

Epidemiological exercises in R

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Prerequisites

To run these exercises, you will need to install the latest version of R (<https://cloud.r-project.org/>) and RStudio (<https://www.rstudio.com/products/rstudio/download/>) on your computer. Please click on the above mentioned links to go to the download pages of R and RStudio.

These exercises are a work in progress, and are an attempt to translate SAS code written by Tim Todd into R.

Chapter 1

Exercise 1

1.1 Load packages

Here is the R code to download the required packages for this exercise.

```
# install package manager 'pacman'  
if (!require(pacman)){  
  install.packages('pacman')  
}
```

Loading required package: pacman

```
# load packages needed for this exercise  
pacman::p_load(tidyverse, # for data manipulation  
               psych, # for `describe()` function  
               ggpubr, # for graphing  
               rlang, # for tidy evaluation  
               ggfortify)
```

1.2 Data

This is equivalent to the data step in SAS. Here, the data is imported from a file `ex1.csv` using the function `read_csv`. This function will download the data file directly from here.

```
# Import data
a <- read_csv("data/ex1.csv") %>%
  # create a new variable `DIFF`
  mutate(DIFF = ESTIMATE - ACTUAL)
```

```
## Parsed with column specification:
## cols(
##   TEST = col_character(),
##   OBS = col_double(),
##   ESTIMATE = col_double(),
##   ACTUAL = col_double()
## )
```

```
# print the data
a
```

```
## # A tibble: 60 x 5
##   TEST      OBS ESTIMATE ACTUAL  DIFF
##   <chr>   <dbl>   <dbl>  <dbl> <dbl>
## 1 BASELINE     1       2      7    -5
## 2 BASELINE     2      25     39   -14
## 3 BASELINE     3      35     49   -14
## 4 BASELINE     4      20     34   -14
## 5 BASELINE     5       5      8    -3
## 6 BASELINE     6      50     57    -7
## 7 BASELINE     7       5     14    -9
## 8 BASELINE     8      50     57    -7
## 9 BASELINE     9      60     73   -13
## 10 BASELINE    10       2      9    -7
## # ... with 50 more rows
```

1.3 Descriptive statistics

```
# use the describe() function to get an idea about different variables
sm_a <- a %>%
  group_by(TEST) %>%
  summarise_at(vars(DIFF),
    list(~mean(., na.rm = TRUE),
         ~mode(.),
         ~median(., na.rm = TRUE),
         ~sd(., na.rm = TRUE),
         ~var(., na.rm = TRUE),
```



```

~IQR(., na.rm = TRUE),
~min(., na.rm = TRUE),
~max(., na.rm = TRUE),
~n(),
~skew(.),
~kurtosi(.)
)) %>%

mutate(CV = (sd/mean)*100,
       std.err = sd/sqrt(n)) %>%

ungroup()

sm_a

```

```

## # A tibble: 3 x 14
##   TEST  mean mode median   sd  var  IQR  min  max    n  skew
##   <chr> <dbl> <chr>  <dbl> <dbl> <dbl> <dbl> <dbl> <dbl> <int> <dbl>
## 1 BASE~ -9.75 nume~   -9.5  4.17  17.4  6.5   -17   -3    20 -0.0599
## 2 POST1 -2.95 nume~    -2   4.43  19.6  7     -14    3    20 -0.804
## 3 POST2 -2.75 nume~   -2.5  8.82  77.8  9.75  -18   12    20  0.0162
## # ... with 3 more variables: kurtosi <dbl>, CV <dbl>, std.err <dbl>

```

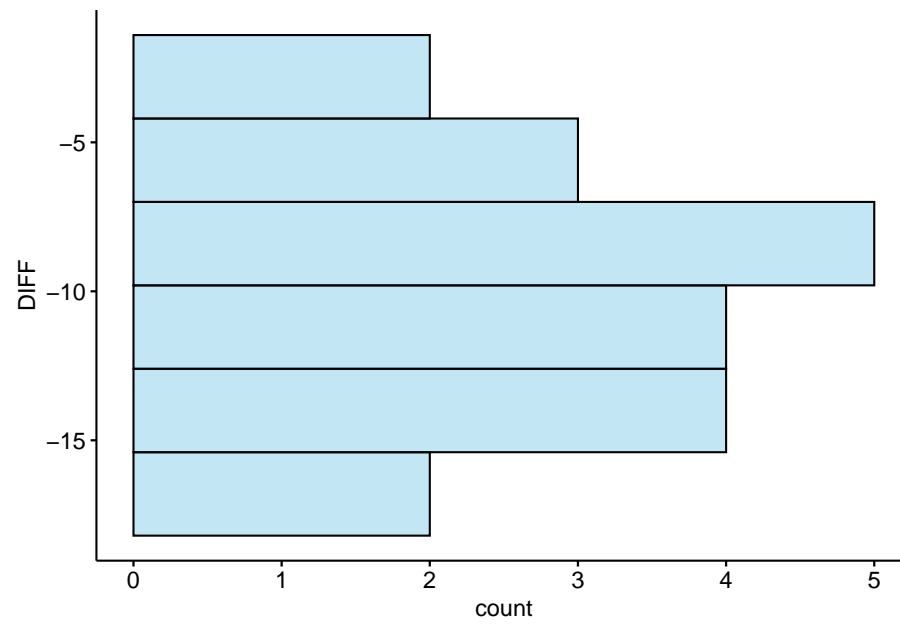
1.4 Create histogram, boxplot, and qqplot

```

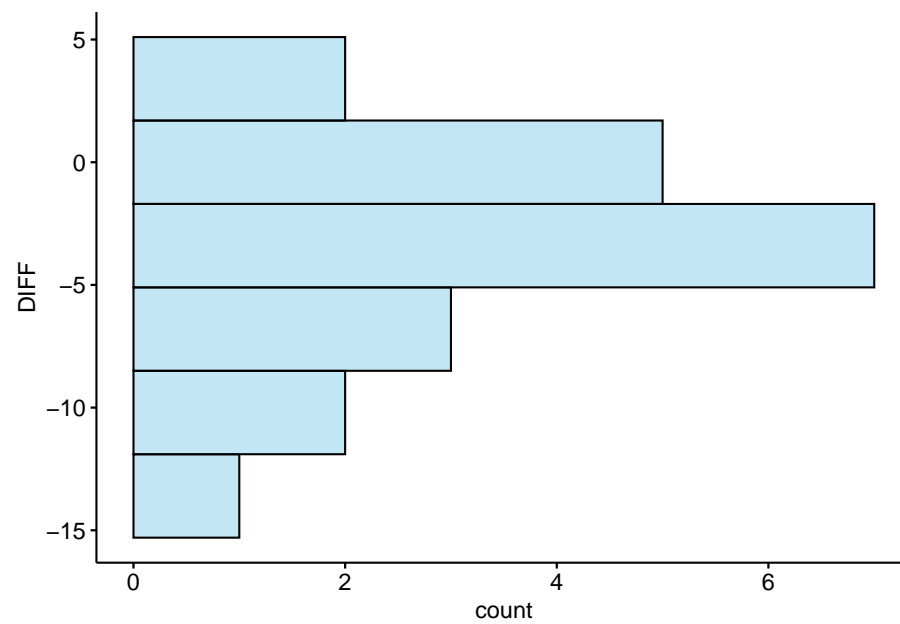
# histograms
create_hist <- function(test_id){
  a %>%
    filter(TEST == {{test_id}}) %>%
    gghistogram("DIFF", bins = 6, fill = "skyblue") +
    coord_flip()
}

create_hist(unique(a$TEST)[1]) # BASELINE

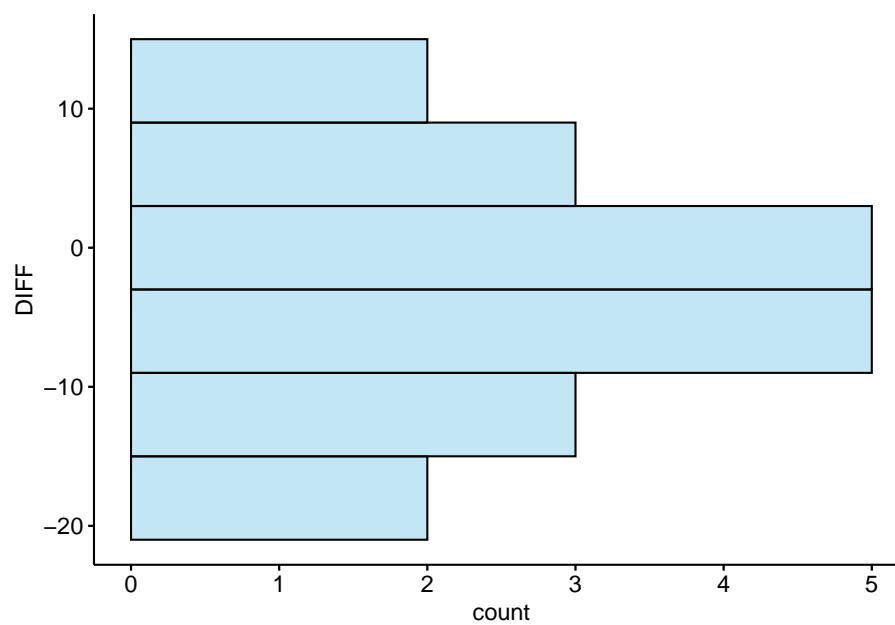
```



```
create_hist(unique(a$TEST)[2]) # POST1
```

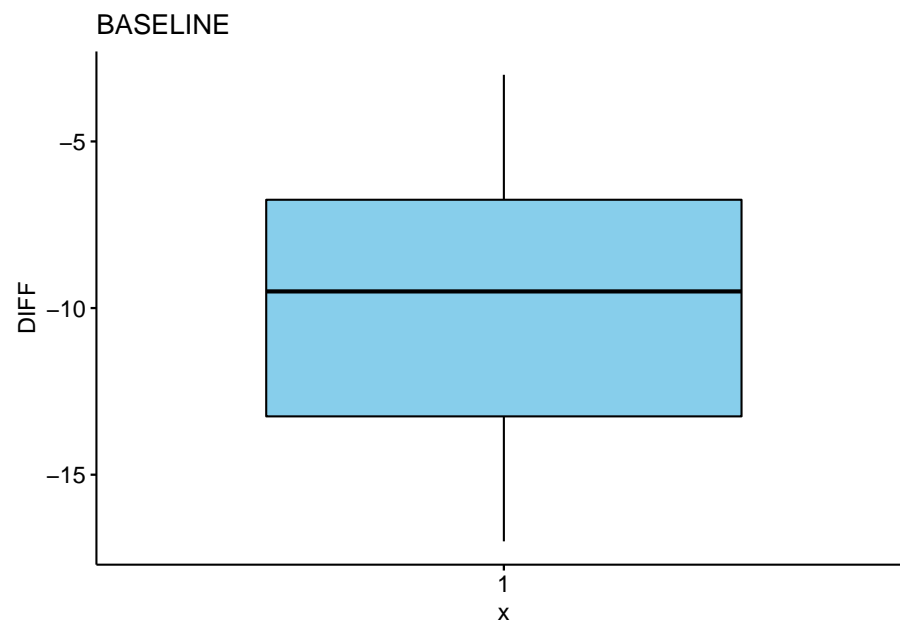


```
create_hist(unique(a$TEST)[3]) # POST2
```

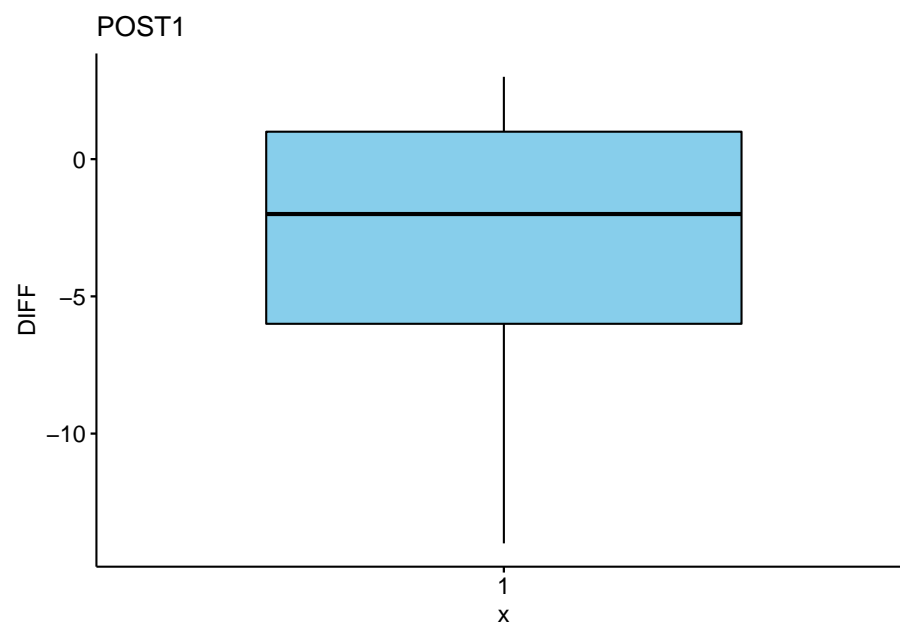


```
# boxplots
create_box <- function(test_id){
  a %>%
    filter(TEST == {{test_id}}) %>%
    ggboxplot(y = "DIFF", fill = "skyblue",
              main = {{test_id}})
}

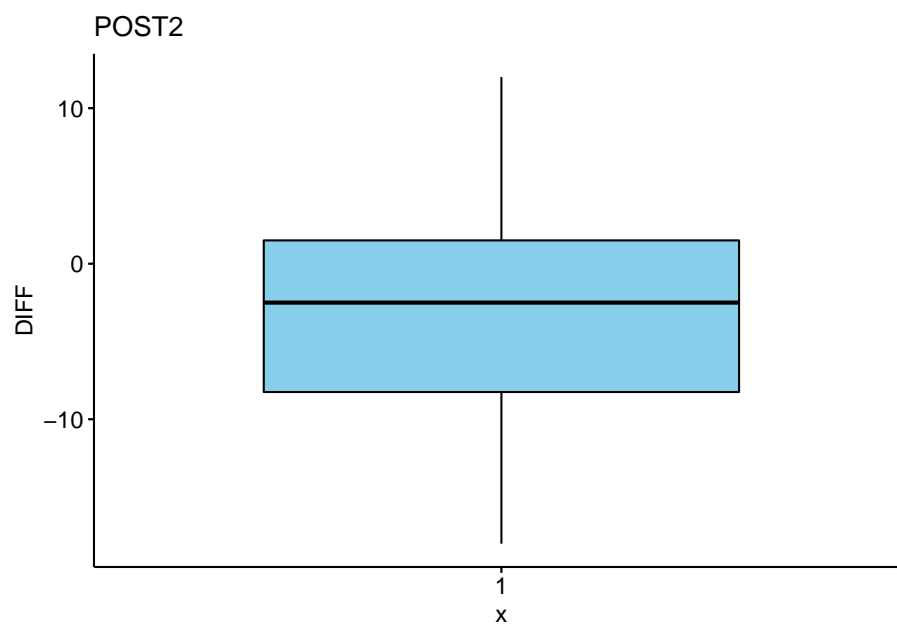
create_box(unique(a$TEST)[1]) # BASELINE
```



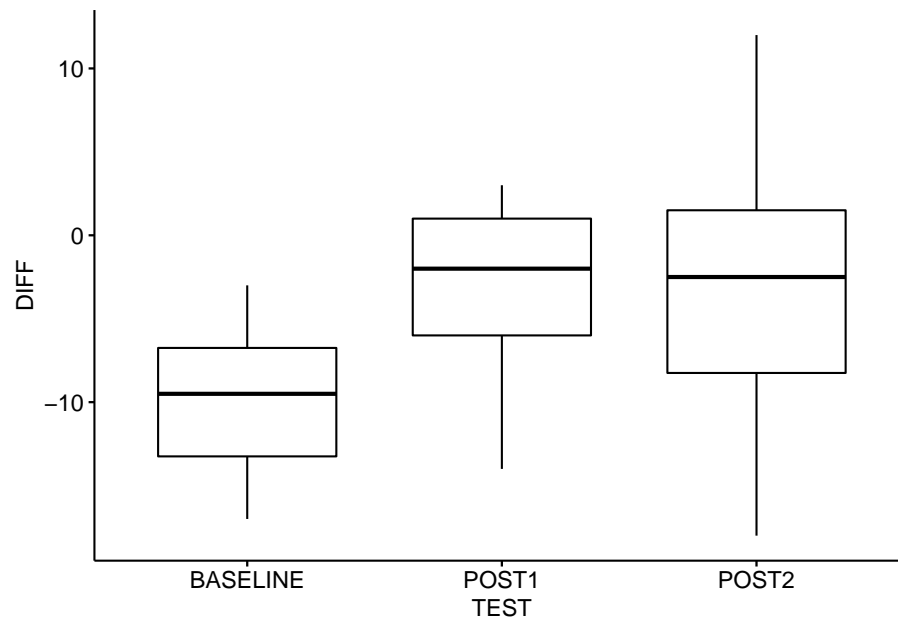
```
create_box(unique(a$TEST)[2]) # POST1
```



```
create_box(unique(a$TEST)[3]) # POST2
```

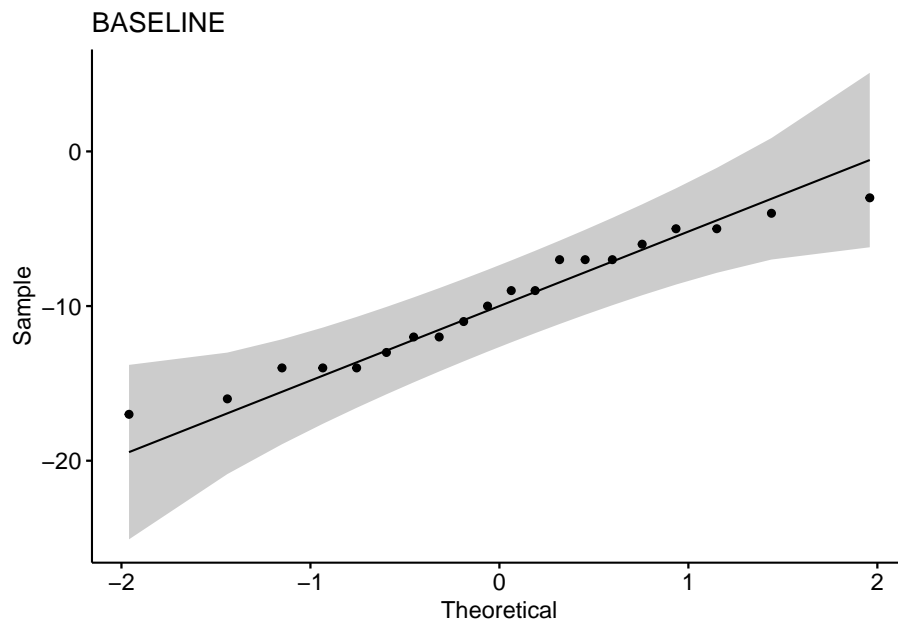


```
# distribution of DIFF by TEST
ggboxplot(data = a,
          x = "TEST", y = "DIFF")
```

