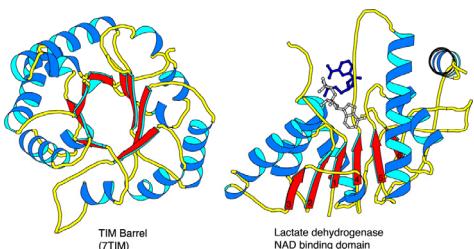


Protein Structure Determination



How are these structures determined?

Why Bother With Structure?

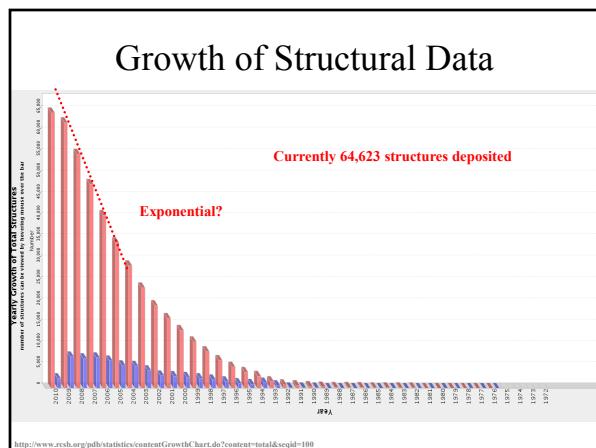
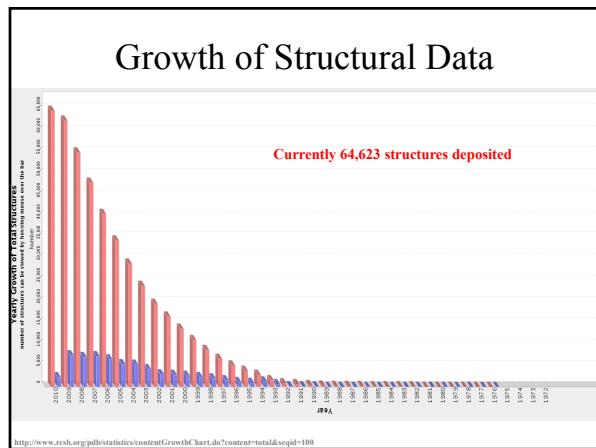
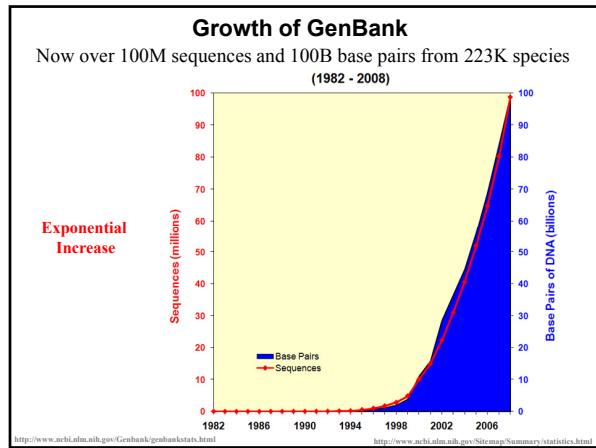
- The amino acid sequence of a protein contains interesting information.
- A protein sequence can be compared to other protein sequences to establish its **evolutionary relationship** to other proteins and protein families.
- However, for the purposes of understanding **protein function**, the 3D structure of the protein is far more useful than the sequence.

Protein Sequences Far Outnumber Structures

- Only a small number of protein structures have been experimentally determined.

PDB ~64,623 protein structures
Genebank ~61,132,599 sequences

- Of the 64,623 structures, only **15,702** are dissimilar in sequence (<30% ID).



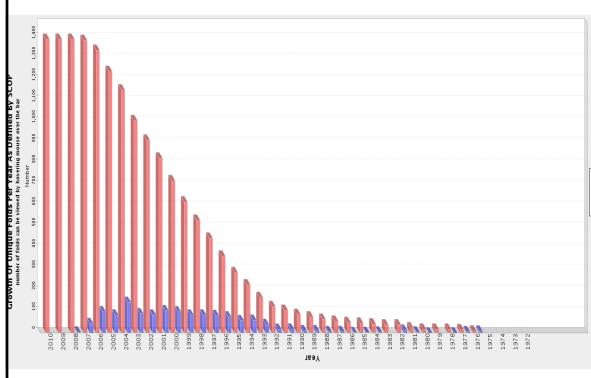
Structural Proteomics

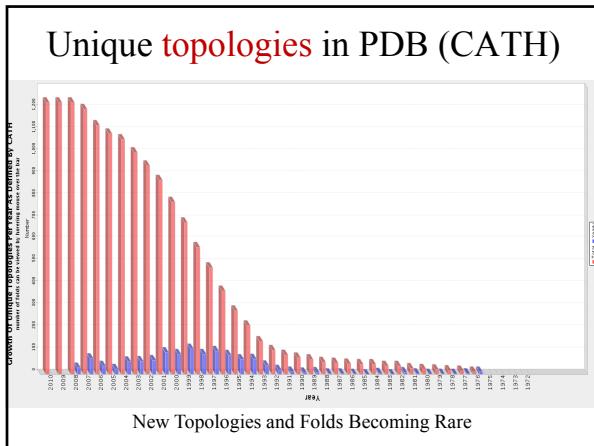
- Use experimentally determined structures to **model** the structures of similar proteins
 - Threading
 - Homology Modeling
 - Fold recognition
- **Avoids *Ab initio* structure determination**
- Need representative protein structures for the total repertoire of **protein folds**
- Provide 3D portraits for all proteins in an organism
- Goal: Use structure to infer function.
 - More sensitive than primary sequence comparisons

Redundancy in PDB (20 April 10)

Sequence identity	Number of non-redundant chains
90%	25615
70%	23116
50%	20306
30%	15702

Unique **folds** in PDB (SCOP)





Structural Genomics

Initiated in 1999 by NIH
Phase I included 9 large centers for high throughput structure determination
Phase I ran from ~2000 - 2005

Goal
The long-range goal of the Protein Structure Initiative (PSI) is to make the three-dimensional atomic-level structures of most proteins **easily obtainable** from knowledge of their corresponding DNA sequences.

