

Supplementary Information

Rapid Discovery of Self-Assembling Peptides with One-Bead One-Compound Peptide Library

Pei-Pei Yang^{1,#}, Yi-Jing Li^{1,2,#}, Yan Cao^{3,#}, Lu Zhang⁴, Jia-Qi Wang¹, Zi-Wei Lai³, Kuo Zhang^{1,2}, Diedra Shorty⁴, Wenwu Xiao⁴, Hui Cao², Lei Wang^{1,*}, Hao Wang^{1,5,*}, Ruiwu Liu^{4,*}, Kit S. Lam^{4,6,*}

¹CAS Center for Excellence in Nanoscience, CAS Key Laboratory for Biomedical Effects of Nanomaterials and Nanosafety, National Center for Nanoscience and Technology (NCNST) No. 11 Beiyitiao, Zhongguancun, Beijing, 100190, China.

²Department of Materials Physics and Chemistry, School of Materials Science and Engineering, University of Science and Technology Beijing, Beijing, 100083, P. R. China.

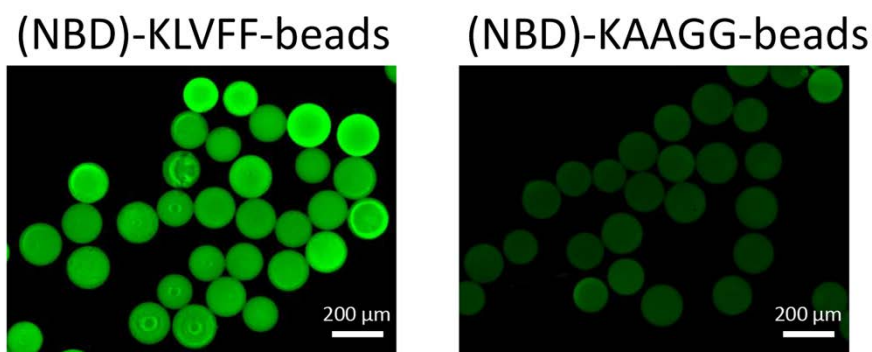
³Institute for Advanced Study, Shenzhen University, Guangdong 518060, China.

⁴Department of Biochemistry and Molecular Medicine, UC Davis NCI-designated Comprehensive Cancer Center, University of California Davis, Sacramento, California 95817, United States.

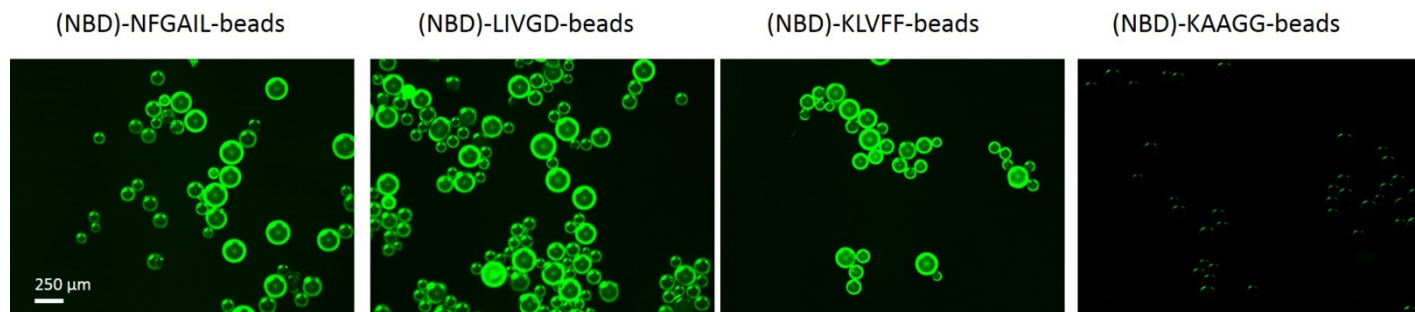
⁵Center of Materials Science and Optoelectronics Engineering, University of Chinese Academy of Sciences, Beijing 100049, P. R. China.

⁶Division of Hematology and Oncology, Department of Internal Medicine, School of Medicine, University of California Davis, Sacramento, California 95817, United States.

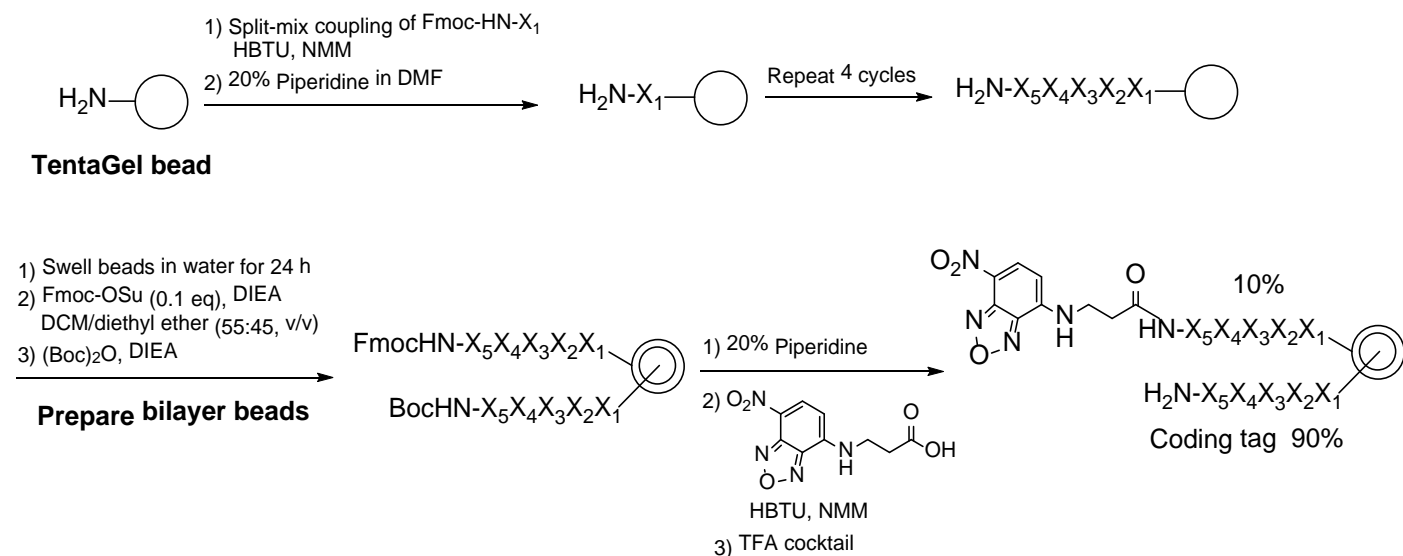
E-mail: kslam@ucdavis.edu; rwliu@ucdavis.edu; wanglei@nanoctr.cn; wanghao@nanoctr.cn.



