

# Task Switching Replication

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## Read in and Check Raw Data

```
#Import current data
task_switching_raw <- read.csv(paste0(workingdir, "/task-switching-replication-recoded-2.csv"))
head(task_switching_raw)
```

	participant	session	condition	trialType	posture	blockNum	trialNum	switchTrialType
## 1	1	1	1	experiment	standing	1	1	buffer
## 2	1	1	1	experiment	standing	1	2	noswitch
## 3	1	1	1	experiment	standing	1	3	switch
## 4	1	1	1	experiment	standing	1	4	noswitch
## 5	1	1	1	experiment	standing	1	5	noswitch
## 6	1	1	1	experiment	standing	1	6	noswitch

	congruentTrialType	cueType	shapeType	shapeColor	response	correctResponse	correct	reactionTime
## 1	incongruent	solid	square	blue	right	left	no	0.9088130
## 2	incongruent	solid	square	blue	left	left	yes	0.5947349
## 3	incongruent	dashed	square	blue	right	right	yes	0.7084870
## 4	incongruent	dashed	square	blue	right	right	yes	0.5995200
## 5	congruent	dashed	square	yellow	right	right	yes	0.4399409
## 6	congruent	dashed	square	yellow	right	right	yes	0.3847258

	date	utcTime
## 1	2021-11-10 10:22:00	1636561737
## 2	2021-11-10 10:22:00	1636561744
## 3	2021-11-10 10:22:00	1636561746
## 4	2021-11-10 10:22:00	1636561748
## 5	2021-11-10 10:22:00	1636561750
## 6	2021-11-10 10:22:00	1636561752

```
#does every person have 392 trials?
ntrials_sub <- task_switching_raw %>%
  group_by(participant) %>%
  summarize(ntrials = n()) %>%
  pull(ntrials)

all(ntrials_sub == 392)

## [1] TRUE

#does every block start with a buffer and have 49 trials?
task_switching_raw <- task_switching_raw %>%
  mutate(condblock = paste0(posture, blockNum))

blocktrials <- task_switching_raw %>%
```

```

group_by(participant, condblock) %>%
  summarize(ntrials = n(), firsttrial = first(switchTrialType))

## `summarise()` has grouped output by 'participant'. You can override using the `.groups` argument.
all(blocktrials$ntrials == 49)

## [1] TRUE

all(blocktrials$firsttrial == "buffer")

## [1] TRUE

```

## Clean Data

```

#Drop buffer trials
task_switching_raw2 <- task_switching_raw %>%
  filter(switchTrialType != "buffer")

#Recode Correct to 1 and Incorrect to 0
task_switching_raw2$correct_bin <- recode(task_switching_raw2$correct,
                                         "no" = 0,
                                         "yes" = 1)

#Calc overall acc by participant
ts_overall_acc <- task_switching_raw2 %>%
  group_by(participant) %>%
  summarize(Accuracy = mean(correct_bin))

#find participants with less than 80% accuracy
#2, 8, 15, 44, 49, 51
low_acc_subs <- ts_overall_acc %>% filter(Accuracy < 0.80) %>%
  pull(participant)

```

```

task_switching_raw3 <- task_switching_raw2 %>%
  filter(!(participant %in% low_acc_subs))

#Calc mean Acc by participant and conditions (posture, con, switch)
#Narrow format
ts_acc_mean <- task_switching_raw3 %>%
  group_by(participant,
            posture,
            congruentTrialType,
            switchTrialType) %>%
  summarize(Accuracy = mean(correct_bin))

```

```

## `summarise()` has grouped output by 'participant', 'posture', 'congruentTrialType'. You can override

#Convert data to wide format (for statview/SPSS/etc)
ts_acc_mean_wide <- ts_acc_mean %>%
  pivot_wider(names_from = c(posture,
                              congruentTrialType,
                              switchTrialType),
              values_from = Accuracy)

```

```

write.csv(ts_acc_mean_wide, file = "new_recoded_for_statview.csv", row.names = F)

ts_acc_mean <- data.frame(ts_acc_mean)
ts_acc_mean$posture <- as.factor(ts_acc_mean$posture)
ts_acc_mean$participant <- as.factor(ts_acc_mean$participant)
ts_acc_mean$congruentTrialType <- as.factor(ts_acc_mean$congruentTrialType)
ts_acc_mean$switchTrialType <- as.factor(ts_acc_mean$switchTrialType)
str(ts_acc_mean)

## 'data.frame': 408 obs. of 5 variables:
## $ participant : Factor w/ 51 levels "1","3","4","5",...: 1 1 1 1 1 1 1 1 2 2 ...
## $ posture : Factor w/ 2 levels "sitting","standing": 1 1 1 1 2 2 2 2 1 1 ...
## $ congruentTrialType: Factor w/ 2 levels "congruent","incongruent": 1 1 2 2 1 1 2 2 1 1 ...
## $ switchTrialType : Factor w/ 2 levels "noswitch","switch": 1 2 1 2 1 2 1 2 1 2 ...
## $ Accuracy : num 0.96 0.978 0.957 0.88 0.981 ...

#Total N = 51 (6 dropped for acc < 80%)
length(unique(ts_acc_mean$participant))

## [1] 51

