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Natural variation in stomata size contributes to the local adaptation of water-use efficiency in *Arabidopsis thaliana*

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Abstract

Stomata control gas exchanges between the plant and the atmosphere. How natural variation in stomata size and density contributes to resolve trade-offs between carbon uptake and water loss in response to local climatic variation is not yet understood. We developed an automated confocal microscopy approach to characterize natural genetic variation in stomatal patterning in 330 fully sequenced *Arabidopsis thaliana* accessions collected throughout the European range of the species. We compared this to variation in water-use efficiency, measured as carbon isotope discrimination ($\delta^{13}\text{C}$). We detect substantial genetic variation for stomata size and density segregating within *Arabidopsis thaliana*. A positive correlation between stomata size and $\delta^{13}\text{C}$ further suggests that this variation has consequences on water-use efficiency. Genome wide association analyses indicate a complex genetic architecture underlying not only variation in stomatal patterning but also to its covariation with carbon uptake parameters. Yet, we report two novel QTL affecting $\delta^{13}\text{C}$ independently of stomatal patterning. This suggests that, in *A. thaliana*, both morphological and physiological variants contribute to genetic variance in water-use efficiency. Patterns of regional differentiation and covariation with climatic parameters indicate that natural selection has contributed to shape some of this variation, especially in Southern Sweden, where water availability is more limited in spring relative to summer. These conditions are expected to favour the evolution of drought avoidance mechanisms over drought escape strategies.

KEYWORDS

Arabidopsis thaliana, genomewide association study, local adaptation to climate, Q_{ST} - F_{ST} analysis, stomata, water-use efficiency

1 | INTRODUCTION

In plants, carbon uptake and water loss are intimately linked by a trade-off between growth and water conservation (Cowan, 1986; Cowan & Farquhar, 1977; Field, Merino, & Mooney, 1983). Stomata, the microscopic pores embedded in the epidermis of plant leaves, play a key role in the resolution of this trade-off. Their density,

distribution and regulation control the rate of CO_2 and water exchange (Raven, 2002). As a result, they impact the ratio of photosynthetic carbon assimilation to water loss via transpiration. This ratio defines water-use efficiency (WUE), a physiological parameter that directly determines plant productivity when the water supply is limited. Variation in density, distribution and regulation of stomata may thus have played a pivotal role in shaping the diversity of plant

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communities throughout the globe (Lambers, Chapin, & Pons, 1998; McDowell et al., 2008).

The density of stomata on the leaf surface is expected to correlate positively with the rate of gas exchanges between the leaf and the atmosphere, also called “conductance.” Models based on gas diffusion theory predict that small stomata in high density can best maximize conductance (Franks & Beerling, 2009). A positive relationship between stomata density and conductance has been reported in a majority of studies looking at natural variation between species (Anderson & Briske, 1990; Pearce, Millard, Bray, & Rood, 2006) as well as within species (Carlson, Adams, & Holsinger, 2016; Muchow & Sinclair, 1989; Reich, 1984). Yet, higher stomata density does not always translate into higher rates of gas exchanges: In a diversity panel of rice (Ohsumi, Kanemura, Homma, Horie, & Shiraiwa, 2007) or within several vegetable crop species (Bakker, 1991), for example, the relationship was not observed.

Molecular mutants in genes promoting stomata development show that reduced stomata density translates into decreased water loss and increased ability to survive after exposure to drought (Franks, Doheny-Adams, Britton-Harper, & Gray, 2015; Yoo et al., 2010).

Yet, decreased stomata density does not necessarily associate with increased demands on WUE imposed by water limitation. In the *Mimulus guttatus* species complex, accessions from drier inland populations showed decreased stomata density and increased WUE, compared to accessions collected in humid coastal populations (Wu, Lowry, Nutter, & Willis, 2010). By contrast, in 19 *Protea repens* populations measured in a common garden experiment, stomata density increased with decreasing summer rainfall at the source location (Carlson et al., 2016).

In fact, stomata density is not the only parameter modulating the balance between water loss and carbon uptake. Variation in stomata size also impacts the efficiency of stomata regulation (Raven, 2014). Stomata open and close in response to environmental and internal signals (Chater et al., 2011; Kinoshita et al., 2011). This ensures that plants do not desiccate when water evaporation is maximal and spares water when photosynthesis is not active (Daszkowska-Golec & Szarejko, 2013). The speed of stomata closure is higher in smaller stomata (Drake, Froend, & Franks, 2013; Raven, 2014). Stomatal responses are an order of magnitude slower than photosynthetic changes, so any increase in closure time lag may result in unnecessary water loss and reduce WUE (T. Lawson, Kramer, & Raines, 2012; Raven, 2014). However, it is often observed that decreases in stomata size occur at the expense of increased stomata density (reviewed in Hetherington & Woodward, 2003). This leads to a correlation that may at first be counterintuitive: An increase in stomata density can result in improved WUE because of indirect effects on stomata size. In *Eucalyptus globulus*, however, plants from the drier sites had smaller stomata and higher WUE but no concomitant change in stomata density (Franks, Drake, & Beerling, 2009). This suggested that the developmental effect correlating stomata size and density may sometimes be alleviated. Altogether, these studies highlight interconnections between stomata size, stomata density and WUE that change across species or populations. How and whether variation in these traits and their

connections support or constrain adaptive processes, however, is not clearly established.

Eco-evolutionary studies, for example, the analysis of evolutionary forces shaping genetic variation in natural populations, can determine whether phenotypic variance has a significant impact on the ecology of species (Carroll, Hendry, Reznick, & Fox, 2007; Hendry, 2016). By drawing on the elaborate toolbox of population genetics and genomics, it is not only possible to determine the genetic architecture of any given trait but also to ask whether it is optimized by natural selection and to investigate the ecological determinants of selective forces at work (Hendry, 2016; Weinig, Ewers, & Welch, 2014). In this effort, the annual species *Arabidopsis thaliana*, which thrives as a pioneer species in disturbed habitats, has a privileged position (Gaut, 2012). Genome-wide patterns of nucleotide variation can be contrasted to phenotypic variation, and both the genetic architecture and the adaptive history of the traits can be reconstructed (Alonso-Blanco et al., 2016; Atwell et al., 2010; Fournier-Level et al., 2011). Environmental variation has a documented impact on local adaptation in this species (Debieu et al., 2013; Hamilton, Okada, Korves, & Schmitt, 2015; Hancock et al., 2011; Kronholm, Picó, Alonso-Blanco, Goudet, & de Meaux, 2012; Lasky et al., 2014; Postma & Ågren, 2016). In addition, natural variation in stomatal patterning is known to segregate among *A. thaliana* accessions (Delgado, Alonso-Blanco, Fenoll, & Mena, 2011). This species thus provides the ideal evolutionary context in which the adaptive contribution of variation in stomatal patterning can be dissected.

Here, we developed an automated confocal microscopy approach that overcomes the technical limitations which have so far complicated the phenotyping of stomatal variation on larger samples. We characterized genetic variation in stomatal patterning in 330 fully sequenced accessions, across a north–south transect of the European range. Additionally, we measured $\delta^{13}\text{C}$, a commonly used estimate of water-use efficiency (WUE; Juenger et al., 2005; Martin & Thorstenson, 1988; McKay et al., 2008; Mojica et al., 2016), for all genotypes. Combined with public genomic and environmental resources, this data set allows us to ask: (a) How variable are natural *A. thaliana* accessions in stomatal patterning? (b) Does variation in stomatal patterning influence the carbon–water trade-off? (c) What is the genetic architecture of traits describing stomatal patterning? and (d) Is stomatal patterning optimized by natural selection?

By combining a genomewide association approach with $Q_{\text{ST}}/F_{\text{ST}}$ analyses and associations with environmental parameters, we show that, in *A. thaliana*, variation in stomatal patterning plays a role in local adaptation. Our results further indicate that natural variation in stomata size is one of the adaptive traits contributing to the optimization of WUE.

2 | METHODS

2.1 | Plant material, plant genotypes and growth conditions

In total, 330 accessions, spanning a wide geographical range, were selected from the 1001 collection of fully sequenced genotypes

(Supporting Information Table S1). Accessions were assigned to five groups based on their geographical origin and genetic clustering (Alonso-Blanco et al., 2016): Spain, Western Europe, Central Europe, Southern Sweden and Northern Sweden (Supporting Information Figure S1). In 20 cases, for which genetic information contradicted geographical information, we prioritized geographical information since we are focusing on local adaptation and expect that geography, as opposed to demographic history, reflects the scale at which local adaptation proceeds. To avoid oversampling, we randomly reduced the number of plants sampled at the same location to one for the analysis of heritability, regional differentiation (Q_{ST} - F_{ST}) and climatic correlation, resulting in 287 accessions.

The genome sequences of the 330 genotypes included in the analysis were downloaded from the 1001 genome database (Alonso-Blanco et al., 2016) on May 12, 2017. Single nucleotide polymorphism (SNP) data were extracted using *VCFtools* (Danecek et al., 2011). Genomic data were thinned to one SNP picked randomly in each 1,000 bp window to reduce computational load. In *A. thaliana*, linkage disequilibrium extends beyond 1 kb (Nordborg et al., 2002). Thus, this data-size reduction should not impact statistics describing the geographical structure of genomic variation. Additionally, minimum minor allele frequency was set to 5% and sites exceeding 5% missing data were removed, resulting in 70,410 SNPs among all genotypes. SNP information was loaded into R using the *vcfR* package (Knaus, Grunwald, Anderson, Winter, & Kamvar, 2017). For genome-wide association studies, the full, unthinned SNP data set was used and missing SNPs were imputed using *BEAGLE* version 3.0 (Browning & Browning, 2009).

Seeds were stratified on wet paper for 6 days at 4°C in darkness. Plants were grown on soil in 5 × 5 cm paper pots in three replicates with one plant per pot. Genotypes were randomized within each of three blocks of 12 trays containing 8 × 4 pots. Plants were grown for 7 weeks in growth chambers (one per block) under the following conditions: 16 hr light; 95 μmol s⁻¹ mm⁻² light intensity; and 20°C day- and 18°C night temperature. Plants were watered twice a week and trays shuffled and rotated every two to three days to account for variable conditions within the chambers.