<http://www.nigms.nih.gov/psi/mission.html>

Structural Genomics

Benefits

Structural descriptions will help researchers illuminate **structure-function relationships** and thus formulate better hypotheses and design better experiments.

The PSI collection of structures will serve as the starting point for structure-based drug development by permitting faster identification of lead compounds and their optimization.

The design of better therapeutics will result from comparisons of the structures of proteins that are from pathogenic and host organisms and from normal and diseased human tissues.

The PSI collection of structures will assist biomedical investigators in research studies of key biophysical and biochemical problems, such as **protein folding, evolution, structure prediction, and the organization of protein families and folds**.

Technical developments, the availability of reagents and materials, and experimental outcome data in protein production and crystallization will directly benefit all structural biologists and provide valuable assistance to a broad range of biomedical researchers.

Structural Genomics Centers in US

PSI-1 Winners

The Joint Center for Structural Genomics (JCSG)

During PSI-2, the JCSG has contributed to the overall goal of **maximizing structural coverage of protein families** with no structural representation and has continued to develop and disseminate innovative new technologies for structural biology. The JCSG consortium theme is the “central machinery of life”—proteins that are conserved in all kingdoms of life.

The Midwest Center for Structural Genomics (MCSG)

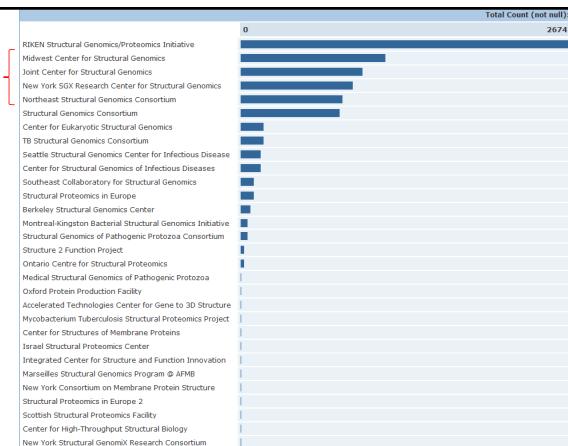
In PSI-2, the multi-institutional consortium is rapidly determining the structures of large numbers of strategically selected proteins using x-ray crystallography both to provide **structural coverage of major protein superfamilies** and to elucidate the entire protein folding space.

The New York Structural Genomics Research Consortium (NYSGRC)

During PSI-2, the consortium's individual project focuses on new targets, principally protein phosphatases and multidomain eukaryotic proteins.

The Northeast Structural Genomics Consortium (NEGS)

In PSI-2, the consortium is solving both prokaryotic and eukaryotic structural representatives from the major domain families constituting the eukaryotic proteome.



Structural Genomics Centers in US

PSI-1 Losers

Center for Eukaryotic Structural Genomics (CESG)

The CESG was founded as a collaborative effort to develop the technologies needed for economical high-throughput structure determination of biologically important eukaryotic proteins and to extend the knowledge of fold-function space. This project also aims to further the research of biologically important proteins in Arabidopsis. The protein structures are being determined via X-ray crystallography or NMR spectroscopy.

The Berkeley Structural Genomics Center (BSCG) The BSCG is pursuing an integrated structural genomics program designed to obtain a near-complete structural complement of two minimal genomes, *Mycoplasma genitalium* and *Mycoplasma pneumoniae*, two related human and animal pathogens. Both NMR spectroscopy and X-ray crystallography are being used for structural determination.

The Seattle Collaboratory for Structural Genomics (SECSG)

The objective of the SECSG is to develop and test experimental and computational strategies for high throughput structure determination of proteins by X-ray crystallography and NMR methods and to apply these strategies to scan the entire genome of an organism at a rapid pace. The eukaryotic organisms, *Caenorhabditis elegans*, *Homo sapiens* and an ancestrally-related prokaryotic microorganism having a small genome, *Pyrococcus furiosus*, have been selected as representative genomes.

Structural Genomics of Pathogenic Protozoa Consortium (SGPP)

The SGPP consortium aims to determine and analyze the structures of a large number of proteins from major global pathogenic protozoa including *Leishmania major*, *Trypanosoma brucei*, *Trypanosoma cruzi* and *Plasmodium falciparum*. These organisms are responsible for the diseases: leishmaniasis, sleeping sickness, Chagas' disease and malaria. X-ray crystallography is being used for structural determination.

The TB Structural Genomics Consortium (TB)

The goal of the TB consortium is to determine the structures of over 400 proteins from *M. tuberculosis*, and to analyze these structures in the context of functional information that currently exists and that is generated by the project. These structures will include about 40 novel folds and 200 new families of protein structures. The protein structures are being determined using X-ray crystallography.

