

# Bayesian Model Averaging for Ensemble-Based Estimates of Solvation Free Energies

Luke J. Gosink,<sup>†</sup> Christopher C. Overall,<sup>†</sup> Sarah M. Reehl,<sup>†</sup> Paul D. Whitney,<sup>‡</sup>

David L. Mobley,<sup>¶</sup> and Nathan A. Baker<sup>\*,‡,§</sup>

<sup>†</sup>*Computational and Statistical Analytics Division, Pacific Northwest National Laboratory,  
Richland, WA 99352, USA*

<sup>‡</sup>*Advanced Computing, Mathematics, and Data Division, Pacific Northwest National  
Laboratory, Richland, WA 99352, USA*

<sup>¶</sup>*UC*

<sup>§</sup>*Division of Applied Mathematics, Brown University, Providence, RI 02912, USA*

E-mail: [nathan.baker@pnnl.gov](mailto:nathan.baker@pnnl.gov)

Phone: +1-509-375-3997

## Abstract

This paper applies the Bayesian Model Averaging (BMA) statistical ensemble technique to estimate small molecule solvation free energies. There is a wide range of methods for predicting solvation free energies, ranging from empirical statistical models to *ab initio* quantum mechanical approaches. Each of these methods are based on a set of conceptual assumptions that can affect a method’s predictive accuracy and transferability. Using an iterative statistical process, we have selected and combined solvation energy estimates using an ensemble of 17 diverse methods from the SAMPL4 blind prediction study to form a single, aggregated solvation energy estimate. The ensemble design process evaluates the statistical information in each individual method as well as the performance of the aggregate estimate obtained from the ensemble as a whole. Methods that possess minimal or redundant information are pruned from the ensemble and the evaluation process repeats until aggregate predictive performance can no longer be improved. We show that this process results in a final aggregate estimate that outperforms all individual methods by reducing estimate errors by 28-91% to a level of  $1.2 - 9.4 \text{ kcal mol}^{-1}$  accuracy. We also compare our iterative refinement approach to other statistical ensemble approaches and demonstrate that this process reduces estimate errors by 25%-61%. This work provides a new approach for accurate solvation free energy prediction and lays the foundation for future work on aggregate models that can balance computational cost with predictive accuracy.

## Introduction

Accurate calculation of solvent-solute interactions is an important component of robust molecular simulation including protein structure prediction,<sup>1-3</sup> conformational ensemble calculations,<sup>4-7</sup> and binding free energy calculations.<sup>8-10</sup> Solvation energy methods for atomically detailed molecular models have a long history of development<sup>11-19</sup> and have recently benefited from the curation of experimental small molecule solvation data for blind prediction challenges such as the Statistical Assessment of Modeling of Proteins and Ligands

(SAMPL) challenge studies.<sup>10,20-25</sup> By presenting a larger range of solvation free energies and molecular weights than seen in common public data sets, these challenges are helping to advance the development of solvation methods to become more robust and accurate in their estimates for a variety of molecular targets.<sup>26-28</sup>

Consistent across many of these challenge studies is the observation that the top-performing methods typically come from a wide range of different strategies.<sup>10,20,21,25</sup> Across different challenges, top performers have included explicit solvation methods,<sup>22,29,30</sup> implicit solvation methods,<sup>4,31</sup> and hybrid methods that combine mixed quantum mechanics (QM) with molecular mechanical (MM) approaches.<sup>32,33</sup> In this context, there is a significant degree of uncertainty associated with how to best select, specify, and evaluate the set of parameters and mathematical systems needed to accurately estimate solvation free energies: e.g., uncertainties due to hydrophobicity,<sup>34</sup> surface effects,<sup>35</sup> and solvent asymmetries.<sup>36</sup> This type of *method selection uncertainty* affects a wide range of scientific and mathematical disciplines and is arguably the greatest source of error and risk associated with estimation tasks.<sup>37-40</sup> One of the most powerful ways to address this uncertainty is by combining an ensemble of varied methods (e.g., through a weighted average) to form a single aggregated estimate.<sup>41-44</sup> The motivation behind ensemble approaches is based on two principles: (1) most methods in the ensemble possess some unique, useful information and (2) no single method is sufficient to account for all uncertainties. Ensemble-based estimates rely on the combination of information and strengths from individual methods, with their corresponding weaknesses and biases overcome by the strength of the group, resulting in more accurate and robust predictions. Ensemble-based estimates are therefore expected to be more reliable and accurate than individual methods, an expectation that has been upheld in numerous examples.<sup>41-44,44-48,48-52</sup>

This paper demonstrates the ability of an ensemble approach called Bayesian Model Averaging (BMA)<sup>44</sup> to estimate solvation free energies for 45 small molecules by combining predictions from 17 diverse methods provided by the SAMPL4 challenge dataset.<sup>25</sup> Though BMA has been applied successfully for prediction tasks in many other domains<sup>45,47,49,50,53</sup>

this is the first application of the BMA approach solvation energy prediction.

## Methods

Throughout this paper, we use the term “methods” to describe the individual approaches to calculate solvation energies and the term “models” to describe statistical ensembles of method predictions.

### Model Specification with Bayesian Model Averaging

A basic BMA approach is to consider a set of methods as a linear system.<sup>44,51,52</sup> Let  $y_i$  be a series of solvation free energy observations for a collection of  $N$  molecules and let  $x_{ij}$  denote the  $i$ -th estimate obtained from the  $j$ -th of  $P$  prediction method for these observations. For example, given that  $y_i$  is the experimentally measured solvation free energy of benzaldehyde, each  $x_{ij}$  for  $j = 1, \dots, P$  would be a specific method’s estimate for this value. Given  $P$  solvation methods, the combination of all  $x_{ij}$  forms the numerical ensemble estimate matrix that, along with  $y_i$ , defines a linear regression model

$$y_i = \sum_{j=1}^P x_{ij} \beta_j + \epsilon_i, \tag{1}$$

where the parameter vector  $\beta_j$  defines the unknown relationship between the ensemble’s  $P$  constituents and  $\epsilon_i$  is a disturbance term that captures all factors (e.g., noise and measurement error) that influence the dependent variable  $y_i$  *other* than the regressors  $x_{ij}$ . The objective is to estimate the values  $\beta_j$  that will fit the known solvation free energy data in  $y_i$  and facilitate the ability to predict the solvation free energy of unknown molecules. Many different regression techniques can estimate  $\beta_j$ ;<sup>54–57</sup> however, these techniques commonly generate estimates that vary in their ability to model and predict.<sup>44,51,52,58,59</sup> The risk and uncertainty associated with using one solvation energy method’s estimates over any other

method is called *statistical model uncertainty*. Like method selection uncertainty, a common source of error in predictive modeling.<sup>44,49,51,52,60</sup>

BMA addresses the challenge of statistical model uncertainty by first evaluating all  $2^P - 1$  possible statistical models that can be formed from the  $P$  estimation methods and combining each model’s  $\beta_j$  through a weighted average into an aggregated parameter vector,  $\beta_j^{\text{BMA}}$  (Equation 2). There are  $k = 1, \dots, 2^P - 1$  distinct combinations of the  $P$  estimation methods, each with a corresponding statistical model,  $M^{(k)}$ , and parameter vector,  $\beta_j^{(k)}$ . BMA combines each  $\beta_j^{(k)}$  through an average that weights each  $\beta_j^{(k)}$  by the probability that its statistical model,  $M^{(k)}$ , is the “true” model:

$$\beta_j^{\text{BMA}} = E[\beta_j | \mathbf{y}] = \sum_{k=1}^{2^P-1} E[\beta_j^{(k)} | \mathbf{y}, M^{(k)}] \Pr(M^{(k)} | \mathbf{y}) \quad (2)$$

where  $E[\beta_j^{(k)} | \mathbf{y}, M^{(k)}]$  is the expected value of the posterior distribution of  $\beta_j^{(k)}$ . This distribution is weighted by the posterior probability  $\Pr(M^{(k)} | \mathbf{y})$  that  $M^{(k)}$  is the *true* statistical model given the data  $\mathbf{y}$ . The expected posterior distribution of  $\beta_j^{(k)}$  is approximated through the linear least squares solution of the given model  $M^{(k)}$  and solvation energy response variable  $\mathbf{y}$ . The posterior probability term is estimated from information criteria<sup>52</sup>

$$\Pr(M^{(k)} | \mathbf{y}) \propto \frac{e^{-\frac{1}{2}B^{(k)}}}{\sum_{l=1}^{2^P-1} e^{-\frac{1}{2}B^{(l)}}} \quad (3)$$

where  $B^{(k)}$  is the Bayesian Information Criteria for model  $M^{(k)}$  estimated as

$$B^{(k)} \approx N \log(1 - R^{2(k)}) + p^{(k)} \log N. \quad (4)$$

Here  $R^{2(k)}$  is the adjusted  $R^2$  for model  $M^{(k)}$  that indicates the model’s goodness of fit for the observations,  $p^{(k)}$  is the number of methods used by the model (not including the intercept), and  $N$  is the number of solvation free energy values to be predicted (i.e., the number of molecules).

The parameter vector  $\beta_j^{\text{BMA}}$  obtained from Equation 2 helps to address model uncertainty by accounting for all systems of linear equations that can model the relationship between the measured solvation free energy values  $y_i$  and values  $x_{ij}$  predicted by each solvation method  $j$ . Over-fitting by more complicated models is addressed through the Bayes information criterion which penalizes models with more variables. Perhaps more importantly,  $\beta_j^{\text{BMA}}$  can be used to estimate new solvation free energy values for unmeasured molecules by combining new  $x_{ij}$  estimates.