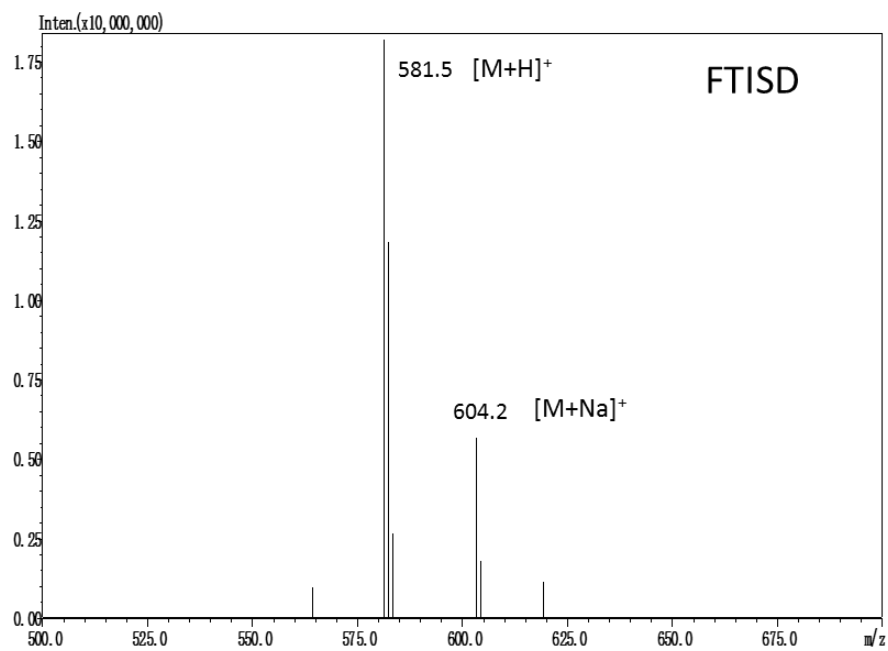
Supplementary Figure 1. Under aqueous condition, TentaGel beads displaying self-assembling peptide KLVFF N-capped by NBD was found to fluoresce strongly. TentaGel beads displaying negative control non-assembling peptide KAAGG did not fluoresce, thus validating the screening assay for self-assembling peptide discovery.



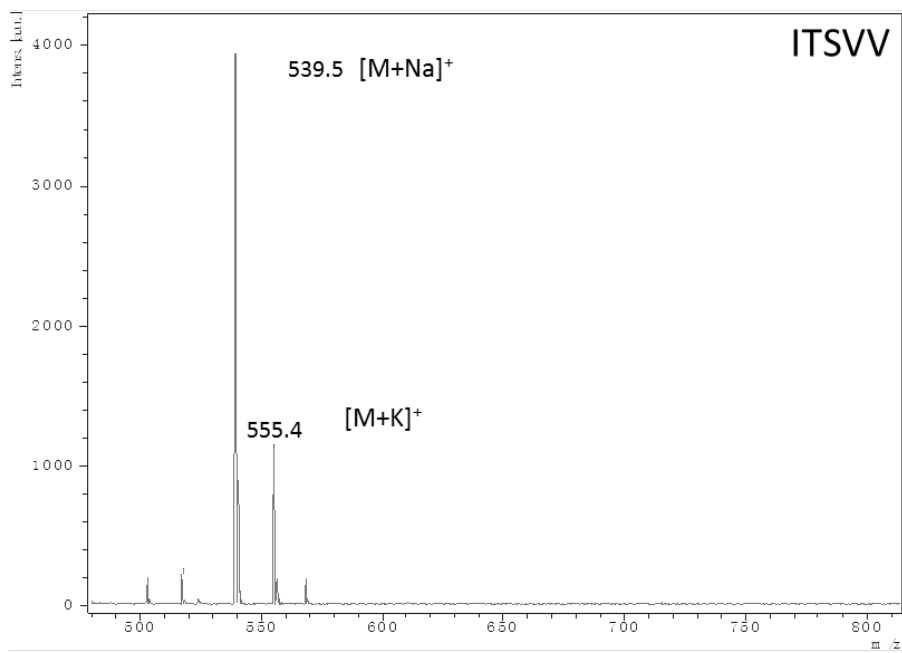
Supplementary Figure 2. Photomicrographs of TentaGel beads displaying NFGAIL, LIVGD, KLVFF, and KAAGG N-terminally capped with NBD at 10% level. NFGAIL, LIVGD, KLVFF are known self-assembling peptides under aqueous condition.



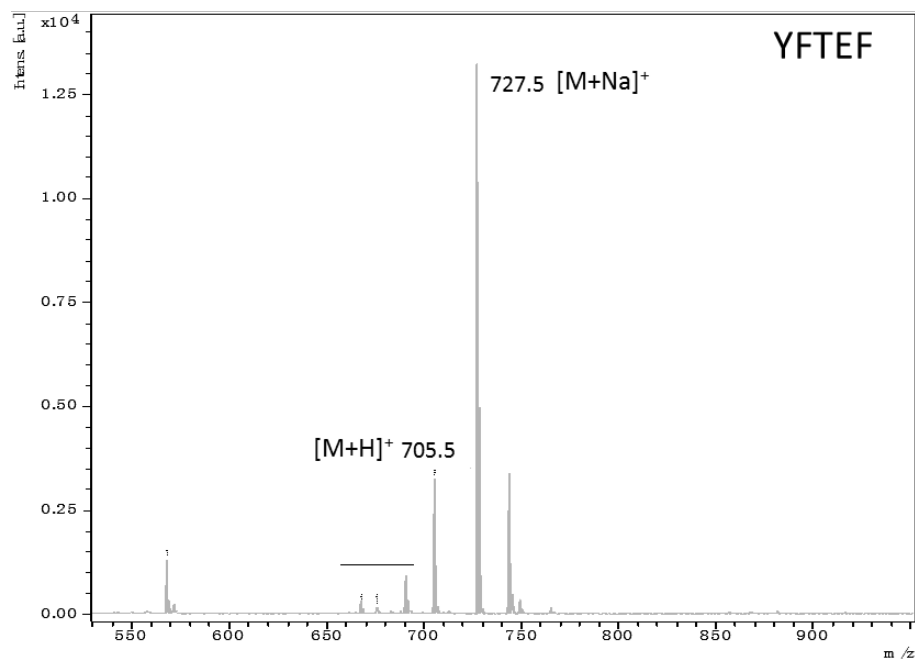
Supplementary Figure 3.. Synthetic scheme of OBOC peptide library N-capped with nitro-1,2,3-benzoxadiazole (NBD), for self-assembling peptide discovery. Only the outer layer of the random peptides was N-capped with the dye. The peptides in the bead interior remained N-terminally free, which is needed for Edman sequencing.



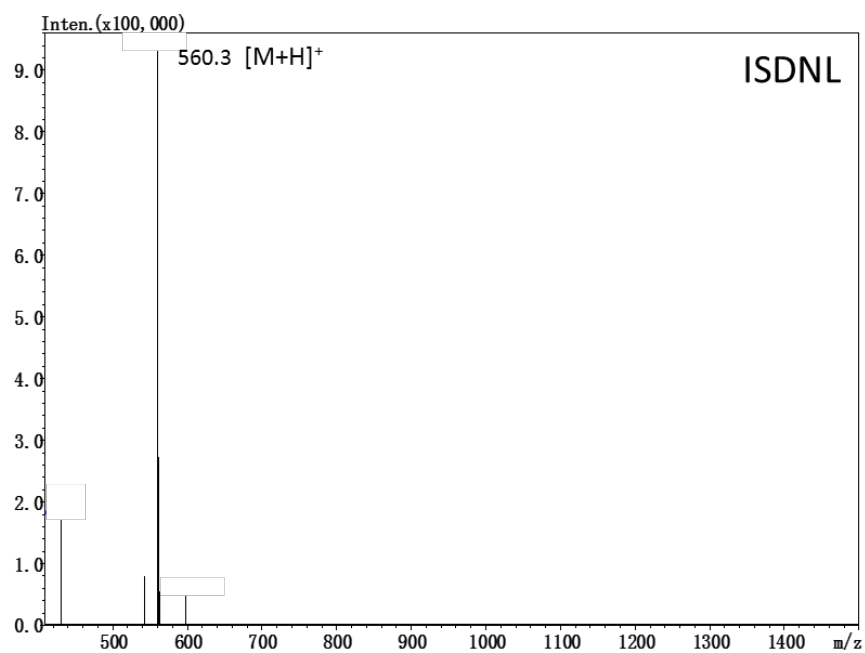
Supplementary Figure 4. The MALDI-TOF spectrum of peptide FTISD