Current PSI Centers

Large-Scale Centers
 Joint Center for Structural Genomics
 Midwest Center for Structural Genomics
 New York SGX Research Center for Structural Genomics
 Northeast Structural Genomics Consortium

Specialized Centers
 Accelerated Technologies Center for Gene to 3D Structure
 Center for Eukaryotic Structural Genomics
 Center for High-Throughput Structural Biology
 Center for Structures of Membrane Proteins
 Integrated Center for Structure and Function Innovation
 New York Consortium on Membrane Protein Structure

Homology Modeling Centers
 Joint Center for Molecular Modeling
 New Methods for High-Resolution Comparative Modeling

Resource Centers
 PSI-Materials Repository
 PSI Knowledgebase

60,000 plasmid clones

2008 Structural Genomics Progress

Status	Total Number of Targets	(%) Relative to "Cloned" Targets	(%) Relative to "Expressed" Targets	(%) Relative to "Purified" Targets	(%) Relative to "Crystallized" Targets
Cloned	61522	100.00	-	-	-
Expressed	39540	64.27	100.00	-	-
Soluble	18221	29.62	46.08	-	-
Purified	14031	22.81	35.49	100.00	-
Crystallized	5616	9.13	14.20	40.03	100.00
Diffraction-quality Crystals	2999	4.73	7.36	20.73	51.80
Diffraction	2429	3.95	6.14	17.31	43.25
NMR Assigned	1051	1.71	2.66	7.49	-
HSQC	1890	3.07	4.78	13.47	-
Crystal Structure	2291	3.72	5.79	16.33	40.79
NMR Structure	953	1.55	2.41	6.79	-
In PDB	2849	4.63	7.21	20.31	35.15
Work Stopped	14137	-	-	-	-
Test Target	4	-	-	-	-
Other	10	-	-	-	-

~40% of structures are from SG in Europe and Asia

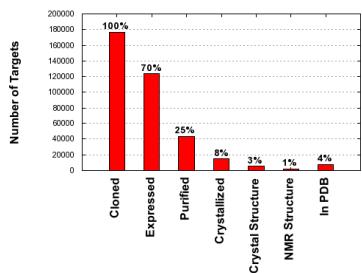
2010 Structural Genomics Progress

Status	Total Number of Targets	(%) Relative to "Cloned" Targets	(%) Relative to "Expressed" Targets	(%) Relative to "Purified" Targets	(%) Relative to "Crystallized" Targets
Cloned	176710	100.0	-	-	-
Expressed	123905	70.1	100.0	-	-
Soluble	47572	26.9	38.4	-	-
Purified	43669	24.7	35.2	100.0	-
Crystallized	14641	8.3	11.8	33.6	100.0
Diffraction-quality Crystals	7708	4.4	6.2	17.7	52.6
Diffraction	6628	3.8	5.3	15.2	45.3
NMR Assigned	2154	1.2	1.7	4.9	-
HSQC	3929	2.2	3.2	9.0	-
Crystal Structure	5215	3.0	4.2	12.0	35.6
NMR Structure	2050	1.2	1.7	4.7	-
In PDB	7569	4.3	6.1	17.4	38
Work Stopped	38962	-	-	-	-
Test Target	93	-	-	-	-
Other	8178	-	-	-	-

~36% of structures are from SG in Europe and Asia

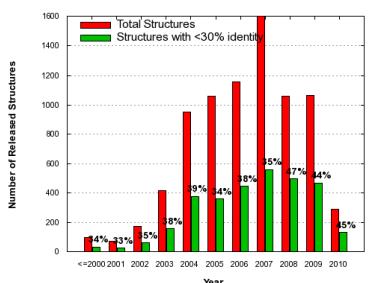
<http://targetdb.ncbi.nlm.nih.gov/TargetStatistics.html>

Project Attrition



<http://targetdb.ncbi.nlm.nih.gov/statistics/TargetStatistics.html>

Unique Folds?



<http://targetdb.ncbi.nlm.nih.gov/statistics/TargetStatistics.html>

Protein Structure Databases

- Where does protein structural information reside?
 - **PDB:**
 - <http://www.rcsb.org/pdb/>
 - **MMDB:**
 - <http://www.ncbi.nlm.nih.gov/Structure/>
 - **FSSP:**
 - <http://www.ebi.ac.uk/dali/fssp/>
 - **SCOP:**
 - <http://scop.mrc-lmb.cam.ac.uk/scop/>
 - **CATH:**
 - http://www.biochem.ucl.ac.uk/bsm/cath_new/

} Jon
} Ingo

RCB Protein Data Bank - Microsoft Internet Explorer provided by RSCB Information Systems

POB ID or Text Search Advanced Search

A Resource for Studying Biological Macromolecules

The PDB archive contains information about experimentally-determined structures of proteins, nucleic acids, and complexes of these molecules. As of Tuesday April 13, 2010 at 5 PM PDT there are 64623 Structures | PoB Statistics

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Molecule of the Month: Concanavalin A and Circular Permutation

In two new NMR structures of an unusually small phytochrome, researchers

Featured Molecules (Previous Features: HOM | PDB)

Molecule of the Month: Concanavalin A and Circular Permutation

Exhibiting at Experimental Biology 2010

Full Article...

