panda: pangenome annotations data analysis

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Warning Messages

With impg 0.2.0 it is normal to get a warning message stating:

thread 'main' panicked at src/main.rs:205:56:

Target name not found in index

This is related to the behaviour of wfmash who, given to fastas A and B, does not align both A vs B and B vs A but instead only aligns A vs B.

TODO

Retrieve the sequences of each block and save them? Probably this is not a wise thing to do. With the nnew output format it is easy to do this on the fly.

Input Files and Formats

The input files are: 1) an alignment, 2) the annotations of the contigs aligned, 3) a properly (priority) sorted list of ids.

The alignment file is in paf format. The delimiter (see https://github.com/pangenome/PanSN-spec) is "#". The annotations are in gff format. The file are named using the "-" separator. E.g. in the alignment we may have in line 1 and column 1: "SGDref#0#chrI". The corresponding annotations file is named "SGDref-0-features.gff" and it reports all the annotations for all the corresponding contigs.

Quick Run

Before running the markdown remember to:

- check the input data folder of the annotation files (dir_anno)
- check all chunks with eval = FALSE
- make a copy of the input alignment in paf format ("/.../aln/graph-name.paf")
- make the input ("/.../ids/ids-ps.txt") file with the ids (e.g. "SGDref-0", "AIF-1", "AIF-2") of the strains used to build the graph induced from the alignment
- choose if mitochondrial features should be used (attached to the first available haplotype)

Analysis

```
## settings ------
### with mitochondrial sequence in one haplotype?
with_mito="FALSE"

### the gff files repository
```

```
dir_anno="${HOME}/data/nano-assemblies-pansn-2024/annotations"
## clmnt -----
### folders
dir_base=$(dirname "${PWD}")
dir_out="${dir_base}/anno/gff"
if [[ -d "${dir_out}" ]]; then rm -rf "${dir_out}"; fi
mkdir -p "${dir_out}"
### the vector with the strain-haplotypes ids
declare -a gff_ids
file_ids="${dir_base}/ids/ids-ps.txt"
while IFS="\n" read -r one_line; do
   gff_ids+=("${one_line}")
done < "${file_ids}"</pre>
for strainhaplo_id in "${gff_ids[0]}"; do
  path_gff=$(find "${dir_anno}" -name "${strainhaplo_id}*gff")
  name_gff=$(basename "${path_gff}")
  strain_id=$(basename "${path_gff}" | cut -f 1 -d "-") # e.g. ADE
  mito_name="${strain_id}-mt-features.gff"
  path_mito_gff=$(find "${dir_anno}" -name "${mito_name}")
  gff_type=$(basename "${path_gff}" | cut -f 2 -d "-") # e.g. 0 or 1
  if [[ "${with mito}" == "TRUE" && \
        -f "${path_mito_gff}" && \
        "${asse_type}" != "2" ]]; then
    cat <(grep -v "^#" "${path_gff}") <(grep -v "^#" "${path_mito_gff}") \</pre>
   > "${dir_out}/${name_gff}"
  else
    grep -v "^#" "${path_gff}" > "${dir_out}/${name_gff}"
  fi
done
```