## Ensemble Pruning and Statistical Design

The inclusion of all  $2^P - 1$  models in Equation 2 is not necessarily beneficial for predictive performance. While some models provide accurate information that will boost the ensemble’s predictive accuracy and robustness, many models will be misspecified. The cumulative effect of these misspecified models, despite the fact that they are down-weighted via low posterior probabilities, can erode the ensemble’s overall performance.<sup>44,47,51,61–64</sup> Madigan and Raftery<sup>64</sup> developed the Occam’s Window ensemble-pruning approach that eliminates under-performing models based on Bayesian information criteria. Occam’s Window defines a set of models

$$\mathbf{A} := \{M^{(k)} \in \mathbf{M} | BIC^{(k)} - BIC^{(min)} < 6\}, \quad (5)$$

where  $BIC^{(min)}$  denotes the BIC of the model  $M^{(k)}$  with the lowest  $BIC$ ; low BIC indicates higher information content. The value 6 is based on Jeffreys’<sup>65</sup> and Raftery’s<sup>52</sup> assessment of Bayes factors for model comparison and ensures all  $m_k \in \mathbf{A}$  meet a minimum statistical information criteria for the aggregation process. Constraining Equation 5 to the set  $\mathbf{A}$  accelerates the evaluation of Equation 5 and improves BMA’s predictive capability:<sup>51,64</sup>

$$\beta_j^{\text{BMA}} = E[\beta_j | \mathbf{y}] = \sum_{M^{(k)} \in \mathbf{A}} E[\beta_j^{(k)} | \mathbf{y}, M^{(k)}] Pr(M^{(k)} | \mathbf{y}) \quad (6)$$

BMA estimates method  $j$ ’s utility for explaining a set of observations,  $\mathbf{y}$ , by assessing

---

**Algorithm 1**  
 Ensemble Design
 

---

**Require:** vector  $\mathbf{y}^{n \times 1}$ , matrix  $\mathbf{x}^{n \times p}$

```

1:  $r \leftarrow 1$ 
2:  $rmse \leftarrow \infty$ 
3:  $\mathbf{x\_solution} \leftarrow \mathbf{x}$ 
4:  $\beta_{BMA}^{1 \times j} \leftarrow \mathbf{0}^{1 \times j}$ 
5: while ( $r < p$ ) do
6:    $\beta_{BMA}, s, \mathbf{v} \leftarrow AssessMethods(\mathbf{y}, \mathbf{x})$ 
7:   if ( $s < rmse$ ) then
8:      $\mathbf{x\_solution} \leftarrow \mathbf{x}$ 
9:      $rmse \leftarrow s$ 
10:  end if
11:   $m \leftarrow 1$ 
12:  for ( $methods \in \mathbf{x}$ ) do
13:    if ( $\mathbf{v}[methods] < \mathbf{v}[m]$ ) then
14:       $m \leftarrow methods$ 
15:    end if
16:  end for
17:   $\mathbf{x} \leftarrow \mathbf{x.delete}(m)$  {remove the least informative method}
18:   $r \leftarrow (r + 1)$ 
19: end while
20: return  $\mathbf{x\_solution}, \beta_{BMA}, rmse$ 

```

---



---

**Algorithm 2**  
 AssessMethods
 

---

**Require:** vector  $\mathbf{y}^{n \times 1}$ , matrix  $\mathbf{m}^{n \times j} : j \leq p$

```

1:  $cv \leftarrow 100$  {perform 100 rounds of 2-fold cross-validation}
2:  $\beta_{BMA}^{1 \times j} \leftarrow \mathbf{0}^{1 \times j}$ 
3:  $error \leftarrow 0$ 
4:  $\mathbf{v}^{1 \times j} \leftarrow \mathbf{0}^{1 \times j}$ 
5: while ( $index < cv$ ) do
6:    $\mathbf{m}_t^{q \times j}, \mathbf{m}_v^{r \times j} \leftarrow \mathbf{m}^{n \times j}$  {split  $\mathbf{m}$  into train and validate}
7:    $\beta_{BMA} \leftarrow \beta_{BMA} + BMA(\mathbf{m}_t)$  {find  $\beta$ : Equation 6}
8:    $error \leftarrow error + RMSE(\mathbf{Y}, \mathbf{m}_v \times \beta_{BMA})$ 
9:    $\mathbf{v} \leftarrow \mathbf{v} + P \neq 0(\mathbf{m}_t)$  {assess method utility: Equation 7}
10:   $index \leftarrow index + 1$ 
11: end while
12: return  $\frac{\beta_{BMA}}{cv}, \frac{error}{cv}, \frac{\mathbf{v}}{cv}$ 

```

---

the probability that the method's coefficient term,  $\beta_j^{BMA}$  will receive a non-zero value. The estimate of this probability is based on the conditioned, cumulative sum of all model posteriors

$$\Pr(\beta_j^{BMA} \neq 0) = \sum_{M^{(k)} \in \mathbf{A}} \Pr(M^{(k)} | \mathbf{y}) I(j) \quad (7)$$

where

$$I(j) := \begin{cases} 1, & \text{if model } M^{(k)} \text{ specifies } j \text{ as a regressor} \\ 0, & \text{otherwise.} \end{cases} \quad (8)$$

Equation 7 can be used to sort and prioritize the methods based on their utility; i.e., the probability that the coefficient term weighting a method's estimate will not be 0. The combination of Equations 6 and Equation 7 therefore provide a statistical framework to support an iterative, statistical process for designing an ensemble to identify the best combination of models and methods that can be used to construct an aggregate estimate for solvation free energy. This design process is shown in Algorithm 1. The process assumes an initial set of  $n$  estimates made by  $p$  methods,  $\mathbf{x}^{n \times p}$  for observations,  $\mathbf{y}^{n \times 1}$ . The second and third lines

initialize the root mean squared error (RMSE) and the solution for the best ensemble of models and methods. The algorithm will iterate over the following tasks while the algorithm still has more than two methods to evaluate (line 5 - line 19).

First, the performance of the current ensemble is evaluated through *AssessMethods* (line 6). The *AssessMethods* function, shown in Algorithm 2, takes as input the same observation data,  $\mathbf{y}$ , as Algorithm 1, as well as a set of  $N$  estimates made by a subset of  $j \leq p$  methods. Algorithm 2 performs 100 iterations of a 2-fold cross-validation and begins by partitioning the estimate matrix  $\mathbf{m}$  equally into training and validation data (line 6). Using the training data, the algorithm uses Equation 6 to estimate  $\beta_{BMA}$  and then calculates the RMSE of this coefficient vector based on the validation data (lines 7 and 8). Next, the  $\Pr(\beta_j^{BMA} \neq 0)$  of each method are calculated and saved in the vector,  $\mathbf{v}$  (line 9). After 100 iterations, the function returns an estimate for  $\beta_j^{BMA}$ , the mean RMSE of the current ensemble, as well as a list of all methods and their corresponding probability values for  $\Pr(\beta_j^{BMA} \neq 0)$ . Next, Algorithm 1 compares and conditionally updates the current "best" performing model, **x\_solution**, with the model returned by *AssessMethods* (lines 8 and 9). The algorithm then identifies the method with the lowest  $\Pr(\beta_j^{BMA} \neq 0)$  and removes this method from the current model (line 17). With this method removed, the process repeats until there are just two methods left. The output of this process is an ensemble of methods,  $\mathbf{x}_{ij}$  that are statistically determined to be the best methods for estimating the observations in  $\mathbf{y}$  and a statistical model  $\beta_{BMA}$  that specifies how to best combine these methods.

## Solvation free energy data and solvation methods

The SAMPL4 challenge consists of 49 submissions representing a total of 19 different research groups.<sup>25</sup> Each of these methods provides solvation free energy estimates for 45 small molecule compounds. The challenge is blind in the sense that the solvation free energy values for these molecules were hidden from participants; e.g., energy values are not found in standard solvation free energy test sets, and their values are not readily available in the



literature.<sup>66</sup> In this work, we restrict our analysis to a subset of 17 method submissions based on the fact that many groups made multiple submissions that were strongly correlated. In these cases, we chose only a single variant to ensure that multicollinearity did not inflate the significance of specific methods during model selection and averaging; such bias can create unstable estimates for  $\beta_j^{\text{BMA}}$  that can reduce BMA’s predictive accuracy.<sup>67</sup> These methods are summarized in Table I based on the methods used to calculate the solvation energy:

- **Group 1:** Single-conformation implicit solvent methods<sup>26,68,69</sup> based on Poisson-Boltzmann and related methods;<sup>70–72</sup>
- **Group 2:** Multi-conformational implicit solvent methods;<sup>73–77</sup>
- **Group 3:** Methods based on molecular dynamics-based free energy calculations in explicit solvent<sup>22,27,30,78</sup> with small molecule force fields;<sup>79</sup> and,
- **Group 4:** Hybrid solvent methods.<sup>80</sup>

From this initial set of 17 methods, we iteratively assessed and pruned methods as described in Algorithm 1. The approach began by randomly sampling (without replacement) 26 of the original 52 experimental solvation free energy measurements to form training and validation datasets (Algorithm 2, line 6). Collectively, these sampled values formed the observation vector  $y_i$  and the estimates from each of the prediction methods in Table I for these measurements defined the ensemble estimate matrix,  $x_{ij}$ . The observation vector and the ensemble estimate matrix were used to form the linear system in Equation 1.