Protein Structure Initiative Featured Molecule: Phytochrome

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Tools File Downloads FTP Services

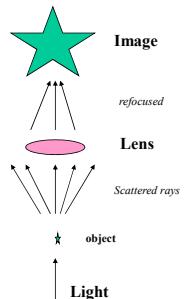
PDB PROTEIN DATA BANK

PDB Contents 20 April 2010

Exp.Method	Proteins	Nucleic Acids	Protein/NA Complexes	Other	Total
X-RAY	52212	1206	2401	17	55836
NMR	7279	896	154	7	8336
ELECTRON MICROSCOPY	195	17	76	0	288
HYBRID	16	1	1	1	19
Other	123	4	4	13	144
Total	59825	2124	2636	38	64623

X-ray Crystallography

Optical Microscope



Atomic Resolution

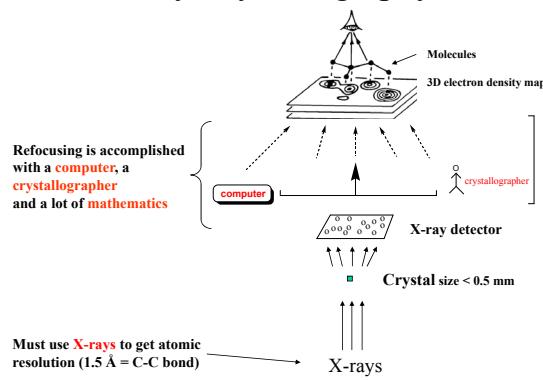
We want to resolve inter-atomic distances ($\sim 1.5 \text{ \AA}$, 0.15 nM)

Visible light has a wavelength of $\sim 500 \text{ nm}$ (5000 \AA)

Electron beam: $\lambda_e \sim 0.001 \text{ \AA}$ (if e^- is moving at c)
Electron velocity is less in electron microscopes
Typical resolution is $\sim 10 \text{ \AA}$, but can be improved

X-ray generators produce photons of $\lambda = 0.5 - 2.5 \text{ \AA}$
Use $\lambda = 1.542 \text{ \AA}$

X-ray Crystallography



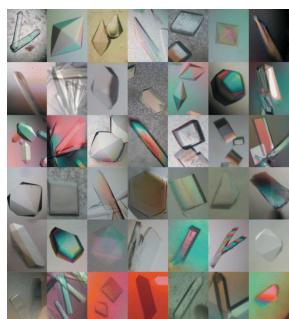
X-Ray Crystallography

1. Make crystals of your protein
0.3-1.0mm in size
Proteins must be in an ordered, repeating pattern.
2. X-ray beam is aimed at crystal and data is collected.
3. Structure is determined from the diffraction data.

X-Ray Crystallography

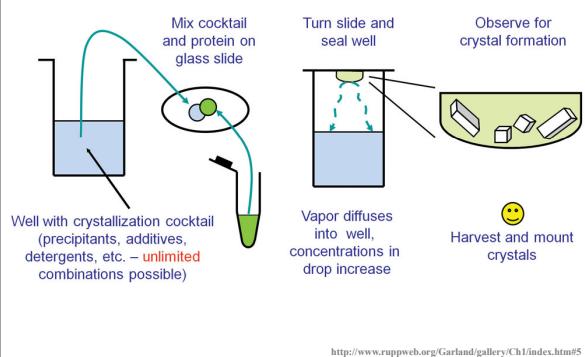
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Protein Crystals



Schmid, M. Trends in Microbiology, 10:s27-s31.

Protein Crystals

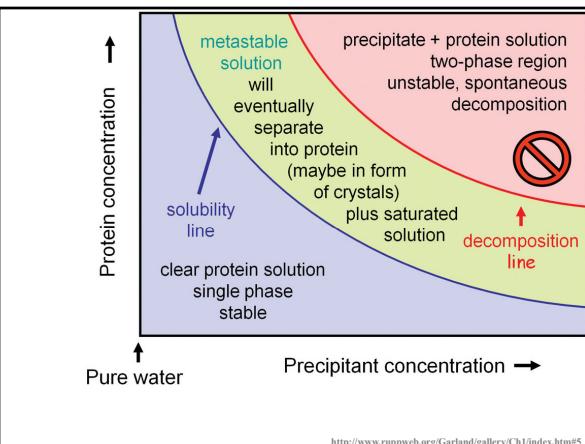


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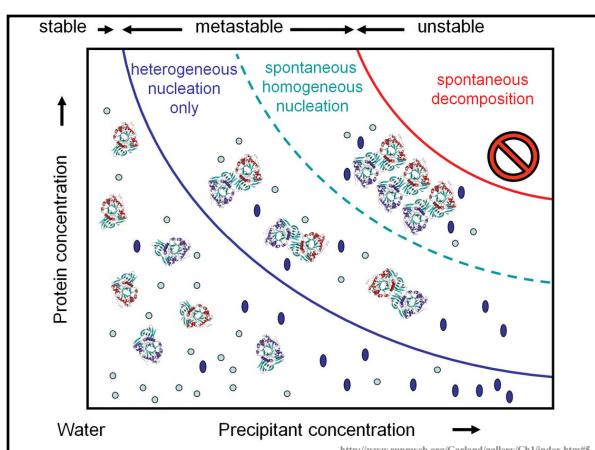
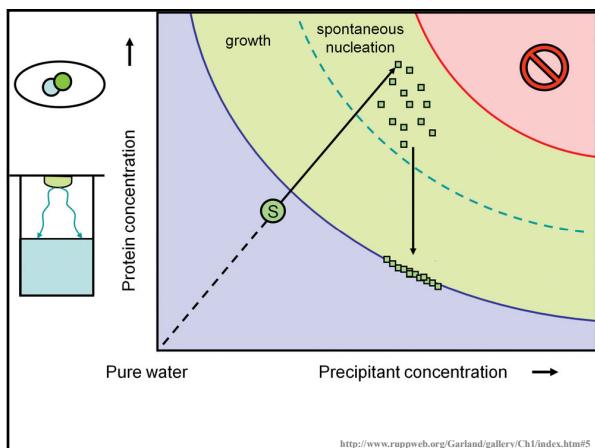
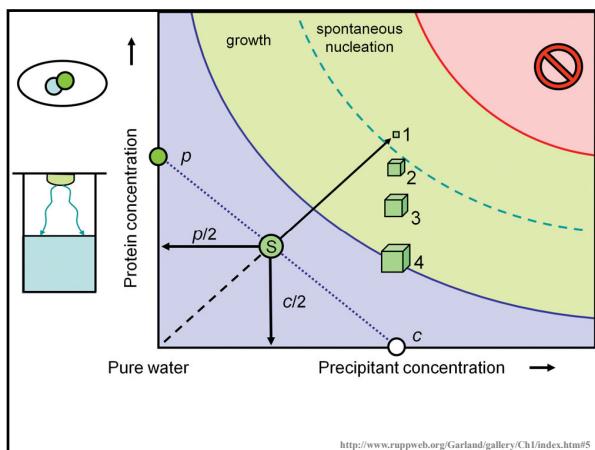
Protein Crystals



