



Funded by
the European Union

Bioinformatics Summer School

Long-reads Transcriptomics

Satrio Wibowo

University of Nottingham,
Nottingham, UK

1 Library Preparations

2 Basecalling

Section 1

Library preparation (lib-prep)

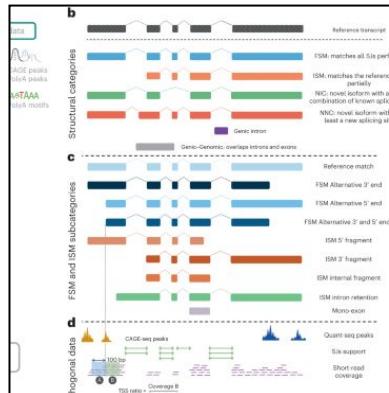
Library preparation strategies for long-reads RNA-seq experiments

Library preparation

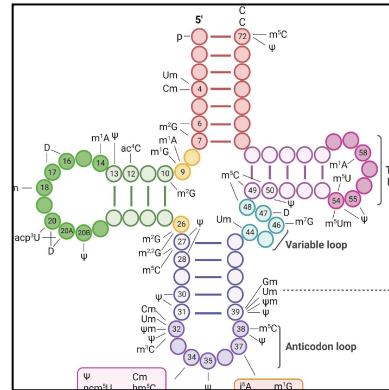
Long TREC

Knowing your goal

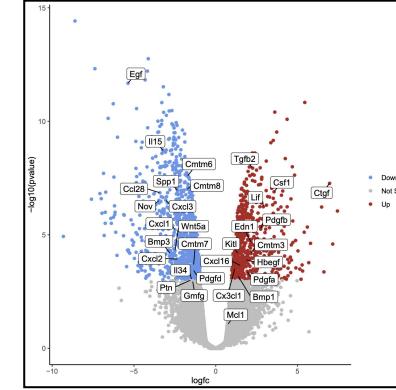
What is your goal?



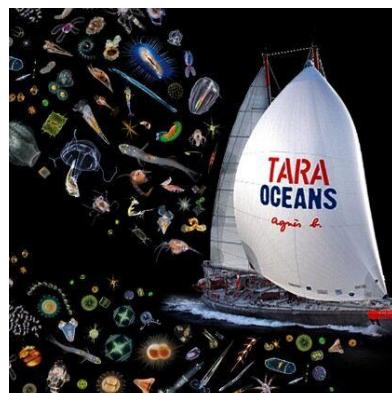
Transcriptomics



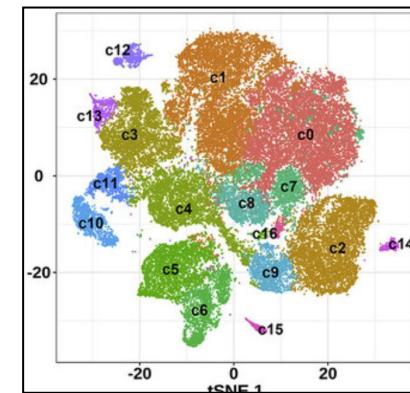
Epi-transcriptomics



Bulk



Meta-transcriptomics



Single-cell



Spatial

RNA-seq

Main difference with short-reads lib-prep

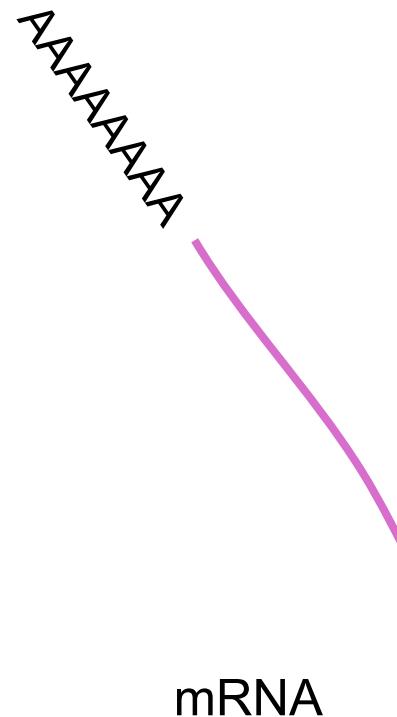


Image: PacBio

RNA-seq

Main difference with short-reads lib-prep

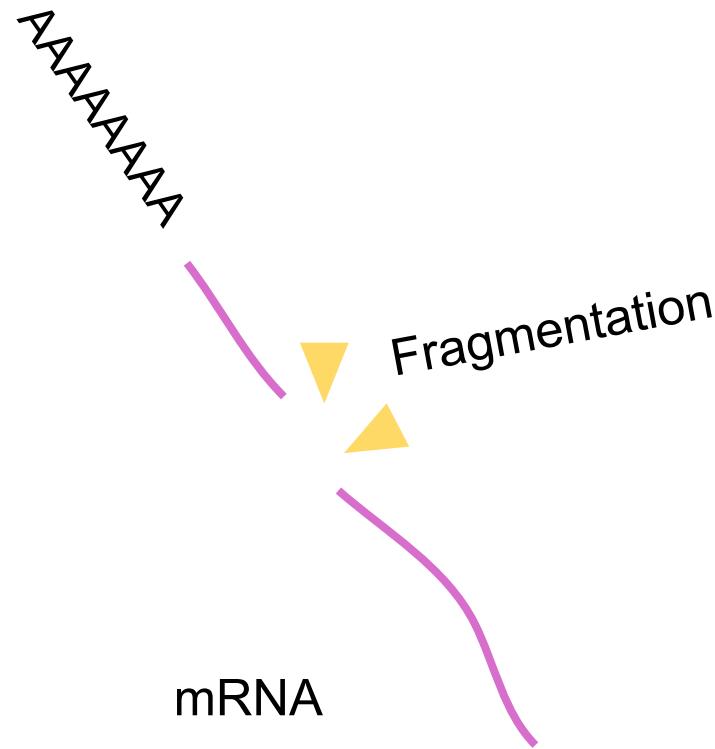
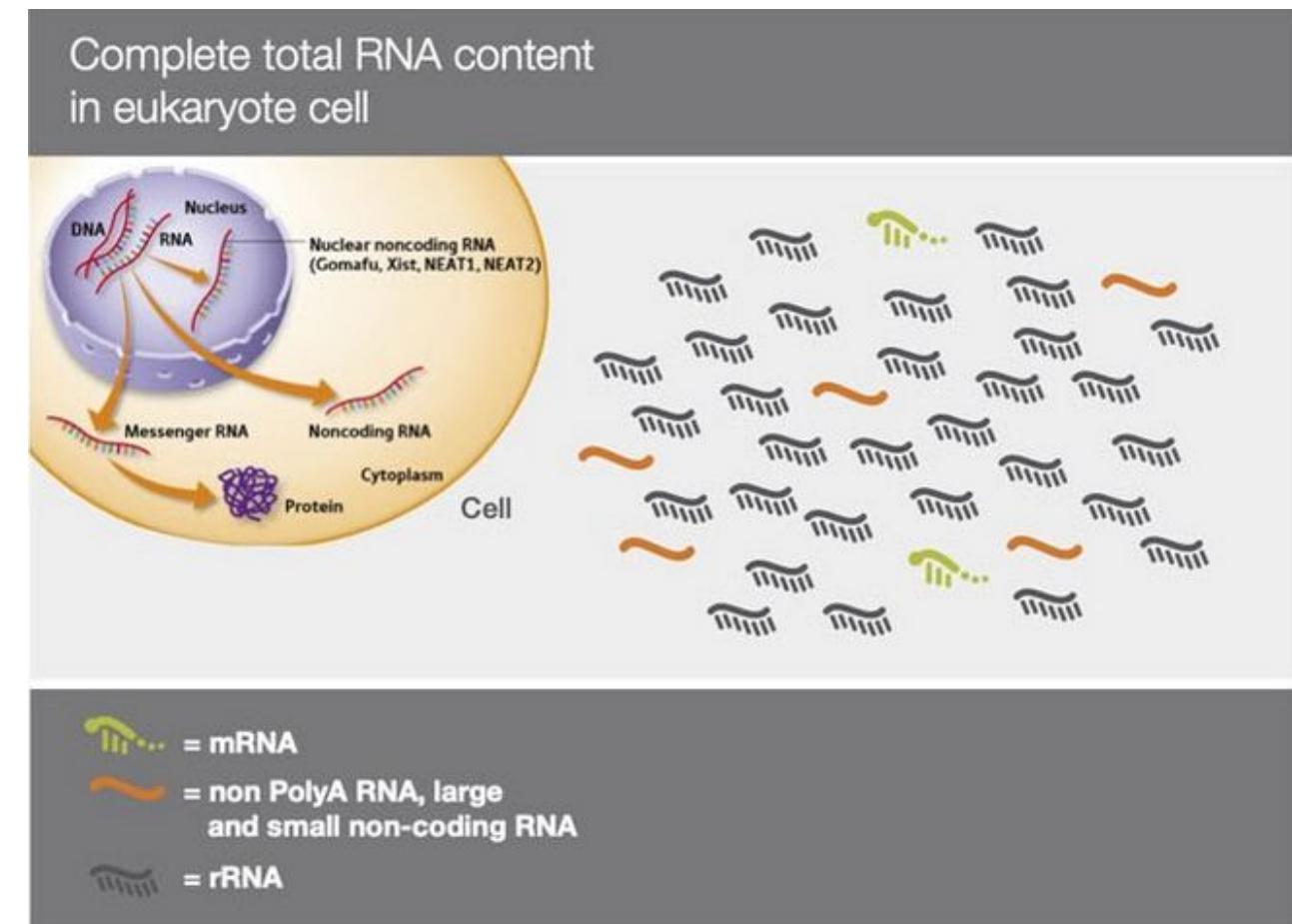


Image: Thermo

RNA-seq

Main difference with short-reads lib-prep

AAAAAAA
mRNA



RNA-seq

Main difference with short-reads lib-prep

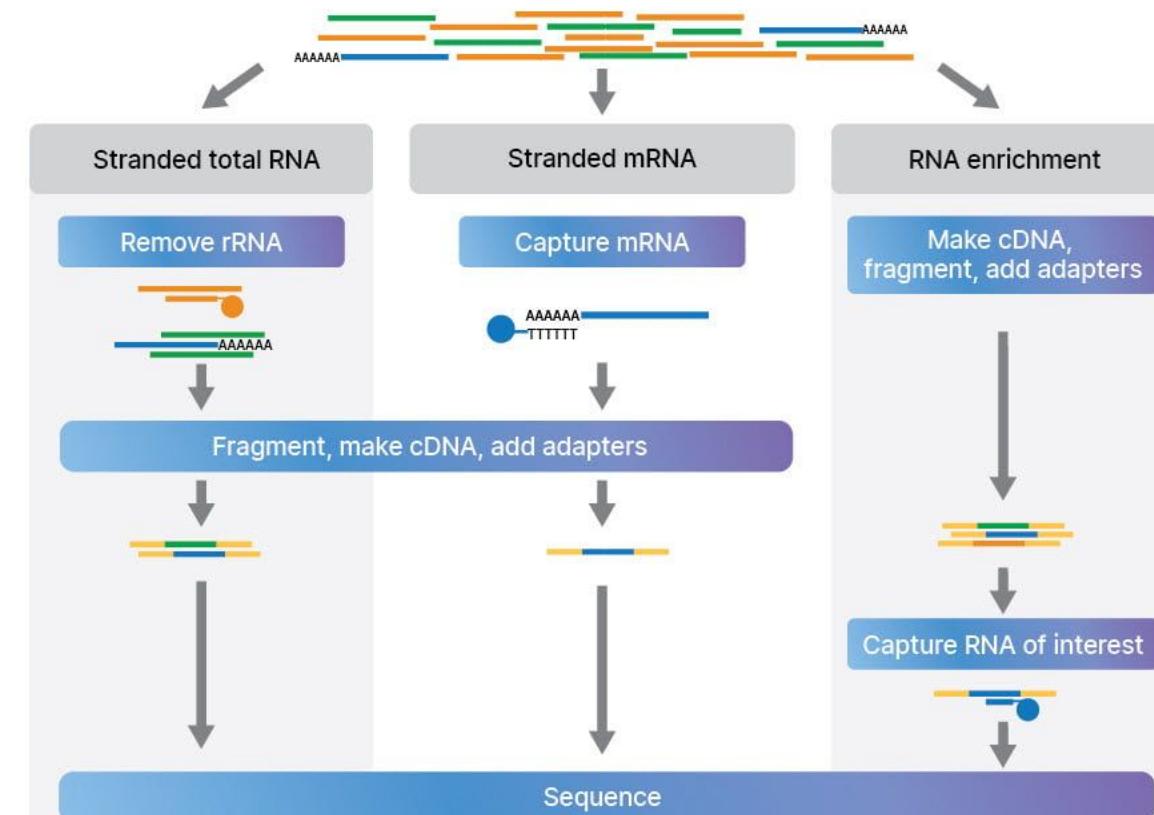
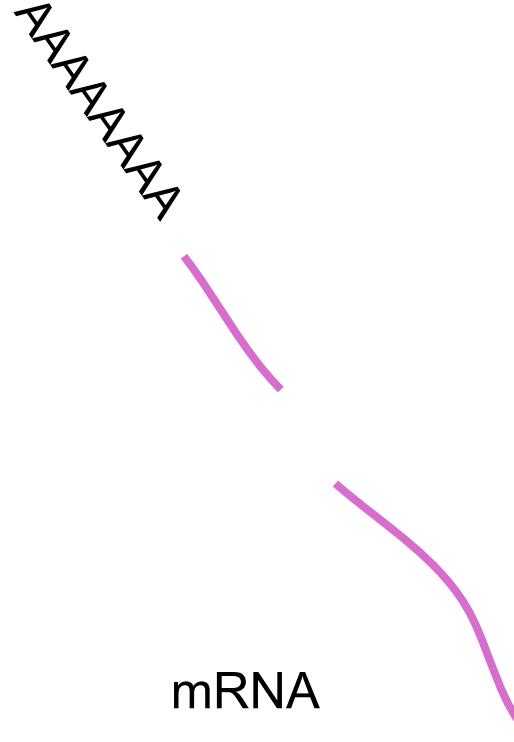


