

World_insects_estimate

Load libraries and ../Data

```
library(SpadeR)
library(dplyr)
library(stringr)
library(pairwiseCI)
#library(remotes)
#install_version("CARE1", "1.1.0")
library(CARE1)

## Load Data

#Microgastrinae

reared <- read.csv("../Data/Microgastrinae ACG Reared.csv") %>%
  select(BOLD.code, Frequency)

core <- read.csv("../Data/Microgastrinae ACG Core Malaise.csv") %>%
  select(BOLD.code, Frequency)

peri <- read.csv("../Data/Microgastrinae ACG Peripheral Malaise.csv") %>%
  select(BOLD.code, Frequency)

# All insecta core

insec <- read.csv("../Data/Insecta ACG Core Malaise.csv") %>%
  select(BOLD.code, Frequency)

## highlight estimator

estimator <- "      iChao1 (Chiu et al. 2014)"
```

```

## adding in the trees

#TA = number of tree species in ACG
TA <- 1500

#Tc = number of tree species in CR

TC <- 2500
#Tw = number of tree species on Earth

## estimated tree species

TW <- 73274
TW_CI <- c(73271, 73276)

## described number of tree species

TW_d <- 60065

### Number of Saturniid moths

SA <- 106
SW <- 4535

```

Approach 1: Calculate insect richness worldwide using the CARE estimator for Microgastrinae

```

### Total number of BINS

reared %>% nrow()

```

```
[1] 886
```

```
core %>% nrow()
```

```
[1] 388
```

```
peri %>% nrow()
```

```
[1] 576
```

```
###
```

Step 1: Combining Malaise and Reared samples of Microgastrinae and estimating the true richness of ACG Microgastrinae

```
full_micro_table <- reared %>%  
  rename(reared = Frequency) %>%  
  full_join(rename(core, core = Frequency)) %>%  
  full_join(rename(peri, peri = Frequency)) %>%  
  mutate(reared = ifelse(is.na(reared), 0, 1)) %>%  
  mutate(core = ifelse(is.na(core), 0, 1)) %>%  
  mutate(peri = ifelse(is.na(peri), 0, 1))
```

```
Joining with `by = join_by(BOLD.code)`
```

```
Joining with `by = join_by(BOLD.code)`
```

```
head(full_micro_table)
```

	BOLD.code	reared	core	peri
1	BOLD:AAA1266	1	0	0
2	BOLD:AAA1634	1	1	1
3	BOLD:AAA2143	1	0	0
4	BOLD:AAA2511	1	0	0
5	BOLD:AAA2668	1	0	0
6	BOLD:AAA3229	1	0	1

```
nrow(full_micro_table)
```

```
[1] 1411
```

```
# Transform observed ../Data to CARE1 input format  
micro_care <- as.record(full_micro_table[, -1])  
  
micro_care
```

```
001 010 011 100 101 110 111
246 133 146 663 114 39 70
```

```
# Main step to obtain population size estimates
CARE1.print(micro_care)
```

```
Warning in ni/nhat: Recycling array of length 1 in vector-array arithmetic is deprecated.
Use c() or as.vector() instead.
Warning in ni/nhat: Recycling array of length 1 in vector-array arithmetic is deprecated.
Use c() or as.vector() instead.
Warning in ni/nhat: Recycling array of length 1 in vector-array arithmetic is deprecated.
Use c() or as.vector() instead.
```

(1) NUMBER OF IDENTIFIED CASES IN EACH LIST:

```
n1 n2 n3
886 388 576
```

(2) ESTIMATES BASED ON ANY PAIR OF SAMPLES:

	Petersen	Chapman	se	cil	ciu
pa12	3154	3136	236	2725	3655
pa13	2774	2765	149	2501	3087
pa23	1035	1033	37	970	1115

Note1: Refer to Seber(1982,pages 59 and 60) for Petersen estimator and Chapman estimators as well as s.e formula.

Note2: A log-transformation is used is used to obtain the confidence interval so that the lower limit is always greater than the number of ascertained. Refer to Chao(1987,Biometrics,43,783-791) for the construction of the confidence interval.