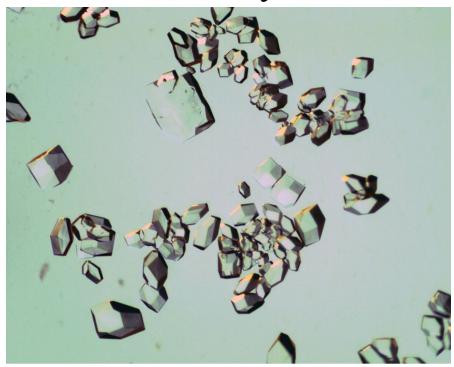
<http://www.ruppweb.org/Garland/gallery/Ch1/index.htm#5>



<http://www.ruppweb.org/Garland/gallery/Ch1/index.htm#5>

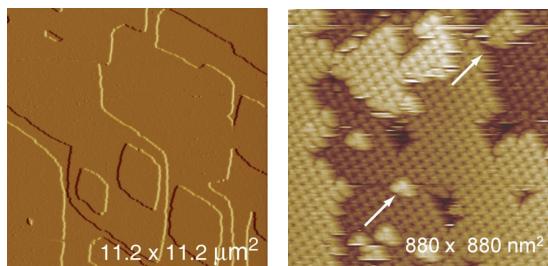


Protein Crystals



<http://www.ruppweb.org/Garland/gallery/Ch1/index.htm#5>

Protein Crystals

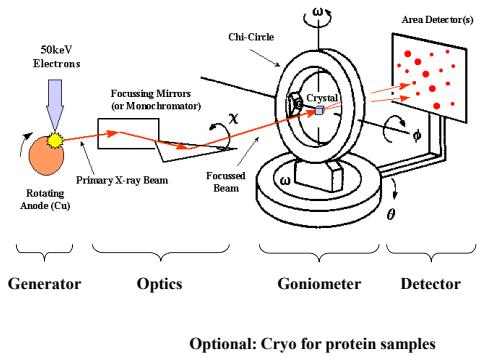


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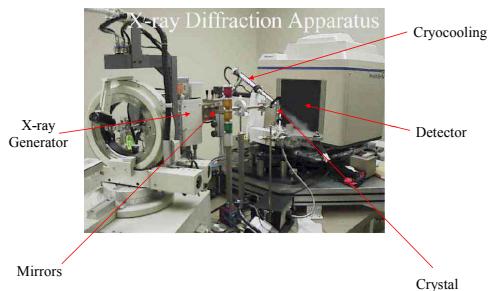
X-Ray Crystallography

1. Make crystals of your protein
0.3-1.0mm in size
Proteins must be in an ordered, repeating pattern.
2. X-ray beam is aimed at crystal and data is collected.
3. Structure is determined from the diffraction data.

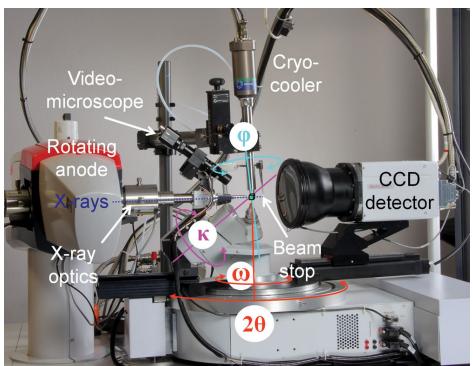
X-Ray Diffraction Experiment



X-ray Crystallography Equipment



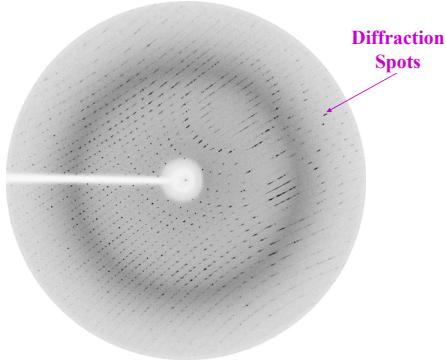
X-ray Crystallography Equipment



X-Ray Crystallography

1. Make crystals of your protein
0.3-1.0mm in size
Proteins must be in an ordered, repeating pattern.
2. X-ray beam is aimed at crystal and data is collected.
3. **Structure is determined from the diffraction data.**

Protein Diffraction Image



Why Spots?

X-ray scattering from individual proteins is diffuse

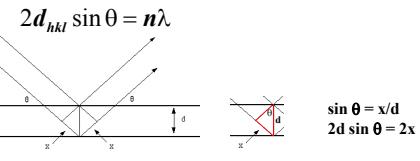
Spots arise from a phenomenon called diffraction that is based on the crystal lattice

Location of reflections indicates **how** an object crystallized
230 possibilities

Intensity of reflections contains information about the **structure** of the object in the crystal

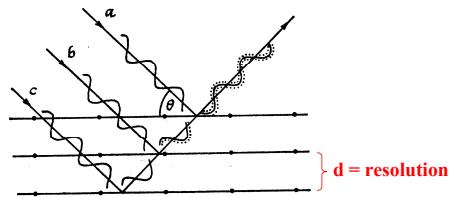
Bragg's Law

Why do we get spots (reflections) and not a diffuse pattern of scattered x-rays?