```

## Summarize Demographics

```

demo_raw <- read.csv(paste0(workingdir, "/Task Switching_February 24, 2022_13.05.csv"), skip = 1) %>%
  slice(-1) %>%
  select(-c(Response.Type, IP.Address, Recipient.Last.Name:Distribution.Channel))
colnames(demo_raw)[10:15] <- c("Gender.Pick", "Gender.Text", "Age", "Race.Pick", "Race.Text", "Eng.First")

dim(demo_raw)

## [1] 59 15

#59 records
#first two are test data
# need to match up the 6 dropped participants from behavioral data
demo_df <- demo_raw %>%
  filter(!(X %in% c("test", low_acc_subs)))
dim(demo_df)

## [1] 51 15

demo_df <- demo_df %>%
  mutate(Gender.New = ifelse(Gender.Pick %in% c("Man", "Woman"), Gender.Pick, Gender.Text),
         Eng.First = toupper(Eng.First))

#gender breakdown
gender_table <- demo_df %>%
  group_by(Gender.New) %>%
  summarize(n = n())
gender_table

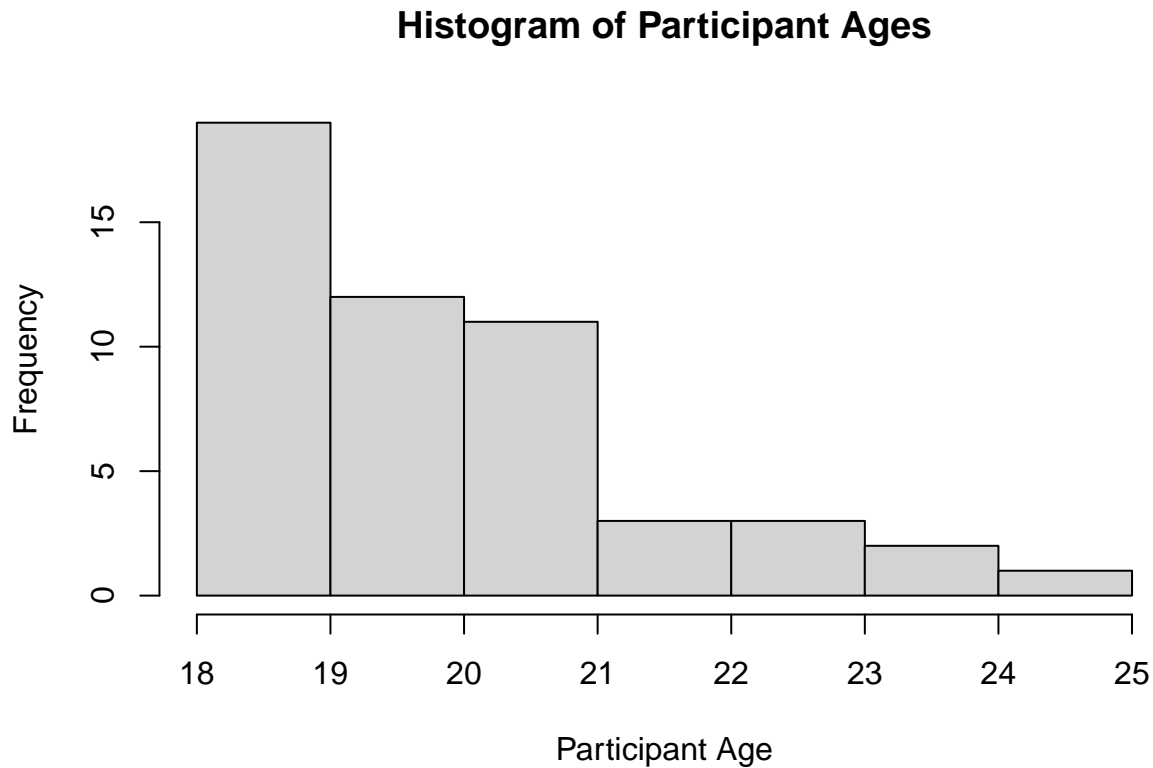
## # A tibble: 3 x 2
##   Gender.New      n
##   <chr>         <int>
## 1 Man           23

```

```
## 2 non binaary      1
## 3 Woman            27
```

```
#age breakdown
```

```
hist(as.numeric(demo_df$Age),
     main = "Histogram of Participant Ages",
     xlab = "Participant Age")
```



```
age_table <- demo_df %>%
  group_by(Age) %>%
  summarize(n = n())
age_table
```

```
## # A tibble: 8 x 2
##   Age      n
##   <chr> <int>
## 1 18      9
## 2 19     10
## 3 20     12
## 4 21     11
## 5 22      3
## 6 23      3
## 7 24      2
## 8 25      1
```

```
#race breakdown
```

```
race_table <- demo_df %>%
  group_by(Race.Pick) %>%
```

```

  summarize(n = n()) %>%
  arrange(desc(n))
race_table

```

```

## # A tibble: 6 x 2
##   Race.Pick          n
##   <chr>          <int>
## 1 White /European American      22
## 2 Black / African American      11
## 3 Hispanic/Latino/Latina/Latinx  11
## 4 Asian /South Pacific Islander   3
## 5 Central Asian /Indian /Pakistani 3
## 6 Native American / American Indian 1

```

```

#language breakdown
lang_table <- demo_df %>%
  group_by(Eng.First) %>%
  summarize(n=n())
lang_table

```

```

## # A tibble: 2 x 2
##   Eng.First    n
##   <chr>    <int>
## 1 NO         8
## 2 YES        43