(3) SAMPLE COVERAGE APPROACH:

	M	D	Chat	est	se	cil	ciu
Nhat-0	1411	1063.667	0.494	2153	58	2048	2277
Nhat	1411	1063.667	0.494	3480	526	2678	4790
Nhat-1	1411	1063.667	0.494	2402	97	2230	2610

Warning: The estimated sample coverage(overlapping information)is too low so that Nhat is unstable. Recommend the use of Nhat-0 or Nhat-1.

Parameter estimates:

	u1	u2	u3	r12	r13	r23
Nhat-0	0.41	0.18	0.27	-0.32	-0.22	1.08

```
Nhat    0.25 0.11 0.17  0.10  0.25 2.36
Nhat-1  0.37 0.16 0.24 -0.24 -0.13 1.32
```

Definitions for the sample coverage approach:

M: number of individuals ascertained in at least one list.

D: the average of the number of individuals listed in the combination of any two lists omitting the other one.

\hat{C} : sample coverage estimate, see Equation (14) of Chao and Tsay(1998).
est: population size estimate.

se: estimated standard error of the population size estimation based on bootstrap replications.

cil: 95% confidence interval lower limit(using a log-transformation).

ciu: 95% confidence interval upper limit(using a log-transformation).

Nhat: Population size estimate for sufficiently high sample coverage cases, see Equation (20) of Chao and Tsay (1998).

Nhat-1: One-step population size estimate for low sample coverage cases; see Equation (2.21) of Chao et al. (1996). This estimator is suggested for use when the estimated se of Nhat is relatively large.

u1,u2,u3: estimated mean probabilities depending on the estimate of N.

r12,r13,r23 etc.: estimated coefficient of covariation(CCV) depending on the estimate of N.

```
# Print the CARE1 input format

## n1 = reared = 886
## n2 = core = 388
## n3 = peripheral = 576

## Using Nhat-1

Nhat1 <- round(estN(micro_care, method = "LSC", se = TRUE, nboot = 200))

micro_all_estimate <- Nhat1[1]
micro_all_CI <- c(Nhat1[3], Nhat1[4])
```

Step 2. Estimating the undersampling ratio for Core Microgastrinae.

```
## proportion of core malaise of the ichao total
```

```
proportion_core <- nrow(core)/micro_all_estimate
```

```
proportion_core
```

```
[1] 0.1615321
```

```
CI_proportion_core <- nrow(core)/micro_all_CI
```

```
CI_proportion_core
```

```
[1] 0.1729826 0.1496336
```

Step 3 Calculate total number of insects in ACG

```
## Total number of insecta bins in ACG core malaise
```

```
n_insects_core <- nrow(incsc)
```

```
n_insects_core
```

```
[1] 53945
```

```
total_insec_acg <- n_insects_core*(1/proportion_core)
```

```
total_insec_acg
```

```
[1] 333958.5
```

```
CI_total_insect_acg <- n_insects_core*(1/round(CI_proportion_core,3))
```

```
n_insects_core*(1/round(CI_proportion_core,3))
```

```
[1] 311820.8 359633.3
```

```
n_insects_core*(1/CI_proportion_core)
```

```
[1] 311852.2 360513.9
```

```
CI_total_insect_acg
```

```
[1] 311820.8 359633.3
```

Step 4. Estimating global insect richness – based on trees

```
##### Total insects
```

```
print("Total world insects based on estimated tree richness")
```

```
[1] "Total world insects based on estimated tree richness"
```

```
total_insect_world <- (TW*total_insec_acg)/TA  
total_insect_world
```

```
[1] 16313649
```

```
CI_total_insect_world <- (TW*CI_total_insect_acg)/TA  
CI_total_insect_world
```

```
[1] 15232239 17567849
```

```
print("Total world insects based on described trees")
```

```
[1] "Total world insects based on described trees"
```

```
total_insect_world <- (TW_d*total_insec_acg)/TA  
total_insect_world
```

```
[1] 13372811
```

```
CI_total_insect_world <- (TW_d*CI_total_insect_acg)/TA  
CI_total_insect_world
```

```
[1] 12486345 14400917
```