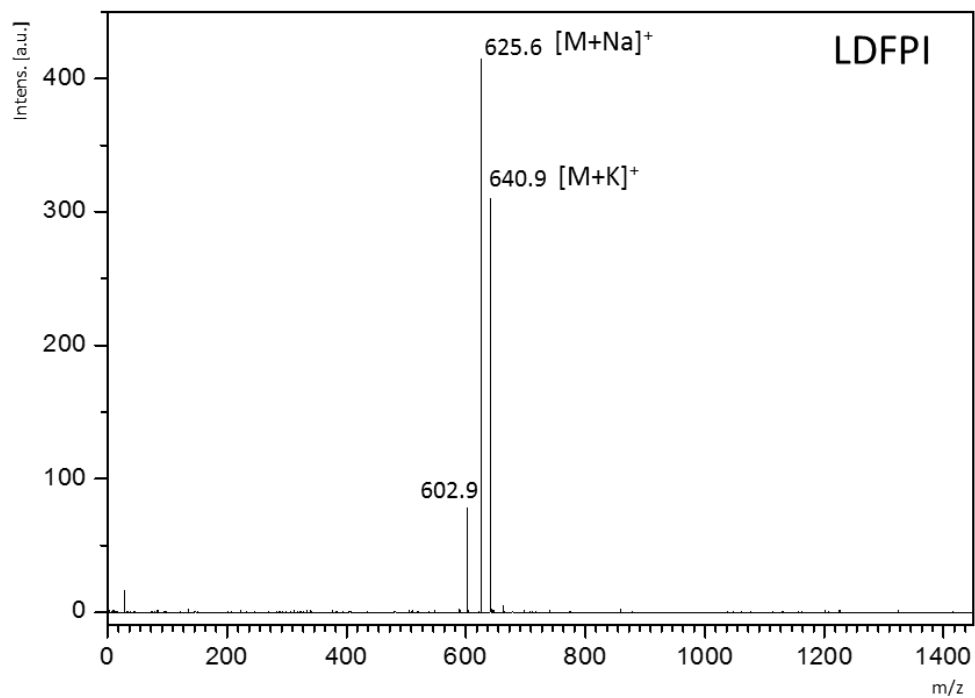
Supplementary Figure 5. The MALDI-TOF spectrum of peptide ITSVV



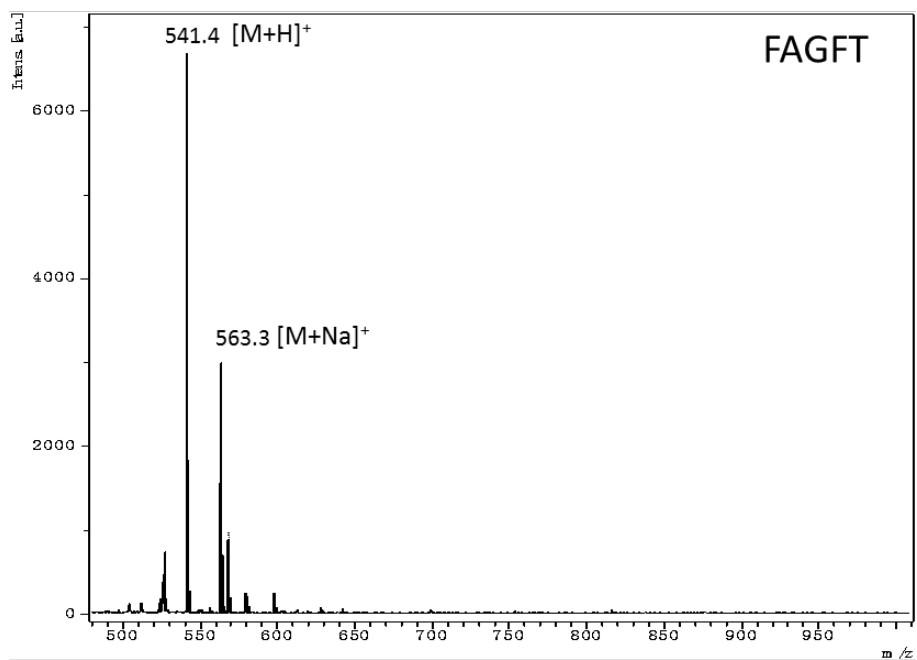
Supplementary Figure 6. The MALDI-TOF spectrum of peptide YFTEF.



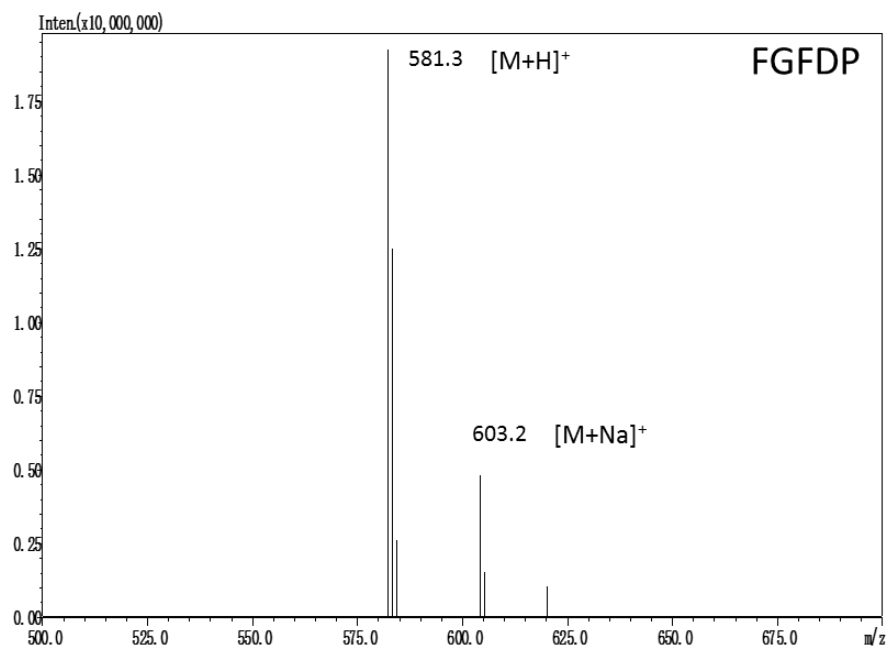
Supplementary Figure 7. The MALDI-TOF spectrum of peptide ISDNL.