Next, we estimated the  $\beta_j^{\text{BMA}}$  parameter from Equation 2 by assessing all possible  $(2^{17} - 1)$  statistical models  $M^{(k)}$ . Each model’s information criteria  $B^{(k)}$  was used to identify a reduced set of most informative models per Equation 5. Based on this reduced modeling space, the coefficient terms  $\beta_j^{\text{BMA}}$  were estimated through weighted averages of each statistical model’s ordinary least squares solution (Equation 6). We used  $\beta_j^{\text{BMA}}$  to estimate the remaining 26 solvation free energy measurements that were *not* used to train the BMA model. This task

Table I: This table lists the solvation methods used in our ensemble design process. Method ID indicates the identification number of the method that is referenced throughout this paper. Sampling strategies include quantum mechanical (QM), molecular dynamics (MD), and molecular mechanics with Poisson-Boltzmann surface area solvation (MM-PBSA). The listed performance is based on 100 iterations of a 2-fold cross-validation study as described in the text. This performance is also shown graphically in Figure 1. The last column is a comparison of each method to the optimal BMA ensemble, BMA (Stage 16). This column indicates that the ensemble design approach presented in this work is able to reduce estimation errors by 29% to 91% (i.e., 1.2 – 9.4 kcal mol<sup>-1</sup>) in comparison to the individual methods. The final BMA ensemble is indicated by the two blue highlighted methods: imp-2 and alc-3. Wilcoxon based *p*-values for BMA’s mean RMSE distribution vs. the best method’s mean RMSE distribution (imp-2) are shown in Table II.

Reference	Method ID	Methodology	Ensemble Mean RMSE (kcal mol <sup>-1</sup> ) and Standard Deviation	Performance Improvement Provided by BMA (Stage 16)
Coleman et al. <sup>81</sup>	imp-6	multi-conformation implicit	9.40 ± 2.03	91%
Parsod et al.	alc-5	alchemical MD	4.50 ± 0.71	82%
Sharp et al. <sup>82</sup>	imp-4	single conformation implicit	3.80 ± 0.45	78%
Jiafu et al.	exp-3	QM/MD	2.80 ± 0.45	71%
Purisima et al. <sup>75</sup>	imp-5	single conformation implicit	2.55 ± 0.49	68%
Mark et al. 2011	alc-2	alchemical MD	2.40 ± 0.36	66%
Genheden et al. <sup>83</sup>	exp-1	MM-PB/SA	2.00 ± 0.22	59%
Weyang et al.	exp-4	MM-PB/SA	1.84 ± 0.31	55%
Biorga et al. <sup>84,85</sup>	alc-4	alchemical MD	1.65 ± 0.13	50%
Elingson et al. <sup>26</sup>	imp-7	single conformation implicit	1.52 ± 0.39	46%
Fennell et al. <sup>80</sup>	exp-2	hybrid	1.52 ± 0.13	46%
Jambeck et al. <sup>86</sup>	alc-1	alchemical MD	1.52 ± 0.20	46%
Park <sup>87</sup>	imp-3	single conformation implicit	1.44 ± 0.20	43%
Klamt et al. <sup>88</sup>	imp-1	single conformation implicit	1.36 ± 0.30	40%
Gilson et al. <sup>27</sup>	alc-3	alchemical MD	1.24 ± 0.17	34%
Geballe et al. <sup>26</sup>	imp-8	single conformation implicit	1.17 ± 0.17	30%
Sandberg et al. <sup>73</sup>	imp-2	multi-conformation implicit	1.15 ± 0.23	29%
BMA (Stage 16)	NA	ensemble	0.82 ± 0.15	0

was accomplished by combining the estimates of all methods in Table I for the validation data with  $\beta_j^{\text{BMA}}$  to produce an aggregated estimate. A root-mean-squared error was obtained for the 26 estimates made on validation data.

This process was repeated 100 times (Algorithm 2) so that performance for any estimation method could be reported as a mean RMSE: i.e., the mean of 100 RMSEs that each represent performance for a 2-fold cross-validation using the 26 member validation set described above. In addition to the mean RMSE, the information content provided by each method in the ensemble is also returned.

Algorithm 1 used this information to perform two tasks. First, if the performance of this

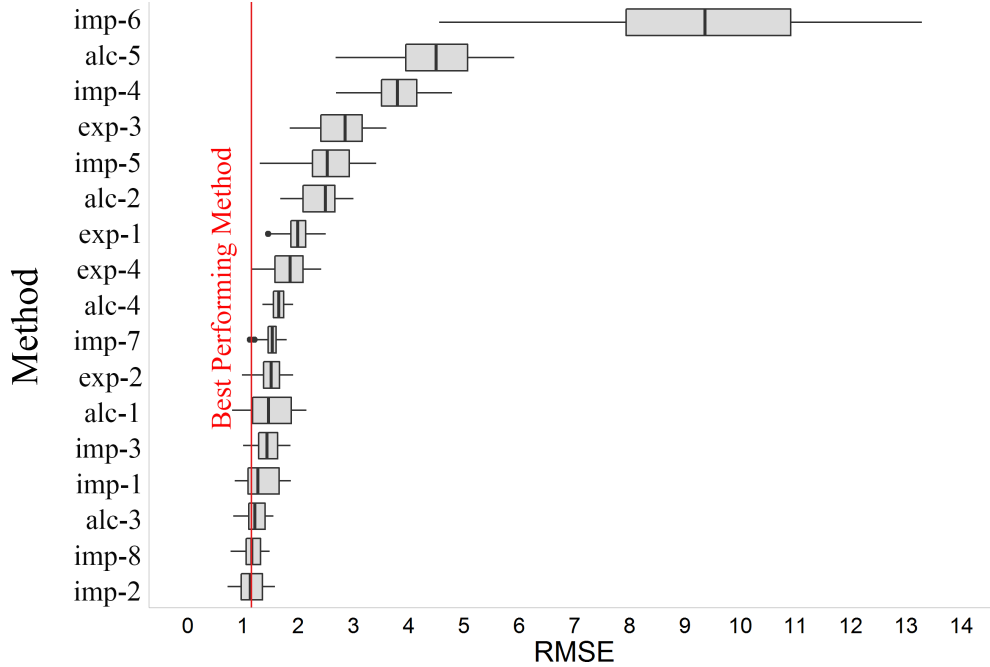


Figure 1: This figure depicts the mean root mean squared error, min, max, first and third quartiles for the 17 initial methods used in our ensemble design process. Method performance is based on 100 iterations of the 2-fold cross-validation experiment detailed in Algorithm 2. The red line is used to indicate the mean performance of the best method, imp-2; this line is referenced again in Figure 2 to show how the ensemble tuning process compares to the performance of this best solvation method.

aggregated estimate was better than all previously examined aggregates, the statistical model combining these methods was saved as the new optimal model (Algorithm 1, lines 7-9). Next, Algorithm 1 pruned method from the ensemble that provided the least amount of information to the aggregated estimate. The pruning process was repeated until just two methods are left (Algorithm 1, lines 12-18). The ensemble of methods whose aggregate forecast has the lowest mean RMSE was saved and returned as the final model for the ensemble. This final model, and the set of methods that correspond to this model, were the final products of the statistically driven ensemble design process, referred to as the BMA-based optimal ensemble.

## Results and Discussion

We report the results of our BMA ensemble predictions in three stages. First, we contrast the performance of the initial 17 SAMPL4 methods to the performance of the BMA-based optimal ensemble. We then present data on the iterative design process used to create this optimal ensemble. Second, we examine the *conditional* performance of this optimal ensemble according to the individual molecules used in the SAMPL4 challenge. In this stage, we show how the BMA-based optimal ensemble provides more reliable estimates in comparison to individual methods, especially against the more challenging set of small molecules from the SAMPL4 dataset. Finally, we complete the analysis of the optimal ensemble by contrasting its performance to the performance of alternate statistical techniques that can be used to form aggregated estimates.

The performance of all methods is based on a mean RMSE. We report the statistical significance of all performance data through a Wilcoxon rank sum paired comparison test.<sup>89</sup> This non-parametric approach tests the hypothesis that the mean RMSE distributions of two approaches are equal:  $H_0 : \mu_Y = \mu_X$ . To control the family-wise error rate of our tests, we applied a Bonferroni correction to determine corrected  $p$ -values with a threshold of  $\alpha = 0.05$ . Thus when comparing BMA to a given method, a Wilcoxon generated  $p$ -value greater than 0.05 indicates we fail to reject  $H_0$ : the distributions are thus equal and we conclude that BMA and the method are equivalent in their performance. On the other hand, Wilcoxon-generated  $p$ -values that are less than 0.05 indicate we should reject  $H_0$ . In this latter case, we then compare the mean RMSE for BMA and the given method to assess performance.

### Comparing estimates from BMA’s optimal ensemble to SAMPL4 challenge methods

The performance of all methods used in this work is shown in Table I and Figure 1. Column 3 in Table I lists the specific method for each estimation approach. The performance results in

column 4 of this table are consistent with results reported by Mobley et al.<sup>25</sup> The performance of the optimal ensemble is shown in the last row. The corresponding methods that are constituents in this optimal ensemble, “alc-3” and “imp-2”, are highlighted in blue. The final column in Table I lists a direct comparison of each method’s performance to the performance of the optimal ensemble: e.g., the ensemble reduces estimation errors by as much as 91% in comparison to imp-6 and by 29% in comparison to imp-2.

The box plots in Figure 1 illustrate the performance variability across the different methods. These plots also underscore the amount of uncertainty inherent in estimating solvation free energies: method-selection uncertainty plays a confounding factor in estimating solvation free energies. Finally, the red line indicates the mean RMSE of the best-performing method: imp-2. This line is also used in Figure 2 to contrast the iterative improvements obtained during the optimal ensemble’s design process. The ensemble’s iterative design process based on Algorithms 1 and 2 is shown in Table II and Figure 2. The process starts with the 17 initial methods shown in Table I. Each subsequent row in Table II represents an iteration through the pruning process. The second column lists the mean root mean squared error (RMSE) of each stage’s ensemble based on its aggregated estimate. At the end of each stage, a method is selected to be pruned from the existing ensemble before proceeding to the next iteration; the specific method that was selected is shown in column 3. For example, at Stage 1 there were 17 methods in the ensemble and imp-6 was selected to be removed. During the next step, Stage 2, imp-6 was removed from the ensemble so that there are only 16 methods used to create an aggregated estimate. At the end of this stage, alc-4 was selected to be pruned for Stage 3. In the final stage, the only remaining methods in the ensemble were imp-2 and alc-3. This iterative design process is also graphically illustrated in Figure 2 showing how selective pruning increases the performance of each successive ensemble. The red line in this figure indicates the performance of the best performing method, imp-2. The benefits of the aggregated estimates become apparent after Stage 6 where the ensembles outperform imp-2.

