



A Computational Pipeline for Investigating Protein Pathway Diversification Post-Genome Duplication

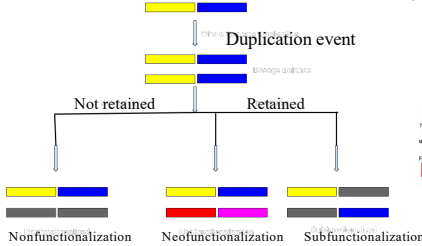
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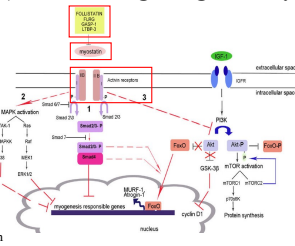


Background

(A) Whole genome duplication



(B) TGF-Beta Signaling Pathway



(C) Species Tree

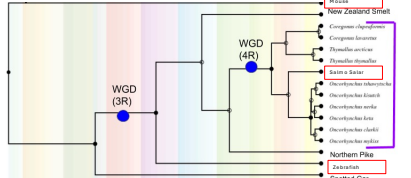


Figure 1: (A) Whole Genome Duplication (WGD) event followed by loss function of one of the copies (neofunctionalization), OR followed by the retention of these genes by changing the function of one of the copies (neofunctionalization) or the two copies sharing parts of the original function (subfunctionalization).

(B) Myostatin is a protein that functions as a skeletal muscle growth inhibitor and is a part of the TGF- β Signaling Pathway. When Myostatin is knocked out, it leads to excess growth of skeletal muscles.

(C) The species tree of interest, with two recent WGD's marked on the branches and salmonids marked by the purple bracket

Hypothesis

Did the mechanisms involved in maintaining duplicated genes after consecutive whole genome duplication events in the Salmonid lineage lead to pathway diversification and increased specificity of the interactions between molecules in the myostatin pathway?

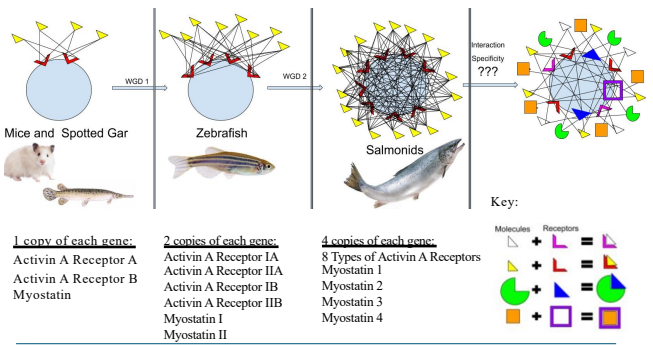
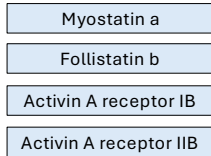


Figure 2: A representation of the effect consecutive WGDs had on myostatin and its interacting partners and the species that represent each pathway.

Phylogenetic Analysis Pipeline and Results

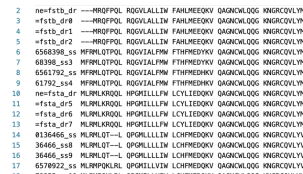
1. Find Homologs (BLASTP)

Query myostatin genes (and interacting partners) in mouse, zebrafish, and salmon genomes



2. Multiple Sequence Alignment (MAFFT)

Align matches for each gene



3. Model Selection (PhyML)

Estimate maximum likelihood phylogenies

Best model: JTT + G+I

Indelimitation model

Evolutionary Processes

Properties of nucleotide sites

Number of indelimitation site categories

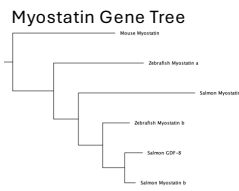
Number shape parameters

- JTT
- G+I
- Indelimitation (n: 20)
- K
- Indelimitation (n: 10)

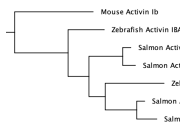
| Model | Discrimination | lnL | AIC | BIC | |
|-------|----------------|-----|-------------|-------------|-------------|
| JTT | +0.01 | 79 | -4860.51380 | 14937.02760 | 14411.55917 |
| JTT | +0.0 | 79 | -4867.53882 | 14910.87724 | 14403.90132 |
| JTT | +0.01eF | 98 | -4893.42708 | 14110.88412 | 14000.78492 |
| JTT | +0.01eF | 97 | -4890.26040 | 14124.52340 | 14010.48051 |
| WAG | +0.01eF | 98 | -7008.40843 | 14205.31286 | 14096.44966 |
| Flu | +0.01eF | 98 | -7005.92686 | 14207.85292 | 14097.78472 |
| WAG | +0.01 | 79 | -7003.67963 | 14261.35726 | 14076.68923 |
| US | +0.01 | 79 | -7003.36408 | 14264.68808 | 14079.15952 |
| WAG | +0.01eF | 98 | -7018.32073 | 14268.68142 | 14058.19222 |

4. Gene Tree Reconstruction

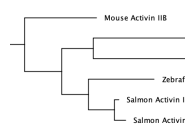
Construct and reconcile gene trees



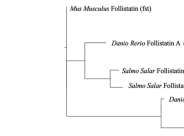
Activin IB Gene Tree



Activin IIB Gene Tree

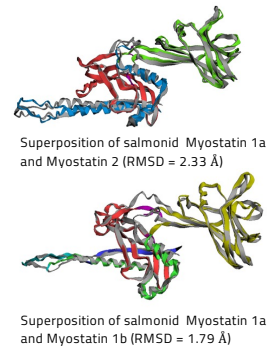


Follistatin Gene Tree



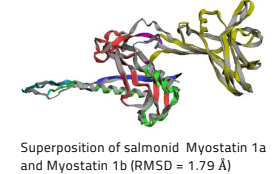
5. Structural Alignment (FATCAT)

Visualize superposition of structures
Determine degree of structural divergence



6. Species Specific Docking (Replica Dock)

Investigate interactions between interacting partners



Conclusion & Next Steps

- Using the high throughput method of building models with FATCAT, the preliminary results show that there is significant structural similarity between Atlantic salmon myostatin 2 with the myostatin 1a. Additionally, myostatin 1a and 1b do not show structural divergence.
- Future work involves more detailed structural modeling as well as modeling of all interacting partners, coupled to docking to look at potential interaction changes and a gain of specificity.
- Further future work involves detailed (dN/dS-based) selection analysis on myostatin and all interacting partners, which when coupled with the protein structural analysis, will illuminate the patterns of pathway and protein regulatory diversification.

References

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