```

## Plots and Analyses

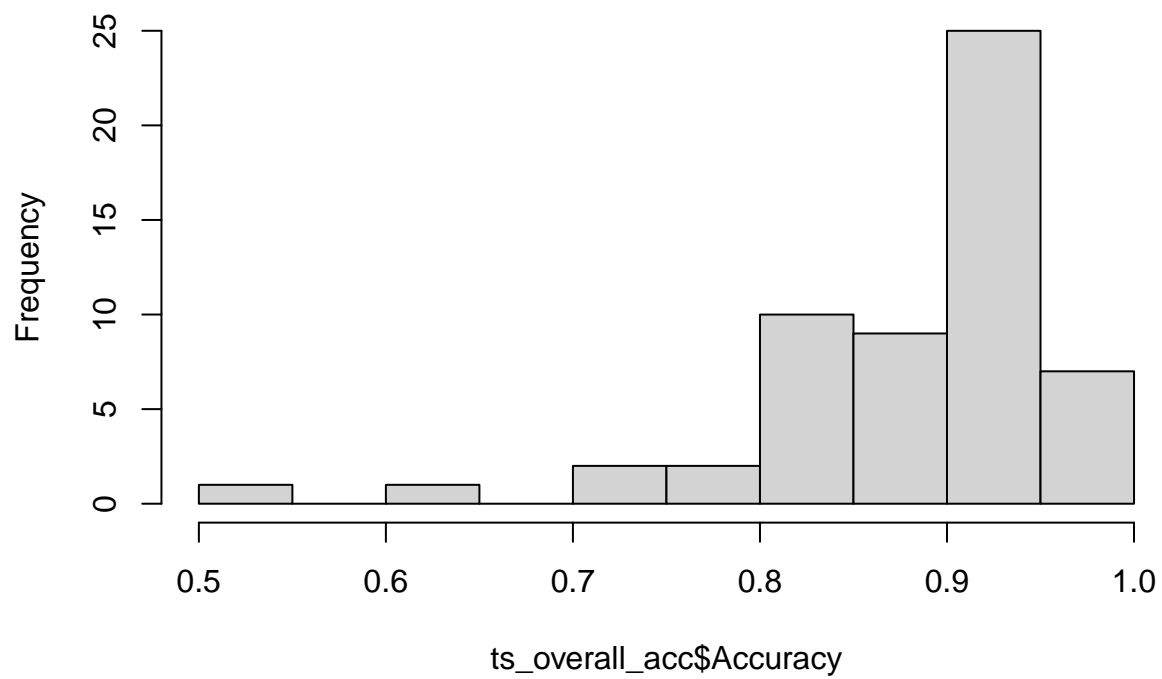
### Accuracy

```

#Accuracy by participant, before drops
hist(ts_overall_acc$Accuracy)

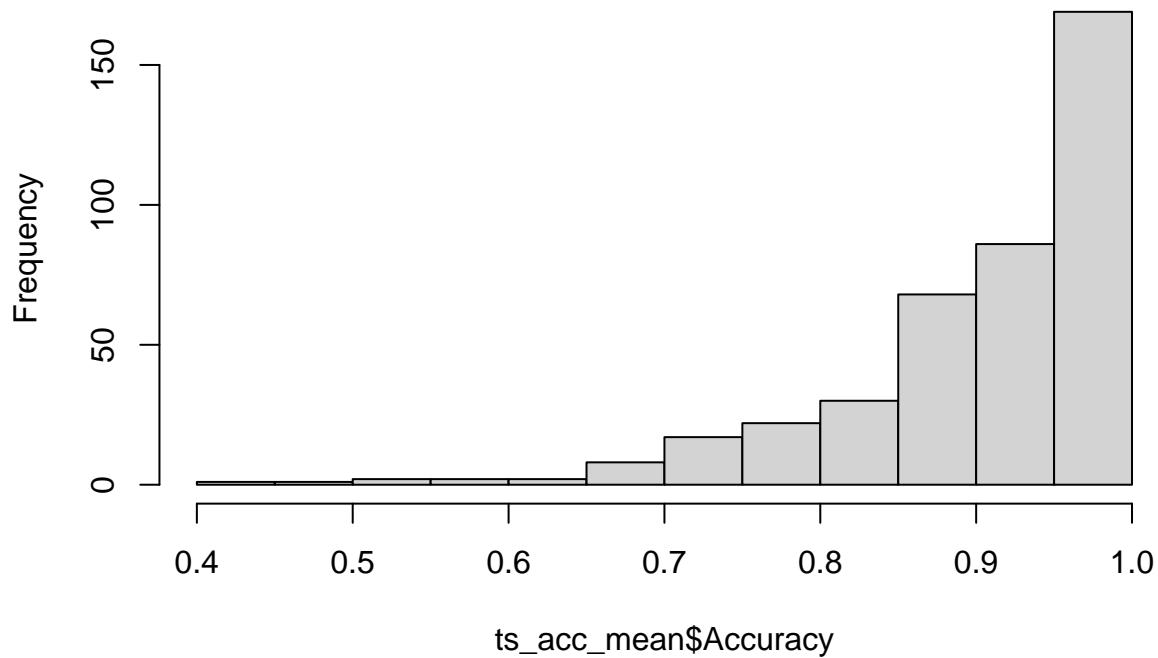
```