Image: Illumina

Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

Garbage in, garbage out



Consideration Working with RNA

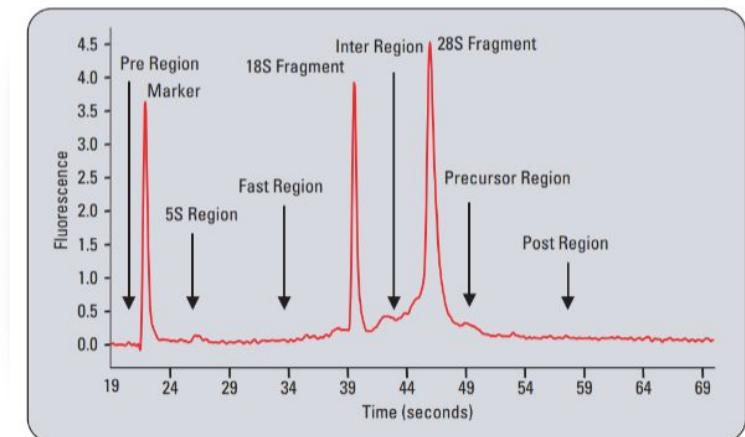
Garbage in, garbage out



RIN < 7

Consideration Working with RNA

Garbage in, garbage out

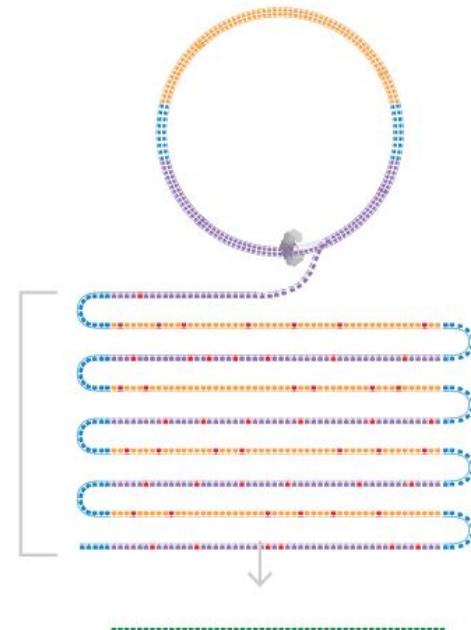


RIN < 7

cDNA Iso-Seq

PacBio

Description	Full-length transcript isoform sequencing
Kit	SMRTbell prep kit 3.0
Multiplex	up to 12
Preparation time	8 Hours 2 Safe stop point
Input	300 ng Total RNA
Output	1M (Sequel I); 8M (Sequel II); Full-length HiFi transcripts



HiFi long reads generated from
PacBio sequencing

Image: PacBio

Library preparation

LongTREC

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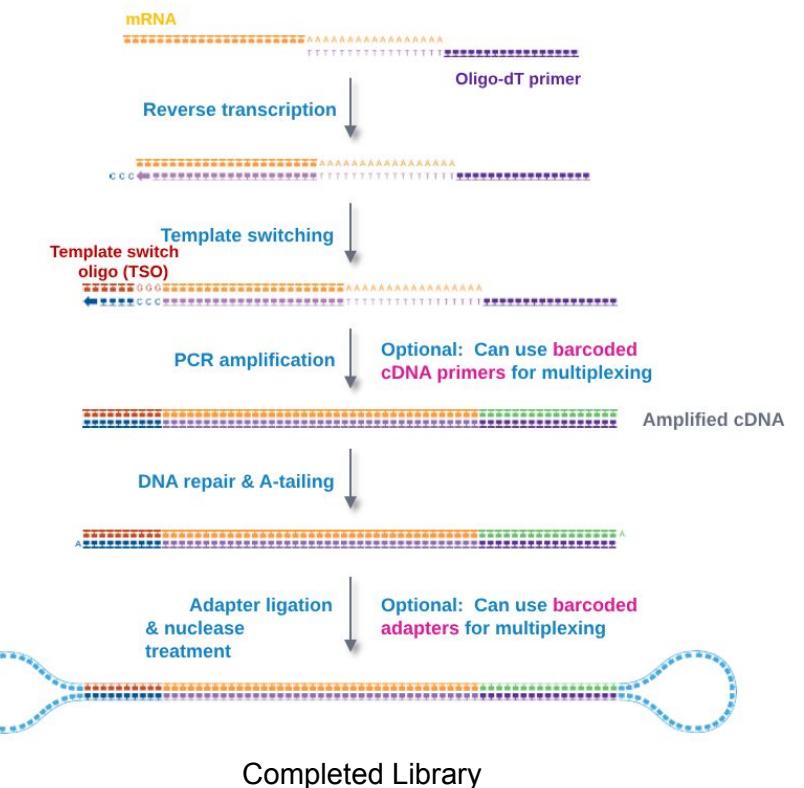
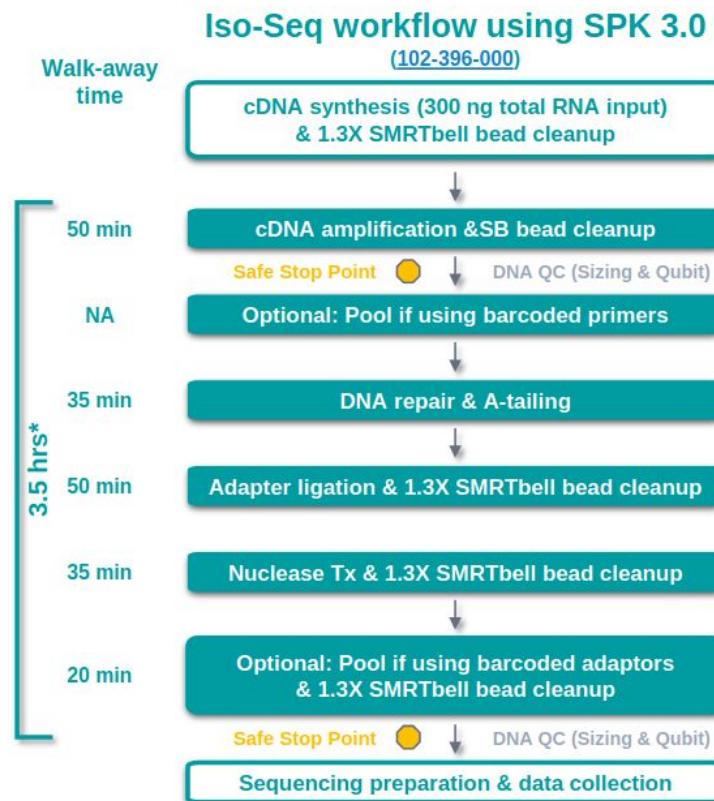


Image: PacBio

cDNA Iso-Seq

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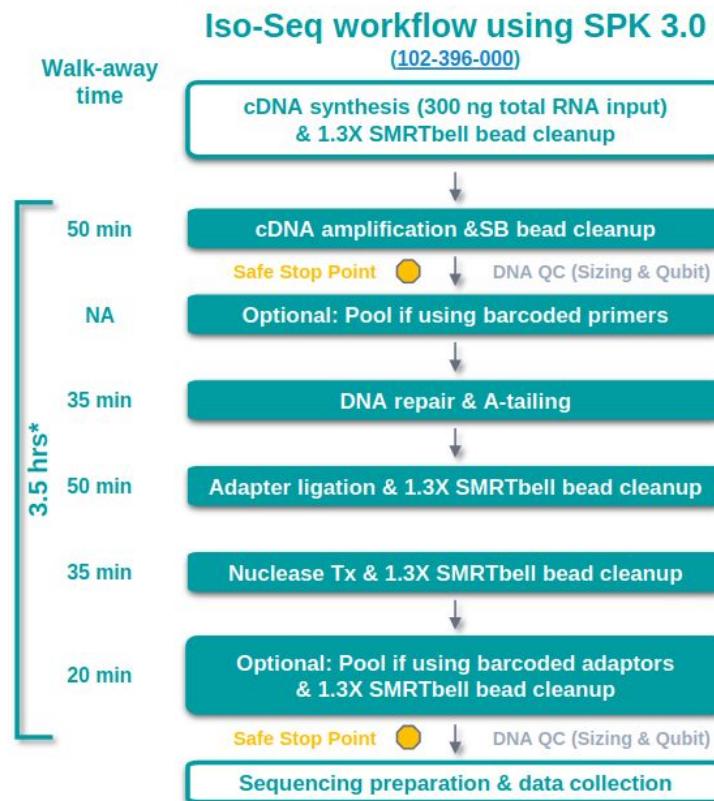
Complete Iso-Seq library preparation workflow for cDNA sequencing

Image: PacBio

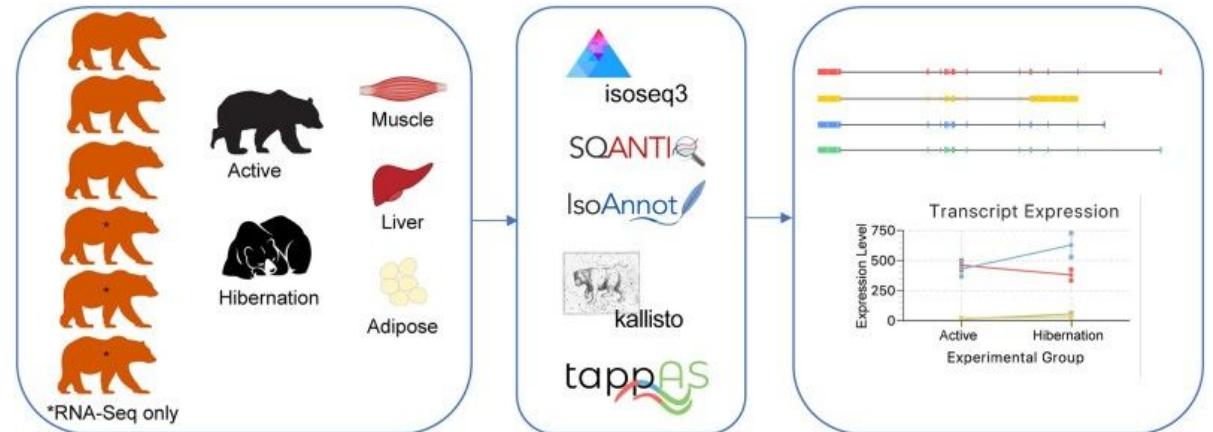
Library preparation

cDNA Iso-Seq

PacBio



Complete Iso-Seq library preparation workflow for cDNA sequencing



Adipose is the most dynamic tissue during hibernation with the highest number of genes with differential isoform usage and isoform switching as compared to liver and muscle.

Tseng et al.(2022). Long-read isoform sequencing reveals tissue-specific isoform expression between active and hibernating brown bears (*Ursus arctos*).