Here we transform the annotation files in gff format to bed files (without header, otherwise impg crashes). The format of the annotations file for the reference genome are slightly different compared to the phenovar data. So we have to process them separately.

```
dirOut <- file.path(dirBase, "anno", "bed")</pre>
unlink(dirOut, recursive = T)
dir.create(dirOut, showWarnings = F)
idRef <- "SGDref"</pre>
hdGff <- c("Chr_id", "Strain_id", "Feat_type", "S_coord",
            "E_coord", "S_val", "Strand_id", "Frame_id", "Attribute_str")
vtClassSrt <- c("gene",
                 "pseudogene")
# vtClassSrt <- c("gene",</pre>
#
                   "pseudogene",
#
                   "intron",
#
                   "five_prime_UTR_intron",
#
                   "noncoding_exon",
#
                   "plus_1_translational_frameshift",
#
                   "blocked_reading_frame",
#
                   "external_transcribed_spacer_region",
#
                   "internal_transcribed_spacer_region",
#
                   "ARS",
#
                   "ARS_consensus_sequence",
#
                   "TY1",
#
                   "TY1/TY2_soloLTR",
#
                   "TY1_truncated",
#
                   "TY2",
#
                   "TY3_soloLTR",
#
                   "TY4 soloLTR",
#
                   "TSU4_soloLTR",
#
                   "TY4 truncated",
                   "TY5",
#
#
                   "TY5 soloLTR",
#
                   "LTR_retrotransposon",
#
                   "long_terminal_repeat",
#
                   "W_region",
#
                   "Z1_region",
#
                   "Z2_region",
#
                   "centromere",
#
                   "centromere_DNA_Element_I",
#
                   "centromere_DNA_Element_II",
#
                   "centromere_DNA_Element_III",
#
                   "matrix_attachment_site",
#
                   "X element",
#
                   "X_element_partial",
#
                   "X_element_combinatorial_repeat",
#
                   "X region",
#
                   "Y_prime_element",
#
                   "Y_region",
#
                   "recombination\_enhancer",
#
                   "silent_mating_type_cassette_array",
#
                   "mating_type_region",
#
                   "tRNA",
#
                   "pseudogenic_transcript",
#
                   "ncRNA",
#
                   "ncRNA_gene",
#
                   "non_transcribed_region",
```

```
"rRNA",
#
                    "rRNA_gene",
#
                    "snRNA",
#
                    "snRNA_gene",
#
                    "snoRNA".
#
                    "snoRNA gene",
#
                   "intein_encoding_region")
### just in case there's some redundancy
vtClassSrt <- unique(vtClassSrt)</pre>
## clmnt -----
### script name
myName <- current_filename()</pre>
cat("[", myName, "] ",
    "Transforming the gff to bed. ",
    "\n", sep = "")
## [] Transforming the gff to bed.
### read strain-haplotypes ids from file
pathIds <- list.files(path = dirBase, pattern = "ids-ps.txt",</pre>
                       full.names = T, recursive = T)
vtStrainHaplo <- as.character(fread(file = pathIds, header = F)[[1]])</pre>
vtRef <- grep(idRef, vtStrainHaplo, value = T)</pre>
### remove the reference
vtStrainHaplo <- grep(idRef, vtStrainHaplo, value = T, invert = T)</pre>
### loop for reference annotations
for (indR in vtRef) {
  ### read the gff
  pathAnnoGff <- list.files(path = dirAnnoGff, pattern = indR,</pre>
                              full.names = T, recursive = T)