**Histogram of ts\_overall\_acc\$Accuracy**



```
#Accuracy for all cells  
hist(ts_acc_mean$Accuracy)
```

## Histogram of ts\_acc\_mean\$Accuracy



```
exp1_anova <- ezANOVA(ts_acc_mean,
  dv = Accuracy,
  wid = participant,
  within = .(posture, congruentTrialType, switchTrialType),
  type = 3,
  detailed = TRUE,
  return_aov = T
)

exp1_stats <- ezStats(ts_acc_mean,
  dv = Accuracy,
  wid = participant,
  within = .(posture, congruentTrialType, switchTrialType),
  type = 3
)

write.csv(exp1_stats[, -7], file = "Task_Switching_Descriptives_ACC.csv", row.names = F)

#data.frame(exp1_anova$ANOVA)
output_acc <- aovEffectSize(exp1_anova, effectSize = "pes")
output_acc <- data.frame(output_acc$ANOVA)
write.csv(output_acc, "Task_switching_ANOVA_acc.csv")
output_acc
```

	Effect	DFn	DFd	SSn	SSd	F
## 1	(Intercept)	1	50	3.331616e+02	0.84392719	1.973876e+04
## 2	posture	1	50	9.386943e-03	0.44204149	1.061772e+00

```
## 3          congruentTrialType 1 50 8.577579e-01 0.43034136 9.966017e+01
## 4          switchTrialType 1 50 2.633377e-01 0.14305407 9.204131e+01
## 5      posture:congruentTrialType 1 50 1.038526e-04 0.20920092 2.482125e-02
## 6      posture:switchTrialType 1 50 1.767731e-03 0.12000107 7.365482e-01
## 7      congruentTrialType:switchTrialType 1 50 8.851220e-02 0.07573624 5.843451e+01
## 8 posture:congruentTrialType:switchTrialType 1 50 2.360230e-03 0.09401427 1.255251e+00
##          p p..05          pes
## 1 1.301952e-66 * 0.9974733137
## 2 3.077703e-01 0.0207938669
## 3 1.702444e-13 * 0.6659097700
## 4 6.367282e-13 * 0.6479897331
## 5 8.754478e-01 0.0004961787
## 6 3.948644e-01 0.0145171124
## 7 5.925302e-10 * 0.5388921804
## 8 2.679047e-01 0.0244901935
```

```
#Calculate confidence interval: PES for posture x switch/condition interaction
#using ANOVA results (partial eta-squared)
interaction_effect_CI <-
get.ci.partial.eta.squared(exp1_anova$ANOVA$F[6],
                           exp1_anova$ANOVA$DFn[6],
                           exp1_anova$ANOVA$DFd[6],
                           conf.level = 0.90) #90% CI is the convention for PES
interaction_effect_CI
```

```
## $LL
## [1] 0
##
## $UL
## [1] 0.1073579
```

```
# This almost works for getting CIs around all effects....
test.model <-
# aov_car(Accuracy ~
#         posture*congruentTrialType*switchTrialType +
#         Error(participant/posture*congruentTrialType *switchTrialType),
#         ts_acc_mean, anova_table = (es = "pes"),
#         return = "aov",
#         include_aov = T,
#         type = 3)
# # test.model
# anova_stats(test.model)

#The CIs around effect sizes are calculated using Type I sums of squares?
#' ## Type of Sums of Squares
#' The sums of squares (or *F* statistics) used for the computation of the
#' effect sizes is based on those returned by `anova(model)` (whatever those may
#' be - for `aov` and `aovlist` these are *type-1* sums of squares; for
#' `lmerMod` (and `lmerModLmerTest`) these are *type-3* sums of squares)
test.pes<-
# effectsize::eta_squared(test.model,
#                           alternative = "two.sided",
#                           ci = 0.90,
#                           partial = T,
#                           verbose = T)
```



```
#
#https://mran.microsoft.com/snapshot/2018-06-30/web/packages/sjstats/vignettes/anova-statistics.html

#Below function won't work with an afex or aov object for an rm ANOVA
#car::Anova(..., type = 3)

congruent.labs <- c("Congruent", "Incongruent")
names(congruent.labs) <- c("1", "2")

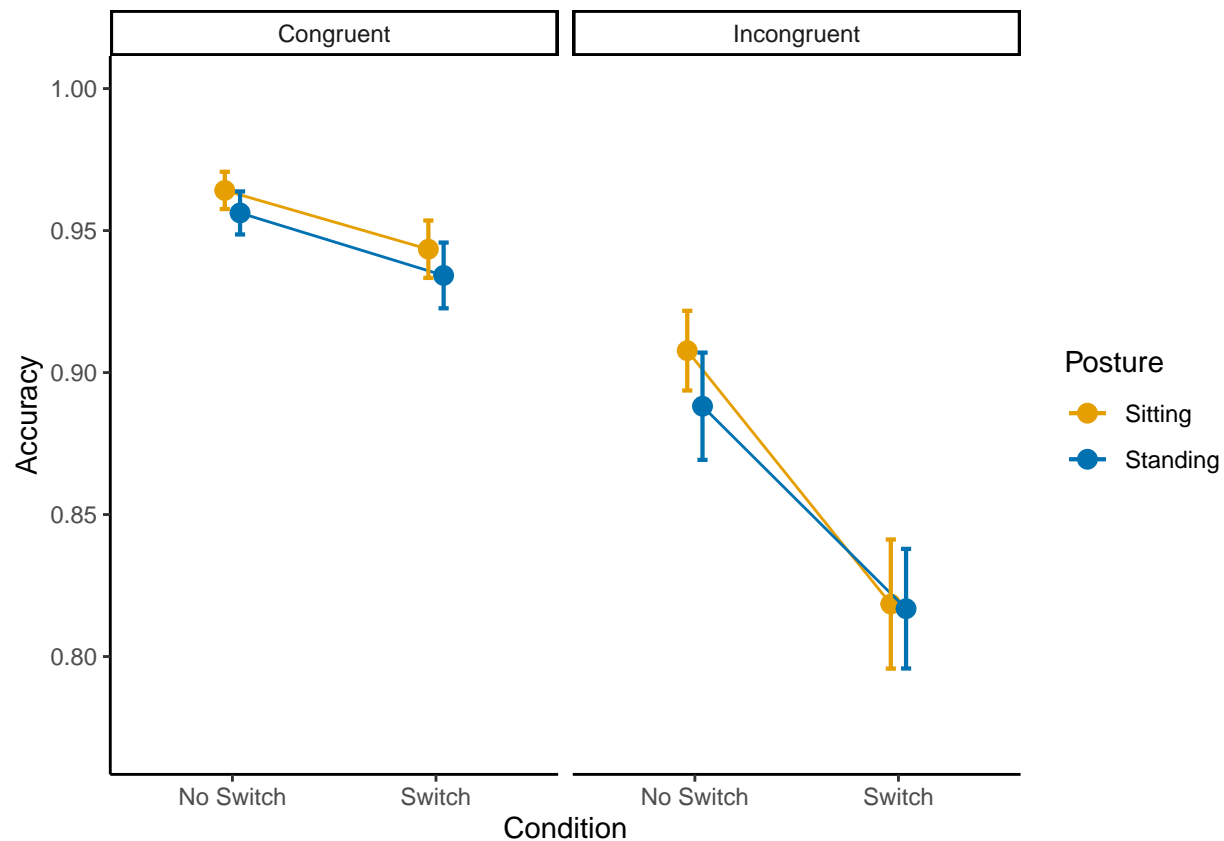
#make plot like Smith et al's
acc_plot <-
superbPlot(ts_acc_mean_wide,
           WSFactors = c("Condition(2)", "Congruent(2)", "Posture(2)"),
           variables = colnames(ts_acc_mean_wide)[2:9],
           errorbar = "SE", #Tempted to change to CI, should stay SE to be consistent with SMith
           plotStyle = "line",
           factorOrder = c("Condition","Posture","Congruent"),
           adjustments = list(purpose = "difference"))+
theme_classic() +
ylim(0.77, 1) + #Trying to make ylim same as the Smith w/o cutting off error bars
facet_wrap(vars(Congruent), labeller = labeller(Congruent = congruent.labs)) +
scale_x_discrete(labels=c("1" = "No Switch", "2" = "Switch"))+
scale_color_manual(values=c("#E69F00", "#0072B2"),
                   labels = c("Sitting", "Standing")) +
labs(y = "Accuracy")
```

