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Outline



Background



Methods



Results & discussion



Conclusions & future studies

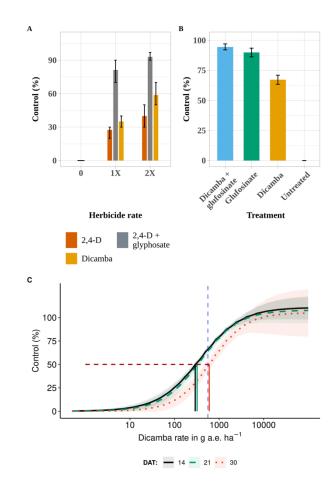


Background

Waterhemp (Amaranthus tuberculatus (Moq.) Sauer) is one of the most troublesome agronomic weeds in the midwestern US.

CHR population:

- Identified in 2012 in Champaign County, IL
- Resistant to 6 sites of action (WSSA groups 2, 4, 5, 14, 15 and 27)
- No history of dicamba or 2,4-D application in CHR field
- Ineffective control of CHR was observed in the field after dicamba application
- Preliminary field experiments show a reduction in the effectiveness of dicamba -> dicamba resistance evolution





Objectives

1. Quantify the dicamba resistance level and investigate its inheritance in CHR

2. Identify putative candidate genes involved with dicamba resistance via RNA-seq



Methods – Experiments















Population development

- Resistant (R) plants selected from original CHR field (after 560 g ae ha⁻¹ of dicamba)
- RxR crosses for R parental line
- Select susceptible parent (S)
- Reciprocal crosses for F₁ generation
- Pseudo- F₂ and Backcross generations

Dose response

- Experimental design: RCBD
 + 6 reps/rate + 9 rates + 2
 experiment replications
- Dicamba rates: 0, 1.18, 3.92, 11.8, 39.2, 118, 392, 1,180 and 2,350 g ae ha⁻¹
- Populations used: Parental (R and S) and F₁ lines
- Analysis: DRC and Dominance degree calculation in R
- Define delimiting rate for segregation analysis

Segregation analysis

- Quantify dicamba resistance inheritance pattern
- Populations: All populations
- Dicamba damage: Using a delimiting rate damage quantification via machine learning
- Analysis: chi-square (x²) and broad sense heritability in R
- Hypothesis: Dicamba resistance is caused by a single gene

RNA-seq

- RNA extraction (T = 0h)
- Dicamba 560 g ae ha¹
- Phenotyping
- Apply machine learning model to selected plants
- Sequencing
- Giacomini et al. 2019

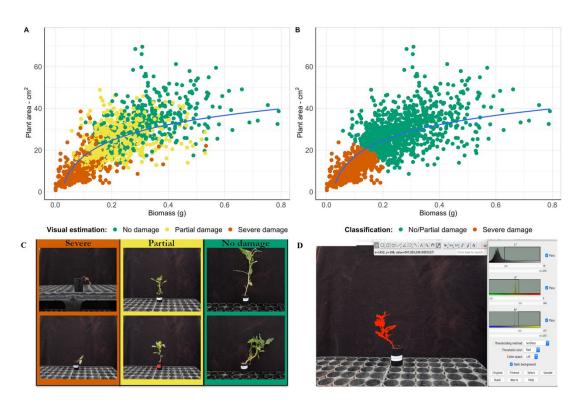
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Methods – Dicamba damage estimation

- Image analysis: ImageJ + Python
- Analysis based on:
 - Plant area
 - Biomass
 - Visual estimation
- Unsupervised machine learning: Bayesian random forest model to classify samples
- 2,000 samples used as training dataset for the model
- 85% accuracy / 88% specificity





