

R -Assignment

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Load necessary libraries

```
library(ggplot2)
```

```
## Warning: package 'ggplot2' was built under R version 4.4.3
```

```
library(dplyr)
```

```
## Warning: package 'dplyr' was built under R version 4.4.3
```

```
##
```

```
## Attaching package: 'dplyr'
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
## filter, lag
```

```
## The following objects are masked from 'package:base':
```

```
##
```

```
## intersect, setdiff, setequal, union
```

```
library(tidyverse)
```

```
## Warning: package 'tidyverse' was built under R version 4.4.3
```

```
## Warning: package 'readr' was built under R version 4.4.3
```

```
## Warning: package 'forcats' was built under R version 4.4.3
```

```
## Warning: package 'lubridate' was built under R version 4.4.3
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
```

```
## v forcats 1.0.0      v stringr 1.5.1
```

```
## v lubridate 1.9.4    v tibble 3.2.1
```

```
## v purrr 1.0.2       v tidyr 1.3.1
```

```
## v readr 2.1.5
```

```
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
library(tidyr)
library(purrr)
```

Load SNP genotype data and SNP position data from input files using read_tsv

read_tsv() reads a tab-separated values (TSV) file into a data frame (or tibble). fang_data will store the contents of fang_et_al_genotypes.txt.

```
#fang_data <- read_tsv("~/ISU/SPRING 2025/BCB4560/UNIX_Assignment/fang_et_al_genotypes.txt")
#snp_data <- read_tsv("~/ISU/SPRING 2025/BCB4560/UNIX_Assignment/snp_position.txt")
```

Alternative means of loading the data

```
fang_data <- read_tsv("https://raw.githubusercontent.com/EEOB-BioData/BCB546_Spring2025/main/assignment1/fang_et_al_genotypes.txt")
```

```
## Rows: 2782 Columns: 986
## -- Column specification -----
## Delimiter: "\t"
## chr (986): Sample_ID, JG_OTU, Group, abph1.20, abph1.22, ae1.3, ae1.4, ae1.5...
##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
```

```
snp_data <- read_tsv("https://raw.githubusercontent.com/EEOB-BioData/BCB546_Spring2025/main/assignments/snp_position.txt")
```

```
## Rows: 983 Columns: 15
## -- Column specification -----
## Delimiter: "\t"
## chr (9): SNP_ID, Chromosome, Position, alt_pos, mult_positions, amplicon, cd...
## dbl (6): cdv_marker_id, Genaissance_daa_id, Sequenom_daa_id, count_amplicons...
##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
```

Data Inspection for Genotype data (fang_et_al_genotypes)

head(fang_data) will display the first few rows of each dataset.

tail(fang_data) will display the last few rows of each dataset.

object.size(fang_data): This function returns the memory size of the fang_data object in bytes.

nrow(fang_data) returns the number of rows in the fang_data data frame.

ncol(fang_data) returns the number of columns (features or variables) in the data frame.

cat() prints the messages in a clean format without quotes.

The slash “n” adds a new line for readability.

`dim(fang_data)` returns the dataset’s dimensions as a vector

`dim(fang_data)[1]` is the number of rows and `dim(fang_data)[2]` is the number of columns.

Count the number of missing values represented as “?/?”

`fang_data == “?/?”` creates a logical matrix where each element is TRUE if it equals “?/?”, and FALSE otherwise.

`sum(..., na.rm = TRUE)` counts the number of TRUE values, effectively giving the total number of missing values.

`na.rm = TRUE` ensures that any existing NA values in `fang_data` do not interfere with the sum.

```
# Display the first few rows of the fang_data dataset to get an overview of the data.
head(fang_data)
```

```
## # A tibble: 6 x 986
##   Sample_ID JG_OTU   Group abph1.20 abph1.22 ae1.3 ae1.4 ae1.5 an1.4 ba1.6 ba1.9
##   <chr>      <chr>   <chr> <chr>      <chr>      <chr> <chr> <chr> <chr> <chr> <chr>
## 1 SL-15     T-aust-1 TRIPS  ??/?      ??/?      T/T   G/G   T/T   C/C   ??/?   G/G
## 2 SL-16     T-aust-2 TRIPS  ??/?      ??/?      T/T   ??/?  T/T   C/C   A/G    G/G
## 3 SL-11     T-brav-1 TRIPS  ??/?      ??/?      T/T   G/G   T/T   ??/?  G/G    G/G
## 4 SL-12     T-brav-2 TRIPS  ??/?      ??/?      T/T   G/G   T/T   C/C   G/G    G/G
## 5 SL-18     T-cund   TRIPS  ??/?      ??/?      T/T   G/G   T/T   C/C   ??/?  G/G
## 6 SL-2      T-dact-1 TRIPS  ??/?      ??/?      T/T   G/G   T/T   C/C   A/G    G/G
## # i 975 more variables: bt2.5 <chr>, bt2.7 <chr>, bt2.8 <chr>, Fea2.1 <chr>,
## #   Fea2.5 <chr>, id1.3 <chr>, lg2.11 <chr>, lg2.2 <chr>, pbf1.1 <chr>,
## #   pbf1.2 <chr>, pbf1.3 <chr>, pbf1.5 <chr>, pbf1.6 <chr>, pbf1.7 <chr>,
## #   pbf1.8 <chr>, PZA00003.11 <chr>, PZA00004.2 <chr>, PZA00005.8 <chr>,
## #   PZA00005.9 <chr>, PZA00006.13 <chr>, PZA00006.14 <chr>, PZA00008.1 <chr>,
## #   PZA00010.5 <chr>, PZA00013.10 <chr>, PZA00013.11 <chr>, PZA00013.9 <chr>,
## #   PZA00015.4 <chr>, PZA00017.1 <chr>, PZA00018.5 <chr>, ...
```

```
# Display the last few rows of the fang_data dataset to inspect how the data ends.
tail(fang_data)
```

```
## # A tibble: 6 x 986
##   Sample_ID JG_OTU   Group abph1.20 abph1.22 ae1.3 ae1.4 ae1.5 an1.4 ba1.6 ba1.9
##   <chr>      <chr>   <chr> <chr>      <chr>      <chr> <chr> <chr> <chr> <chr> <chr>
## 1 SYN262    Zmm-IL~ ZMMIL C/C      A/A      T/T   G/G   C/C   C/C   G/G    G/G
## 2 S0398      Zmm-IL~ ZMMIL G/G      A/A      T/T   G/G   C/C   C/C   G/G    G/G
## 3 S1636      Zmm-IL~ ZMMIL G/G      A/A      T/T   G/G   C/C   C/C   G/G    G/G
## 4 CU0201     Zmm-IL~ ZMMIL C/C      A/A      T/T   G/G   C/C   C/C   G/G    G/G
## 5 S0215      Zmm-IL~ ZMMIL G/G      A/A      T/T   ??/?  C/C   C/C   G/G    G/G
## 6 CU0202     Zmm-IL~ ZMMIL C/C      A/A      T/T   G/G   C/C   C/C   ??/?  G/G
## # i 975 more variables: bt2.5 <chr>, bt2.7 <chr>, bt2.8 <chr>, Fea2.1 <chr>,
## #   Fea2.5 <chr>, id1.3 <chr>, lg2.11 <chr>, lg2.2 <chr>, pbf1.1 <chr>,
## #   pbf1.2 <chr>, pbf1.3 <chr>, pbf1.5 <chr>, pbf1.6 <chr>, pbf1.7 <chr>,
## #   pbf1.8 <chr>, PZA00003.11 <chr>, PZA00004.2 <chr>, PZA00005.8 <chr>,
## #   PZA00005.9 <chr>, PZA00006.13 <chr>, PZA00006.14 <chr>, PZA00008.1 <chr>,
## #   PZA00010.5 <chr>, PZA00013.10 <chr>, PZA00013.11 <chr>, PZA00013.9 <chr>,
## #   PZA00015.4 <chr>, PZA00017.1 <chr>, PZA00018.5 <chr>, ...
```

```
# Check the size of the dataset
```

```
cat("The fang_data object occupies approximately",
    format(object.size(fang_data), units = "MB"), "of memory.\n")
```

```
## The fang_data object occupies approximately 22.1 Mb of memory.
```

```
# Calculate the number of rows in the fang_data dataset
num_lines_fang_data <- nrow(fang_data)
# Calculate the number of columns in the fang_data dataset.
num_columns_fang_data <- ncol(fang_data)
cat("The fang_et_al_genotypes dataset contains", num_lines_fang_data, "rows (samples) and",
    num_columns_fang_data, "columns .\n")
```

```
## The fang_et_al_genotypes dataset contains 2782 rows (samples) and 986 columns .
```

```
# Display the dimensions of the dataset
#dim(fang_data)

cat("The dimensions of the dataset are:", dim(fang_data)[1], "rows and", dim(fang_data)[2], "columns.\n")
```

```
## The dimensions of the dataset are: 2782 rows and 986 columns.
```

```
num_missing_fang <- sum(fang_data == "?/?", na.rm = TRUE)
cat("The sum of the missing values is:", num_missing_fang)
```

```
## The sum of the missing values is: 135452
```

```
# Count the number of missing values represented as "?/?" in the dataset
num_missing_fang <- sum(fang_data == "?/?", na.rm = TRUE)

# Print the total number of missing values
cat("The sum of the missing values in the fang_data:", num_missing_fang)
```

```
## The sum of the missing values in the fang_data: 135452
```

Data Inspection for Genotype data (fang_et_al_genotypes)

view(fang_data) opens the data in a spreadsheet-like viewer within RStudio.

```
# View the dataset (This will open it in RStudio's Data Viewer)
cat("Opening the fang_data dataset in the Viewer...\n")
```

```
## Opening the fang_data dataset in the Viewer...
```

```
View(fang_data)
```

Data Inspection for Genotype data (fang_et_al_genotypes)

str(fang_data) displays the structure of the fang_data object including data type of each column

```
# Display the structure of the dataset
cat("Here is the structure of the fang_data dataset:\n")
```

```
## Here is the structure of the fang_data dataset:
```

```
#str(fang_data)
```

The same file inspections were don for SNP Position data

Data Inspection for SNP Position data (snp_position)

```
# Display the first few rows of the snp_data dataset to get an overview of the data.
head(snp_data)
```

```
## # A tibble: 6 x 15
##   SNP_ID   cdv_marker_id Chromosome Position  alt_pos mult_positions amplicon
##   <chr>         <dbl> <chr>      <chr>      <chr>   <chr>          <chr>
## 1 abph1.20      5976 2        27403404 <NA>    <NA>          abph1
## 2 abph1.22      5978 2        27403892 <NA>    <NA>          abph1
## 3 ae1.3         6605 5        167889790 <NA>    <NA>          ae1
## 4 ae1.4         6606 5        167889682 <NA>    <NA>          ae1
## 5 ae1.5         6607 5        167889821 <NA>    <NA>          ae1
## 6 an1.4         5982 1        240498509 <NA>    <NA>          an1
## # i 8 more variables: cdv_map_feature.name <chr>, gene <chr>,
## #   'candidate/random' <chr>, Genaissance_daa_id <dbl>, Sequenom_daa_id <dbl>,
## #   count_amplicons <dbl>, count_cmf <dbl>, count_gene <dbl>
```

```
# Display the last few rows of the snp_data dataset to inspect how the data ends.
tail(snp_data)
```

```
## # A tibble: 6 x 15
##   SNP_ID   cdv_marker_id Chromosome Position  alt_pos mult_positions amplicon
##   <chr>         <dbl> <chr>      <chr>      <chr>   <chr>          <chr>
## 1 zap1.2       3514 2        233128584 <NA>    <NA>          zap1
## 2 zen1.1       3519 unknown unknown  <NA>    <NA>          zen1
## 3 zen1.2       3520 unknown unknown  <NA>    <NA>          zen1
## 4 zen1.4       3521 unknown unknown  <NA>    <NA>          zen1
## 5 zfl2.6       6463 2        12543294 <NA>    <NA>          zfl2
## 6 zmm3.4       3527 9        16966348 <NA>    <NA>          zmm3
## # i 8 more variables: cdv_map_feature.name <chr>, gene <chr>,
## #   'candidate/random' <chr>, Genaissance_daa_id <dbl>, Sequenom_daa_id <dbl>,
## #   count_amplicons <dbl>, count_cmf <dbl>, count_gene <dbl>
```

```
# Check the size of the dataset
cat("The snp_data object occupies approximately",
    format(object.size(snp_data), units = "MB"), "of memory.\n")
```

```
## The snp_data object occupies approximately 0.3 Mb of memory.
```

```
# Calculate the number of rows in the snp_data dataset
num_lines_snp_data <- nrow(snp_data)

# Calculate the number of columns in the snp_data dataset.
num_columns_snp_data <- ncol(snp_data)

cat("The snp_position dataset contains", num_lines_snp_data, "rows (SNP positions) and",
    num_columns_snp_data, "columns.\n")
```

```
## The snp_position dataset contains 983 rows (SNP positions) and 15 columns.
```

```
# Display the dimensions of the dataset
cat("The dimensions of the dataset are:", dim(snp_data)[1], "rows and", dim(snp_data)[2], "columns.\n")
```

```
## The dimensions of the dataset are: 983 rows and 15 columns.
```

```
# Count the number of missing values represented as "?/?" in the dataset
num_missing_snp <- sum(snp_data == "?/?", na.rm = TRUE)

# Print the total number of missing values
cat("The sum of the missing values in snp_data is:", num_missing_snp)
```

```
## The sum of the missing values in snp_data is: 0
```

Data Inspection for SNP Position data (snp_position)

```
# View the snp_position dataset (This will open it in RStudio's Data Viewer)
cat("Opening the snp_position in the Viewer...\n")
```

```
## Opening the snp_position in the Viewer...
```

```
View(snp_data)
```

Data Inspection for SNP Position data (snp_position)

```
# Display the structure of the dataset
cat("Here is the structure of the snp_position:\n")
```

```
## Here is the structure of the snp_position:
```

```
str(snp_data)
```

```
## spc_tbl_ [983 x 15] (S3: spec_tbl_df/tbl_df/tbl/data.frame)
## $ SNP_ID : chr [1:983] "abph1.20" "abph1.22" "ae1.3" "ae1.4" ...
## $ cdv_marker_id : num [1:983] 5976 5978 6605 6606 6607 ...
## $ Chromosome : chr [1:983] "2" "2" "5" "5" ...
## $ Position : chr [1:983] "27403404" "27403892" "167889790" "167889682" ...
## $ alt_pos : chr [1:983] NA NA NA NA ...
## $ mult_positions : chr [1:983] NA NA NA NA ...
## $ amplicon : chr [1:983] "abph1" "abph1" "ae1" "ae1" ...
## $ cdv_map_feature.name: chr [1:983] "AB042260" "AB042260" "ae1" "ae1" ...
## $ gene : chr [1:983] "abph1" "abph1" "ae1" "ae1" ...
## $ candidate/random : chr [1:983] "candidate" "candidate" "candidate" "candidate" ...
## $ Genaissance_daa_id : num [1:983] 8393 8394 8395 8396 8397 ...
## $ Sequenom_daa_id : num [1:983] 10474 10475 10477 10478 10479 ...
## $ count_amplicons : num [1:983] 1 0 1 0 0 1 1 0 1 0 ...
## $ count_cmf : num [1:983] 1 0 1 0 0 1 0 0 1 0 ...
## $ count_gene : num [1:983] 1 0 1 0 0 1 1 0 1 0 ...
## - attr(*, "spec")=
## .. cols(
## .. SNP_ID = col_character(),
## .. cdv_marker_id = col_double(),
## .. Chromosome = col_character(),
## .. Position = col_character(),
## .. alt_pos = col_character(),
## .. mult_positions = col_character(),
## .. amplicon = col_character(),
## .. cdv_map_feature.name = col_character(),
## .. gene = col_character(),
## .. 'candidate/random' = col_character(),
## .. Genaissance_daa_id = col_double(),
## .. Sequenom_daa_id = col_double(),
## .. count_amplicons = col_double(),
## .. count_cmf = col_double(),
## .. count_gene = col_double()
## .. )
## - attr(*, "problems")=<externalptr>
```

Data Processing

`t(fang_data)` will transpose the dataset, swapping rows and columns.