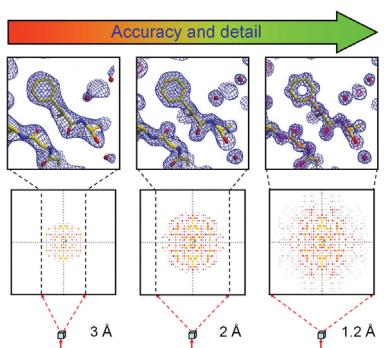
Difference in path (2x) must equal integral number of wavelengths (nλ)

Constructive Interference



- Condition for reflection

Resolution



<http://www.ruppweb.org/Garland/gallery/Ch1/index.htm#5>

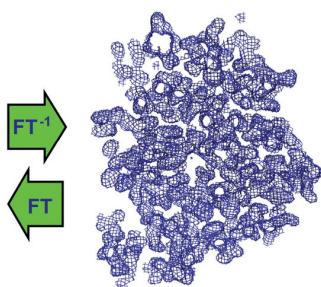
Phase Problem

- The intensities are recorded by the detector
- The phases are lost
- Must have **both** to reconstruct the image (structure)



Phase Problem

h	k	l	$F(hkl)$	$\varphi(hkl)$
2	0	0	228.0	180.0
1	0	1	10.4	90.0
2	0	1	901.8	270.0
1	1	1	367.0	332.1
1	2	3	149.3	37.8
8	9	1	97.9	255.1
7	7	2	111.5	139.7

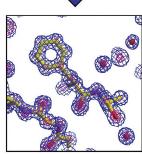


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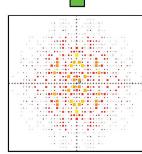


Phase Problem

$$\rho(x, y, z) = \frac{1}{V} \sum_{-h}^h \sum_{-k}^k \sum_{-l}^l F_{hkl} \exp[-2\pi i(hx + ky + lz - \alpha_{hkl})]$$



Electron Density



Intensities

The crystallographic phase problem



The crystallographic phase problem



Solutions to the Phase Problem

Molecular replacement

- Use **known structure** of close homologue
- Rotational and translational search for solution

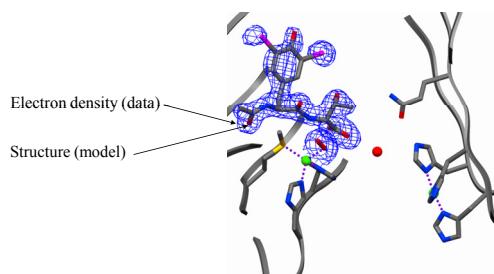
Heavy atom labeling

- Label the protein with **electron dense atoms** (Hg)
- Compare independent datasets collected from native and labeled protein
- Heavy atom substructure provides initial phases

Anomalous diffraction

- Crystal must contain atoms with **absorption edges** between 0.5 and 2.5 Å
- Compare independent datasets collected at pre-edge and post-edge x-ray energies

Model Building



Crystallography Pros/Cons

Advantages

- can be "fast" – down to a few months
- large structures possible (ribosome)
- very low resolution (down to 0.5 Å)
- observables typically > refinement parameters

Disadvantages

- requires crystal formation
- non-physiological conditions
- crystal contacts can limit protein motion

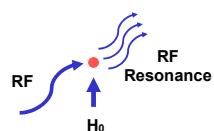
Nuclear Magnetic Resonance

Nuclear Magnetic Resonance

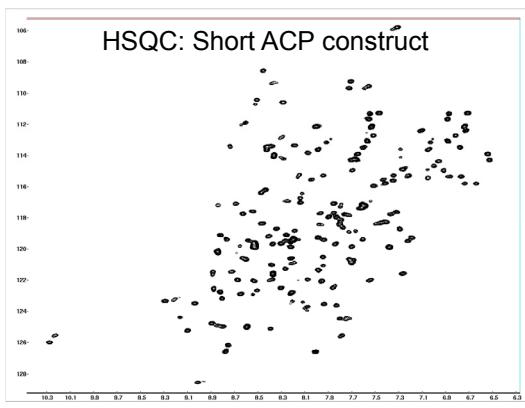
Magnetically align unpaired proton spins (H_0)

Probe with radio frequency (RF)

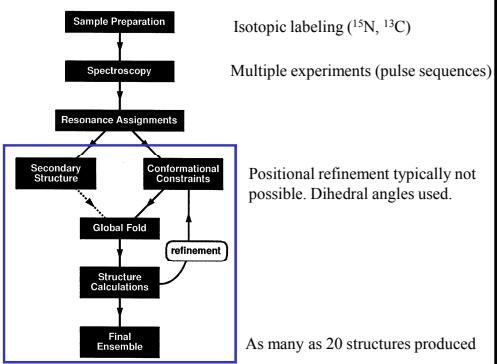
Observe resonance



HSQC: Short ACP construct



NMR Overview



NMR Experimental Observables

- Backbone conformation from chemical shifts (Chemical Shift Index- CSI)
- Distance constraints from NOEs
- Hydrogen bond constraints
- Backbone and side chain dihedral angle constraints from scalar couplings
- Orientation constraints from residual dipolar couplings