## superb::FYI: Here is how the within-subject variables are understood:

```
## Condition Congruent Posture variable
##      1      1      1 sitting_congruent_noswitch
##      2      1      1 sitting_congruent_switch
##      1      2      1 sitting_incongruent_noswitch
##      2      2      1 sitting_incongruent_switch
##      1      1      2 standing_congruent_noswitch
##      2      1      2 standing_congruent_switch
##      1      2      2 standing_incongruent_noswitch
##      2      2      2 standing_incongruent_switch
```

```
ggsave(acc_plot,
       file = "plots/acc_plot.pdf",
       units = "in",
       width = 6.62,
       height = 5.50,
       dpi = 600)
```

```
acc_plot
```



### RT for correct trials only

```
#look at reaction time for correct trials?
ts_correct_only <- task_switching_raw3 %>%
  filter(correct_bin == 1)
```

```
ts_rt_mean <- ts_correct_only %>%
  group_by(participant,
            posture,
            congruentTrialType,
            switchTrialType) %>%
  summarize(mean_rt = mean(reactionTime))
```

## `summarise()` has grouped output by 'participant', 'posture', 'congruentTrialType'. You can override

```
#Convert data to wide format
```

```
ts_rt_mean_wide <- ts_rt_mean %>%
  pivot_wider(names_from = c(posture,
                              congruentTrialType,
                              switchTrialType),
              values_from = mean_rt)
```

```
RT_plot <-
  superbPlot(ts_rt_mean_wide,
             WSFactors = c("Condition(2)", "Congruent(2)", "Posture(2)"),
             variables = colnames(ts_acc_mean_wide)[2:9],
```

```

    errorbar = "SE",
    plotStyle = "line",
    factorOrder = c("Condition", "Posture", "Congruent"),
    adjustments = list(purpose = "difference"))+
theme_classic()+
facet_wrap(vars(Congruent), labeller = labeller(Congruent = congruent.labs)) +
scale_x_discrete(labels=c("1" = "No Switch", "2" = "Switch"))+
scale_color_manual(values=c("#E69F00", "#0072B2"), labels = c("Sitting", "Standing")) +
labs(y = "Reaction Time (ms)")