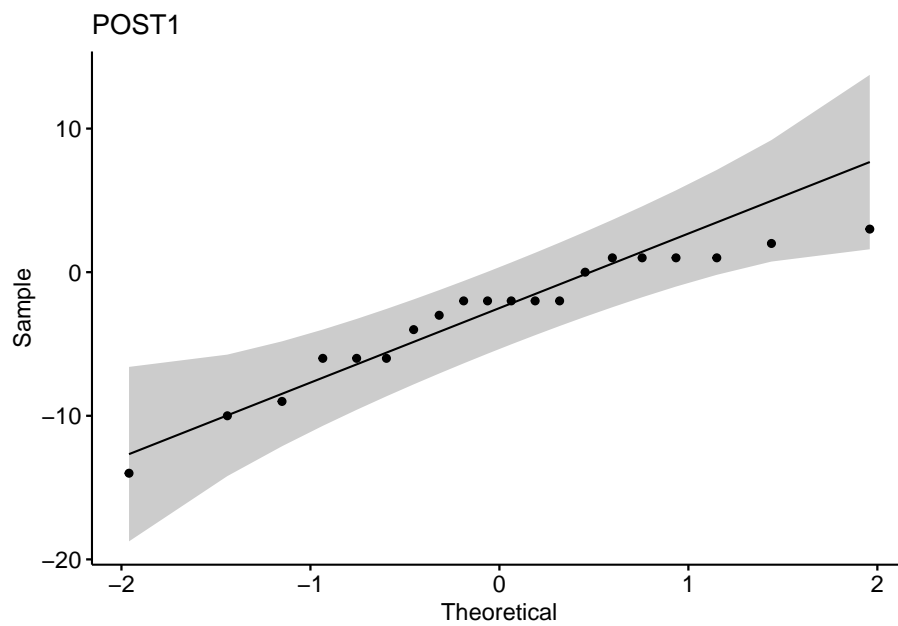


```
# qqplots
create_qq <- function(test_id){
  a %>%
    filter(TEST == {{test_id}}) %>%
    ggqqplot("DIFF",
              main = {{test_id}})
}

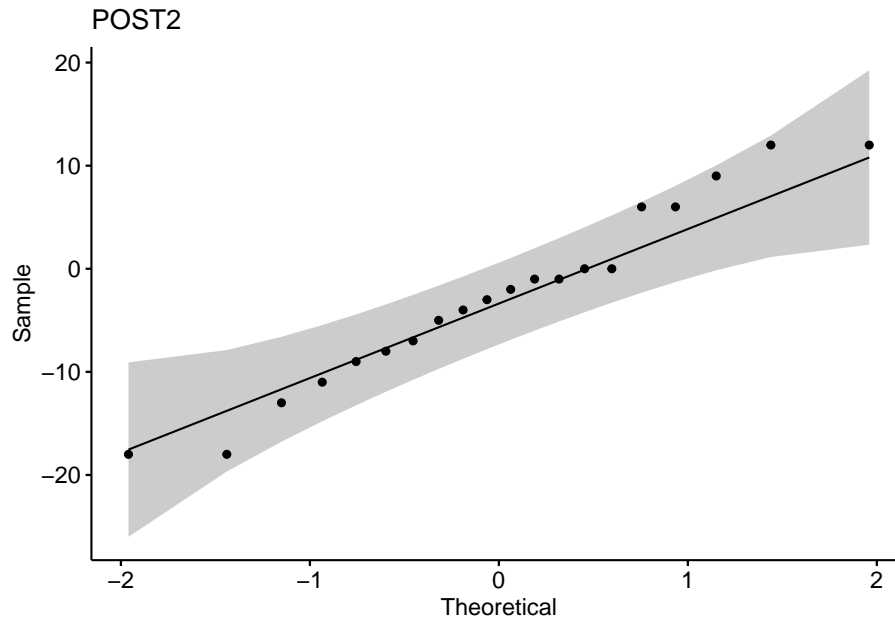
create_qq(unique(a$TEST)[1]) # BASELINE
```



```
create_qq(unique(a$TEST)[2]) # POST1
```



```
create_qq(unique(a$TEST)[3]) # POST2
```



1.5 Normality test

```
st <- shapiro.test(a$DIFF)
data.frame(st$method, st$statistic, st$p.value)
```

```
##                               st.method st.statistic st.p.value
## W Shapiro-Wilk normality test    0.9783355  0.3622644
```

```
do_shp_test <- function(df){
  st <- shapiro.test(df$DIFF)
  data.frame(st$method, st$statistic, st$p.value)
}
```

```
a %>%
  group_by(TEST) %>%
  do(do_shp_test(.)) %>%
  ungroup()
```



```
## # A tibble: 3 x 4
##   TEST      st.method      st.statistic st.p.value
##   <chr>    <fct>          <dbl>      <dbl>
## 1 BASELINE Shapiro-Wilk normality test    0.958    0.511
## 2 POST1    Shapiro-Wilk normality test    0.919    0.0958
## 3 POST2    Shapiro-Wilk normality test    0.965    0.638
```

1.6 Linear regression

```
# fit linear regression by TEST
# Response: ESTIMATE
est_lm <- function(df){
  model <- lm(ESTIMATE ~ ACTUAL, data = df)

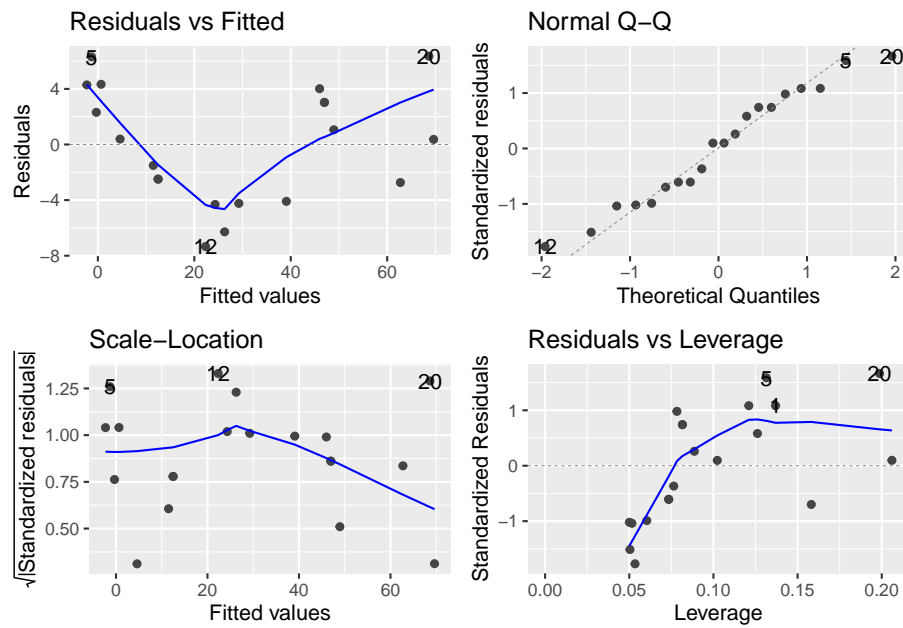
  # fit plot
  fit_plot <- ggplot(data = df, aes(x = ACTUAL, y = ESTIMATE)) +
    geom_point()+
    stat_smooth(method = "lm", col = "blue")

  return(list(summary(model), autoplot(model), fit_plot))
}

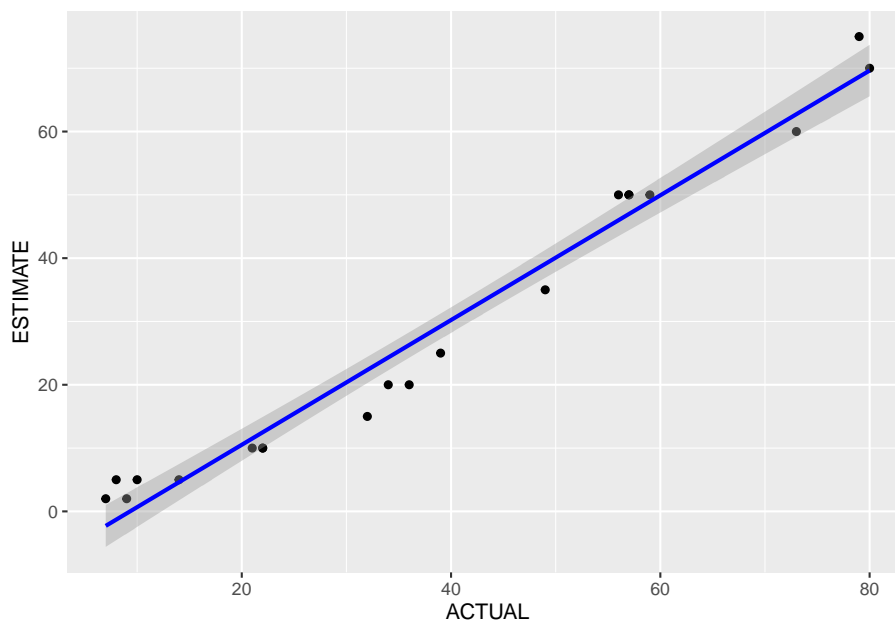
by(a, a$TEST, est_lm)
```

```
## a$TEST: BASELINE
## [[1]]
##
## Call:
## lm(formula = ESTIMATE ~ ACTUAL, data = df)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -7.3419 -3.0730  0.3805  3.2750  6.3549
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -9.1837     1.8107  -5.072 7.95e-05 ***
## ACTUAL         0.9852     0.0403  24.448 2.93e-15 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.264 on 18 degrees of freedom
## Multiple R-squared:  0.9708, Adjusted R-squared:  0.9691
```

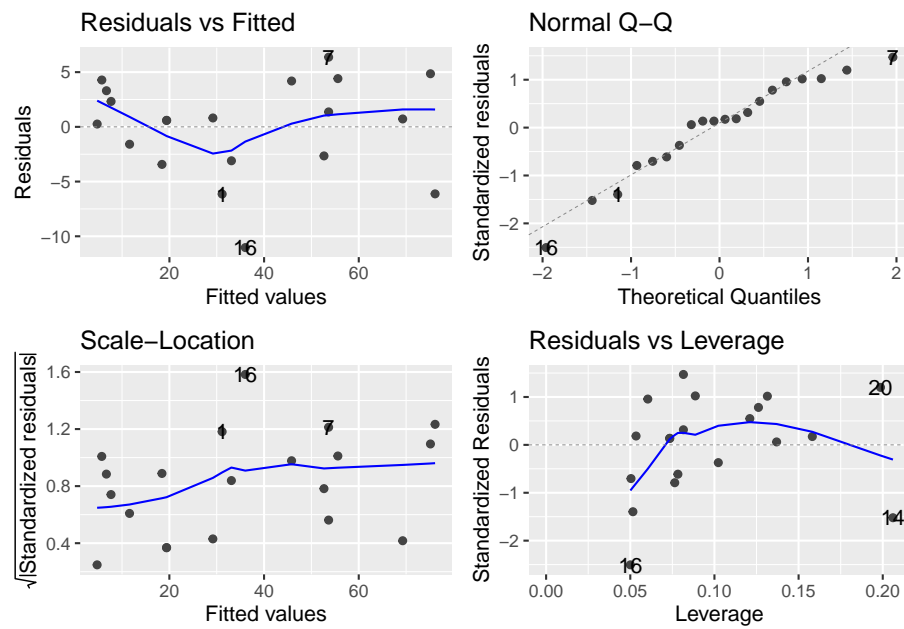
```
## F-statistic: 597.7 on 1 and 18 DF,  p-value: 2.933e-15
##
##
## [[2]]
```



```
##
## [[3]]
```

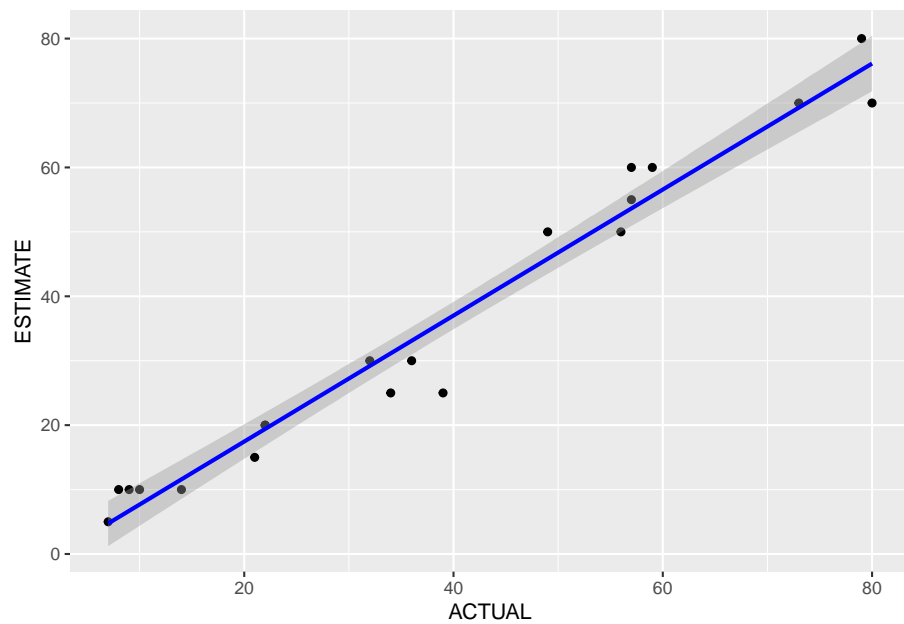


```
##
## -----
## a$TEST: POST1
## [[1]]
##
## Call:
## lm(formula = ESTIMATE ~ ACTUAL, data = df)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -11.0323  -2.7663   0.6561   3.5244   6.3667
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -2.1033     1.9186  -1.096   0.287
## ACTUAL         0.9778     0.0427  22.901 9.18e-15 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.518 on 18 degrees of freedom
## Multiple R-squared:  0.9668, Adjusted R-squared:  0.965
## F-statistic: 524.5 on 1 and 18 DF,  p-value: 9.183e-15
##
##
## [[2]]
```



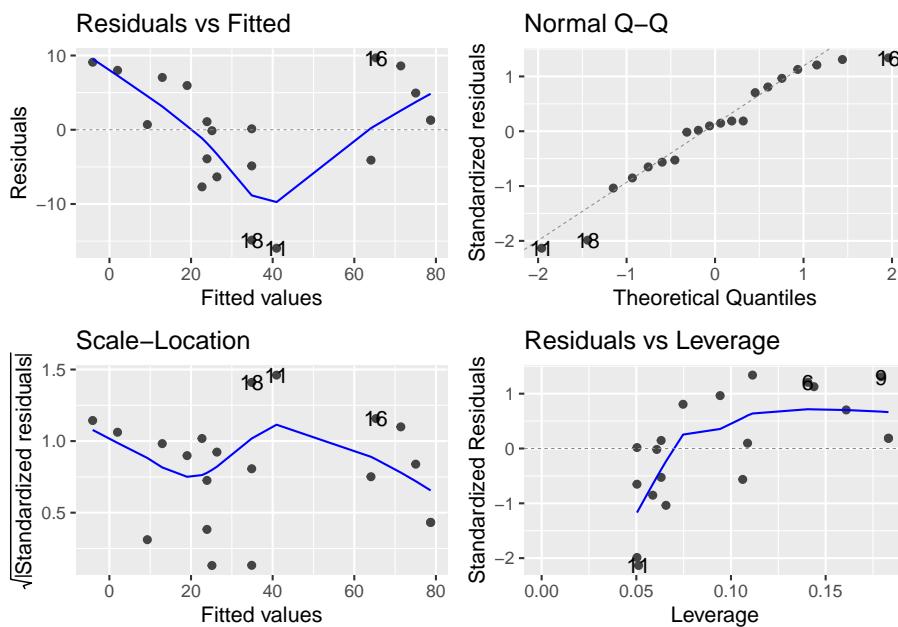
##

[[3]]

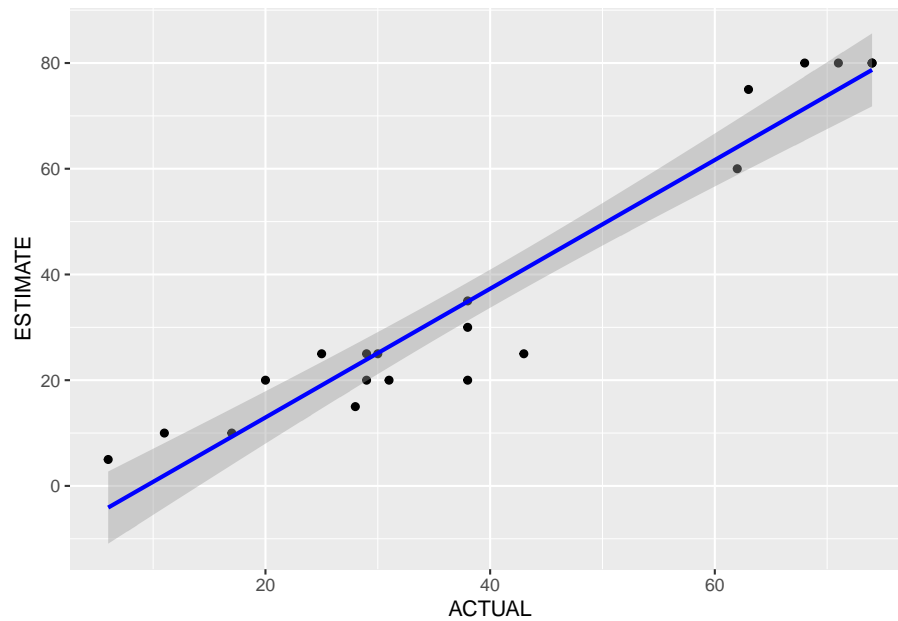


##

```
## -----
## a$TEST: POST2
## [[1]]
##
## Call:
## lm(formula = ESTIMATE ~ ACTUAL, data = df)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -15.9575  -4.2873   0.8961   6.2328   9.6890
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -11.40269    3.67614  -3.102  0.00615 **
## ACTUAL       1.21768    0.08178  14.889 1.46e-11 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 7.675 on 18 degrees of freedom
## Multiple R-squared:  0.9249, Adjusted R-squared:  0.9207
## F-statistic: 221.7 on 1 and 18 DF,  p-value: 1.46e-11
##
## [[2]]
```



```
##
## [[3]]
```



```
# Response: DIFF
diff_lm <- function(df){
  model <- lm(DIFF ~ ACTUAL, data = df)

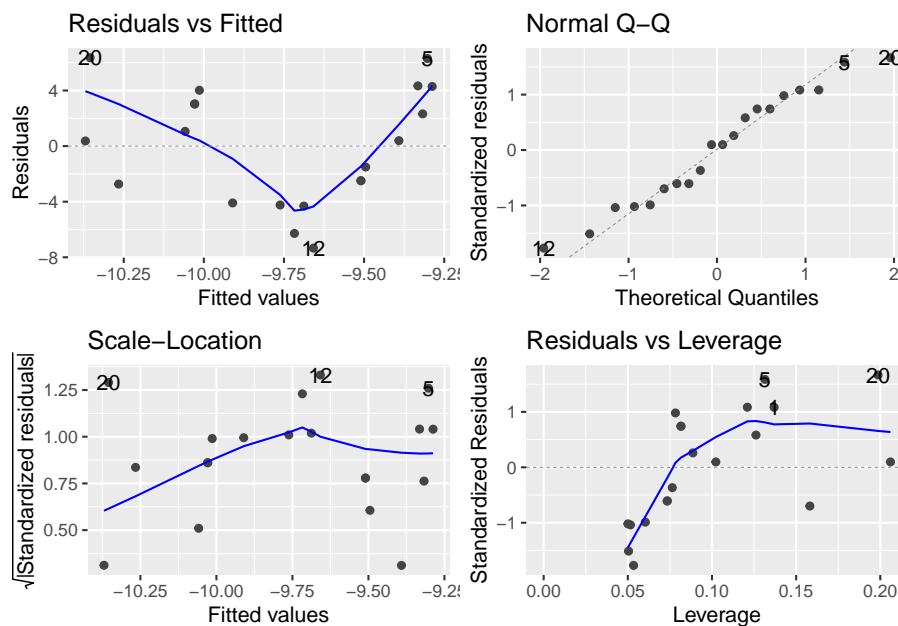
  # fit plot
  fit_plot <- ggplot(data = df, aes(x = ACTUAL, y = DIFF)) +
    geom_point() +
    stat_smooth(method = "lm", col = "blue")

  return(list(summary(model), autoplot(model)))
}

by(a, a$TEST, diff_lm)
```