`as.data.frame(..., stringsAsFactors = FALSE)` will convert the transposed matrix back into a data frame while preventing automatic conversion of text to factors. `fang_transposed[3,]` is used to extract the third row to use as column names, and `colnames(fang_transposed) <- ...` is used to assign the third row values as the new column names for the transposed data.

`rownames(fang_transposed)`: Retrieves the original row names from the transposed data and `cbind(...)` is used to combine the original row names as a new first column (Original_Colnames).

`fang_transposed[-c(1:3),]`: removes the first 3 rows, which are no longer needed after setting column names. `colnames(fang_transposed)[1] <- "SNP_ID"`: Renames the first column to "SNP_ID".

`rownames(fang_transposed) <- NULL`: resets row names to sequential integers starting from 1.

```
# Transpose the fang_data dataset to switch rows and columns
fang_transposed <- as.data.frame(t(fang_data), stringsAsFactors = FALSE)
```

```

# View the transposed data (optional, for checking)
view(fang_transposed)

# Convert the third row into column names (assuming row 3 contains SNP names)
colnames(fang_transposed) <- fang_transposed[3, ]

# Add the original row names as a new first column to preserve them
fang_transposed <- cbind(Original_Colnames = rownames(fang_transposed), fang_transposed)

# Remove the first 3 rows as they are not needed after setting column names
fang_transposed <- fang_transposed[-c(1:3), ]

# Rename the first column to "SNP_ID" for clarity
colnames(fang_transposed)[1] <- "SNP_ID"

# View the cleaned transposed data (optional, for checking)
view(fang_transposed)

# Reset row names to ensure they are sequential and start from 1
rownames(fang_transposed) <- NULL

```

Joining the transposed genotype dat and the SNP position data by SNP

`select(snp_data, SNP_ID, Chromosome, Position)`: Extracts `SNP_ID`, `Chromosome`, and `Position` columns from `snp_data`.

`make.unique(colnames(fang_transposed))`: Ensures unique column names in `fang_transposed`.

`left_join(snp_info, fang_transposed, by = "SNP_ID")`: Merges `snp_info` with `fang_transposed` based on `SNP_ID`.

`write.table(df_joined, "joined_data.txt", sep = "\t", row.names = FALSE, quote = FALSE)`: Saves the joined dataset as a tab-separated text file without row names or quotes.

```

# Select relevant columns (SNP_ID, Chromosome, Position) from snp_data
snp_info <- select(snp_data, SNP_ID, Chromosome, Position)

# Ensure unique column names in fang_transposed to avoid duplication issues
colnames(fang_transposed) <- make.unique(colnames(fang_transposed))

# Perform a left join to merge snp_info with fang_transposed based on SNP_ID
df_joined <- left_join(snp_info, fang_transposed, by = "SNP_ID")

# View the resulting dataframe in RStudio
view(df_joined)

# Save the joined dataset as a tab-separated text file
write.table(df_joined, "joined_fang_snp_data.txt", sep = "\t", row.names = FALSE, quote = FALSE)

```


Maize

Extract relevant columns for maize

The code below uses `select()` to extract relevant columns from `df_joined`.

Keeps `SNP_ID`, `Chromosome`, and `Position` columns.

Selects all columns whose names start with “ZMMIL”, “ZMMLR”, or “ZMMMR” (presumably maize-related data).

The `write.table(...)` saves `df_joined` as a tab-separated text file named `maize_data.txt` inside the “Maize” directory

`row.names = FALSE` prevents row numbers from being written.

`quote = FALSE` ensures that values are not enclosed in quotes.

```
# Extract relevant columns for maize, including SNP_ID, Chromosome, Position, and columns starting with
df_maize <- df_joined %>% select(SNP_ID, Chromosome = Chromosome, Position = Position,
                                starts_with("ZMMIL"), starts_with("ZMMLR"), starts_with("ZMMMR"))

# View the maize-specific dataset in RStudio
view(df_maize)

write.table(df_joined, "Maize/maize_data.txt", sep = "\t", row.names = FALSE, quote = FALSE)
```

Extract rows with ‘multiple’ or ‘unknown’ in Chromosome column for Maize

The uses `filter()` to filter `df_maize` to keep only rows where the `Chromosome` column has the value “multiple” and saves the result in `df_multiple`.

Similarly filters `df_maize` to keep only rows where the `Chromosome` column has the value “unknown”. The filtered `df_multiple` and `df_unknown` datasets as tab-separated .txt files, named `maize_chromo_multiple.txt` and `maize_chromo_unknown.txt` are respectively saved in the “Maize” directory

```
# Extract rows with 'multiple' in the Chromosome column
df_multiple <- df_maize %>% filter(Chromosome == "multiple")

# Extract rows with 'unknown' in the Chromosome column
df_unknown <- df_maize %>% filter(Chromosome == "unknown")

# Save the 'multiple' chromosome data to a tab-separated text file
write.table(df_multiple, "Maize/maize_chromo_multiple.txt", sep = "\t", row.names = FALSE, quote = FALSE)

# Save the 'unknown' chromosome data to a tab-separated text file
write.table(df_unknown, "Maize/maize_chromo_unknown.txt", sep = "\t", row.names = FALSE, quote = FALSE)
```

Sort by ascending order of SNP position

The code below uses `arrange()` to sort the `df_maize` dataset in ascending order based on the `Position` column.

Extracts and combines SNP data from chromosomes 1-10 in `df_maize`. Converts the `Position` column to a numeric type. The result is saved as `df_maize_sorted_incre`. the sorted dataset (`df_maize_sorted_incre`) as a tab-separated.txt file called `maize_sorted_incre_data.txt` in the “Maize” directory

```

# Sort by SNP position in increasing order
# Sort by SNP position
df_maize_chr_filtered <- data.frame() # Initialize an empty data frame to store sorted SNPs

for (chr_num in 1:10) { # Loop through chromosome numbers 1 to 10
  df_chr <- df_maize %>% # Filter the data for the current chromosome
    filter(Chromosome == chr_num) # Ensure column name matches the dataset structure

  df_maize_chr_filtered <- bind_rows(df_maize_chr_filtered, df_chr) # Append filtered data for each chromosome
}

# Convert Position column to numeric type and sort in increasing order
df_maize_sorted_incre <- df_maize_chr_filtered %>%
  mutate(Position = suppressWarnings(as.numeric(Position))) %>% # Convert Position to numeric type for sorting
  arrange(Position) # Sort the data by Position in increasing order

# View the sorted dataset in RStudio
view(df_maize_sorted_incre)

# Save the sorted maize dataset as a tab-separated text file
write.table(df_maize_sorted_incre, "Maize/maize_sorted_incre_data.txt", sep = "\t", row.names = FALSE, col.names = TRUE)

```

Create separate chromosome files for Increasing order of SNP Position for maize

The uses for loop to over chromosome numbers from 1 to 10.

the code filters df_maize_sorted_incre to include only rows where the chromosome column matches the current chromosome number (chr_num) and saves the filtered result as df_chr.

The filtered chromosome dataset (df_chr) are saved to a tab-separated text file. The file name includes the chromosome number (chr_num), such as Maize_incre_chromo1.txt for chromosome 1.

```

#Increasing SNP Position (Maize)
# Create separate chromosome files for chromosomes 1 to 10
for (chr_num in 1:10) {
  # Filter rows for each chromosome number
  df_chr <- df_maize_sorted_incre %>% filter(Chromosome == chr_num)

  # Save the filtered chromosome data to a tab-separated text file without row names or quotes
  write.table(df_chr, paste0("Maize/Maize_Increasing_Chromo/Maize_incre_chromo", chr_num, ".txt"), sep = "\t", row.names = FALSE, col.names = TRUE)
}

```

Create separate chromosome files for decreasing order of SNP Position for maize

Repalce “?/?” by “-/-” and Sort by decreasing order of SNP position

This code performs the following steps:

`df_maize_chr_filtered_dash[df_maize_chr_filtered_dash == “?/?”] <- “-/-”`. This step directly replaces the “?/?” placeholder with “-/-” in the dataset.

The modified dataset is saved as a tab-separated .txt file without row names and quotes. Extracts and combines SNP data from chromosomes 1-10 in `df_maize_dash`. Converts the Position column to a numeric type.

The `arrange(desc(Position))` is used to sort the dataset by the Position column in descending order. The result is saved as `df_maize_sorted_decre`. The sorted dataset is then saved as a tab-separated .txt file.

```
# maize decreasing SNP Positions
# Create a copy of the maize dataset
df_maize_chr_filtered_dash <- df_maize_chr_filtered

# Replace all occurrences of "?/?" with "-/-" in the dataset
df_maize_chr_filtered_dash[df_maize_chr_filtered_dash == "?/?"] <- "-/-"

# View the modified dataset in RStudio
view(df_maize_chr_filtered_dash)

# Save the modified dataset to a tab-separated text file
write.table(df_maize_chr_filtered_dash, "Maize/maize_chr_filtered_dash.txt", sep = "\t", row.names = FALSE)

# Sort the modified dataset by Position in descending order
#df_maize_sorted_decre_dash <- df_maize_dash %>% arrange(desc(Position))
# Convert Position column to numeric type and sort in decreasing order
df_maize_sorted_decre_dash <- df_maize_chr_filtered_dash %>%
  mutate(Position = suppressWarnings(as.numeric(Position))) %>% # Convert Position to numeric type for
  arrange(desc(Position)) # Sort the data by Position in decreasing order
# View the sorted dataset in RStudio
view(df_maize_sorted_decre_dash)

# Save the sorted dataset to a tab-separated text file
write.table(df_maize_sorted_decre_dash, "Maize/maize_sorted_decre_dash.txt", sep = "\t", row.names = FALSE)
```

Generate separate chromosome files for decreasing order of SNP Position for maize

The uses for loop to over chromosome numbers from 1 to 10.

the code filters `df_maize_sorted_incre` to include only rows where the chromosome column matches the current chromosome number (`chr_num`) and saves the filtered result as `df_chr`.

The filtered chromosome dataset (`df_chr`) are saved to a tab-separated text file. The file name includes the chromosome number (`chr_num`), such as `Maize_decre_chrom01.txt` for chromosome 1.

```

# Create separate chromosome files for chromosomes 1 to 10
for (chr_num in 1:10) {
  # Filter rows for each chromosome number
  df_chr <- df_maize_sorted_decre_dash %>% filter(Chromosome == chr_num)

  # Save the filtered chromosome data to a tab-separated text file
  write.table(df_chr, paste0("Maize/Maize_Decreasing_Chromo/Maize_decre_chromo", chr_num, ".txt"), sep = "\t", as.is = TRUE)
}

```

Teosinte

Similar to the codes used in maize, the codes below do same in teosinte. The summary of what the codes is below with detailed explanation seen in maize

Extract relevant columns for Teosinte

Extract Relevant Columns for Teosinte

The script extracts SNP data for Teosinte from a combined dataset (df_joined).

It selects SNP_ID, Chromosome, Position, and columns starting with “ZMPBA”, “ZMPIL”, or “ZMPJA”.

Saves the extracted data as “data_teosinte.txt”

```

# Define the directory path for saving the output file
file_dir = "~/ISU/SPRING 2025/BCB4560/R_Assignment_Final/Teosinte/"

# Extract relevant columns for Teosinte from the joined dataframe
# Selecting SNP_ID, Chromosome, Position, and columns that start with "ZMPBA", "ZMPIL", or "ZMPJA"
df_teosinte <- df_joined %>% select(SNP_ID, Chromosome = Chromosome, Position = Position,
                                   starts_with("ZMPBA"), starts_with("ZMPIL"), starts_with("ZMPJA"))

# View the extracted dataframe in RStudio
view(df_teosinte)

# Save the extracted Teosinte data to a text file with tab-separated values
write.table(df_teosinte, paste0(file_dir, "data_teosinte.txt"), sep = "\t", row.names = FALSE, quote = FALSE)

```

Extract rows with ‘multiple’ or ‘unknown’ in Chromosome column (Teosinte)

Identifies SNPs where Chromosome is labeled as “multiple” or “unknown”. Saves them separately as: “Teosinte_chromo_multiple.txt” and “Teosinte_chromo_unknown.txt”

```

# Extract rows where the Chromosome column has the value "multiple"
df_multiple <- df_teosinte %>% filter(Chromosome == "multiple")

# Extract rows where the Chromosome column has the value "unknown"
df_unknown <- df_teosinte %>% filter(Chromosome == "unknown")

# Save the extracted data for "multiple" chromosomes to a text file with tab-separated values
write.table(df_multiple, paste0(file_dir, "Teosinte_chromo_multiple.txt"), sep = "\t", row.names = FALSE, quote = FALSE)
write.table(df_unknown, paste0(file_dir, "Teosinte_chromo_unknown.txt"), sep = "\t", row.names = FALSE, quote = FALSE)

```

```
write.table(df_multiple, paste0(file_dir, "Teosinte_chromo_multiple.txt"),
            sep = "\t", row.names = FALSE, quote = FALSE)

# Save the extracted data for "unknown" chromosomes to a text file with tab-separated values
write.table(df_unknown, paste0(file_dir, "Teosinte_chromo_unknown.txt"),
            sep = "\t", row.names = FALSE, quote = FALSE)
```

Sort by ascending order of SNP position (Teosinte)

Sorts SNPs by Position in ascending order.