2.2 | High-throughput phenotyping

After 7 weeks, one fully expanded, intact, adult leaf (one of the largest leaves developed after leaf 4) was selected from each plant for microscopic analysis. Stomata density and size as well as leaf size were measured using our high-throughput microscopy pipeline (for details, see Supporting Information Appendix S1). Stomata density was also determined manually on a random set of 14 individuals and on a set of 32 independently grown individuals. Automatic and manual measurements were strongly correlated (Pearson correlation coefficient $r^2 = 0.88$, $p < 0.01$ and $r^2 = 0.81$, $p < 0.01$, for the 14 and 32 see individuals Supporting Information Figures S2 and S3). The algorithm was conservative and tended to slightly underestimate stomata numbers, resulting in a low false-positive rate. This ensured that stomata area was generally quantified on objects that

corresponded to real stomata. Due to quality filters in our pipeline, the number of analysed images differed between samples (Supporting Information Figure S4). We found a significant correlation between the number of images analysed and stomata density ($r = 0.21$, $p < 0.01$, Supporting Information Figure S5), but not stomata size ($r = 0.02$, $p > 0.05$). Thus, we included the number of images as a cofactor in all statistical models for stomata density (see below). Carbon isotope discrimination measurements ($\delta^{13}C$) of whole rosettes were performed for all plants in block 1 (for details, see Supporting Information Appendix S1).

2.3 | Heritability estimates

Broad-sense heritability H^2 , the proportion of the observed phenotypic variance that is genetic, was estimated as:

$$H^2 = \text{VarG} / (\text{VarG} + \text{VarE}),$$

where VarG is the genetic variance, and VarE is the environmental variance. Because we worked with inbred lines, VarE and VarG could be estimated as the variance between replicates of a genotype and the variance between genotypes, respectively, with a linear mixed model using block as fixed effect and genotype as random effect. We created linear mixed model using the *lme* function from the *NLME* package (Pinheiro, Bates, DebRoy, & Sarkar, 2015; Supporting Information Appendix S2). For $\delta^{13}C$, no replicates were available but a pseudoheritability estimate was extracted from the GWAS mixed model including the kinship matrix (Atwell et al., 2010).

2.4 | Genomewide association study (GWAS)

For GWAS, SNPs with minor allele count <5 were removed, leaving a data set of 2.8–3M SNPs, depending on missing data for the phenotypes. Minor allele frequency spectra for all three data sets show that the subset of 261 genotypes, for which all three phenotypes were determined, has a lower proportion of rare SNPs (Supporting Information Figure S6). GWAS was performed with a mixed model correcting for population structure using a kinship matrix calculated under the assumption of the infinitesimal model. SNPs were first analysed with a fast approximation (Kang et al., 2010), and the 1000 topmost associated SNPs were reanalysed with the complete model that estimates the respective variance components for each SNP separately (Kang et al., 2008).

For trait pairs measured on the same plant, a multitrait mixed model (MTMM) was applied to distinguish common and trait-specific SNP–phenotype association (Korte et al., 2012).

The MTMM performs three different statistical tests on a bivariate phenotype including each trait pair. The first model tests whether a given SNP has the same effect on both traits. This model has increased power to detect significant associations, which may fall under the significance threshold when traits are analysed in isolation. The second model identifies SNPs having distinct effects on the two traits. It is well suited to detect SNPs with antagonistic effects on both traits. The last model combines both trait-specific and common effects and is particularly powerful for detecting markers affecting

both traits with different intensity. The MTMM analysis also provides estimates of the genetic and environmental correlation for each pair of traits. The statistical details of the models are described in Korte et al. (2012).

For all analyses (GWAS and MTMM), the significance threshold for QTL identification was determined as a 5% Bonferroni threshold, that is, 0.05 divided by the number of SNPs in the data set.

2.5 | Climatic data

Climatic data included average precipitation, temperature, water vapour pressure (humidity), wind speed and solar radiation estimates with 2.5-min grid resolution (WorldClim2 database (Fick & Hijmans, 2017) on May 30th, 2017) and soil water content (Trabucco & Zomer, 2010). For each variable and accession, we extracted a mean over the putative growing season, that is, the months in the year with average temperature $>5^{\circ}\text{C}$ and average soil water content over 25% (Supporting Information Table S1). We further computed historical drought frequencies at *A. thaliana* collection sites using 30+ years of the remotely sensed Vegetation Health Index (VHI). The VHI is a drought detection method that combines the satellite measured Vegetative Health and Thermal Condition Indices to identify drought-induced vegetative stress globally at weekly 4 km^2 resolution (Kogan, 1995). This is a validated method for detecting drought conditions in agriculture. Specifically, we used VHI records to calculate the historic frequency of observing drought conditions (VHI <40) during the spring (quarter surrounding spring equinox) and summer (quarter surrounding summer solstice). These are the typical reproductive seasons of *Arabidopsis* populations (reviewed in Burghardt, Metcalf, Wilczek, Schmitt, & Donohue, 2015). The drought regime in each location was quantified as the log-transformed ratio of spring over summer drought frequency. Positive values of this drought regime measure reflect environments where the frequency of drought decreases over the typical reproductive growing season and vice versa for negative values. This ratio quantifies the seasonality of water availability. It correlates with the ratio of soil water content of the first and third month of the reproductive season ($r = 0.54$, $p < 0.01$), which we defined as the first and third growing month in the year, giving similar estimates as Burghardt et al. (2015).

Because the seven climate variables are correlated, we combined them in seven principal components (PCs) for 316 *A. thaliana* collection sites (Supporting Information Figures S7–S9, loadings described in Supporting Information Appendix S2). Fourteen genotypes with missing climate data were excluded. Climatic distance between each region pair was estimated as the F -statistic of a multivariate analysis of variance (MANOVA) with climatic PCs as response variables and region of origin as predictor.

2.6 | Population genomic analysis

Population genomic analysis was conducted using R (R Development Core Team, 2008; R Markdown documentation in Supporting Information Appendix S2). Principal component analysis (PCA) of genomic

data (thinned to 1 kb) was done using the ADEGENET package (Jombart et al., 2016) with missing data converted to the mean (Supporting Information Figures S10 and S11).

Comparing phenotypic differentiation (Q_{ST}) to the distribution of F_{ST} is a useful method to reveal signatures of local adaptation (Leinonen, McCairns, O'hara, & Merilä, 2013; Whitlock & Guillaume, 2009). Genomewide, pairwise F_{ST} estimates between regions were calculated using the HIERFSTAT package (function *basic.stats*, Nei's F_{ST} ; Goudet, 2005). Negative F_{ST} values were set to zero before the 95th percentile was calculated.

For stomata density, stomata size and WUE, the respective phenotypic differentiation between regions, Q_{ST} , was estimated as:

$$Q_{ST} = \text{VarB} / (\text{VarW} + \text{VarB}),$$

where VarW is the genetic variance within regions and VarB the genetic variance between regions as described in Kronholm et al. (2012; for details, see Supporting Information Appendix S1).

To test whether Q_{ST} estimates significantly exceed the 95th percentile of the F_{ST} distribution, we permuted the phenotypic data by randomizing genotype labels to keep heritability constant. For each permutation and phenotype, we calculated the difference between each Q_{ST} value and the 95th percentile of the F_{ST} distribution. We used the 95th percentile of the maximum $Q_{ST}-F_{ST}$ distance distribution as a threshold for determining if phenotypic differentiation significantly exceeds neutral expectations. Since this test takes the maximum $Q_{ST}-F_{ST}$ distance for all population combinations in each permutation, it does not require multiple testing correction.

2.7 | Statistical analysis

Statistical analysis was conducted using R (R Development Core Team, 2008; R Markdown documentation in Supporting Information Appendix S2). Plots were created using the following libraries: *ggplot2* (Wickham, 2009), *ggthemes* (Arnold et al., 2017), *ggmap* (Kahle & Wickham, 2013), *ggbiplot* (Vu, 2011) and *effects* (Fox et al., 2016).

We used generalized linear models (GLM) to test the effect of block, origin, pot position in tray (edge or centre) and leaf size on each phenotype (stomata density, stomata size and $\delta^{13}\text{C}$). For stomata density, we also included the number of analysed images as a cofactor. The error distribution was a quasi-Poisson distribution for stomata density and size and Gaussian for $\delta^{13}\text{C}$. Stomata density was log-transformed to avoid overdispersion. Significance of each predictor was determined via a type-II likelihood ratio test (ANOVA function of the CAR package). Significant differences between regions were based on GLMs including only significant predictor variables and determined with Tukey's contrasts using the *glht* function of the MULTCOMP package (Hothorn et al., 2017). GLMs were also used to test the impact of all climatic PCs on phenotypic traits while accounting for population structure with the first 20 PCs for genetic variation, which explain 28% of genetic variation (see above). Additionally, for $\delta^{13}\text{C}$, we also tested a simpler model including climatic parameters but not population structure. From the resulting models, we created effect plots for significant environmental PCs using the

EFFECTS package (Fox et al., 2016). Further, we used GLMs with binomial distribution to test whether any of the climatic PCs significantly predicts the allelic states of loci associated with WUE in GWAS.

3 | RESULTS

3.1 | Substantial genetic variation in stomata density and size

We analysed over 31,000 images collected in leaves of 330 *A. thaliana* genotypes and observed high levels of genetic variation in stomatal patterning. Genotypic means ranged from 87 to 204 stomata/mm² for stomata density and from 95.0 to 135.1 μm^2 for stomata size (see Supporting Information Table S2 for raw phenotypic data). Leaf size was not significantly correlated with stomata density ($r = -0.02$, $p = 0.7$, Supporting Information Figure S12) and stomata size ($r = -0.08$, $p = 0.15$, Supporting Information Figure S13), as expected in fully developed leaves. Broad-sense heritability reached 0.41 and 0.30 for stomata size and density, respectively. Mean stomata density and stomata size were negatively correlated ($r = -0.51$, $p < 0.001$; Figure 1). Due to the strong correlation between stomata size and density, we focus primarily on stomata size in the following report, but results for stomata density are in the Supporting Information Appendix S2.