The statistical significance of the iterative design process is listed in columns 4 and 5 in

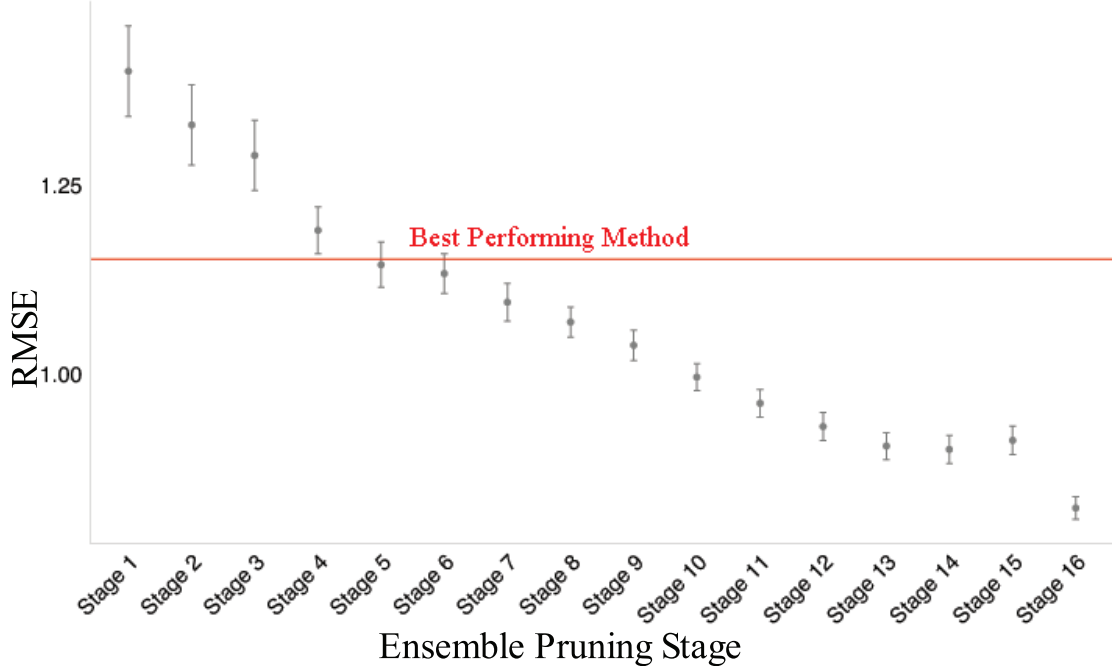


Figure 2: This plot illustrates the iterative pruning process discussed in Section . The y-axis depicts the mean root mean squared error (RMSE) of the different aggregated estimates based on ensembles formed during the different iterations of pruning. The x-axis indicates the stages of pruning. In general, the variance and overall mean RMSE reduces with each iteration. The performance of the different ensembles are compared to the best performing method through the red line at  $y = 1.15$ ; all iterations past Stage 7 outperform the best method in the ensemble. Based on Wilcoxon generated p-values, the significance in the distributions of mean RMSE between the different ensembles are presented in Table II. Based on mean RMSE, the optimal ensemble is created at Stage 16.

Table II; bold  $p$ -values in these columns indicate the performance between two distributions are equivalent. Column 4 lists the Wilcoxon-generated  $p$ -values based on comparisons of mean RMSE distributions obtained from sequential ensembles. For example, in the second row the  $p$ -value for the comparison of Stage 2 vs. Stage 1 indicate that these distributions are equivalent. Contrariwise, in row 4, the  $p$ -value for the comparison between Stage 4 and Stage 3 indicate that the distributions are not equivalent; the mean RMSE in column 2 indicates that the ensemble of Stage 4 outperforms the ensemble built in Stage 3.

As a second analysis of significance, column 5 lists the Wilcoxon-generated  $p$ -values that represent comparisons between each ensemble and the best-performing method, imp-2. From these values, Stages 4-6 are seen to be equivalent to imp-2. However, the mean RMSE distri-

Table II: This table lists the performance of the aggregated estimates obtained from the different ensembles created during the design process in Section . The second column lists the mean root mean squared error (RMSE) for each ensemble’s aggregated estimate based on 100 iterations of a 2-fold cross-validation; this performance is also shown in Figure 2. The third column lists the method that was selected to be pruned from the ensemble at the *next* stage of the design process. The Wilcoxon generated p-values in column four are based on comparisons of mean RMSE distributions obtained from sequential ensembles and their aggregated estimates. *p*-values that are greater than 0.05 are shown and indicate distributions that are equivalent (e.g., Stage 3 and 2) based on an  $\alpha = 0.05$ . Similarly, column five lists the Wilcoxon generated p-values reflecting comparisons between each stage and the best performing method, imp-2. As with column 4, bold *p*-values indicate that the performance between imp-2 and a given stage are equivalent (e.g., Stages 4-6). Based on mean RMSE and p-values from this table, the optimal ensemble is the one created in Stage 16; the final column in this table lists the performance improvement this ensemble provides in comparison to the ensembles generated in previous stages.

Stage	Ensemble mean RMSE (kcal mol <sup>-1</sup> ) ± Standard Deviation	Method Selected To Be Pruned	p-value (sequential stages)	p-value (stage vs. imp-2)	Performance Benefit of BMA (Stage 16)
1	1.40 ± 0.60	imp-6	NA	0.000	41%
2	1.32 ± 0.53	alc-4	<b>0.411</b> (vs. Stage 1)	0.002	38%
3	1.29 ± 0.46	imp-8	<b>1.000</b> (vs. Stage 2)	0.004	36%
4	1.19 ± 0.31	imp-5	0.012 (vs. Stage 3)	<b>0.356</b>	31%
5	1.14 ± 0.29	imp-3	0.000 (vs. Stage 4)	<b>0.438</b>	28%
6	1.13 ± 0.26	imp-7	<b>0.883</b> (vs. Stage 5)	<b>0.306</b>	27%
7	1.09 ± 0.25	alc-5	0.015 (vs. Stage 6)	0.027	25%
8	1.06 ± 0.19	alc-2	<b>0.055</b> (vs. Stage 7)	0.003	23%
9	1.03 ± 0.20	imp-1	<b>1.000</b> (vs. Stage 8)	0.001	20%
10	0.99 ± 0.17	imp-4	0.000 (vs. Stage 9)	0.001	17%
11	0.96 ± 0.18	alc-1	0.000 (vs. Stage 10)	0.000	15%
12	0.93 ± 0.18	exp-4	<b>1.000</b> (vs. Stage 11)	0.000	12%
13	0.90 ± 0.17	exp-2	<b>1.000</b> (vs. Stage 12)	0.000	09%
14	0.90 ± 0.18	exp-1	<b>1.000</b> (vs. Stage 13)	0.000	09%
15	0.91 ± 0.18	exp-3	0.001 (vs. Stage 14)	0.000	10%
16	0.82 ± 0.15	NA	0.000 (vs. Stage 15)	0.000	0%

bution of all subsequent stages are not equivalent: based on mean RMSE listed in column 2, we conclude that these successive ensembles (increasingly) outperform imp-2. The p-values in these columns and the mean RMSE results demonstrate that the optimal ensemble is created in Stage 16. The final column in Table II lists the performance improvement that the Stage 16 ensemble provides in comparison to each other ensemble that was generated in the design process.

Figure 3 depicts a heat-map that shows the statistical information that drove the pruning process for each stage. In this image, the y-axis represents the different methods; the x-axis