  dtGff <- fread(file = pathAnnoGff, sep = "\t", header = F, verbose = F)
  colnames(dtGff) <- hdGff</pre>
  ### id
  strIdPref <- paste0(sub(pattern = "-", replacement = "#", x = indR), "#")
  ### check start and end coordinates: sometimes start = end and impg breaks
  dtGff <- dtGff[S_coord < E_coord]</pre>
  ### filter features
  dtGff <- dtGff[Feat_type %in% vtClassSrt, ]</pre>
  ### make the feature id column
  strFeatId <- sub("^.*Name=([^;]*).*$", "\\1", dtGff$Attribute_str)</pre>
  \textit{### trim after the first ":", e.g. Name=TY3\_soloLTR:chrI:183676-184015:--
  strFeatIdTrm <- sub("^([^:]*).*$", "\\1", strFeatId)</pre>
  \hbox{\it \#\#\# trim trailing redundancy, e.g. \_intron in YAL003W\_intron}
  strFeatIdTrm <- sub("^([^_]*).*$", "\\1", strFeatId)
  ### if strFeatId = dtGff[, Feat_type] set strFeatId = "MN"
  indM <- which(dtGff[, Feat_type] == strFeatIdTrm)</pre>
  if (length(indM) != 0) {
    strFeatIdTrm[indM] <- "MN"</pre>
  ### paste class and feature id (and the strand)
  strName <- pasteO(dtGff[, Feat_type], ":",</pre>
```

```
strFeatIdTrm, "#", dtGff[, Strand_id])
  ### transform the gff into a bed file
  dtBed <- data.table(chrom = pasteO(strIdPref, dtGff[, Chr_id]),</pre>
                       chromStart = dtGff[, S_coord],
                       chromEnd = dtGff[, E_coord],
                       name = strName)
  ### write the bed file
  nameOut <- sub(pattern = "-features.gff$", replacement = ".bed",</pre>
                  x = basename(pathAnnoGff))
  pathOutBed <- file.path(dirOut, nameOut)</pre>
  fwrite(file = pathOutBed, x = dtBed, sep = "\t",
         quote = F, row.names = F, col.names = F)
}
### loop for phenovar annotations
for (indS in vtStrainHaplo) {
  ### dev
  # indS <- vtStrainHaplo[1] ### dev</pre>
  \# indS <- "AKH_1a-0" \#\#\# dev
  ### read the gff
  pathAnnoGff <- list.files(path = dirAnnoGff, pattern = indS,</pre>
                             full.names = T, recursive = T)
  dtGff <- fread(file = pathAnnoGff, sep = "\t", header = F, verbose = F)</pre>
  colnames(dtGff) <- hdGff</pre>
  ### id
  strIdPref <- paste0(sub(pattern = "-", replacement = "#", x = indS), "#")
  ### check start and end coordinates: sometimes start = end and impg breaks
  dtGff <- dtGff[S_coord < E_coord]</pre>
  ### filter features
  dtGff <- dtGff[Feat_type %in% vtClassSrt, ]</pre>
  ### make the feature id column
  strFeatId <- sub("^.*Name=([^;]*).*$", "\\1", dtGff$Attribute_str)</pre>
  ### trim after the first ":", e.g. Name=TY3_soloLTR:chrI:183676-184015:-
  strFeatIdTrm <- sub("^([^:]*).*$", "\\1", strFeatId)
  ### trim "tRNA_" from feature id
  strFeatIdTrm <- sub(pattern = "tRNA_", replacement = "", x = strFeatIdTrm)
  ### if strFeatId = dtGff[, Feat_type] set strFeatId = "MN"
  indM <- which(dtGff[, Feat_type] == strFeatIdTrm)</pre>
  if (length(indM) != 0) {
   strFeatIdTrm[indM] <- "MN"</pre>
  }
  ### paste class and feature id (and the strand)
  strName <- pasteO(dtGff[, Feat_type], ":",</pre>
                     strFeatIdTrm, "#", dtGff[, Strand_id])
  ### transform the gff into a bed file
  dtBed <- data.table(chrom = paste0(strIdPref, dtGff[, Chr_id]),</pre>
                       chromStart = dtGff[, S_coord],
                       chromEnd = dtGff[, E_coord],
                       name = strName)
  ### write the bed file
  nameOut <- sub(pattern = "-features.gff$", replacement = ".bed",</pre>
                  x = basename(pathAnnoGff))
```