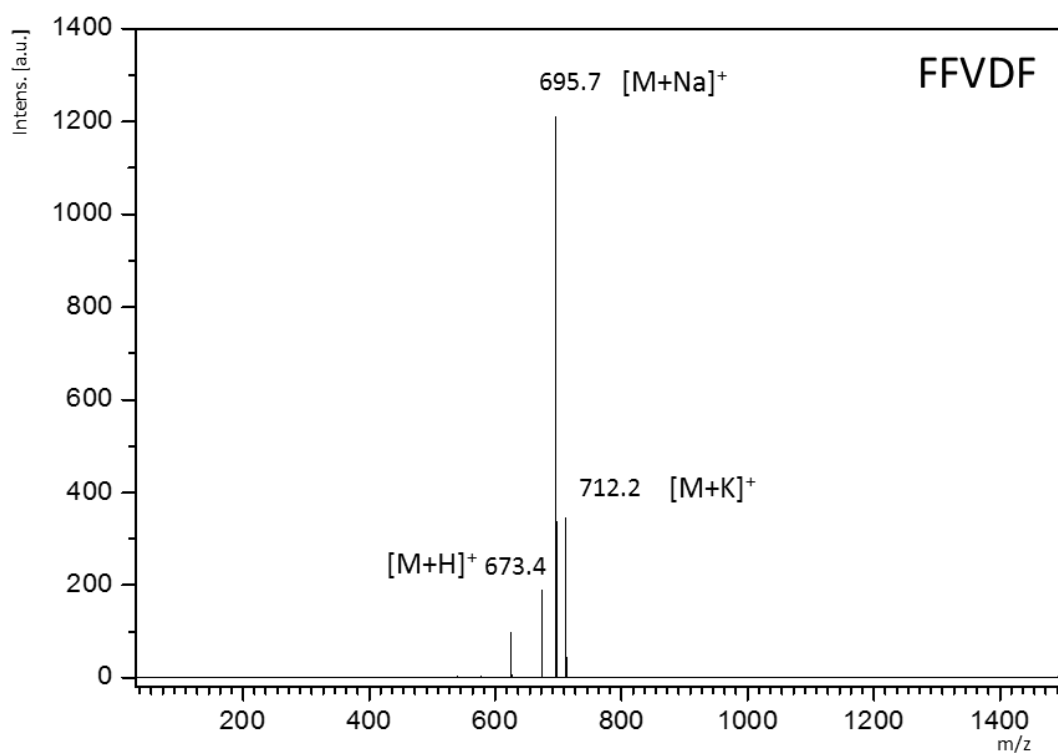
Supplementary Figure 8. The MALDI-TOF spectrum of peptide LDFPI.



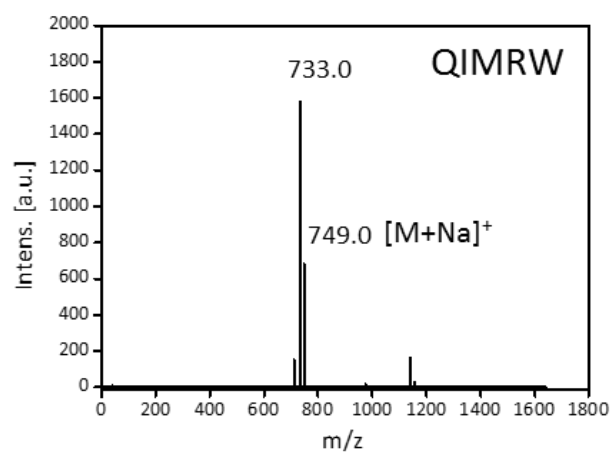
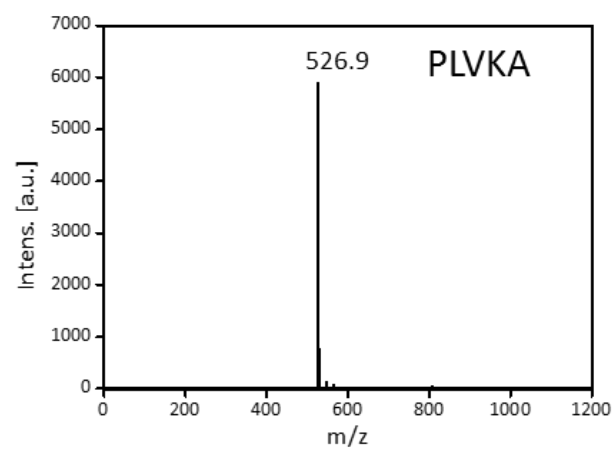
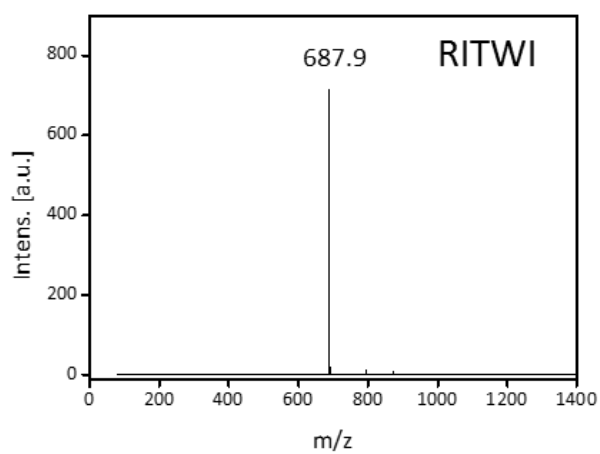
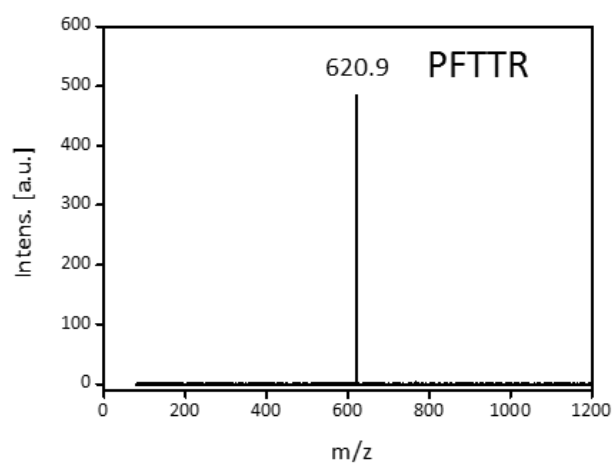
Supplementary Figure 9. The MALDI-TOF spectrum of peptide FAGFT.



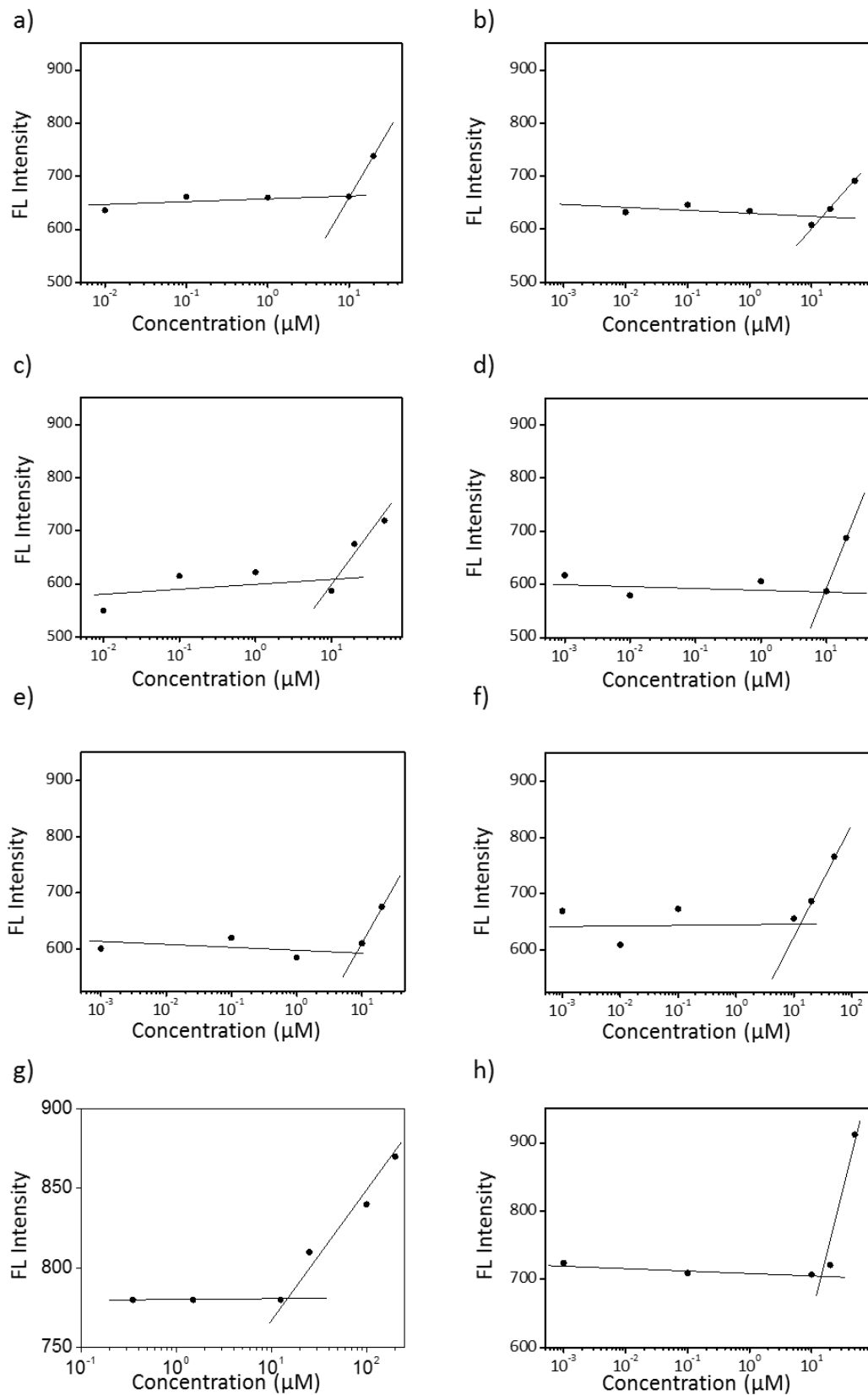
Supplementary Figure 10. The MALDI-TOF spectrum of peptide FGFDp.