Saves the sorted data as “data_teosinte_sorted_incre.txt”.

```
# Sort by SNP position
df_teosinte_chr_filtered <- data.frame() # Initialize an empty data frame to store sorted SNPs

for (chr_num in 1:10) { # Loop through chromosome numbers 1 to 10
  df_chr <- df_teosinte %>% # Filter the data for the current chromosome
    filter(Chromosome == chr_num) # Ensure column name matches the dataset structure

  df_teosinte_chr_filtered <- bind_rows(df_teosinte_chr_filtered, df_chr) # Append filtered data for e
}

# Convert Position column to numeric type and sort in increasing order
df_teosinte_sorted_incre <- df_teosinte_chr_filtered %>%
  mutate(Position = suppressWarnings(as.numeric(Position))) %>% # Convert Position to numeric type for
  arrange(Position) # Sort the data by Position in ascending order
# View the sorted dataframe in RStudio
view(df_teosinte_sorted_incre)

# Save the sorted Teosinte data to a text file with tab-separated values
write.table(df_teosinte_sorted_incre, paste0(file_dir, "data_teosinte_sorted_incre.txt"),
            sep = "\t", row.names = FALSE, quote = FALSE)
```

Increasing SNP Position (Teosinte)

Iterates over chromosomes 1 to 10.

Filters SNPs for each chromosome.

Saves them in the “Teosinte_Increasing_Chromo” directory as:

“Teosinte_incre_chromo1.txt”

“Teosinte_incre_chromo2.txt”, etc.

```
# Create separate chromosome files for Teosinte (Increasing SNP Positions)
# Create separate files for each chromosome (1 to 10) in Teosinte dataset, sorted by increasing SNP pos
for (chr_num in 1:10) {

  # Filter the dataframe to include only rows where Chromosome matches the current chromosome number
  df_chr <- df_teosinte_sorted_incre %>% filter(Chromosome == chr_num)

  # Save the filtered data to a text file, named according to the chromosome number
  write.table(df_chr, paste0(file_dir, "/Teosinte_Increasing_Chromo/Teosinte_incre_chromo", chr_num, ".")
```

```
sep = "\t", row.names = FALSE, quote = FALSE)
}
```

Decreasing SNP Position (Teosinte)

Repalce “?/?” by “-/-” and Sort by decreasing order of SNP position

Creates a copy of the dataset.

Replaces all “?/?” genotypes with “-/-”.

Sorts SNPs by Position in descending order.

Saves the cleaned dataset as “data_teosinte_sorted_decre_dash.txt”.

```
# Create a copy of the Teosinte dataframe to modify the genotype format
df_teosinte_chr_filtered_dash <- df_teosinte_chr_filtered
# Replace all occurrences of "?/?" with "-/-" in the dataframe
df_teosinte_chr_filtered_dash[df_teosinte_chr_filtered_dash == "?/?"] <- "-/-"

# View the modified dataframe in RStudio
view(df_teosinte_chr_filtered_dash)

# Save the modified Teosinte data to a text file with tab-separated values
write.table(df_teosinte_chr_filtered_dash, paste0(file_dir, "teosinte_chr_filtered_dash.txt"),
            sep = "\t", row.names = FALSE, quote = FALSE)

# Sort the modified dataframe by SNP position in decreasing order
df_teosinte_sorted_decre_dash <- df_teosinte_chr_filtered_dash %>% arrange(desc(Position))

# Convert Position column to numeric type and sort in decreasing order
df_teosinte_sorted_decre_dash <- df_teosinte_chr_filtered_dash %>%
  mutate(Position = suppressWarnings(as.numeric(Position))) %>% # Convert Position to numeric type for
  arrange(Position) # Sort the data by Position in ascending order
# View the sorted dataframe in RStudio
view(df_teosinte_sorted_decre_dash)

# Save the sorted Teosinte data to a text file with tab-separated values
write.table(df_teosinte_sorted_decre_dash, paste0(file_dir, "data_teosinte_sorted_decre_dash.txt"),
            sep = "\t", row.names = FALSE, quote = FALSE)
```

Create Separate Files for Each Chromosome (Decreasing SNP Positions)

Similar to Step 4 but with SNPs sorted in descending order.

Saves files in “Teosinte_Decreasing_Chromo” directory.

```
# Create separate chromosome files for Teosinte (Decreasing SNP Positions)
# Create separate files for each chromosome (1 to 10) in the Teosinte dataset, sorted by decreasing SNP
for (chr_num in 1:10) {
```

```

# Filter the dataframe to include only rows where Chromosome matches the current chromosome number
df_chr <- df_teosinte_sorted_decre_dash %>% filter(Chromosome == chr_num)

# Save the filtered data to a text file, named according to the chromosome number, using tab-separated
write_tsv(df_chr, paste0(file_dir, "/Teosinte_Decreasing_Chromo/Teosinte_decre_chromo", chr_num, ".txt"),
}

```

#Part II Visualization

```

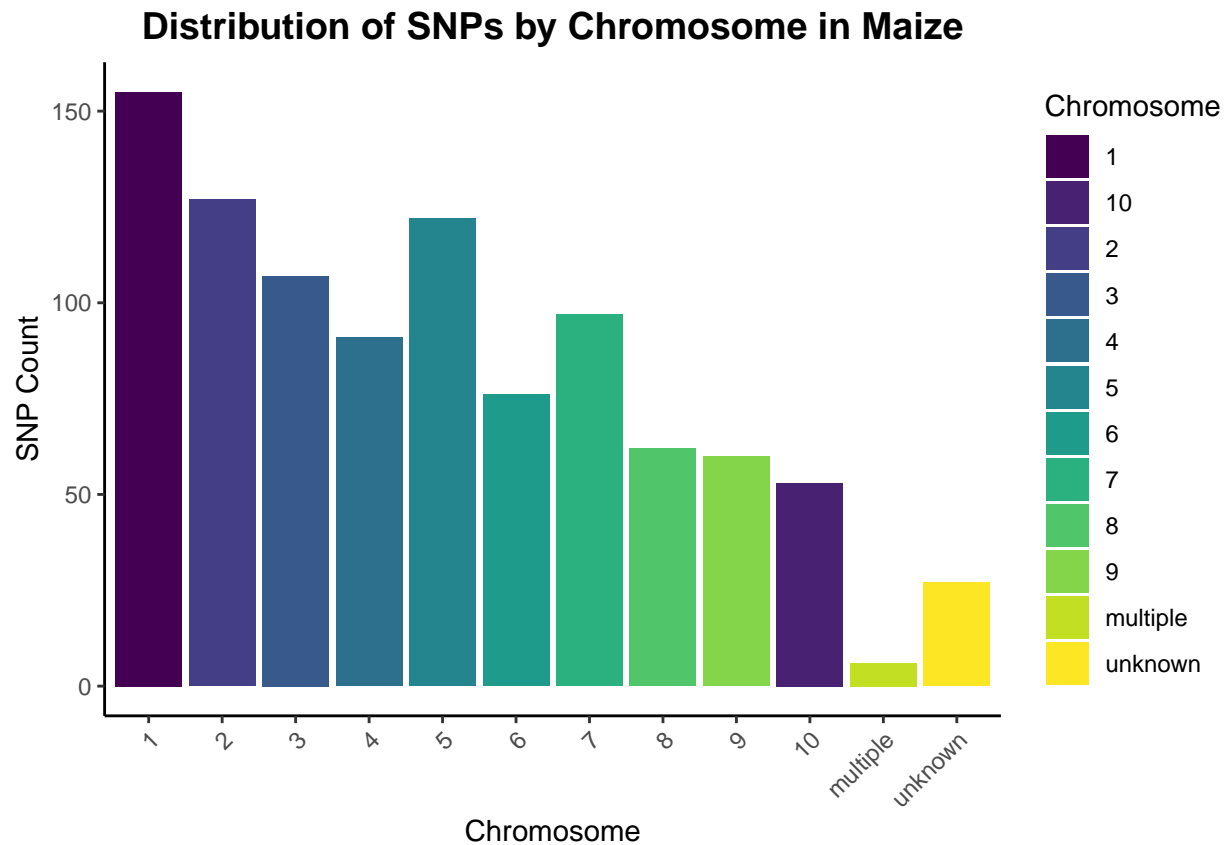
# Bar plot of SNP distribution by Chromosome in Maize
ggplot(data = df_maize, aes(x = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "11", "12")),
                           fill = factor(Chromosome))) +
  geom_bar() +
  ggtitle("Distribution of SNPs by Chromosome in Maize") +

# Improve theme and text positioning
theme_classic() +
theme(plot.title = element_text(hjust = 0.5, size = 14, face = "bold"),
      axis.text.x = element_text(angle = 45, hjust = 1)) +

# Improve axis labels
xlab("Chromosome") +
ylab("SNP Count") +

# Improve legend
scale_fill_viridis_d(name = "Chromosome") # Better color mapping

```

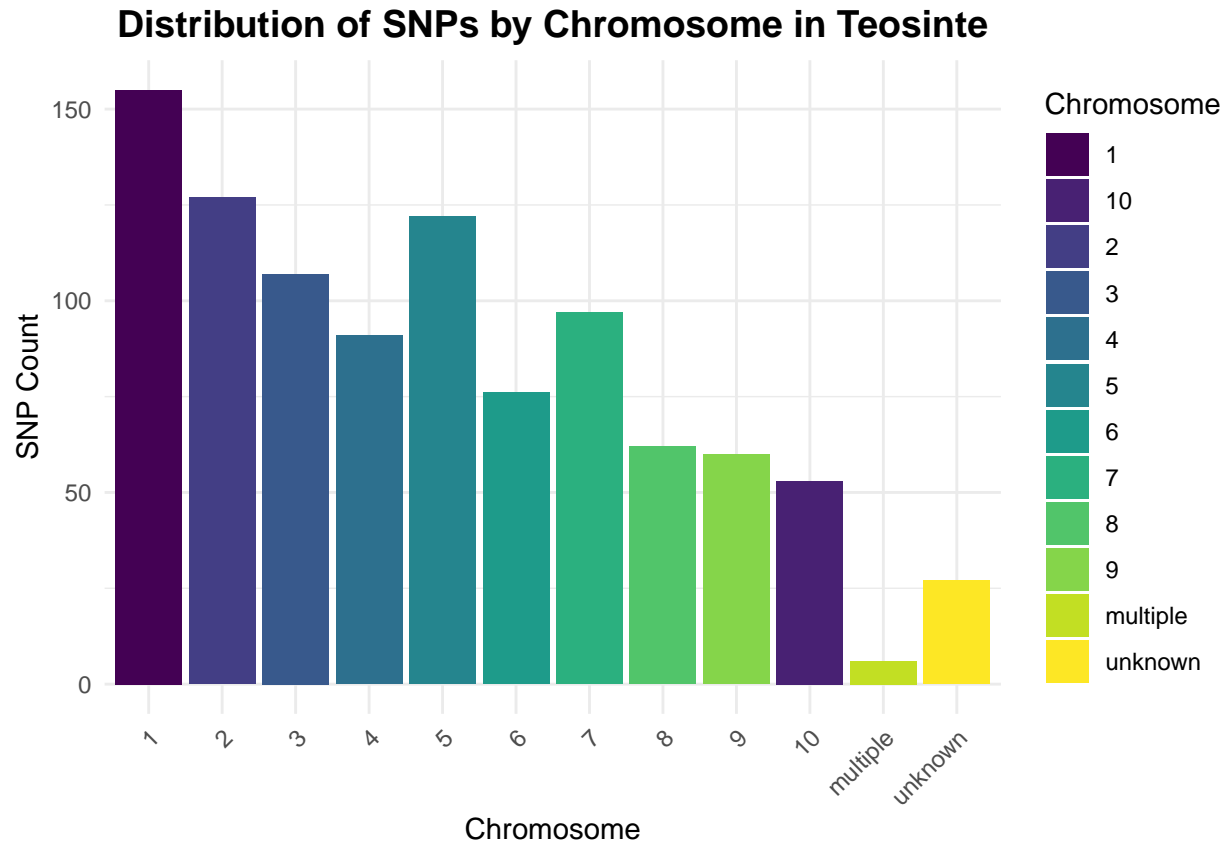


```
# Bar plot of SNP distribution by Chromosome in Teosinte
ggplot(data = df_teosinte, aes(x = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown"),
                                     fill = factor(Chromosome))) +
  geom_bar() +
  ggtitle("Distribution of SNPs by Chromosome in Teosinte") +

  # Improve theme and text positioning
  theme_minimal() +
  theme(plot.title = element_text(hjust = 0.5, size = 14, face = "bold"),
        axis.text.x = element_text(angle = 45, hjust = 1)) +

  # Improve axis labels
  xlab("Chromosome") +
  ylab("SNP Count") +

  # Improve legend
  scale_fill_viridis_d(name = "Chromosome") # Better color mapping
```

The `group_by(Chromosome)` function organizes the dataset (`df_maize`) into groups based on unique values in the `Chromosome` column.

The `summarise(SNP_Count = n())` function calculates the number of rows (SNPs) in each chromosome group and stores the count in a new column called `SNP_Count`.

The `mutate(Group = "Maize")` function adds a new column called `Group` to the resulting dataframe and assigns the value "Maize" to all rows. This helps in labeling the data when combining results from multiple datasets.

Count SNPs per chromosome for maize

```
# Count the number of SNPs per chromosome for the maize dataset
snp_count_maize <- df_maize %>%

# Group the data by the Chromosome column
group_by(Chromosome) %>%

# Count the number of SNPs in each chromosome group
summarise(SNP_Count = n()) %>%

# Add a new column to indicate the dataset source as "Maize"
mutate(Species = "Maize")
```

Count SNPs per chromosome for teosinte

```
# Count the number of SNPs per chromosome for the Teosinte dataset
snp_count_teosinte <- df_teosinte %>%

# Group the data by the Chromosome column
group_by(Chromosome) %>%

# Count the number of SNPs in each chromosome group
summarise(SNP_Count = n()) %>%

# Add a new column to indicate the dataset source as "Teosinte"
mutate(Species = "Teosinte")
```

#Combining Data from Two Dataframes:

The `bind_rows(snp_count_maize, snp_count_teosinte)` function merges the `snp_count_maize` and `snp_count_teosinte` dataframes by stacking them on top of each other.