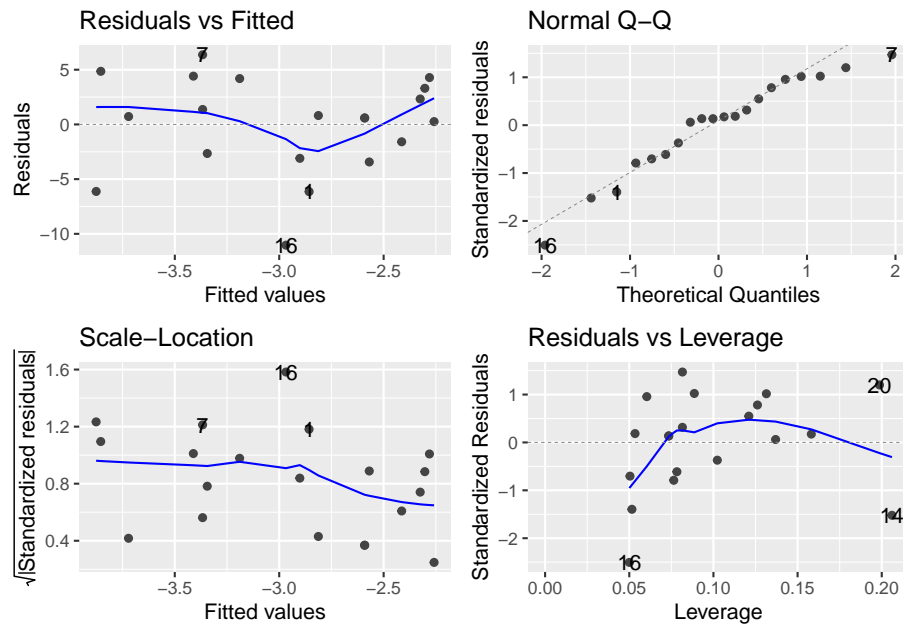
```
## a$TEST: BASELINE
## [[1]]
##
## Call:
## lm(formula = DIFF ~ ACTUAL, data = df)
##
## Residuals:
```

```
##      Min      1Q  Median      3Q      Max
## -7.3419 -3.0730  0.3805  3.2750  6.3549
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -9.18368    1.81073  -5.072 7.95e-05 ***
## ACTUAL      -0.01483    0.04030  -0.368  0.717
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.264 on 18 degrees of freedom
## Multiple R-squared:  0.007463, Adjusted R-squared:  -0.04768
## F-statistic: 0.1353 on 1 and 18 DF, p-value: 0.7172
##
##
## [[2]]
```



```
##
## -----
## a$TEST: POST1
## [[1]]
##
## Call:
## lm(formula = DIFF ~ ACTUAL, data = df)
```

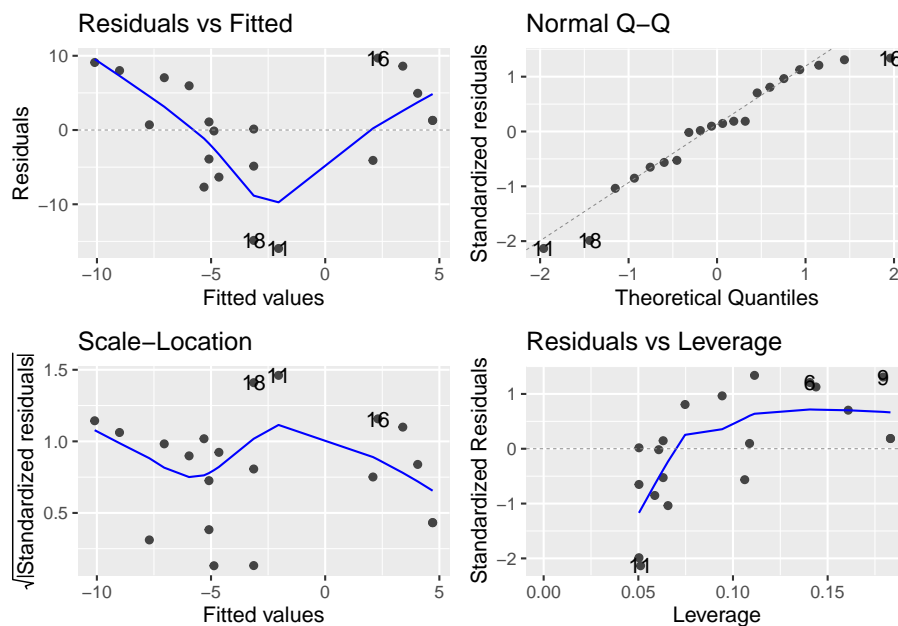
```
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -11.0323  -2.7663   0.6561   3.5244   6.3667
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -2.10325     1.91861  -1.096   0.287
## ACTUAL      -0.02217     0.04270  -0.519   0.610
##
## Residual standard error: 4.518 on 18 degrees of freedom
## Multiple R-squared:  0.01475,    Adjusted R-squared:  -0.03998
## F-statistic: 0.2695 on 1 and 18 DF,  p-value: 0.61
##
##
## [[2]]
```



```
##
## -----
## a$TEST: POST2
## [[1]]
##
## Call:
## lm(formula = DIFF ~ ACTUAL, data = df)
```



```
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -15.9575  -4.2873   0.8961   6.2328   9.6890
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) -11.40269    3.67614  -3.102  0.00615 **
## ACTUAL       0.21768    0.08178   2.662  0.01589 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 7.675 on 18 degrees of freedom
## Multiple R-squared:  0.2824, Adjusted R-squared:  0.2426
## F-statistic: 7.084 on 1 and 18 DF,  p-value: 0.01589
##
##
## [[2]]
```



Chapter 2

Exercise 4

2.1 Load packages

Here is the R code to download the required packages for this exercise.

```
# install package manager 'pacman'  
if (!require(pacman)){  
  install.packages('pacman')  
}
```

Loading required package: pacman

Load the packages needed for this exercise:

```
pacman::p_load(tidyverse,  
               nlme,  
               emmeans)
```

2.2 Import data

Our data is located in `ex4.csv` file, which can be found on my github repo. Import the data and create new variables using the code below.

```
# import data  
a <- read_csv("https://raw.githubusercontent.com/luckymehra/epidem-exercises/master/data/ex4.csv")  
col_types = cols(  
  blk = col_factor(), # parse blk as a factor
```

Table 2.1: The first 6 rows of dataset *a*.

plot	t	blk	trt	pctsev	y	ystar	wt
101	0	1	2	9	0.09	-2.313635	0.0819
102	0	1	1	6	0.06	-2.751535	0.0564
103	0	1	3	2	0.02	-3.891820	0.0196
201	0	2	2	7	0.07	-2.586689	0.0651
202	0	2	3	5	0.05	-2.944439	0.0475
203	0	2	1	3	0.03	-3.476099	0.0291

```

      trt = col_factor() # parse trt as a factor
    ))

# create new variables
a$y <- a$pctsev/100
a$ystar <- log(a$y/(1-a$y))
a$wt <- a$y*(1-a$y)

# print the data
knitr::kable(head(a),
              caption = "The first 6 rows of dataset *a*.")

# get a glimpse of data
glimpse(a)

```

```

## Observations: 72
## Variables: 8
## $ plot    <dbl> 101, 102, 103, 201, 202, 203, 301, 302, 303, 401, 402, ...
## $ t       <dbl> 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 7, 7, 7, 7, 7, 7, 7...
## $ blk     <fct> 1, 1, 1, 2, 2, 2, 3, 3, 3, 4, 4, 4, 1, 1, 1, 2, 2, 2, 3...
## $ trt     <fct> 2, 1, 3, 2, 3, 1, 3, 2, 1, 1, 2, 3, 2, 1, 3, 2, 3, 1, 3...
## $ pctsev  <dbl> 9, 6, 2, 7, 5, 3, 4, 2, 6, 1, 1, 4, 4, 6, 10, 2, 5, 3, ...
## $ y       <dbl> 0.09, 0.06, 0.02, 0.07, 0.05, 0.03, 0.04, 0.02, 0.06, 0...
## $ ystar   <dbl> -2.313635, -2.751535, -3.891820, -2.586689, -2.944439, ...
## $ wt      <dbl> 0.0819, 0.0564, 0.0196, 0.0651, 0.0475, 0.0291, 0.0384, ...

```

2.3 First mixed model

2.3.1 Fit the model

Run the mixed model analysis using **nlme** package in R. The function used to fit the mixed model is called `lme()`.

```

# fit the model

mm_1 <- lme(ystar ~ trt*t, # fixed effects
            data = a,
            random = list(blk = ~ 1, plot = ~ 1), # random effects
            correlation = corAR1(form = (plot = ~ 1)), # specify that observations within a plot
            contrasts = list(trt = "contr.SAS"), # specify this option to get parameter estimates
            weights = ~ I(1/wt))

# output the summary
summary(mm_1)

## Linear mixed-effects model fit by REML
## Data: a
##      AIC      BIC    logLik
## 210.5257 232.4222 -95.26285
##
## Random effects:
## Formula: ~1 | blk
##      (Intercept)
## StdDev: 0.1887117
##
## Formula: ~1 | plot %in% blk
##      (Intercept) Residual
## StdDev: 4.604287e-05 0.2519511
##
## Correlation Structure: AR(1)
## Formula: ~1 | blk/plot
## Parameter estimate(s):
##      Phi
## 0.06205463
## Variance function:
## Structure: fixed weights
## Formula: ~I(1/wt)
## Fixed effects: ystar ~ trt * t
##              Value Std.Error DF   t-value p-value
## (Intercept) -2.5689859 0.3629604 57 -7.077868 0.0000
## trt2         -0.1948084 0.5193013  6 -0.375136 0.7205
## trt1         -0.5406041 0.5136249  6 -1.052527 0.3331
## t            0.0992675 0.0142177 57  6.981964 0.0000
## trt2:t       -0.0221696 0.0202998 57 -1.092109 0.2794
## trt1:t        0.0437431 0.0212717 57  2.056398 0.0443
## Correlation:
##      (Intr) trt2   trt1   t      trt2:t
## trt2  -0.652

```

```
## trt1    -0.658  0.459
## t       -0.888  0.621  0.627
## trt2:t   0.623 -0.924 -0.439 -0.700
## trt1:t   0.592 -0.413 -0.913 -0.667  0.466
##
## Standardized Within-Group Residuals:
##           Min           Q1           Med           Q3           Max
## -2.1518915 -0.6900213 -0.4024653  0.4132408  2.7733450
##
## Number of Observations: 72
## Number of Groups:
##           blk plot %in% blk
##           4      12
```

```
# extract covariance parameter estimates
VarCorr(mm_1)
```

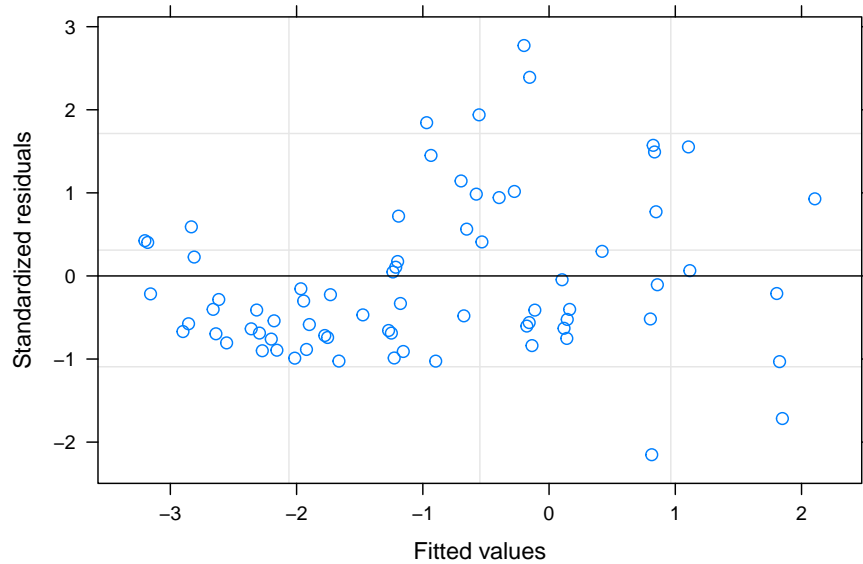
```
##           Variance      StdDev
## blk =      pdLogChol(1)
## (Intercept) 3.561212e-02 1.887117e-01
## plot =      pdLogChol(1)
## (Intercept) 2.119946e-09 4.604287e-05
## Residual    6.347936e-02 2.519511e-01
```

```
# extract type3 fixed effects anova
anova.lme(mm_1, type = 'marginal')
```

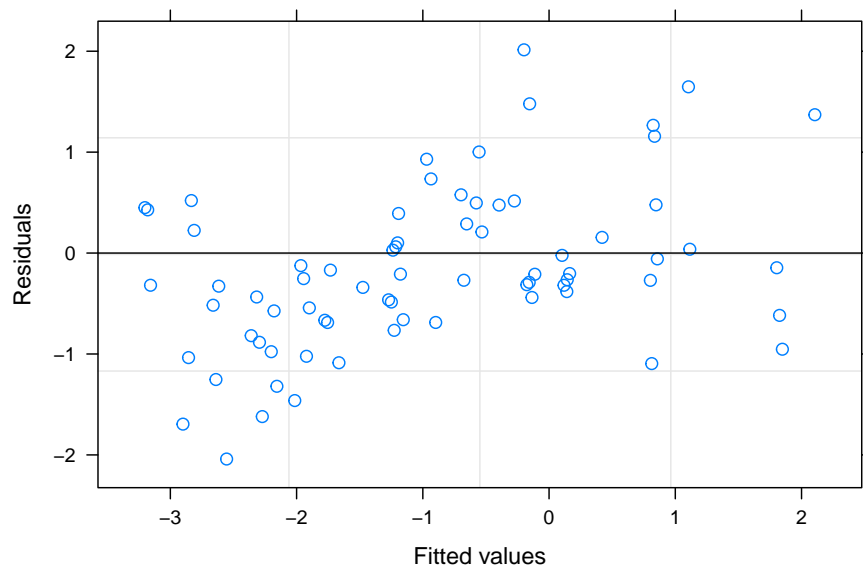
```
##           numDF denDF  F-value p-value
## (Intercept)      1    57 50.09622 <.0001
## trt              2     6  0.56135  0.5977
## t                1    57 48.74782 <.0001
## trt:t            2    57  4.80110  0.0118
```

2.3.2 Diagnostic plots

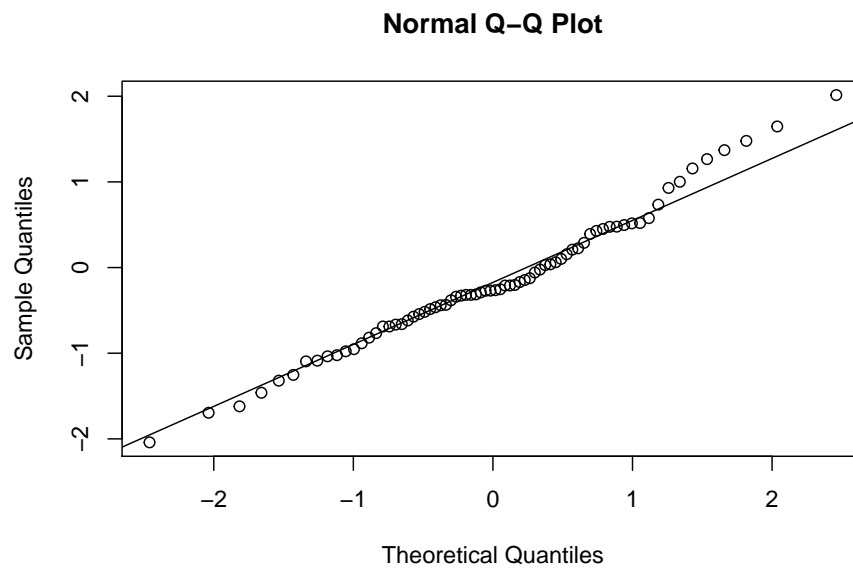
```
# pearson residuals vs. fitted values
plot(mm_1, resid(., type="pearson") ~ fitted(.), abline = 0)
```



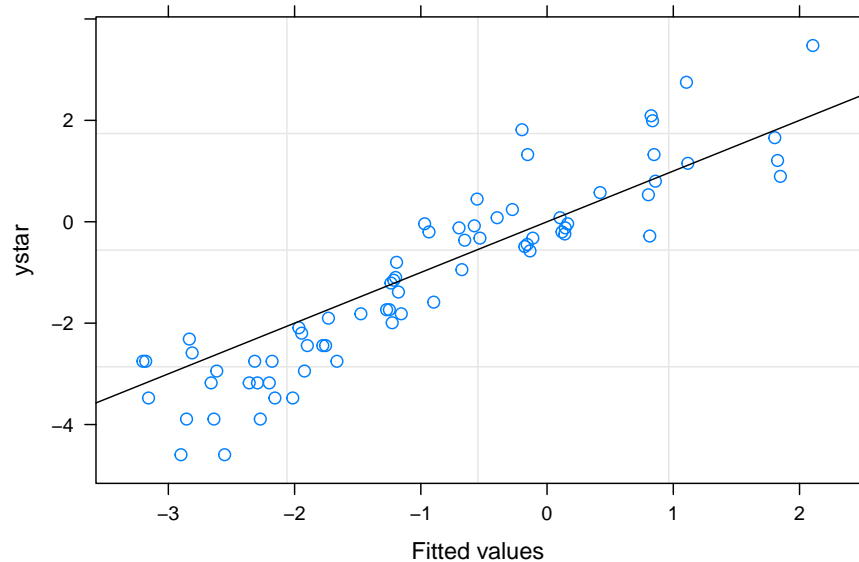
```
# standardized residuals vs. fitted values  
plot(mm_1, resid(., scaled=TRUE) ~ fitted(.), abline = 0)
```



```
# qq plot  
qqnorm(residuals(mm_1))  
qqline(residuals(mm_1))
```



```
#observed vs. fitted values  
plot(mm_1, ystar ~ fitted(.), abline = c(0,1))
```