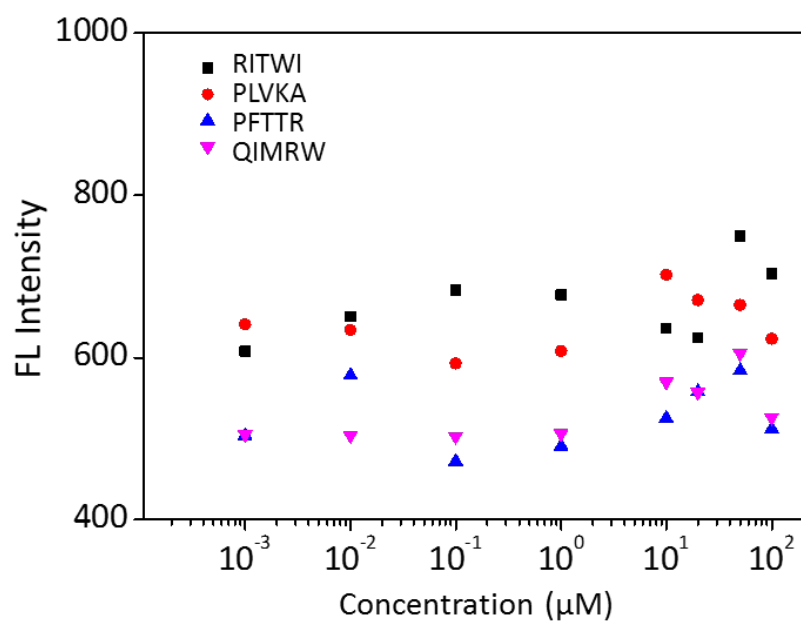
Supplementary Figure 11. The MALDI-TOF spectrum of peptide FFVDF.



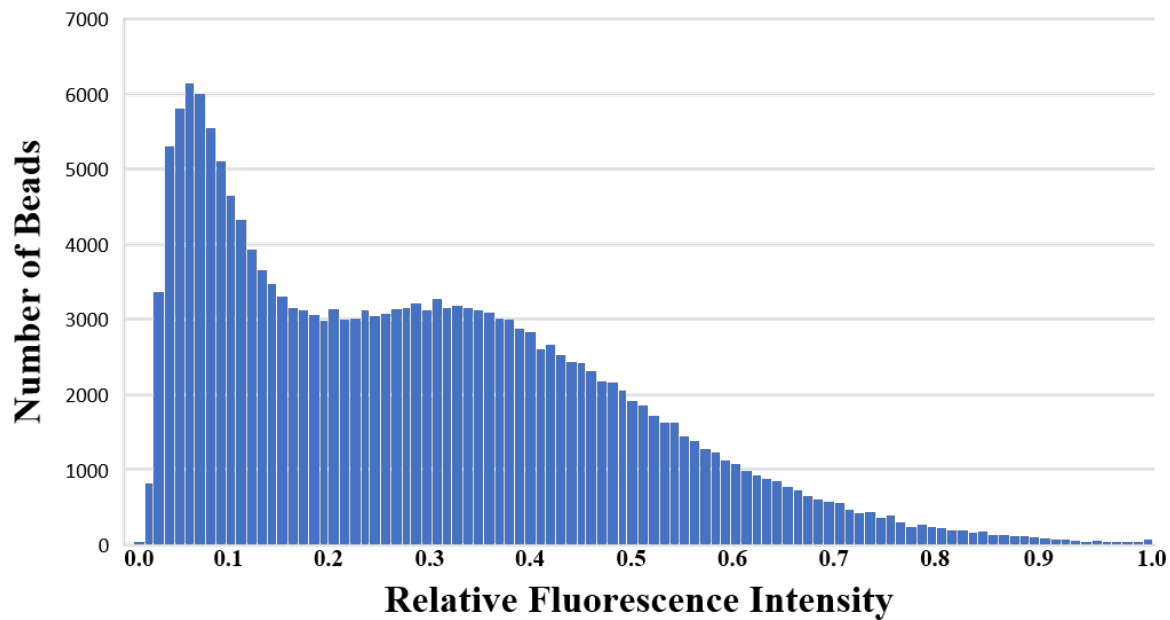
Supplementary Figure 12. The MALDI-TOF spectra of negative non-assembling pentapeptides identified from screening OBOC library.



Supplementary Figure 13. The CMC values of the peptide a)-h) FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP and FFVDF in PBS with 1% DMSO.



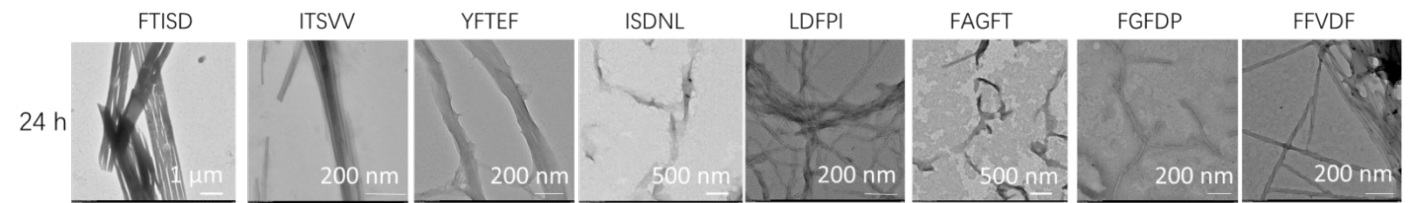
Supplementary Figure 14. The CMC experiment of non-assembling peptide RITWI, PLVKA, PFTTR, and QIMRW in PBS with 1% DMSO.



