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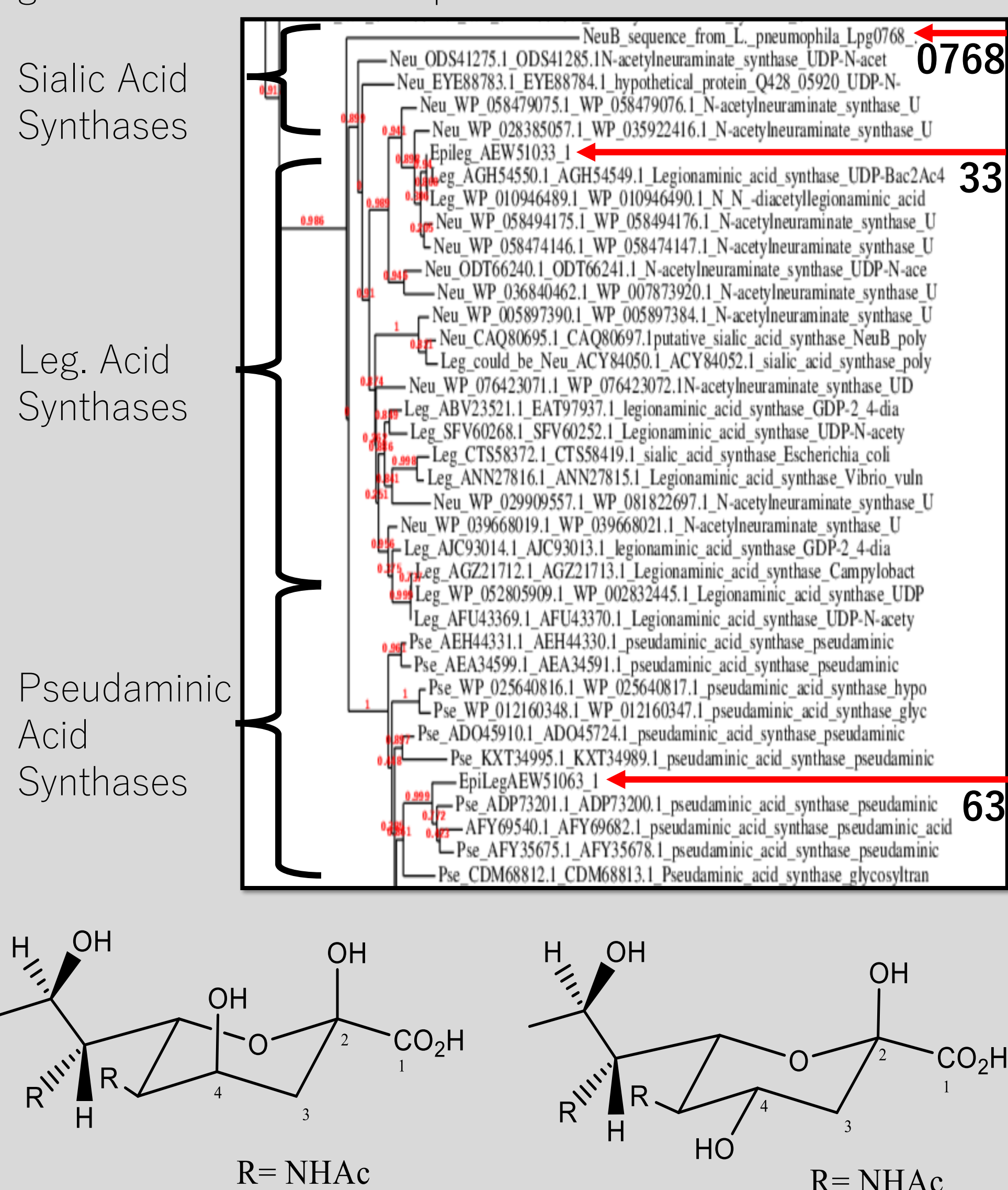
# Characterizing Legionaminic Acid Biosynthesis from Diverse Species