Image: PacBio

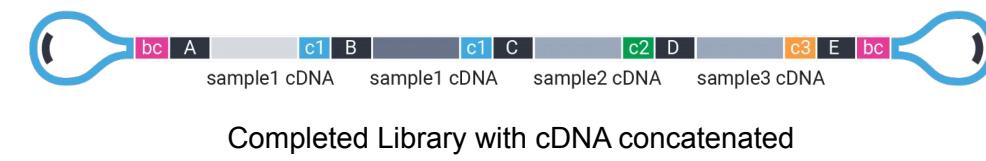


cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio

Description	Full-length isoform discovery with flexible sample multiplexing
Kit	Kinnex full-length RNA kit
Multiplex	5M × 3-plex (Sequel II); 5M x 6-plex (Vega); 10M × 6-plex (Revio SPRQ)
Preparation time	1.5 days 4 Safe stop point
Input	300 ng Total RNA
Output	15-20M (Sequel II); 20-30M (Vega); 50-60M (Revio SPRQ) Full-length HiFi transcripts (Q40)
Notable Studies	Al Khafaji et al., 2023. Nature Portofolio



Completed Library with cDNA concatenated

cDNA

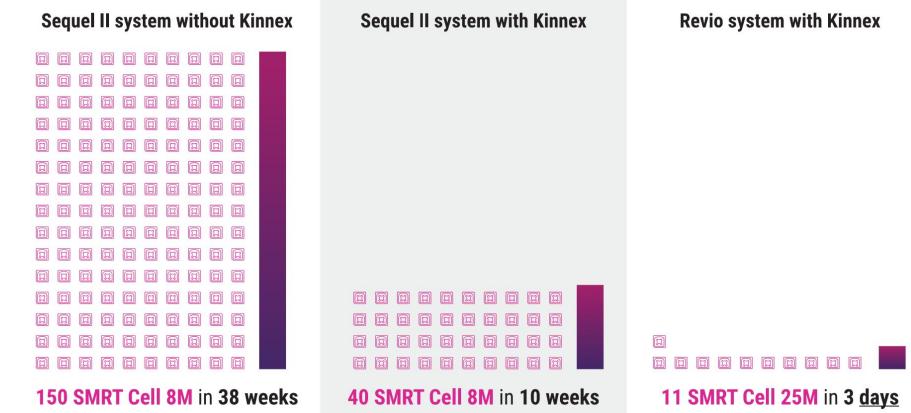
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Completed Library with cDNA concatenated



Kinnex increase throughput significantly

cDNA

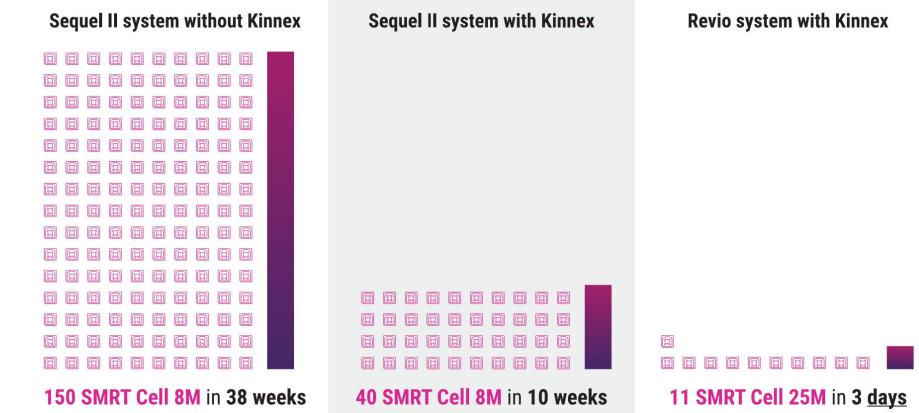
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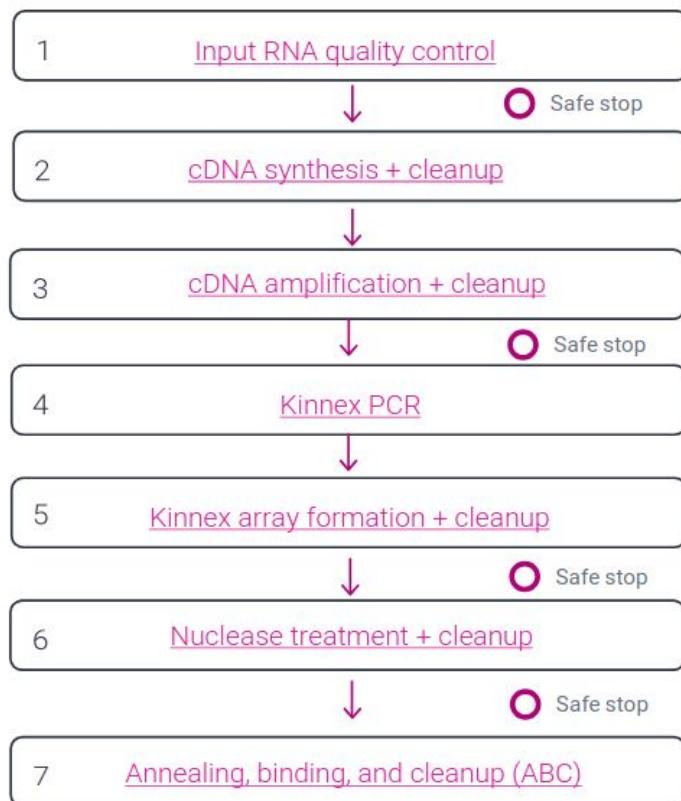
Kinnex increase throughput significantly

Image: PacBio

cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio



SMRTbell® library preparation

Iso-Seq® Express 2.0 Kit	PacBio® 103-071-500
SMRTbell® cleanup beads	PacBio® 102-158-300*
Elution buffer (50 mL)	PacBio® 101-633-500*
Kinnex™ PCR 8-fold kit	PacBio® 103-071-600*
Revio® SPRQ™ polymerase kit or Vega™ polymerase kit or Revio® polymerase kit** or Sequel® II binding kit 3.2**	PacBio® 103-520-100 PacBio® 103-517-600 PacBio® 102-817-600 PacBio® 102-333-300

Complete Kinnex library preparation workflow for cDNA sequencing

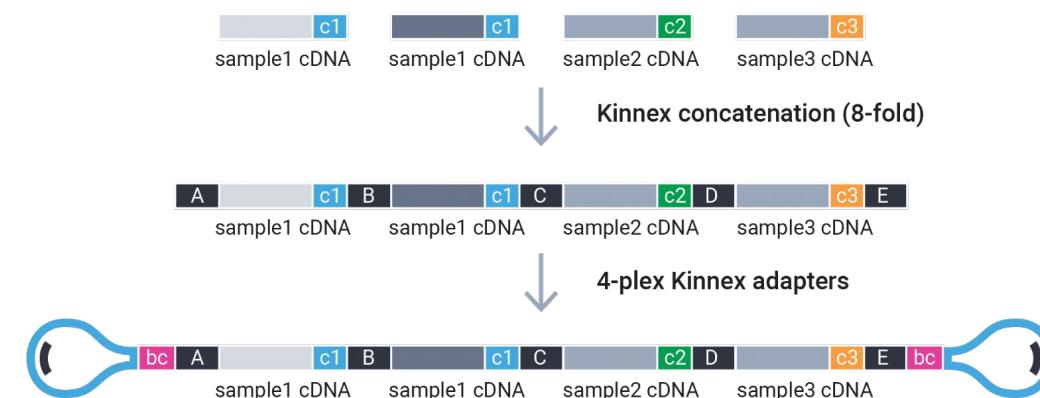
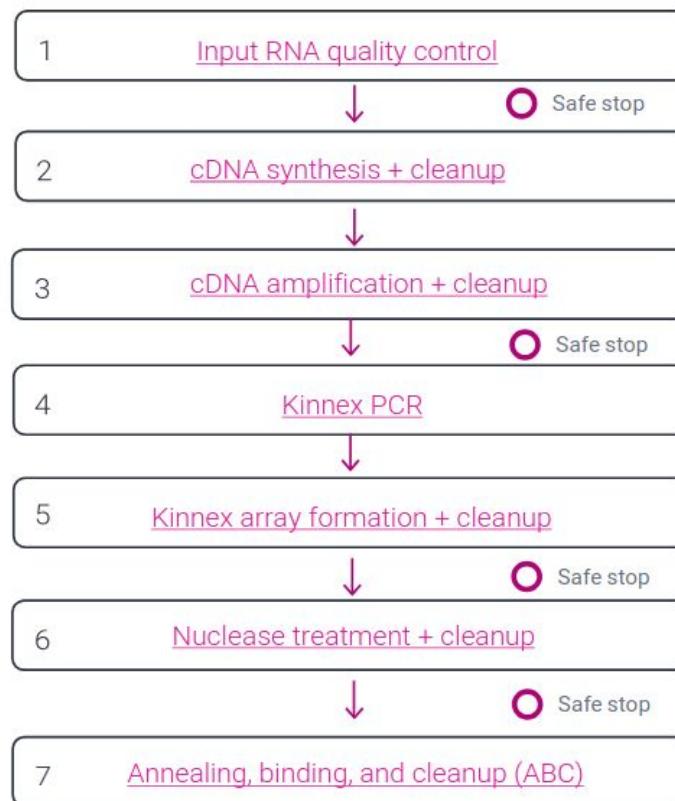
Image: PacBio

Library preparation

cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio



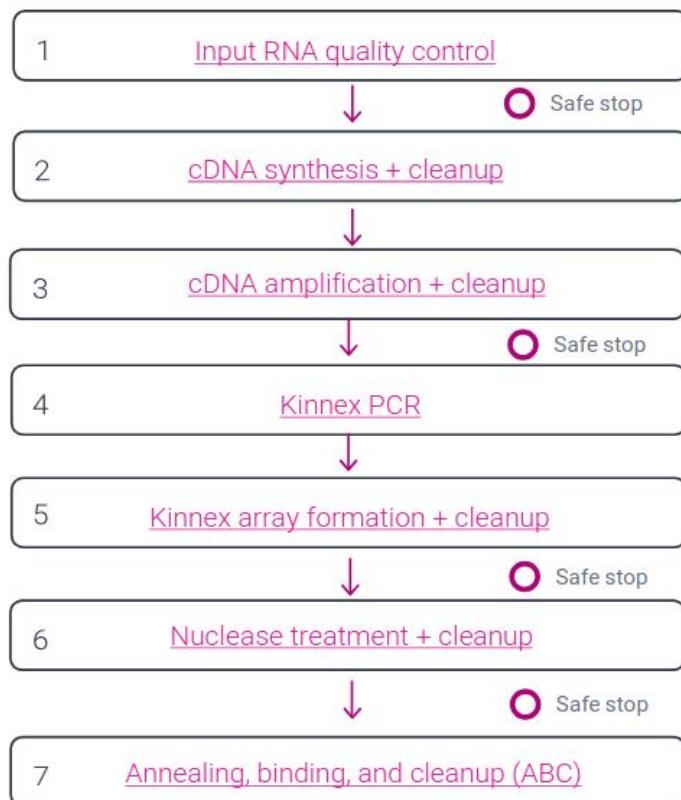
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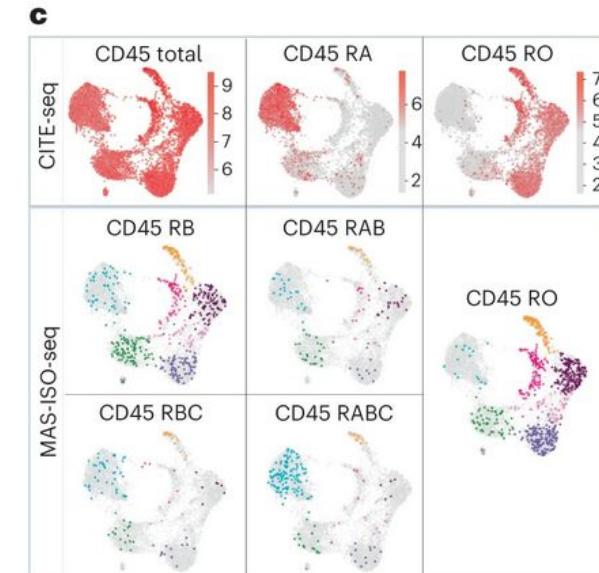
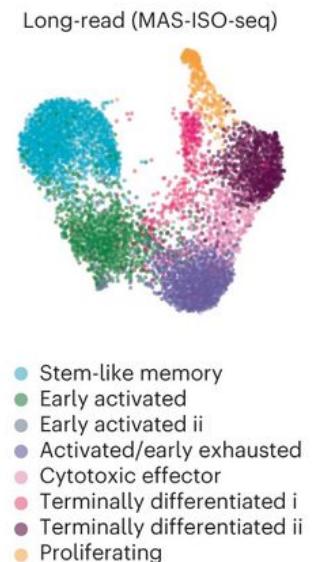
cDNA

Kinnex (Commercial version of MAS-ISO-seq)

PacBio



Complete Kinnex library preparation workflow for cDNA sequencing



Increasing the throughput >15-fold to nearly 40 million cDNA reads per run. MAS-ISO-seq demonstrated a 12- to 32-fold increase in the discovery of differentially spliced genes.

Al'Khafaji, A.M., Smith, J.T., Garimella, K.V. et al (2024). High-throughput RNA isoform sequencing using programmed cDNA concatenation. <https://doi.org/10.1038/s41587-023-01815-7>

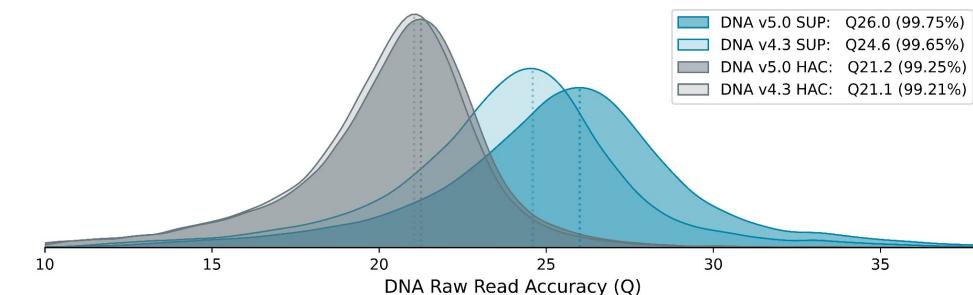
Image: PacBio

cDNA

cDNA-PCR (R10.4.1)

Nanopore

Description	A sequencing kit optimised for identification and quantification of full-length transcripts with highest output.
Kit	cDNA-PCR Sequencing Kit V14
Multiplex	up to 24-plex (cDNA-PCR Barcoding Kit V14)
Preparation time	225 minutes + PCR
Input	10 ng poly(A)+ RNA or 500 ng total RNA
Output	10-15 million cDNA (MinION); 100 million cDNA (PromethION)



Latest transformer model and R10 pores
deliver higher accuracy

cDNA

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Two types of nanopore device with different sequencing throughput (Left: PromethION, Right: MinION)

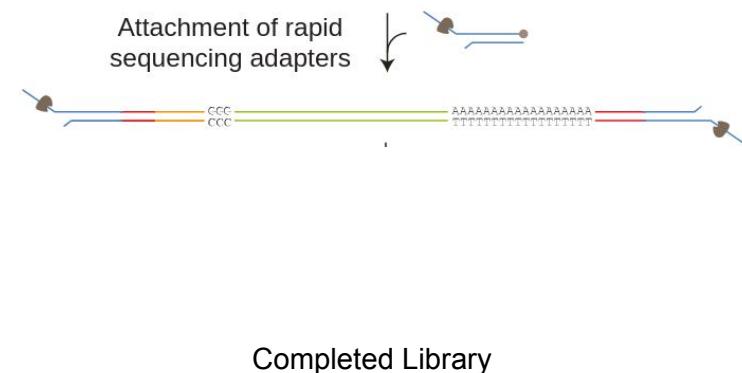
Image: Nanopore

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Common Gotchas

Always perform flow cell QC prior lib prep, ensure you have enough pores!