This works because both dataframes have the same column structure (Chromosome, SNP_Count, and Group).

```
# Combine SNP count data from both Maize and Teosinte datasets
snp_counts <- bind_rows(snp_count_maize, snp_count_teosinte)
```

#Creating the Plot This code generates a bar plot to visualize the SNP distribution across chromosomes for both Maize and Teosinte

```
# Plot SNP count per chromosome for Maize and Teosinte
ggplot(snp_counts, aes(x = Chromosome, y = SNP_Count, fill = Species)) +

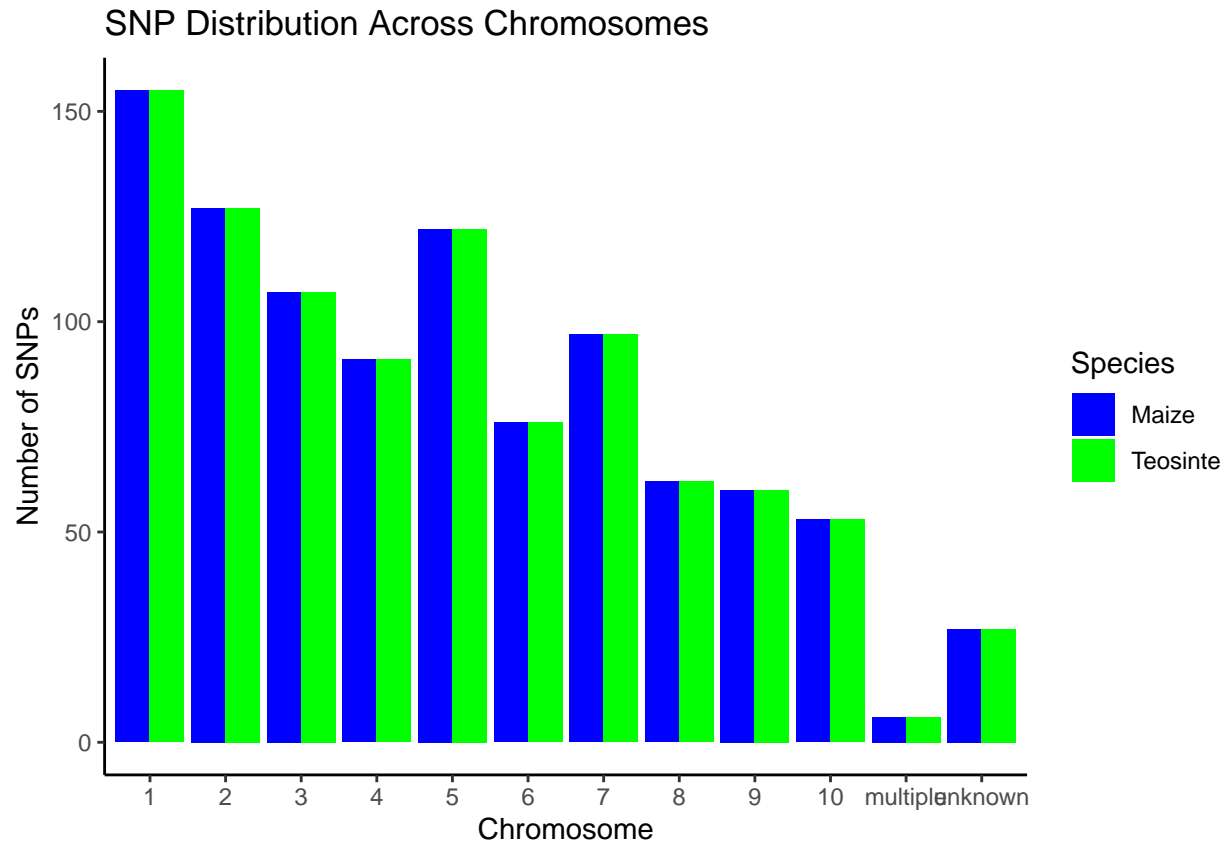
# Create a bar plot with "Chromosome" on the x-axis and "SNP_Count" on the y-axis
# The bars are grouped by "Group" (Maize vs. Teosinte)
geom_bar(stat = "identity", position = "dodge") +

# Add labels to the plot
labs(title = "SNP Distribution Across Chromosomes",
      x = "Chromosome", y = "Number of SNPs") +

# Set custom colors for the "Maize" and "Teosinte" groups
scale_fill_manual(values = c("Maize" = "blue", "Teosinte" = "green")) +

# Ensure chromosome labels appear in the correct order
scale_x_discrete(limits = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown"))

# Use a clean and simple theme for the plot
theme_classic()
```



```
# Scatter plot of SNP count per chromosome for Maize and Teosinte
ggplot(snp_counts, aes(x = Chromosome, y = SNP_Count, color = Species)) +

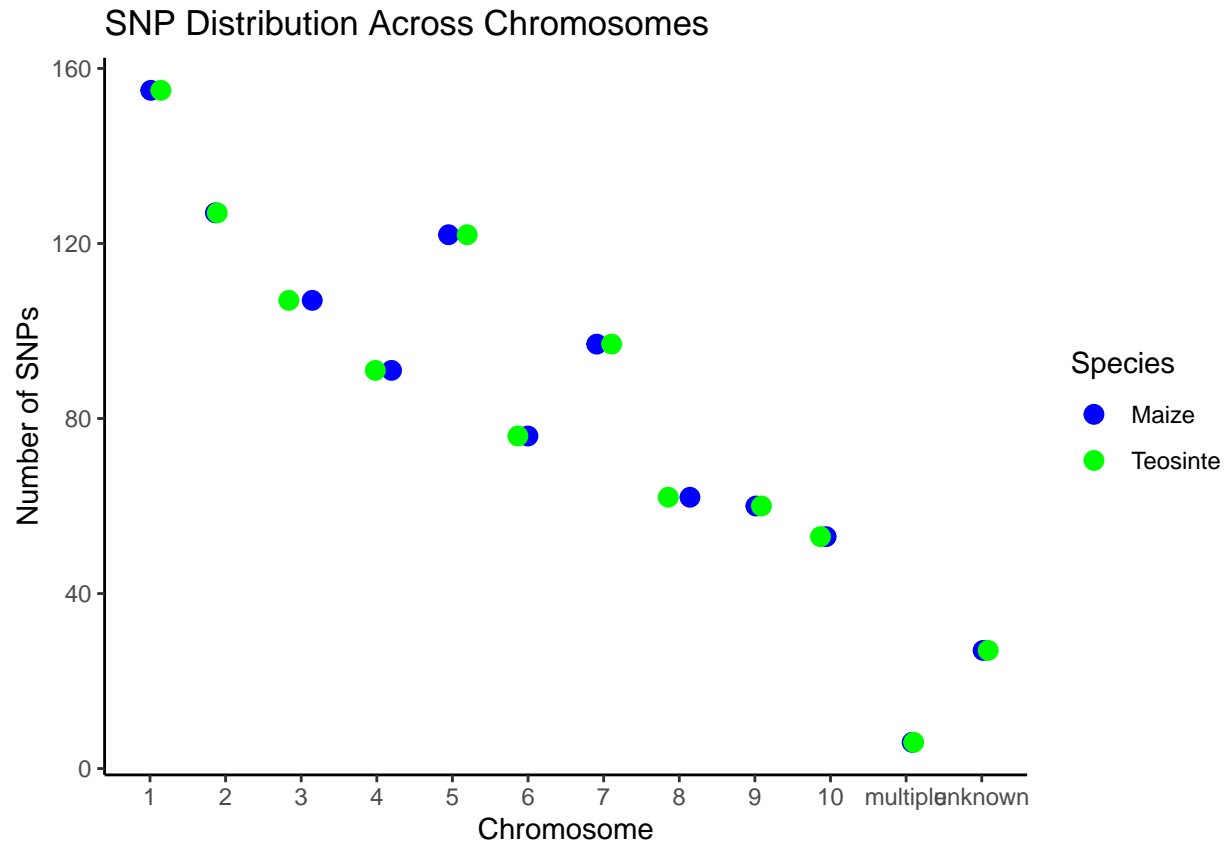
  # Create a scatter plot with "Chromosome" on the x-axis and "SNP_Count" on the y-axis
  geom_point(size = 3, position = position_jitter(width = 0.2, height = 0)) + # Add slight jitter for

  # Add labels to the plot
  labs(title = "SNP Distribution Across Chromosomes",
        x = "Chromosome", y = "Number of SNPs") +

  # Set custom colors for the "Maize" and "Teosinte" groups
  scale_color_manual(values = c("Maize" = "blue", "Teosinte" = "green")) +

  # Ensure chromosome labels appear in the correct order
  scale_x_discrete(limits = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown"))

  # Use a clean and simple theme for the plot
  theme_classic()
```



```
# Scatter plot of SNP distribution by Chromosome in Maize
ggplot(data = df_maize, aes(x = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown")),
                             y = Position,
                             color = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown")),
                             size = 1.5) + # Adjust transparency and size
  geom_point(alpha = 0.7, size = 1.5) +

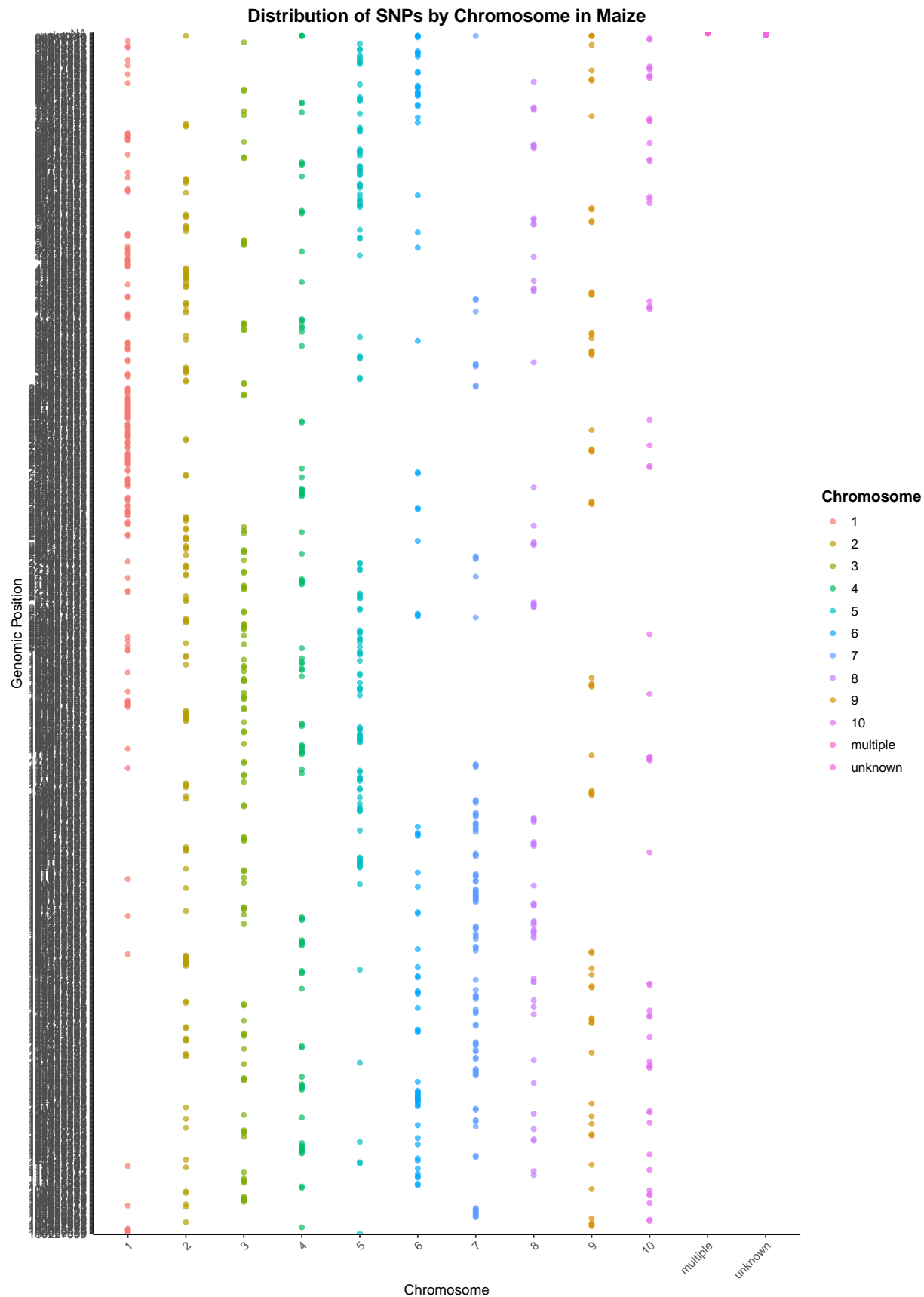
  ggtitle("Distribution of SNPs by Chromosome in Maize") +

  # Improve axis labels
  xlab("Chromosome") +
  ylab("Genomic Position") +

  # Improve theme and text positioning
  theme_classic() +
  theme(plot.title = element_text(hjust = 0.5, size = 14, face = "bold"),
        axis.text.x = element_text(angle = 45, hjust = 1),
        legend.title = element_text(size = 12, face = "bold"),
        legend.text = element_text(size = 10)) +

  # Explicitly define color mapping and ensure correct legend order
  scale_color_manual(name = "Chromosome",
                     values = c("1" = "#F8766D", "2" = "#B79F00", "3" = "#7CAE00", "4" = "#00BE67",
                                "5" = "#00BFC4", "6" = "#00A9FF", "7" = "#619CFF", "8" = "#C77CFF",
                                "9" = "#D89000", "10" = "#E76BF3", "multiple" = "#FF61CC", "unknown" = "#FFCC00"),
                     breaks = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown"))
```

```
guides(color = guide_legend(ncol = 1)) # Ensure legend is in one column
```



```

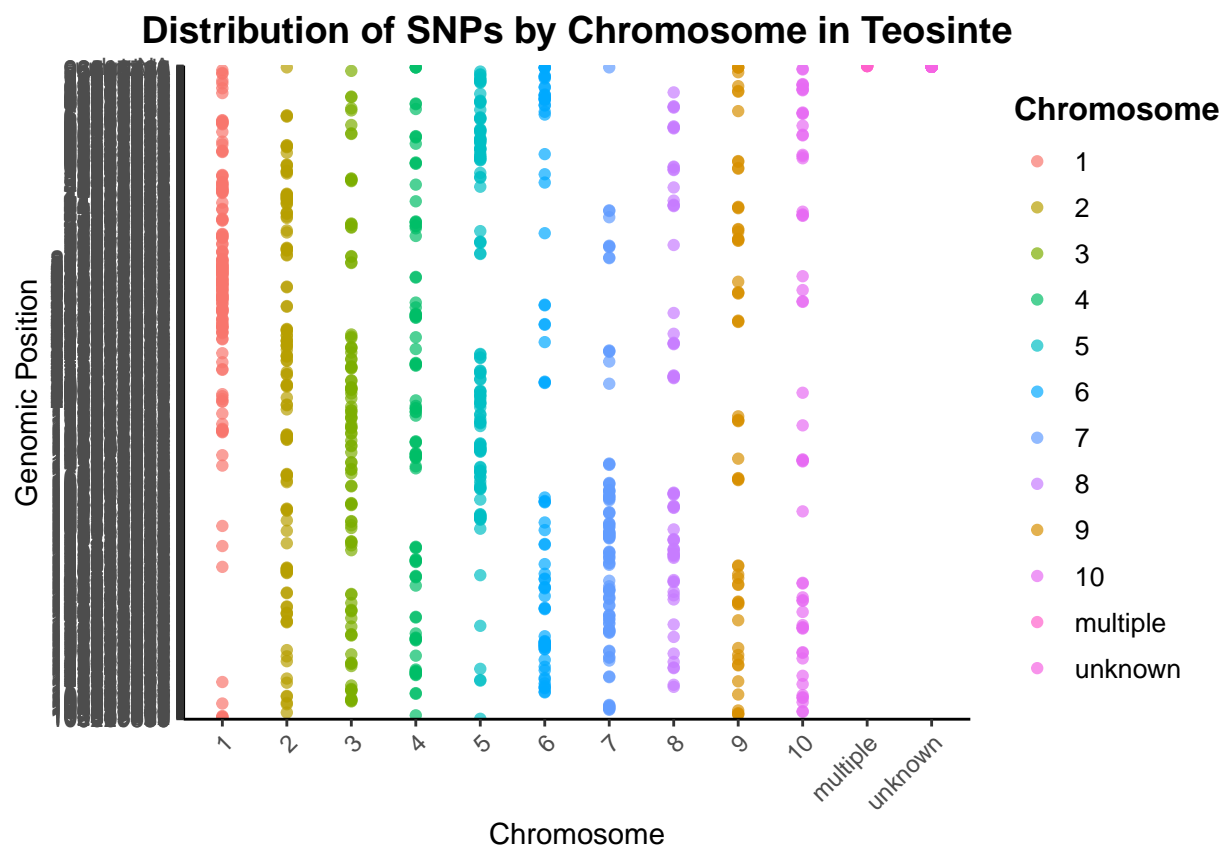
# Scatter plot of the distribution of SNPs by Chromosome in Teosinte
ggplot(data = df_teosinte, aes(x = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown")),
                                y = Position, color = factor(Chromosome, levels = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown")),
                                size = 1.5) + # Adjust point transparency and size for better visibility
  geom_point(alpha = 0.7, size = 1.5) +
  ggtitle("Distribution of SNPs by Chromosome in Teosinte") +

# Improve axis labels
  xlab("Chromosome") +
  ylab("Genomic Position") +

# Improve theme and text positioning
  theme_classic() +
  theme(plot.title = element_text(hjust = 0.5, size = 14, face = "bold"),
        axis.text.x = element_text(angle = 45, hjust = 1),
        legend.title = element_text(size = 12, face = "bold"),
        legend.text = element_text(size = 10)) +

# Explicitly define color mapping and ensure correct legend order
  scale_color_manual(name = "Chromosome",
                     values = c("1" = "#F8766D", "2" = "#B79F00", "3" = "#7CAE00", "4" = "#00BE67",
                                "5" = "#00BFC4", "6" = "#00A9FF", "7" = "#619CFF", "8" = "#C77CFF",
                                "9" = "#D89000", "10" = "#E76BF3", "multiple" = "#FF61CC", "unknown" = "#FF0000"),
                     breaks = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown")) +
  guides(color = guide_legend(ncol = 1)) # Ensure legend is in one column