2.4 Second mixed model

2.4.1 Fit the model

Run the mixed model analysis using **nlme** package in R. The function used to fit the mixed model is called `lme()`. Here we will specify no intercept. We will also use **emmeans** package to get least squared means and contrasts.

```
# fit the model
#library(nlme)
mm_2 <- update(mm_1, fixed = ystar ~ - 1 + trt + trt:t) # update fixed effects in mm_1, -1 indicates no intercept

# output the summary
summary(mm_2)

## Linear mixed-effects model fit by REML
## Data: a
##      AIC      BIC    logLik
## 210.5257 232.4222 -95.26285
##
## Random effects:
## Formula: ~1 | blk
```

```
##          (Intercept)
## StdDev:   0.1887117
##
## Formula: ~1 | plot %in% blk
##          (Intercept) Residual
## StdDev: 4.603147e-05 0.2519511
##
## Correlation Structure: AR(1)
## Formula: ~1 | blk/plot
## Parameter estimate(s):
##          Phi
## 0.06205463
## Variance function:
## Structure: fixed weights
## Formula: ~I(1/wt)
## Fixed effects: ystar ~ trt + trt:t - 1
##          Value Std.Error DF   t-value p-value
## trt2    -2.7637943 0.3944803   6 -7.006165 4e-04
## trt1    -3.1095900 0.3877657   6 -8.019250 2e-04
## trt3    -2.5689859 0.3629604   6 -7.077868 4e-04
## trt2:t    0.0770979 0.0144893  58  5.321034 0e+00
## trt1:t    0.1430106 0.0158560  58  9.019328 0e+00
## trt3:t    0.0992675 0.0142177  58  6.981964 0e+00
## Correlation:
##          trt2   trt1   trt3   trt2:t trt1:t
## trt1      0.057
## trt3      0.062  0.065
## trt2:t   -0.901  0.001  0.001
## trt1:t    0.001 -0.881 -0.002 -0.001
## trt3:t    0.000 -0.002 -0.888  0.000  0.002
##
## Standardized Within-Group Residuals:
##          Min          Q1          Med          Q3          Max
## -2.1518915 -0.6900213 -0.4024653  0.4132408  2.7733450
##
## Number of Observations: 72
## Number of Groups:
##          blk plot %in% blk
##          4          12
```

```
# extract covariance parameter estimates
VarCorr(mm_2)
```

```
##          Variance      StdDev
## blk =      pdLogChol(1)
```

```
## (Intercept) 3.561212e-02 1.887117e-01
## plot =      pdLogChol(1)
## (Intercept) 2.118896e-09 4.603147e-05
## Residual    6.347936e-02 2.519511e-01
```

```
# extract type3 fixed effects anova
anova.lme(mm_2, type = 'marginal')
```

```
##      numDF denDF  F-value p-value
## trt      3      6 48.57698 1e-04
## trt:t    3     58 52.74601 <.0001
```

```
# compare the slopes for different treatments
#library(emmeans)
```

```
emtrends(mm_2, pairwise ~ trt, var="t", adjust = "none")
```

```
## $emtrends
##   trt t.trend      SE df lower.CL upper.CL
## 2    0.0771 0.0145 58    0.0481    0.106
## 1    0.1430 0.0159 58    0.1113    0.175
## 3    0.0993 0.0142 58    0.0708    0.128
##
## d.f. method: containment
## Confidence level used: 0.95
##
## $contrasts
##   contrast estimate      SE df t.ratio p.value
## 2 - 1      -0.0659 0.0215 58   -3.067  0.0033
## 2 - 3      -0.0222 0.0203 58   -1.092  0.2793
## 1 - 3       0.0437 0.0213 58    2.056  0.0443
```

```
# get the treatment difference at various time points
```

```
emmeans(mm_2, pairwise ~ trt|t, nesting = NULL, at = list(t = c(0, 7, 14, 21, 28, 35)), adjust =
```

```
## $emmeans
## t = 0:
##   trt emmean      SE df lower.CL upper.CL
## 2    -2.7638 0.394   6    -3.729  -1.7985
## 1    -3.1096 0.388   6    -4.058  -2.1608
## 3    -2.5690 0.363   6    -3.457  -1.6809
##
## t = 7:
##   trt emmean      SE df lower.CL upper.CL
```

```

## 2 -2.2241 0.306 6 -2.974 -1.4746
## 1 -2.1085 0.295 6 -2.830 -1.3873
## 3 -1.8741 0.278 6 -2.555 -1.1931
##
## t = 14:
## trt emmean SE df lower.CL upper.CL
## 2 -1.6844 0.229 6 -2.246 -1.1232
## 1 -1.1074 0.219 6 -1.644 -0.5712
## 3 -1.1792 0.207 6 -1.687 -0.6719
##
## t = 21:
## trt emmean SE df lower.CL upper.CL
## 2 -1.1447 0.179 6 -1.582 -0.7072
## 1 -0.1064 0.184 6 -0.556 0.3437
## 3 -0.4844 0.168 6 -0.896 -0.0726
##
## t = 28:
## trt emmean SE df lower.CL upper.CL
## 2 -0.6051 0.179 6 -1.042 -0.1680
## 1 0.8947 0.210 6 0.380 1.4095
## 3 0.2105 0.183 6 -0.237 0.6581
##
## t = 35:
## trt emmean SE df lower.CL upper.CL
## 2 -0.0654 0.229 6 -0.626 0.4948
## 1 1.8958 0.282 6 1.207 2.5850
## 3 0.9054 0.242 6 0.314 1.4968
##
## d.f. method: containment
## Confidence level used: 0.95
##
## $contrasts
## t = 0:
## contrast estimate SE df t.ratio p.value
## 2 - 1 0.3458 0.537 6 0.644 0.5435
## 2 - 3 -0.1948 0.519 6 -0.375 0.7205
## 1 - 3 -0.5406 0.514 6 -1.053 0.3331
##
## t = 7:
## contrast estimate SE df t.ratio p.value
## 2 - 1 -0.1156 0.404 6 -0.286 0.7843
## 2 - 3 -0.3500 0.392 6 -0.893 0.4062
## 1 - 3 -0.2344 0.383 6 -0.613 0.5625
##
## t = 14:
## contrast estimate SE df t.ratio p.value

```

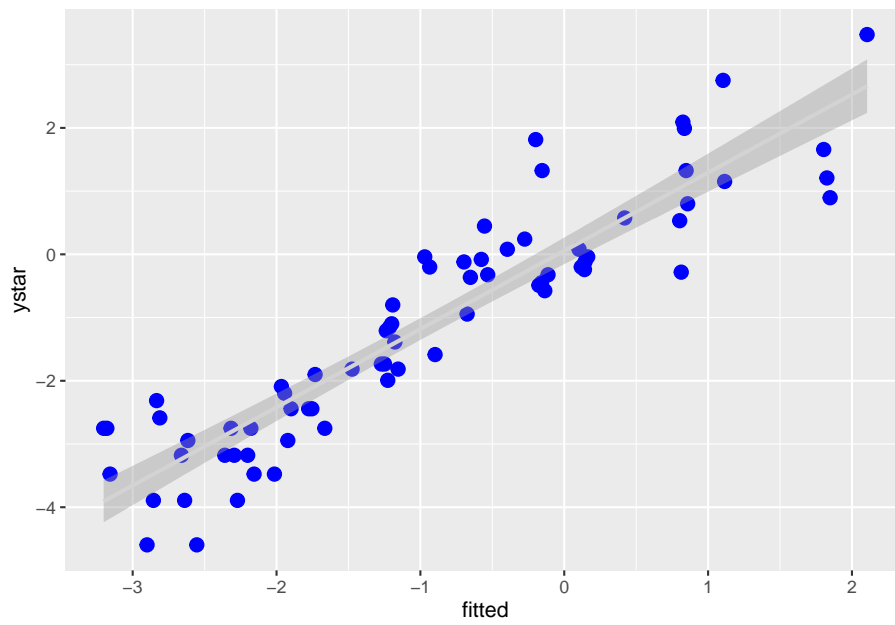
```
## 2 - 1      -0.5770 0.288  6 -2.004  0.0919
## 2 - 3      -0.5052 0.279  6 -1.811  0.1201
## 1 - 3       0.0718 0.271  6  0.265  0.7996
##
## t = 21:
## contrast estimate      SE df t.ratio p.value
## 2 - 1      -1.0384 0.219  6 -4.739  0.0032
## 2 - 3      -0.6604 0.206  6 -3.204  0.0185
## 1 - 3       0.3780 0.211  6  1.794  0.1229
##
## t = 28:
## contrast estimate      SE df t.ratio p.value
## 2 - 1      -1.4998 0.242  6 -6.204  0.0008
## 2 - 3      -0.8156 0.218  6 -3.741  0.0096
## 1 - 3       0.6842 0.245  6  2.795  0.0314
##
## t = 35:
## contrast estimate      SE df t.ratio p.value
## 2 - 1      -1.9611 0.338  6 -5.806  0.0011
## 2 - 3      -0.9707 0.305  6 -3.184  0.0190
## 1 - 3       0.9904 0.346  6  2.861  0.0288
```

2.4.2 Plot observed versus predicted model values

```
# add fitted and residuals in to a new dataset called b
b = cbind(a, resid = resid(mm_2), fitted = fitted(mm_2))

# fit linear regression
b.lm <- lm(ystar ~ fitted, data=b)

# plot using ggplot2 package
ggplot(b, aes(x=fitted, y = ystar)) +
  geom_point(color="blue", size = 3) +
  geom_smooth(method = lm, color = "lightgrey")
```



Chapter 3

Exercise 9.4

3.1 Load packages

Here is the R code to download the required packages for this exercise.

```
# install package manager 'pacman'  
if (!require(pacman)){  
  install.packages('pacman')  
}
```

Loading required package: pacman

```
# load packages needed for this exercise  
library(pacman)  
p_load(tidyverse,  
       lctools, # to calculate Moran's I  
       spdep, # to calculate geary's c  
       geoR, # to compute variogram  
       gridExtra, # to stack plots  
       gstat, automap, # packages for variogram model selection  
       sp # need a function called `coordinates`  
       )
```

3.2 Data

This is equivalent to data step in SAS. Here, the data is entered inside a function called `tibble`.

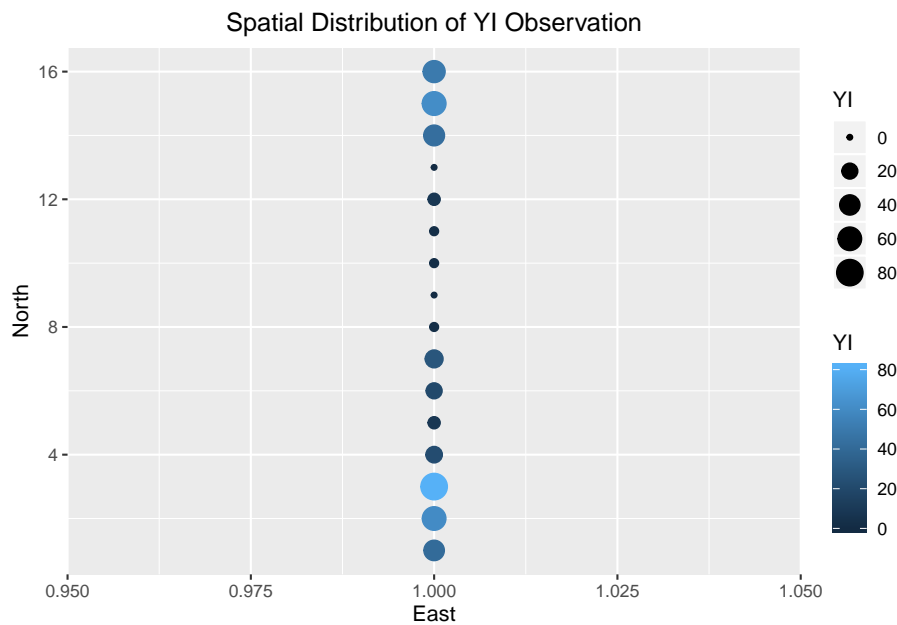
```
# Enter data
a <- tibble(I = 1:16, YI = c(41, 60, 81, 22, 8, 20, 28, 2,
                             0, 2, 2, 8, 0, 43, 61, 50)) %>%
  # creat new variable East and North
  mutate(East = 1,
         North = I)

# print the data
a
```

```
## # A tibble: 16 x 4
##       I     YI East North
##   <int> <dbl> <dbl> <int>
## 1     1    41     1     1
## 2     2    60     1     2
## 3     3    81     1     3
## 4     4    22     1     4
## 5     5     8     1     5
## 6     6    20     1     6
## 7     7    28     1     7
## 8     8     2     1     8
## 9     9     0     1     9
## 10    10     2     1    10
## 11    11     2     1    11
## 12    12     8     1    12
## 13    13     0     1    13
## 14    14    43     1    14
## 15    15    61     1    15
## 16    16    50     1    16
```

3.3 Autocorrelation statistics

```
# visualize the data
ggplot(data = a) +
  geom_point(mapping = aes(x = East, y = North, size = YI, color = YI)) +
  ggtitle("Spatial Distribution of YI Observation") +
  theme(plot.title = element_text(hjust = 0.5))
```

```
# calculate Moran's I
Coords <- a %>%
  dplyr::select(East, North)

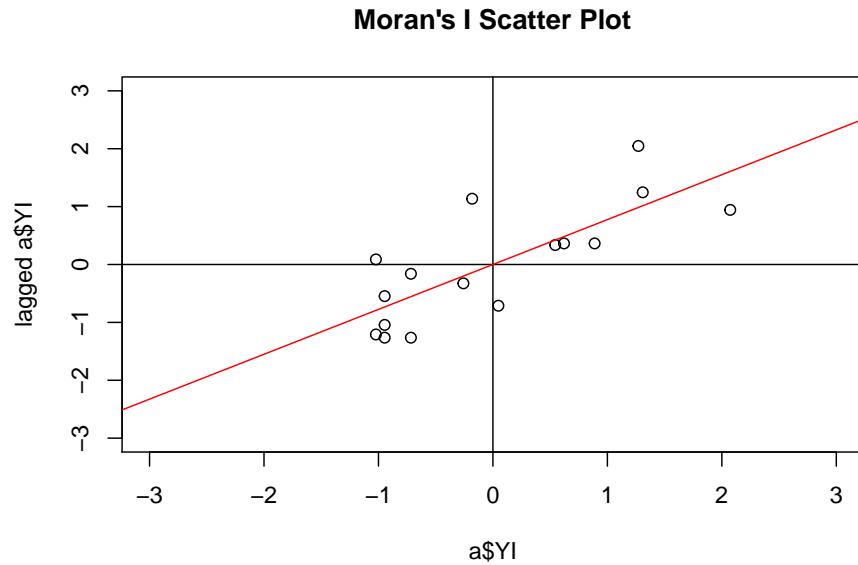
mI <- moransI(Coords, Bandwidth = 1, a$YI)

# print Moran's I table
moran.table <- tribble(
  ~`Moran's I`, ~`Expected I`, ~`Z randomization`, ~`P value randomization`,
  #-----/-----/-----/-----
  mI$Morans.I, mI$Expected.I, mI$z.randomization, mI$p.value.randomization
)

moran.table
```

```
## # A tibble: 1 x 4
##   `Moran's I` `Expected I` `Z randomization` `P value randomization`
##   <dbl>      <dbl>      <dbl>      <dbl>
## 1      0.625      -0.0667      2.81      0.00499
```

```
# create Moran's I scatter plot
l.moran <- l.moransI(Coords, Bandwidth = 1, a$YI)
```



```
# calculate geary's c
Coords_num <- coordinates(Coords)

# create an object of class 'nb' so that it can be used with function from package 'sp'
Coords_nb <- knn2nb(knearneigh(Coords_num))

# create a 'listw' object for use in the function 'geary.test'
coords_listw <- nb2listw(Coords_nb)

gearyC <- geary.test(a$YI, coords_listw, alternative = "two.sided")
gearyC
```

```
##
## Geary C test under randomisation
##
## data: a$YI
## weights: coords_listw
##
## Geary C statistic standard deviate = 2.5826, p-value = 0.009806
## alternative hypothesis: two.sided
## sample estimates:
## Geary C statistic      Expectation      Variance
##      0.37085605      1.00000000      0.05934473
```

3.4 First variogram

We will use the package `geoR` to construct empirical variogram, and then draw them using package `ggplot2`.

```
v1 <- variog(coords = Coords_num, data = a$YI, breaks = seq(0.5, 15.5),
             max.dist = 11)
```

```
## variog: computing omnidirectional variogram
```

```
# extract data from object v1 for plotting
v1_plot_data <- cbind(v1$u, v1$v, v1$n) %>%
  as.data.frame() %>%
  dplyr::rename(Distance = V1,
                Semivariance = V2,
                Pair_count = V3)

# in the table below, gamma is semivariance
v1_plot_data
```

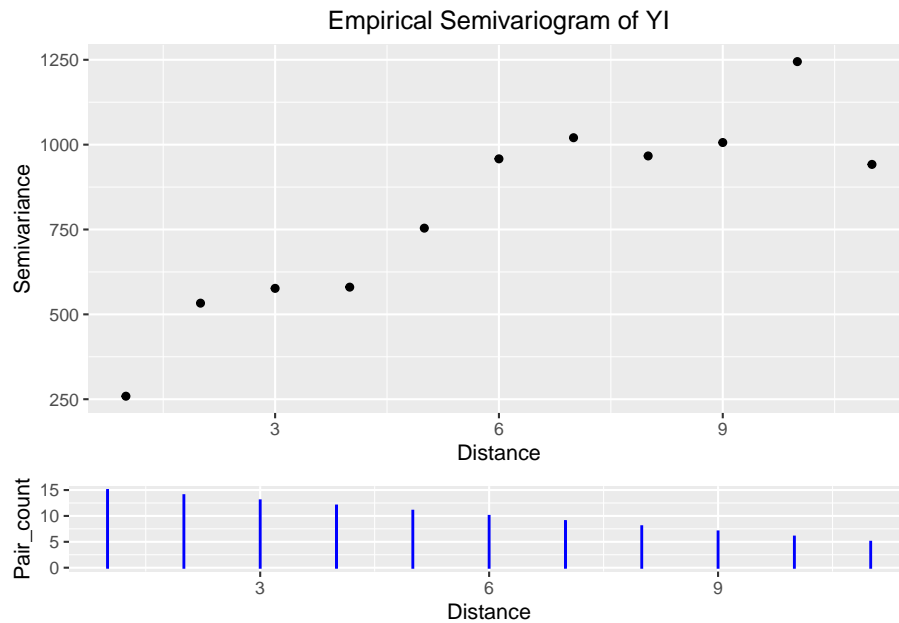
##	Distance	Semivariance	Pair_count
## 1	1	258.8333	15
## 2	2	533.0000	14
## 3	3	576.6154	13
## 4	4	580.1667	12
## 5	5	754.0000	11
## 6	6	958.2000	10
## 7	7	1020.4444	9
## 8	8	966.7500	8
## 9	9	1006.2857	7
## 10	10	1244.6667	6
## 11	11	941.8000	5

```
# plot variogram
v1_plot_vario <- ggplot(data = v1_plot_data) +
  geom_point(mapping = aes(x = Distance, y = Semivariance)) +
  ggtitle("Empirical Semivariogram of YI") +
  theme(plot.title = element_text(hjust = 0.5))

# plot pair counts
v1_plot_pair_count <- ggplot(data = v1_plot_data) +
  geom_col(mapping = aes(x = Distance, y = Pair_count), width = 0.01, color = "blue")

# stack two plots
```

```
grid.arrange(v1_plot_vario, v1_plot_pair_count,
              ncol = 1, heights = c(3, 1))
```



3.5 Second variogram

Plot robust and classical variogram together.

```
# fit robust variogram
v1_robust <- variog(coords = Coords_num, data = a$YI, breaks = seq(0.5, 15.5),
                  max.dist = 11, estimator.type = "modulus")
```

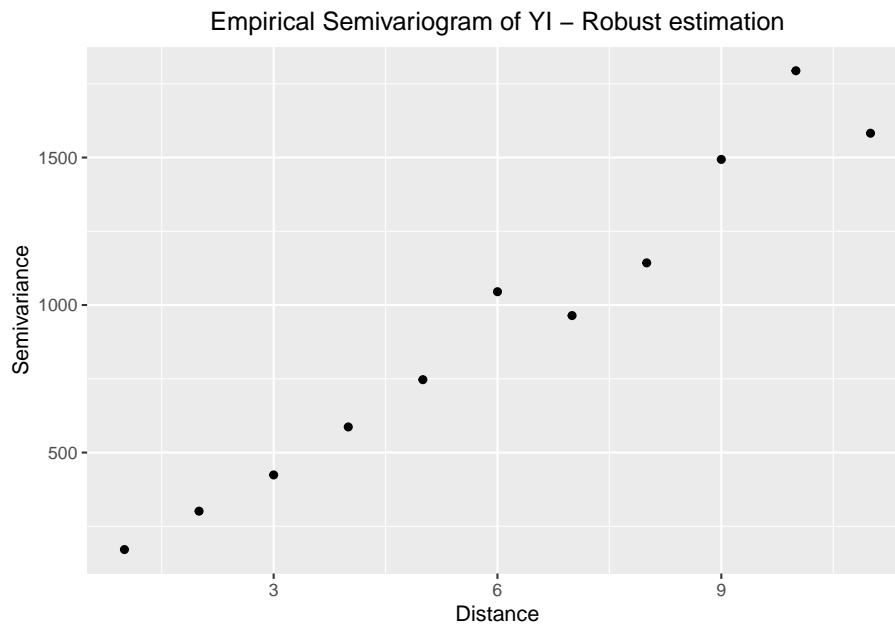
```
## variog: computing omnidirectional variogram
```

```
# extract the data
v1_robust_data <- cbind(v1_robust$u, v1_robust$v, v1_robust$n) %>%
  as.data.frame() %>%
  dplyr::rename(Distance = V1,
                Semivariance = V2,
                Pair_count = V3)

# plot robust variogram
```

```
v1_robust_vario <- ggplot(data = v1_robust_data) +
  geom_point(mapping = aes(x = Distance, y = Semivariance)) +
  ggtitle("Empirical Semivariogram of YI - Robust estimation") +
  theme(plot.title = element_text(hjust = 0.5))

v1_robust_vario
```