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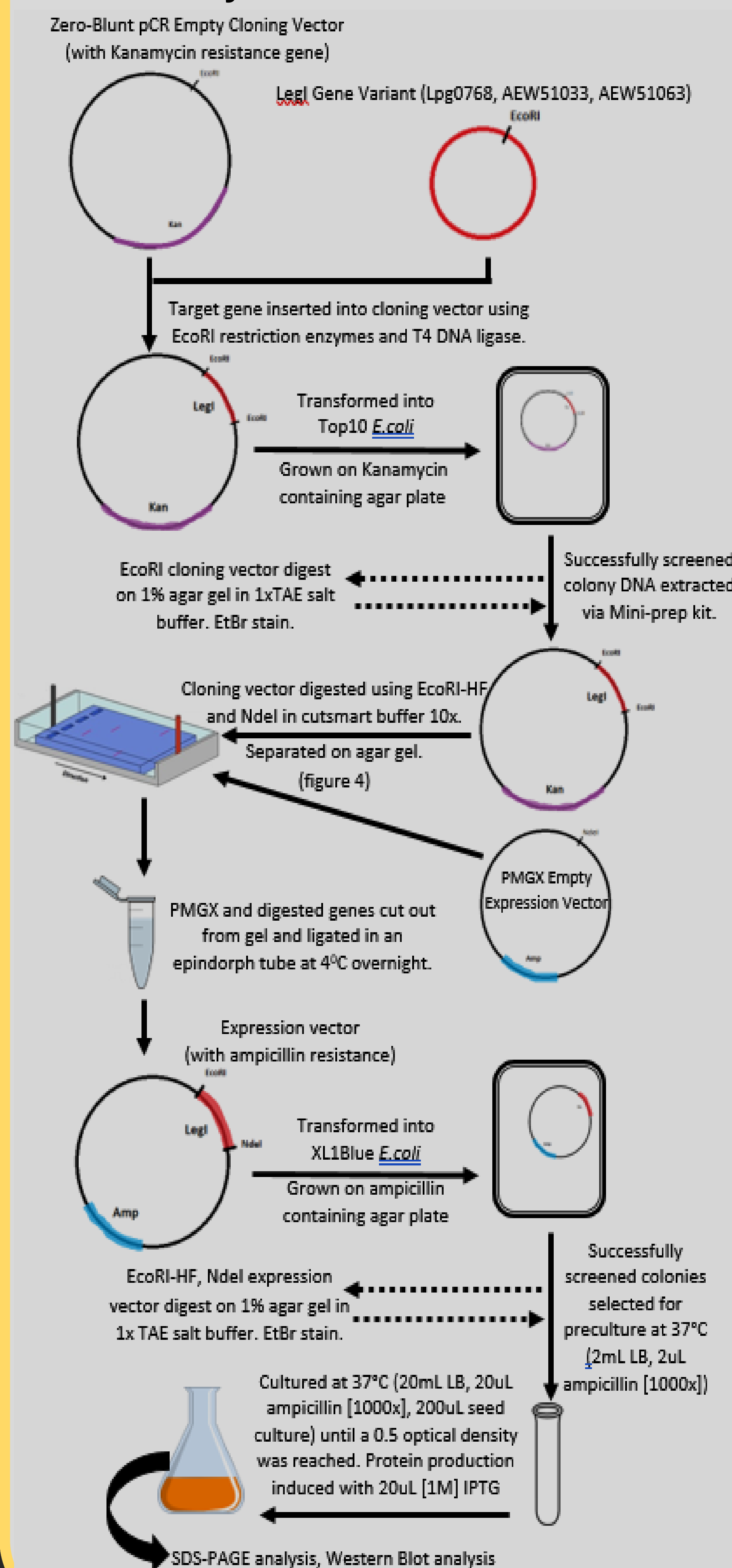
## I. Background

Legionaminic acid belongs to a family of carbohydrates called nonulosonic acids (nine carbon alpha keto acids) and are 5,7-diamino derivatives of the more commonly known sialic acid, Neu5Ac. These carbohydrates are found in cell surface glycoconjugates of gram-negative bacteria such as *H. pylori*, *C. jejuni*, and *L. pneumophila*, and contribute to their pathogenicity. The physiological role of legionaminic acid in bacterial pathogenicity is not yet well understood since there is no reliable source of it to support its study.<sup>1</sup> This project helps examine the role of synthase genes in a biosynthetic pathway initially characterized in *C. jejuni* in 2009.