```

## superb::FYI: Here is how the within-subject variables are understood:

```

## Condition Congruent Posture variable
##      1      1      1 sitting_congruent_noswitch
##      2      1      1 sitting_congruent_switch
##      1      2      1 sitting_incongruent_noswitch
##      2      2      1 sitting_incongruent_switch
##      1      1      2 standing_congruent_noswitch
##      2      1      2 standing_congruent_switch
##      1      2      2 standing_incongruent_noswitch
##      2      2      2 standing_incongruent_switch

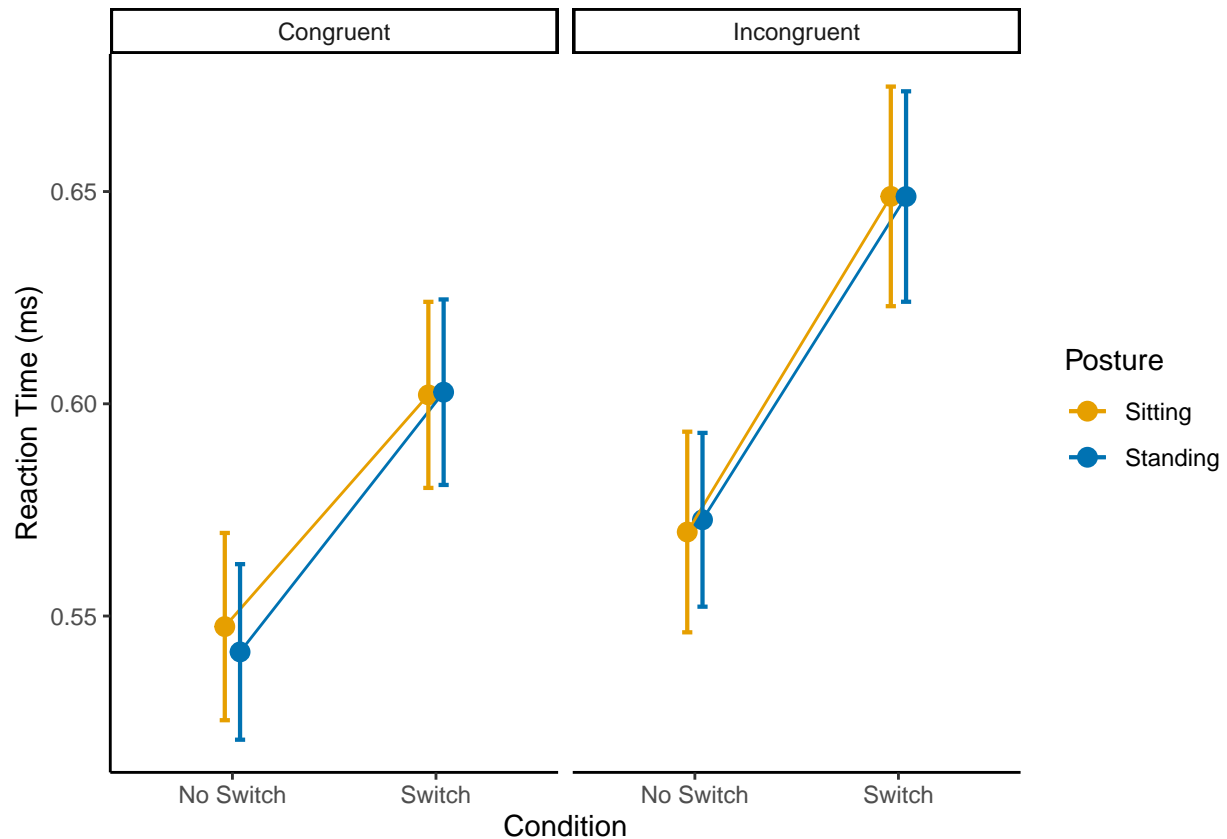
```

```

ggsave(RT_plot,
  file = "plots/RT_plot.pdf",
  units = "in",
  width = 6.62,
  height = 5.50,
  dpi = 600)

```

RT\_plot



```
exp1_anova_rt <- ezANOVA(ts_rt_mean,
  dv = mean_rt,
  wid = participant,
  within = .(posture, congruentTrialType, switchTrialType),
  type = 3,
  detailed = TRUE,
  return_aov = T
)
```

```
## Warning: Converting "participant" to factor for ANOVA.
## Warning: Converting "posture" to factor for ANOVA.
## Warning: Converting "congruentTrialType" to factor for ANOVA.
## Warning: Converting "switchTrialType" to factor for ANOVA.
```

```
exp1_stats_rt <- ezStats(ts_rt_mean,
  dv = mean_rt,
  wid = participant,
  within = .(posture, congruentTrialType, switchTrialType),
  type = 3
)
```

```
## Warning: Converting "participant" to factor for ANOVA.
## Warning: Converting "posture" to factor for ANOVA.
## Warning: Converting "congruentTrialType" to factor for ANOVA.
```

```
## Warning: Converting "switchTrialType" to factor for ANOVA.
write.csv(exp1_stats_rt[, -7], file = "Task_Switching_Descriptives_RT.csv", row.names = F)

#data.frame(exp1_anova$ANOVA)
output_rt <- aovEffectSize(exp1_anova_rt, effectSize = "pes")
output_rt <- data.frame(output_rt$ANOVA)
write.csv(output_rt, "Task_switching_ANOVA_RT.csv")
output_rt
```

```
##              Effect DFn DFd          SSn          SSd          F
## 1      (Intercept)    1   50 1.428705e+02 4.29367155 1.663733e+03
## 2      posture      1   50 3.979595e-05 0.43347417 4.590349e-03
## 3 congruentTrialType 1   50 1.363938e-01 0.14478068 4.710359e+01
## 4      switchTrialType 1   50 4.682022e-01 0.16485963 1.420003e+02
## 5 posture:congruentTrialType 1   50 4.266607e-04 0.06545830 3.259027e-01
## 6 posture:switchTrialType 1   50 8.700576e-05 0.05211061 8.348180e-02
## 7 congruentTrialType:switchTrialType 1   50 9.914733e-03 0.04607197 1.076005e+01
## 8 posture:congruentTrialType:switchTrialType 1   50 5.769395e-04 0.06995820 4.123459e-01
##              p p..05      pes
## 1 4.809845e-40      * 9.708239e-01
## 2 9.462532e-01      9.179855e-05
## 3 9.823661e-09      * 4.850860e-01
## 4 3.195952e-16      * 7.395837e-01
## 5 5.706388e-01      6.475845e-03
## 6 7.738279e-01      1.666853e-03
## 7 1.893419e-03      * 1.770909e-01
## 8 5.237146e-01      8.179462e-03
```

