

# Stop and Think about Basal Ganglia Functional Organization: The Pallido-Striatal “Stop” Route

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The “arkypallidal” neurons of the globus pallidus (external segment) emit feedback GABAergic projections to the striatum. In this issue of *Neuron*, Mallet et al. (2016) show that “arkypallidal” neurons provide a Stop signal, suppressing the development of Go-related striatal activity.

Most of neurology and neuroscience textbooks describe basal ganglia (BG) network as two segregated internal BG pathways (Figure 1A). Both BG pathways start in the striatum and converge on the BG output structures (i.e., the internal segment of the globus pallidus and the substantia nigra pars reticulata, GPi and SNr, respectively). The “direct pathway” (striatum-GPi/SNr) is mono-synaptic GABAergic inhibitory, whereas the “indirect pathway” (striatum-GPe-STN-GPi/SNr) is poly-synaptic dis-inhibitory through the GABAergic projections of the striatum and the external segment of the globus pallidus (GPe) and the glutamatergic (excitatory) subthalamic nucleus (STN). Thus, GPe is considered a relay station in the “indirect pathway” between the striatum and the STN (Albin et al., 1989; Gerfen et al., 1990; Figure 1A). The notion of two segregated BG pathways has been strongly supported by the finding of two non-overlapping populations of striatal medium spiny (projection) neurons (MSNs). MSNs with D1 dopamine receptors give rise to the “direct pathway,” whereas MSNs expressing D2 dopamine receptors project to GPe and constitute the origin of the “indirect pathway” (Figure 1A).

This “box and arrow” model dividing the BG into two segregated functional routes has been useful for understanding the computational physiology and pathophysiology of the BG network, as well as for the development of pharmacological and surgical treatments of common BG disorders (Bergman et al., 1990). However, it does not reflect the extreme

complexity of the BG circuitry, notably the intrinsic neuronal heterogeneity of GPe and its full connectivity within the BG network.

In vivo electrophysiological recordings in non-human primates (DeLong, 1971; Elias et al., 2007) and rodents (Benhamou et al., 2012) revealed that GPe neurons can be divided into two distinct populations based on their discharge rate and pattern. Most GPe neurons (~60%–85%) exhibit a high-frequency discharge (50–70 spikes/s) interrupted by long-duration (0.3–2 s) pauses (HFD-P neurons). The remainder exhibits a low-frequency discharge (10–30 spikes/s) with occasional brief high-frequency bursts (LFD-B neurons).

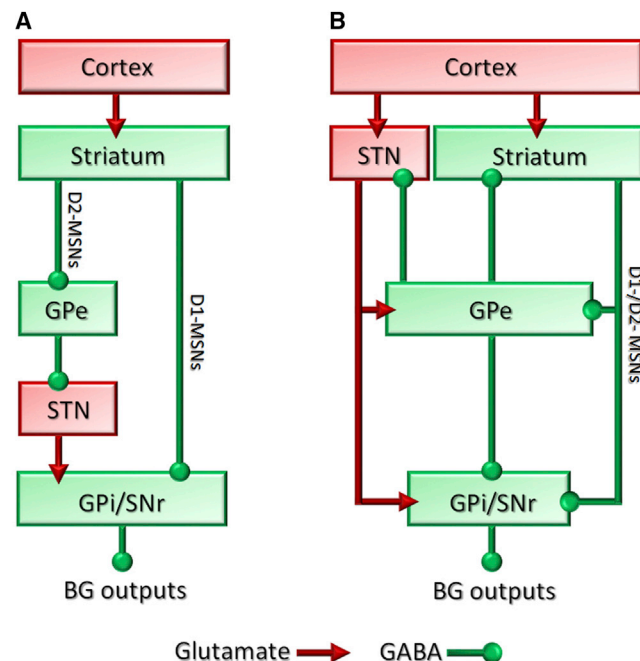
In line with this electrophysiological dichotomous organization of the GPe, recent studies from Peter Magill’s group used juxtacellular recording-labeling technique in dopamine-depleted (Mallet et al., 2012) and dopamine-intact (Abdi et al., 2015) rats, to identify two categories of GABAergic GPe neurons. These two categories of GPe neurons exhibit distinct neurochemical and structural features and have been labeled as “prototypic” and “arkypallidal” neurons. The “prototypic” neurons comprise approximately two-thirds of all GPe neurons and coexpress parvalbumin (PV) and transcription factors Nkx2-1 and Lhx6, while the “arkypallidal” neurons account for only one-fourth of GPe neurons and coexpress preproenkephalin (PPE) and transcription factors FoxP2 and Meis2. Remarkably, the “prototypic” neurons only projects to the STN and the