3.2 | Stomata size correlates with water-use efficiency

We expected variation in stomatal traits to influence the trade-off between carbon uptake and transpiration. Thus, we measured

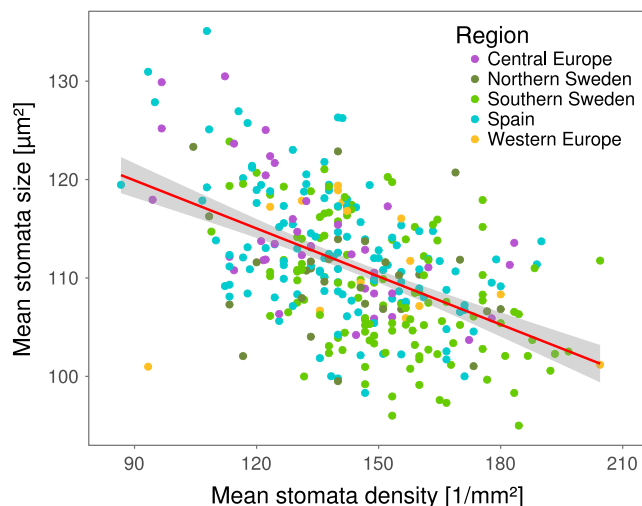


FIGURE 1 Natural variation in stomatal patterning. Stomata density and size were measured for 330 natural genotypes of *Arabidopsis thaliana*. The plot shows genotypic means of stomata density and stomata size. Dots are coloured based on the geographical origin of each accession. The red line shows a linear fit, and grey shadows indicate the error of the fit. Pearson's product-moment correlation $r = -0.51$, $p < 0.001$

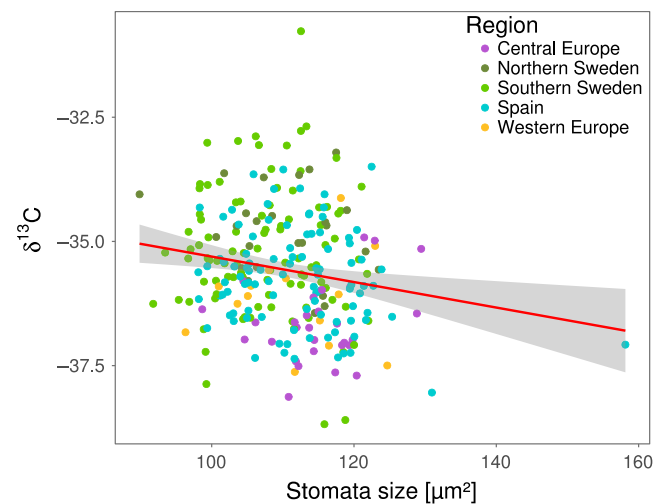


FIGURE 2 Stomata size correlates with water-use efficiency. $\delta^{13}\text{C}$ was measured for all plants in block 1. Plots show correlation of stomata size (block 1 only) with $\delta^{13}\text{C}$. $\delta^{13}\text{C}$ is expressed as ‰ against the Vienna Pee Dee Belemnite (VPDB) standard. The red line shows a linear fit, and grey shadows indicate the error of the fit. Pearson's product-moment correlation: $r = -0.18$, $p = 0.004$. Correlation of $\delta^{13}\text{C}$ and stomata size is not only driven by the Spanish outlier (correlation without outlier: $r = -0.16$, $p = 0.009$). Genetic correlation was calculated using the MTMM approach: $r = -0.58$, $p < 0.05$

isotopic carbon discrimination, $\delta^{13}\text{C}$, an estimator that increases with water-use efficiency (WUE; Farquhar, Hubick, Condon, & Richards, 1989; McKay et al., 2008). $\delta^{13}\text{C}$ ranged from -38.7‰ to -30.8‰ (Supporting Information Table S2) and was significantly correlated with stomata size ($r = -0.18$, $p = 0.004$; Figure 2), indicating that accessions with smaller stomata have higher WUE. About ~4% of the total phenotypic variation (i.e., the sum of phenotypic and genetic variance) in $\delta^{13}\text{C}$ is explained by genetic variance in stomata size. We found no significant correlation between stomata density and $\delta^{13}\text{C}$ ($r = -0.007$, $p = 0.9$, Supporting Information Figure S14).

3.3 | Common genetic basis of stomata size and $\delta^{13}\text{C}$

To identify the genetic basis of the phenotypic variance we observe, we conducted a genome-wide association study (GWAS) for each phenotype. We calculated for each phenotype a pseudoheritability, which is the fraction of phenotypic variance explained by the empirically estimated relatedness matrix (e.g., kinship matrix computed on genome-wide SNP typing). Pseudoheritability estimates were 0.59 for stomata density, 0.56 for stomata size and 0.69 for $\delta^{13}\text{C}$, indicating that differences in stomatal patterning and carbon physiology decreased with increasing relatedness. Despite considerable levels of heritability, we did not detect any variant associating with stomata density at a significance above the Bonferroni-corrected p -value of 0.05 ($\log_{10}(p) = 7.78$). For stomata size, we detected one QTL with two SNPs significantly associating at positions 8567936 and

8568437 (Supporting Information Figure S15). These SNPs have an allele frequency of 1.5% (five counts) and 2.1% (seven counts), respectively, and map to gene AT4G14990.1, which encodes for a protein annotated with a function in cell differentiation. The former SNP is a synonymous coding mutation, while the latter is in an intron.

For $\delta^{13}\text{C}$, one genomic region on chromosome 2 position 15094310 exceeded the Bonferroni significance threshold ($\log_{10}(p) = 7.97$, Supporting Information Figure S16). Allele frequency at this SNP was 9.7% (30 counts), and all accessions carrying this allele, except four, were from Southern Sweden (three Northern Sweden and one Central Europe). Southern Swedish lines carrying the allele showed significantly increased $\delta^{13}\text{C}$ compared to the remaining Southern Swedish lines ($W = 1868$, p -value = $6.569\text{e-}05$, Supporting Information Figure S17). A candidate causal mutation is a nonsynonymous SNP at position 15109013 in gene AT2G35970.1, which codes for a protein belonging to the late embryogenesis-abundant (LEA) hydroxyproline-rich glycoprotein family. This SNP also shows elevated association with the phenotype. However, its significance was below the Bonferroni threshold ($\log(p) = 7$). Since this SNP is not in linkage disequilibrium with the highest associating SNP in the region (Supporting Information Figure S18), it is possible that another, independent SNP in this region is causing the association.

We used multitrait mixed model (MTMM) analysis to disentangle genetic and environmental determinants of the phenotypic correlations. We found that the significant correlation between stomata density and stomata size ($r = -0.5$) had no genetic basis, but had a significant ($r = -0.9$, $p < 0.05$) residual correlation. This suggests that the correlation was not determined by common loci controlling the two traits, but by other, perhaps physical, constraints or by epistatic alleles at distinct loci. By contrast, the correlation between stomata size and $\delta^{13}\text{C}$ ($r = -0.18$) had a significant genetic basis (kinship-based correlation, $r = -0.58$, $p < 0.05$). Thus, in contrast to the phenotypic variation, genetic variation in stomata size roughly explains over 33% of the genetic variation in $\delta^{13}\text{C}$.

To further investigate the genetic basis for the correlation between stomata size and $\delta^{13}\text{C}$, we performed MTMM GWAS, which tests three models: The first model tests whether a SNP has the same effect on both traits; the second model tests whether a SNP has differing effects on both traits; and the third model is a combination of the first two to identify SNPs which have effects of different magnitude on the traits (Korte et al., 2012). We did not observe variants with same or differing effects on $\delta^{13}\text{C}$ and stomata size. However, with the combined model, we observed a marginally significant association on chromosome 4, which had an effect on $\delta^{13}\text{C}$ but not on stomata size. GWAS of $\delta^{13}\text{C}$ restricted to the 261 individuals used for the MTMM analysis confirmed the QTL on chromosome 4. GWAS applied to different but overlapping sets of accessions yield similar results but can sometimes differ in the set of significant associations, since marginal changes in SNP frequency can affect significance levels (Supporting Information Figure S6). Indeed, the p -values of associations with $\delta^{13}\text{C}$ for the two data sets (310 and 261 accessions) are highly correlated ($r = 0.87$, $p < 0.0001$,

Supporting Information Figure S19). In this set of genotypes, two SNPs, at position 7083610 and 7083612, exceeded the Bonferroni-corrected significance threshold ($\alpha = 0.05$; both $p = 4.8\text{e-}09$, Supporting Information Figure S20) although they were under the significance threshold in the larger data set. Allele frequency is 14% (37 counts) at these two loci and explains 11% of the phenotypic variation. The association is probably due to complex haplotype differences since it coincides with a polymorphic deletion and contains several imputed SNPs. Thirty-five of the 37 accessions carrying the minor allele originated from Southern Sweden and showed significantly higher $\delta^{13}\text{C}$ compared to other Southern Swedish accessions (mean difference = 1.34; $W = 1707$, $p = 1.15\text{e-}06$; Supporting Information Figure S21). In summary, we detected two genetic variants significantly associating with $\delta^{13}\text{C}$, independent of stomata size, despite the common genetic basis of the two traits.

3.4 | Stomata size and stomata density correlate with geographical patterns of climatic variation

We used PCA to describe multivariate variation in climatic conditions reported for the locations of origins of the genotypes. We tested the correlation of each measured phenotype with climatic principal components (PCs) using a GLM which accounted for genetic population structure (see Methods). We found a significant, negative relationship between genetic variation in stomata size and climatic PC2 (likelihood ratio test chi-square (LRT X^2) = 9.2784, degrees of freedom (df) = 1, $p = 0.005$) and PC5 (LRT X^2 = 5.7335, df = 1, $p = 0.02$, Figure 3). Climatic PC 2 explained 23.8% of climatic variation and had the strongest loadings (both negative) from temperature and water vapour pressure (humidity). Climatic PC 5 explained 9% of the climatic variation and mostly increased with increasing spring–summer drought probability ratio and increasing solar radiation. We also found significant climatic predictors for the distribution of genetic variation in stomata density (PC 2: LRT X^2 = 8.6612, df = 1, $p = 0.003$; PC 5: LRT X^2 = 7.3773, df = 1, $p = 0.007$; PC 7: LRT X^2 = 6.6033, df = 1, $p = 0.01$; Supporting Information Figure S22). $\delta^{13}\text{C}$ did not correlate with any of the climatic PCs. However, removing population structure covariates from the model revealed significant correlations of $\delta^{13}\text{C}$ with climatic PC2 (+, LRT X^2 = 7.3564, df = 1, $p = 0.006$), PC3 (–, LRT X^2 = 3.8889, df = 1, $p = 0.048$) and PC4 (+, LRT X^2 = 6.6885, df = 1, $p = 0.009$; Supporting Information Figure S23). PC3 explained 13.7% of climatic variation and principally increased with rainfall and decreased with spring–summer drought probability ratio. PC4 explained 11.4% of the total variation and mostly increased with wind speed. Therefore, the covariation of $\delta^{13}\text{C}$ with climatic parameters describing variation in water availability and evaporation in *A. thaliana* is strong but confounded with the demographic history of the species. To test whether alleles associating with increased $\delta^{13}\text{C}$ in GWAS are involved in adaptation to local climate, we checked whether any climatic PC is a significant predictor of the allelic state of Southern Swedish accessions. However, none of the climatic PCs was a significant predictor for one of the two loci.