Step 4. Estimating global insect richness – based on saturniid moths

```
##### Total insects
```

```
print("Total world insects based on Saturniid moths")
```

```
[1] "Total world insects based on Saturniid moths"
```

```
total_insect_world <- (SW*total_insec_acg)/SA  
total_insect_world
```

```
[1] 14287752
```

```
CI_total_insect_world <- (SW*CI_total_insect_acg)/SA  
CI_total_insect_world
```

```
[1] 13340636 15386200
```

Additional Information

For Pooled Microgastrinae CV and Sample Coverage: $1 - [\text{number singletons}] / [\text{number specimens}]$.

```
### Combine all microgastrinae
```

```
micro_all <- rbind(reared, core, peri) %>%  
  group_by(BOLD.code) %>%  
  summarise(new_freq = sum(Frequency)) %>%  
  select(BOLD.code, new_freq)
```

```
## number of pooled micros  
nrow(micro_all)
```

```
[1] 1411
```



```
## use chao on microgastrinae
```

```
micro_all_estimate <- ChaoSpecies(micro_all$new_freq,"abundance",k=10,conf=0.95)  
micro_all_estimate$Species_table[estimator,]
```

```
Estimate      s.e. 95%Lower 95%Upper  
1911.739    35.582 1846.715 1986.466
```

```
## from spade R
```

```
micro_all_estimate$Basic_data_information[c("    CV for entire dataset",  
                                             "    Coverage estimate for entire dataset"),]
```

	Variable	Value
CV for entire dataset	CV	3.01
Coverage estimate for entire dataset	C	0.981

```
## based on coverage spreadsheet
```

```
total_bins <- nrow(micro_all)  
total_bins
```

```
[1] 1411
```

```
total_singletons_micro_all <- sum(micro_all$new_freq ==1)  
total_singletons_micro_all
```

```
[1] 406
```

```
total_doubletons_micro_all <- sum(micro_all$new_freq ==2)  
total_doubletons_micro_all
```

```
[1] 202
```

```
# total samples  
sum(micro_all$new_freq)
```

```
[1] 21708
```

```
coverage_micro_all <- 1-(total_singletons_micro_all/sum(micro_all$new_freq))
coverage_micro_all
```

```
[1] 0.9812972
```

```
CV_micro_all <- sd(micro_all$new_freq)/mean(micro_all$new_freq)
CV_micro_all
```

```
[1] 2.990786
```

Sample Coverage For Core Malaise Microgastrinae.

```
## from Spade R
micro_core_estimate <- ChaoSpecies(core$Frequency,"abundance",k=10,conf=0.95)

micro_core_estimate$Basic_data_information[c("    CV for entire dataset",
      "    Coverage estimate for entire dataset"),]
```

	Variable	Value
CV for entire dataset	CV	3.39
Coverage estimate for entire dataset	C	0.953

```
## calculating from the spreadsheet
```

```
total_bins <- nrow(core)
total_bins
```

```
[1] 388
```

```
total_singletons_core <- sum(core$Frequency ==1)
total_singletons_core
```

```
[1] 176
```

```
total_doubletons_core <- sum(core$Frequency ==2)
total_doubletons_core
```

```
[1] 54
```

```
# total samples
sum(core$Frequency)
```

```
[1] 3781
```

```
coverage_core <- 1-(total_singletons_core/sum(core$Frequency))
coverage_core
```

```
[1] 0.9534515
```

```
CV_core <- sd(core$Frequency)/mean(core$Frequency)
CV_core
```

```
[1] 3.322479
```

Sample Coverage For Core Insecta

```
## from spade R

insec_acg_estimate <- readRDS("../Data/insecta_acg_chao.rds")

insec_acg_estimate$Basic_data_information[c("    CV for entire dataset",
                                             "    Coverage estimate for entire dataset"),]
```