## Reproduce ANOVA for Smith et al. task-switching data

```
#load acc data
Smith_Exp2_acc <- read_excel("StandingData.xlsx",
                             sheet = "Exp2Acc")

head(Smith_Exp2_acc)

## # A tibble: 6 x 9
##   subj sit_congruent_noswitch sit_congruent_sw~ sit_incongruent~ sit_incongruent~ stand_congruent~
##   <chr>          <dbl>          <dbl>          <dbl>          <dbl>          <dbl>
## 1 1              1              1              0.957          0.951          0.98
## 2 2              0.976          0.978          0.981          0.796          0.98
## 3 3              0.977          0.979          0.98           0.935          0.980
## 4 4              0.893          0.884          0.816          0.549          0.980
## 5 5              1              0.98           0.925          0.933          0.978
## 6 6              1              0.95           0.957          0.868          1
## # ... with 3 more variables: stand_congruent_switch <dbl>, stand_incongruent_noswitch <dbl>,
## #   stand_incongruent_switch <dbl>

#Drop last row with comment
Smith_Exp2_acc <- Smith_Exp2_acc[1:30,]

#Restructure from wide to narrow, using tidyr
Smith_exp2_acc_narrow <- Smith_Exp2_acc %>%
  gather(v, accuracy, colnames(Smith_Exp2_acc)[2:9]) %>%
```

```

separate(v, c("posture", "con", "switch")) %>%
  arrange(subj)

Smith_exp2_acc_anova <- ezANOVA(Smith_exp2_acc_narrow,
                                dv = accuracy,
                                wid = subj,
                                within = .(posture, con, switch),
                                type = 3,
                                detailed = TRUE,
                                return_aov=F
                                )

## Warning: Converting "subj" to factor for ANOVA.
## Warning: Converting "posture" to factor for ANOVA.
## Warning: Converting "con" to factor for ANOVA.
## Warning: Converting "switch" to factor for ANOVA.
Smith_exp2_acc_anova

## $ANOVA
##           Effect DFn DFd          SSn          SSd          F          p p<.05          ges
## 1 (Intercept)    1  29 2.048909e+02 0.24225540 2.452716e+04 5.184222e-44 * 0.9965856538
## 2 posture        1  29 1.215410e-02 0.12309940 2.863286e+00 1.013416e-01 0.0170197030
## 3 con            1  29 2.719991e-01 0.11703974 6.739569e+01 4.729349e-09 * 0.2792700999
## 4 switch         1  29 1.403032e-01 0.06464470 6.294084e+01 9.482835e-09 * 0.1665777615
## 5 posture:con    1  29 2.734424e-03 0.04708210 1.684256e+00 2.045882e-01 0.0038802688
## 6 posture:switch 1  29 7.099180e-03 0.03715534 5.540959e+00 2.556408e-02 * 0.0100120392
## 7 con:switch     1  29 3.248637e-02 0.04037101 2.333617e+01 4.059829e-05 * 0.0442321480
## 8 posture:con:switch 1 29 5.244392e-04 0.03031748 5.016491e-01 4.844309e-01 0.0007465437

output_Smith_exp2_acc <- aovEffectSize(Smith_exp2_acc_anova, effectSize = "pes")
output_Smith_exp2_acc <- data.frame(output_Smith_exp2_acc$ANOVA)
write.csv(output_Smith_exp2_acc, "Smith_Exp2_ANOVA_acc.csv")
output_Smith_exp2_acc