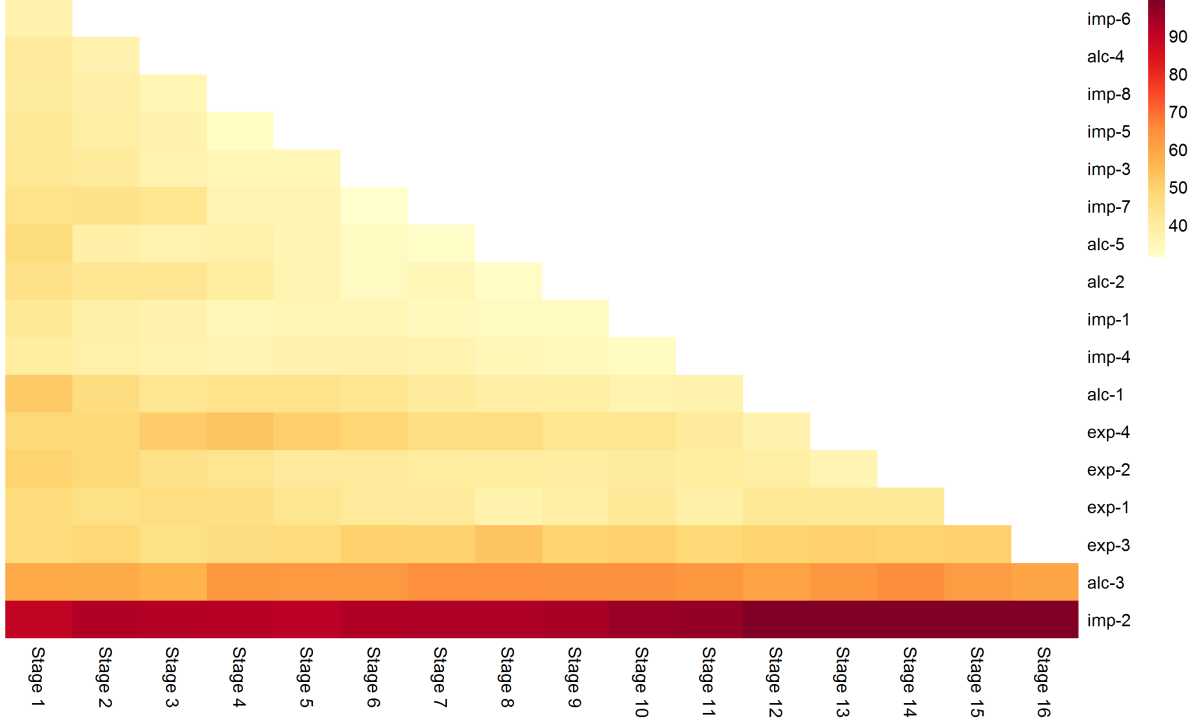


Figure 3: This image provides a graphical depiction of the ensemble design process performed on the initial ensemble of 17 methods. The color scale represents the probability, 0 - 100%, that a given method's coefficient term,  $\beta_j$  will not be zero. The stages in the design process, starting at the left and moving to the right, are shown on the x-axis. The methods in each ensemble are shown on the y-axis; note that the methods are listed (from top to bottom) in the order that they are pruned in the design process. Thus at Stage 1, all methods are used in the ensemble and their  $\Pr(\beta_j^{\text{BMA}} \neq 0)$  range from 40% (e.g., imp-6) to 90% (imp-2). By Stage 3, imp-6 and alc-4 have been pruned from the ensemble and the probability values have adjusted accordingly as shown in the colored column above Stage 3. In general the trend across the different stages of pruning illustrates that methods become increasingly lighter, i.e., the ensemble design process becomes increasingly confident that these methods (e.g., imp-7 and exp-4) are not needed in the ensemble. Contrariwise, the  $\Pr(\beta_j^{\text{BMA}} \neq 0)$  for a few methods (e.g., alc-3, and imp-2) remain above 70% and even increase throughout the design process indicating a high degree of confidence in the statistical significance of these methods.

(starting from left) indicates the successive stages in the design process. The color scale represents the mean probability, 0 - 100%, that a given method's coefficient term,  $\beta_j$  will not be zero. At Stage 1, all methods were used in the ensemble and their  $\Pr(\beta_j^{\text{BMA}} \neq 0)$  ranged from 40% (imp-6) to 80% (imp-2). As imp-6 had the lowest mean probability of not being zero, imp-6 was pruned after Stage 1; the color map colors this method white in Stage 2 to indicate it has been eliminated. The methods listed on the legend of the y-axis are ordered



by the sequence that they were eliminated in the design process: imp-6 first, alc-4 second, imp-8 third, etc.

## Performance Analysis Based on Compounds

Figures 4 and 5 depict performance of different methods according to specific SAMPL4 challenge small molecule compounds. In addition to the optimal ensemble, we also show the performance of the first-, second-, and third best-performing methods from the SAMPL4 challenge: imp-2, imp-8, and alc-3. Methods imp-2 and alc-3 are the methods used in the optimal BMA ensemble and exp-3 is the final method eliminated from the ensemble (Stage 15 in Table II).

The analyses of Mobley et al. identified certain compounds that were especially difficult to estimate in the SAMPL4 challenge: SAMPL4.022 (mefenamic acid), SAMPL4.023 (diphenhydramine), SAMPL4.027 (1,3-bis-(nitroxy)propane), SAMPL4.009 (2,6-dichlorosyringaldehyde), and SAMPL4.001 (mannitol).<sup>25</sup> Figures 4 and 5 show that the optimal ensemble outperforms all methods in estimating SAMPL4.022 and SAMPL4.001, and provides the second best estimates for SAMPL4.009, SAMPL4.023 and SAMPL4.027. Aside from the optimal ensemble, there is no clear best method for estimating these compounds. For example, while imp-8 is best at estimating SAMPL4.009, it does not do well at estimating either SAMPL4.023 or SAMPL4.027. Similarly while imp-2 performs well at estimating SAMPL4.023, it does not do as well at estimating SAMPL4.009 or SAMPL4.027. While there are certain compounds that challenge the ensemble (e.g., SAMPL4.017), the general trend in performance suggests that the optimal ensemble provides more consistent and accurate estimates than any specific method.

## Performance Analysis of Alternate Ensemble Techniques

There are other approaches besides BMA that can combine an ensemble of methods into an aggregate prediction. In our cross-validation study, we evaluated four common approaches

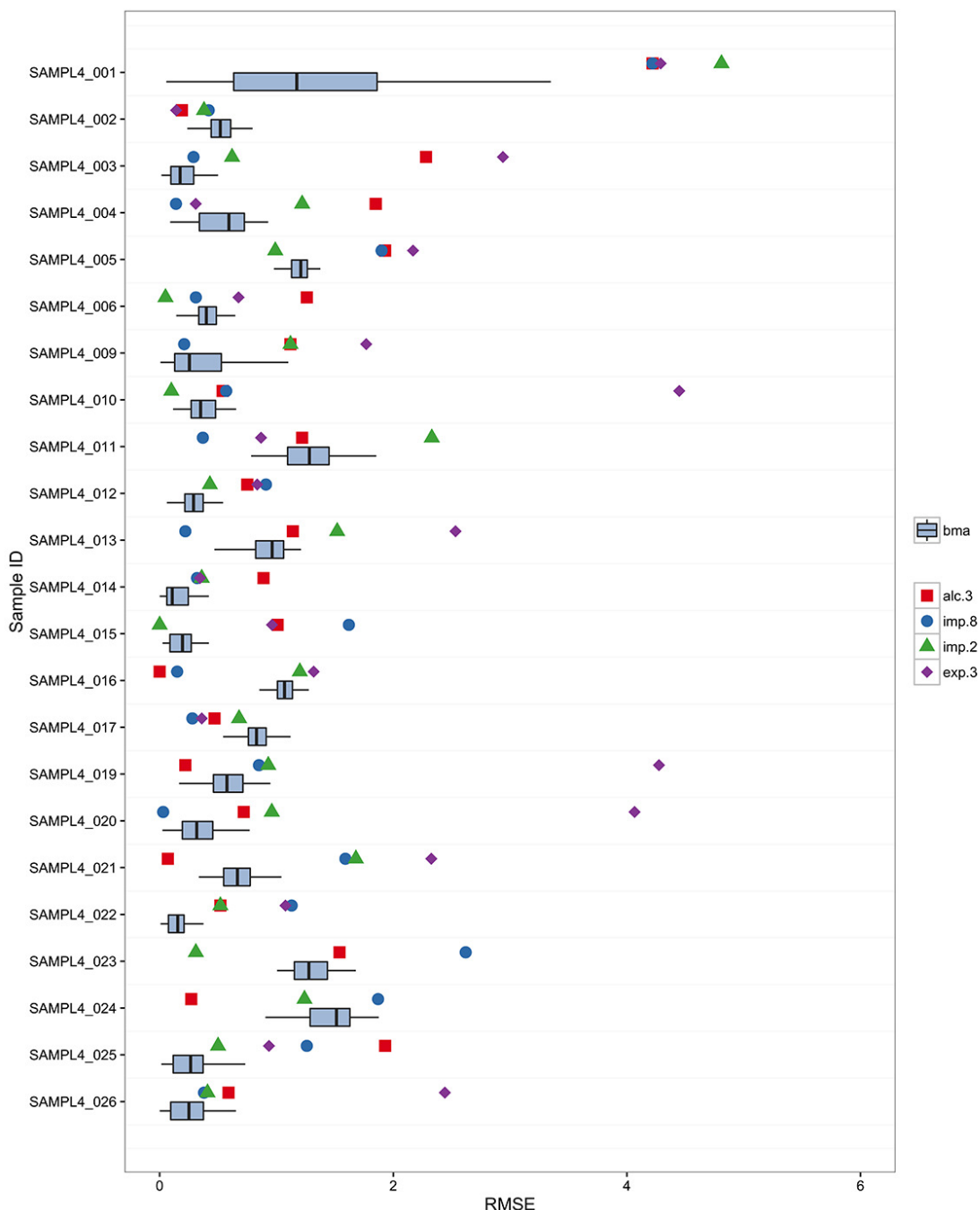


Figure 4: This figure is one of two figures (Figures 4 and 5) that depict the RMSE (kcal mol<sup>-1</sup>) performance of several methods based on the individual compounds taken from the SAMPL4 challenge: the first, second and third performing methods (i.e., imp-2, imp-8, and alc-3) as well as exp-3. Note that imp-2 and alc-3 are the methods used in the optimal BMA ensemble (Stage 15) and exp-3 is the final method eliminated from the ensemble (Table II) in Stage 16. BMA’s performance based on the optimal ensemble is shown based on its distribution of the mean root mean squared error for estimates made in our 2-fold cross-validation analysis. Of note is the performance of the different methods for SAMPL4.022 (mefenamic acid), SAMPL4.023 (diphenhydramine), SAMPL4.027 (1,3-bis-(nitroxy)propane), SAMPL4.009 (2,6-dichlorosyringaldehyde), and SAMPL4.001 (mannitol). These are the most challenging compounds for methods to estimate based on the analysis of Mobley et al. of the SAMPL4 data.<sup>25</sup> The benefits of the ensemble is clearly demonstrated here as the BMA ensemble outperforms all methods in estimating SAMPL4.022, SAMPL4.009, and SAMPL4.001. For SAMPL4.023 and SAMPL4.027 the ensemble provides