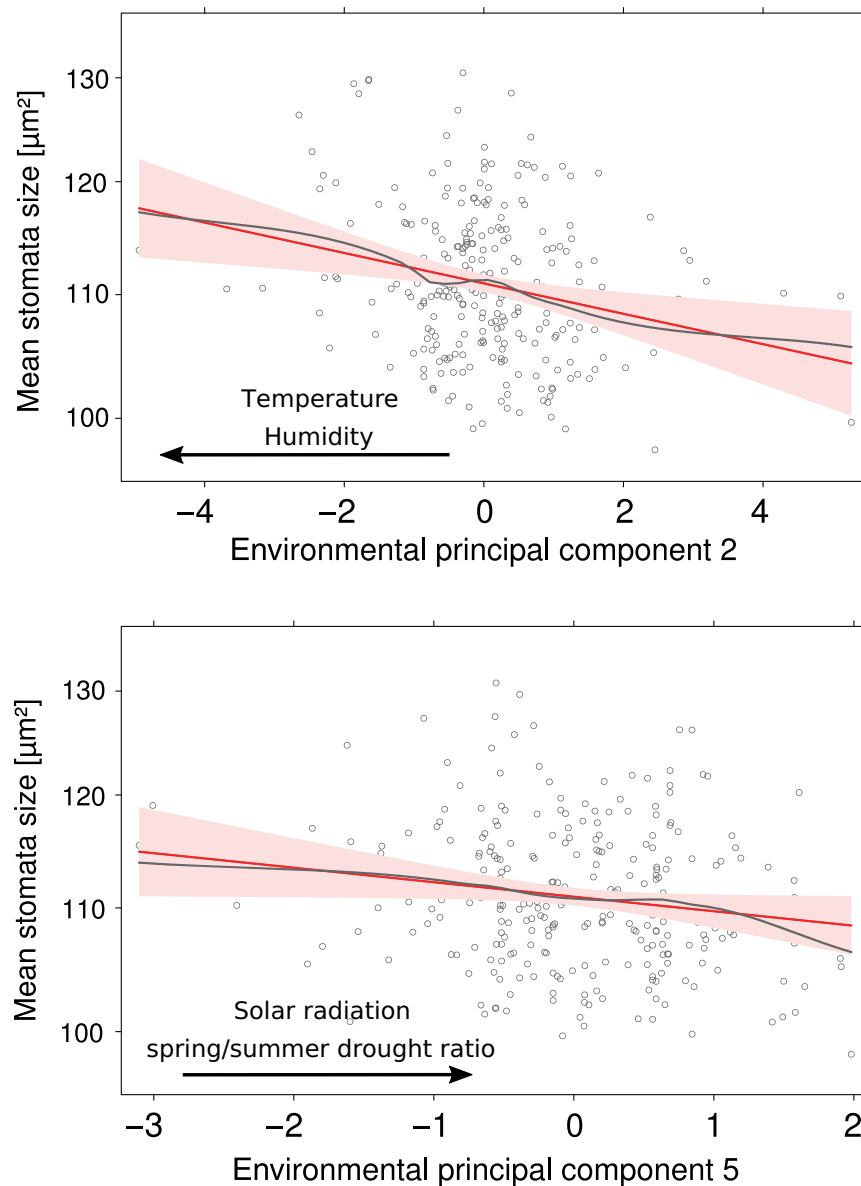


FIGURE 3 Stomata patterns correlate with geographical patterns of climatic variation. Correlation between stomata patterns and seven climatic principal components (PCs) was tested for each phenotype using a generalized linear model (GLM) including genetic population structure as described by the 20 first genetic PCs. Plots are effect plots based on the GLM (see Methods), showing the correlation between stomata size and two climatic PCs. Black arrows indicate correlation with the climatic variables showing the strongest loadings for the respective PC. Plots show the linear fit (red solid line) and the smoothed fit of partial residuals (grey) of the specific predictor. Grey dots are partial residuals. The red shade shows the error of the linear fit. Both PCs shown here are significant predictors of the respective response variable ($p < 0.05$)

3.5 | Patterns of regional differentiation depart from neutral expectations

Genotypes were divided into five regions based on genetic clustering (Alonso-Blanco et al., 2016) and their geographical origin (Supporting Information Figure S1, see Methods). We detected significant phenotypic differentiation among these regions for stomata size (LRT $X^2 = 52.852$, $df = 4$, $p = 9.151e-11$, Figure 4). Stomata size was significantly lower in Southern Sweden (mean = $108 \mu\text{m}^2$) compared to Central Europe (mean = $114 \mu\text{m}^2$, generalized linear hypothesis test (GLHT) $z = -6.24$, $p < 0.001$), Western Europe (mean = $111 \mu\text{m}^2$, GLHT $z = 2.769$, $p = 0.04$) and Spain (mean = $113 \mu\text{m}^2$, GLHT

$z = 6.709$, $p < 0.001$), which did not significantly differ from each other. Northern Sweden showed an intermediate phenotype and did not differ significantly from any region (mean = $110 \mu\text{m}^2$). Variation for stomata density showed a similar but inverted pattern (Supporting Information Figure S24).

Furthermore, we found significant regional differentiation in $\delta^{13}\text{C}$ measurements (LR $X^2 = 58.029$, $df = 4$, $p = 7.525e-12$, Figure 4). Highest $\delta^{13}\text{C}$ levels (highest WUE) were found in accessions from Northern Sweden (mean = -34.8) and Southern Sweden (mean = -35.2), which were significantly higher than in accessions from Spain (mean = -35.7 ; GLHT Southern Sweden $z = -3.472$, $p = 0.008$; GLHT North Sweden $z = -3.49$, $p = 0.001$) and Western

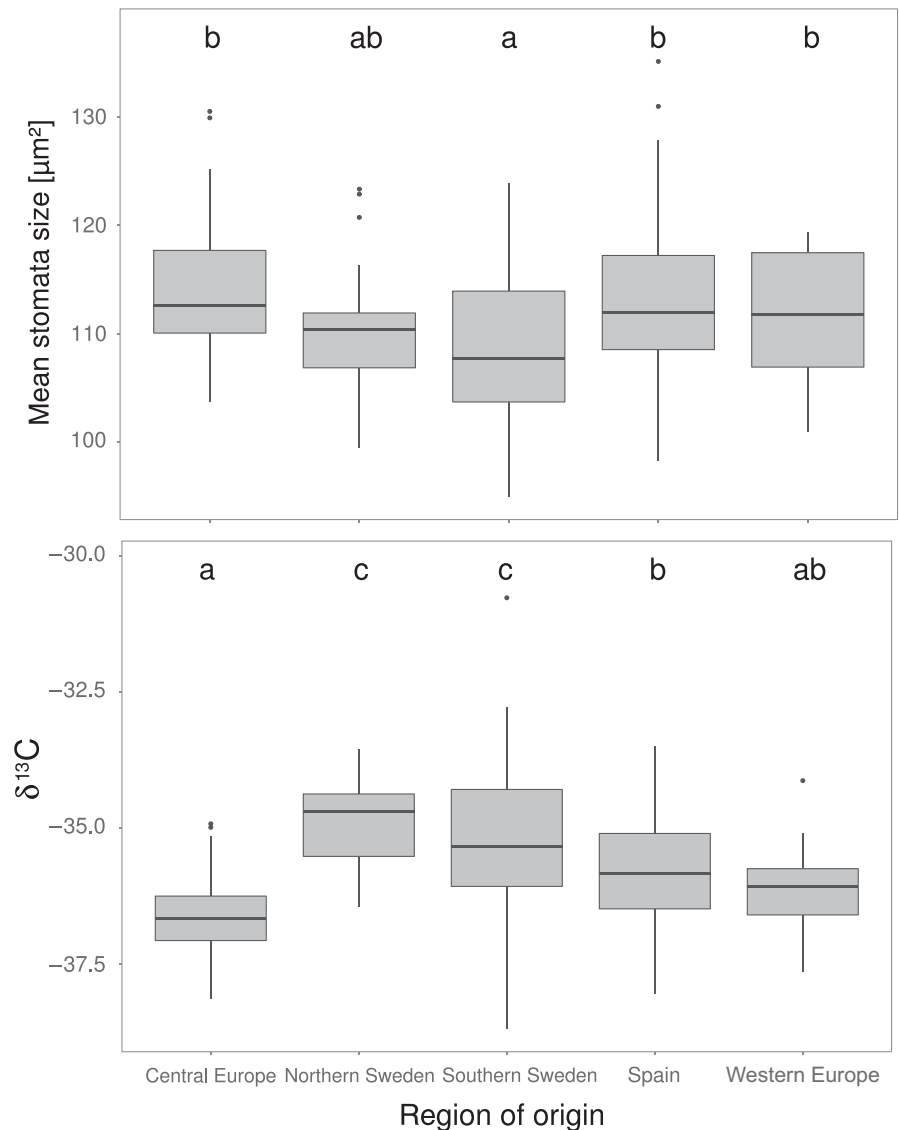


FIGURE 4 Significant regional differentiation of stomata size and $\delta^{13}\text{C}$. *Arabidopsis thaliana* accessions were grouped based on their geographical origin. Box plots show regional differentiation of stomata size (top) and $\delta^{13}\text{C}$ (bottom). Significance of differentiation was tested using generalized linear models followed by a post hoc test. Statistical significance is indicated by letters on top: Groups that do not share a common letter are significantly different. Significance levels: top) a–c, a–bc: $p < 0.001$; ab–c: $p < 0.05$; bottom) a–c, a–b: $p < 0.001$, b–c: $p < 0.01$, ab–c: $p < 0.05$

Europe (mean = -36.06 ; GLHT Southern Sweden $z = -2.8$, $p = 0.03$; GLHT Northern Sweden $z = -3.28$, $p = 0.008$). Lowest $\delta^{13}\text{C}$ levels were found in lines from Central Europe (mean = -36.6), which were significantly lower than in lines from Northern Sweden (GLHT $z = 5.676$, $p < 0.001$), Southern Sweden (GLHT $z = 6.992$, $p < 0.001$) and Spain (GLHT $z = 3.714$, $p = 0.002$).