Image: Nanopore

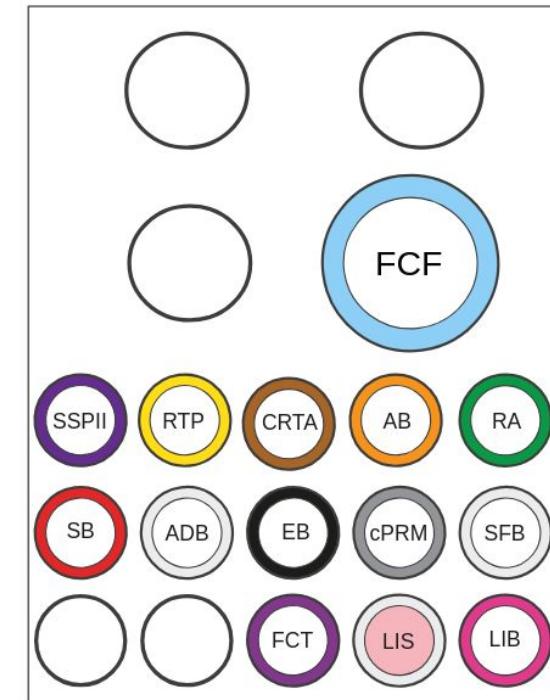
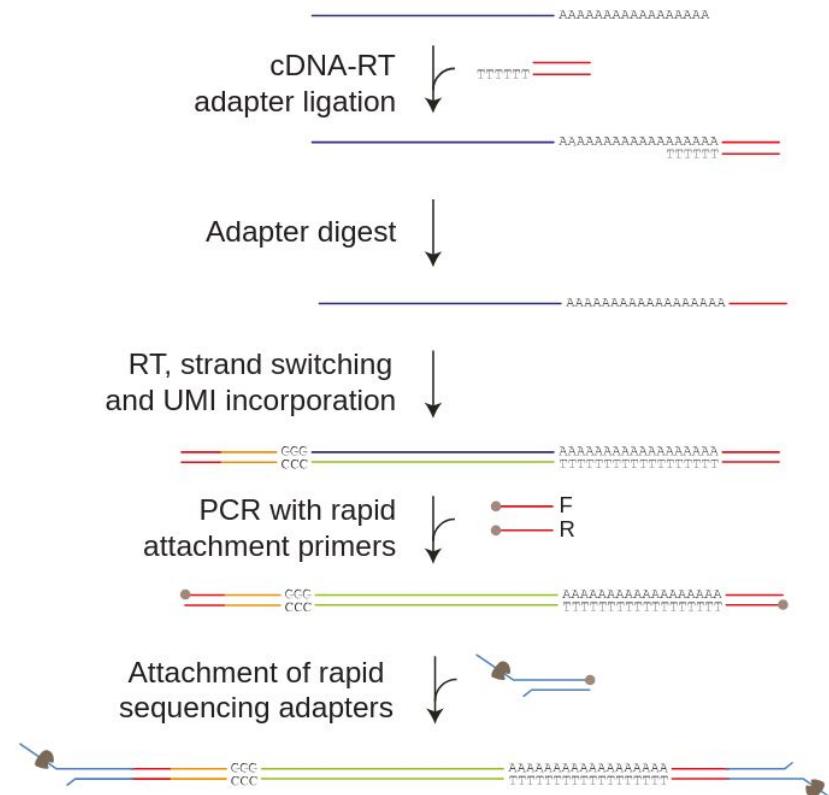
Library preparation

LongTREC

cDNA

cDNA-PCR (R10.4.1)

Nanopore

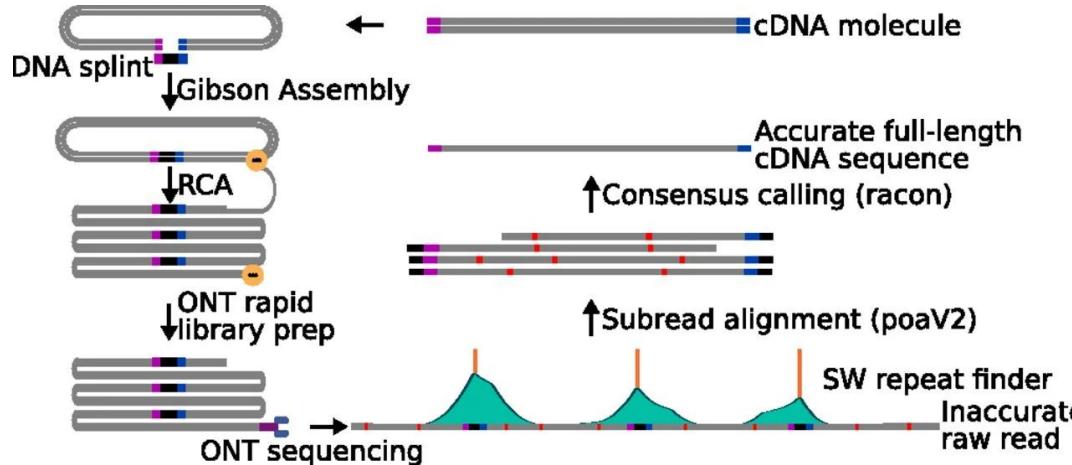


Complete library workflow for cDNA sequencing with cDNA-PCR kit

Image: Nanopore

cDNA

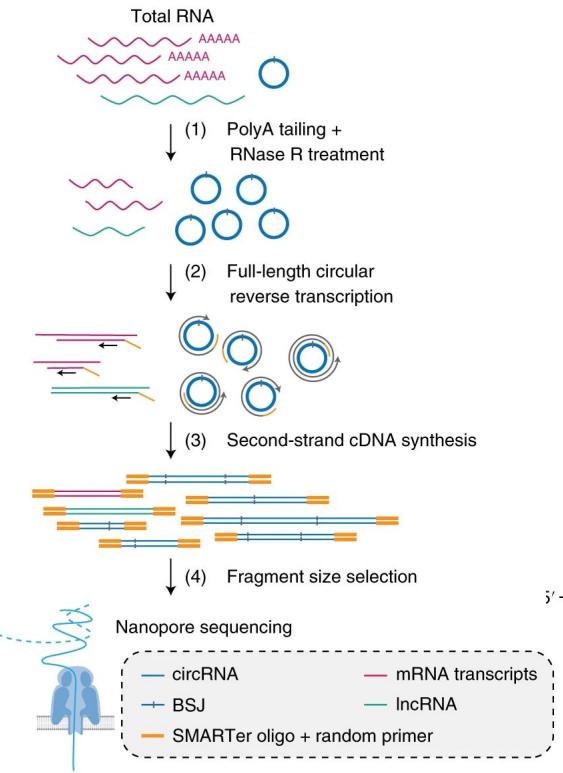
cDNA-PCR (R10.4.1)



Mandalorian was used to analyze the R2C2 reads of the 96 B cells and identified several surface receptor isoforms expressed by 96 distinct single human B cells.



R. Volden, et. al. Improving nanopore read accuracy with the R2C2 method enables the sequencing of highly multiplexed full-length single-cell cDNA. <https://doi.org/10.1073/pnas.1806447115> (2018).



Nanopore
Community

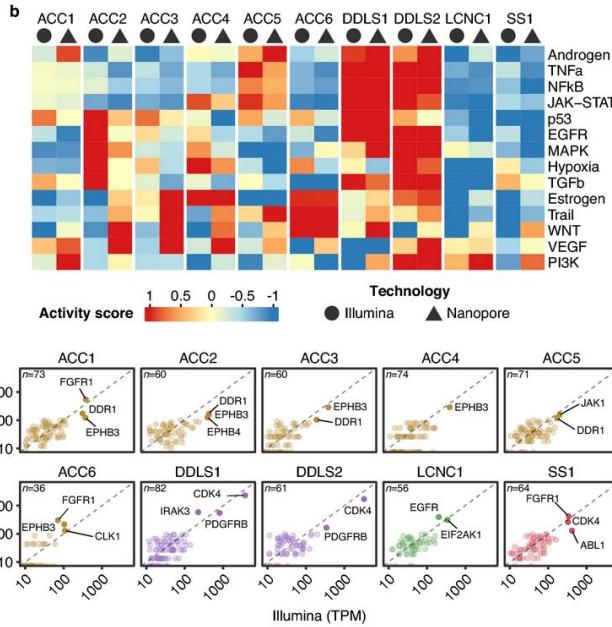
Circular reverse transcription and size selection achieves a 20-fold higher enrichment of circRNAs from total RNA compared to previous methods.

Zhang, J., Hou, L., Zuo, Z. et al. Comprehensive profiling of circular RNAs with nanopore sequencing and CIRI-long. *Nat Biotechnol* 39, 836–845 (2021). <https://doi.org/10.1038/s41587-021-00842-6>

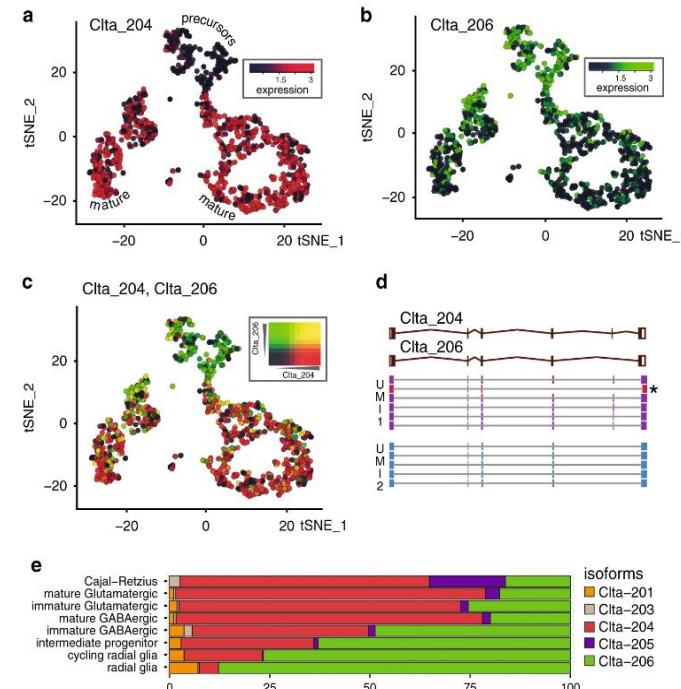
Library preparation

LongTREC

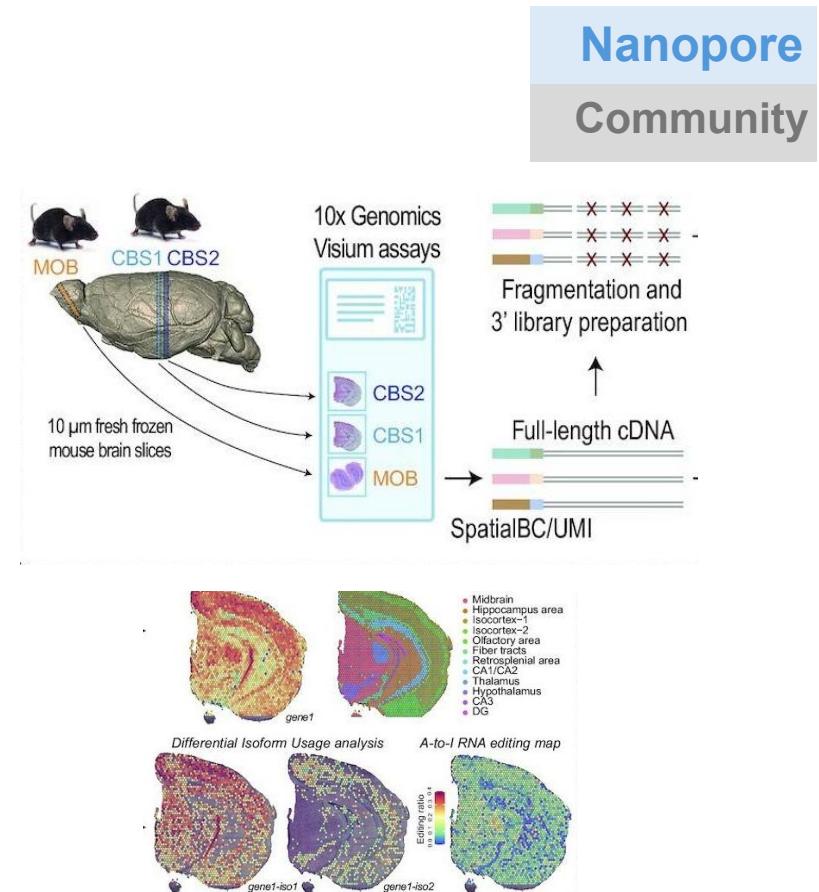
cDNA cDNA-PCR (R10.4.1)



Shallow nanopore RNA-seq enables rapid and biologically meaningful transcriptome profiling of tumors (Mock et al 2023)



Nanopore scRNA-seq reveals transcript isoform diversity (Lebrigand et al 2020).