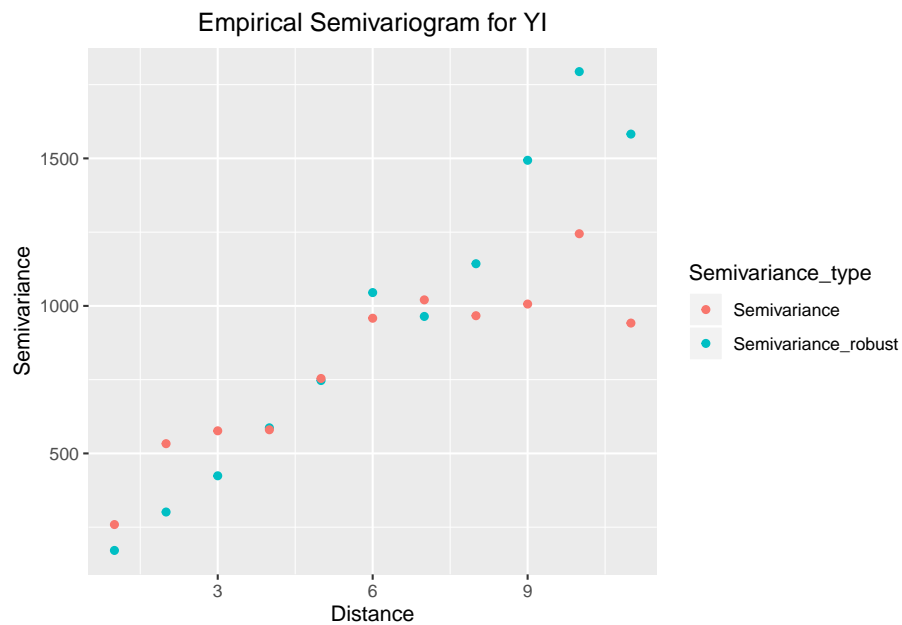


```
# combine robust and classical variogram
var_comb <- v1_robust_data %>%

  # combine robust and classical variogram datasets
  dplyr::rename(Semivariance_robust = Semivariance) %>%
  bind_cols(dplyr::select(v1_plot_data, Semivariance)) %>%
  gather(key = "Semivariance_type", value = "Semivariance", -c(Distance, Pair_count)) %>%

  # plot
  ggplot() +
  geom_point(mapping = aes(x = Distance, y = Semivariance, color = Semivariance_type)) +
  ggtitle("Empirical Semivariogram for YI") +
  theme(plot.title = element_text(hjust = 0.5))

var_comb
```



3.6 Variogram model selection

We will use the package `gstat` and `automap` for variogram model selection

```
# specify coordinates in the dataset
coordinates(a) = ~East+North

# select the best model out of exponential, spherical, and gaussian
autofitVariogram(YI ~ East + North, a, model = c("Sph", "Exp", "Gau"))
```

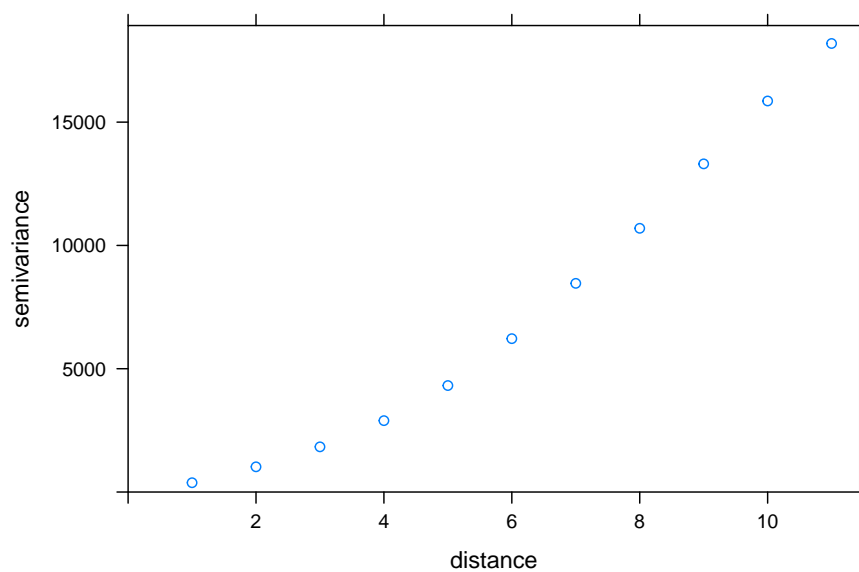
```
## $exp_var
##   np dist    gamma dir.hor dir.ver   id
## 1  15    1 259.9333      0      0 var1
## 2  14    2 536.4286      0      0 var1
## 3  13    3 574.6538      0      0 var1
## 4  12    4 571.5000      0      0 var1
## 5  11    5 743.7727      0      0 var1
##
## $var_model
##   model  psill  range
## 1  Nug  0.0000 0.000000
## 2  Exp 835.8984 2.488287
##
```

```
## $sserr
## [1] 31108.23
##
## attr("class")
## [1] "autofitVariogram" "list"
```

```
# fit empirical variogram
v_emp <- variogram(YI ~ East + North, data = a, cutoff = 11)
v_emp
```

```
##      np dist      gamma dir.hor dir.ver   id
## 1  15    1  377.2333      0      0 var1
## 2  14    2 1022.1429      0      0 var1
## 3  13    3 1832.0000      0      0 var1
## 4  12    4 2894.8333      0      0 var1
## 5  11    5 4317.6364      0      0 var1
## 6  10    6 6219.0000      0      0 var1
## 7   9    7 8462.2222      0      0 var1
## 8   8    810694.7500      0      0 var1
## 9   7    913308.0000      0      0 var1
##10   6   1015858.0000      0      0 var1
##11   5   1118189.8000      0      0 var1
```

```
plot(v_emp)
```



```
# fit exponential variogram
v_exp <- fit.variogram(v_emp, vgm("Exp"))
```

```
## Warning in fit.variogram(v_emp, vgm("Exp")): No convergence after 200
## iterations: try different initial values?
```

```
v_exp
```

```
##   model      psill    range
## 1   Nug    1587.279  0.0000
## 2   Exp  206333.966 176.1914
```

```
# fit spherical and gaussian
v_sph <- fit.variogram(v_emp, vgm("Sph"))
```

```
## Warning in fit.variogram(v_emp, vgm("Sph")): No convergence after 200
## iterations: try different initial values?
```

```
v_gau <- fit.variogram(v_emp, vgm("Gau"))
```

```
# extract plotting data from fitted variograms
v_exp_line <- variogramLine(v_exp, maxdist = 11)
v_sph_line <- variogramLine(v_sph, maxdist = 11)
v_gau_line <- variogramLine(v_gau, maxdist = 11)
```

```
# plot empirical and fitted variograms together
```

```
# specify color for legends
```

```
legend_color <- c("Empirical" = "blue", "Exponential" = "blue",
                  "Spherical" = "orange", "Gaussian" = "green")
```

```
ggplot(data = v_emp) +
```

```
  geom_point(mapping = aes(x = dist, y = gamma, fill = "Empirical"), color = "blue") +
```

```
  geom_line(data = v_exp_line, mapping = aes(x = dist, y = gamma, color = "Exponential")
```

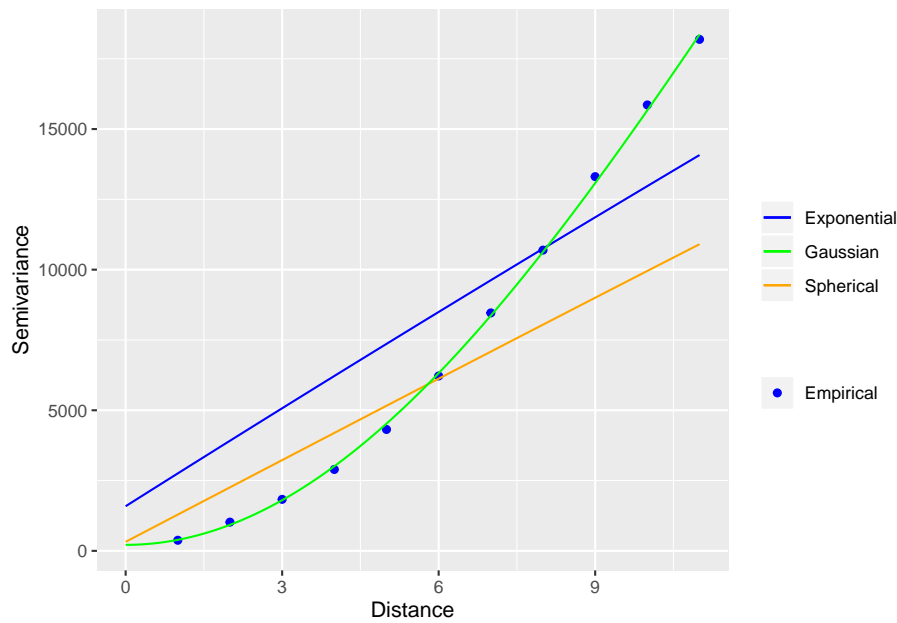
```
  geom_line(data = v_sph_line, mapping = aes(x = dist, y = gamma, color = "Spherical"))
```

```
  geom_line(data = v_gau_line, mapping = aes(x = dist, y = gamma, color = "Gaussian"))
```

```
  scale_color_manual(name = "", values = legend_color) +
```

```
  scale_fill_manual(name = "", values = legend_color) +
```

```
  labs(x = "Distance",
       y = "Semivariance")
```

Chapter 4

Exercise 9.5

4.1 Load packages

Here is the R code to download the required packages for this exercise.

```
# install package manager 'pacman'  
if (!require(pacman)){  
  install.packages('pacman')  
}
```

Loading required package: pacman

```
# load packages needed for this exercise  
library(pacman)  
p_load(tidyverse,  
       lctools, # to calculate Moran's I  
       spdep, # to calculate geary's c  
       geoR, # to compute variogram  
       gridExtra, # to stack plots  
       gstat, automap, # packages for variogram model selection  
       sp # need a function called `coordinates`  
)
```

4.2 Data

This is equivalent to data step in SAS. Here, the data is imported from a file `data.csv` using the function `read_csv`. This function will download the file directly from here.

```
# Import data
a <- read_csv("https://raw.githubusercontent.com/luckymehra/epidem-exercises/master/data.csv")

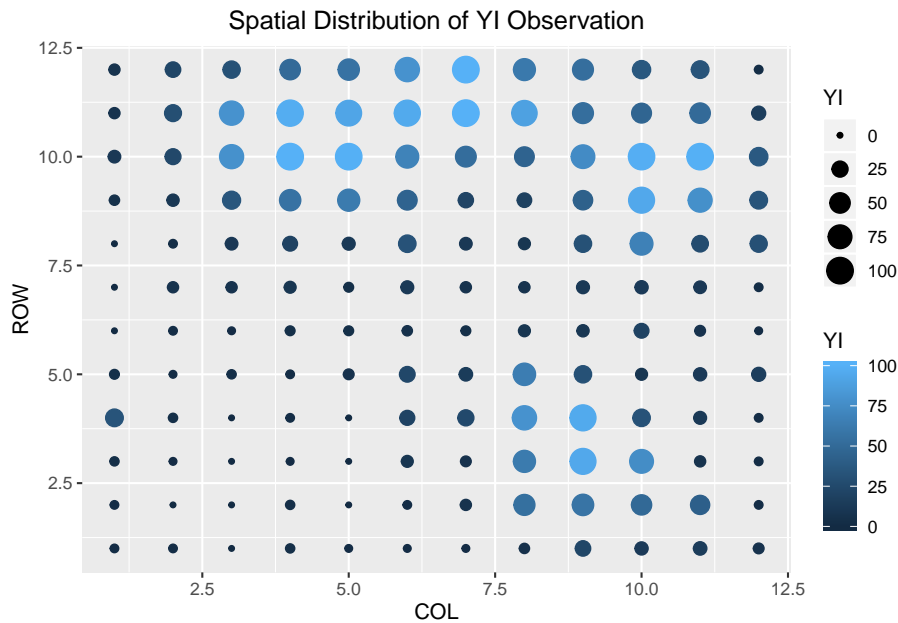
## Parsed with column specification:
## cols(
##   COL = col_double(),
##   ROW = col_double(),
##   YI = col_double()
## )

# print the data
a
```

```
## # A tibble: 144 x 3
##   COL ROW  YI
##   <dbl> <dbl> <dbl>
## 1     1     1     2
## 2     2     1     2
## 3     3     1     0
## 4     4     1     3
## 5     5     1     1
## 6     6     1     1
## 7     7     1     1
## 8     8     1     5
## 9     9     1    22
## 10    10     1    13
## # ... with 134 more rows
```

4.3 Autocorrelation statistics

```
# visualize the data
ggplot(data = a) +
  geom_point(mapping = aes(x = COL, y = ROW, size = YI, color = YI)) +
  ggtitle("Spatial Distribution of YI Observation") +
  theme(plot.title = element_text(hjust = 0.5))
```



```
# calculate Moran's I
Coords <- a %>%
  dplyr::select(COL, ROW)

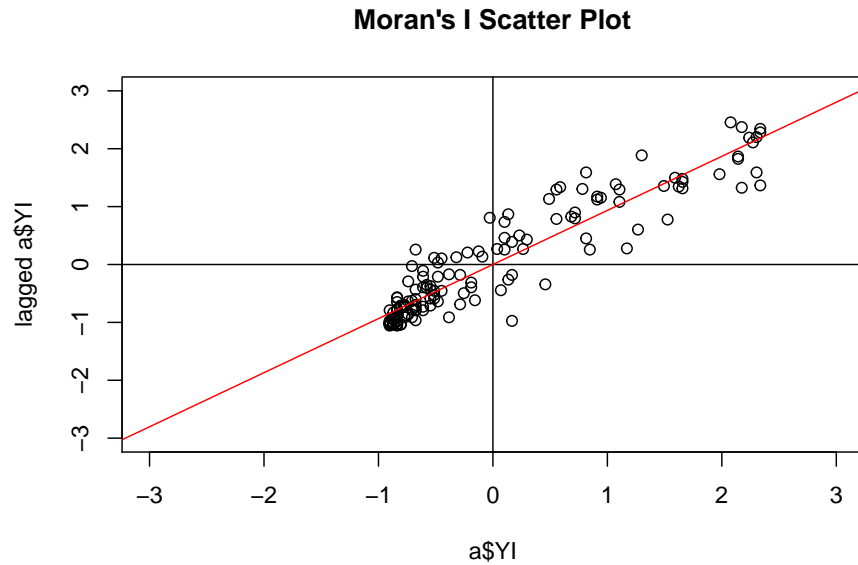
mI <- moransI(Coords, Bandwidth = 1, a$YI)

# print Moran's I table
moran.table <- tribble(
  ~`Moran's I`, ~`Expected I`, ~`Z randomization`, ~`P value randomization`,
  #-----/-----/-----/-----
  mI$Morans.I, mI$Expected.I, mI$z.randomization, mI$p.value.randomization
)

moran.table
```

```
## # A tibble: 1 x 4
##   `Moran's I` `Expected I` `Z randomization` `P value randomization`
##   <dbl>      <dbl>      <dbl>      <dbl>
## 1    0.782    -0.00699      13.0      1.28e-38
```

```
# create Moran's I scatter plot
l.moran <- l.moransI(Coords, Bandwidth = 1, a$YI)
```



```
# calculate geary's c
Coords_num <- coordinates(Coords)

# create an object of class 'nb' so that it can be used with function from package 'sp'
Coords_nb <- knn2nb(knearneigh(Coords_num))

# create a 'listw' object for use in the function 'geary.test'
coords_listw <- nb2listw(Coords_nb)

gearyC <- geary.test(a$YI, coords_listw, alternative = "two.sided")
gearyC
```

```
##
## Geary C test under randomisation
##
## data: a$YI
## weights: coords_listw
##
## Geary C statistic standard deviate = 8.8657, p-value < 2.2e-16
## alternative hypothesis: two.sided
## sample estimates:
## Geary C statistic      Expectation      Variance
##      0.235058006      1.000000000      0.007444457
```

4.4 First variogram

We will use the package `geoR` to construct empirical variogram, and then draw them using package `ggplot2`.

```
v1 <- variog(coords = Coords_num, data = a$YI, breaks = seq(0.5, 15.5),
             max.dist = 12)
```

```
## variog: computing omnidirectional variogram
```

```
# extract data from object v1 for plotting
v1_plot_data <- cbind(v1$u, v1$v, v1$n) %>%
  as.data.frame() %>%
  dplyr::rename(Distance = V1,
                Semivariance = V2,
                Pair_count = V3)

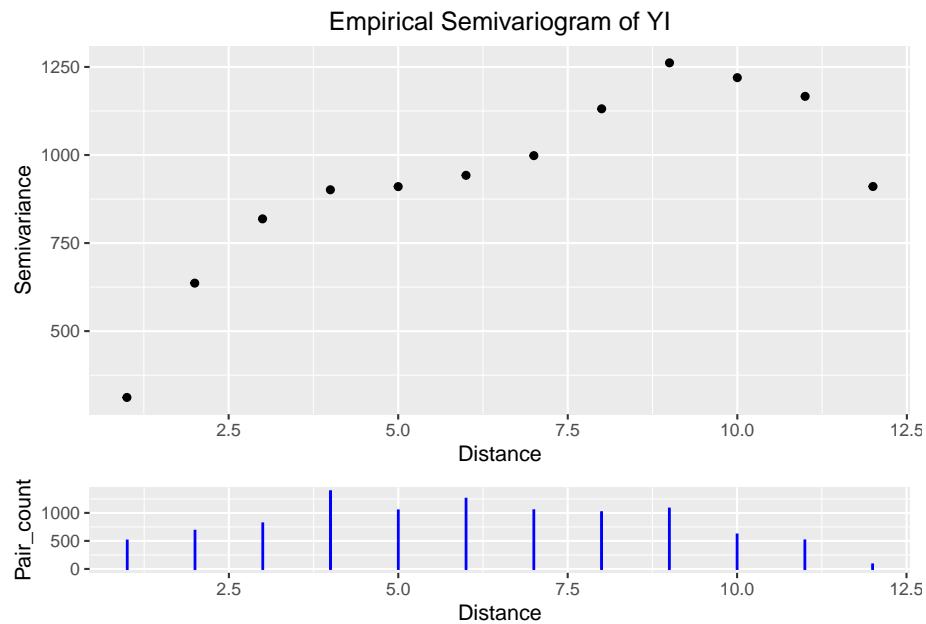
# in the table below, gamma is semivariance
v1_plot_data
```

##	Distance	Semivariance	Pair_count
## 1	1	311.7154	506
## 2	2	636.2074	680
## 3	3	818.7044	812
## 4	4	901.3218	1386
## 5	5	910.2773	1044
## 6	6	942.3219	1252
## 7	7	998.2290	1046
## 8	8	1131.2105	1012
## 9	9	1261.6817	1076
## 10	10	1219.6067	614
## 11	11	1166.5541	508
## 12	12	910.6250	80

```
# plot variogram
v1_plot_vario <- ggplot(data = v1_plot_data) +
  geom_point(mapping = aes(x = Distance, y = Semivariance)) +
  ggtitle("Empirical Semivariogram of YI") +
  theme(plot.title = element_text(hjust = 0.5))

# plot pair counts
v1_plot_pair_count <- ggplot(data = v1_plot_data) +
  geom_col(mapping = aes(x = Distance, y = Pair_count), width = 0.01, color = "blue")
```

```
# stack two plots
grid.arrange(v1_plot_vario, v1_plot_pair_count,
             ncol = 1, heights = c(3, 1))
```



4.5 Second variogram

Plot robust and classical variogram together.

```
# fit robust variogram
v1_robust <- variog(coords = Coords_num, data = a$YI, breaks = seq(0.5, 15.5),
                  max.dist = 12, estimator.type = "modulus")
```