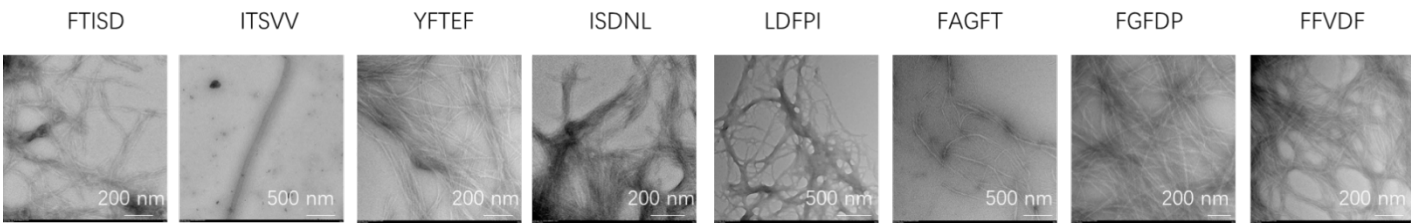
Supplementary Figure 15. Fluorescence profile of an entire immobilized random OBOC pentapeptide library N-capped with NBD dye under aqueous condition, with relative fluorescence unit ranging between 0 to 1.0. Of the 186,288 beads immobilized and screened, 79 or $\sim 0.04\%$ beads displayed a relative fluorescence intensity ≥ 0.99 , and 559 or 0.3% beads displayed a relative fluorescence intensity ≥ 0.9 . Each vertical bar depicts the number of beads present within a 0.01 relative fluorescence unit range.

Supplementary Table 1. q values, Miller indexes, observed d -spacings (d_{obs} , nm) and calculated d -spacings (d_{cal} , nm) of FFVDF crystals in the nanofibers.

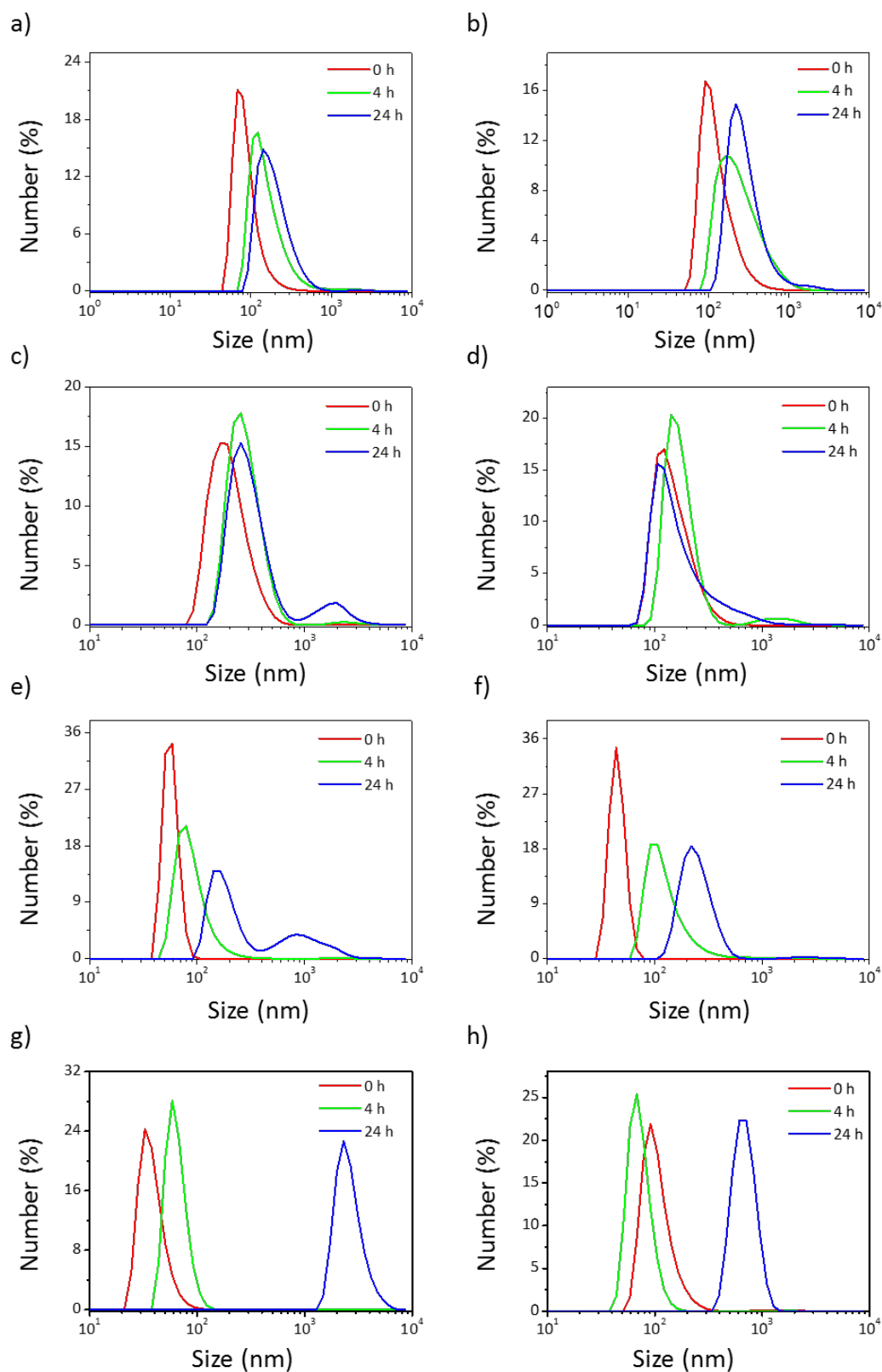
$q(\text{nm}^{-1})$	h	k	l	$d_{obs} \text{ (nm)}$	$d_{cal} \text{ (nm)}$
1.81	0	1	0	3.47	3.53
2.77	0	0	2	2.27	2.28
3.41	0	1	2	1.84	1.92
5.64	0	0	4	1.11	1.14
7.24	0	4	0	0.87	0.88
8.94	0	4	4	0.70	0.70
10.21	1	3	4	0.61	0.62
11.49	0	2	8	0.55	0.54
13.20	2	0	0	0.48	0.47
13.73	2	0	2	0.46	0.47
14.15	2	2	2	0.44	0.45
15.54	0	8	4	0.40	0.41
15.75	2	4	4	0.40	0.39
17.24	2	4	6	0.36	0.37
20.22	2	8	4	0.31	0.31



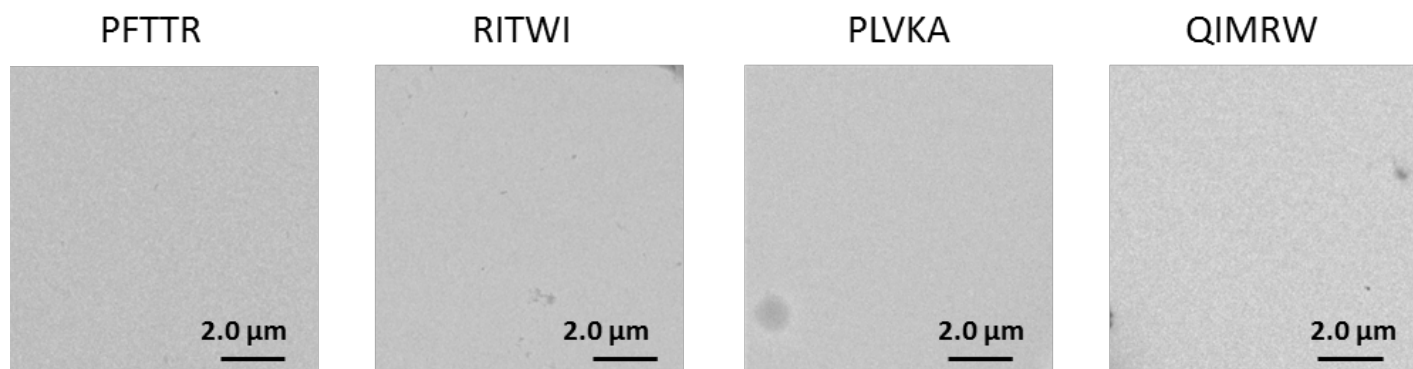
Supplementary Figure 16. The TEM images of pentapeptides FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP, and FFVDF at 50 μM , 24 h after solubilized in water with 1% DMSO. The scale bar: 1 μm .