```



For optimized mapping

This code reshapes the genotype data from both Maize and Teosinte, classifies each SNP as “Homozygous”, “Heterozygous”, or “Missing”, and then combines both datasets into a single dataframe (df_combined) that includes both Maize and Teosinte genotypes.

`pivot_longer`: Converts the maize data from wide format (where each sample is a column) to long format (where each row represents a genotype for a specific SNP and sample). The `SNP_ID`, `Chromosome`, and `Position` columns are kept intact, while the rest of the columns (representing samples) are “pivoted” into two new columns: “Sample” and “Genotype”.

`mutate(SNP_Type = map_chr(Genotype, classify_snp))`: Uses the `classify_snp` function to classify the Genotype for each row. The result is stored in a new column called `SNP_Type`.

`bind_rows`: Combines the `df_maize_long` and `df_teosinte_long` datasets into one dataframe. The `mutate(Group = “Maize”)` and `mutate(Group = “Teosinte”)` commands add a new column called “Group” to each dataset to label whether the data comes from “Maize” or “Teosinte”.

`df_combined`: This combined dataframe contains data for both Maize and Teosinte, with columns: `SNP_ID`, `Chromosome`, `Position`, `Sample`, `Genotype`, `SNP_Type`, and `Group`.

```
# Function to classify SNPs
classify_snp <- function(genotype) {
  switch(genotype,
    "?/?" = "Missing",
    ifelse(grepl("A|A|C|C|G|G|T|T", genotype), "Homozygous", "Heterozygous"))
}

df_maize_long <- df_maize %>%
  pivot_longer(cols = -c(SNP_ID, Chromosome, Position), # Exclude metadata columns
    names_to = "Group", values_to = "Genotype") %>%

  # Standardize Group names (remove .1, .2 suffixes)
  mutate(Group = str_remove(Group, "\\.\d+$")) %>%

  filter(str_starts(Group, "ZMMIL") | str_starts(Group, "ZMLLR") | str_starts(Group, "ZMMMR")) %>%
  mutate(Species = "Maize", SNP_Type = map_chr(Genotype, classify_snp)) # Assign species and classify
head(df_maize_long)
```

```
## # A tibble: 6 x 7
##   SNP_ID   Chromosome Position Group Genotype Species SNP_Type
##   <chr>    <chr>      <chr>  <chr> <chr>    <chr>  <chr>
## 1 abph1.20 2          27403404 ZMMIL G/G      Maize   Homozygous
## 2 abph1.20 2          27403404 ZMMIL C/C      Maize   Homozygous
## 3 abph1.20 2          27403404 ZMMIL C/C      Maize   Homozygous
## 4 abph1.20 2          27403404 ZMMIL C/C      Maize   Homozygous
## 5 abph1.20 2          27403404 ZMMIL C/C      Maize   Homozygous
## 6 abph1.20 2          27403404 ZMMIL G/G      Maize   Homozygous
```

```
# Convert teosinte data to long format
df_teosinte_long <- df_teosinte %>%
  pivot_longer(cols = -c(SNP_ID, Chromosome, Position), # Exclude metadata columns
    names_to = "Group", values_to = "Genotype") %>%

  # Standardize Group names (remove .1, .2 suffixes)
  mutate(Group = str_remove(Group, "\\.\d+$")) %>%
```



```

  filter(str_starts(Group, "ZMPBA") | str_starts(Group, "ZMPIL") | str_starts(Group, "ZMPJA")) %>%
  mutate(Species = "Teosinte", SNP_Type = map_chr(Genotype, classify_snp)) # Assign species and clas
head(df_teosinte_long)

```

```

## # A tibble: 6 x 7
##   SNP_ID   Chromosome Position Group Genotype Species  SNP_Type
##   <chr>    <chr>      <chr>  <chr> <chr>   <chr>    <chr>
## 1 abph1.20 2          27403404 ZMPBA C/G    Teosinte Heterozygous
## 2 abph1.20 2          27403404 ZMPBA C/G    Teosinte Heterozygous
## 3 abph1.20 2          27403404 ZMPBA G/G    Teosinte Homozygous
## 4 abph1.20 2          27403404 ZMPBA G/G    Teosinte Homozygous
## 5 abph1.20 2          27403404 ZMPBA C/G    Teosinte Heterozygous
## 6 abph1.20 2          27403404 ZMPBA C/G    Teosinte Heterozygous

```

```

# Combine both datasets into one
df_combined <- bind_rows(df_maize_long, df_teosinte_long)
head(df_combined)

```

```

## # A tibble: 6 x 7
##   SNP_ID   Chromosome Position Group Genotype Species  SNP_Type
##   <chr>    <chr>      <chr>  <chr> <chr>   <chr>    <chr>
## 1 abph1.20 2          27403404 ZMMIL G/G    Maize    Homozygous
## 2 abph1.20 2          27403404 ZMMIL C/C    Maize    Homozygous
## 3 abph1.20 2          27403404 ZMMIL C/C    Maize    Homozygous
## 4 abph1.20 2          27403404 ZMMIL C/C    Maize    Homozygous
## 5 abph1.20 2          27403404 ZMMIL C/C    Maize    Homozygous
## 6 abph1.20 2          27403404 ZMMIL G/G    Maize    Homozygous

```

```

# Merge with filtered SNP metadata
df_snp_filtered <- merge(df_combined, by = "SNP_ID")

# Summarize SNP types by species (Maize or Teosinte)
df_snp_summary <- df_snp_filtered %>%
  group_by(Species, SNP_Type, Group) %>%
  summarise(Count = n(), .groups = "drop") %>%
  mutate(Proportion = Count / sum(Count)) # Calculate proportion of SNPs

# Print SNP summary table
print(df_snp_summary)

```

```

## # A tibble: 18 x 5
##   Species  SNP_Type   Group Count Proportion
##   <chr>    <chr>    <chr> <int>    <dbl>
## 1 Maize    Heterozygous ZMMIL   1017  0.000406
## 2 Maize    Heterozygous ZMLLR  206037  0.0823
## 3 Maize    Heterozygous ZMMMR    759  0.000303
## 4 Maize    Homozygous   ZMMIL 259176  0.103
## 5 Maize    Homozygous   ZMLLR 981471  0.392
## 6 Maize    Homozygous   ZMMMR  22854  0.00912
## 7 Maize    Missing      ZMMIL  24877  0.00993
## 8 Maize    Missing      ZMLLR  47140  0.0188

```

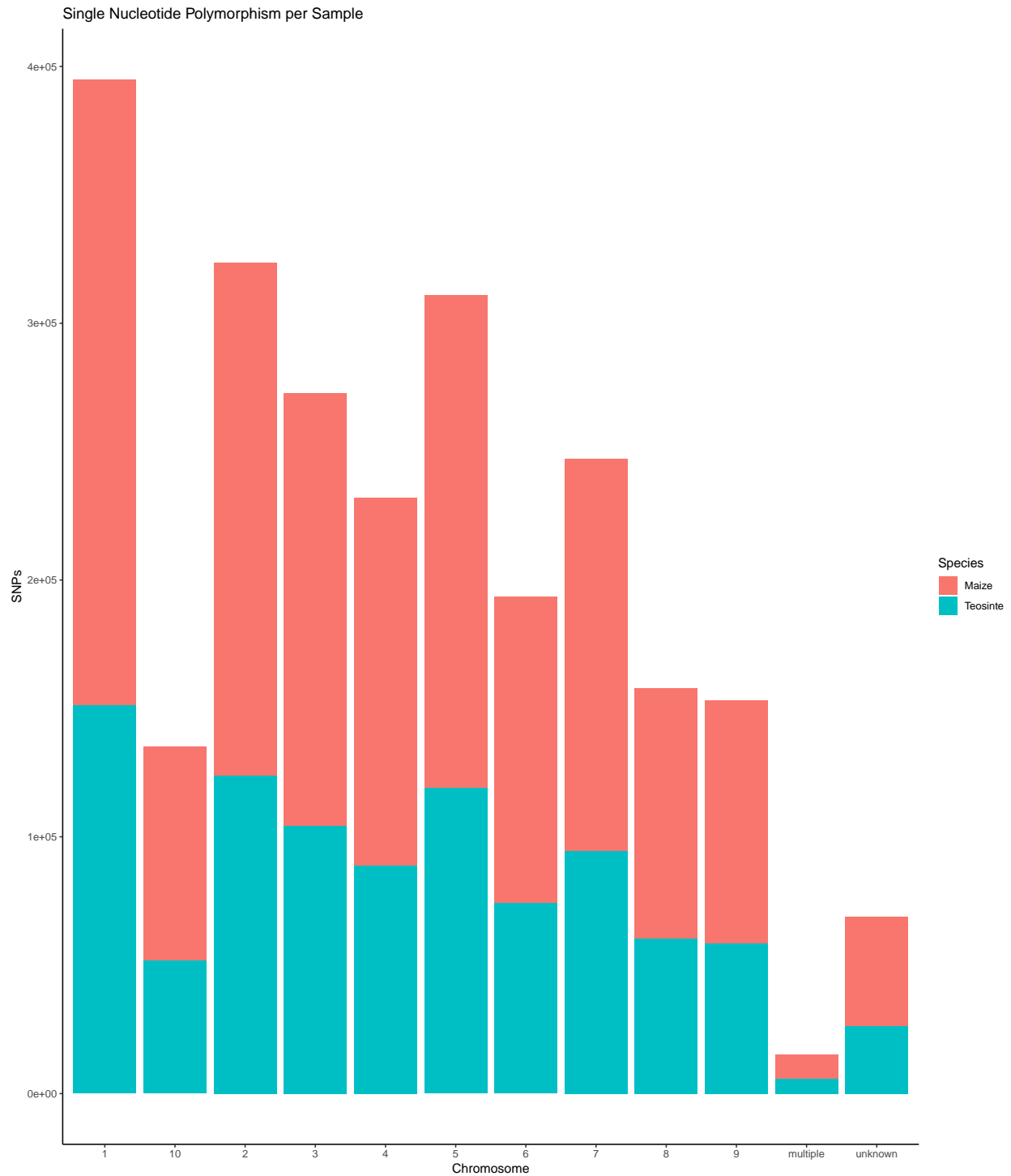
```
## 9 Maize      Missing      ZMMMR    2928    0.00117
## 10 Teosinte Heterozygous ZMPBA 195961 0.0782
## 11 Teosinte Heterozygous ZMPIL    731 0.000292
## 12 Teosinte Heterozygous ZMPJA   5394 0.00215
## 13 Teosinte Homozygous   ZMPBA 648658 0.259
## 14 Teosinte Homozygous   ZMPIL  37841 0.0151
## 15 Teosinte Homozygous   ZMPJA  27393 0.0109
## 16 Teosinte Missing      ZMPBA  40081 0.0160
## 17 Teosinte Missing      ZMPIL   1731 0.000691
## 18 Teosinte Missing      ZMPJA    635 0.000254
```

```
#####
```

```
# Plot SNP distribution by chromosome for maize vs teosinte
by_species_plot <- ggplot(data = df_combined) +
  geom_bar(mapping = aes(x = Chromosome, fill = Species)) +
  xlab("Chromosome") +
  ylab("SNPs") +
  ggtitle("Single Nucleotide Polymorphism per Sample") +

  theme_classic()

# Display the plot
print(by_species_plot)
```



```
### Distribution of Homozygous Genotypes by Group in Maize
# Select only homozygous genotypes
maize_homozygous_snp <- df_maize_long %>%
  filter(SNP_Type == "Homozygous")

# Convert "Group" into a factor with the correct order
maize_homozygous_snp$Group <- factor(
```