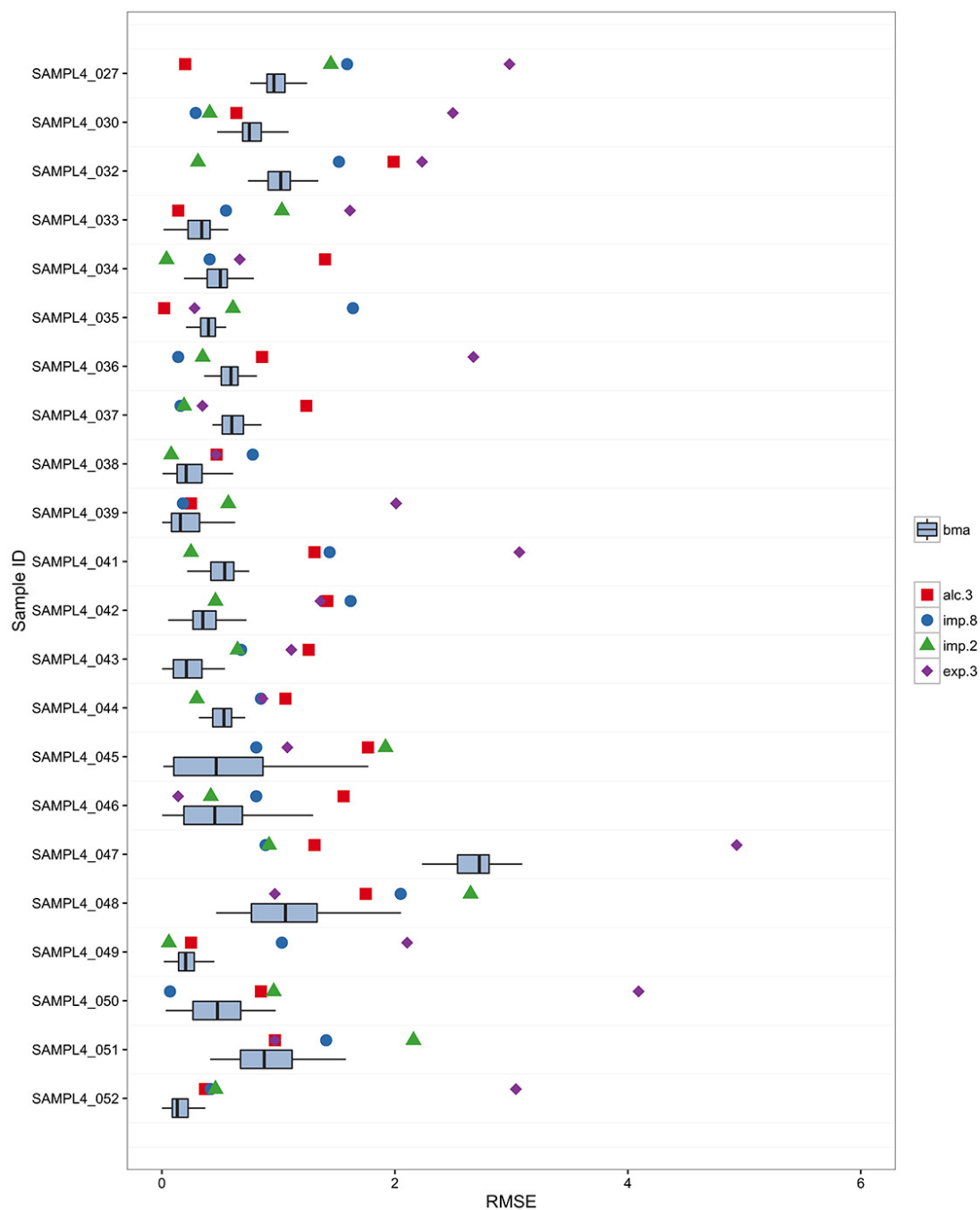


Figure 5: This figure is one of two figures (Figures 4 and 5) that depict the RMSE (kcal mol<sup>-1</sup>) performance of several methods based on the individual compounds taken from the SAMPL4 challenge: the first, second and third performing methods (i.e., imp-2, imp-8, and alc-3) as well as exp-3. See Figure 4 for more information about both plots.

for aggregating an ensemble and evaluated their predictive benefits in comparison to BMA. These methods are listed in Table III and include: Random Forest,<sup>90</sup> Ridge Regression,<sup>91</sup> Lasso,<sup>92</sup> and stepwise regression via forward selection. These techniques were chosen as they

**Table III:** This table lists the performance of different ensemble approaches in comparison to the optimal ensemble designed in this work through BMA. The performance of these ensemble approaches, given as the mean root mean squared error with standard deviation, are based on the 100 iterations of the 2-fold cross-validation experiment discussed in Section . Based on an  $\alpha = 0.05$ , the Wilcoxon based  $p$ -values indicate that BMA’s improved performance is statistically significant to the other ensemble approaches for combining methods to make an aggregated estimate. The last column indicates the improvement in estimation that the optimal ensemble provides to these alternate techniques: estimation accuracy is improved from 25% to 61%.

Ensemble Model	Ensemble Mean RMSE and Standard Deviation	p-value	Performance Improvement Provided by BMA (Stage 16)
Random Forest	2.08 $\pm$ 1.00	0.00	61%
Ridge	2.06 $\pm$ 0.75	0.00	60%
Lasso	1.12 $\pm$ 0.34	0.00	27%
Forward Selection	1.09 $\pm$ 0.26	0.00	25%
BMA (Stage 16)	0.82 $\pm$ 0.17	NA	0%

have all been used successfully for a variety of inference tasks and are readily available for use.<sup>93,94</sup> As all of these approaches construct an estimate for  $\beta$ , training and predicting with these approaches was performed identically to BMA, using the cross-validation detailed in Section . We also follow the same procedure for comparing BMA’s predictive capability to these alternate ensemble-based techniques. Figure 6 and Table III provide an overview of the cross-validation errors for the various ensemble-based approaches and the BMA-based optimal ensemble. The statistical significance of BMA’s performance in Figure 6 is based on  $p$ -values shown in Table III. Based on an  $\alpha = 0.05$ , Table III indicates that we reject the null hypothesis for all paired comparison tests. BMA’s mean RMSE distribution is therefore not equivalent to the mean RMSE distribution of any other ensemble-based technique. As the distributions are not equal, we compared mean RMSE distributions of BMA to the other ensemble-based approaches in Figure 6 and Table III. From these mean RMSE, it is clear that the BMA-based approach outperforms all other ensemble-based prediction approaches:

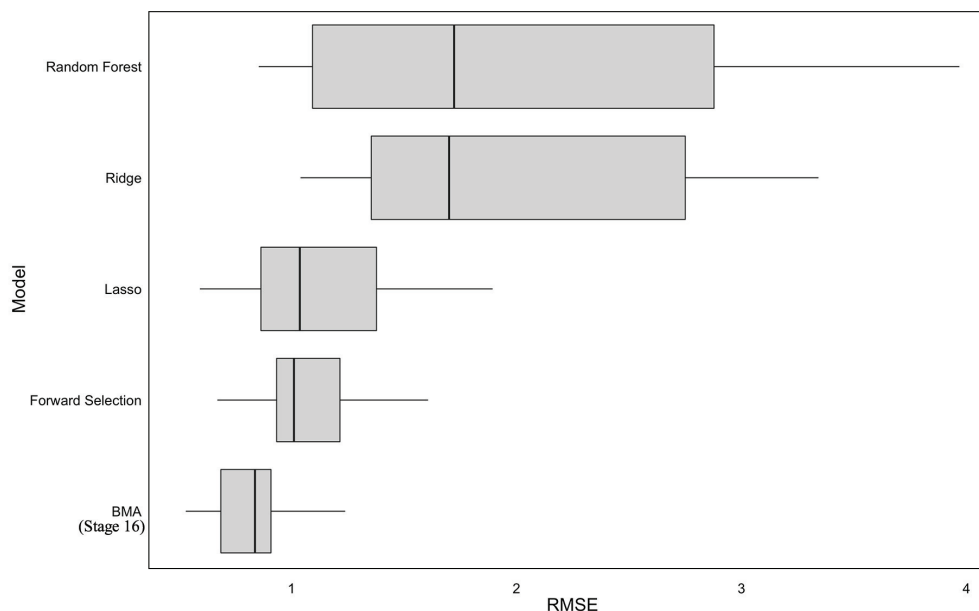


Figure 6: This figure displays the performance of different ensemble approaches in comparison to the optimal ensemble designed in this work, BMA (Stage 16). The mean root mean squared error, min, max, first and third quartiles ( $\text{kcal mol}^{-1}$ ) of these ensemble approaches are shown based on the 100 iterations of the 2-fold cross-validation experiment discussed in Section . Based on an  $\alpha = 0.05$ , the Wilcoxon based p-values (Table III) indicate that BMA’s improved performance is statistically significant to the other approaches that can combine an ensemble of methods to make an aggregated estimate.

BMA-based estimates reduced error by approximately 60% in comparison to Random Forest and Ridge regression methods. In comparison to Lasso, BMA reduces estimation error by approximately 27%. Finally, in comparison to stepwise regression via forward selection, BMA reduces error by approximately 25%.

## Conclusions

This study demonstrates a proof-of-principle application of how to statistically design and aggregate an ensemble of methods for estimating solvation free energies in small molecules. While the performance of BMA is expected to generalize to a much broader set of small molecule estimation problems, the specific BMA model trained in this study is likely to be dependent on the small molecules used in the SAMPL4 challenge. The BMA approach is purely statistical in nature, but could also be trained to modify the aggregation process based on structural and environmental features (e.g., only look at ensembles of empirical methods for certain structural features and consider all methods for other structures). In future work we will look at penalizing computationally expensive methods that provide minimal accuracy benefits to optimize both the accuracy of the estimates and the efficiency of the calculation.

## Acknowledgments

This research was funded by NIH grant R01 GM069702 to NAB.

★★ **Need to fix several of the references** ★★

## References

- (1) Levy, R.; Zhang, L.; Gallicchio, E.; Felts, A. On the Nonpolar Hydration Free Energy of Proteins: Surface Area and Continuum Solvent Models for the Solute - Solvent Interaction Energy. *J. AM. CHEM. SOC* **2003**, *125*, 9523–9530.