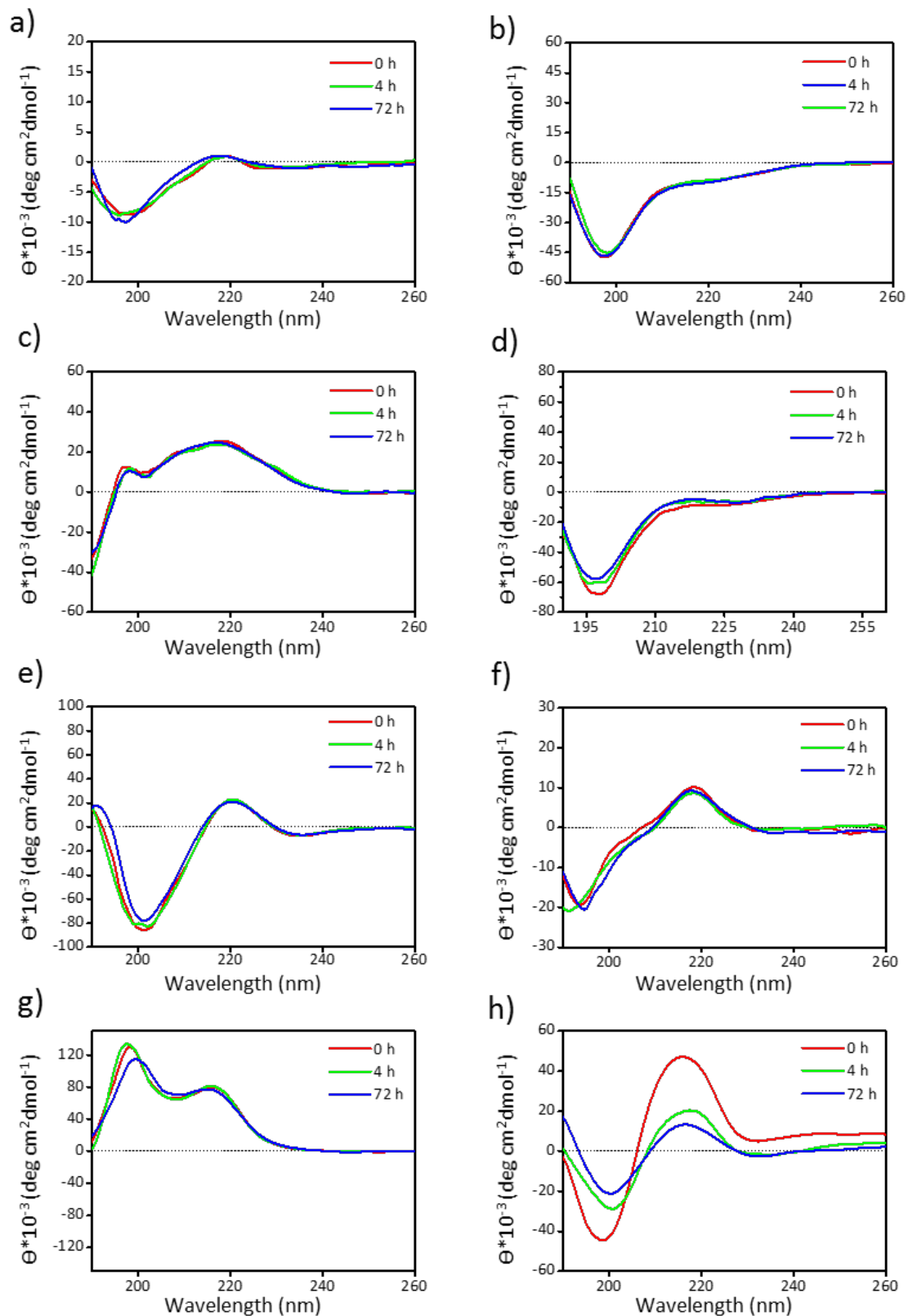
Supplementary Figure 17. The TEM images of pentapeptides FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP and FFVDF that had undergone thermal annealing at 90 $^{\circ}\text{C}$ for 5 h.



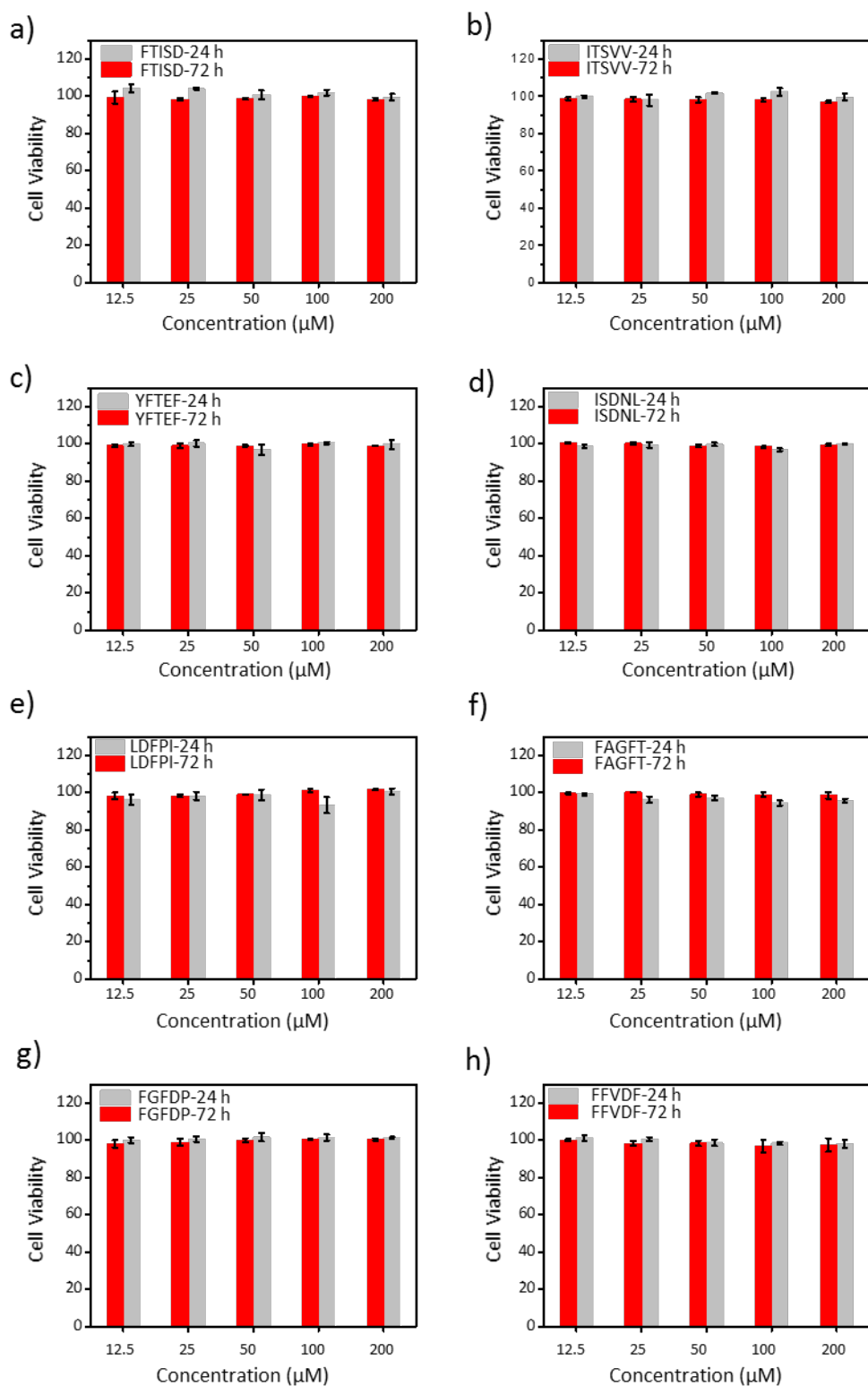
Supplementary Figure 18. The DLS spectra of the pentapeptides a) - h) FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP, and FFVDF at 50 μ M in water with 1% DMSO, obtained at 0, 4 and 24 h.