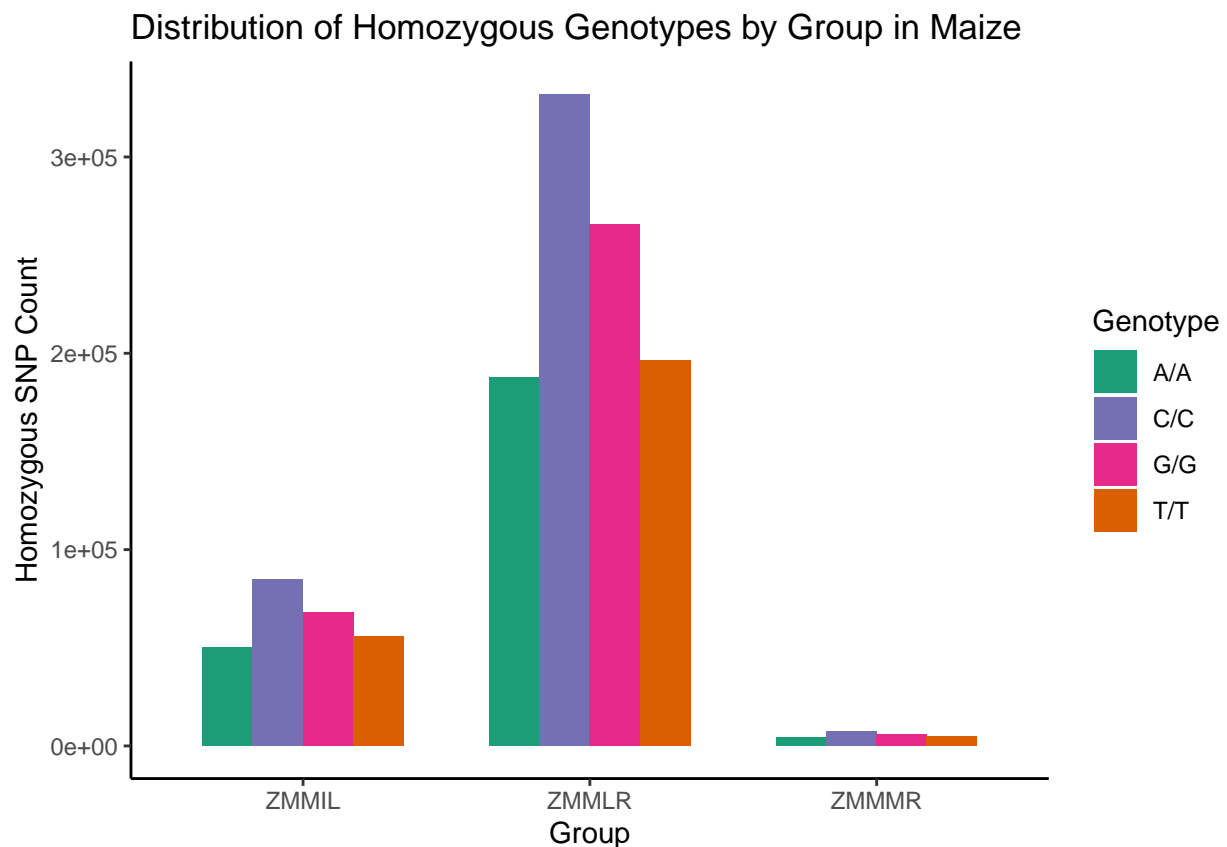
```

maize_homozygous_snp$Group,
levels = c("ZMMIL", "ZMMLR", "ZMMMR")
)

# Exclude NA groups from the plot
maize_homozygous_snp <- maize_homozygous_snp %>%
  filter(!is.na(Group))

# Plot: Homozygous Genotype Distribution by Group
maize_homozygous_plot <- ggplot(maize_homozygous_snp, aes(x = Group, fill = Genotype)) +
  geom_bar(position = "dodge", width = 0.7) +
  scale_fill_manual(
    values = c("A/A" = "#1b9e77", "T/T" = "#d95f02", "C/C" = "#7570b3", "G/G" = "#e7298a"),
    name = "Genotype"
  ) +
  xlab("Group") +
  ylab("Homozygous SNP Count") +
  ggtitle("Distribution of Homozygous Genotypes by Group in Maize") +
  theme_classic()
maize_homozygous_plot

```



```

### Distribution of Homozygous Genotypes by Group in Teosinte
# Select only homozygous genotypes
teosinte_homozygous_snp <- df_teosinte_long %>%
  filter(SNP_Type == "Homozygous")

```

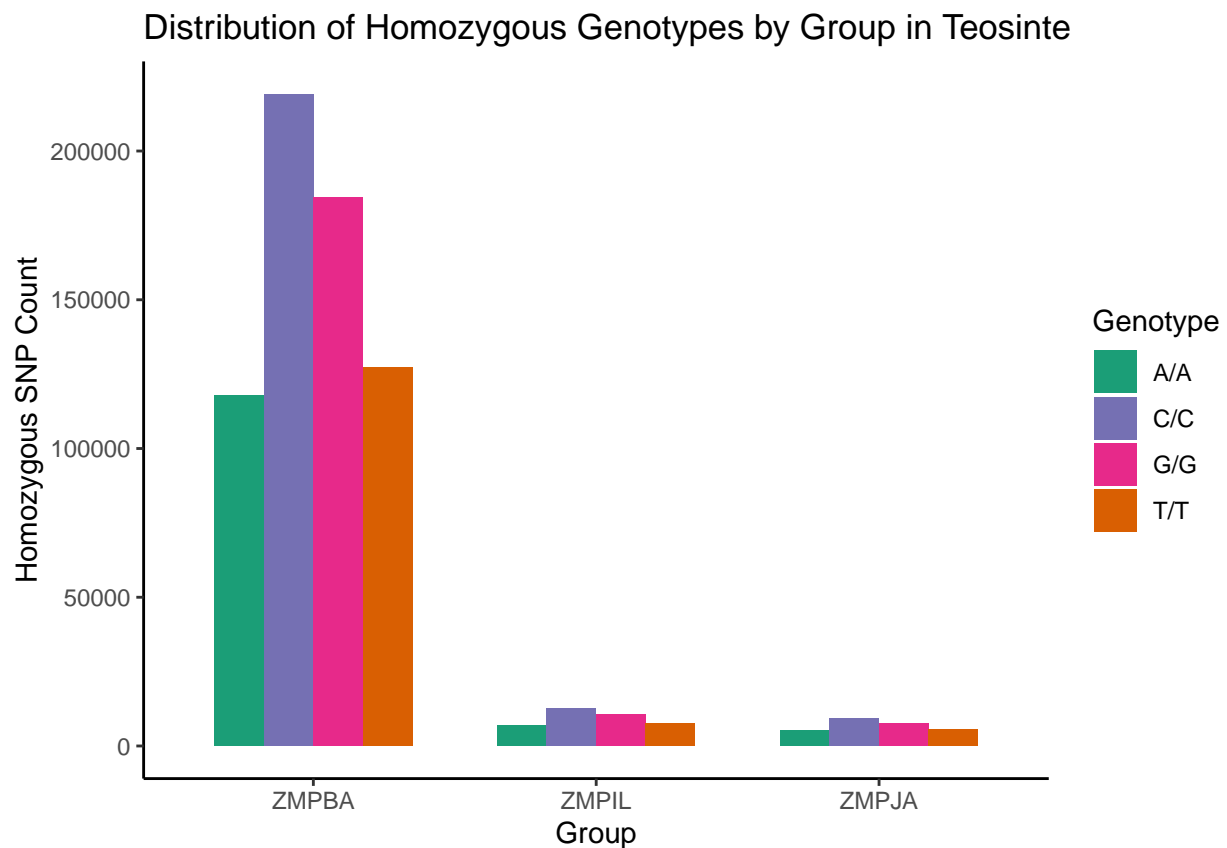
```

# Convert "Group" into a factor with the correct order
teosinte_homozygous_snp$Group <- factor(
  teosinte_homozygous_snp$Group,
  levels = c("ZMPBA", "ZMPIL", "ZMPJA")
)

# Exclude NA groups from the plot
teosinte_homozygous_snp <- teosinte_homozygous_snp %>%
  filter(!is.na(Group))

# Plot: Homozygous Genotype Distribution by Group
teosinte_homozygous_plot <- ggplot(teosinte_homozygous_snp, aes(x = Group, fill = Genotype)) +
  geom_bar(position = "dodge", width = 0.7) +
  scale_fill_manual(
    values = c("A/A" = "#1b9e77", "T/T" = "#d95f02", "C/C" = "#7570b3", "G/G" = "#e7298a"),
    name = "Genotype"
  ) +
  xlab("Group") +
  ylab("Homozygous SNP Count") +
  ggtitle("Distribution of Homozygous Genotypes by Group in Teosinte") +
  theme_classic()
teosinte_homozygous_plot

```



```

# Select only heterozygous genotypes in maize groups
maize_heterozygous_snp <- df_maize_long %>%
  filter(SNP_Type == "Heterozygous")

# Convert "Group" into a factor with the correct order
maize_heterozygous_snp$Group <- factor(
  maize_heterozygous_snp$Group,
  levels = c("ZMMIL", "ZMMLR", "ZMMMR")
)

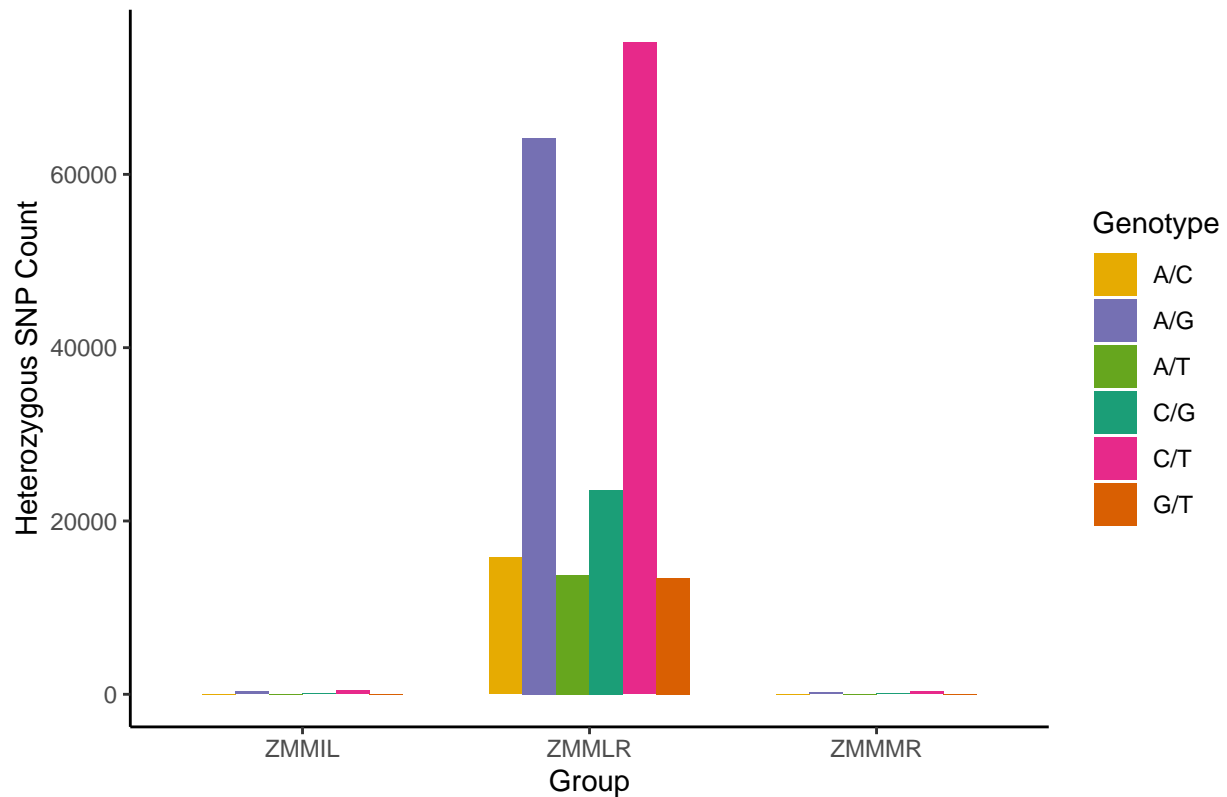
# Exclude NA groups from the plot
maize_heterozygous_snp <- maize_heterozygous_snp %>%
  filter(!is.na(Group))

# Plot: Heterozygous Genotype Distribution by Group
maize_heterozygous_plot <- ggplot(maize_heterozygous_snp, aes(x = Group, fill = Genotype)) +
  geom_bar(position = "dodge", width = 0.7) +
  scale_fill_manual(
    values = c("C/G" = "#1b9e77", "G/T" = "#d95f02", "A/G" = "#7570b3",
              "C/T" = "#e7298a", "A/T" = "#66a61e", "A/C" = "#e6ab02"),
    name = "Genotype"
  ) +
  xlab("Group") +
  ylab("Heterozygous SNP Count") +
  ggtitle("Distribution of Heterozygous Genotypes by Group in Maize") +
  theme_classic()

maize_heterozygous_plot