- (2) Robinson, G.; Cho, C. Role of Hydration Water in Protein Unfolding. *Biophys J* **1999**, *77*, 3311–3318.
- (3) Rakhmanov, S.; Makeev, V. Atomic hydration potentials using a Monte Carlo Reference State (MCRS) for protein solvation modeling. *BMC Struct Biol.* **2007**, 7–19.
- (4) Jorgensen, W.; Ulmschneider, J.; Tirado-Rives, J. Free Energies of Hydration from a Generalized Born Model and an All-Atom Force Field. *J. Phys. Chem. B* **2004**, *108*, 16264–16270.
- (5) Cui, Q.; Smith, V. Solvation Structure, Thermodynamics, and Molecular Conformational Equilibria for n-Butane in Water Analyzed by Reference Interaction Site Model Theory Using an All-Atom Solute Model. *J. Phys. Chem. B*, **2002**, *106*, 6554–6565.
- (6) Ashbaugh, H.; Garde, S.; Hummer, G.; Kaler, E.; Paulaitis, M. Conformational equilibria of alkanes in aqueous solution: Relationship to water structure near hydrophobic solutes. *Biophysical Journal* **1999**, *77*, 645–654.
- (7) Ashbaugh, H.; Kaler, E.; Paulaitis, M. Conformational equilibria of polar and charged flexible polymer chains in water. *Polymer* **2002**, *43*, 559–565.
- (8) Yang, C.; Sun, H.; Chen, J.; Nikolovska-Coleska, Z.; Wang, S. Importance of ligand reorganization free energy in protein-ligand binding-affinity prediction. *J Am Chem Soc.* **2009**, *131*, 13709–13721.
- (9) Whalen, K.; Spies, M. Flooding Enzymes: Quantifying the Contributions of Interstitial Water and Cavity Shape to Ligand Binding Using Extended Linear Response Free Energy Calculations. *J Chem Inf Model.* **2013**, *53*, 2349–2359.
- (10) Mobley, D.; Dill, K. Binding of Small-Molecule Ligands to Proteins: “What You See” Is Not Always “What You Get”. *Structure* **2009**, *17*, 489–498.

- (11) Eisenberg, D.; McLachlan, A. Solvation energy in protein folding and binding. *Nature* **1986**, *319*, 199–203.
- (12) Kang, Y.; Nemethy, G.; Scheraga, H. Free energies of hydration of solute molecules. 1. Improvement of the hydration shell model by exact computations of overlapping volumes. *J. Phys. Chem* **1987**, *91*, 4105–4109.
- (13) Kang, Y.; Nemethy, G.; Scheraga, H. Free energies of hydration of solute molecules. 2. Application of the hydration shell model to nonionic organic molecules. *J. Phys. Chem* **1987**, *91*, 4109–4117.
- (14) Kang, Y.; Nemethy, G.; Scheraga, H. Free energies of hydration of solute molecules. 3. Application of the hydration shell model to charged organic molecules. *J. Phys. Chem* **1987**, *91*, 4118–4120.
- (15) Kang, Y.; Gibson, D.; Nemethy, G.; Scheraga, H. Free energies of hydration of solute molecules. 4. Revised treatment of the hydration shell mode. *J. Phys. Chem* **1988**, *92*, 4739–4742.
- (16) Tan, C.; Yang, L.; Lou, R. How well does Poisson-Boltzmann implicit solvent agree with explicit solvent? A quantitative analysis. *J. Phys. Chem.* **2006**, *110*, 18680–18687.
- (17) Gallicchio, E.; Zhang, L.; Levy, R. The SGB/NP hydration free energy model based on the surface generalized born solvent reaction field and novel nonpolar hydration free energy estimators. *J. Comput. Phys.* **2002**, *23*, 517–529.
- (18) Thomas, D.; Chun, J.; Chen, Z.; Wei, G.; Baker, N. Parameterization of a Geometric Flow Implicit Solvation Model. *Journal of Computational Chemistry* **2013**, *34*, 687–695.
- (19) Lei, H.; Yang, X.; Zheng, B.; Baker, N. Constructing Surrogate Models of Complex Sys-



- tems with Enhanced Sparsity: Quantifying the influence of conformational uncertainty in biomolecular solvation. *Multiscale Modeling and Simulation* **2015**, *13*, 1327–1353.
- (20) Nicholls, A.; Mobley, D.; Guthrie, J.; Chodera, J.; Bayly, C.; Cooper, M.; Pande, V. Predicting small-molecule solvation free energies: an informal blind test for computational chemistry. *J. Med. Chem.* **2008**, *51*, 769–779.
- (21) Mobley, D.; Bayly, C.; Cooper, M.; Dill, K. Predictions of Hydration Free Energies from All-Atom Molecular Dynamics Simulations. *J Phys Chem B.* **2009**, *113*, 4533–4537.
- (22) Klimovich, P.; Mobley, D. Predicting hydration free energies using all-atom molecular dynamics simulations and multiple starting conformations. *J Comput Aided Mol Des.* **2010**, *24*, 307–316.
- (23) Geballe, M.; Guthrie, J. The SAMPL3 blind prediction challenge: transfer energy overview. *J Comput Aided Mol Des.* **2012**, *26*, 489–496.
- (24) Geballe, M.; Skillman, A.; Nicholls, A.; Guthrie, J.; Taylor, P. The SAMPL2 blind prediction challenge: introduction and overview. *J Comput Aided Mol Des* **2010**, *24*, 259–279.
- (25) Mobley, D. L.; Wymer, K. L.; Lim, N. M.; Guthrie, J. P. Blind prediction of solvation free energies from the SAMPL4 challenge. *J. Comput. Aided Mol. Des.* **2014**, *3*, 135–150.
- (26) Ellingson, B.; Gaballe, M.; Wlodek, S.; Bayly, C.; Skillman, A.; Nicholls, A. Efficient calculation of SAMPL4 hydration free energies using OMEGA, SZYBKI, QUACPAC, and Zap TK. *J. Comput. Aided Mol. Des.* **2014**, *28*, 289–298.
- (27) Muddana, H.; Sapra, N.; Fenley, A.; Gilson, M. The SAMPL4 hydration challenge: Evaluation of partial charge sets with explicit-water molecular dynamics simulations. *J Comput Aided Mol Des* **2014**, *28*, 277–287.

- (28) Fu, J.; Liu, Y.; Wu, J. Fast prediction of hydration free energies for SAMPL4 blind test from a classical density functional theory. *J Comput Aided Mol Des* **2014**, *28*, 299–304.
- (29) Levy, R.; Gallicchio, E. Computer simulations with explicit solvent: recent progress in the thermodynamic decomposition of free energies and in modeling electrostatic effects. *Annu Rev Phys Chem* **1998**, *49*, 531–567.
- (30) Mobley, D.; Bayly, C.; Cooper, M.; Shirts, M.; Dill, K. Small Molecule Hydration Free Energies in Explicit Solvent: An Extensive Test of Fixed-Charge Atomistic Simulations. *J Chem Theory Comput.* **2009**, *5*, 350–358.
- (31) Mennucci, B., Camm, R., Eds. *Continuum solvation models in chemical physics: from theory to applications*; John Wiley & Sons, 2007.
- (32) Konig, G.; Pickard, C.; Mei, Y.; Brooks, R. Predicting hydration free energies with a hybrid QM/MM approach: an evaluation of implicit and explicit solvation models in SAMPL4. *J Comput Aided Mol Des* **2014**, *28*, 245–57.
- (33) Kamerlin, S.; Haranczyk, M.; Warshel, A. Are Mixed Explicit/Implicit Solvation Models Reliable for Studying Phosphate Hydrolysis? A Comparative Study of Continuum, Explicit and Mixed Solvation Models. *Chemphysche* **2009**, *10*, 1125–1134.
- (34) Ashbaugh, H. S.; Kaler, E. W.; Paulaitis, M. E. A universal surface area correlation for molecular hydrophobic phenomena. *J. Am. Chem. Soc.* **1999**, *39*, 9243–9244.
- (35) Chorny, I.; Dill, K. A.; Jacobson, M. P. A universal surface area correlation for molecular hydrophobic phenomena. *J. Phys. Chem. B.* **2005**, *50*, 24056–24060.
- (36) Mobley, D. L.; Barber, A. E.; Fennell, C. J.; Dill, K. A. Charge Asymmetries in Hydration of Polar Solutes. *J. Phys. Chem. B.* **2008**, *8*, 2405–2414.
- (37) Rojas, R.; Batelaan, O.; Feyen, L.; Dassargues, A. Assessment of conceptual model

- uncertainty for the regional aquifer Pampa del Tamarugal – North Chile. *Hydrology and Earth System Sciences* **2010**, *14*, 171–192.
- (38) Apostolakis, G. The concept of probability in safety assessments of technological systems. *Science* **1990**, *250*, 1359–1364.
- (39) Devooght, J. Model uncertainty and model inaccuracy. *Reliability Engineering and System Safety* **1998**, *59*, 171–185.
- (40) Neuman, S.; Wierenga, P. A Comprehensive Strategy of Hydrogeologic Modeling and Uncertainty Analysis for Nuclear Facilities and Sites. 2003.
- (41) Bates, J. M.; Granger, C. W. J. The combination of forecasts. *Operational Research Quarterly* **1969**, *20*, 451–468.
- (42) Opitz, D.; Maclin, R. Popular Ensemble Methods: An Empirical Study. *Journal of Artificial Intelligence Research* **1999**, *11*, 169–198.
- (43) Rokach, L. Ensemble-based classifiers. *Artif. Intell. Rev.* **2010**, *33*, 1–39.
- (44) Hoeting, J.; Madigan, D.; Raftery, A.; Volinsky, C. Bayesian model averaging: a tutorial. *Statistical Science* **1999**, *14*, 382–417.
- (45) Gosink, L.; Hogan, E.; Pulsipher, T.; Baker, N. Bayesian model aggregation for ensemble-based estimates of protein pKa values. *Proteins* **2014**, *82*, 354–363.
- (46) Zhang, G. P. Time series forecasting using a hybrid ARIMA and neural network model. *Neurocomputing* **2003**, *50*, 159–175.
- (47) Morales-Casique, E.; Neuman, S. P.; Vesselinov, V. V. Maximum likelihood Bayesian averaging of airflow models in unsaturated fractured tuff using Occam and variance windows. *Stochastic Environmental Research and Risk Assessment* **2010**, *24*, 863–880.