SiT can be used to profile isoform expression and sequence heterogeneity in different areas of the tissue (Lebrigand et al 2023).

mRNA (Native RNA)

Direct RNA sequencing (RNA001, RNA002)

Nanopore

Description	A sequencing kit optimised for sequencing native RNA with improved output and accuracy.
Kit	Direct RNA Sequencing Kit
Multiplex	Under development
Preparation time	135 minutes
Input	300 ng poly(A)+ RNA or 1 ug total RNA
Output	2 GB cDNA (MinION); 30 million cDNA (PromethION)

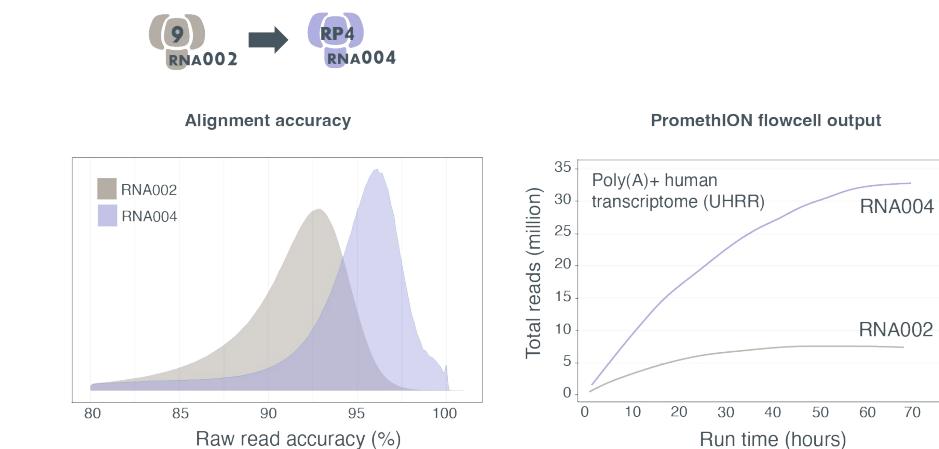
Image: Nanopore

mRNA (Native RNA)

Direct RNA sequencing (RNA004)

Nanopore

Description	A sequencing kit optimised for sequencing native RNA with improved output and accuracy.
Kit	Direct RNA Sequencing Kit
Multiplex	Under development
Preparation time	135 minutes
Input	300 ng poly(A)+ RNA or 1 ug total RNA
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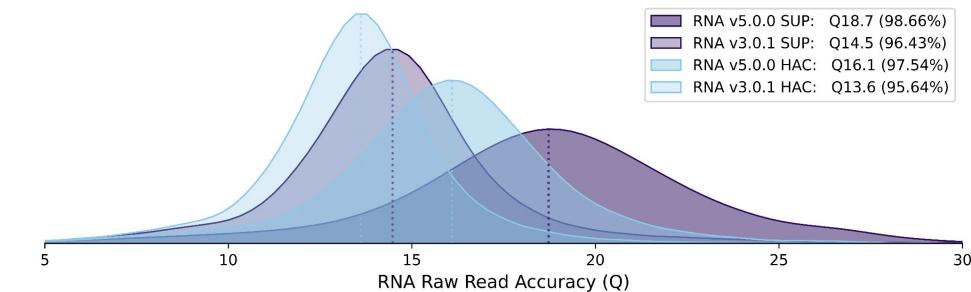
Overall quality and throughput significantly improved in RNA004. Libprep step is the same.

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Latest transformer model and RP4 pores
deliver higher accuracy

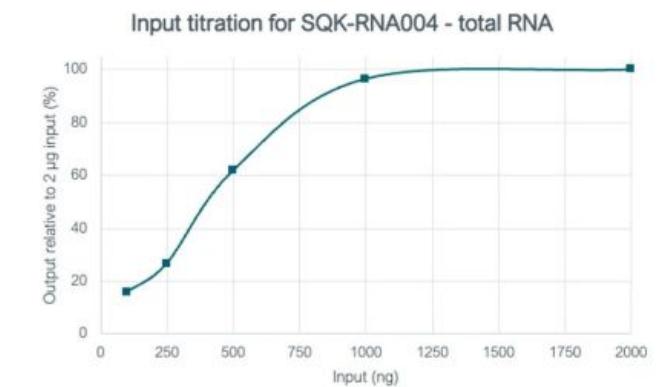
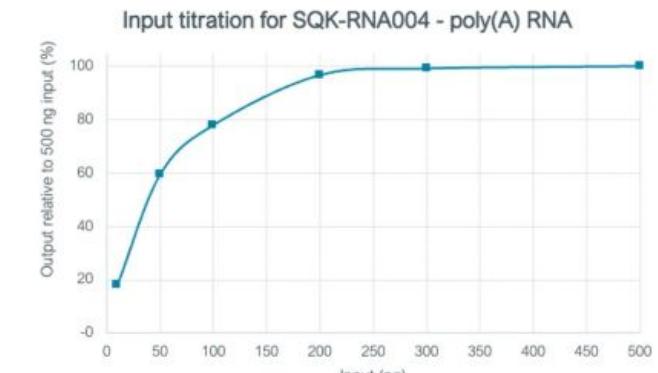
Image: Nanopore

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Direct RNA kit support poly A+ RNA or total RNA

Image: Nanopore

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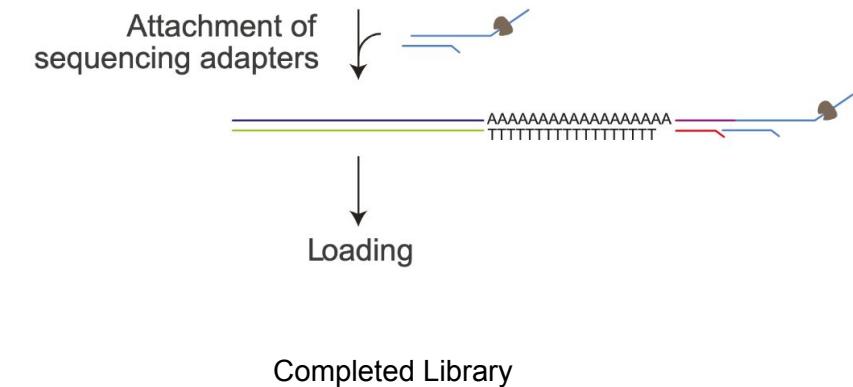
Image: Nanopore

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Common Gotchas

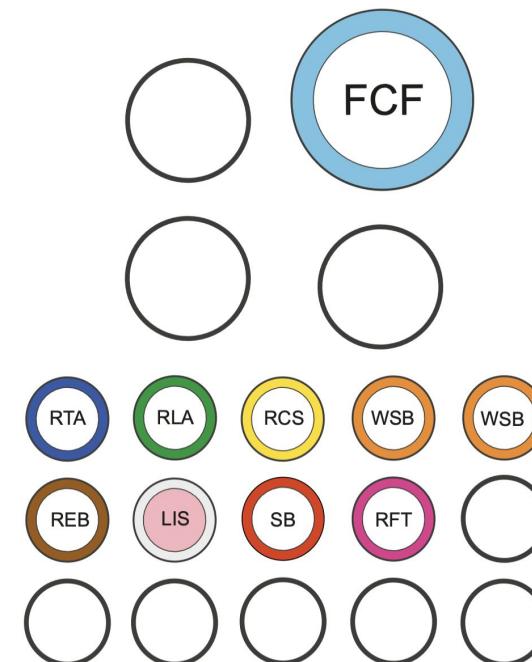
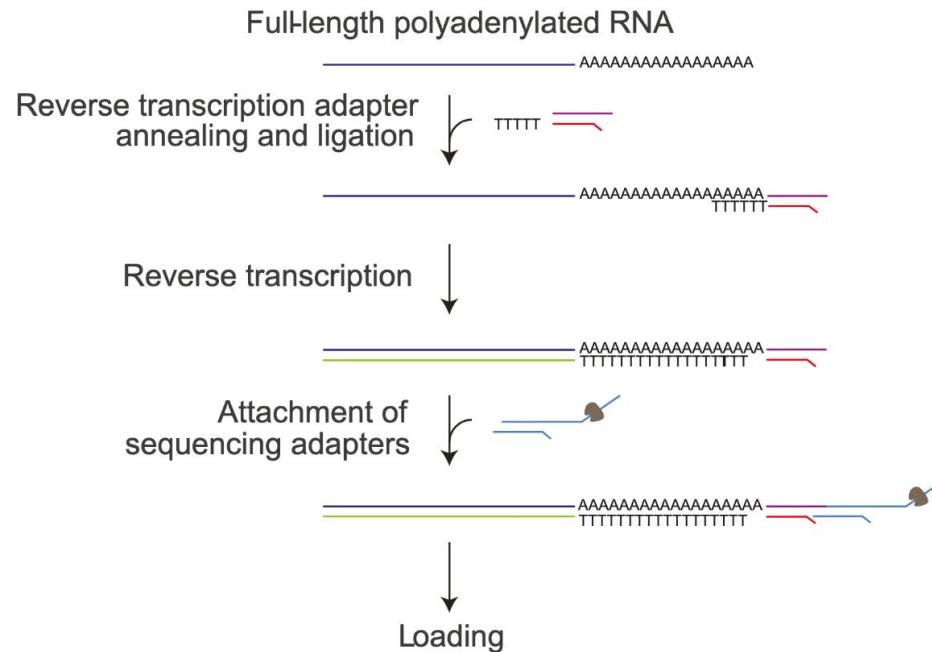
Always perform flow cell QC prior lib prep, ensure you have enough pores!

Image: Nanopore

mRNA (Native RNA)

Direct RNA sequencing (RNA004)

Nanopore



FCF : Flow Cell Flush
 RTA: RT Adapter
 RLA : RNA Ligation Adapter
 RCS : RNA CS
 WSB : Wash Buffer

SB : Sequencing Bu ffer
 RFT : RNA Flush Tether
 REB : RNA Elution Buffer
 LIS : Library Solution

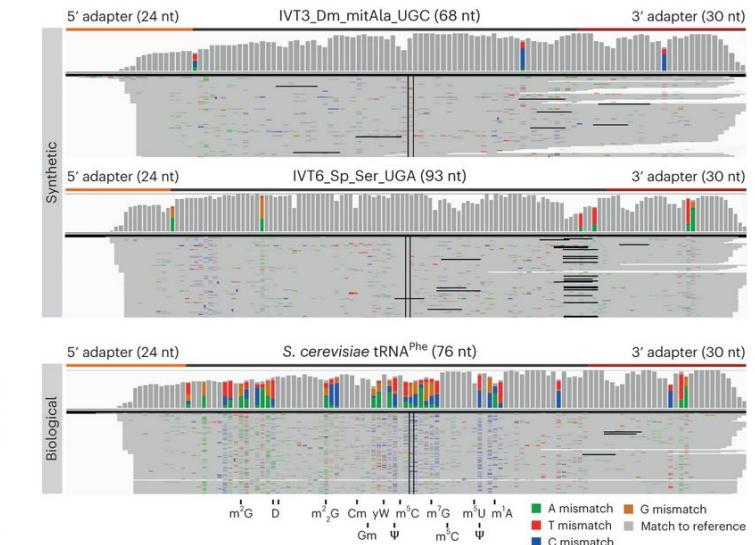
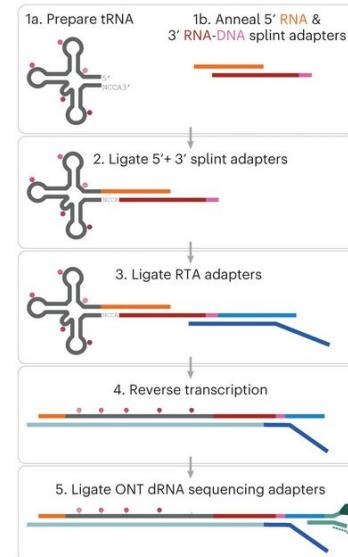
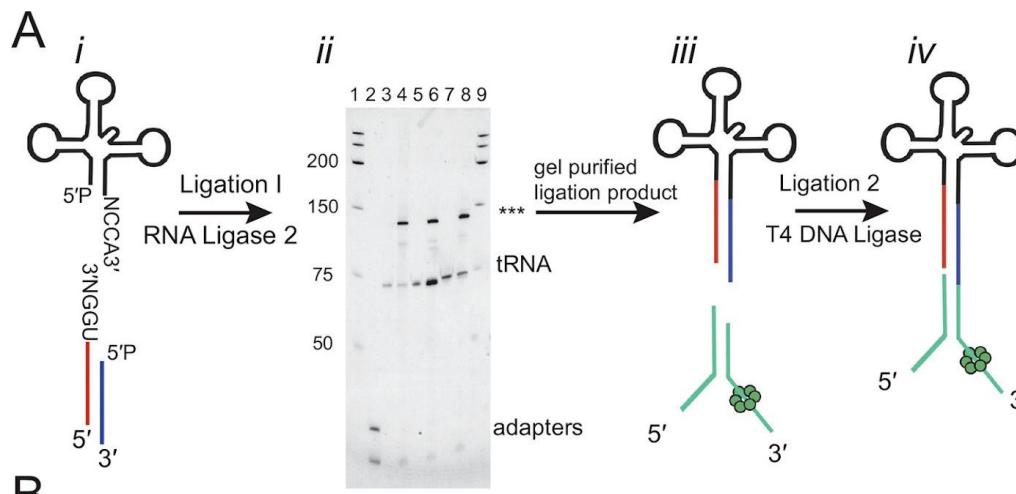
Complete library workflow for Direct RNA sequencing with Direct RNA Sequencing kit