```

Distribution of Heterozygous Genotypes by Group in Maize



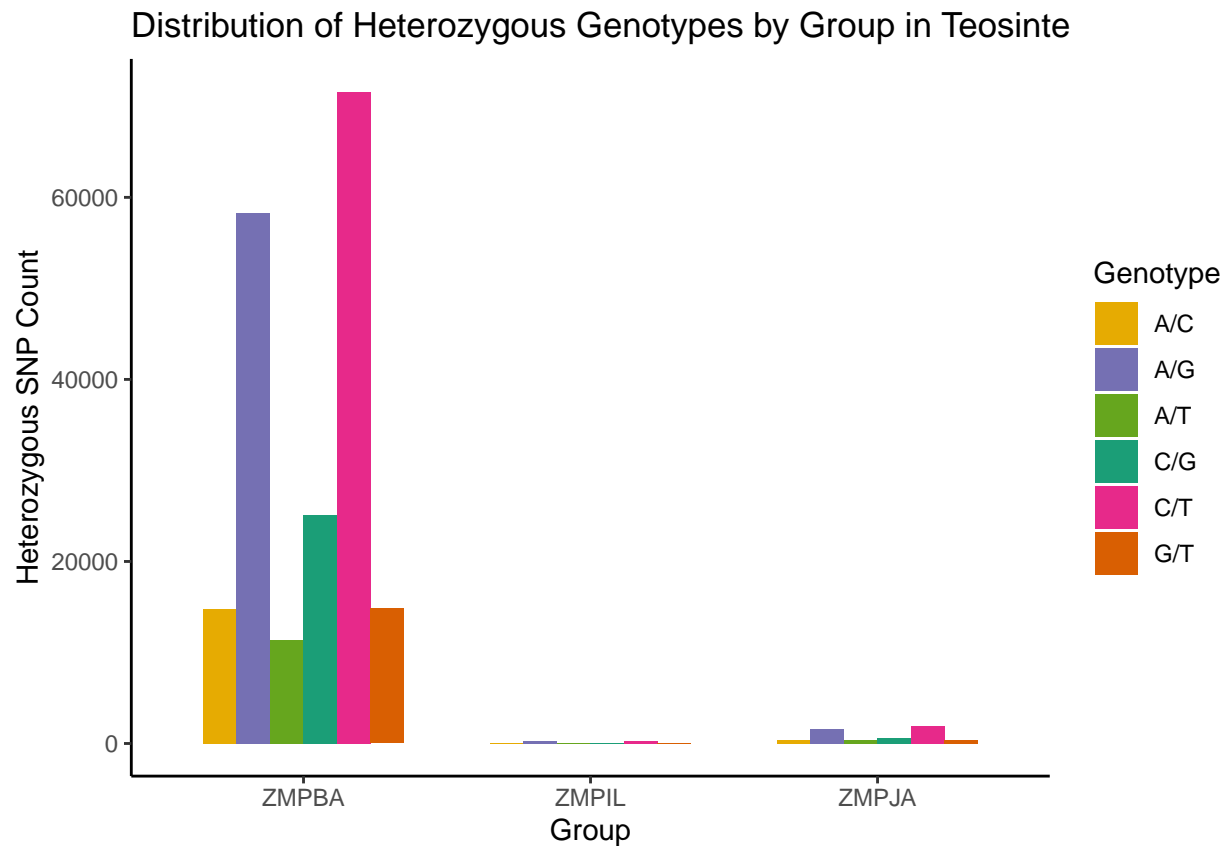
```
# Select only heterozygous genotypes
teosinte_heterozygous_snp <- df_combined %>%
  filter(SNP_Type == "Heterozygous")

# Convert "Group" into a factor with the correct order
teosinte_heterozygous_snp$Group <- factor(
  teosinte_heterozygous_snp$Group,
  levels = c("ZMPBA", "ZMPIL", "ZMPJA")
)

# Exclude NA groups from the plot
teosinte_heterozygous_snp <- teosinte_heterozygous_snp %>%
  filter(!is.na(Group))

# Plot: Heterozygous Genotype Distribution by Group
teosinte_heterozygous_plot <- ggplot(teosinte_heterozygous_snp, aes(x = Group, fill = Genotype)) +
  geom_bar(position = "dodge", width = 0.7) +
  scale_fill_manual(
    values = c("C/G" = "#1b9e77", "G/T" = "#d95f02", "A/G" = "#7570b3",
              "C/T" = "#e7298a", "A/T" = "#66a61e", "A/C" = "#e6ab02"),
    name = "Genotype"
  ) +
  xlab("Group") +
  ylab("Heterozygous SNP Count") +
  ggtitle("Distribution of Heterozygous Genotypes by Group in Teosinte") +
  theme_classic()
```

```
teosinte_heterozygous_plot
```



Count proportions of SNP types per group

`df_long <- bind_rows(maize_long, teosinte_long)`: Combines the long-format maize and teosinte datasets into a single dataframe (`df_long`). `group_by(Group, SNP_Type)`: Groups the data by two columns: `Group` (which differentiates between Maize and Teosinte) and `SNP_Type` (Homozygous, Heterozygous, or Missing).

`summarise(Count = n(), .groups = "drop")`: For each group and SNP type combination, it counts the number of occurrences (i.e., how many times each SNP type appears for a given group). The `.groups = "drop"` argument ensures that the grouping structure is removed after summarizing.

`mutate(Proportion = Count / sum(Count))`: Adds a new column (`Proportion`) that calculates the proportion of each SNP type within each group by dividing the count of each SNP type by the total count within the group.

`print(df_snp_summary)`: Displays the summarized results, showing the count and proportion of each SNP type (Homozygous, Heterozygous, Missing) for both Maize and Teosinte.

```
# Summarize SNP types by group (Maize or Teosinte)
df_snp_summary <- df_combined %>%
```

```
  # Group the data by "Group" (Maize or Teosinte) and "SNP_Type" (Homozygous, Heterozygous, Missing)
  group_by(Species, SNP_Type) %>%
```



```

# Count the number of occurrences for each SNP type in each group
summarise(Count = n(), .groups = "drop") %>%

# Calculate the proportion of each SNP type within each group
mutate(Proportion = Count / sum(Count))

# View the summarized results (SNP counts and proportions)
print(df_snp_summary)

```

```

## # A tibble: 6 x 4
##   Species SNP_Type      Count Proportion
##   <chr>   <chr>      <int>     <dbl>
## 1 Maize   Heterozygous  207813    0.0830
## 2 Maize   Homozygous   1263501    0.504
## 3 Maize   Missing       74945    0.0299
## 4 Teosinte Heterozygous  202086    0.0807
## 5 Teosinte Homozygous   713892    0.285
## 6 Teosinte Missing     42447    0.0169

```

Plot SNP Type distribution per Chromosome

`ggplot(df_combined, aes(x = Chromosome, fill = SNP_Type))`: Initializes the plot using `df_combined`, with Chromosome on the x-axis and different colors for each SNP_Type (e.g., Homozygous, Heterozygous, Missing).

`geom_bar(position = "fill")`: Creates a bar chart where the height of each bar represents the proportion of each SNP type, not the count, scaling the bars to show percentages.

`facet_wrap(~ Species)`: Creates separate subplots for each group (Maize and Teosinte), showing the SNP distribution across chromosomes for each group.

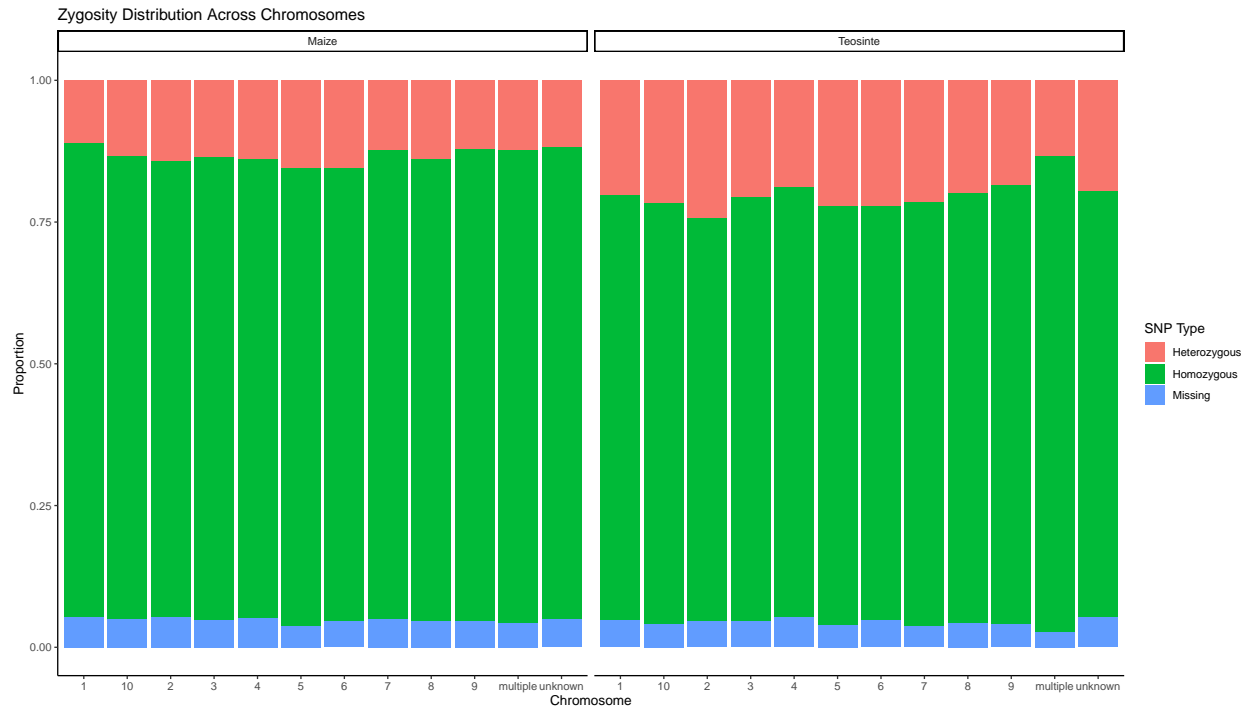
`labs(...)`: Adds titles and axis labels: Title: "Zygoty Distribution Across Chromosomes", X-axis: "Chromosome" Y-axis: "Proportion", Legend: "SNP Type" `theme_classic()`: Applies a clean, minimalist theme to the plot, removing gridlines for a simpler appearance.

This creates a clear, proportion-based bar chart of SNP distribution across chromosomes for both Maize and Teosinte.

```

ggplot(df_combined, aes(x = Chromosome, fill = SNP_Type)) +
  geom_bar(position = "fill") + # "fill" makes proportions instead of counts
  facet_wrap(~ Species) + # Separate plots for Maize & Teosinte
  labs(title = "Zygoty Distribution Across Chromosomes",
       x = "Chromosome", y = "Proportion",
       fill = "SNP Type") +
  theme_classic()

```

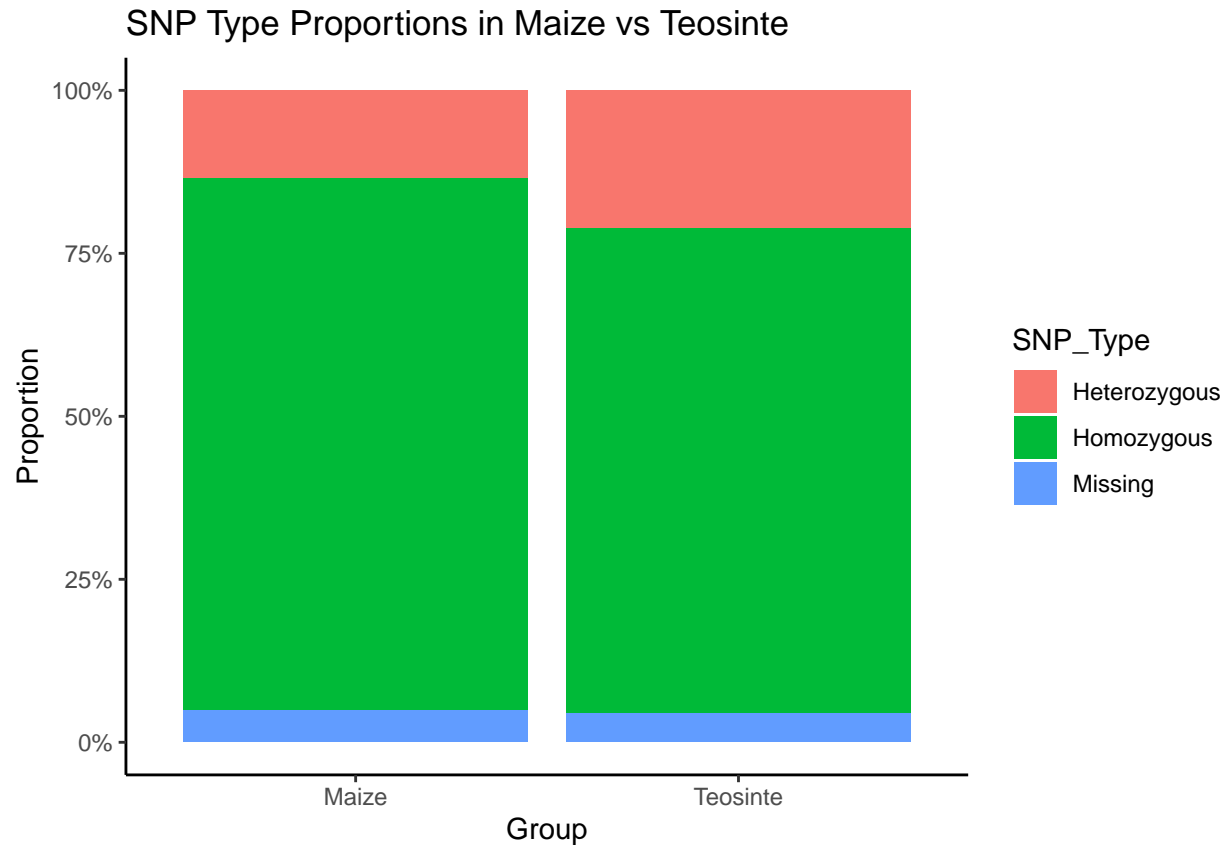


#Proportional Bar Plot (SNP Distribution by Group) Shows the proportions of SNP types (Homozygous, Heterozygous) in Maize vs Teosinte.

Uses a stacked bar chart where the height represents the proportion of each SNP type.

Y-axis labels are formatted as percentages.

```
#Bar Plot - SNP Distribution by Group
ggplot(df_snp_summary, aes(x = Species, y = Proportion, fill = SNP_Type)) +
  geom_bar(stat = "identity", position = "fill") + # Stacked proportional bar chart
  labs(title = "SNP Type Proportions in Maize vs Teosinte",
        x = "Group", y = "Proportion") +
  scale_y_continuous(labels = scales::percent) +
  theme_classic()
```

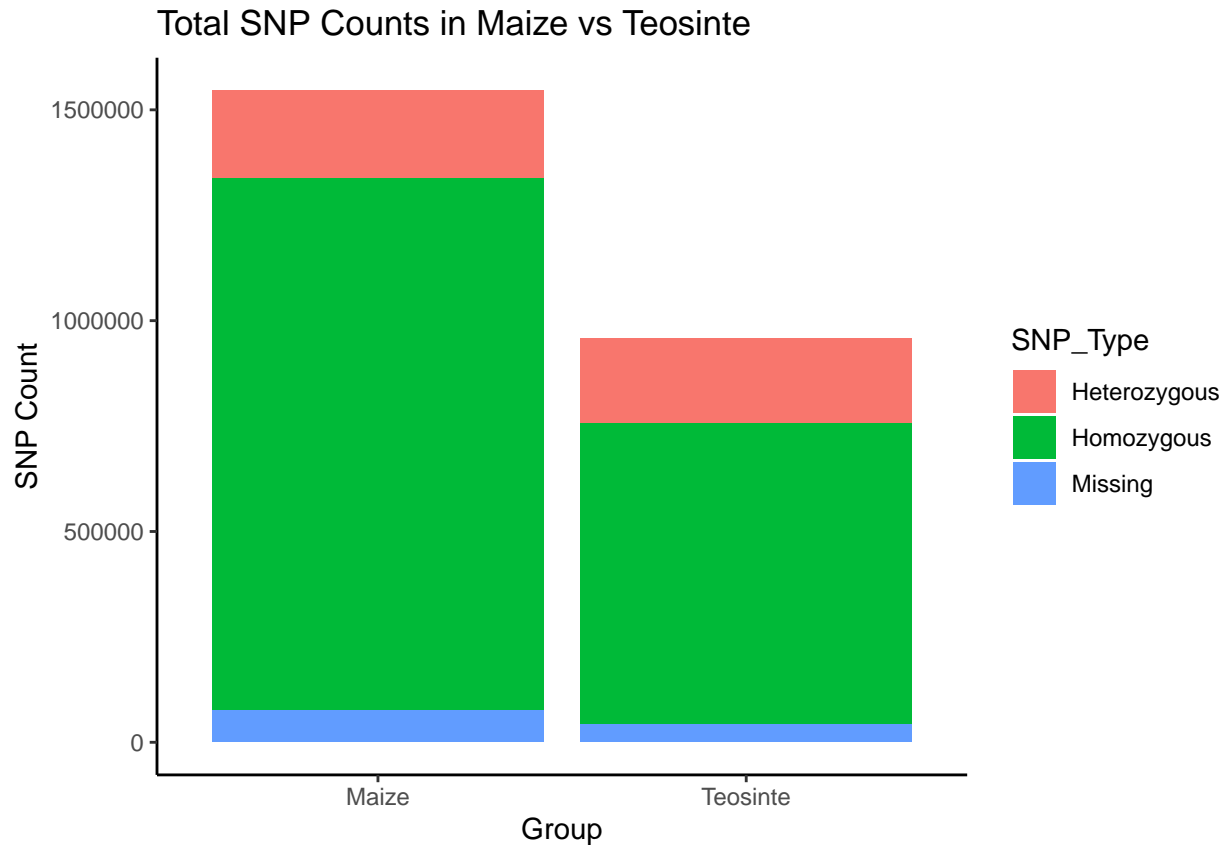


Stacked Bar Plot (Absolute SNP Counts):

Displays the total SNP counts for each SNP type in Maize vs Teosinte.