The observed regional differences result either from the demographic history of the regions or from the action of local selective forces. To tease these possibilities apart, the phenotypic differentiation (Q_{ST}) can be compared to nucleotide differentiation (F_{ST} ; Kronholm et al., 2012; Leinonen et al., 2013). We examined each pair of regions separately, since they are not equidistant from each other and calculated F_{ST} distributions for over 70,000 SNP markers (spaced at least 1 kb apart, see methods). For each trait, Q_{ST} exceeded the 95th percentile of the F_{ST} distribution in at least two pairs of regions (Table 1A–C). We used permutations to calculate a significance threshold for the Q_{ST}/F_{ST} difference (see methods). Significant regional differentiation was pervasive in our sample, with Central Europe and Southern Sweden being significantly

differentiated for all three phenotypes. This analysis suggests that natural selection has contributed to shape the phenotypic differentiation between regions.

Regional differences in climate may have imposed divergence in stomatal patterning. Thus, we estimated climatic distances between regions using estimates of regional effects extracted from a MANOVA. We did not observe significant correlations between adaptive phenotypic divergence ($Q_{ST}-F_{ST}$) and the climatic distance of the respective regions (Mantel test $p > 0.05$ for each of the three traits). Regional divergence in $\delta^{13}\text{C}$, stomata density and stomata size was therefore not proportional to climatic divergence.

4 | DISCUSSION

4.1 | Genetic variation for stomata density and size segregates in *Arabidopsis thaliana*

We used high-throughput confocal imaging to characterize stomatal patterning in over 31,000 images from 870 samples collected from

TABLE 1 (A–C) Patterns of regional differentiation depart from neutral expectations

$Q_{ST}/Q_{ST}-F_{ST}$	Central Europe	North. Sweden	South. Sweden	Spain	West. Europe
(A) Stomata size					
Central Europe		–0.32	0.29	–0.17	–0.13
Northern Sweden	0.15		–0.31	–0.38	–0.51
Southern Sweden	0.41	0.09		0.12	–0.03
Spain	0.01	0.06	0.32		–0.18
West. Europe	0.02	<0.01	0.21	<0.01	
(B) Stomata density					
Central Europe		–0.37	0.31	–0.16	0.17
Northern Sweden	0.09		–0.24	–0.44	–0.49
Southern Sweden	0.44	0.16		0.17	–0.19
Spain	0.01	0.01	0.36		0.07
West. Europe	0.32	0.02	<0.01	0.26	
(C) $\delta^{13}C$					
Central Europe		0.21	0.28	0.13	–0.07
Northern Sweden	0.7		–0.40	–0.16	–0.01
Southern Sweden	0.4	0.01		–0.08	–0.01
Spain	0.30	0.28	0.11		–0.12
West. Europe	0.07	0.40	0.17	0.05	

Note. Pairwise Q_{ST} estimates were derived from linear mixed models for all regions. Genomewide, pairwise F_{ST} distribution was calculated based on 70,000 SNPs for all regions. In the top half of each table, the difference $Q_{ST}-F_{ST}$ for each pair of regions is shown. In the bottom half of each table, the Q_{ST} estimate for each pair of regions is shown. Each table represents one phenotype as indicated by table headlines. Significant $Q_{ST}-F_{ST}$ differences are written in bold. The significance threshold is based on the 95th percentile of a distribution of maximum $Q_{ST}-F_{ST}$ values from 1,000 random permutations of phenotypic data.

330 genotypes. Our high-throughput pipeline could characterize stomata density and stomata size with a reliable accuracy, confirmed by high correlation with manual measurements. Broad-sense heritability and pseudoheritability estimates for stomata density, which are 30% and 58%, respectively, are slightly lower than in a previous report of manually counted stomata diversity across a smaller sample chosen to maximize genetic diversity (Delgado et al., 2011). Despite the clear impact of environmental (random) variance on both observed phenotypes, stomata size and stomata density showed a strong negative correlation. This is consistent with earlier reports of studies manipulating regulators of stomata development (Doheny-Adams, Hunt, Franks, Beerling, & Gray, 2012; Franks et al., 2015), but also with studies analysing stomatal trait variation in a wide range of species (Franks & Beerling, 2009; Hetherington & Woodward, 2003).

The extensive genomic resources available in *A. thaliana* enabled us to investigate the genetic basis of trait variation and covariation, with the help of GWAS (Atwell et al., 2010). Much is known about the molecular pathways that control the differentiation of stomata in *Arabidopsis thaliana*, providing a set of candidate genes expected to control genetic variation in stomatal patterning (Bergmann &

Sack, 2007; Pillitteri & Torii, 2012). However, we did not detect any genomic region that associated with stomata density at a p -value beyond the Bonferroni significance threshold. For stomata size, there was only one significant association on chromosome 4, albeit with very low minor allele frequency in a gene that has not been reported previously in stomata development. GWAS studies can detect small-effect loci only if they segregate at high frequency, whereas rare alleles only give detectable signals when they are of large effect (Korte & Farlow, 2013; Wood et al., 2014). Given that variance for both stomata size and stomata density is clearly heritable, the genetic variants controlling these traits are not causing strong association signals in GWAS. Theoretically, the presence of a large effect QTL impacting local adaptation can be masked by correction for population structure. However, not correcting for population structure is known to lead to a high number of false positives and is thus not a reliable alternative (Vilhjálmsdóttir & Nordborg, 2012). Nevertheless, we can conclude that variation in stomatal patterning is controlled by a combination of (a) alleles of moderate effect size segregating at frequencies too low to be detected by GWAS and/or (b) alleles segregating at high frequency but with effect size too small to be detected and/or (c) rare alleles of small effect. In addition, it is possible that the effect of associated loci is weakened by epistatic interactions among loci. In *A. thaliana*, the genetic architecture of natural variation in stomata traits is therefore not caused by a handful of large effect variants but complex and polygenic.

Using MTMM analysis (Korte et al., 2012), we further investigated the impact of genetic variation on the negative covariation between stomata size and density. This analysis revealed that genetic similarity does not influence the pattern of covariation. It implies that either multiple alleles act epistatically on the covariation or that physical or environmental factors explain the correlation.

4.2 | Natural variation in stomatal patterning can contribute to optimize physiological performance

Both stomata development and reactions to drought stress are being intensively investigated in *A. thaliana* (Bergmann & Sack, 2007; Krasensky & Jonak, 2012; Pillitteri & Torii, 2012; Verslues, Govinal Badiger, Ravi, & Nagaraj, 2013). Mutants in stomata density or size have recently been shown to have a clear impact on carbon physiology (Franks et al., 2015; Hepworth, Doheny-Adams, Hunt, Cameron, & Gray, 2015; Hughes et al., 2017; S. S. Lawson, Pijut, & Michler, 2014; Masle, Gilmore, & Farquhar, 2005; Yoo et al., 2010; Yu et al., 2008). Yet, the relevance of natural variation in stomatal patterning for facing local limitations in water availability had not been documented in this species so far. We provide here concomitant measures of morphological and physiological variation to examine the impact of variation in stomatal patterning on natural variation in WUE. By including genomewide patterns of nucleotide diversity, our analysis presents two major findings: (a) The decrease in stomata size associates with an increase in WUE in *A. thaliana*, and (b) this pattern of covariation has a genetic basis. This shows that, in *A.*

thaliana, variation in stomata size has the potential to be involved in the optimization of physiological processes controlling the trade-off between growth and water loss. Interestingly, in the close relative *A. lyrata* ssp. *lyrata*, stomata were observed to grow smaller in experimental drought compared to well-watered conditions, which coincided with increased WUE (Paccard, Fruleux, & Willi, 2014). This suggests that the consequences of decreased stomata size are conserved in the genus.

While variation for stomata size and density is likely shaped by a complex genetic architecture that hindered QTL detection, we detected two regions in the genome that associated significantly with carbon isotope discrimination. Three previous QTL-mapping analyses, including one between locally adapted lines from Sweden and Italy, identified 16 distinct QTLs controlling $\delta^{13}\text{C}$ (Juenger et al., 2005; McKay et al., 2008; Mojica et al., 2016). One of these is caused by a rare allele in the root-expressed gene *mitogen-activated protein kinase 12* (MPK-12; Campitelli, Des Marais, & Juenger, 2016; Juenger et al., 2005). While QTL-mapping approaches can only reveal the variance shown by the parental lines, GWAS approaches fail to detect rare alleles unless they have a very strong impact. It is therefore not surprising that the loci that stand out in GWAS do not overlap with the QTL previously mapped. In fact, one of the mapping populations used the parental genotype Cvi-0, a genotypic and phenotypic outlier.

The two QTL we report here on chromosomes 2 and 4 add two novel loci, raising the number of genomic regions known to impact $\delta^{13}\text{C}$ in *A. thaliana* to 18. The novel loci we report are locally frequent. Individuals carrying the minor alleles of both loci are almost exclusively from Southern Sweden and display significantly higher $\delta^{13}\text{C}$ than other Southern Swedish accessions. However, we did not find any climatic factor significantly correlated with the allelic states of our QTLs. This suggests that other factors, like soil composition, play a role in drought adaptation. Alternatively, locally adapted alleles may not yet be fixed within the region.

Interestingly, the accessions with the minor allele associating with high $\delta^{13}\text{C}$ in both QTL did not show decreased stomata size compared to other accessions. Multitrait GWAS confirmed that these QTL are associated with $\delta^{13}\text{C}$ variants that are independent of genetic variation for stomatal patterning. We therefore can conclude that stomatal patterning is only one of the traits contributing to the optimization of WUE. A large array of molecular and physiological reactions is indeed known to contribute to tolerance to drought stress (Krasensky & Jonak, 2012; Verslues et al., 2013). The close vicinity of the chromosome 2 QTL to a nonsynonymous mutation in a gene encoding an LEA protein, known to act as a chaperone when cells dehydrate, suggests one possible mechanism by which WUE might be optimized independently of stomata size and density (Candau et al., 2014; Eriksson, Kutzer, Procek, Gröbner, & Harryson, 2011; Reyes et al., 2005). Variation in rates of proline accumulation in the presence of drought stress or in nutrient acquisition in the root is also among the physiological mechanism that appears to have contributed to improve drought stress tolerance in this species (Campitelli et al., 2016; Kesari et al., 2012).

4.3 | Adaptive evolution of stomatal patterning is suggested by the geographical distribution of genetic variation

Phenotypic variation for stomatal patterning and carbon uptake is not uniformly distributed throughout the species range. All three phenotypes we report in this study were significantly differentiated between the five broad regions defined in our sample of 330 genotypes. We performed a comparison of phenotypic and nucleotide levels of divergence to evaluate the putative role of past selective events in shaping the distribution of diversity we report (Leinonen et al., 2013; Whitlock & Guillaume, 2009). Because these regions are not equally distant, $F_{\text{ST}}/Q_{\text{ST}}$ comparisons averaged over all populations may mask local patterns of adaptation (Leinonen et al., 2013). We therefore measured Q_{ST} between pairs of regions and compared them to the distribution of pairwise F_{ST} , using permutations to establish the significance of outlier Q_{ST} . This analysis showed that, for all three traits, differentiation between some regions was stronger than expected from genomewide patterns of diversity, suggesting local adaptation. This is further supported by our finding that stomata density and stomata size correlated with climatic PCs, which are most strongly driven by temperature, humidity, solar radiation and historic drought regimen.