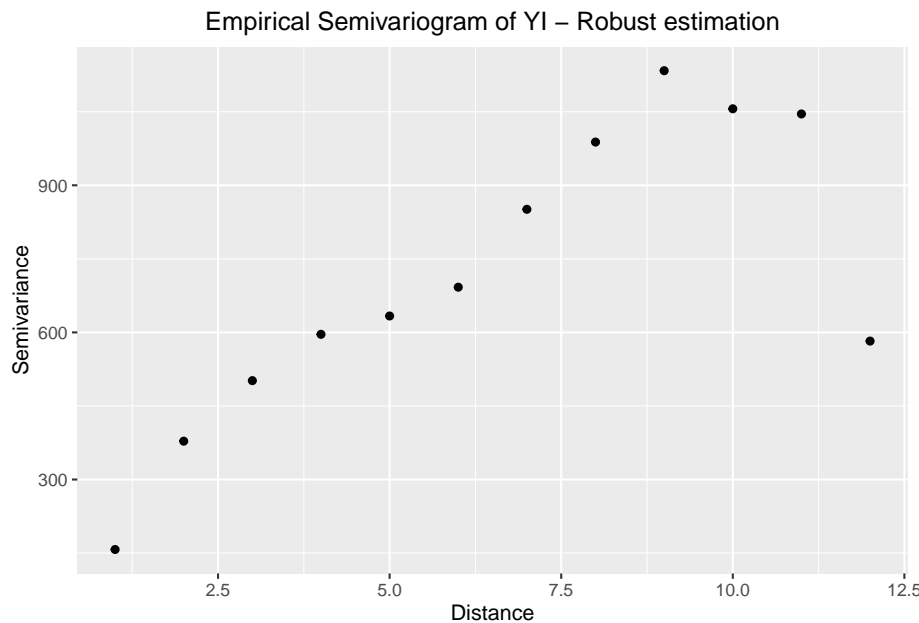
```
## variog: computing omnidirectional variogram
```

```
# extract the data
v1_robust_data <- cbind(v1_robust$u, v1_robust$v, v1_robust$n) %>%
  as.data.frame() %>%
  dplyr::rename(Distance = V1,
                Semivariance = V2,
                Pair_count = V3)
```



```
# plot robust variogram
v1_robust_vario <- ggplot(data = v1_robust_data) +
  geom_point(mapping = aes(x = Distance, y = Semivariance)) +
  ggtitle("Empirical Semivariogram of YI - Robust estimation") +
  theme(plot.title = element_text(hjust = 0.5))

v1_robust_vario
```

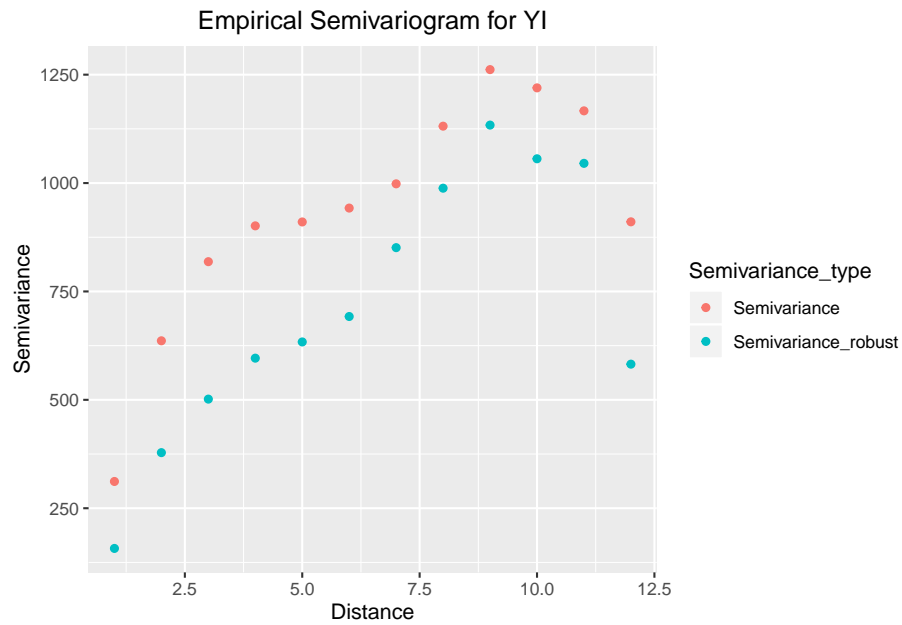


```
# combine robust and classical variogram
var_comb <- v1_robust_data %>%

  # combine robust and classical variogram datasets
  dplyr::rename(Semivariance_robust = Semivariance) %>%
  bind_cols(dplyr::select(v1_plot_data, Semivariance)) %>%
  gather(key = "Semivariance_type", value = "Semivariance", -c(Distance, Pair_count)) %>%

  # plot
  ggplot() +
  geom_point(mapping = aes(x = Distance, y = Semivariance, color = Semivariance_type)) +
  ggtitle("Empirical Semivariogram for YI") +
  theme(plot.title = element_text(hjust = 0.5))

var_comb
```



4.6 Variogram model selection

We will use the package `gstat` and `automap` for variogram model selection

```
# specify coordinates in the dataset
coordinates(a) = ~COL+ROW

# select the best model out of exponential, spherical, and gaussian
autofitVariogram(YI ~ COL + ROW, a, model = c("Sph", "Exp", "Gau"), cutoff = 12)
```

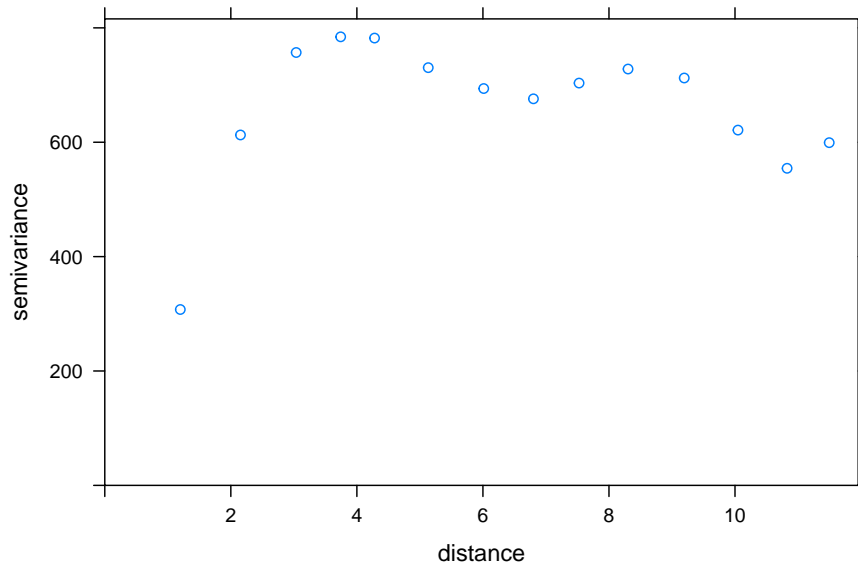
```
## $exp_var
##      np      dist      gamma dir.hor dir.ver  id
## 1  264  1.000000  233.2400      0      0 var1
## 2  242  1.414214  388.5222      0      0 var1
## 3  680  2.152750  612.6985      0      0 var1
## 4  812  3.036881  756.8971      0      0 var1
## 5 1066  3.944315  783.1461      0      0 var1
## 6 1364  4.977586  742.6252      0      0 var1
##
## $var_model
##      model      psill      range
## 1  Nug     0.0000  0.000000
## 2  Sph    782.9935  4.019145
```

```
##
## $sserr
## [1] 1247749
##
## attr("class")
## [1] "autofitVariogram" "list"
```

```
# fit empirical variogram
v_emp <- variogram(YI ~ COL + ROW, data = a, cutoff = 12)
v_emp
```

```
##      np      dist      gamma dir.hor dir.ver  id
## 1   506  1.198102 307.5054      0      0 var1
## 2   680  2.152750 612.6985      0      0 var1
## 3   812  3.036881 756.8971      0      0 var1
## 4   552  3.742751 784.3027      0      0 var1
## 5   834  4.280245 782.1560      0      0 var1
## 6  1044  5.132514 730.3844      0      0 var1
## 7  1028  6.012860 693.9058      0      0 var1
## 8   878  6.801676 675.9157      0      0 var1
## 9   836  7.525735 703.4337      0      0 var1
## 10  852  8.302717 728.0099      0      0 var1
## 11  792  9.194510 712.3311      0      0 var1
## 12  542 10.047104 621.2100      0      0 var1
## 13  452 10.826377 554.3985      0      0 var1
## 14  208 11.494850 599.2237      0      0 var1
```

```
plot(v_emp)
```



```
# fit exponential variogram
v_exp <- fit.variogram(v_emp, vgm("Exp"))
```

```
# fit spherical and gaussian
v_sph <- fit.variogram(v_emp, vgm("Sph"))
v_sph
```

```
##  model    psill    range
## 1  Nug     0.0000 0.000000
## 2  Sph    745.8602 3.765221
```

```
v_gau <- fit.variogram(v_emp, vgm("Gau"))
```

```
## Warning in fit.variogram(v_emp, vgm("Gau")): No convergence after 200
## iterations: try different initial values?
```

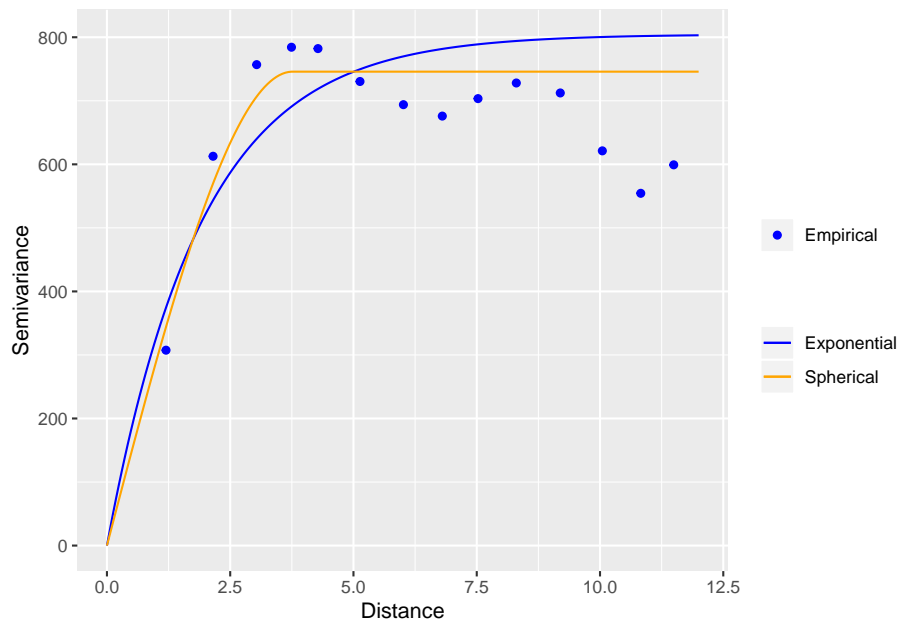
```
# extract plotting data from fitted variograms
v_exp_line <- variogramLine(v_exp, maxdist = 12)
v_sph_line <- variogramLine(v_sph, maxdist = 12)
# v_gau_line <- variogramLine(v_gau, maxdist = 12)

# plot empirical and fitted variograms together
# specify color for legends
```

```

legend_color <- c("Empirical" = "blue", "Exponential" = "blue",
                  "Spherical" = "orange")
ggplot(data = v_emp) +
  geom_point(mapping = aes(x = dist, y = gamma, fill = "Empirical"), color = "blue") +
  geom_line(data = v_exp_line, mapping = aes(x = dist, y = gamma, color = "Exponential")) +
  geom_line(data = v_sph_line, mapping = aes(x = dist, y = gamma, color = "Spherical")) +
  # geom_line(data = v_gau_line, mapping = aes(x = dist, y = gamma, color = "Gaussian")) +
  scale_color_manual(name = "", values = legend_color) +
  scale_fill_manual(name = "", values = legend_color) +
  labs(x = "Distance",
       y = "Semivariance")

```



Chapter 5

Yield loss

5.1 Load packages

Here is the R code to download the required packages for this exercise.

```
# install package manager 'pacman'  
if (!require(pacman)){  
  install.packages('pacman')  
}
```

Loading required package: pacman

```
# load packages needed for this exercise  
pacman::p_load(tidyverse,  
               nlme,  
               emmeans,  
               predictmeans  
)
```

5.2 Data

This is equivalent to the data step in SAS. Here, the data is imported from a file `yield_loss.csv` using the function `read_csv`. This function will download the data file directly from here.

```
# Import data  
a <- read_csv("https://raw.githubusercontent.com/luckymehra/epidem-exercises/master/data/yield_loss.csv")
```

```
## Parsed with column specification:
## cols(
##   WP = col_double(),
##   SP = col_character(),
##   BLK = col_double(),
##   TRT = col_double(),
##   FUNG = col_double(),
##   DS = col_double(),
##   YIELD = col_double()
## )
```

```
# print the data
a
```

```
## # A tibble: 24 x 7
##       WP SP      BLK TRT FUNG    DS YIELD
##   <dbl> <chr> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1  101 A      1     1     0    43   205
## 2  101 B      1     1     1     1   399
## 3  102 A      1     2     1     2   426
## 4  102 B      1     2     0    92   102
## 5  103 A      1     3     1     2   385
## 6  103 B      1     3     0     7   355
## 7  201 A      2     2     1     4   412
## 8  201 B      2     2     0    75   224
## 9  202 A      2     3     1     3   425
## 10 202 B      2     3     0    10   352
## # ... with 14 more rows
```

```
# specify that FUNG, TRT, and BLK are factors
a$FUNG <- as.ordered(as.factor(a$FUNG))
a$TRT <- as.ordered(as.factor(a$TRT))
a$BLK <- as.ordered(as.factor(a$BLK))
```

5.3 Mixed model for response variable DS

```
# fit the model
mm_1 <- lme(DS ~ TRT*FUNG, # fixed effects
            data = a,
            random = ~1|BLK/TRT) # read mm_1 as mixed model 1

# summary output
summary(mm_1)
```



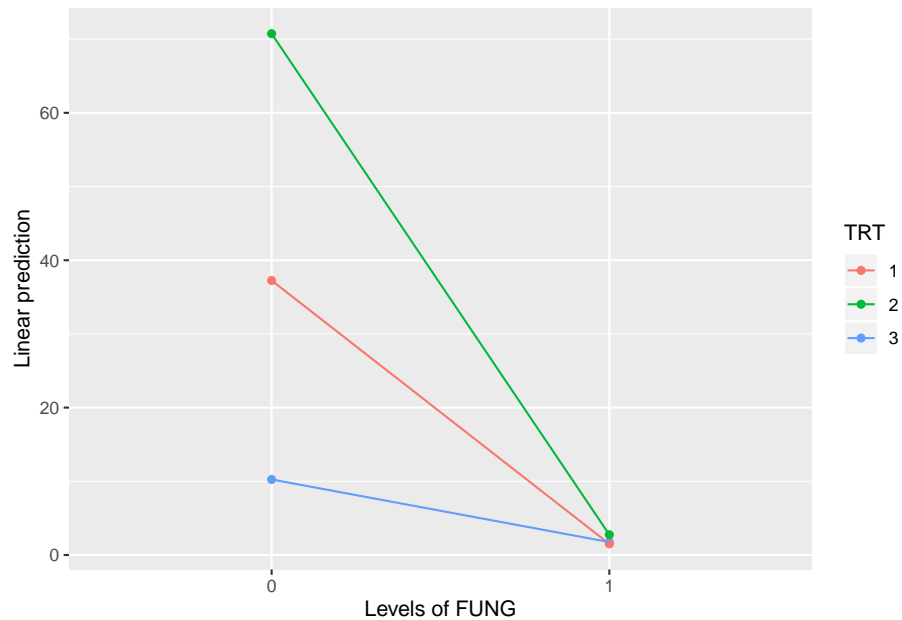
```
## Linear mixed-effects model fit by REML
## Data: a
##      AIC      BIC    logLik
## 156.1691 164.1825 -69.08456
##
## Random effects:
## Formula: ~1 | BLK
##      (Intercept)
## StdDev: 0.0009561632
##
## Formula: ~1 | TRT %in% BLK
##      (Intercept) Residual
## StdDev: 0.001007113 7.918859
##
## Fixed effects: DS ~ TRT * FUNG
##              Value Std.Error DF   t-value p-value
## (Intercept) 20.708333  1.616430  9 12.811150  0.0000
## TRT.L       -9.457553  2.799740  6  -3.378012  0.0149
## TRT.Q       -19.646949  2.799740  6  -7.017420  0.0004
## FUNG.L      -26.457579  2.285978  9 -11.573857  0.0000
## TRT.L:FUNG.L 13.625000  3.959430  9   3.441152  0.0074
## TRT.Q:FUNG.L 26.485944  3.959430  9   6.689333  0.0001
## Correlation:
##      (Intr) TRT.L TRT.Q FUNG.L TRT.L:
## TRT.L      0
## TRT.Q      0      0
## FUNG.L      0      0      0
## TRT.L:FUNG.L 0      0      0      0
## TRT.Q:FUNG.L 0      0      0      0      0
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -2.241484e+00 -9.471064e-02 -1.590814e-08  1.736361e-01  2.683467e+00
##
## Number of Observations: 24
## Number of Groups:
##      BLK TRT %in% BLK
##      4      12
```

```
# type 3 tests of fixed effects
anova(mm_1)
```

```
##      numDF denDF   F-value p-value
## (Intercept)    1     9 164.12557 <.0001
## TRT           2     6  30.32757  7e-04
```

```
## FUNG          1      9 133.95416 <.0001
## TRT:FUNG      2      9  28.29435  1e-04
```

```
# visualize interaction
emmip(mm_1, TRT ~ FUNG)
```



```
# to do anova for random effects, we need to compare mm_1 with a model that only has f
# we can use `gls()` function in `nlme` to fit the fixed effects model
fixed_model <- gls(DS ~ TRT * FUNG,
                   data = a)
```

```
# test the random effects in the model
anova(mm_1, fixed_model)
```

```
##          Model df      AIC      BIC    logLik  Test      L.Ratio
## mm_1          1  9 156.1691 164.1825 -69.08456
## fixed_model    2  7 152.1691 158.4017 -69.08456 1 vs 2 1.250038e-08
##          p-value
## mm_1
## fixed_model      1
```

```
# least square means
test(emmeans(mm_1, "TRT"))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## TRT emmean SE df t.ratio p.value
## 1      19.4 2.8 3  6.920  0.0062
## 2      36.8 2.8 3 13.126  0.0010
## 3       6.0 2.8 3  2.143  0.1215
##
## Results are averaged over the levels of: FUNG
## d.f. method: containment
```

```
test(emmeans(mm_1, "FUNG"))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## FUNG emmean SE df t.ratio p.value
## 0      39.4 2.29 3 17.243  0.0004
## 1       2.0 2.29 3  0.875  0.4460
##
## Results are averaged over the levels of: TRT
## d.f. method: containment
```

```
# pairwise difference
test(emmeans(mm_1, pairwise ~ TRT), adjust = "none")
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## $emmeans
## TRT emmean SE df t.ratio p.value
## 1      19.4 2.8 3  6.920  0.0062
## 2      36.8 2.8 3 13.126  0.0010
## 3       6.0 2.8 3  2.143  0.1215
##
## Results are averaged over the levels of: FUNG
## d.f. method: containment
##
## $contrasts
## contrast estimate SE df t.ratio p.value
## 1 - 2      -17.4 3.96 6 -4.388  0.0046
## 1 - 3       13.4 3.96 6  3.378  0.0149
## 2 - 3       30.8 3.96 6  7.766  0.0002
##
## Results are averaged over the levels of: FUNG
```

```
test(emmeans(mm_1, pairwise ~ FUNG))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## $emmeans
##   FUNG emmean    SE df t.ratio p.value
## 0      39.4 2.29 3 17.243 0.0004
## 1       2.0 2.29 3 0.875 0.4460
##
## Results are averaged over the levels of: TRT
## d.f. method: containment
##
## $contrasts
## contrast estimate    SE df t.ratio p.value
## 0 - 1          37.4 3.23 9 11.574 <.0001
##
## Results are averaged over the levels of: TRT
```