We invoke impg through a system command to compute the features pangenome. Then we remove the redundancy generated by impg by checking the overlap of the regions computed by impg. To do this, every annotation interval is used to generate a block of homology. Each block of homology is decomposed in sub-blocks which are defined by a feature class. The reduced (i.e. nonredundant) sub-block is then transformed (i.e. formatted) and appended to output.

```
## header -----
options(scipen = 999)
options(stringsAsFactors = F)
rm(list = ls())
library(data.table)
library(this.path)
library(scriptName)
library(GenomicRanges)
## Caricamento del pacchetto richiesto: stats4
## Caricamento del pacchetto richiesto: BiocGenerics
##
## Caricamento pacchetto: 'BiocGenerics'
## I seguenti oggetti sono mascherati da 'package:stats':
##
##
       IQR, mad, sd, var, xtabs
## I seguenti oggetti sono mascherati da 'package:base':
##
##
       Filter, Find, Map, Position, Reduce, anyDuplicated, aperm, append,
##
       as.data.frame, basename, cbind, colnames, dirname, do.call,
##
       duplicated, eval, evalq, get, grep, grepl, intersect, is.unsorted,
##
       lapply, mapply, match, mget, order, paste, pmax, pmax.int, pmin,
##
       pmin.int, rank, rbind, rownames, sapply, setdiff, sort, table,
##
       tapply, union, unique, unsplit, which.max, which.min
## Caricamento del pacchetto richiesto: S4Vectors
##
## Caricamento pacchetto: 'S4Vectors'
## I seguenti oggetti sono mascherati da 'package:data.table':
##
##
       first, second
## Il seguente oggetto è mascherato da 'package:utils':
##
##
       findMatches
## I seguenti oggetti sono mascherati da 'package:base':
##
       I, expand.grid, unname
##
## Caricamento del pacchetto richiesto: IRanges
```

```
##
## Caricamento pacchetto: 'IRanges'
## Il seguente oggetto è mascherato da 'package:data.table':
##
      shift
## Caricamento del pacchetto richiesto: GenomeInfoDb
## Warning: il pacchetto 'GenomeInfoDb' è stato creato con R versione 4.3.3
library(tictoc)
##
## Caricamento pacchetto: 'tictoc'
## Il seguente oggetto è mascherato da 'package:GenomicRanges':
##
##
      shift
## Il seguente oggetto è mascherato da 'package: IRanges':
##
##
      shift
## Il seguente oggetto è mascherato da 'package:data.table':
##
      shift
## function(s) -----
SplitSubCol <- function(x, n, s) {</pre>
 y <- sapply(strsplit(x, split = s, fixed = T), "[[", n)
 return(y)
}
### It splits a character column of a data-table and collects one sub-column.
###
### Arguments:
### (1) x: a character column of a data-table
### (2) n: the number of sub-column to print in output
### (3) s: the string to apply the split
###
### Returns:
### (1) y: a character vector
## settings ------
### fixed settings
dirBase <- dirname(this.dir())</pre>
### dev dirBase <- "/Users/Lorenzo/dev/panda"</pre>
dirAnnoBed <- file.path(dirBase, "anno", "bed")</pre>
dirOut <- file.path(dirBase, "png")</pre>
unlink(dirOut, recursive = T)
dir.create(dirOut, showWarnings = F)
pathGen <- file.path(dirOut, "generators.txt")</pre>
cat("class:feature", "\n", file = pathGen)
idRef <- "SGDref"</pre>
hdImpg <- c("Query_id", "Query_start", "Query_end",
          "Target_id", "Target_start", "Target_end",
```

```
"Target_clsfeat", "N_score", "Query_alndir", "Target_alndir")
# vtClassSrt <- c("gene",</pre>
                   "pseudogene",
#
                   "intron",
#
                   "five_prime_UTR_intron",
#
                   "noncoding_exon",