The strongest $Q_{\text{ST}}-F_{\text{ST}}$ differences are found across regional pairs including Central Europe. Particularly, WUE is significantly differentiated between Central Europe and Spain as well as both Swedish regions, due to low WUE in Central Europe. It is tempting to speculate that the significantly lower WUE observed in Central Europe results from selection for life cycling at latitudes where two life cycles can be completed each year, as high WUE is usually associated with a reduction in photosynthetic rate (Blum, 2009; Field et al., 1983; Kimball et al., 2014). Interestingly, Central Europe and Southern Sweden are significantly differentiated for all three traits and Southern Sweden and Spain are significantly differentiated for both stomata traits. Combined with the fact that Swedish genotypes show the highest values for WUE, this suggests that stomata size is involved in drought adaptation of Swedish accessions. This result is somewhat counterintuitive because Sweden is not known to be a region experiencing intense drought. However, our result is supported by an independent study showing that Northern and Southern Swedish genotypes maintain photosynthetic activity under terminal drought stress longer than other, especially Central and Western European, accessions (Exposito-Alonso et al., 2017). Additionally, locally adapted genotypes from Northern Sweden (which showed high WUE in our study, as well) have been shown to display higher WUE than Italian genotypes (Mojica et al., 2016).

This regional difference in *A. thaliana* further coincides with the satellite measurements of historic drought regimen, which show that Sweden is a region where drought frequency is changing throughout the season: It is relatively more frequent in the early growing season (spring) than in the late growing season (summer). Drought episodes occurring earlier in the growth season may favour the evolution of drought avoidance traits (e.g., morphological or physiological stress

adaptations) over that of escape strategies mediated by, for example, seed dormancy (Kooyers, 2015; Passioura, 1996). Indeed, in Northern Europe, increased negative covariation between flowering time and seed dormancy suggested that the narrow growth season imposes a strong selection on life history traits (Debieu et al., 2013). In Southern European regions, *A. thaliana* appeared to rely on escape strategies provided by increased seed dormancy (Kronholm et al., 2012). Taken together, this suggests that decreased stomata size and, consequently, increased $\delta^{13}\text{C}$ have contributed to adaptation to water limitations in spring in a region where the narrow growth season leaves no room for escape strategies. Indeed, both stomata size and $\delta^{13}\text{C}$ associate with historic drought regimen. For $\delta^{13}\text{C}$, however, this association disappears when genetic population structure is included as a covariate. This indicates that local adaptation for WUE might have contributed to shape current population structure.

Finally, the coarse regional contrasts used in the present study cannot resolve patterns of local adaptation occurring at a fine-grained scale within regions (as e.g., local adaptation to specific soil patches). In fact, we observe most variation for all three phenotypes within regions. It is therefore possible that we underestimate the magnitude of adaptive differentiation across the species' European range, which could further explain why Q_{ST}/F_{ST} differences did not covary with environmental divergence in our data set.

5 | CONCLUSION

This work provides a comprehensive description of the variation in stomata size and density that segregates throughout the European range of *A. thaliana*. It shows that stomata size covaries with water-use efficiency and may contribute to local adaptation. Several reports indicate that plants can also change stomatal development in water-limiting conditions (Fraser, Greenall, Carlyle, Turkington, & Friedman, 2009; Paccard et al., 2014; Xu & Zhou, 2008). Future work will have to investigate whether this variation in stomata size and number also contributes to adaptive plasticity to drought stress.

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AUTHOR CONTRIBUTION

J.d.M., A.K. and H.D. conceived the study. H.D. conducted the experiment and produced phenotypic data for stomata traits. T.M. and A.W. were responsible for $\delta^{13}\text{C}$ measurements. G.M. provided

data on historic drought regimen. J.d.M., A.K. and H.D. were responsible for the statistical analyses of the data. J.d.M. and H.D. wrote the manuscript with significant contributions from A.K., T.M., A.W. and G.M.

DATA ACCESSIBILITY

Raw image data and image analysis scripts are stored in a Dryad repository (<https://doi.org/10.5061/dryad.n068q74>). Phenotypic data are provided as supplemental material and included in the Dryad repository. Additionally, genotypic means of each phenotype were uploaded to the AraPheno database (<https://arapheno.1001genomes.org>, (Seren et al., 2017)). We provide an R Markdown file (Supporting Information Appendix S2), which contains all figures (except GWAS and MTMM) and the corresponding R code used to create the figures and statistics in the supplemental material. GWAS scripts are available at <https://github.com/arthurkorte/GWAS>. MTMM scripts are available at <https://github.com/Gregor-Mendel-Institute/mtmm>. Genomic data used are publicly available in the 1001 genomes database (Alonso-Blanco et al., 2016).

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REFERENCES

- Alonso-Blanco, C., Andrade, J., Becker, C., Bemm, F., Bergelson, J., Borgwardt, K. M., ... Zhou, X. (2016). 1,135 genomes reveal the global pattern of polymorphism in *Arabidopsis thaliana*. *Cell*, 166(2), 481–491. <https://doi.org/10.1016/j.cell.2016.05.063>
- Anderson, V. J., & Briske, D. D. (1990). Stomatal distribution, density and conductance of three perennial grasses native to the southern true prairie of Texas. *The American Midland Naturalist*, 123(1), 152–159. <https://doi.org/10.2307/2425768>
- Arnold, J. B., Daroczi, G., Werth, B., Weitzner, B., Kunst, J., Auguie, B., ... London, J. (2017). ggthemes: Extra Themes, Scales and Geoms for "ggplot2" (Version 3.4.0). Retrieved from <https://cran.r-project.org/web/packages/ggthemes/index.html>
- Atwell, S., Huang, Y. S., Vilhjálmsson, B. J., Willems, G., Horton, M., Li, Y., ... Nordborg, M. (2010). Genome-wide association study of 107 phenotypes in *Arabidopsis thaliana* inbred lines. *Nature*, 465(7298), 627–631. <https://doi.org/10.1038/nature08800>
- Bakker, J. C. (1991). Effects of humidity on stomatal density and its relation to leaf conductance. *Scientia Horticulturae*, 48(3), 205–212. [https://doi.org/10.1016/0304-4238\(91\)90128-L](https://doi.org/10.1016/0304-4238(91)90128-L)
- Bergmann, D. C., & Sack, F. D. (2007). Stomatal development. *Annual Review of Plant Biology*, 58(1), 163–181. <https://doi.org/10.1146/annurev.arplant.58.032806.104023>
- Blum, A. (2009). Effective use of water (EUW) and not water-use efficiency (WUE) is the target of crop yield improvement under drought stress. *Field Crops Research*, 112(2), 119–123. <https://doi.org/10.1016/j.fcr.2009.03.009>
- Browning, B. L., & Browning, S. R. (2009). A unified approach to genotype imputation and haplotype-phase inference for large data sets of trios and unrelated individuals. *The American Journal of Human Genetics*, 84(2), 210–223. <https://doi.org/10.1016/j.ajhg.2009.01.005>