Uses a stacked bar chart where the total height represents the sum of SNP counts, segmented by SNP type.

```
#Stacked Bar Plot - Absolute SNP Counts
ggplot(df_snp_summary, aes(x = Species, y = Count, fill = SNP_Type)) +
  geom_bar(stat = "identity", position = "stack") + # Stacked bar chart
  labs(title = "Total SNP Counts in Maize vs Teosinte",
        x = "Group", y = "SNP Count") +
  theme_classic()
```



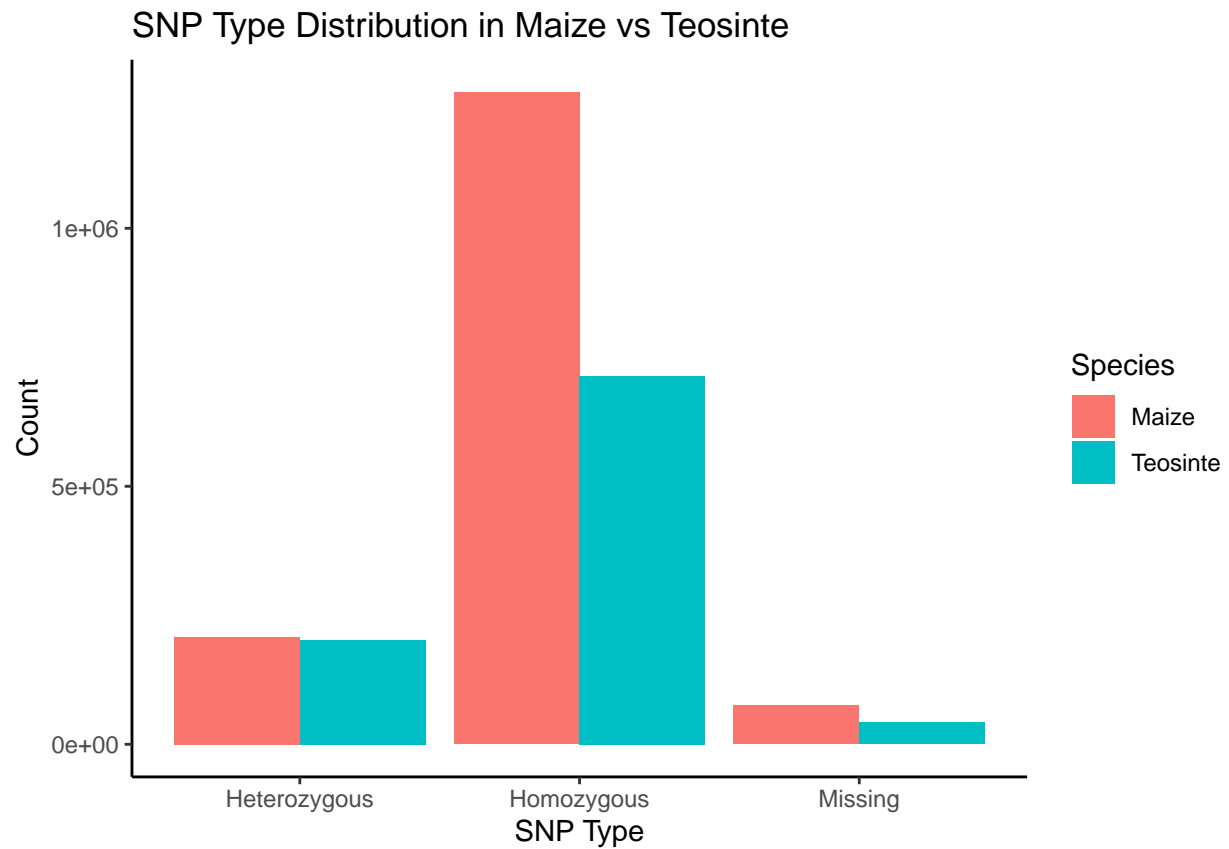
#SNP Distribution Bar Plot per Sample:

Displays the count of each SNP Type (Homozygous, Heterozygous, etc.) in Maize vs Teosinte. Uses a dodge position for a side-by-side comparison of SNP types across the two groups (Maize and Teosinte). Labels include the title: "SNP Type Distribution in Maize vs Teosinte", X-axis: "SNP Type", and Y-axis: "Count".

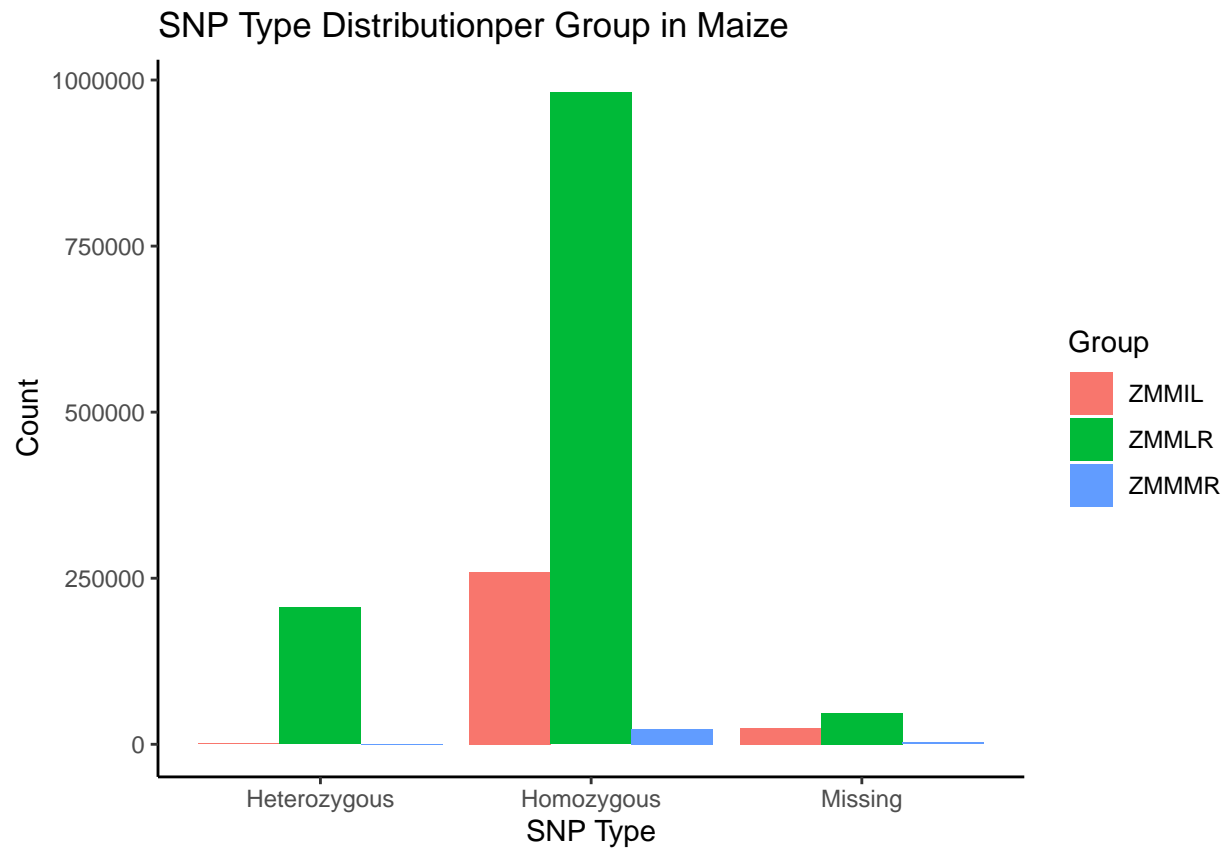
The plot uses a clean theme (theme_classic()).

```
# Bar Plot - SNP Distribution per Species
ggplot(df_combined, aes(x = SNP_Type, y = ..count.., fill = Species)) +
  geom_bar(position = "dodge") +
  labs(title = "SNP Type Distribution in Maize vs Teosinte",
       x = "SNP Type", y = "Count") +
  theme_classic()
```

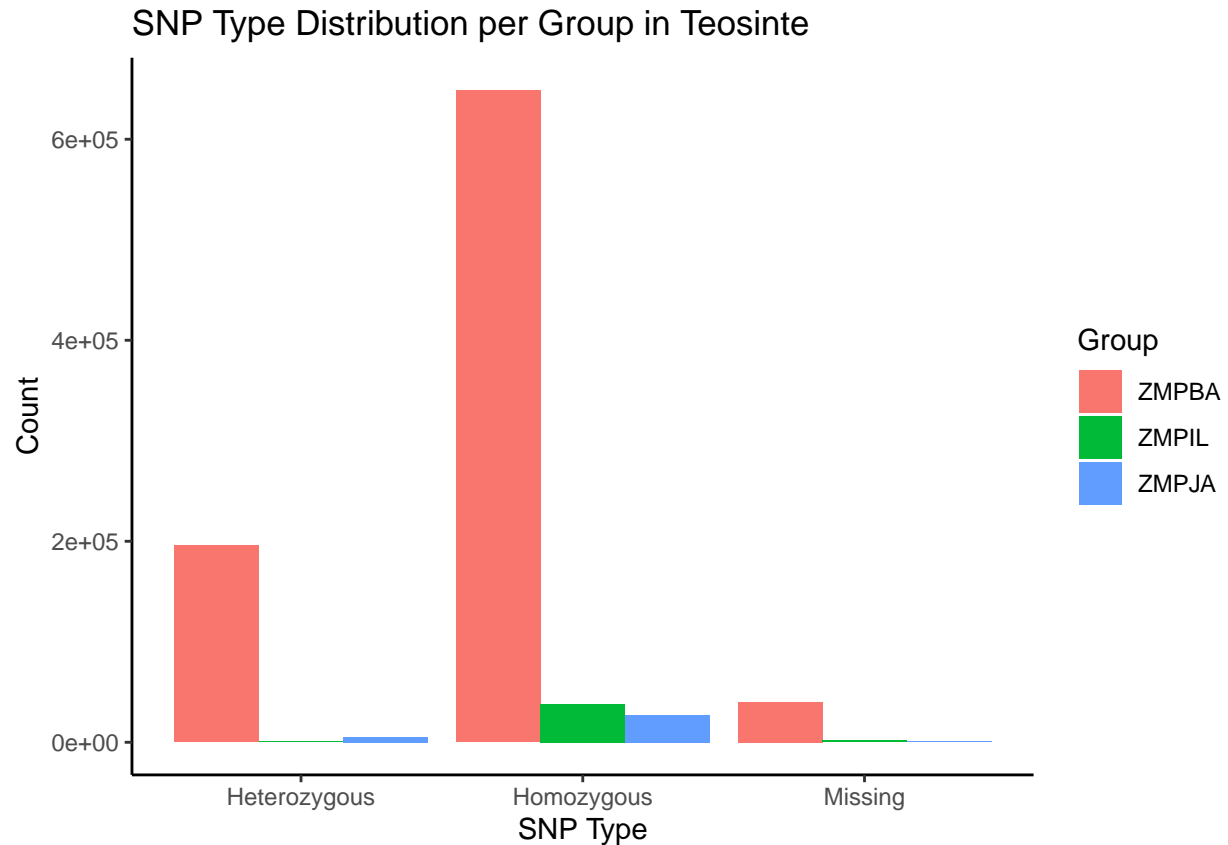
```
## Warning: The dot-dot notation ('..count..') was deprecated in ggplot2 3.4.0.
## i Please use 'after_stat(count)' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```



```
# Bar Plot - SNP Distribution per Group in Maize
ggplot(df_maize_long, aes(x = SNP_Type, y = ..count.., fill = Group)) +
  geom_bar(position = "dodge") +
  labs(title = "SNP Type Distributionper Group in Maize",
       x = "SNP Type", y = "Count") +
  theme_classic()
```



```
# Bar Plot - SNP Distribution per Group in Teosinte
ggplot(df_teosinte_long, aes(x = SNP_Type, y = ..count.., fill = Group)) +
  geom_bar(position = "dodge") +
  labs(title = "SNP Type Distribution per Group in Teosinte",
       x = "SNP Type", y = "Count") +
  theme_classic()
```



```
# Heatmap of SNP distribution across chromosomes for Maize and Teosinte
ggplot(snp_counts, aes(x = Chromosome, y = Species, fill = SNP_Count)) +

  # Create a heatmap using geom_tile()
  geom_tile(color = "white ") +

  # Add labels to the plot
  labs(title = "SNP Distribution Heatmap Across Chromosomes",
        x = "Chromosome", y = "Species", fill = "SNP Count") +

  # Use a gradient color scale to represent SNP counts
  scale_fill_gradient(low = "yellow", high = "blue") +

  # Ensure chromosome labels appear in the correct order
  scale_x_discrete(limits = c("1", "2", "3", "4", "5", "6", "7", "8", "9", "10", "multiple", "unknown"))

# Use a clean and simple theme
theme_classic()
```