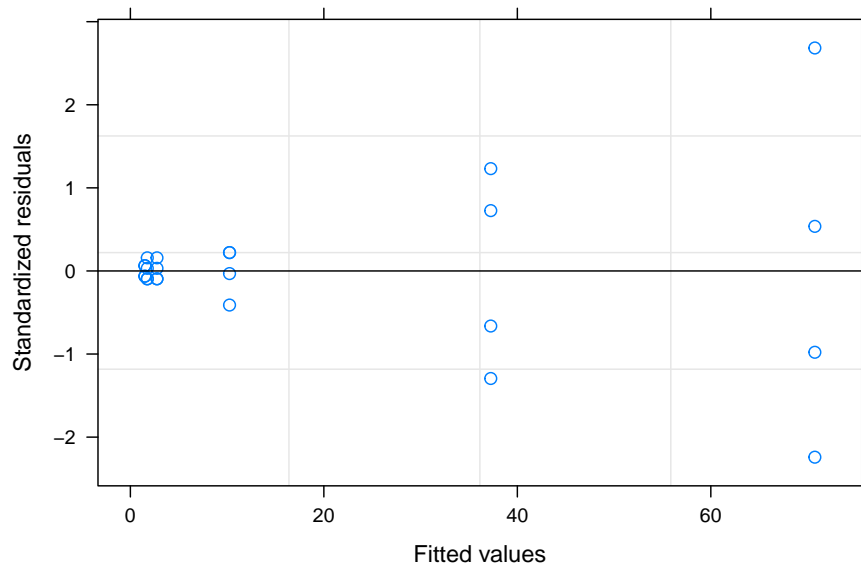
```
test(emmeans(mm_1, pairwise ~ TRT*FUNG), adjust = "none")
```

```
## $emmeans
## TRT FUNG emmean    SE df t.ratio p.value
## 1 0      37.25 3.96 3 9.408 0.0025
## 2 0      70.75 3.96 3 17.869 0.0004
## 3 0      10.25 3.96 3 2.589 0.0812
## 1 1       1.50 3.96 3 0.379 0.7300
## 2 1       2.75 3.96 3 0.695 0.5373
## 3 1       1.75 3.96 3 0.442 0.6884
##
## d.f. method: containment
##
## $contrasts
## contrast estimate    SE df t.ratio p.value
## 1,0 - 2,0    -33.50 5.6 6 -5.983 0.0010
## 1,0 - 3,0     27.00 5.6 6 4.822 0.0029
## 1,0 - 1,1     35.75 5.6 9 6.385 0.0001
## 1,0 - 2,1     34.50 5.6 6 6.161 0.0008
## 1,0 - 3,1     35.50 5.6 6 6.340 0.0007
## 2,0 - 3,0     60.50 5.6 6 10.805 <.0001
## 2,0 - 1,1     69.25 5.6 6 12.367 <.0001
## 2,0 - 2,1     68.00 5.6 9 12.144 <.0001
## 2,0 - 3,1     69.00 5.6 6 12.323 <.0001
## 3,0 - 1,1      8.75 5.6 6 1.563 0.1692
## 3,0 - 2,1      7.50 5.6 6 1.339 0.2289
```

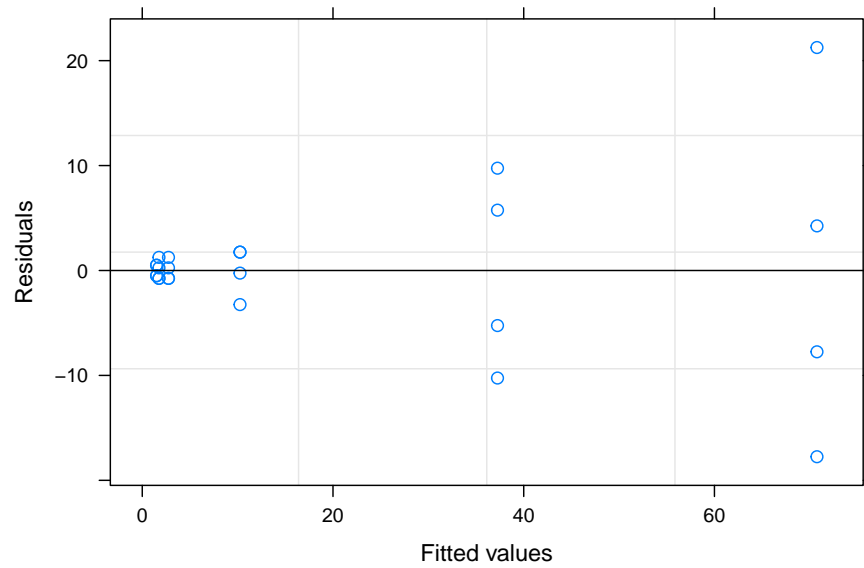
```
## 3,0 - 3,1      8.50 5.6  9  1.518  0.1633
## 1,1 - 2,1     -1.25 5.6  6 -0.223  0.8308
## 1,1 - 3,1     -0.25 5.6  6 -0.045  0.9658
## 2,1 - 3,1      1.00 5.6  6  0.179  0.8641
```

5.3.1 Diagnostic plots

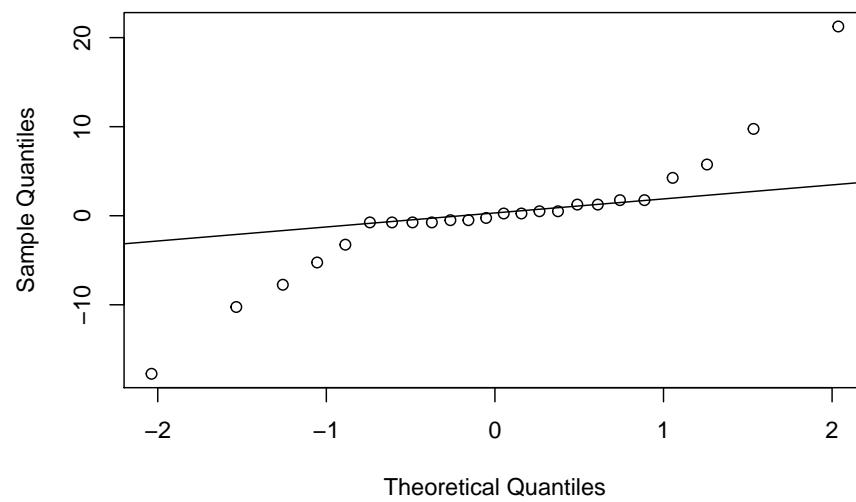
```
# pearson residuals vs. fitted values
plot(mm_1, resid(., type="pearson") ~ fitted(.), abline = 0)
```



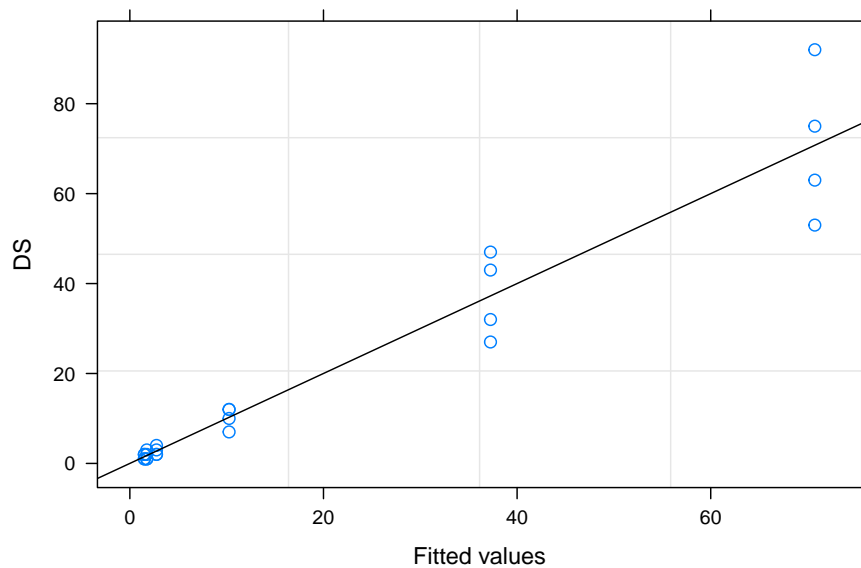
```
# standardized residuals vs. fitted values
plot(mm_1, resid(., scaled=TRUE) ~ fitted(.), abline = 0)
```



```
# qq plot  
qqnorm(residuals(mm_1))  
qqline(residuals(mm_1))
```

Normal Q-Q Plot

```
#observed vs. fitted values
plot(mm_1, DS ~ fitted(.), abline = c(0,1))
```



5.4 Mixed model for response variable YIELD

```
# fit the model
mm_2 <- lme(YIELD ~ TRT*FUNG, # fixed effects
            data = a,
            random = ~1|BLK/TRT) # read mm_2 as mixed model 2
```

```
# summary output
summary(mm_2)
```

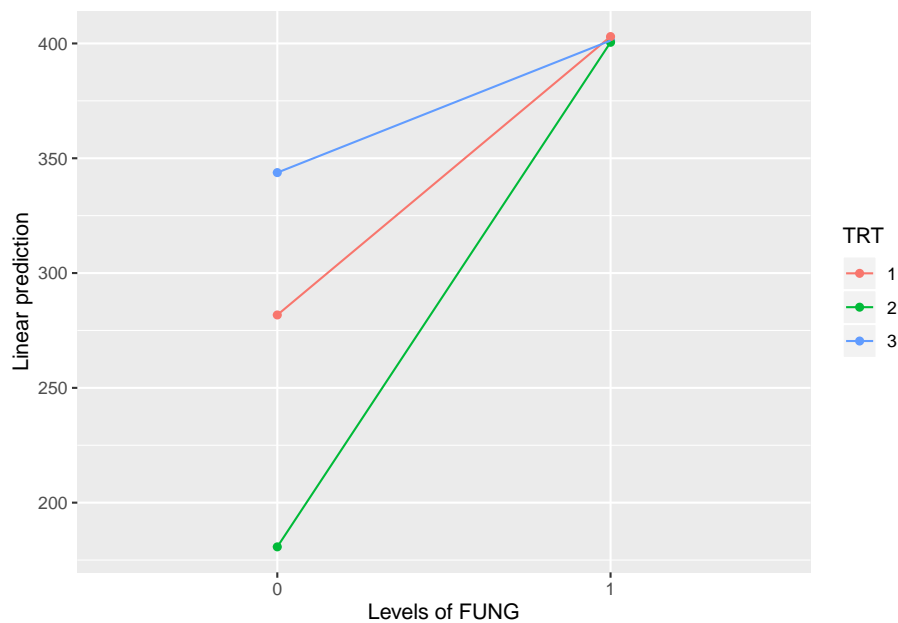
```
## Linear mixed-effects model fit by REML
## Data: a
##      AIC      BIC   logLik
## 209.9214 217.9348 -95.9607
##
## Random effects:
## Formula: ~1 | BLK
##      (Intercept)
```

```
## StdDev:    11.99815
##
## Formula: ~1 | TRT %in% BLK
##           (Intercept) Residual
## StdDev: 0.001914579 33.61779
##
## Fixed effects: YIELD ~ TRT * FUNG
##              Value Std.Error DF   t-value p-value
## (Intercept) 335.1667  9.114752  9 36.77189  0.0000
## TRT.L       21.3016 11.885682  6  1.79221  0.1233
## TRT.Q       54.5522 11.885682  6  4.58974  0.0037
## FUNG.L      93.9274  9.704619  9  9.67862  0.0000
## TRT.L:FUNG.L -31.8750 16.808893  9 -1.89632  0.0904
## TRT.Q:FUNG.L -75.2720 16.808893  9 -4.47811  0.0015
## Correlation:
##              (Intr) TRT.L TRT.Q FUNG.L TRT.L:
## TRT.L         0
## TRT.Q         0      0
## FUNG.L         0      0      0
## TRT.L:FUNG.L  0      0      0      0
## TRT.Q:FUNG.L  0      0      0      0      0
##
## Standardized Within-Group Residuals:
##              Min      Q1      Med      Q3      Max
## -2.04399017 -0.33265713  0.07191314  0.49972251  1.20545632
##
## Number of Observations: 24
## Number of Groups:
##              BLK TRT %in% BLK
##              4      12
```

```
# type 3 tests of fixed effects
anova(mm_2)
```

```
##              numDF denDF   F-value p-value
## (Intercept)      1     9 1352.1720 <.0001
## TRT             2     6  12.1389  0.0078
## FUNG            1     9  93.6757 <.0001
## TRT:FUNG        2     9  11.8247  0.0030
```

```
# visualize interaction
emmip(mm_2, TRT ~ FUNG)
```

```
# to do anova for random effects, we need to compare mm_1 with a model that only has fixed effects
# we can use `glsl()` function in `nlme` to fit the fixed effects model
fixed_model_YIELD <- glsl(YIELD ~ TRT * FUNG,
                           data = a)
```

```
# test the random effects in the model
anova(mm_2, fixed_model_YIELD)
```

```
##           Model df      AIC      BIC    logLik  Test  L.Ratio
## mm_2           1  9 209.9214 217.9348 -95.96070
## fixed_model_YIELD  2  7 206.3763 212.6089 -96.18815 1 vs 2 0.4548877
##           p-value
## mm_2
## fixed_model_YIELD 0.7966
```

```
# least square means
test(emmeans(mm_2, "TRT"))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## TRT emmean SE df t.ratio p.value
## 1      342 13.3  3 25.716 0.0001
## 2      291 13.3  3 21.829 0.0002
```

```
## 3      372 13.3  3 27.978  0.0001
```

```
##
```

```
## Results are averaged over the levels of: FUNG
```

```
## d.f. method: containment
```

```
test(emmeans(mm_2, "FUNG"))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## FUNG emmean    SE df t.ratio p.value
```

```
## 0      269 11.4  3 23.556  0.0002
```

```
## 1      402 11.4  3 35.198  0.0001
```

```
##
```

```
## Results are averaged over the levels of: TRT
```

```
## d.f. method: containment
```

```
# pairwise difference
```

```
test(emmeans(mm_2, pairwise ~ TRT), adjust = "none")
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

```
## $emmeans
```

```
## TRT emmean    SE df t.ratio p.value
```

```
## 1      342 13.3  3 25.716  0.0001
```

```
## 2      291 13.3  3 21.829  0.0002
```

```
## 3      372 13.3  3 27.978  0.0001
```

```
##
```

```
## Results are averaged over the levels of: FUNG
```

```
## d.f. method: containment
```

```
##
```

```
## $contrasts
```

```
## contrast estimate    SE df t.ratio p.value
```

```
## 1 - 2      51.8 16.8  6  3.079  0.0217
```

```
## 1 - 3     -30.1 16.8  6 -1.792  0.1233
```

```
## 2 - 3     -81.9 16.8  6 -4.871  0.0028
```

```
##
```

```
## Results are averaged over the levels of: FUNG
```

```
test(emmeans(mm_2, pairwise ~ FUNG))
```

```
## NOTE: Results may be misleading due to involvement in interactions
```

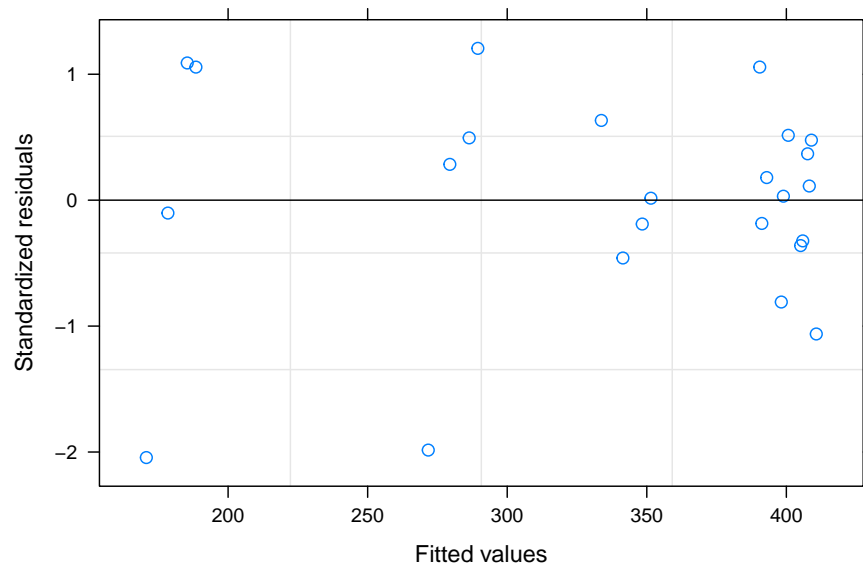
```
## $emmeans
##      FUNG emmean    SE df t.ratio p.value
##    0      269 11.4   3 23.556  0.0002
##    1      402 11.4   3 35.198  0.0001
##
## Results are averaged over the levels of: TRT
## d.f. method: containment
##
## $contrasts
## contrast estimate    SE df t.ratio p.value
## 0 - 1          -133 13.7   9 -9.679  <.0001
##
## Results are averaged over the levels of: TRT
```

```
test(emmeans(mm_2, pairwise ~ TRT*FUNG), adjust = "none")
```

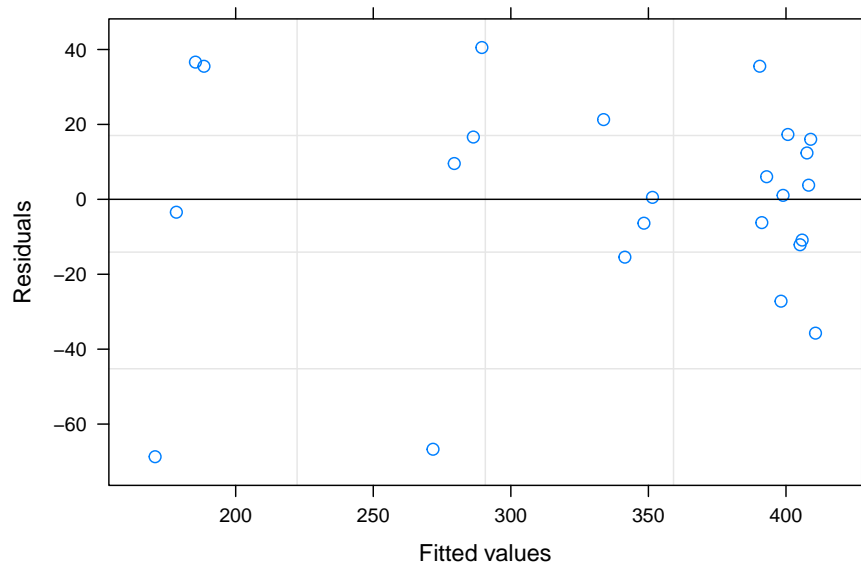
```
## $emmeans
##      TRT FUNG emmean    SE df t.ratio p.value
##    1    0      282 17.8   3 15.787  0.0006
##    2    0      181 17.8   3 10.128  0.0021
##    3    0      344 17.8   3 19.261  0.0003
##    1    1      403 17.8   3 22.580  0.0002
##    2    1      400 17.8   3 22.440  0.0002
##    3    1      401 17.8   3 22.482  0.0002
##
## d.f. method: containment
##
## $contrasts
## contrast estimate    SE df t.ratio p.value
## 1,0 - 2,0    101.00 23.8   6  4.249  0.0054
## 1,0 - 3,0    -62.00 23.8   6 -2.608  0.0402
## 1,0 - 1,1   -121.25 23.8   9 -5.101  0.0006
## 1,0 - 2,1   -118.75 23.8   6 -4.996  0.0025
## 1,0 - 3,1   -119.50 23.8   6 -5.027  0.0024
## 2,0 - 3,0   -163.00 23.8   6 -6.857  0.0005
## 2,0 - 1,1   -222.25 23.8   6 -9.349  0.0001
## 2,0 - 2,1   -219.75 23.8   9 -9.244  <.0001
## 2,0 - 3,1   -220.50 23.8   6 -9.276  0.0001
## 3,0 - 1,1    -59.25 23.8   6 -2.492  0.0470
## 3,0 - 2,1    -56.75 23.8   6 -2.387  0.0542
## 3,0 - 3,1    -57.50 23.8   9 -2.419  0.0387
## 1,1 - 2,1      2.50 23.8   6  0.105  0.9197
## 1,1 - 3,1      1.75 23.8   6  0.074  0.9437
## 2,1 - 3,1     -0.75 23.8   6 -0.032  0.9759
```

5.4.1 Diagnostic plots

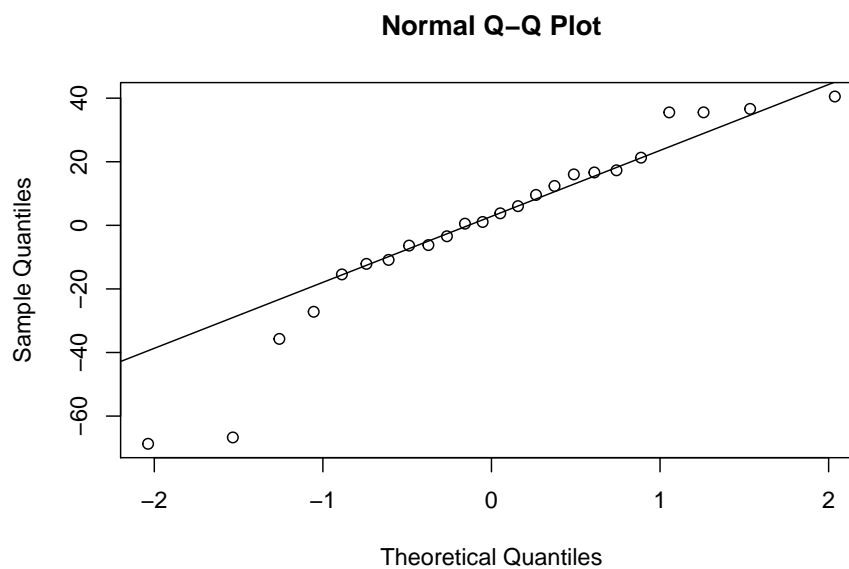
```
# pearson residuals vs. fitted values  
plot(mm_2, resid(., type="pearson") ~ fitted(.), abline = 0)
```



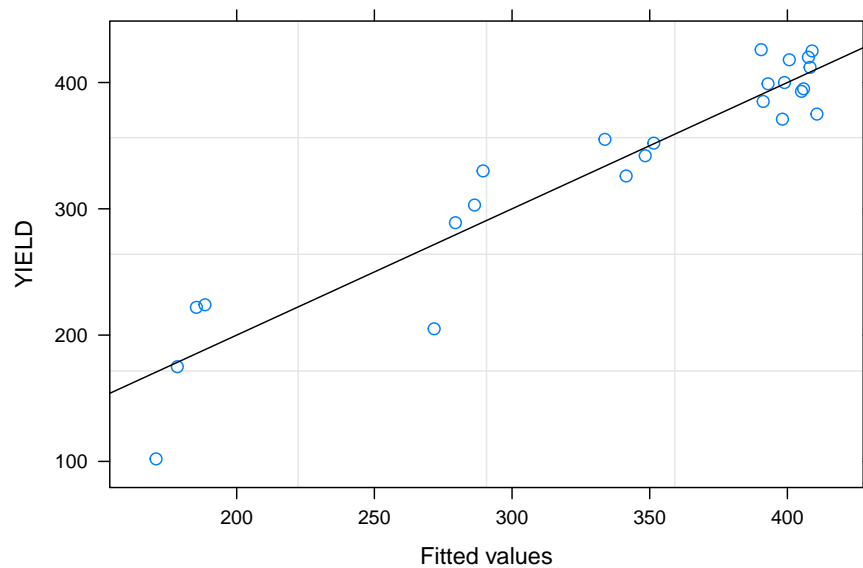
```
# standardaized residuals vs. fitted values  
plot(mm_2, resid(., scaled=TRUE) ~ fitted(.), abline = 0)
```