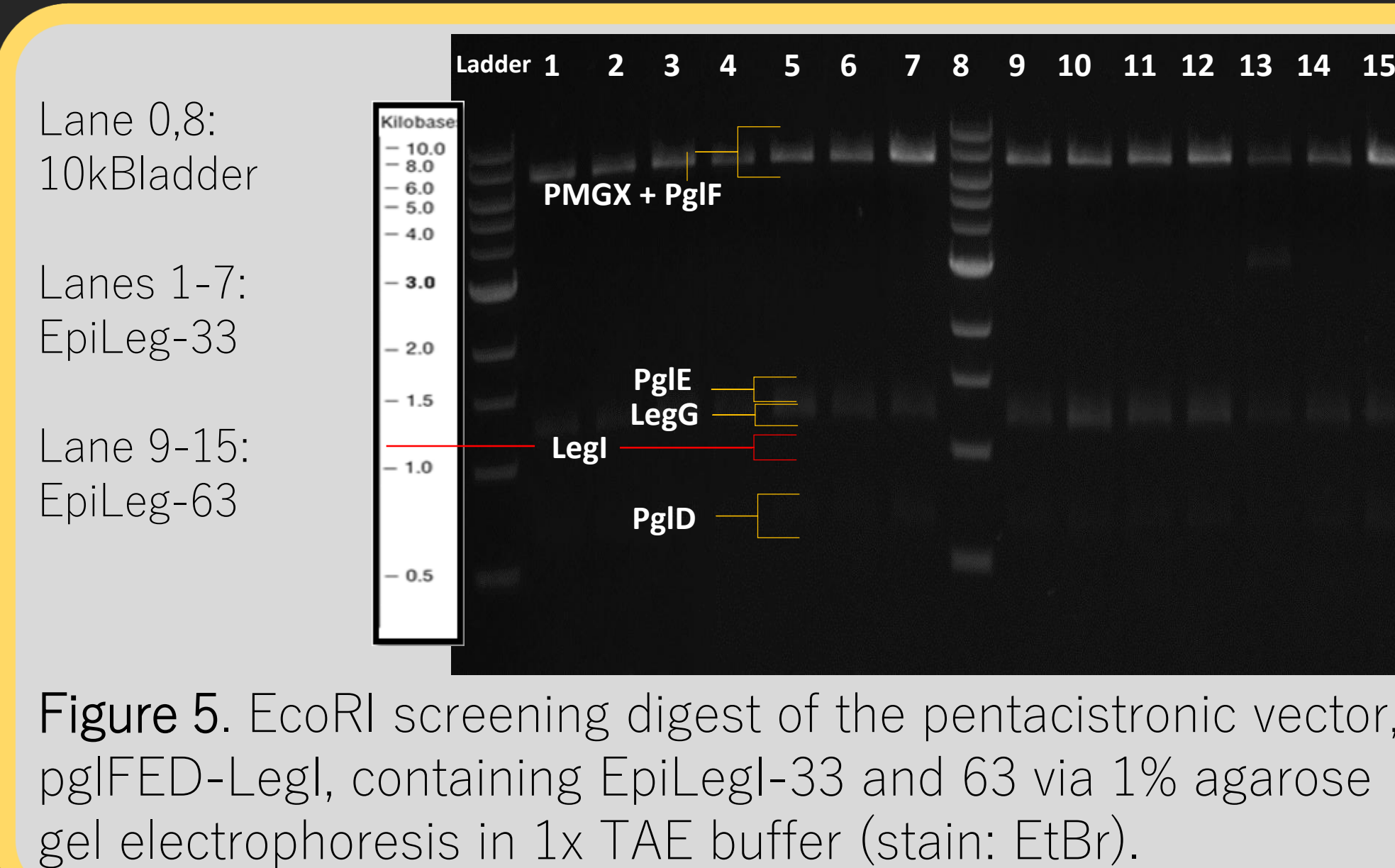
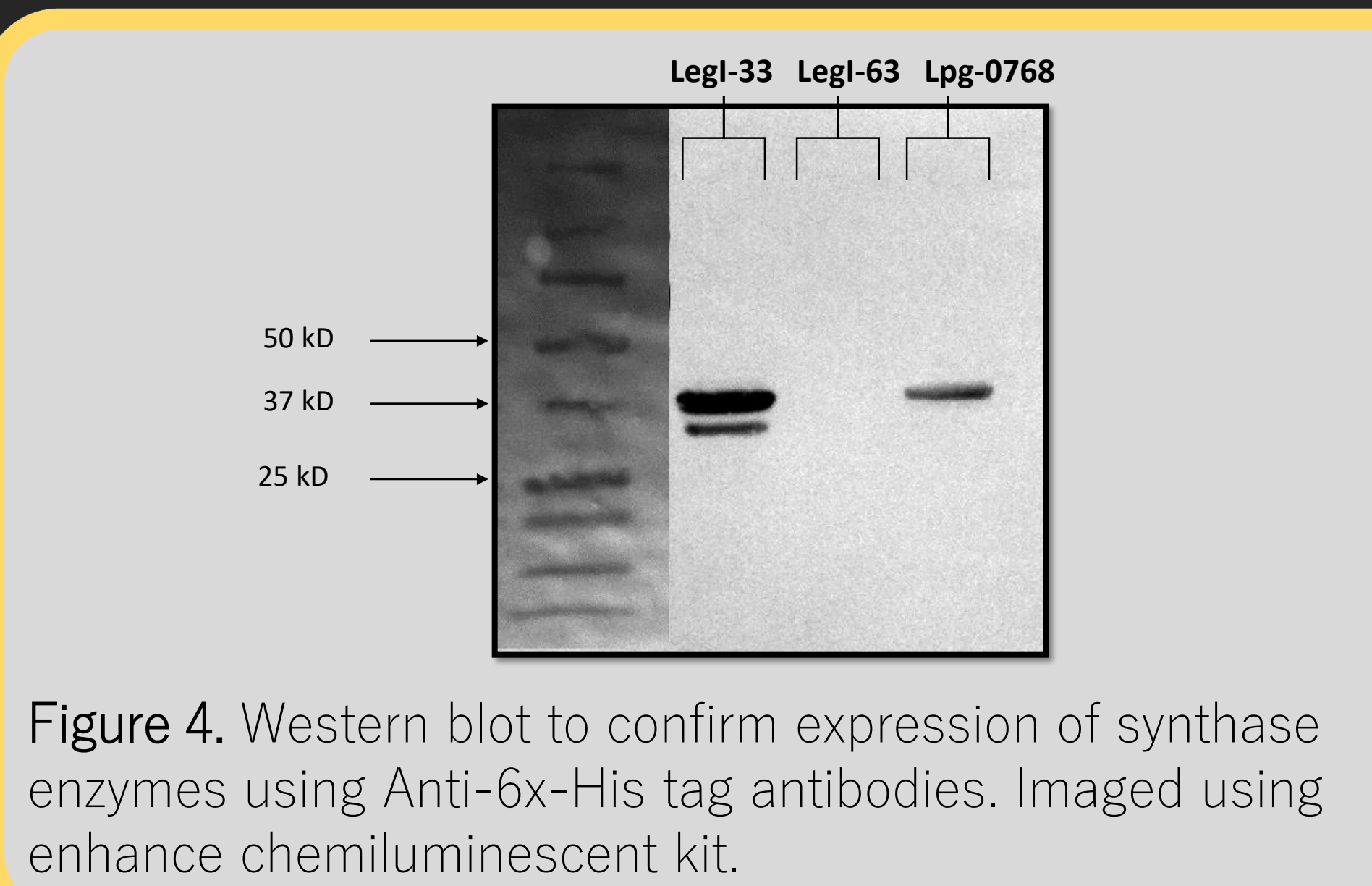
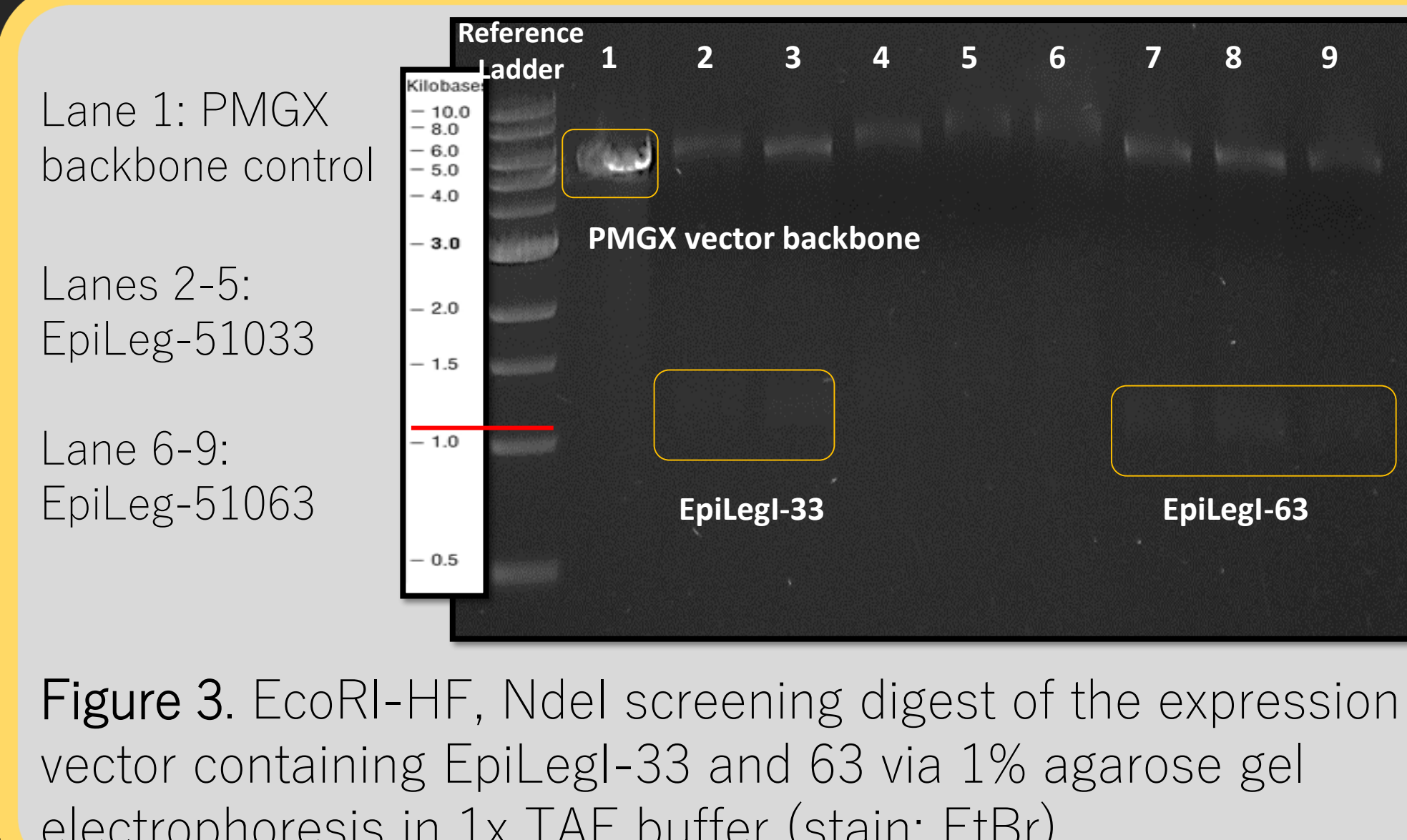
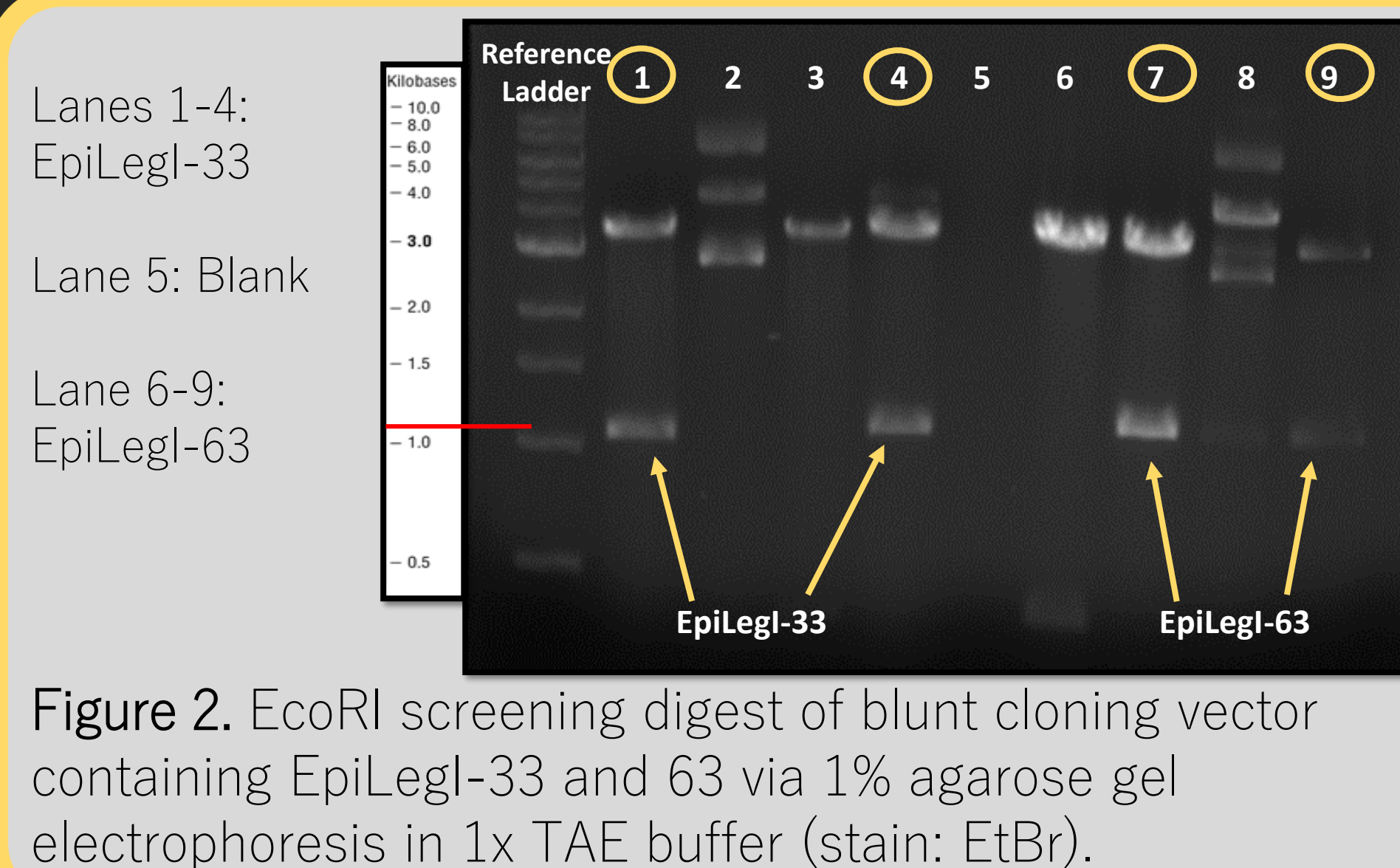
The biosynthetic pathway for LA contains five genes: three bacterial protein glycosylation (PglE, PglF, PglD) genes from *C. jejuni*, and two LA synthase genes (LegF, LegI) from *L. pneumophila*. Bioinformatic analysis revealed three closely related LegI genes that might generate a modified 4-epi LA stereoisomer, which has been isolated in previous studies but no dedicated biosynthetic pathway exists. Natively these are synthase genes for sialic acid and pseudaminic acid.