“arkypallidal” neurons exclusively innervate the striatum. These GPe feedback projections to the striatum parallelized the well-known feedback GPe to STN projections (Figure 1B, although the cellular origin of GPe neuronal populations projecting to the STN is not known yet). Therefore, GPe can no longer be considered as a mere relay station in the “indirect pathway” (Figure 1B). Instead, the emerging view is that GPe plays a central/pivotal role in the orchestration of neuronal activity along the BG network. This GPe central role is revealed by providing a massive GABAergic input to the striatum, targeting both striatal projection neurons and interneurons, as well as to the STN and to the BG output structures—GPi/SNr.

Fortunately, “prototypic” and “arkypallidal” GPe neurons have distinct electrophysiological signatures that allow us to safely discriminate them based on their firing rate and pattern properties across different brain states (Abdi et al., 2015; Mallet et al., 2012). In anesthetized rats, the “prototypic” neurons have a high and regular firing rate regardless of brain state (i.e., EEG slow-wave activity, which is similar to activity observed during natural sleep, or EEG spontaneous activation, which is close to activity observed during awake/behaving state). In contrast, the “arkypallidal” neurons have lower and irregular firing rate that is reduced during slow-wave activity. Finally, in dopamine-depleted animals, the “prototypic” neurons exhibit anti-phasic firing compared to cortical slow-wave (~1 Hz) and beta (~13–30 Hz) oscillations, but the

“arkypallidal” neurons fire in-phase (Mallet et al., 2008, 2012). These electrophysiological signatures enable the studies of the different physiological roles and functions of the two distinct populations of GPe neurons.

In this issue of *Neuron*, Mallet et al. (2016) investigate the role of the cortex-BG network in suppressing ongoing actions. To do so, they recorded the neuronal activity of the “prototypic” and “arkypallidal” GPe neurons in rats engaged in a Stop Signal Task. First, they showed that “prototypic” and “arkypallidal” neurons can be identified through their distinctive firing properties across natural sleep/wake cycle. This is a step beyond the previous classification of GPe neurons based on experimentally induced anesthesia state. Once this objective classification of the GPe neurons was established, Mallet et al. (2016) discovered that the “arkypallidal” neurons selectively respond to Stop cue signaling the sudden cancellation of the action-in-preparation. In addition, these “arkypallidal” Stop responses temporally fit with the timing of the suppression of developing striatal Go-related activity. This is the first demonstration of a distinct physiological action of the “arkypallidal” neurons compared to the “prototypic” neurons in the GPe. Moreover, the discovery of this pallido-striatal motor-suppressing route within the BG network compels us to rethink the role of the GPe in cancellation or suppression of inappropriate behaviors. Schmidt et al. (2013) have previously suggested that cancelling action involves a race in information processing between BG pathways. Indeed, these authors showed that the fast subthalamic activation (probably via hyper-direct cortical projections to STN, Figure 1B) in response to Stop/No-Go signal occurs before the transmission of the striatal Go related-activity to GPi/SNr (via the direct pathway), thus stopping action release.



**Figure 1. The Functional Connectivity of the Basal Ganglia Network: Yesterday and Today**

(A) The classic D1/D2 direct/indirect model of the BG. The GPe is considered as a relay station in the “indirect pathway” between the striatum and the STN. (B) The revised functional connectivity of the BG. Like the striatum, the STN is an input stage of the BG that innervates GPe and the BG output structures (GPi and SNr). In return, GPe projects back to the striatum and the STN and innervates the GPi and the SNr. Glutamatergic and GABAergic connections are shown in red and green, respectively. Abbreviations: BG, basal ganglia; D1-/D2-MSNs, striatal medium spiny (projection) neurons expressing D1/D2 dopamine receptors; GPe and GPi, external and internal segment of the globus pallidus; STN, subthalamic nucleus; SNr, substantia nigra pars reticulata.

However, this fast Stop/No-Go subthalamic response seems too transient to completely cancel the action-in-preparation (Mallet et al., 2016). Therefore, based on their results, Mallet et al. (2016) now proposed a two-step model for successful complete action suppression: actions-in-preparation are first paused via a subthalamic fast activation (directly transmitted to the BG output nuclei—GPi/SNr) and then cancelled via GPe “arkypallidal” Stop GABAergic responses that suppress the Go process within the striatum.