```
# qq plot  
qqnorm(residuals(mm_2))  
qqline(residuals(mm_2))
```



```
#observed vs. fitted values
plot(mm_2, YIELD ~ fitted(.), abline = c(0,1))
```



5.5 Linear regression between YIELD and DS

```
# fit `lm` model
lm_1 <- lm(YIELD ~ DS, data = a)
summary(lm_1)
```

```
##
## Call:
## lm(formula = YIELD ~ DS, data = a)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -61.196 -18.565   0.856  22.676  56.812
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  399.2384     8.0711   49.47  < 2e-16 ***
## DS          -3.0940     0.2399  -12.90 9.81e-12 ***
```

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 31.17 on 22 degrees of freedom
## Multiple R-squared:  0.8832, Adjusted R-squared:  0.8779
## F-statistic: 166.4 on 1 and 22 DF,  p-value: 9.809e-12
```

```
anova(lm_1)
```

```
## Analysis of Variance Table
##
## Response: YIELD
##           Df Sum Sq Mean Sq F value    Pr(>F)
## DS           1 161600  161600   166.38 9.809e-12 ***
## Residuals   22  21368     971
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# diagnostic plots
residplot(lm_1)
```

5.5.1 Linear regression between RY1 and DS

```
b <- a %>%
  mutate(RY1 = YIELD/399.23843)
```

```
# fit linear regression model
lm_2 <- lm(RY1 ~ DS, data = b)
summary(lm_2)
```

```
##
## Call:
## lm(formula = RY1 ~ DS, data = b)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.153282 -0.046502  0.002143  0.056798  0.142301
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  1.0000000  0.0202162   49.47  < 2e-16 ***
## DS          -0.0077498  0.0006008  -12.90 9.81e-12 ***
```

```
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.07806 on 22 degrees of freedom
## Multiple R-squared:  0.8832, Adjusted R-squared:  0.8779
## F-statistic: 166.4 on 1 and 22 DF,  p-value: 9.809e-12
```

```
anova(lm_2)
```

```
## Analysis of Variance Table
##
## Response: RY1
##           Df Sum Sq Mean Sq F value    Pr(>F)
## DS           1  1.01385   1.01385   166.38 9.809e-12 ***
## Residuals    22  0.13406   0.00609
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# diagnostic plots
residplot(lm_2)
```

5.5.2 Transform dataset a

```
a_yield <- a %>%
  dplyr::select(BLK, TRT, YIELD) %>%
  arrange(BLK, TRT, YIELD) %>%
  group_by(BLK, TRT) %>%
  summarise(RY2 = YIELD[1]/YIELD[2]) %>%
  ungroup()

a_ds <- a %>%
  dplyr::select(BLK, TRT, DS) %>%
  arrange(BLK, TRT, DS) %>%
  group_by(BLK, TRT) %>%
  summarise(CDS = DS[2]) %>%
  ungroup()

a_new <- a_yield %>%
  inner_join(a_ds) %>%
  ungroup() %>%
  mutate(BLK = parse_factor(as.character(BLK)),
         TRT = parse_factor(as.character(TRT)))
```



```
## Joining, by = c("BLK", "TRT")
```

```
# print the data
a_new
```

```
## # A tibble: 12 x 4
##   BLK   TRT   RY2   CDS
##   <fct> <fct> <dbl> <dbl>
## 1 1     1     0.514 43
## 2 1     2     0.239 92
## 3 1     3     0.922 7
## 4 2     1     0.88 27
## 5 2     2     0.544 75
## 6 2     3     0.828 10
## 7 3     1     0.721 47
## 8 3     2     0.565 63
## 9 3     3     0.866 12
## 10 4     1     0.691 32
## 11 4     2     0.472 53
## 12 4     3     0.815 12
```

5.6 Mixed model for RY2

```
# fit the model
mm_3 <- lme(RY2 ~ TRT, # fixed effects
            data = a_new,
            random = ~1|BLK) # read mm_3 as mixed model 3

# summary output
summary(mm_3)
```

```
## Linear mixed-effects model fit by REML
## Data: a_new
##      AIC      BIC    logLik
## 2.04077 3.026893 3.979615
##
## Random effects:
## Formula: ~1 | BLK
##      (Intercept) Residual
## StdDev: 0.05297273 0.1134963
##
## Fixed effects: RY2 ~ TRT
```

```
##              Value Std.Error DF   t-value p-value
## (Intercept)  0.7016501 0.06262490  6 11.204012  0.0000
## TRT2        -0.2467228 0.08025398  6 -3.074274  0.0218
## TRT3         0.1561339 0.08025398  6  1.945497  0.0997
## Correlation:
##      (Intr) TRT2
## TRT2 -0.641
## TRT3 -0.641  0.500
##
## Standardized Within-Group Residuals:
##      Min          Q1          Med          Q3          Max
## -1.50508335 -0.38516867 -0.01698779  0.58195830  1.29565814
##
## Number of Observations: 12
## Number of Groups: 4
```

```
# type 3 tests of fixed effects
anova(mm_3)
```

```
##              numDF denDF   F-value p-value
## (Intercept)      1     6 254.00333  <.0001
## TRT              2     6  12.81141  0.0068
```

```
# to do anova for random effects, we need to compare mm_1 with a model that only has f
# we can use `gls()` function in `nlme` to fit the fixed effects model
fixed_model_RY2 <- gls(RY2 ~ TRT,
                       data = a_new)
```

```
# test the random effects in the model
anova(mm_3, fixed_model_RY2)
```

```
##              Model df        AIC        BIC   logLik   Test   L.Ratio
## mm_3              1  5 2.0407705 3.026893 3.979615
## fixed_model_RY2   2  4 0.3057644 1.094663 3.847118 1 vs 2 0.2649939
##
##              p-value
## mm_3
## fixed_model_RY2  0.6067
```

```
# pairwise difference
test(emmeans(mm_3, pairwise ~ TRT), adjust = "none")
```

```
## $emmeans
## TRT emmean      SE df t.ratio p.value
## 1      0.702 0.0626  3 11.204  0.0015
```

```
## 2    0.455 0.0626  3  7.264  0.0054
## 3    0.858 0.0626  3 13.697  0.0008
##
## d.f. method: containment
##
## $contrasts
## contrast estimate      SE df t.ratio p.value
## 1 - 2          0.247 0.0803  6  3.074  0.0218
## 1 - 3         -0.156 0.0803  6 -1.945  0.0997
## 2 - 3         -0.403 0.0803  6 -5.020  0.0024
```

```
# diagnostic plots
residplot(mm_3)
```

5.7 Linear regression between RY2 and CDS

```
# fit linear regression model
lm_3 <- lm(RY2 ~ CDS, data = a_new)
summary(lm_3)
```

```
##
## Call:
## lm(formula = RY2 ~ CDS, data = a_new)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -0.13338 -0.05085 -0.01090  0.06530  0.12439
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.9386111  0.0467991  20.056 2.09e-09 ***
## CDS         -0.0067778  0.0009845  -6.884 4.28e-05 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.09061 on 10 degrees of freedom
## Multiple R-squared:  0.8258, Adjusted R-squared:  0.8083
## F-statistic: 47.39 on 1 and 10 DF,  p-value: 4.275e-05
```

```
anova(lm_3)
```

```
## Analysis of Variance Table
```

```
##
## Response: RY2
##           Df Sum Sq Mean Sq F value    Pr(>F)
## CDS         1 0.38914  0.38914   47.394 4.275e-05 ***
## Residuals  10 0.08211  0.00821
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
# diagnostic plots
residplot(lm_3)
```

Appendix A

SAS code

A.1 Exercise 4

Copy and paste the below code into a SAS editor, and hit run to see the output.

```
DATA A;
INPUT PLOT AN T BLK TRT PCTSEV;
Y=PCTSEV/100;
YSTAR=LOG(Y/(1-Y));
WT=Y*(1-Y);
DROP AN;
CARDS;
101 1 0 1 2 9
102 1 0 1 1 6
103 1 0 1 3 2
201 1 0 2 2 7
202 1 0 2 3 5
203 1 0 2 1 3
301 1 0 3 3 4
302 1 0 3 2 2
303 1 0 3 1 6
401 1 0 4 1 1
402 1 0 4 2 1
403 1 0 4 3 4
101 2 7 1 2 4
102 2 7 1 1 6
103 2 7 1 3 10
201 2 7 2 2 2
202 2 7 2 3 5
203 2 7 2 1 3
```

```

301 2 7 3 3 11
302 2 7 3 2 6
303 2 7 3 1 4
401 2 7 4 1 8
402 2 7 4 2 3
403 2 7 4 3 6
101 3 14 1 2 8
102 3 14 1 1 20
103 3 14 1 3 15
201 3 14 2 2 13
202 3 14 2 3 12
203 3 14 2 1 14
301 3 14 3 3 15
302 3 14 3 2 8
303 3 14 3 1 25
401 3 14 4 1 17
402 3 14 4 2 14
403 3 14 4 3 49
101 4 21 1 2 24
102 4 21 1 1 38
103 4 21 1 3 61
201 4 21 2 2 31
202 4 21 2 3 42
203 4 21 2 1 79
301 4 21 3 3 48
302 4 21 3 2 23
303 4 21 3 1 86
401 4 21 4 1 52
402 4 21 4 2 45
403 4 21 4 3 56
101 5 28 1 2 28
102 5 28 1 1 89
103 5 28 1 3 44
201 5 28 2 2 41
202 5 28 2 3 49
203 5 28 2 1 79
301 5 28 3 3 45
302 5 28 3 2 47
303 5 28 3 1 63
401 5 28 4 1 94
402 5 28 4 2 52
403 5 28 4 3 64
101 6 35 1 2 36
102 6 35 1 1 77
103 6 35 1 3 88

```

```

201 6    35  2  2  42
202 6    35  2  3  69
203 6    35  2  1  71
301 6    35  3  3  43
302 6    35  3  2  39
303 6    35  3  1  84
401 6    35  4  1  97
402 6    35  4  2  47
403 6    35  4  3  76
;
PROC MIXED DATA=A COVTEST;
CLASS BLK TRT;
MODEL YSTAR=TRT|T/ SOLUTION DDFM=bw RESIDUAL;
RANDOM BLK;
WEIGHT WT;
REPEATED/SUBJECT=BLK*TRT TYPE=AR(1) R RCORR;
quit;

PROC MIXED DATA=A;
CLASS BLK TRT;
MODEL YSTAR=TRT TRT*T/NOINT SOLUTION DDFM=bw OUTPM=B;
RANDOM BLK;
WEIGHT WT;
REPEATED/SUBJECT=BLK*TRT TYPE=AR(1);
LSMEANS TRT/DIFF AT T=0;
LSMEANS TRT/DIFF AT T=7;
LSMEANS TRT/DIFF AT T=14;
LSMEANS TRT/DIFF AT T=21;
LSMEANS TRT/DIFF AT T=28;
LSMEANS TRT/DIFF AT T=35;

ESTIMATE 'TRT1 S VS TRT2 S' TRT*T 1 -1 0;
ESTIMATE 'TRT1 S VS TRT3 S' TRT*T 1 0 -1;
ESTIMATE 'TRT2 S VS TRT3 S' TRT*T 0 1 -1;
quit;

PROC PRINT DATA=B;

PROC REG DATA=B;
MODEL YSTAR=PRED;

RUN;

```

A.2 Exercise 9.4

Copy and paste the below code into a SAS editor, and hit run to see the output.

```
DATA A;
INPUT I YI;
EAST=1;
NORTH=I;
CARDS;
1 41
2 60
3 81
4 22
5 8
6 20
7 28
8 2
9 0
10 2
11 2
12 8
13 0
14 43
15 61
16 50
;
PROC VARIOGRAM PLOTS=MORAN OUTVAR=B;
COMPUTE LAGD=1 MAXLAG=11 AUTOCORR(ASSUM=RANDOM);
COORDINATES XC=EAST YC=NORTH;
VAR YI;

PROC PRINT;
run;

PROC VARIOGRAM DATA=A PLOTS=FIT;
COMPUTE LAGD=1 MAXLAG=11 CL ROBUST;
COORDINATES XC=EAST YC=NORTH;
MODEL FORM=AUTO(MLIST=(SPH EXP GAU) NEST=1);
VAR YI;

RUN;
```


A.3 Exercise 9.5

Copy and paste the below code into a SAS editor, and hit run to see the output.

```
DATA A;  
INPUT COL ROW YI;  
CARDS;  
1 1 2  
2 1 2  
3 1 0  
4 1 3  
5 1 1  
6 1 1  
7 1 1  
8 1 5  
9 1 22  
10 1 13  
11 1 14  
12 1 6  
1 2 2  
2 2 0  
3 2 0  
4 2 3  
5 2 0  
6 2 2  
7 2 7  
8 2 54  
9 2 57  
10 2 49  
11 2 42  
12 2 2  
1 3 3  
2 3 1  
3 3 0  
4 3 1  
5 3 0  
6 3 9  
7 3 6  
8 3 62  
9 3 94  
10 3 75  
11 3 7  
12 3 2  
1 4 33  
2 4 3
```

```
3 4 0
4 4 2
5 4 0
6 4 20
7 4 25
8 4 79
9 4 95
10 4 32
11 4 12
12 4 2
1 5 4
2 5 1
3 5 3
4 5 2
5 5 6
6 5 23
7 5 14
8 5 64
9 5 31
10 5 9
11 5 13
12 5 16
1 6 0
2 6 2
3 6 1
4 6 4
5 6 4
6 6 5
7 6 4
8 6 9
9 6 10
10 6 19
11 6 6
12 6 1
1 7 0
2 7 7
3 7 7
4 7 9
5 7 4
6 7 12
7 7 7
8 7 7
9 7 12
10 7 13
11 7 11
```

12	7	2
1	8	0
2	8	2
3	8	11
4	8	19
5	8	12
6	8	32
7	8	11
8	8	9
9	8	31
10	8	67
11	8	27
12	8	30
1	9	5
2	9	10
3	9	35
4	9	56
5	9	62
6	9	45
7	9	21
8	9	18
9	9	43
10	9	94
11	9	77
12	9	33
1	10	11
2	10	24
3	10	78
4	10	100
5	10	99
6	10	68
7	10	52
8	10	45
9	10	74
10	10	98
11	10	99
12	10	37
1	11	7
2	11	29
3	11	79
4	11	97
5	11	92
6	11	95
7	11	100
8	11	89

```

9  11  53
10 11  46
11 11  50
12 11  16
1  12  7
2  12  22
3  12  31
4  12  50
5  12  56
6  12  79
7  12  100
8  12  61
9  12  53
10 12  36
11 12  33
12 12  2
;

PROC VARIOGRAM DATA=A PLOTS=MORAN OUTVAR=B;
COMPUTE LAGD=1 MAXLAG=12 AUTOCORR(ASSUM=RANDOM);
COORDINATES XC=COL YC=ROW;
VAR YI;

PROC PRINT;

PROC VARIOGRAM DATA=A PLOTS=FIT;
COMPUTE LAGD=1 MAXLAG=12 CL ROBUST;
COORDINATES XC=COL YC=ROW;
MODEL FORM=AUTO(MLIST=(SPH EXP GAU) NEST=1);
VAR YI;

RUN;

```

A.4 Yield loss

Copy and paste the below code into a SAS editor, and hit run to see the output.

```

DATA A;
INPUT WP SP $ BLK TRT FUNG DS YIELD;
CARDS;
101 A 1 1 0 43 205
101 B 1 1 1 1 399
102 A 1 2 1 2 426

```

```

102 B 1 2 0 92 102
103 A 1 3 1 2 385
103 B 1 3 0 7 355
201 A 2 2 1 4 412
201 B 2 2 0 75 224
202 A 2 3 1 3 425
202 B 2 3 0 10 352
203 A 2 1 0 27 330
203 B 2 1 1 2 375
301 A 3 1 1 2 420
301 B 3 1 0 47 303
302 A 3 3 0 12 342
302 B 3 3 1 1 395
303 A 3 2 0 63 222
303 B 3 2 1 3 393
401 A 4 3 0 12 326
401 B 4 3 1 1 400
402 A 4 1 0 32 289
402 B 4 1 1 1 418
403 A 4 2 1 2 371
403 B 4 2 0 53 175
;
PROC MIXED COVTEST METHOD=TYPE3;
CLASS BLK TRT FUNG;
MODEL DS=TRT|FUNG/RESIDUAL;
RANDOM BLK BLK*TRT;
LSMEANS TRT|FUNG/DIFF;
run;

PROC MIXED COVTEST METHOD=TYPE3;
CLASS BLK TRT FUNG;
MODEL YIELD=TRT|FUNG/RESIDUAL;
RANDOM BLK BLK*TRT;
LSMEANS TRT|FUNG/DIFF;

PROC REG;
MODEL YIELD=DS;

DATA B;
SET A;
RY1=YIELD/399.23843;

PROC REG;
MODEL RY1=DS;

```

```
PROC SORT DATA=A; BY BLK TRT FUNG;
PROC TRANSPOSE DATA=A OUT=T1A; BY BLK TRT;
VAR YIELD;
run;

DATA T2A;
SET T1A;
RY2=COL1/COL2;
DROP _NAME_ COL1 COL2;
run;

PROC TRANSPOSE DATA=A OUT=T1B; BY BLK TRT;
VAR DS;
run;

DATA T2B;
SET T1B;
CDS=COL1;
DROP _NAME_ COL1 COL2;
run;

DATA T3;
MERGE T2A T2B;
BY BLK TRT;

PROC PRINT;
run;

PROC MIXED COVTEST;
CLASS BLK TRT;
MODEL RY2=TRT/RESIDUAL;
RANDOM BLK;
LSMEANS TRT/DIFF;

PROC REG;
MODEL RY2=CDS;

RUN;
```