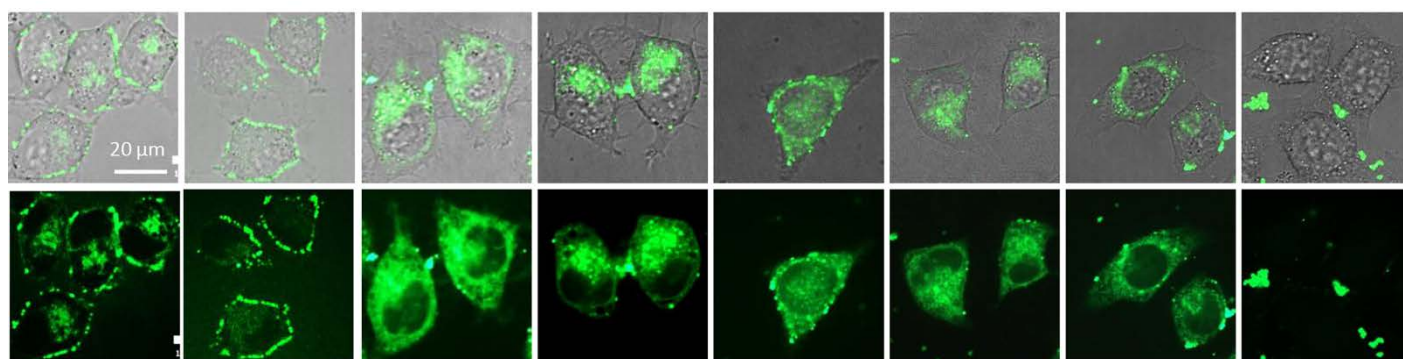
Supplementary Figure 19. The TEM images of the non-assembling pentapeptides PFTTR, RITWI, PLVKA, and QIMRW at 50 μ M in water with 1% DMSO after 24 h. No nanostructure was detected.



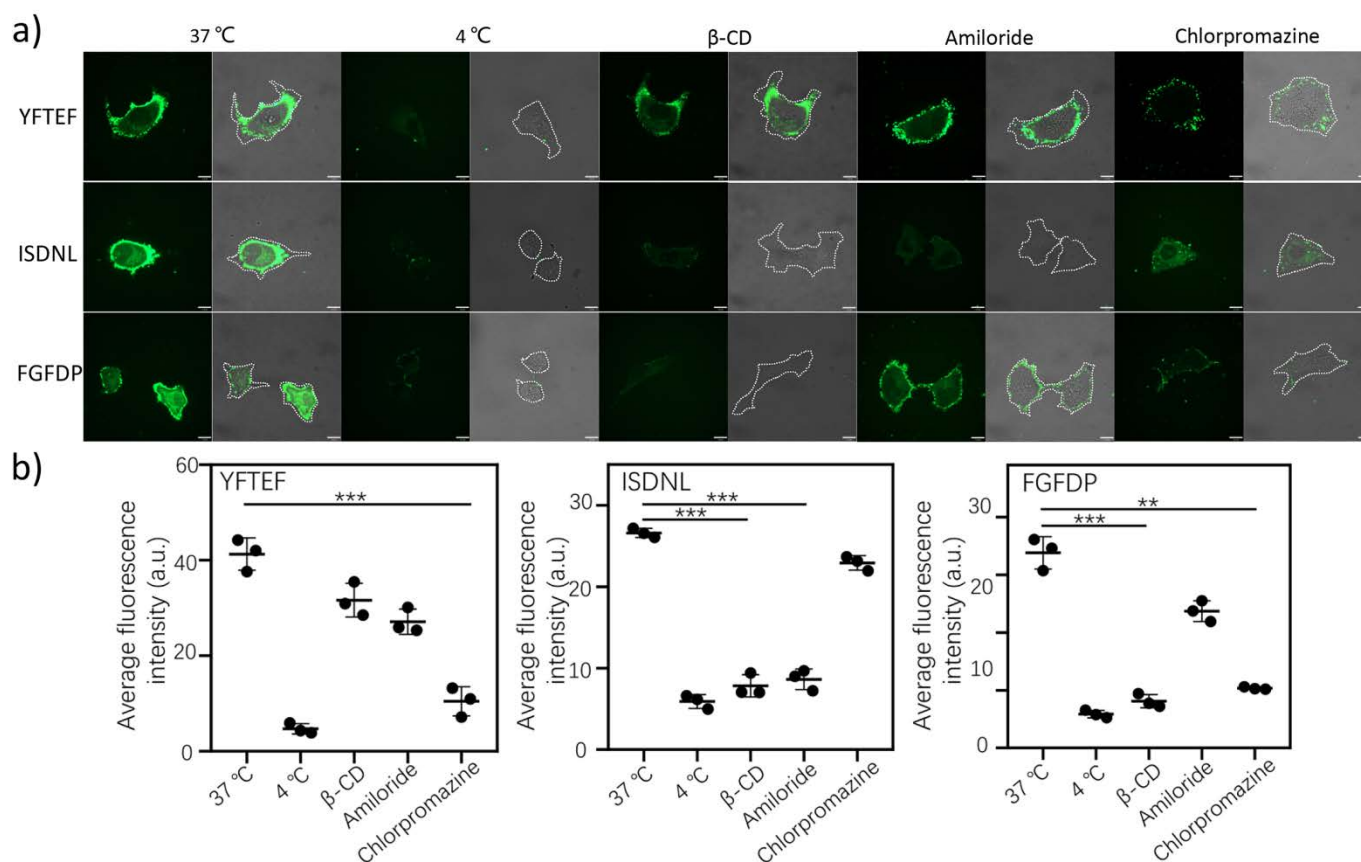
Supplementary Figure 20. The time-dependent CD spectra of the pentapeptides FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP, FFVDF at 40 μ M corresponding to a)-h) in water with 1% DMSO. Significant changes in CD-spectra were observed over time in FFVDF peptide.



Supplementary Figure 21. a-h. CCK-8 cell viability assay for 8 pentapeptides (FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDG, FFVDF) after incubation with HeLa cells for 24 and 72 h. No cell toxicity was observed, Error bars represent Standard Deviation (n=3).



Supplementary Figure 22. Confocal microscopy of HeLa cells after incubation with FITC-labeled pentapeptide assemblies (from left to right: FTISD, ITSVV, YFTEF, ISDNL, LDFPI, FAGFT, FGFDP, FFVDF) for 4 h; peptide concentration used was 50 μ M. 3% of the assembled peptides was N-terminally labeled with FITC.



Supplementary Figure 23. Cell uptake pathway analysis for FITC-labeled pentapeptide assemblies YFTEF, ISDNL and FDFDP. a) CLSM images of HeLa cells incubated with pentapeptides (50 μ M) at different temperatures (37 or 4 °C) and in the presence of various endocytosis inhibitors, such as amiloride (2 mM), M- β -CD (5 mM), and chlorpromazine (50 μ M). The scale bar is 10 μ m. b) Relevant quantitative analysis of images from 'a'. 3% of the assembled peptides was N-terminally labeled with FITC, Error bars represent Standard Deviation (n=3).