- (48) Seni, G.; Elder, J. F. Ensemble Methods in Data Mining: Improving Accuracy Through Combining Predictions. *Synthesis Lectures on Data Mining and Knowledge Discovery* **2010**, *2*, 1–126.
- (49) Raftery, A.; Gneiting, T.; Balabdaoui, F.; Polakowski, M. Using Bayesian Model Averaging to Calibrate Forecast Ensembles. *Monthly Weather Review* **2005**, *133*, 1155–1174.
- (50) Vlachopoulou, M.; Gosink, L.; Pulsipher, T.; Ferryman, T.; Zhou, N.; Tong, J. An Ensemble Approach for Forecasting Net Interchange Schedule. IEEE PES General Meeting. 2013; p n. pag.
- (51) Raftery, A.; Madigan, D.; Hoeting, J. Bayesian Model Averaging for Linear Regression Models. *Journal of the American Statistical Association* **1998**, *92*, 179–191.
- (52) Raftery, A. Bayesian Model Selection in Social Research. *Sociological Methodology* **1995**, *25*, 111–163.
- (53) Ye, M.; Neuman, S. P.; Meyer, P. D. Maximum likelihood Bayesian averaging of spatial variability models in unsaturated fractured tuff. *Water Resources Research* **2004**, *40*, n. pag.
- (54) Hosmer, D. W.; Lemeshow, S. *Applied Logistic Regression*; Wiley: New York, 1989.
- (55) Reiss, P. T.; Huang, L.; Cavanaugh, J. E.; Roy, A. K. Resampling-based information criteria for best-subset regression. *Annals of the Institute of Statistical Mathematics* **2012**, *64*, 1161–1186.
- (56) Mallows, C. L. Some comments on  $C_p$ . *Technometrics* **1973**, *15*, 661–675.
- (57) Candes, E.; Tao, T. The Dantzig selector: Statistical estimation when  $p$  is much larger than  $n$ . *Annals of Statistics* **2007**, *35*, 2313–51.

- (58) Genell, A.; Nemes, S.; Steineck, G.; Dickman, P. W. Model selection in Medical Research: A simulation study comparing Bayesian Model Averaging and Stepwise Regression. *BMC Medical Research Methodology* **2010**, *10*, 108.
- (59) Davidson, I.; Fan, W. *Knowledge Discovery in Databases: PKDD 2006*; Lecture Notes in Comput. Science; Springer Berlin Heidelberg, 2006; Vol. 4213; pp 478–486.
- (60) Volinsky, C.; Madigan, D.; Raftery, A.; Kronmal, R. Bayesian Model Averaging in Proportional Hazard Models: Assessing the Risk of a Stroke. *Journal of the Royal Statistical Society: Series C (Applied Statistics)* **1997**, *46*, 433–448.
- (61) Quan, C.; Yu, Y.; Zhou, Z. Pareto Ensemble Pruning. Conference on Artificial Intelligence. 2015; pp 2935– 2941.
- (62) Martinez-Munoz, G.; Hernandez-Lobato, D.; Suarez, A. An Analysis of Ensemble Pruning Techniques Based on Ordered Aggregation. *Trans. Pattern Analysis and Machine Intelligence* **2009**, *31*, 245–259.
- (63) Onorante, L.; Raftery, A. *Dynamic Model Averaging in Large Model Spaces Using Dynamic Occam’s Window*; 2014.
- (64) Madigan, D.; Raftery, A. Model selection and accounting for model uncertainty in graphical models using Occam’s Window. *J. Am. Stat. Assoc.* **1994**, *89*, 1335–1346.
- (65) Jeffreys, H. *Theory of Probability*; Oxford: Oxford University Press, 1961.
- (66) Guthrie, J. SAMPL4, a blind challenge for computational solvation free energies: the compounds considered. *J Comput Aided Mol Des.* **2014**, *28*, 151–168.
- (67) Clyde, M. Bayesian Model Averaging and Model Search Strategies. *Bayesian Statistics*. 1999; pp 157–185.
- (68) Nicholls, A.; Wlodek, S.; Grant, J. SAMPL2 and continuum modeling. *J Comput Aided Mol Des.* **2010**; **24(4):293–306.**, *24*, 293–306.

- (69) Hawkins, G.; Giesen, D.; Lynch, G.; Chambers, C.; Rossi, I.; Storer, J.; Li, J.; Zhu, T.; Thompson, J.; Winget, P. et al. AMSOL.
- (70) Fixman, M. The Poisson–Boltzmann equation and its application to polyelectrolytes. *Journal of Chemical Physics* **1979**, *70*, 4995–146.
- (71) Honig, B.; Nicholls, A. Classical electrostatics in biology and chemistry. *Science* **1995**, *268*, 1144–1149.
- (72) Davis, M.; McCammon, A. Electrostatics in biomolecular structure and dynamics. *Chem. Rev.* **1990**, *90*, 509–521.
- (73) Sandberg, L. Predicting hydration free energies with chemical accuracy: The SAMPL4 challenge. *J Comput Aided Mol Des.* **2013**,
- (74) Klamt, A.; Eckert, F.; Diedenhofen, M. Prediction of the Free Energy of Hydration of a Challenging Set of Pesticide-Like Compounds. *J Phys Chem B.* **2009**,
- (75) Hogues, H.; Sulea, T.; Purisima, E. Exhaustive docking and solvated interaction energy scoring: Lessons learned from the SAMPL4 challenge. *J Comput Aided Mol Des.* **2014**,
- (76) Sulea, T.; Purisima, E. Predicting hydration free energies of polychlorinated aromatic compounds from the SAMPL-3 data set with FiSH and LIE models. *J Comput Aided Mol Des.* **2011**, *26*, 661–667.
- (77) Reinisch, J.; Klamt, A. Prediction of free energies of hydration with COSMO-RS on the SAMPL4 data set. *J Comput Aided Mol Des* **2014**,
- (78) Mobley, D.; Dunmont, E.; Chodera, J.; Dill, K. Comparison of charge models for fixed-charge force fields: small-molecule hydration free energies in explicit solvent. *J. Phys. Chem. B.* **2007**, *11*, 2242–2254.
- (79) Wang, J.; Wolf, R.; Caldwell, J.; Kollman, P.; Case, D. Development and testing of a general AMBER force field. *J Comput Chem.* **2004**, *25*, 1157–1174.

- (80) Li, L.; Dill, K.; Fennell, C. Testing the semi-explicit assembly model of aqueous solvation in the SAMPL4 challenge. *J Comput Aided Mol Des.* **2014**, *28*, 259–264.
- (81) Coleman, R.; Sterling, T.; Weiss, D. SAMPL4 & DOCK3.7: lessons for automated docking procedures. *J. Comput. Aided Mol. Des.* **2014**, *28*, 201–209.
- (82) Yang, Q.; Sharp, K. Atomic Charge Parameters for the Finite Difference Poisson-Boltzmann Method Using Electronegativity Neutralization. *J. Chem. Theory Comput.* **2006**, *4*, 1152–1167.
- (83) Genheden, S.; Cabedo, M.; Criddle, M.; Essex, J. Extensive all-atom Monte Carlo sampling and QM/MM corrections in the SAMPL4 hydration free energy challenge. *J. Comput. Aided Mol. Des.* **2014**, *28*, 187–200.
- (84) Beckstein, O.; Iorga, B. Prediction of hydration free energies for aliphatic and aromatic chloro derivatives using molecular dynamics simulations with the OPLS-AA force field. *J Comput Aided Mol Des* **2012**, *26*, 635–645.
- (85) Beckstein, O.; Fourier, A.; Iorga, B. Prediction of hydration free energies for the SAMPL4 diverse set of compounds using molecular dynamics simulations with the OPLS-AA force field. *J. Comput. Aided Mol. Des.* **2014**, *28*, 265–276.
- (86) Jambeck, J.; Mocci, F.; Lyubartsev, A.; Laaksonen, A. Partial atomic charges and their impact on the free energy of solvation. *J. Comput. Chem.* **2013**, *34*, 187–197.
- (87) Park, H. Extended solvent-contact model approach to SAMPL4 blind prediction challenge for hydration free energies. *J. Comput. Aided Mol. Des.* **2014**, *28*, 175–186.
- (88) Klamt, A.; Diedenhofen, M. Blind prediction test of free energies of hydration with COSMO-RS. *J. Comput. Aided Mol. Des.* **2010**, *24*, 357–360.
- (89) Wilcoxon, F. Individual Comparisons by Ranking Methods. *Biometrics Bulletin* **1945**, *1*, 80–83.

- (90) Breiman, L. Random Forests. *Machine Learning* **2001**, *45*, 5–32.
- (91) Hoerl, A.; Kennard, R. Ridge Regression: Biased Estimation for Nonorthogonal Problems. *Technometrics* **2000**, *42*, 80–86.
- (92) Tibshirani, R. Regression Shrinkage and Selection Via the Lasso. *J. Royal Stats. Soc.* **1994**, *58*, 267–288.
- (93) R Development Core Team, R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing: Vienna, Austria, 2008.
- (94) Buitinck, L.; Louppe, G.; Blondel, M.; Pedregosa, F.; Mueller, A.; Grisel, O.; Niculae, V.; Prettenhofer, P.; Gramfort, A.; Grobler, J. et al. API design for machine learning software: experiences from the scikit-learn project. ECML PKDD Workshop: Languages for Data Mining and Machine Learning. 2013; pp 108–122.