#
                   "plus 1 translational frameshift",
#
                   "blocked_reading_frame",
#
                   "external_transcribed_spacer_region",
#
                   "internal_transcribed_spacer_region",
#
                   "ARS",
#
                   "ARS_consensus_sequence",
#
                   "TY1",
#
                   "TY1/TY2_soloLTR",
#
                   "TY1_truncated",
#
                   "TY2",
#
                   "TY3_soloLTR",
#
                   "TY4_soloLTR",
#
                   "TSU4_soloLTR",
#
                   "TY4_truncated",
#
                   "TY5",
#
                   "TY5 soloLTR",
#
                   "LTR_retrotransposon",
#
                   "long terminal repeat",
#
                   "W region",
#
                   "Z1 region",
#
                   "Z2_region",
#
                   "centromere",
#
                   "centromere_DNA_Element_I",
#
                   "centromere_DNA_Element_II",
#
                   "centromere_DNA_Element_III",
#
                   "matrix_attachment_site",
#
                   "X_element",
#
                   "X_element_partial",
#
                   "X_element_combinatorial_repeat",
#
                   "X_region",
#
                   "Y_prime_element",
#
                   "Y_region",
#
                   "recombination_enhancer",
#
                   "silent_mating_type_cassette_array",
#
                   "mating_type_region",
#
                   "tRNA",
#
                   "pseudogenic_transcript",
#
                   "ncRNA",
#
                   "ncRNA_gene",
#
                   "non_transcribed_region",
#
                   "rRNA",
#
                   "rRNA_ gene",
#
                   "snRNA",
                   "snRNA_qene",
#
#
                   "snoRNA",
#
                   "snoRNA_gene",
```

```
"intein_encoding_region")
### the path to the paf file
dirAlnPaf <- file.path(dirBase, "aln")</pre>
pathAlnPaf <- list.files(path = dirAlnPaf, pattern = "paf$",</pre>
                          recursive = T, full.names = T)
if (length(pathAlnPaf) > 1) {
 stop("Multiple paf files found.")
} else if (length(pathAlnPaf) == 0) {
  stop("No paf file found.")
}
## clmnt -----
### script name
myName <- current_filename()</pre>
cat("[", myName, "] ",
    "Making the pangenome ",
    "\n", sep = "")
## [] Making the pangenome
### read strain-haplotypes ids from file (the order is maintained in the output)
pathIds <- file.path(dirBase, "ids", "ids-ps.txt")</pre>
vtStrainHaplo <- as.character(fread(file = pathIds, header = F)[[1]])</pre>
vtStrainHaploHash <- gsub(pattern = "-", replacement = "#", x = vtStrainHaplo)
### initialise the output features table
nHaplos <- length(vtStrainHaploHash)</pre>
dtPanFeats <- data.table(matrix(character(length = 0), ncol = nHaplos))</pre>
setnames(dtPanFeats, vtStrainHaploHash)
### dev
# vtStrainHaploHash <- c("SGDref#0", "AFI#0", "S288C#0",
                          "XXX-h3", "stoca-zo", "DBVPG6765#0")
### dev indS <- vtStrainHaplo[1]
indL <- 1
lsImpg <- list()</pre>
### for each strain-haplotypes (hyphen-separated)
for (indS in vtStrainHaplo) {
  ### pick the strain-haplotypes bed file (query)
  pathAnnoBed <- list.files(path = dirAnnoBed, pattern = indS, full.names = T)</pre>
  ### string for bash
  strBashImpg <- paste0("impg -I -p ", pathAlnPaf, " -b ", pathAnnoBed)
  ### run "$ impg -p file.paf -b file.bed" which is
  ### (50x faster than going line-by-line)
  ### with direct load of the output
  strOut <- system(strBashImpg, intern = T)</pre>