- Burghardt, L. T., Metcalf, C. J. E., Wilczek, A. M., Schmitt, J., & Donohue, K. (2015). Modeling the influence of genetic and environmental variation on the expression of plant life cycles across landscapes. *The American Naturalist*, 185(2), 212–227. <https://doi.org/10.1086/679439>
- Campitelli, B. E., Des Marais, D. L., & Juenger, T. E. (2016). Ecological interactions and the fitness effect of water-use efficiency: Competition and drought alter the impact of natural MPK12 alleles in *Arabidopsis*. *Ecology Letters*, 19(4), 424–434. <https://doi.org/10.1111/ele.12575>
- Candat, A., Paszkiewicz, G., Neveu, M., Gautier, R., Logan, D. C., Avelange-Macherel, M.-H., & Macherel, D. (2014). The ubiquitous distribution of late embryogenesis abundant proteins across cell compartments in *Arabidopsis* offers tailored protection against abiotic stress. *The Plant Cell*, 26(7), 3148–3166. <https://doi.org/10.1105/tpc.114.127316>
- Carlson, J. E., Adams, C. A., & Holsinger, K. E. (2016). Intraspecific variation in stomatal traits, leaf traits and physiology reflects adaptation along aridity gradients in a South African shrub. *Annals of Botany*, 117(1), 195–207. <https://doi.org/10.1093/aob/mcv146>
- Carroll, S. P., Hendry, A. P., Reznick, D. N., & Fox, C. W. (2007). Evolution on ecological time-scales. *Functional Ecology*, 21(3), 387–393. <https://doi.org/10.1111/j.1365-2435.2007.01289.x>
- Chater, C., Kamisugi, Y., Movahedi, M., Fleming, A., Cumming, A. C., Gray, J. E., & Beerling, D. J. (2011). Regulatory mechanism controlling stomatal behavior conserved across 400 million years of land plant evolution. *Current Biology: CB*, 21(12), 1025–1029. <https://doi.org/10.1016/j.cub.2011.04.032>
- Cowan, I. R. (1986). Economics of carbon fixation in higher plants. *On the Economy of Plant Form and Function: Proceedings of the Sixth Maria Moors Cabot Symposium, Evolutionary Constraints on Primary Productivity, Adaptive Patterns of Energy Capture in Plants, Harvard Forest, August 1983*. Retrieved from <http://agris.fao.org/agris-search/search.do?recordID=US201301399273>
- Cowan, I. R., & Farquhar, G. D. (1977). Stomatal function in relation to leaf metabolism and environment. *Symposia of the Society for Experimental Biology*, 31, 471–505.
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., ... Durbin, R. (2011). The variant call format and VCFtools. *Bioinformatics*, 27(15), 2156–2158. <https://doi.org/10.1093/bioinformatics/btr330>
- Daszkowska-Golec, A., & Szarejko, I. (2013). Open or close the gate – stomata action under the control of phytohormones in drought stress conditions. *Frontiers in Plant Science*, 4, 138. <https://doi.org/10.3389/fpls.2013.00138>
- Debieu, M., Tang, C., Stich, B., Sikosek, T., Effgen, S., Josephs, E., ... de Meaux, J. (2013). Co-variation between seed dormancy, growth rate and flowering time changes with latitude in *Arabidopsis thaliana*. *PLoS ONE*, 8(5), e61075. <https://doi.org/10.1371/journal.pone.0061075>
- Delgado, D., Alonso-Blanco, C., Fenoll, C., & Mena, M. (2011). Natural variation in stomatal abundance of *Arabidopsis thaliana* includes cryptic diversity for different developmental processes. *Annals of Botany*, 107(8), 1247–1258. <https://doi.org/10.1093/aob/mcr060>
- Doheny-Adams, T., Hunt, L., Franks, P. J., Beerling, D. J., & Gray, J. E. (2012). Genetic manipulation of stomatal density influences stomatal size, plant growth and tolerance to restricted water supply across a growth carbon dioxide gradient. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 367(1588), 547–555. <https://doi.org/10.1098/rstb.2011.0272>
- Drake, P. L., Froend, R. H., & Franks, P. J. (2013). Smaller, faster stomata: Scaling of stomatal size, rate of response, and stomatal conductance. *Journal of Experimental Botany*, 64(2), 495–505. <https://doi.org/10.1093/jxb/ers347>
- Eriksson, S. K., Kutzer, M., Procek, J., Gröbner, G., & Harryson, P. (2011). Tunable membrane binding of the intrinsically disordered dehydrin Lti30, a cold-induced plant stress protein. *The Plant Cell*, 23(6), 2391–2404. <https://doi.org/10.1105/tpc.111.085183>
- Exposito-Alonso, M., Vasseur, F., Ding, W., Wang, G., Burbano, H. A. A., & Weigel, D. (2017). Genomic basis and evolutionary potential for extreme drought adaptation in *Arabidopsis thaliana*. *BioRxiv*, 118067. <https://doi.org/10.1101/118067>
- Farquhar, G. D., Hubick, K. T., Condon, A. G., & Richards, R. A. (1989). Carbon isotope fractionation and plant water-use efficiency. In P. W. Rundel, J. R. Ehleringer, & K. A. Nagy (Eds.), *Stable isotopes in ecological research* (pp. 21–40). New York, NY: Springer, New York. <https://doi.org/10.1007/978-1-4612-3498-2>
- Fick, S. E., & Hijmans, R. J. (2017). WorldClim 2: New 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology*, 37(12), 4302–4315.
- Field, C., Merino, J., & Mooney, H. A. (1983). Compromises between water-use efficiency and nitrogen-use efficiency in five species of California Evergreens. *Oecologia*, 60(3), 384–389. <https://doi.org/10.1007/BF00376856>
- Fournier-Level, A., Korte, A., Cooper, M. D., Nordborg, M., Schmitt, J., & Wilczek, A. M. (2011). A map of local adaptation in *Arabidopsis thaliana*. *Science*, 334(6052), 86–89. <https://doi.org/10.1126/science.1209271>
- Fox, J., Weisberg, S., Friendly, M., Hong, J., Andersen, R., Firth, D., & Taylor, S. (2016). effects: Effect Displays for Linear, Generalized Linear, and Other Models (Version 3.1-2). Retrieved from <https://cran.r-project.org/web/packages/effects/index.html>
- Franks, P. J., & Beerling, D. J. (2009). Maximum leaf conductance driven by CO₂ effects on stomatal size and density over geologic time. *Proceedings of the National Academy of Sciences*, 106(25), 10343–10347. <https://doi.org/10.1073/pnas.0904209106>
- Franks, P. J., Doheny-Adams, T. W., Britton-Harper, Z. J., & Gray, J. E. (2015). Increasing water-use efficiency directly through genetic manipulation of stomatal density. *New Phytologist*, 207(1), 188–195. <https://doi.org/10.1111/nph.13347>
- Franks, P. J., Drake, P. L., & Beerling, D. J. (2009). Plasticity in maximum stomatal conductance constrained by negative correlation between stomatal size and density: An analysis using *Eucalyptus globulus*. *Plant, Cell and Environment*, 32(12), 1737–1748. <https://doi.org/10.1111/j.1365-3040.2009.02031.x>
- Fraser, L. H., Greenall, A., Carlyle, C., Turkington, R., & Friedman, C. R. (2009). Adaptive phenotypic plasticity of *Pseudoroegneria spicata*: Response of stomatal density, leaf area and biomass to changes in water supply and increased temperature. *Annals of Botany*, 103(5), 769–775. <https://doi.org/10.1093/aob/mcn252>
- Gaut, B. (2012). *Arabidopsis thaliana* as a model for the genetics of local adaptation. *Nature Genetics*, 44(2), 115–116. <https://doi.org/10.1038/ng.1079>
- Goudet, J. (2005). hierfstat, a package for R to compute and test hierarchical F-statistics. *Molecular Ecology Notes*, 5(1), 184–186. <https://doi.org/10.1111/j.1471-8286.2004.00828.x>
- Hamilton, J. A., Okada, M., Korves, T., & Schmitt, J. (2015). The role of climate adaptation in colonization success in *Arabidopsis thaliana*. *Molecular Ecology*, 24(9), 2253–2263. <https://doi.org/10.1111/mec.13099>
- Hancock, A. M., Brachi, B., Faure, N., Horton, M. W., Jarymowycz, L. B., Sperone, F. G., ... Bergelson, J. (2011). Adaptation to climate across the *Arabidopsis thaliana* genome. *Science*, 334(6052), 83–86. <https://doi.org/10.1126/science.1209244>
- Hendry, A. P. (2016). *Eco-evolutionary dynamics*. Princeton, NJ: Princeton University Press.
- Hepworth, C., Doheny-Adams, T., Hunt, L., Cameron, D. D., & Gray, J. E. (2015). Manipulating stomatal density enhances drought tolerance without deleterious effect on nutrient uptake. *The New Phytologist*, 208(2), 336–341. <https://doi.org/10.1111/nph.13598>
- Hetherington, A. M., & Woodward, F. I. (2003). The role of stomata in sensing and driving environmental change. *Nature*, 424(6951), 901–908. <https://doi.org/10.1038/nature01843>