NMR Pros/Cons

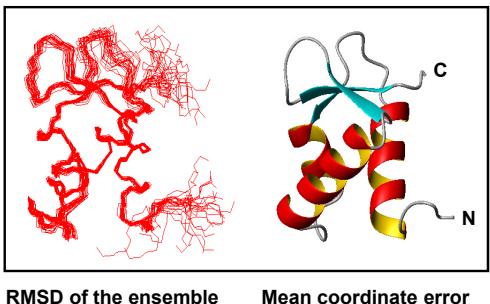
Advantages

- no crystal formation needed
- more physiological conditions

Disadvantages

- results in a set of models that are compatible with data
- size limitation to 200-300 residues (extended recently)
- must label protein with ^{15}N and ^{13}C
- observables typically < refinement parameters

Precision NMR vs. X-ray



RMSD of the ensemble Mean coordinate error

A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

```
HEADER OXIDOREDUCTASE 03-OCT-02 1MXT
TITLE ATOMIC RESOLUTION STRUCTURE OF CHOLESTEROL OXIDASE
TITLE 2(STREPTOMYCES SP SA-COO)
COMPND_MOL_ID_1_
COMPND 2 MOLECULE: CHOLESTEROL OXIDASE
COMPND 3 SCW-1_A;
COMPND 4 SYNONYM: CHOD;
COMPND 5 EC: 1.1.3.6;
COMPND 6 ENGINEERED: YES;
COMPND 7 OTHER_DETAILS: FAD COFACTOR NON-COVALENTLY BOUND TO THE
COMPND 8 ENZYME
```

A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

```
SOURCE MOL_ID_1;
SOURCE 2 ORGANISM_SCIENTIFIC: STREPTOMYCES SP.;
SOURCE 3 ORGANISM_COMMON: BACTERIA;
SOURCE 4 SOURCE_TYPE: CRYSTALLOGRAPHY;
SOURCE 5 EXPRESSION_SYSTEM: ESCHERICHIA COLI;
SOURCE 6 EXPRESSION_SYSTEM_COMMON:BACTERIA;
SOURCE 7 EXPRESSION_SYSTEM_STRAIN: BL21(DE3)LYSS;
SOURCE 8 EXPRESSION_SYSTEM_VECTOR_TYPE: PLASMID;
SOURCE 9 EXPRESSION_SYSTEM_PLASMID: PC0202
```

A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

```
AUTHOR A.VRIELINK,P.LARIO  
REVDAT 25-APR-2003  
JRNL AUTHOR P.LARIO,N.SAMPSON,A.VRIELINK  
JRNL TITL SUB-ATOMIC RESOLUTION CRYSTAL STRUCTURE OF  
JRNL TITL 2 CHOLESTEROL OXIDASE: WHAT ATOMIC RESOLUTION  
JRNL TITL 3 CRYSTALLOGRAPHY REVEALS ABOUT ENZYME MECHANISM AND  
JRNL TITL 4 THE ROLE OF FAD COFACTOR IN REDOX ACTIVITY  
JRNL REF J.MOL.BIOL. V.326 1635 2003  
JRNL REPN ASTM JMOBAK UK ISSN 0022-2836
```

A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

```
REMARK 3 DATA USED IN REFINEMENT:  
REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS) : 0.95  
REMARK 3 RESOLUTION RANGE LOW (ANGSTROMS) : 28.00  
REMARK 3 DATA CUTOFF (SIGMA(F)) : 0.000  
REMARK 3 COMPLETENESS FOR RANGE (%) : 94.1  
REMARK 3 CROSS-VALIDATION METHOD : FREE R  
REMARK 3 FREE R VALUE TEST SET SELECTION : RANDOM  
REMARK 3  
REMARK 3 FIT TO DATA USED IN REFINEMENT (NO CUTOFF)  
REMARK 3 R VALUE (WORKING + TEST SET, NO CUTOFF) : 0.110  
REMARK 3 R VALUE (WORKING SET, NO CUTOFF) : 0.110  
REMARK 3 FREE R VALUE (NO CUTOFF) : 0.132  
REMARK 3 FREE R VALUE TEST SET SIZE (% NO CUTOFF) : 5.000  
REMARK 3 FREE R VALUE TEST SET COUNT (NO CUTOFF) : 13180  
REMARK 3 TOTAL NUMBER OF REFLECTIONS (NO CUTOFF) : 263551
```

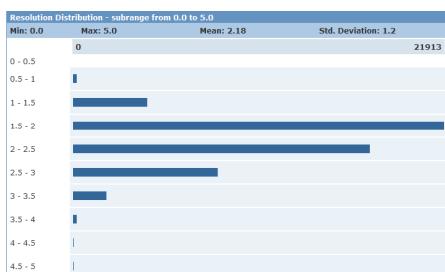
Resolution:

Low > 3 Å
Mid 2-3 Å
High 1.5-2 Å
Very High < 1.5 Å

R factor (residual):

Low resolution ~ 27%
Mid resolution ~ 22 %
High resolution ~ 29 %
Very High res ~ 15%

Resolution



A PDB File

Header contains information about protein and structure
 date of the entry, references, crystallographic data,
 contents and positions of secondary structure elements

```

HELIx 14 14 ALA A 289 THR A 304 1          16
HELIx 15 15 THR A 402 GLN A 405 5          4
HELIx 16 16 ASN A 426 VAL A 425 1          20
HELIx 17 17 ASP A 474 ILE A 475 5          5
HELIx 18 18 PRO A 486 VAL R 506 1          21
SHEET 1 A 6 HIS A 248 GLN A 255 0
SHEET 2 A 6 TYR A 261 LYS A 268 1 O GLU A 266 N GLN A 249
SHEET 3 A 6 LEU A 274 LEU A 287 1 O LEU A 275 N GLN A 267
SHEET 4 A 6 TYR A 10 ILE A 16 1 N VAL A 14 O PHE A 286
SHEET 5 A 6 THR A 36 GLU A 40 1 O LEU A 37 N VAL A 15
SHEET 6 A 6 VAL A 242 THR A 246 1 O THR A 243 N MET A 38