Image: Nanopore

tRNA (Native RNA)

Direct RNA sequencing (RNA004)

Nanopore
Community



Nanopore sequencing detected all 43 expected isoacceptors in total *E. coli* MRE600 tRNA

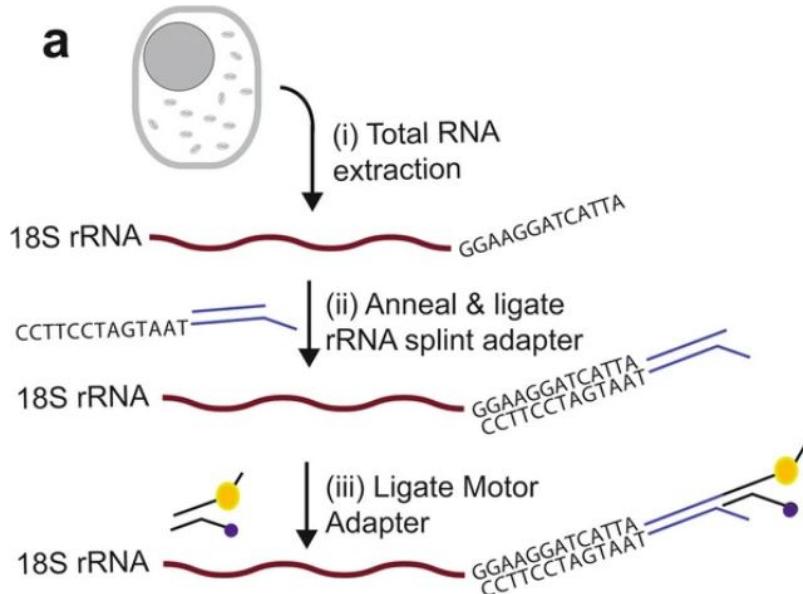
Thomas, N. K., Poodari, V. C., Jain, M., Olsen, H. E., Akeson, M., & Abu-Shumays, R. L. (2021). Direct Nanopore Sequencing of Individual Full Length tRNA Strands. *ACS nano*, 15(10), 16642–16653. <https://doi.org/10.1021/acsnano.1c06488>

Nano-tRNAsq can efficiently sequence both IVT and native tRNA populations

Lucas, M.C., Pryszcz, L.P., Medina, R. et al. Quantitative analysis of tRNA abundance and modifications by nanopore RNA sequencing. *Nat Biotechnol* 42, 72–86 (2024). <https://doi.org/10.1038/s41587-023-01743-6>

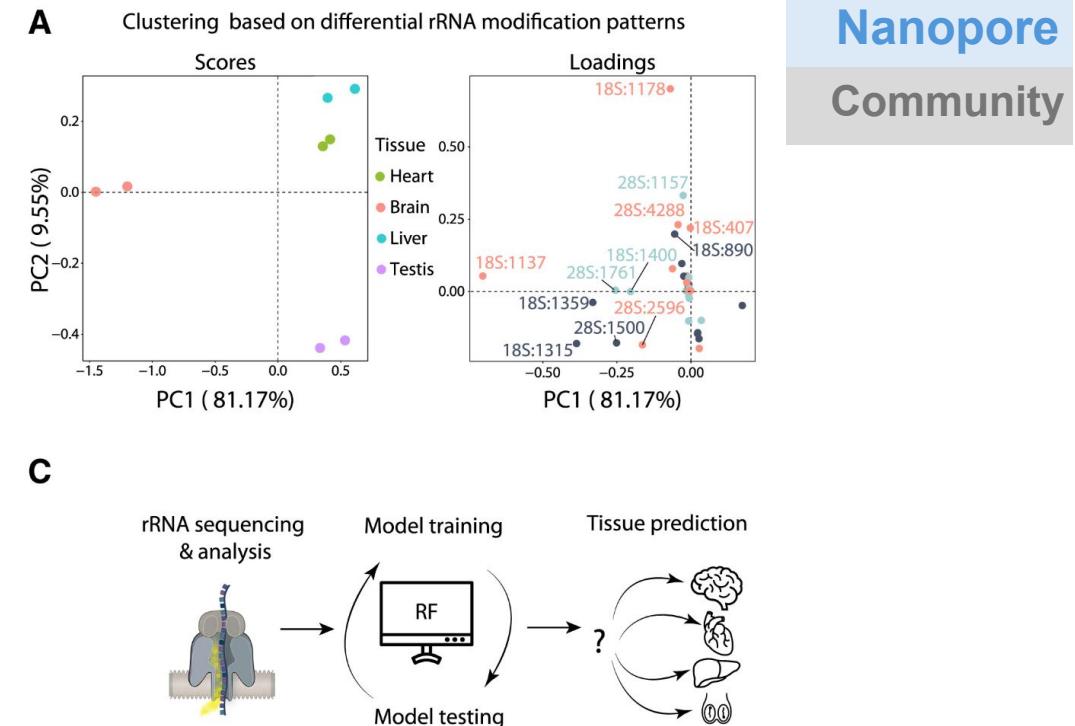
rRNA (Native RNA)

Direct RNA sequencing (RNA004)



Differential rRNA modification patterns can be used to predict tissue types and developmental stages

Jain, M., Olsen, H.E., Akeson, M., Abu-Shumays, R. (2021). Adaptation of Human Ribosomal RNA for Nanopore Sequencing of Canonical and Modified Nucleotides. In https://doi.org/10.1007/978-1-0716-1374-0_4



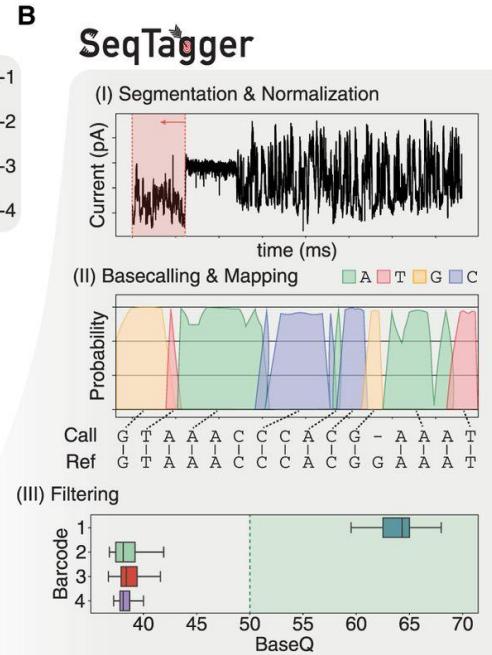
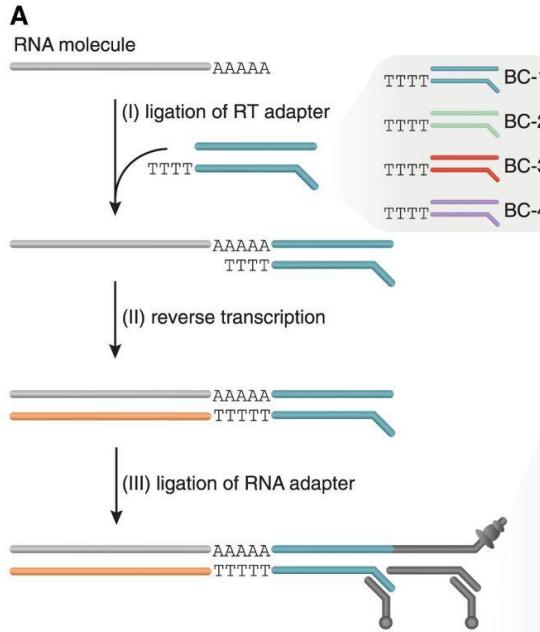
Differential rRNA modification patterns can be used to predict tissue types and developmental stages

Milenkovic, et al. (2025). Epitranscriptomic rRNA fingerprinting reveals tissue-of-origin and tumor-specific signatures. Molecular cell, 85(1), 177–190.e7. <https://doi.org/10.1016/j.molcel.2024.11.014>

Nanopore
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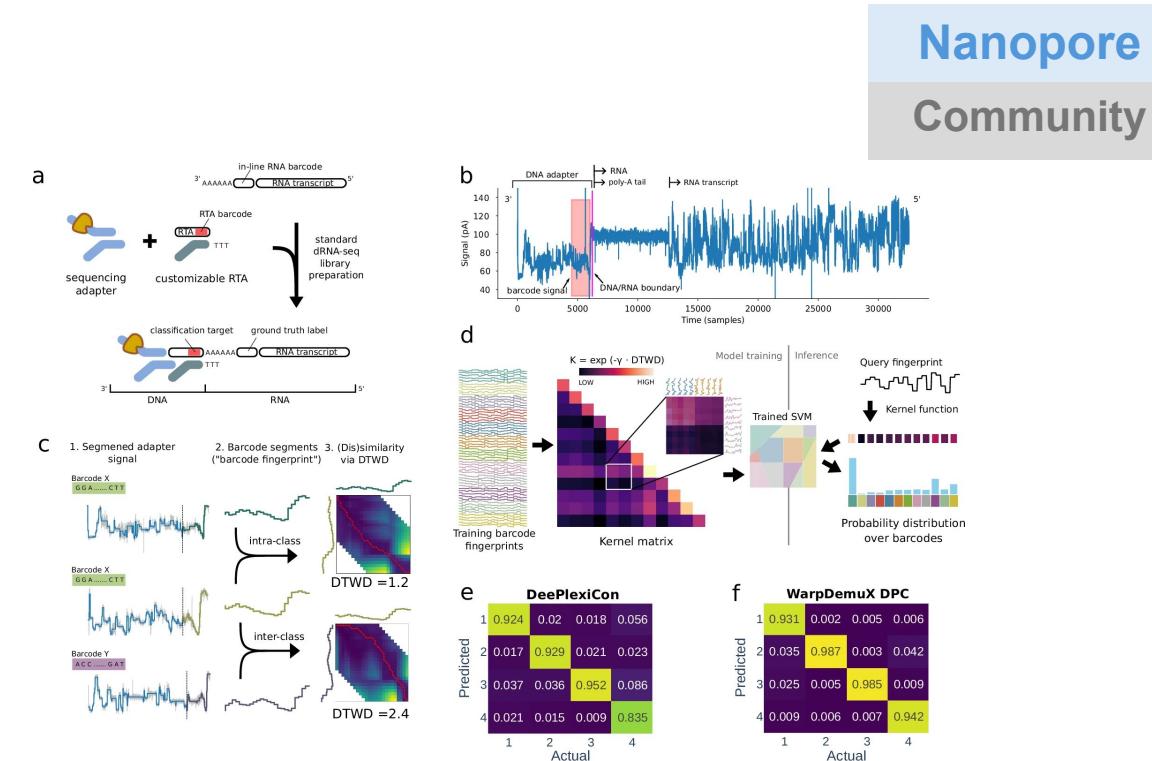
Multiplexing

Direct RNA sequencing (RNA004)



SeqTagger, a rapid and robust method that can demultiplex DRS data sets with 99% precision and 95% recall.

Thomas, N. K., Poodari, V. C., Jain, M., Olsen, H. E., Akeson, M., & Abu-Shumays, R. L. (2021). Direct Nanopore Sequencing of Individual Full Length tRNA Strands. *ACS nano*, 15(10), 16642–16653. <https://doi.org/10.1021/acsnano.1c06488>

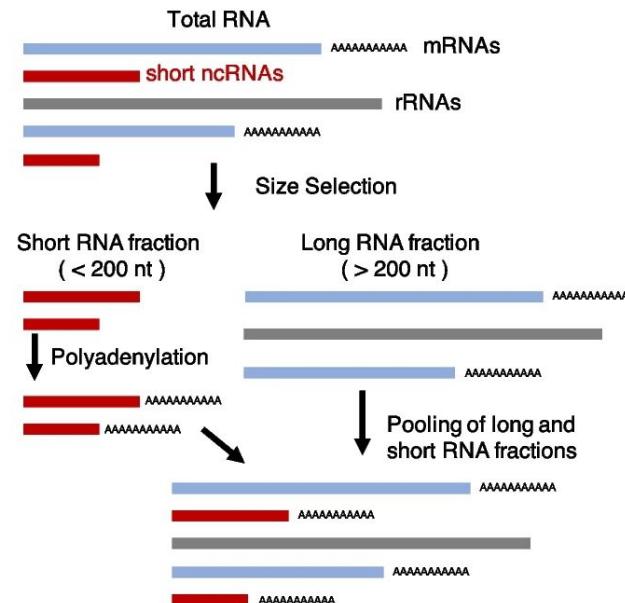


Rapid phenotypic profiling of different SARS-CoV-2 viruses through multiplexed sequencing of longitudinal samples on a single flowcell, identifying systematic differences in transcript abundance and poly(A) tail lengths

van der Toorn, W., Bohn, P., Liu-Wei, W. et al. Demultiplexing and barcode-specific adaptive sampling for nanopore direct RNA sequencing. *Nat Commun* 16, 3742 (2025). <https://doi.org/10.1038/s41467-025-59102-9>

Improving Representation Direct RNA sequencing (RNA004)

Nanopore
Community



NERD-seq expands representation of frequently modified non-coding RNAs, such as snoRNAs, snRNAs, scRNAs, srpRNAs, tRNAs, and rRFs.