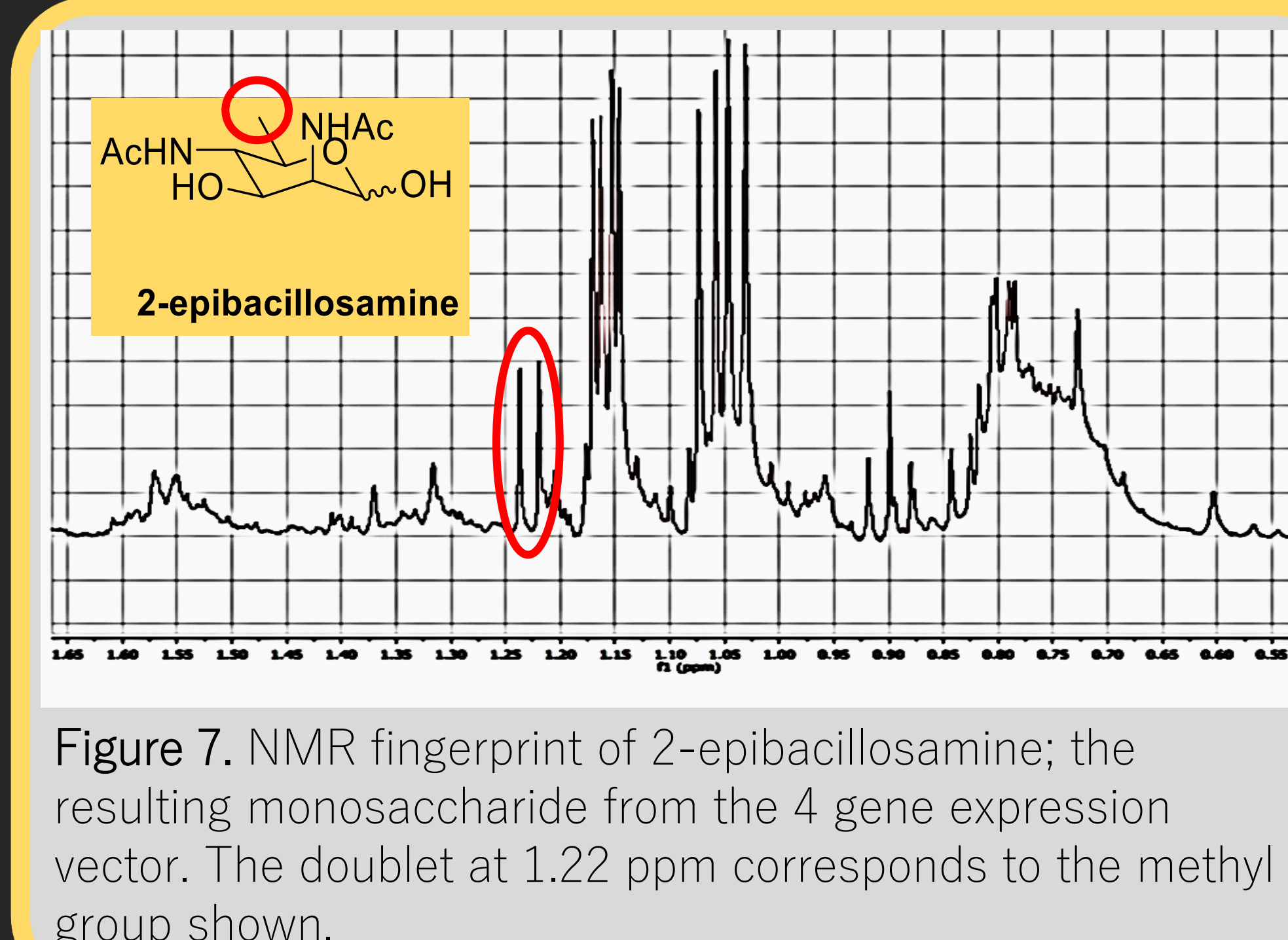
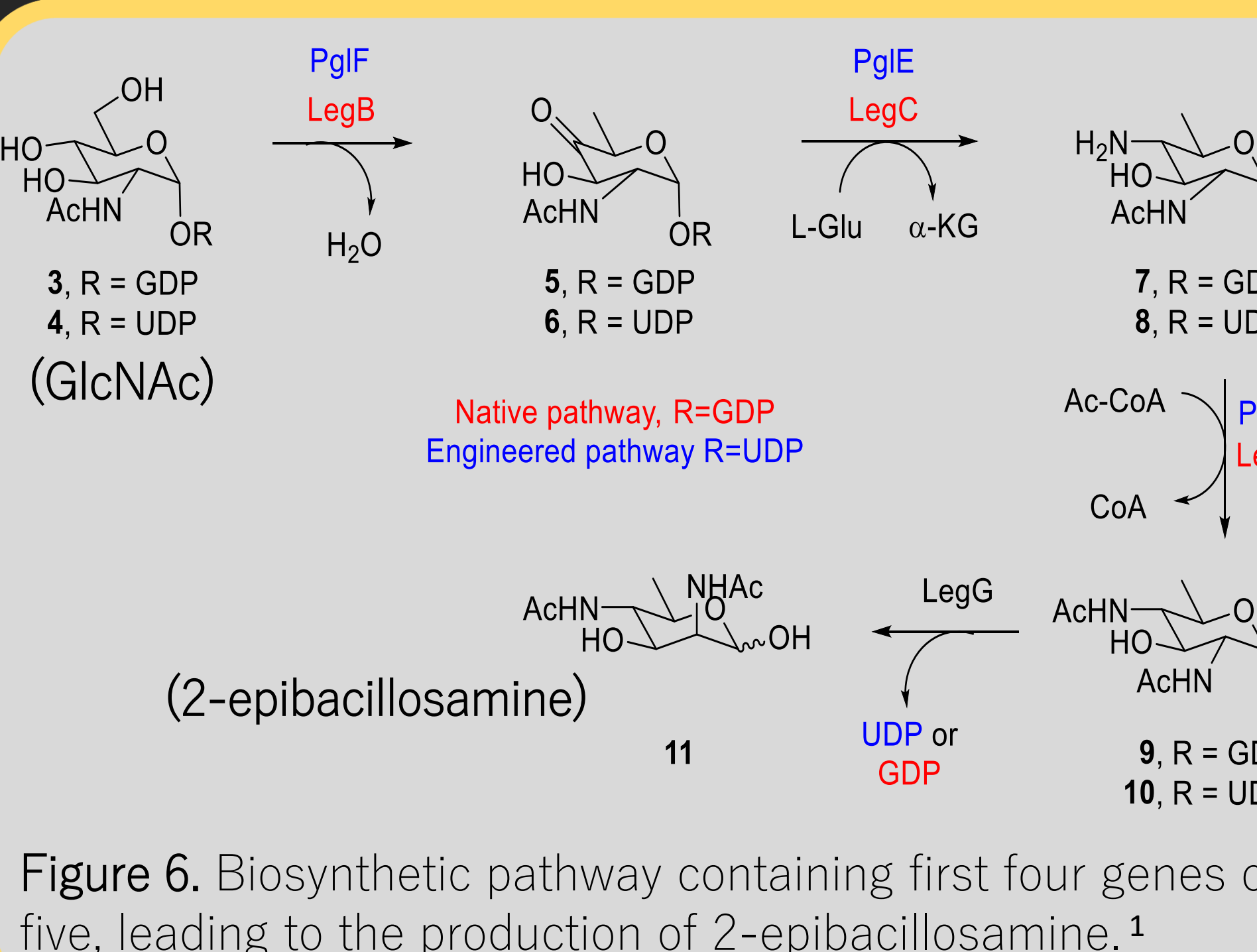
## II. Methods – Confirming LegI Synthase Production