```

A PDB File

Body of PDB file contains information about the atoms in the structure

```

ATOM 76 N PRO A 12 31.129 -4.659 43.245 1.00 9.00 N
ATOM 77 CA PRO A 12 32.426 -4.662 42.542 1.00 9.00 C
ATOM 78 C PRO A 12 32.423 -4.009 41.182 1.00 8.02 C
ATOM 79 O PRO A 12 33.267 -3.177 40.892 1.00 8.31 O
ATOM 80 CB PRO A 12 32.791 -6.126 42.592 1.00 10.02 C
ATOM 81 CG PRO A 12 32.190 -6.663 43.857 1.00 10.12 C
ATOM 82 CD PRO A 12 30.850 -6.126 42.595 1.00 8.77 C
ATOM 90 N ALA A 13 31.485 -4.468 40.316 1.00 8.06 N
ATOM 91 CA ALA A 13 31.357 -3.854 39.004 1.00 7.28 C
ATOM 92 C ALA A 13 29.947 -3.309 38.814 1.00 7.21 C
ATOM 93 O ALA A 13 28.969 -3.932 39.200 1.00 7.56 O
ATOM 94 CB ALA A 13 31.636 -4.879 37.897 1.00 8.54 C

```

Atom number Residue name Residue number
 Atom name

A PDB File

Body of PDB file contains information about the atoms in the structure

```

ATOM 76 N PRO A 12 31.129 -4.659 43.245 1.00 9.00 N
ATOM 77 CA PRO A 12 32.426 -4.662 42.542 1.00 9.00 C
ATOM 78 C PRO A 12 32.423 -4.009 41.182 1.00 8.02 C
ATOM 79 O PRO A 12 33.267 -3.177 40.892 1.00 8.31 O
ATOM 80 CB PRO A 12 32.791 -6.126 42.592 1.00 10.02 C
ATOM 81 CG PRO A 12 32.190 -6.663 43.857 1.00 10.12 C
ATOM 82 CD PRO A 12 30.850 -6.126 42.595 1.00 8.77 C
ATOM 90 N ALA A 13 31.485 -4.468 40.316 1.00 8.06 N
ATOM 91 CA ALA A 13 31.357 -3.854 39.004 1.00 7.28 C
ATOM 92 C ALA A 13 29.947 -3.309 38.814 1.00 7.21 C
ATOM 93 O ALA A 13 28.969 -3.932 39.200 1.00 7.56 O
ATOM 94 CB ALA A 13 31.636 -4.879 37.897 1.00 8.54 C

```

Coordinates in Å

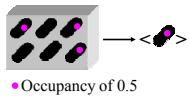
(X, Y, Z)

Mean coordinate error:
 Low > 3 Å .4 Å
 Mid 2-3 Å .3 Å
 High 1.5-2 Å .2 Å
 Very High < 1.5 Å .1 Å

A PDB File

Body of PDB file contains information about the atoms in the structure

ATOM	76	N	PRO	A	12	31.129	-4.659	43.245	1.00	9.00	N
ATOM	77	CA	PRO	A	12	32.426	-4.662	42.542	1.00	9.00	C
ATOM	78	C	PRO	A	12	32.423	-4.009	41.182	1.00	8.02	C
ATOM	79	O	PRO	A	12	33.129	-4.659	40.892	1.00	8.11	O
ATOM	80	CB	PRO	A	12	32.791	-6.126	42.592	1.00	10.02	C
ATOM	81	CG	PRO	A	12	32.190	-6.663	43.857	1.00	10.12	C
ATOM	82	CD	PRO	A	12	30.850	-5.927	43.925	1.00	9.87	C
ATOM	90	N	ALA	A	13	31.485	-4.468	40.316	1.00	8.06	N
ATOM	91	CA	ALA	A	13	31.357	-3.854	39.304	1.00	7.28	C
ATOM	92	C	ALA	A	13	29.947	-3.309	38.814	1.00	7.21	C
ATOM	93	O	ALA	A	13	28.969	-3.332	39.200	1.00	7.56	O
ATOM	94	CB	ALA	A	13	31.636	-4.879	37.897	1.00	8.54	C



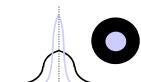
• Occupancy of 0.5

Fractional occupancy

A PDB File

Body of PDB file contains information about the atoms in the structure

ATOM	76	N	PRO	A	12	31.129	-4.659	43.245	1.00	9.00	N
ATOM	77	CA	PRO	A	12	32.426	-4.662	42.542	1.00	9.00	C
ATOM	78	C	PRO	A	12	32.423	-4.009	41.182	1.00	8.02	C
ATOM	79	O	PRO	A	12	33.267	-3.177	40.892	1.00	8.31	O
ATOM	80	CB	PRO	A	12	32.791	-6.126	42.592	1.00	10.02	C
ATOM	81	CG	PRO	A	12	32.190	-6.663	43.857	1.00	10.12	C
ATOM	92	C	ALA	A	12	30.947	-3.309	42.395	1.00	7.77	C
ATOM	90	N	ALA	A	13	31.485	-4.468	40.316	1.00	8.06	N
ATOM	91	CA	ALA	A	13	31.357	-3.854	39.304	1.00	7.28	C
ATOM	92	C	ALA	A	13	29.947	-3.309	38.814	1.00	7.21	C
ATOM	93	O	ALA	A	13	28.969	-3.332	39.200	1.00	7.56	O
ATOM	94	CB	ALA	A	13	31.636	-4.879	37.897	1.00	8.54	C



B-factor Å²