	Variable	Value
CV for entire dataset	CV	9.009
Coverage estimate for entire dataset	C	0.988

```
insecta <- read.csv("../Data/Insecta ACG Core Malaise.csv")
```

```
## from the spreadsheet
```

```
total_bins <- nrow(insecta)
total_bins
```

```
[1] 53945
```

```
total_singletons_insecta <- sum(insecta$Frequency ==1)
total_singletons_insecta
```

```
[1] 19374
```

```
total_doubletons_insecta <- sum(insecta$Frequency ==2)
total_doubletons_insecta
```

```
[1] 7402
```

```
coverage_insecta <- 1-(total_singletons_insecta/sum(insecta$Frequency))
coverage_insecta
```

```
[1] 0.9881422
```

```
CV_insecta <- sd(insecta$Frequency)/mean(insecta$Frequency)
CV_insecta
```

```
[1] 8.956866
```

Number of singletons and sample coverage for Core Malaise for each taxonomic level (Braconidae, Ichneumonoidea, and Hymenoptera).

Braconidae

```
braconidae <- read.csv("../Data/Braconidae ACG Core Malaise.csv")

## from the spreadsheet

total_bins <- nrow(braconidae)
total_bins
```

```
[1] 2160
```

```
total_singletons_braconidae <- sum(braconidae$Frequency ==1)
total_singletons_braconidae
```

```
[1] 1022
```

```
total_doubletons_braconidae <- sum(braconidae$Frequency ==2)
total_doubletons_braconidae
```

```
[1] 342
```

```
## total number of samples
sum(braconidae$Frequency)
```

```
[1] 15108
```

```
coverage_braconidae <- 1-(total_singletons_braconidae/sum(braconidae$Frequency))
coverage_braconidae
```

```
[1] 0.9323537
```

```
CV_braconidae <- sd(braconidae$Frequency)/mean(braconidae$Frequency)
CV_braconidae
```

```
[1] 3.270035
```

Ichneumonoidea

```
ichneumonoidea <- read.csv("../Data/Ichneumonoidea ACG Core Malaise.csv")

## from the spreadsheet

total_bins <- nrow(ichneumonoidea)
total_bins
```

```
[1] 3402
```

```
total_singletons_ichneumonoidea <- sum(ichneumonoidea$Frequency ==1)
total_singletons_ichneumonoidea
```

```
[1] 1496
```

```
total_doubletons_ichneumonoidea <- sum(ichneumonoidea$Frequency ==2)
total_doubletons_ichneumonoidea
```

```
[1] 530
```

```
## total number of samples
sum(ichneumonoidea$Frequency)
```

```
[1] 33207
```

```
coverage_ichneumonoidea <- 1-(total_singletons_ichneumonoidea/sum(ichneumonoidea$Frequency))
coverage_ichneumonoidea
```

```
[1] 0.9549493
```

```
CV_ichneumonoidea <- sd(ichneumonoidea$Frequency)/mean(ichneumonoidea$Frequency)
CV_ichneumonoidea
```

```
[1] 3.862408
```

Hymenoptera

```
hymenoptera <- read.csv("../Data/Hymenoptera ACG Core Malaise.csv")
## from the spreadsheet
total_bins <- nrow(hymenoptera)
total_bins
```

```
[1] 13934
```

```
total_singletons_hymenoptera <- sum(hymenoptera$Frequency ==1)
total_singletons_hymenoptera
```

```
[1] 5689
```

```
total_doubletons_hymenoptera <- sum(hymenoptera$Frequency ==2)
total_doubletons_hymenoptera
```

```
[1] 2061
```

```
## total number of samples

sum(hymenoptera$Frequency)
```

```
[1] 235248
```

```
coverage_hymenoptera <- 1-(total_singletons_hymenoptera/sum(hymenoptera$Frequency))
coverage_hymenoptera
```

```
[1] 0.975817
```

```
CV_hymenoptera <- sd(hymenoptera$Frequency)/mean(hymenoptera$Frequency)
CV_hymenoptera
```

```
[1] 7.55354
```