As mentioned by Mallet et al. (2016) in their discussion, the afferent projections of the “arkypallidal” neurons are still unknown. Consequently, to date, it is impossible to know the relative contribution of network versus intrinsic properties of “arkypallidal” neurons to the Stop signals. Given the importance of their conclusions for the neuro-scientific community,

it seems legitimate to hope that researchers will use bio-molecular or optogenetic tools to provide causal results (e.g., effects of an inactivation of the “arkypallidal” neurons on behavior during Stop Signal Task) that strengthen those reported up to now.

Interestingly, a recent study suggests that “prototypic” neurons selectively express Lhx6 or PV, thus dividing the homogeneous population of “prototypic” neurons described by Mallet and colleagues into two distinct populations (Gittis et al., 2014; Mastro et al., 2014). In this study, the authors demonstrated that Lhx6- and PV-GPe neurons differentially innervate a large number of brain areas within and outside the BG. In particular, they showed striatal projections of both Lhx6- and PV-GPe neurons that are selective for striatal GABAergic interneurons (i.e., fast spiking interneurons [FSIs]), suggesting that they are distinct from “arkypallidal neurons.” Therefore, neuronal heterogeneity of the GPe might be

even more complex than the bipartite “prototypic” versus “arkypallidal” organization. Further studies should clarify the role of the striatal projections of both Lhx6- and PV-GPe “prototypic” neurons that, in contrast to “arkypallidal” neurons, selectively target the FSIs.

In Parkinson’s disease (PD), it is established that GPe neurons fire at an abnormal low rate and exhibit pathological discharge pattern, such as an oscillatory activity (Wichmann and DeLong, 2003). Given the GABAergic nature of the GPe neurons, this hypo-activity of the GPe is congruent with the abnormal elevation of the firing rate of the subthalamic neurons observed in PD patients and animal models of PD. From now on, computational models of PD should also consider the evidence of feedback GABAergic projections from GPe to striatum, as well as the reciprocal connectivity between STN and GPe (Figure 1B).

Based on their firing rate and pattern features, it is tempting to speculate that the “prototypic” and “arkypallidal” neurons respectively correspond to the HFD-P and LFD-B neurons previously described in the GPe of both behaving rodents (Benhamou et al., 2012) and non-human primates (DeLong, 1971). If “dreams come true,” the activity of the “prototypic/HFD-P” and “arkypallidal/LFD-B” neurons might also be examined in non-human primates under several behavioral conditions in health and disease. Therefore, the discovery of a pallido-striatal motor-suppressing route by Mallet et al. (2016) provides a major new insight into the understanding of the physiology and pathophysiology of the BG network.

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## Small Immunoglobulin Domain Proteins at Synapses and the Maintenance of Neuronal Features

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The integrity of neural circuits must be maintained throughout the lifetime of an organism. In this issue of *Neuron*, Cherra and Jin (2016) characterize a small, two-Ig domain protein, ZIG-10, and its role in maintaining synaptic density in a specific set of *C. elegans* neurons.

It represents a formidable challenge to preserve the architectural integrity of the nervous system as an animal grows extensively after birth and as newly born neurons are added to pre-existing neuronal circuits (Bénard and Hobert, 2009). Nervous systems also face mechanical strain due to movement and stresses from the environment that can disrupt its organization. Apart from morphology and placement, neurons must also maintain synaptic connections with their targets over time. This can be particularly challenging as subsets of synapses in a circuit are remodeled to accommodate the integration of newly

born neurons (White et al., 1978). Over the past decade, several studies have identified “maintenance factors” that are important for maintaining nervous system integrity. The immunoglobulin domain superfamily (IgSF) of proteins has been shown to play a role in a variety of aspects of neural circuitry control, including maintenance (Rougon and Hobert, 2003). Notably, several small IgSF proteins are specifically dedicated to the maintenance of neural circuits and not their establishment (Bénard and Hobert, 2009).

Members of the *zig* (zwei [two] Ig domain) family of proteins in the nematode *C. elegans* were the first dedicated

maintenance factors identified (Aurelio et al., 2002; Bénard et al., 2009) (Figure 1). The worm genome encodes ten *zig* genes, all of which contain two Ig domains and no other recognizable protein domains; eight of the *zigs* are predicted to be secreted, whereas the other two contain transmembrane domains (Hobert, 2013). Several of the *zig* genes are expressed in PVT, a guidepost interneuron that extends its axon along the length of the animal in its ventral nerve cord (Aurelio et al., 2002). Loss of *zig-4*, which is predicted to be secreted from PVT, causes a phenotype in which axonal tracts in the ventral nerve cord are not properly