##           Effect DFn DFd          SSn          SSd          F          p p.05          pes
## 1 (Intercept)    1  29 2.048909e+02 0.24225540 2.452716e+04 5.184222e-44 * 0.99881903
## 2 posture        1  29 1.215410e-02 0.12309940 2.863286e+00 1.013416e-01 0.08986162
## 3 con            1  29 2.719991e-01 0.11703974 6.739569e+01 4.729349e-09 * 0.69915667
## 4 switch         1  29 1.403032e-01 0.06464470 6.294084e+01 9.482835e-09 * 0.68457979
## 5 posture:con    1  29 2.734424e-03 0.04708210 1.684256e+00 2.045882e-01 0.05488989
## 6 posture:switch 1  29 7.099180e-03 0.03715534 5.540959e+00 2.556408e-02 * 0.16041707
## 7 con:switch     1  29 3.248637e-02 0.04037101 2.333617e+01 4.059829e-05 * 0.44588988
## 8 posture:con:switch 1 29 5.244392e-04 0.03031748 5.016491e-01 4.844309e-01 0.01700410

#plot acc effect sizes for our results and Smith together
#Check that ANOVA tables (rows) match
output_acc$Effect

## [1] "(Intercept)"          "posture"
## [3] "congruentTrialType"   "switchTrialType"
## [5] "posture:congruentTrialType" "posture:switchTrialType"
## [7] "congruentTrialType:switchTrialType" "posture:congruentTrialType:switchTrialType"

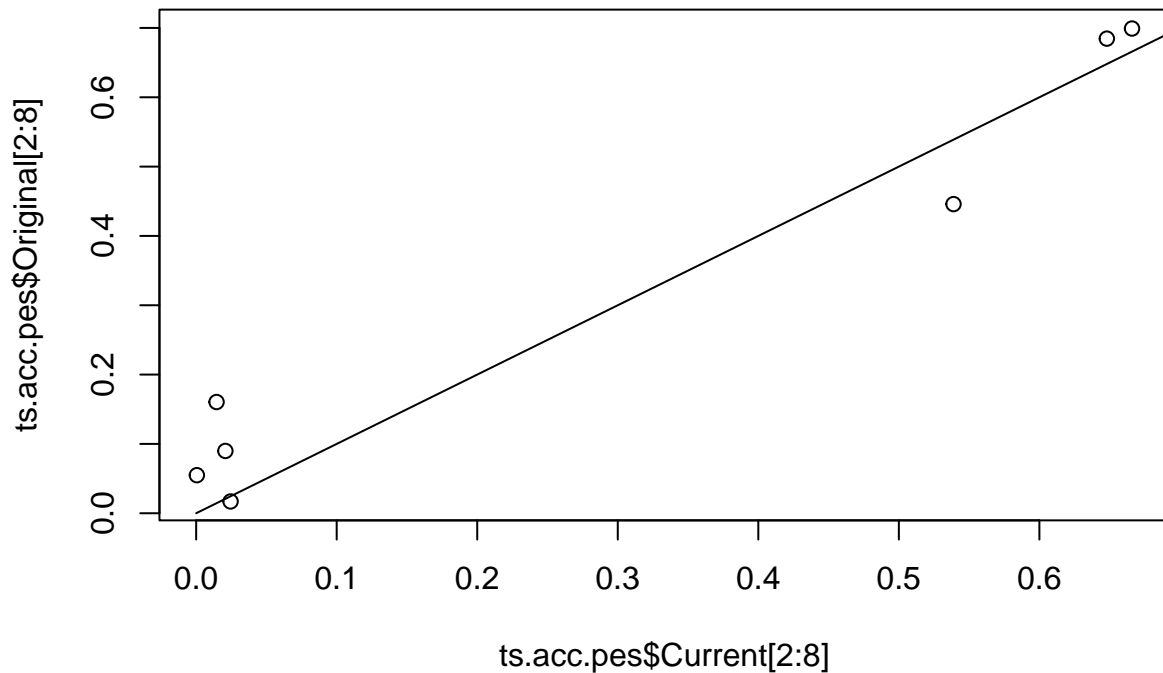
```

```
output_Smith_exp2_acc$Effect
```

```
## [1] "(Intercept)"      "posture"           "con"               "switch"
## [5] "posture:con"       "posture:switch"    "con:switch"        "posture:con:switch"
```

```
ts.acc.pes <- data.frame(matrix(nrow = 8, ncol = 3))
ts.acc.pes[1] <- output_acc$Effect
ts.acc.pes[2] <- output_acc$pes
ts.acc.pes[3] <- output_Smith_exp2_acc$pes

colnames(ts.acc.pes) <- c("Effect", "Current", "Original")
plot(ts.acc.pes$Current[2:8], ts.acc.pes$Original[2:8])
lines(x = c(0,1), y = c(0,1))
```



```
#Exact proportion of current to original effect: ~9%
ts.acc.pes$Current[6]/ts.acc.pes$Original[6]
```

```
## [1] 0.09049606
```

```
#Compare current vs. original effect sizes
```

```
interaction.original <-
get.ci.partial.eta.squared(output_Smith_exp2_acc$F[6],
                           output_Smith_exp2_acc$DFn[6],
                           output_Smith_exp2_acc$DFd[6],
                           conf.level = 0.90)
```

```
ts.acc.pes.int <- data.frame(matrix(nrow = 2, ncol = 5))
```

```

colnames(ts.acc.pes.int) <- c("Exp", "Effect.size", "CI.Lower", "CI.Upper", "name")
ts.acc.pes.int[1] <- c("Current", "Original")
ts.acc.pes.int[1,2:4] <- c(ts.acc.pes$Current[6], data.frame(interaction_effect_CI))
ts.acc.pes.int[2,2:4] <- c(ts.acc.pes$Original[6], data.frame(interaction.original))
ts.acc.pes.int[1:2,5] <- c("Accuracy", "Accuracy")
#Graph comparison of key effects for all three experiments- Similar to forest plot in sig
#filtering paper?
forest.colors <- c("black", "red")

ts.acc.pes.int$Exp <- as.factor(ts.acc.pes.int$Exp)

str(ts.acc.pes.int)

## 'data.frame': 2 obs. of 5 variables:
## $ Exp : Factor w/ 2 levels "Current","Original": 1 2
## $ Effect.size: num 0.0145 0.1604
## $ CI.Lower : num 0 0.011
## $ CI.Upper : num 0.107 0.347
## $ name : chr "Accuracy" "Accuracy"

forest.comp <- mod.forestplot(df = ts.acc.pes.int,
                             estimate = Effect.size,
                             ci.lower = CI.Lower,
                             ci.upper = CI.Upper,
                             colour = Exp,
                             xlab = "Partial Eta-Squared"
) +
  scale_color_manual(values = forest.colors)

## Scale for 'colour' is already present. Adding another scale for 'colour', which will replace the
## existing scale.

forest.comp

```