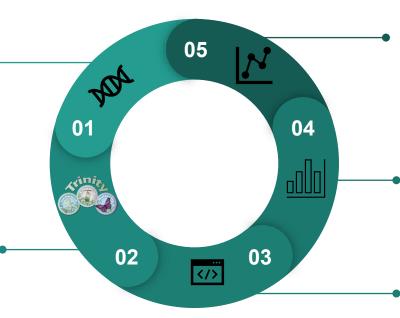
Methods - RNA-seq

Sequencing & data processing

- Illumina NovaSeq 6000 1x100bp
- 16 samples (8 R and 8 S)
- Adapters trimmed and rRNA removed

Transcriptome assembly

- Reads mapped to waterhemp genome using STAR
- Genome guided transcriptome assembly using Trinity



RT-qPCR confirmation

- RNA-seq candidate genes tested via qPCR
- · Two housekeeping genes
- 2^{ΔΔct2} method for relative expression estimation
- 36 F₂ individuals tested (Including individuals used for RNA-seq)

Differential expression analysis

Analysis conducted using the Sleuth and EdgeR for transcript and gene level, respectively.

Expression count

 Expression count done via pseudoalignment using the software Kallisto.



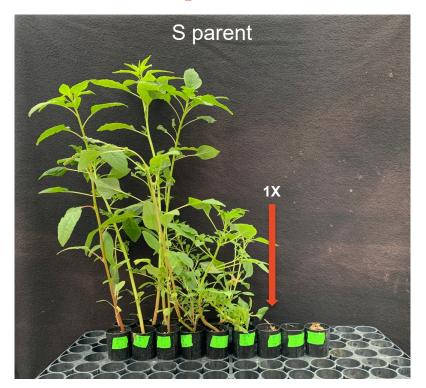
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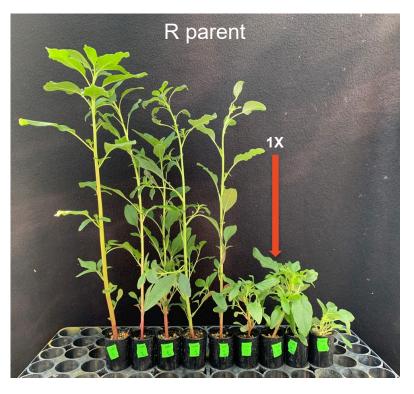
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Dose-response





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Dose-response

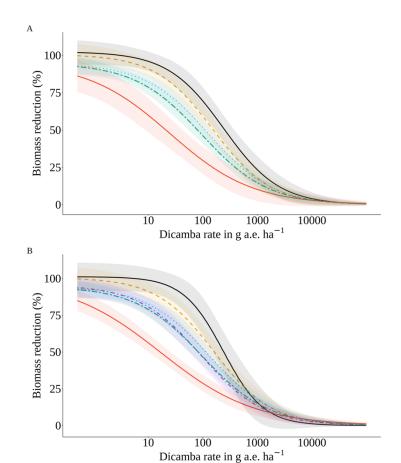
- 3 parameter Log-logistic model
- Two experiments
- R/S = 5 10
- Degree of dominance = 0.25
- Incomplete dominant trait

$$Degree\ of\ dominance = \frac{(2W_3 - W_2 - W_1)}{(W_2 - W_1)}$$

$$W_x = ED_{50}$$

$$1,2,3 = S,\ R,\ F_1$$





Populations: - CHR **-** F1-1 F1-2 F1-3 F1-4 **-** WUS

Conclusions

Segregation analysis

		Observed		Expected		Chi-square	
Population	# plants	No/partial damage	Severe damage	No/partial damage	Severe damage	x ²	p-value
F ₂	431	247	183	282	149	22.74	< 0.001
BC-1	147	93	54	72	75	10.34	<0.001
BC-2	110	68	42	54	56	6.14	0.01
F₁-R♀xS♂	165	140	25	Reject 3:1 ratio (single gene)			

Reject 3:1 ratio (single gene)