  ### make a single-file data-table
  lsImpgOne <- strsplit(strOut, split = "\t")</pre>
  dtImpgOne <- rbindlist(lapply(lsImpgOne,</pre>
                                 function(x) as.data.table(as.list(x))))
  ### append the single-file data-table to a list
  lsImpg[[indL]] <- dtImpgOne</pre>
```

```
indL <- indL + 1
}
### get the redundant data-table
dtImpgAll <- rbindlist(lsImpg)</pre>
colnames(dtImpgAll) <- hdImpg</pre>
### add supplementary columns
dtImpgAll[, c("Query_start", "Query_end") := lapply(.SD, as.numeric),
          .SDcols = c("Query_start", "Query_end")]
dtImpgAll[, Target_cls := SplitSubCol(Target_clsfeat, 1, ":")]
dtImpgAll[, c("Target_start", "Target_end") := lapply(.SD, as.numeric),
          .SDcols = c("Target_start", "Target_end")]
dtImpgAll[, Target_len := Target_end - Target_start]
### size filters
dtImpgAllSzFlt <- dtImpgAll[Target_len > 5 & Query_end - Query_start > 5]
### sorting ascending or descending is not equivalent since the
### overlap is done in two rounds and
### is strand-aware (if two features have overlapping
### coordinates but one is in the + strand and the other is in
### the - strand they do not overlap), thus
### we sort in increasing (ascending) order to avoid spurious
### overlaps that may occur when multiple classes are analysed
### (a spurious overlap will produce a big block that cannot be easily
### decomposed, except for the different classes);
### the YFL066C locus (where we have the Y' region TEL06L) is a good
### example of a spurious overlap
setorder(dtImpgAllSzFlt, Target_len)
### transfer the strand data from Target_clsfeat to Query_id and Target_id
strStrand <- SplitSubCol(dtImpgAllSzFlt[, Target_clsfeat], 2, "#")</pre>
dtImpgAllSzFlt[, Query_id := paste0(Query_id, "#", strStrand)]
dtImpgAllSzFlt[, Target_id := paste0(Target_id, "#", strStrand)]
nBlocks <- 1
vtUnq <- unique(dtImpgAllSzFlt[, Target_clsfeat])</pre>
### dev
# indTarClsFeat <- "Y_prime_element:TEL06L#-"</pre>
# indTarClsFeat <- "gene:YAR014C#-"</pre>
# indTarClsFeat <- vtUng[1]</pre>
# indTarClsFeat <- "gene:YBL037W#+"</pre>
# indTarClsFeat <- "X_element:TELO1L#-"</pre>
# indTarClsFeat <- "gene:YBR140C#-"</pre>
# indTarClsFeat <- "ARS:ARS102#+"</pre>
# indTarClsFeat <- "gene:YCR012W#+"</pre>
### quello con molti haplo-id doppioni, indSb <- "TY1/TY2_soloLTR"
# indTarClsFeat <- "TY1:MN#-"</pre>
# indTarClsFeat <- "qene:YGR296W#+"</pre>
# which(vtUng == "gene:YGR296W#+")
# indTarClsFeat <- "gene:S288C_G0022800#+"</pre>
# which(vtUnq == "gene:S288C_G0022800#+")
```

```
# which(vtUnq == "ARS+ARS102#+")
# which(vtUng == "X_element+TELO1L#+")
# indTarClsFeat <- unique(dtImpqAllSzFlt[, Target_clsfeat])[3]</pre>
# indTarClsFeat <- vtUng[1]</pre>
for (indTarClsFeat in vtUng) {
 ## block calculation -----
 ### dev print(nBlocks)
 ### get the target class-features that will generate the block
 dtTarClsFeat <- dtImpgAllSzFlt[indTarClsFeat, on = "Target_clsfeat"]
 ### check if the corresponding interval is still in dtImpgAllSzFlt
 ### (if it was ovelapping with a former indTarClsFeat
 ### it has already been removed, but dtTarClsFeat will still have a line
 ### e.g. the following):
 ### Query_id Query_start Query_end Target_id Target_start Target_end
 ###
      <char>
                 <num> <num> <char> <num>
 ###
         <NA>
                     NA
                               NA
                                        <NA>
                                                     NA
                                                                 NA