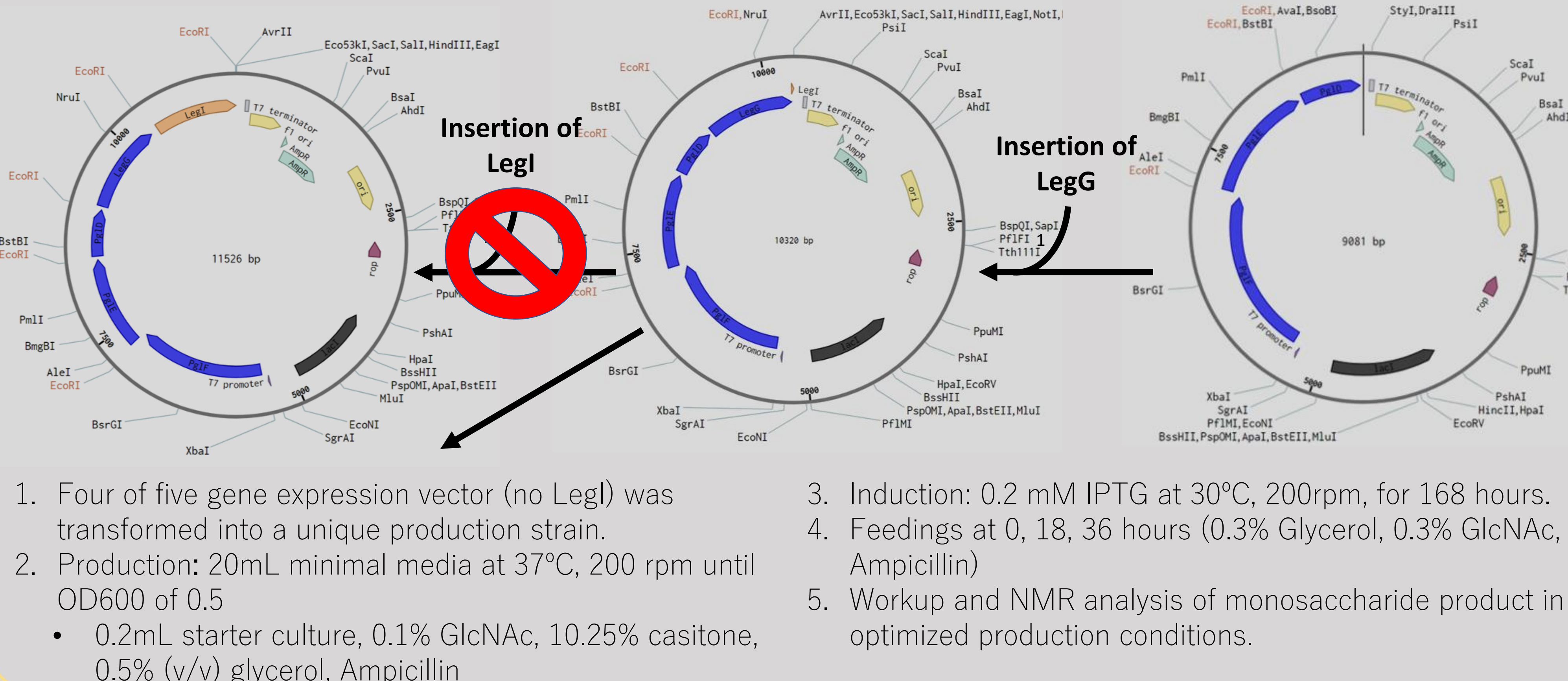
## III. Results – LegI Expression



## III. Results – Biosynthetic Pathway Expression without LegI



## II. Methods – Production of 2-epibacillosamine



## IV. Conclusions

- 2-epibacillosamine, like legionaminic acid, is another difficult to isolate pathogenic factor in gram negative bacteria. The tetracistronic vector, pglFED-LegG, offers the first developed biosynthetic pathway for its production.
- Sub-cloning of the LegI gene in with the pgl operons proved unsuccessful using in house golden gate assembly. The figure 4 digest is missing a DNA fragment at 1.2kB corresponding to the expected LegI insertion (fifth component).
- Monosaccharide production was induced in pglFED-LegG instead. NMR analysis revealed a doublet peak at 1.22ppm, corresponding to the methyl group seen in 2-epibacillosamine confirming its production.
- pglFED-LegG supports hypothesized 2-epibacillosamine pathway (figure 6).
- Glycerol; an energy dense, metabolically significant molecule; provided the greatest potential for cultures to grow. The glycerol rich medium optimized production of glycoconjugates, in this case 2-epibacillosamine,
- Figure 2 illustrates successful cloning vector recombination for EpiLegI-33 and EpiLegI-63. The LegI gene is around 1.2kB in length, and the digested sample shows a fragment at 1.2kB and a cloning vector backbone at 3.5kB.
- Figure 3 illustrates successful recombination of the expression vector in lanes 2, 3 (EpiLegI-33) and 7, 8, 9 (EpiLegI-63). An over dilution of the extracted DNA led to very dim images, but DNA fragments are still marginally visible at 1.2kB.
- Western blot using Anti-6x-His tag antibodies confirmed synthase production for EpiLegI-33 and Lpg-0768, but not for EpiLegI-63. This indicates EpiLegI-63 is not feasible for the full synthetic pathway for Legionaminic acid.
- Initially the third gene, Lpg-0768, was not successfully transformed as the growth media did not produce colonies. Later on in the project it was successfully cloned and expressed but images are not provided on the space limited poster.

## V. Future Directions

- Sub cloning LegI into the full legionaminic acid biosynthetic pathway to express legionaminic acid.
- Bacillosamine synthase targeting antibiotics can be designed and tested to investigate the physiological significance of bacillosamine glycoconjugates and their effects on gram negative prokaryotic pathogenicity.
- Optimization of growth mediums for 2-epibacillosamine production to achieve a more concentrated final product.

## VI. Works Cited

- Hassan, M. I.; Lundgren, B. L.; Chaumun, M.; Whitfield, D. M.; Clark, B.; Schoenhofen, I. C.; Boddy, C. N. *Angew. Total Biosynthesis of Legionaminic Acid, a Bacterial Sialic Acid Analog*. Chem. Int. Ed. 2016, 55, 12018-12021.

## VII. Acknowledgements

Thank you, Mohamed Hassan for your generous contributions, and Christopher N. Boddy for your continued support. This project was funded by the University of Ottawa, UROP, and NSERC.