Saville, L., Wu, L., Habtewold, J. et al. NERD-seq: a novel approach of Nanopore direct RNA sequencing that expands representation of non-coding RNAs. *Genome Biol* 25, 233 (2024).
<https://doi.org/10.1186/s13059-024-03375-8>

Section 1.1

Bonus

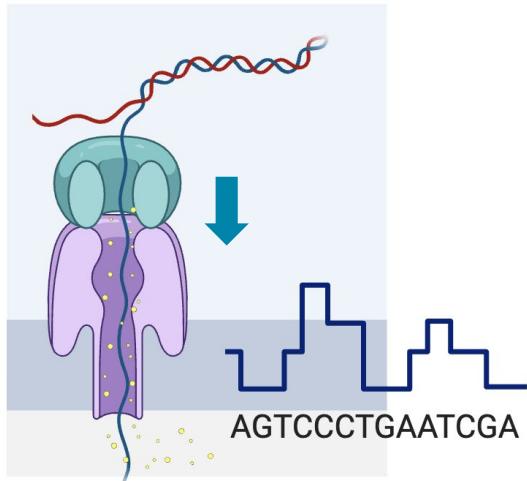
Library preparation strategies for long-reads RNA-seq experiments

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore

Community



Adaptive sampling can be used to enrich or deplete target transcripts.

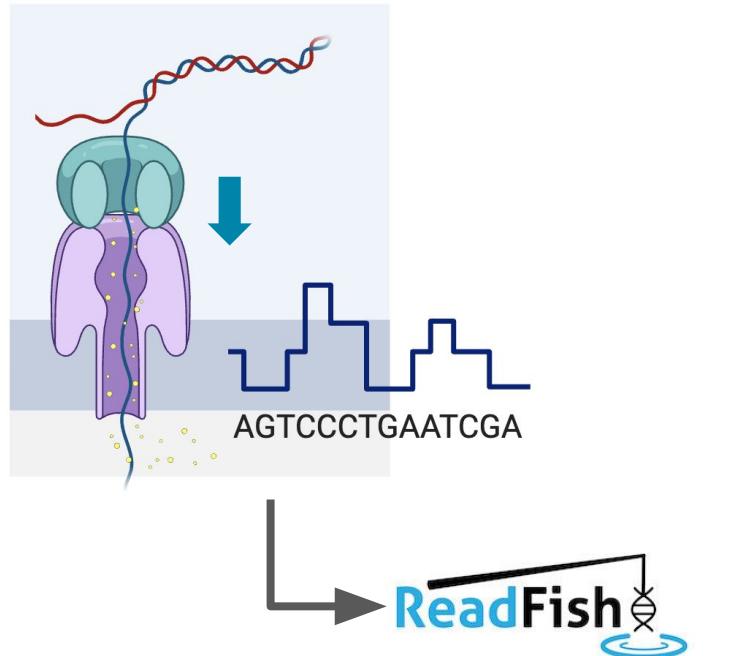
Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore

Community



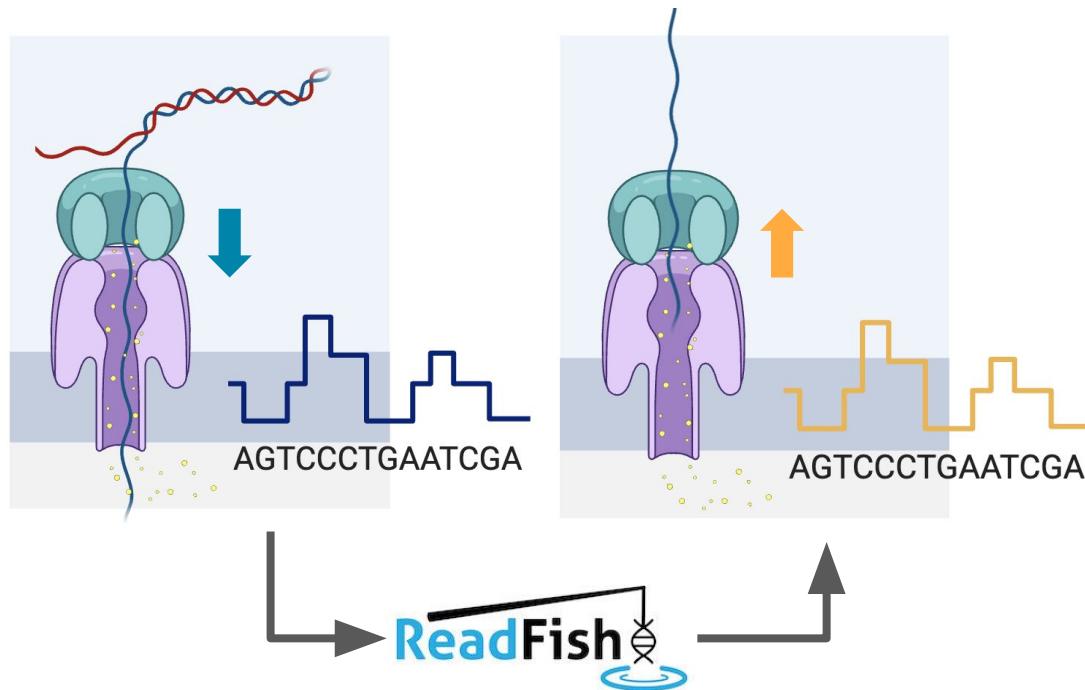
Adaptive sampling can be used to enrich or deplete target transcripts.

Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore
Community



Adaptive sampling can be used to enrich or deplete target transcripts.

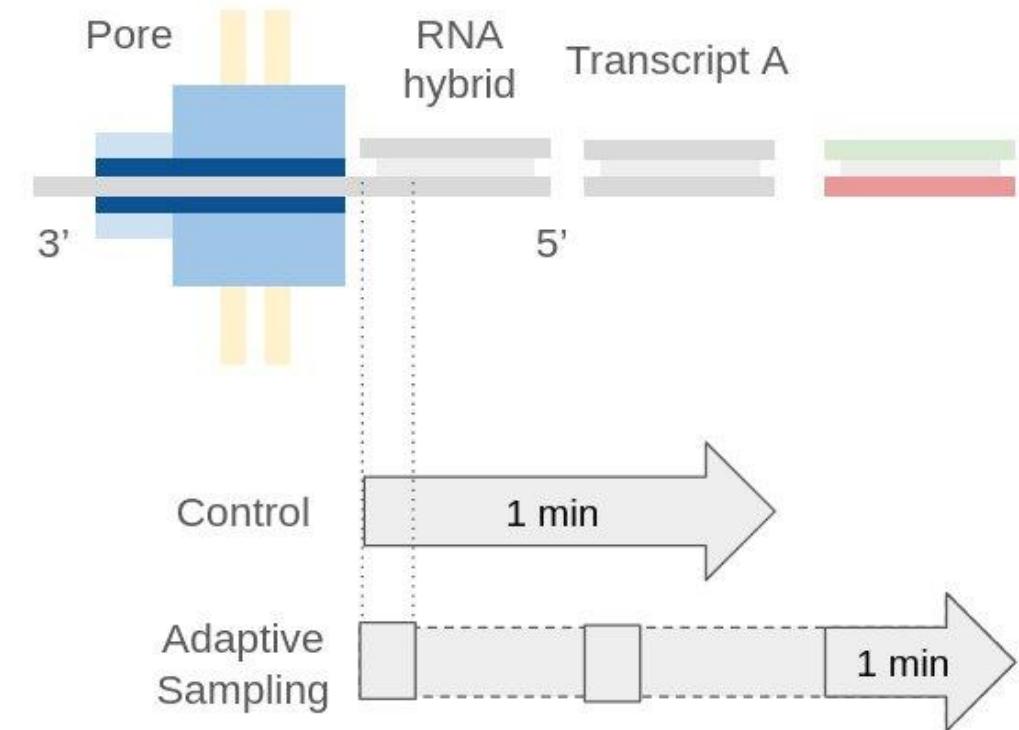


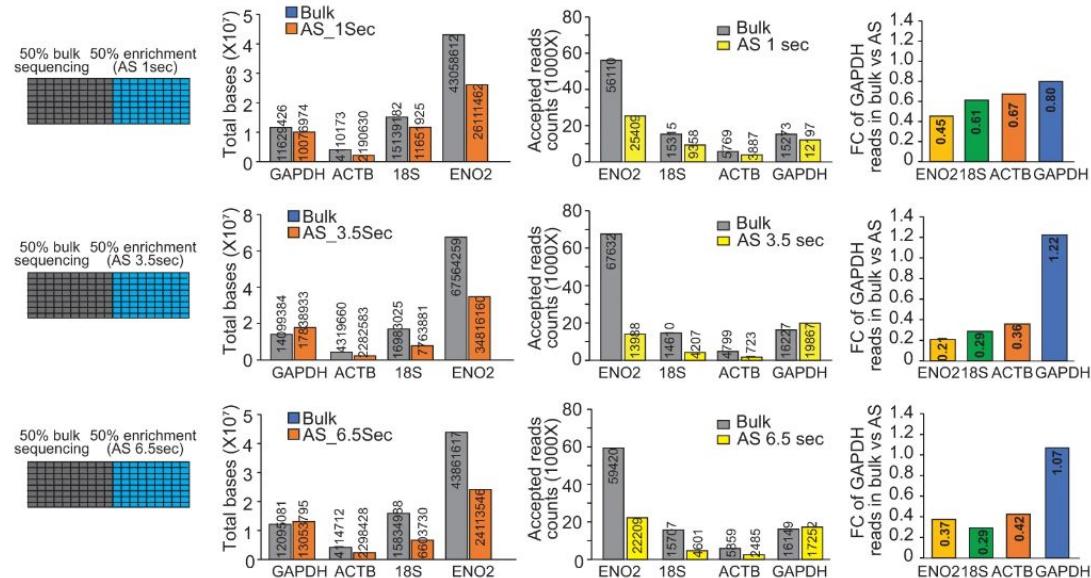
Image: Nanopore

Adaptive Sampling

Direct RNA sequencing (RNA004)

Nanopore

Community



Adaptive sampling performed on a pool of in vitro transcribed RNAs resulted in a net increase of 22-30% in the proportion of transcripts of interest in the population.

Wang, J., et al. (2024). Direct RNA sequencing coupled with adaptive sampling enriches RNAs of interest in the transcriptome.

<https://doi.org/10.1038/s41467-023-44656-3>

Image: Nanopore

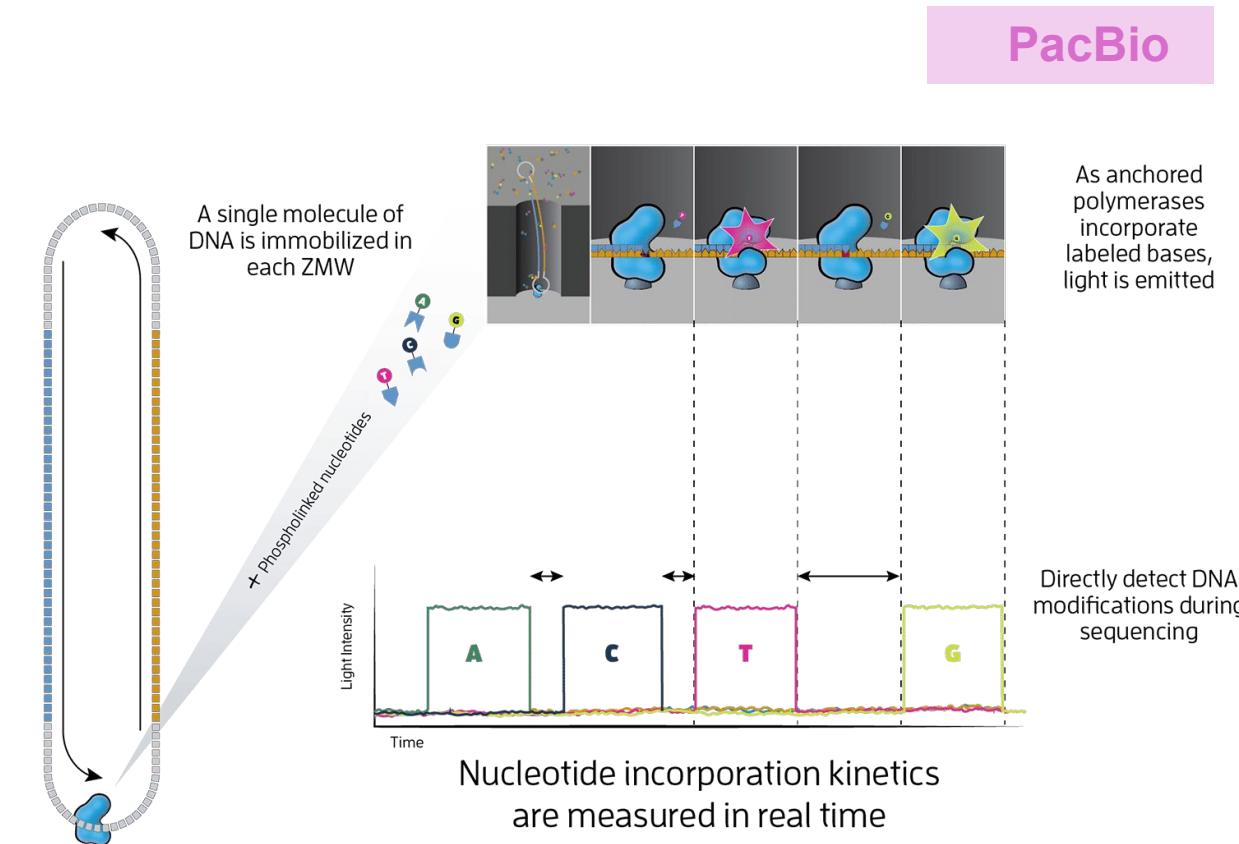
Section 2

Basecalling

Raw data formats and Basecalling

PacBio

Official tool	ICS v13.3 (Revio)
	Base calling HiFi read generation with DeepConsensus Methylation calling Barcode demultiplexing BAM file generation



