data (Millet et al., 2019)

```

# >>> Load wiser library, display package help ,and attach maize data

# Load library
library(wiser)

# Show help for library
help(package = "wiser")

# Attach maize data
data("maize_raw_pheno_data")
data("maize_genomic_data")

# Display maize data
head(maize_raw_pheno_data)
head(maize_genomic_data)[, 1:10]

# Generate row and column variables per environment (i.e. combination of Site, year, Management
# and block in maize_genomic_data)

# NOTE: the Envir variable is not created in maize_genomic_data. Indeed, the two fixed-effect
# variables, row and column number, are fitted with wiser as quantitative variables per environment
# to limit the high number of fixed-effect variables (see Jacquin et al. 2025). Hence, they are
# strongly correlated to each environment for which they are fitted. Therefore, it is not necessary
# to create and fit the Envir variable due to strong redundancy and multicollinearity.
maize_raw_pheno_data <- generate_row_column_variables_by_environment(
  maize_raw_pheno_data
)

```

```

# For the genomic data, assign genotypes as row names (if necessary) to comply with wiser functions,
# and remove the "Genotype" column (if present). Note that "V1" corresponds to "Genotype" column here.
rownames(maize_genomic_data) <- maize_genomic_data$V1
maize_genomic_data <- maize_genomic_data[, -match(
  "V1", colnames(maize_genomic_data)
)]

# Define a trait for phenotypic estimation using wiser (note: some data should be available for
# the trait)
trait_ <- "anthesis"

# Get fixed-effect variables, row and column, which are fitted as quantitative variables for each
# environment (i.e. combination of Site, year, Management and block in maize_genomic_data)
fixed_effect_vars_ <- grep("_row$|_column$", colnames(maize_raw_pheno_data), value = TRUE)

# >>> Compute WISER-estimated phenotypes using estimate_wiser_phenotype() with its default values
# for whitening_method ("ZCA-cor") and alpha_ (0.01). Remark that the overall mean is always fitted
# within the wiser framework

# Estimate "anthesis" (i.e. trait_) phenotypes for genotypes using estimate_wiser_phenotype()
wiser_obj <- estimate_wiser_phenotype(
  maize_genomic_data,
  maize_raw_pheno_data,
  trait_,
  fixed_effect_vars = fixed_effect_vars_,
  fixed_effect_vars_computed_as_factor = NULL,
  envir_var = NULL,
  fixed_effect_vars_computed_as_factor_by_envir = NULL,
  random_effects_vars = "Genotype"
)

# >>> Plot WISER-estimated phenotypes density, display wiser fixed-effect estimates, and verify
# the whitening property

# Plot the density for the estimated phenotypes
dev.new()
plot(density(wiser_obj$wiser_phenotypes$v_hat), main = paste0(trait_, " v_hat"))

# Print the fixed-effect estimates computed from the whitening process and OLS
print(wiser_obj$wiser_fixed_effect_estimates)

# Print the estimated variance components (from ABC)
print(wiser_obj$wiser_abc_variance_component_estimates)

# Print the fundamental whitening property, i.e.  $In = W \Sigma_u W'$ 
id_mat <- wiser_obj$w_mat %*% wiser_obj$sig_mat_u %*% t(wiser_obj$w_mat)
print(id_mat[1:5, 1:5])

```

Phenotypic estimation for rice data (Baerstchi et al., 2021)

```

# >>> Load wiser library, display package help ,and attach rice data

# Load library
library(wiser)

# Show help for library
help(package = "wiser")

# Attach rice data
data("rice_raw_pheno_data")

```



```

data("rice_genomic_data")

# Display rice data
head(rice_raw_pheno_data)
head(rice_genomic_data)[, 1:10]

# For the raw phenotypic data, create the Envir variable in rice_raw_pheno_data by
# combining TRIAL, BLOC, and GENERATION. This variable represents the environments (i.e., levels)
rice_raw_pheno_data$Envir <- paste0(
  rice_raw_pheno_data$TRIAL, "_",
  rice_raw_pheno_data$BLOC, "_",
  rice_raw_pheno_data$GENERATION.
)

# For the genomic data, assign genotypes as row names (if necessary) to comply with wiser functions,
# and remove the "Genotype" column (if present). Note that "V1" corresponds to "Genotype" column here.
rownames(rice_genomic_data) <- rice_genomic_data$V1
rice_genomic_data <- rice_genomic_data[, -match(
  "V1", colnames(rice_genomic_data)
)]

# Define a trait for phenotypic estimation using wiser (note: some data should be available for
# the trait)
trait_ <- "ZN" # Zinc concentration

# NOTE: no spatial information is available in rice_genomic_data. Hence, only Envir is fitted as a
# fixed-effect factor

# >>> Compute WISER-estimated phenotypes using estimate_wiser_phenotype() with its default values
# for whitening_method ("ZCA-cor") and alpha_ (0.01). Remark that the overall mean is always fitted
# within the wiser framework

# Estimate "ZN" (i.e. trait_) phenotypes for genotypes using estimate_wiser_phenotype()
wiser_obj <- estimate_wiser_phenotype(
  rice_genomic_data,
  rice_raw_pheno_data,
  trait_,
  fixed_effect_vars = "Envir",
  fixed_effect_vars_computed_as_factor = "Envir",
  envir_var = NULL,
  fixed_effect_vars_computed_as_factor_by_envir = NULL,
  random_effects_vars = "Genotype"
)

# >>> Plot WISER-estimated phenotypes density, display wiser fixed-effect estimates, and verify
# the whitening property

# Plot the density for the estimated phenotypes
dev.new()
plot(density(wiser_obj$wiser_phenotypes$v_hat), main = paste0(trait_, " v_hat"))

# Print the fixed-effect estimates computed from the whitening process and OLS
print(wiser_obj$wiser_fixed_effect_estimates)

# Print the estimated variance components (from ABC)
print(wiser_obj$wiser_abc_variance_component_estimates)

# Print the fundamental whitening property, i.e.  $In = W \Sigma W'$ 
id_mat <- wiser_obj$w_mat %*% wiser_obj$SIG_mat_u %*% t(wiser_obj$w_mat)
print(id_mat[1:5, 1:5])

```

Author and maintainer

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