- Hothorn, T., Bretz, F., Westfall, P., Heiberger, R. M., Schuetzenmeister, A., & Scheibe, S. (2017). multcomp: Simultaneous Inference in General Parametric Models (Version 1.4-7). Retrieved from <https://cran.r-project.org/web/packages/multcomp/index.html>
- Hughes, J., Hepworth, C., Dutton, C., Dunn, J. A., Hunt, L., Stephens, J., ... Gray, J. E. (2017). Reducing stomatal density in barley improves drought tolerance without impacting on yield. *Plant Physiology*, 174(2), 776–787. <https://doi.org/10.1104/pp.16.01844>
- Jombart, T., Kamvar, Z. N., Lustrik, R., Collins, C., Beugin, M.-P., Knaus, B., ... Calboli, F. (2016). adegenet: Exploratory Analysis of Genetic and Genomic Data (Version 2.0.1). Retrieved from <https://cran.r-project.org/web/packages/adegenet/index.html>
- Juenger, T. E., McKay, J. K., Hausmann, N., Keurentjes, J. J. B., Sen, S., Stowe, K. A., ... Richards, J. H. (2005). Identification and characterization of QTL underlying whole-plant physiology in *Arabidopsis thaliana*: $\delta^{13}\text{C}$, stomatal conductance and transpiration efficiency. *Plant, Cell and Environment*, 28(6), 697–708. <https://doi.org/10.1111/j.1365-3040.2004.01313.x>
- Kahle, D., & Wickham, H. (2013). ggmap: Spatial visualization with ggplot2. *The R Journal*, 5(1), 144–161.
- Kang, H. M., Sul, J. H., Service, S. K., Zaitlen, N. A., Kong, S., Freimer, N. B., ... Eskin, E. (2010). Variance component model to account for sample structure in genome-wide association studies. *Nature Genetics*, 42(4), 348–354. <https://doi.org/10.1038/ng.548>
- Kang, H. M., Zaitlen, N. A., Wade, C. M., Kirby, A., Heckerman, D., Daly, M. J., & Eskin, E. (2008). Efficient control of population structure in model organism association mapping. *Genetics*, 178(3), 1709–1723. <https://doi.org/10.1534/genetics.107.080101>
- Kesari, R., Lasky, J. R., Villamor, J. G., Marais, D. L. D., Chen, Y.-J. C., Liu, T.-W., ... Verslues, P. E. (2012). Intron-mediated alternative splicing of *Arabidopsis* P5CS1 and its association with natural variation in proline and climate adaptation. *Proceedings of the National Academy of Sciences*, 109(23), 9197–9202. <https://doi.org/10.1073/pnas.1203433109>
- Kimball, S., Gremer, J. R., Barron-Gafford, G. A., Angert, A. L., Huxman, T. E., & Venable, D. L. (2014). High water-use efficiency and growth contribute to success of non-native *Erodium cicutarium* in a Sonoran Desert winter annual community. *Conservation Physiology*, 2(1), cou006. <https://doi.org/10.1093/conphys/cou006>
- Kinoshita, T., Ono, N., Hayashi, Y., Morimoto, S., Nakamura, S., Soda, M., ... Shimazaki, K. (2011). FLOWERING LOCUS T regulates stomatal opening. *Current Biology*, 21(14), 1232–1238. <https://doi.org/10.1016/j.cub.2011.06.025>
- Knaus, B. J., Grunwald, N. J., Anderson, E. C., Winter, D. J., & Kamvar, Z. N. (2017). vcfR: Manipulate and Visualize VCF Data (Version 1.4.0). Retrieved from <https://cran.r-project.org/web/packages/vcfR/index.html>
- Kogan, F. N. (1995). Application of vegetation index and brightness temperature for drought detection. *Advances in Space Research*, 15(11), 91–100. [https://doi.org/10.1016/0273-1177\(95\)00079-T](https://doi.org/10.1016/0273-1177(95)00079-T)
- Kooyers, N. J. (2015). The evolution of drought escape and avoidance in natural herbaceous populations. *Plant Science*, 234, 155–162. <https://doi.org/10.1016/j.plantsci.2015.02.012>
- Korte, A., & Farlow, A. (2013). The advantages and limitations of trait analysis with GWAS: A review. *Plant Methods*, 9, 29. <https://doi.org/10.1186/1746-4811-9-29>
- Korte, A., Vilhjálmsson, B. J., Segura, V., Platt, A., Long, Q., & Nordborg, M. (2012). A mixed-model approach for genome-wide association studies of correlated traits in structured populations. *Nature Genetics*, 44(9), 1066–1071. <https://doi.org/10.1038/ng.2376>
- Krasensky, J., & Jonak, C. (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *Journal of Experimental Botany*, 63(4), 1593–1608. <https://doi.org/10.1093/jxb/err460>
- Kronholm, I., Picó, F. X., Alonso-Blanco, C., Goudet, J., & de Meaux, J. (2012). Genetic basis of adaptation in *Arabidopsis thaliana*: Local adaptation at the seed dormancy Qtl Dog1. *Evolution*, 66(7), 2287–2302. <https://doi.org/10.1111/j.1558-5646.2012.01590.x>
- Lambers, H., Chapin, F. S., & Pons, T. L. (Eds.). (1998). Photosynthesis, respiration, and long-distance transport. In *Plant physiological ecology* (pp. 10–153). New York, NY: Springer.
- Lasky, J. R., Marais, D. L. D., Lowry, D. B., Povolotskaya, I., McKay, J. K., ... Juenger, T. E. (2014). Natural variation in abiotic stress responsive gene expression and local adaptation to climate in *Arabidopsis thaliana*. *Molecular Biology and Evolution*, 31(9), 2283–2296. <https://doi.org/10.1093/molbev/msu170>
- Lawson, T., Kramer, D. M., & Raines, C. A. (2012). Improving yield by exploiting mechanisms underlying natural variation of photosynthesis. *Current Opinion in Biotechnology*, 23(2), 215–220. <https://doi.org/10.1016/j.copbio.2011.12.012>
- Lawson, S. S., Pijut, P. M., & Michler, C. H. (2014). The cloning and characterization of a poplar stomatal density gene. *Genes and Genomics*, 36(4), 427–441. <https://doi.org/10.1007/s13258-014-0177-x>
- Leinonen, T., McCairns, R. S., O'hara, R. B., & Merilä, J. (2013). QST–FST comparisons: Evolutionary and ecological insights from genomic heterogeneity. *Nature Reviews Genetics*, 14(3), 179–190. <https://doi.org/10.1038/nrg3395>
- Martin, B., & Thorntenson, Y. R. (1988). Stable carbon isotope composition ($\delta^{13}\text{C}$), water use efficiency, and biomass productivity of *Lycopersicon esculentum*, *Lycopersicon pennellii*, and the F1 hybrid. *Plant Physiology*, 88(1), 213–217. <https://doi.org/10.1104/pp.88.1.213>
- Masle, J., Gilmore, S. R., & Farquhar, G. D. (2005). The ERECTA gene regulates plant transpiration efficiency in *Arabidopsis*. *Nature*, 436(7052), 866. <https://doi.org/10.1038/nature03835>
- McDowell, N., Pockman, W. T., Allen, C. D., Breshears, D. D., Cobb, N., Kolb, T., ... Yezzer, E. A. (2008). Mechanisms of plant survival and mortality during drought: Why do some plants survive while others succumb to drought? *New Phytologist*, 178(4), 719–739. <https://doi.org/10.1111/j.1469-8137.2008.02436.x>
- McKay, J. K., Richards, J. H., Nemali, K. S., Sen, S., Mitchell-Olds, T., Boles, S., ... Juenger, T. E. (2008). Genetics of drought adaptation in *Arabidopsis thaliana* li. Qtl analysis of a new mapping population, Kas-1 \times Tsu-1. *Evolution*, 62(12), 3014–3026. <https://doi.org/10.1111/j.1558-5646.2008.00474.x>
- Mojica, J. P., Mullen, J., Lovell, J. T., Monroe, J. G., Paul, J. R., Oakley, C. G., & McKay, J. K. (2016). Genetics of water use physiology in locally adapted *Arabidopsis thaliana*. *Plant Science*, 251(Suppl C), 12–22. <https://doi.org/10.1016/j.plantsci.2016.03.015>
- Muchow, R. C., & Sinclair, T. R. (1989). Epidermal conductance, stomatal density and stomatal size among genotypes of *Sorghum bicolor* (L.) Moench. *Plant, Cell and Environment*, 12(4), 425–431. <https://doi.org/10.1111/j.1365-3040.1989.tb01958.x>
- Nordborg, M., Borevitz, J. O., Bergelson, J., Berry, C. C., Chory, J., Hagenblad, J., ... Weigel, D. (2002). The extent of linkage disequilibrium in *Arabidopsis thaliana*. *Nature Genetics*, 30(2), 190–193. <https://doi.org/10.1038/ng813>
- Ohsumi, A., Kanemura, T., Homma, K., Horie, T., & Shiraiwa, T. (2007). Genotypic variation of stomatal conductance in relation to stomatal density and length in rice (*Oryza sativa* L.). *Plant Production Science*, 10(3), 322–328. <https://doi.org/10.1626/pp.10.322>
- Paccard, A., Fruleux, A., & Willi, Y. (2014). Latitudinal trait variation and responses to drought in *Arabidopsis lyrata*. *Oecologia*, 175(2), 577–587. <https://doi.org/10.1007/s00442-014-2932-8>
- Passioura, J. B. (1996). Drought and drought tolerance. In E. Belhassen (Ed.), *Drought tolerance in higher plants: Genetical, physiological and molecular biological analysis* (pp. 1–5). Dordrecht, Netherlands: Springer.
- Pearce, D. W., Millard, S., Bray, D. F., & Rood, S. B. (2006). Stomatal characteristics of riparian poplar species in a semi-arid environment. *Tree*

- Physiology*, 26(2), 211–218. <https://doi.org/10.1093/treephys/26.2.211>
- Pillitteri, L. J., & Torii, K. U. (2012). Mechanisms of stomatal development. *Annual Review of Plant Biology*, 63(1), 591–614. <https://doi.org/10.1146/annurev-arplant-042811-105451>
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., & R Core Team. (2015). *nlme: Linear and Nonlinear Mixed Effects Models*. Retrieved from <http://CRAN.R-project.org/package=nlme>
- Postma, F. M., & Ågren, J. (2016). Early life stages contribute strongly to local adaptation in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences*, 113(27), 7590–7595. <https://doi.org/10.1073/pnas.1606303113>
- R Development Core Team. (2008). *R: A language and environment for statistical computing*. Retrieved from <http://www.R-project.org>
- Raven, J. A. (2002). Selection pressures on stomatal evolution. *New Phytologist*, 153(3), 371–386. <https://doi.org/10.1046/j.0028-646X.2001.00334.x>
- Raven, J. A. (2014). Speedy small stomata? *Journal of Experimental Botany*, 65(6), 1415–1424. <https://doi.org/10.1093/jxb/eru032>
- Reich, P. B. (1984). Leaf stomatal density and diffusive conductance in three amphistomatous hybrid poplar cultivars. *New Phytologist*, 98(2), 231–239. <https://doi.org/10.1111/j.1469-8137.1984.tb02733.x>
- Reyes, J. L., Rodrigo, M.-J., Colmenero-Flores, J. M., Gil, J.-V., Garay-Arroyo, A., Campos, F., ... Covarrubias, A. A. (2005). Hydrophilins from distant organisms can protect enzymatic activities from water limitation effects in vitro. *Plant, Cell and Environment*, 28(6), 709–718. <https://doi.org/10.1111/j.1365-3040.2005.01317.x>
- Seren, Ü., Grimm, D., Fitz, J., Weigel, D., Nordborg, M., Borgwardt, K., & Korte, A. (2017). AraPheno: A public database for *Arabidopsis thaliana* phenotypes. *Nucleic Acids Research*, 45(D1), D1054–D1059. <https://doi.org/10.1093/nar/gkw986>
- Trabucco, A., & Zomer, R. (2010). Global soil water balance geospatial database. *CGIAR Consortium for Spatial Information, Published Online*. Retrieved from <http://www.cgiar-csi.org>
- Verslues, P. E., Govinal Badiger, B., Ravi, K., & Nagaraj, M. (2013). Drought tolerance mechanisms and their molecular basis. In T. A. Jenks, & P. M. Hasegawa (Eds.), *Plant abiotic stress* (pp. 15–46). Hoboken, NJ: John Wiley & Sons Inc.
- Vilhjálmsdóttir, B. J., & Nordborg, M. (2012). The nature of confounding in genome-wide association studies [Comments and Opinion]. <https://doi.org/10.1038/nrg3382>
- Vu, V. Q. (2011). A ggplot2 based biplot (Version 0.55).
- Weinig, C., Ewers, B. E., & Welch, S. M. (2014). Ecological genomics and process modeling of local adaptation to climate. *Current Opinion in Plant Biology*, 18(Suppl C), 66–72. <https://doi.org/10.1016/j.pbi.2014.02.007>
- Whitlock, M. C., & Guillaume, F. (2009). Testing for spatially divergent selection: Comparing QST to FST. *Genetics*, 183(3), 1055–1063. <https://doi.org/10.1534/genetics.108.099812>
- Wickham, H. (2009). *Ggplot2: Elegant graphics for data analysis*. New York, NY: Springer. <https://doi.org/10.1007/978-0-387-98141-3>
- Wood, A. R., Esko, T., Yang, J., Vedantam, S., Pers, T. H., Gustafsson, S., ... Frayling, T. M. (2014). Defining the role of common variation in the genomic and biological architecture of adult human height. *Nature Genetics*, 46(11), 1173. <https://doi.org/10.1038/ng.3097>
- Wu, C. A., Lowry, D. B., Nutter, L. I., & Willis, J. H. (2010). Natural variation for drought-response traits in the *Mimulus guttatus* species complex. *Oecologia*, 162(1), 23–33. <https://doi.org/10.1007/s00442-009-1448-0>
- Xu, Z., & Zhou, G. (2008). Responses of leaf stomatal density to water status and its relationship with photosynthesis in a grass. *Journal of Experimental Botany*, 59(12), 3317–3325. <https://doi.org/10.1093/jxb/ern185>
- Yoo, C. Y., Pence, H. E., Jin, J. B., Miura, K., Gosney, M. J., Hasegawa, P. M., & Mickelbart, M. V. (2010). The *Arabidopsis* GTL1 transcription factor regulates water use efficiency and drought tolerance by modulating stomatal density via transrepression of SDD1. *The Plant Cell Online*, 22(12), 4128–4141. <https://doi.org/10.1105/tpc.110.078691>
- Yu, H., Chen, X., Hong, Y.-Y., Wang, Y., Xu, P., Ke, S.-D., ... Xiang, C.-B. (2008). Activated expression of an *Arabidopsis* HD-START protein confers drought tolerance with improved root system and reduced stomatal density. *The Plant Cell*, 20(4), 1134–1151. <https://doi.org/10.1105/tpc.108.058263>

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