 ### Target_clsfeat N_score Query_alndir Target_alndir Target_cls Target_len
                                         <char> <char>
                             <char>
 ###
             <char> <char>
                                                <NA>
 ###
        buciodeculo <NA>
                                   <NA>
                                                           < NA >
 if (any(is.na(dtTarClsFeat[, Query_id]))) {
   next
 }
 ### first round of overlap (on query coordinates)
 setkey(dtTarClsFeat, Query_id, Query_start, Query_end)
 dtIndOverOne <- foverlaps(dtImpgAllSzFlt, dtTarClsFeat,</pre>
                           nomatch = NULL, type = "any", which = T)
 dtBlock <- dtImpgAllSzFlt[dtIndOverOne[, xid], ]</pre>
 print(format(object.size(dtBlock), units = "auto"))
 ### second round of overlap (on target coordinates)
 setkey(dtBlock, Target_id, Target_start, Target_end)
 dtIndOverTwo <- foverlaps(dtImpgAllSzFlt, dtBlock,</pre>
                           nomatch = NULL, type = "any", which = T)
 dtBlock <- rbindlist(list(dtBlock, dtImpgAllSzFlt[dtIndOverTwo[, xid], ]))</pre>
 print(format(object.size(dtBlock), units = "auto"))
 ### block classes
 allClassesInBlock <- unique(dtBlock[, Target_cls])</pre>
 ### dev
 # cat(indTarClsFeat, "\t",
       nBlocks, "\t", length(allClassesInBlock), "\n",
       file = "~/Desktop/check-nsb.txt", append = T)
 ## sub-blocks reduction (class-by-class, strand-aware) ------
 ### dev
  # indSb <- "gene"
```

```
# indSb <- "TY1/TY2_soloLTR"</pre>
# indSb <- "Y_prime_element"</pre>
for (indSb in allClassesInBlock) {
  ### reduce (same as bedtools merge) the rows for each class
  ### on the basis of the query coordinates
  grSblock <- GRanges(seqnames = dtBlock[Target_cls == indSb, Query_id],</pre>
                      ranges = IRanges(start = dtBlock[Target_cls == indSb,
                                                       Query start],
                                       end = dtBlock[Target_cls == indSb,
                                                      Query_end]),
                      mcols = dtBlock[Target_cls == indSb, Target_clsfeat])
  grSblockRed <- reduce(grSblock)</pre>
  ### vector with the features names and class name
  ### cleaned of the "#+" or "#-" at the end of the string
  ### e.g. "gene:S288C_G0022800" and "gene:YGR296W"
  vtClsFeat <- sub(pattern = "#[\\+\\-]$",</pre>
                   replacement = "",
                   x = unique(grSblock$mcols))
  ### features string
  strFeats <- paste(gsub(pattern = paste0(indSb, ":"), replacement = "",</pre>
                         x = vtClsFeat),
                    collapse = ",")
  dtSblockRed <- data.table(as.character(grSblockRed@seqnames),</pre>
                            as.character(grSblockRed@ranges))
  ## transformation of the reduced sub-block -----
  ### e.g. this dtSblockRed:
  ###
                  <char>
                                  <char>
  ### SGDref#0#chrVII#+ 1084864-1090591
  ### S288C#0#chrVII#+ 1085116-1090843
  ###
  ### becomes a two-column data-table with columns:
  ###
                      SGDref#0
                                                    S288C#0
                        <char>
  ### chrVII:1084864-1090591#+ chrVII:1085116-1090843#+
  ### dev
  # vtHaplo <- sub("#chr.*", "", dtSblockRed[, V1])
  # tbHaplo <- table(vtHaplo)</pre>
  # if (length(which(tbHaplo > 1)) != 0) {
  # stop("found it!")
  # }
  ### transposition and collapsing explained with
  ### another example, this dtSblockRed:
  ###
                           V1