• Moderate heritability

199

148

131

• Dicamba resistance: Multi-genic trait

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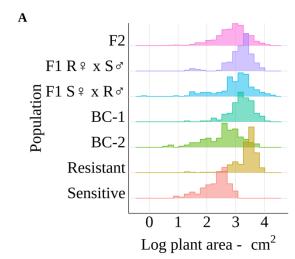
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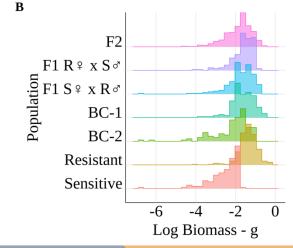


F₁- S♀xR♂

R parent

S parent





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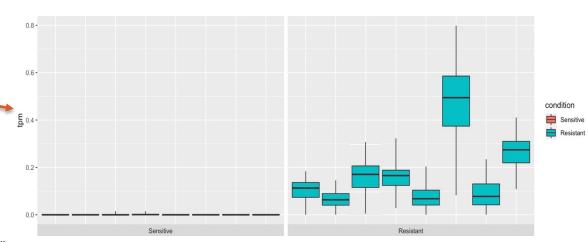
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RNA-seq – Transcript level

- 45 DE transcripts
- 1 major candidate:
 - Auxin efflux gene (TIR3)

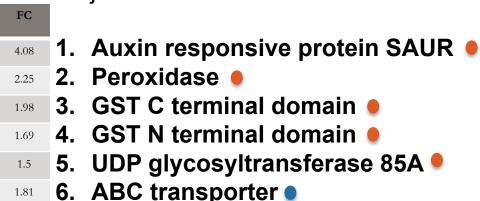
- Aggregation of transcripts:
- 2 major candidates:
 - 1. Transcription activator for cytokine response (ORR)
 - 2. Auxin induced protein (IAA)

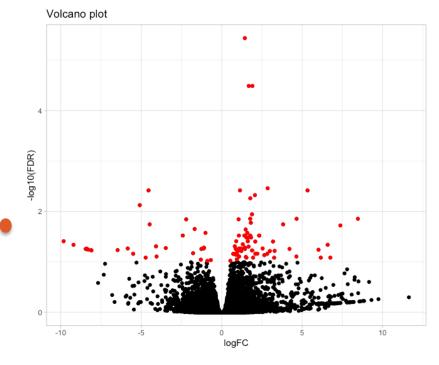




RNA-seq – Gene level

- 103 DE genes
- 7 major candidates:





Upregulated

Down regulated



7. MLP – ABA regulator •

Candidate genes dicamba resistance

Candidates:

- 1. SAUR First Auxin responsive protein
- 2. **Peroxidase** Auxin Catabolism and ROS detoxification
- **3. MLP** ABA regulator when down regulated
- 4. TIR3 Auxin efflux gene
- **5. IAA induced protein** Auxin response transcriptional factors that repress early auxin responses
- **6. UDP-glycosyl transferase** Glycosylation of hormones and exogenous compounds
- 7. Glutathione-S-transferase conjugation of exogenous compound and hormones. Can also work as a peroxidase.
- **8. ABC** transporter Transportation of secondary metabolites



RT qPCR – Gene confirmation

F₂ plants

- 2 housekeeping genes (GAPDH and EF1 α)
- $36 \, \mathrm{F}_2 \, \mathrm{plants}$
- $2^{\Delta\Delta ct2}$ method
- Pairwise t-test

Gene	P-value		
PEROX	< 0.001		
PIN3	0.0032		
GST-N	0.005		
UDP	<0.001		
SAUR	NS		
GST-C	NS		
ABC	NS		
MLP	NS		
IAA	NS		
ORR	NS		