PacBio

Official tool	ICS v13.3 (Revio)
	Base calling HiFi read generation with DeepConsensus Methylation calling Barcode demultiplexing BAM file generation

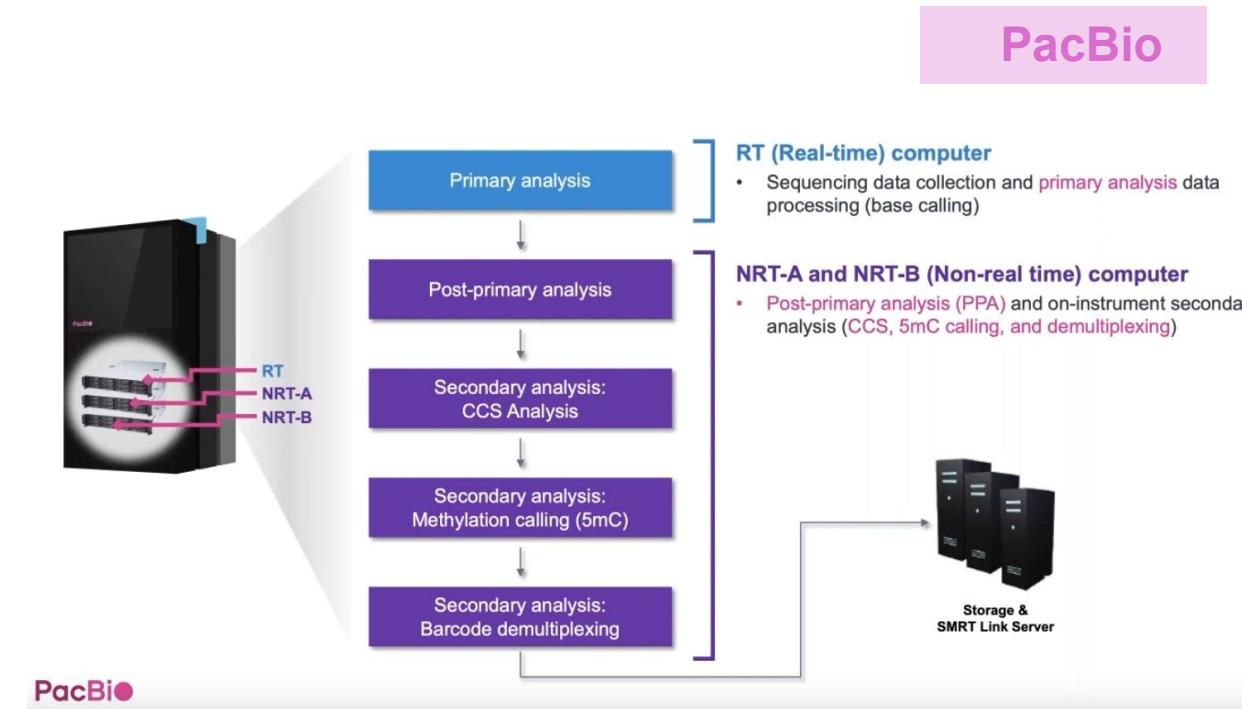


Image: Nanopore

Nanopore Basecalling

cDNA-PCR (R10.4.1)

Nanopore

Official tool	MinKNOW (dorado) Dorado standalone
Model	dna_r10.4.1_e8.2_400bps_fast@v5.2.0 dna_r10.4.1_e8.2_400bps_hac@v5.2.0 dna_r10.4.1_e8.2_400bps_sup@v5.2.0
Resource	PromethION Tower, P2I Professional/Gaming Laptop/Tower

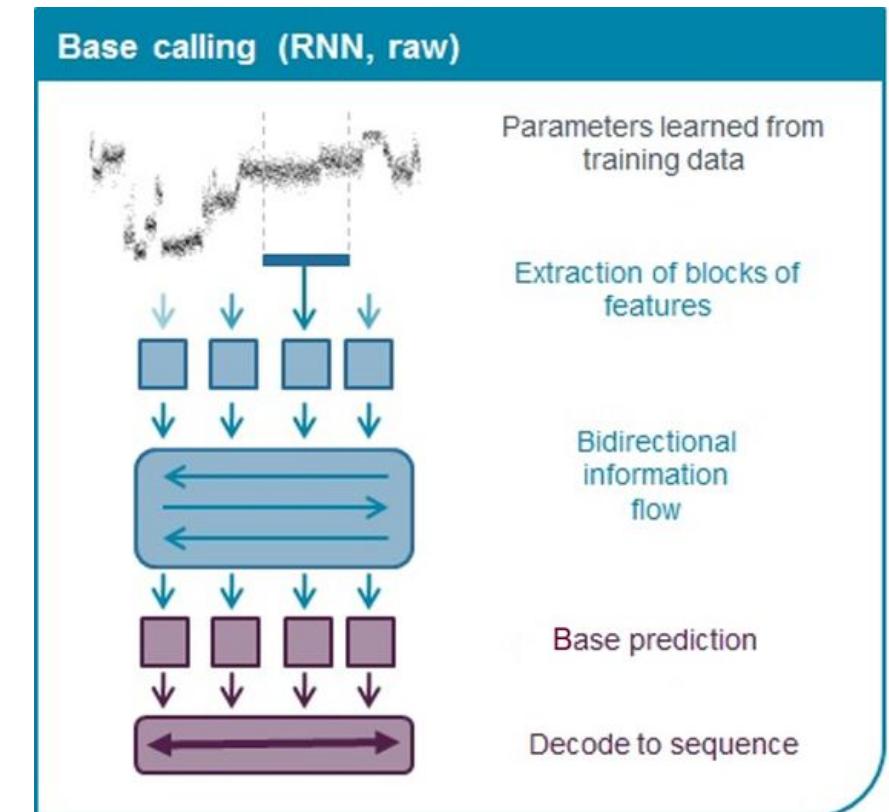


Image: Nanopore

Nanopore Basecalling

Direct RNA sequencing (RNA004)

Nanopore

Official tool	MinKNOW (dorado) Dorado standalone Remora
Model and mods	rna004_130bps_fast@v5.2.0 rna004_130bps_hac@v5.2.0 m5C m6A_DRACH inosine_m6A pseU rna004_130bps_sup@v5.2.0 m5C_2OmeC m6A_DRACH inosine_m6A_2OmeA pseU_2OmeU 2OmeG
Resource	PromethION Tower, P2I Professional/Gaming Laptop/Tower

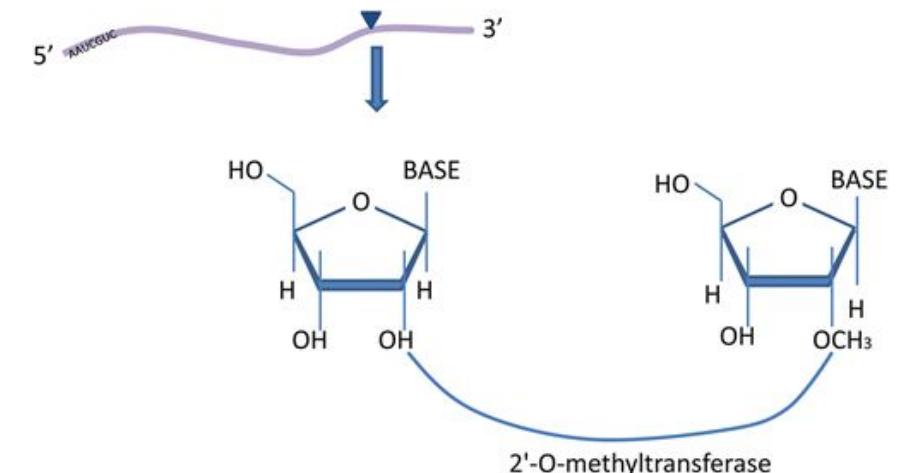
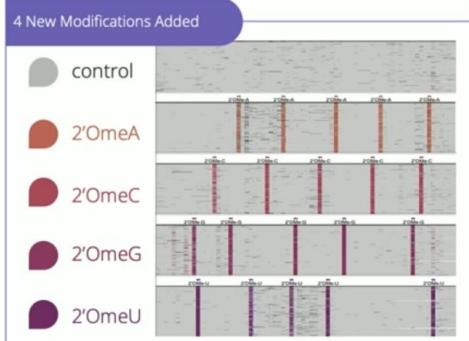
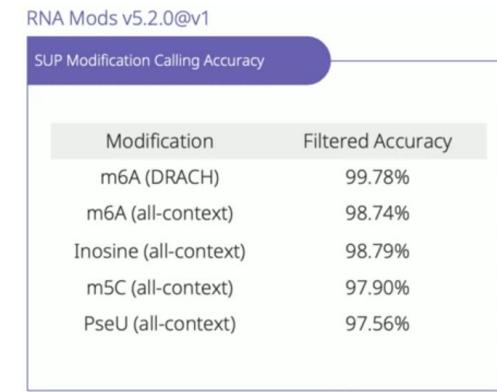


Image: Nanopore

Nanopore Modcalling

Direct RNA sequencing (RNA004)

Nanopore

Community tool

EpiNano (Liu et al, 2019)

m6A

Nanocompose (Leger, 2021)

De novo in comparison with IVT

m6anet (Hendra et al, 2022)

m6A

NanoRMS (Begik, 2022)

De novo, modification sites

m6ABasecaller (Cruciani, 2023)

m6A

and many more ...

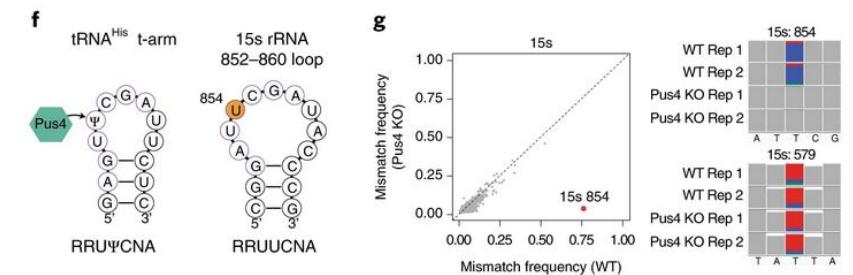
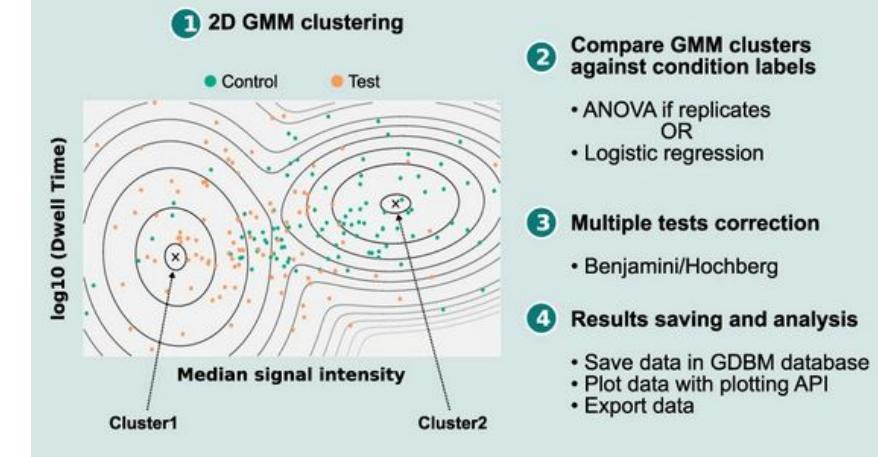


Image: Nanopore

Questions

1. What method kinnex was based on?
2. What improved throughput in Kinnex kit?
3. Why RT is performed in dRNA sequencing?
4. What mods supported by dorado?

Thank You!



For more information about the LongTREC Summer School:

<https://longtrec.eu>