                                        V2
                       <char>
  ###
                                     <char>
  ###
       DBVPG6765#0#chrIV#- 960288-966619
             AFI#0#chrXIII#- 349539-349680
```

```
AFI#0#chrXIII#- 349713-349989
      DBVPG6765#0#chrXIII#- 354903-355179
  ###
  ###
                AFI#0#chrIV#- 960661-961002
  ###
            S288C#0#chrXIII#- 384508-384784
  ###
           SGDref#0#chrXIII#- 378732-379008
  ###
  ### must produce this column:
  ### DBVPG6765#0
  ### chrIV:960288-966619#-,chrXIII:354903-355179#-
  ### formatting dtSblockRed
  dtSblockRed[, Haplo_id := sub("#chr.*", "", dtSblockRed[, V1])]
  vtCoord <- paste(SplitSubCol(x = dtSblockRed[, V1], n = 3, s = "#"),</pre>
                   dtSblockRed[, V2],
                   sep = ":")
 dtSblockRed[, Info_str := paste(vtCoord,
                                    SplitSubCol(dtSblockRed[, V1], 4, "#"),
                                    sep = "#")]
  ### TODO: retrieve and store the sequences with samtools
  ### collapsing with ";" all the Haplo_id elements of a Haplo_id
  dtSblockRedCo <- dtSblockRed[, paste(Info_str, collapse = ";"),</pre>
                               by = Haplo_id]
  ### transpose, producing a data-table
  ### we keep it as data-table even if
  ### qui
 dtTra <- transpose(dtSblockRedCo)</pre>
  ### set column names and format
  setnames(dtTra, as.character(dtTra[1, ]))
 dtTra <- dtTra[-1]</pre>
  ### add class and features columns
 dtTra[, ':='(class = rep(indSb, .N), features = rep(strFeats, .N))]
  ### add the missing columns: not needed
  ### since rbindlist makes it by default using fill = TRUE
  # newCols <- setdiff(vtStrainHaploHash, colnames(dtTra))</pre>
  # dtTra[, (newCols) := lapply(newCols, function(x) NA)]
  # setcolorder(dtTra, vtStrainHaploHash)
  ### append with rbindlist matching (default operation) column names
 dtPanFeats <- rbindlist(list(dtPanFeats, dtTra), fill = T)</pre>
  ### append the generator feature to the output
  cat(rep(indTarClsFeat, nrow(dtTra)), sep = "\n",
      append = T, file = pathGen)
  ### dev print(nrow(dtTra))
}
### delete dtIndOverOne and dtIndOverTwo rows in dtImpgAllSzFlt,
### the (any(is.na(dtTarClsFeat[, Query_id]))) will avoid
### the loop to break
```

```
indOut <- unique(c(dtIndOverOne[, xid], dtIndOverTwo[, xid]))</pre>
  dtImpgAllSzFlt <- dtImpgAllSzFlt[-indOut]</pre>
  nBlocks <- nBlocks + 1
  ### dev if (nBlocks == 20) stop("We did 20 iterations!")
}
## [1] "6.9 Kb"
## [1] "18.7 Kb"
## [1] "4.8 Kb"
## [1] "6.4 Kb"
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- ## [1] "5.6 Kb"
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### format and write dtPanFeats
leftCols <- c("class", "features")</pre>
colOrder <- c(leftCols, setdiff(names(dtPanFeats), leftCols))</pre>
setcolorder(dtPanFeats, colOrder)
pathOutPanFeat <- file.path(dirOut, "pan-features.txt")</pre>
# fwrite(dtPanFeats, file = pathOutPanFeat, append = F, quote = F, sep = "\t",
         col.names = T, na = "MA", nThread = 8, buffMB = 1024)
\#>Error in fwrite(dtPanFeats, file = pathOutPanFeat, append = F, quote = F, :
#> Bad address: '/home/ltattini/prog/graphs/panda/run-11/png/pan-features.txt'
write.table(x = dtPanFeats, file = pathOutPanFeat, append = F, quote = F,
            sep = "\t", col.names = T, row.names = F, na = "MA")
```

```
save(dtPanFeats, file = file.path(dirOut, "pan-features.RData"))
### TODO: write dtPanSeqs
```