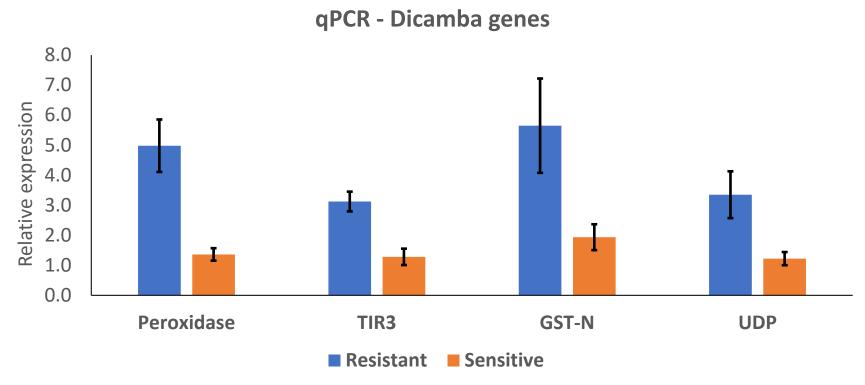
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R & D

Conclusions

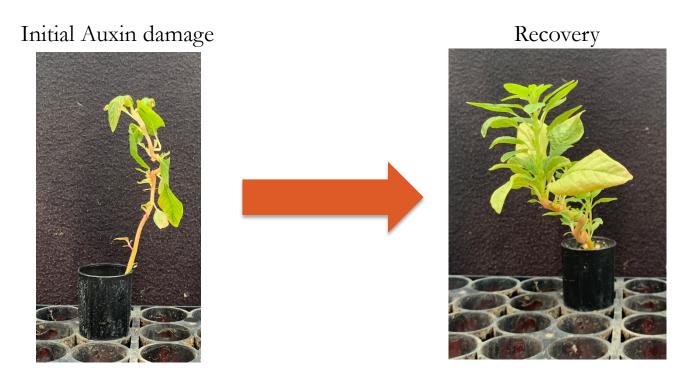
RT qPCR – Gene confirmation





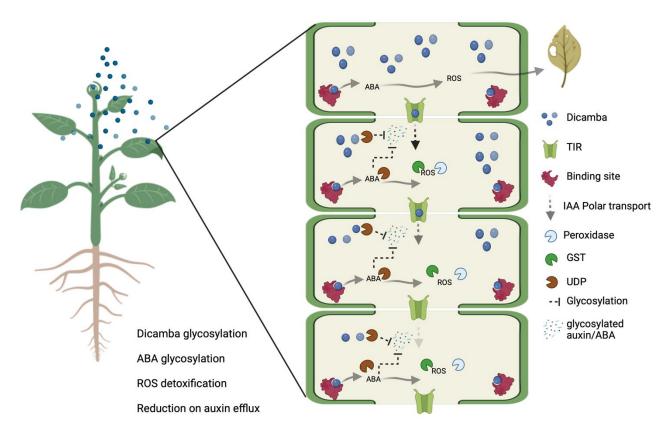
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Hypothesis: Mechanism of resistance





Hypothesis: mechanism of resistance





Background Methods R & D Conclusions Future studies

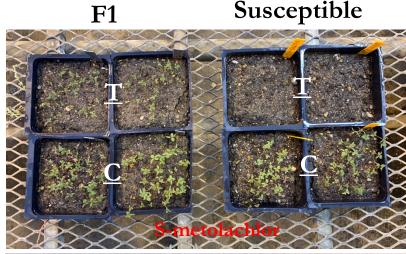
Conclusions

- Dicamba resistance:
 - Incompletely dominant trait
 - Moderate heritability
 - Multi-genic trait
- Multiple candidate genes identified including Glutathione-S-transferases and UDP-glycosyl transferase
- Hypothesis of mechanism of resistance:
 - ROS detoxification via GST and peroxidase
 - Glycosylation and conjugation of ABA and dicamba
 - Changes in dicamba efflux



Future studies

- Physiology studies
- Expression variation after dicamba treatment
- Functional validation of genes
- 3-way resistance QTL mapping:
 - 2,4-D
 - S-metolachlor
 - Dicamba







R&D

Acknowledgements



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- Damilola Alex Rayemo
- Undergrads

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- Dr. Brent Murphy
- Dr. Darci Giacomini
- Jake Montgomery

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