

Final Project – Antibody Response Induced by HIV Vaccines and T-cell Suppression Treatments in Rhesus Macaques – Second Draft

Kan Luo, Shih-Ni Prim, Frederick Davey, Rizwana Rehman

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Introduction

A dominant vaccine development strategy is to induce neutralizing antibodies by immunizing humans with the virus' glycoproteins. However, HIV vaccines that adopted this strategy mostly failed due to the fact that HIV is an RNA virus, which mutates rapidly to escape the inhibition of neutralizing antibodies. By the time the body generates neutralizing antibodies against the glycoproteins of some HIV strains, the RNA virus has already mutated. Thus, the existing neutralizing antibody fails to recognize, bind with, and neutralize the HIV virus. One possible solution is to increase the number of potential neutralizing antibodies that will cycle in the body by releasing a variety of antibodies after glycoprotein immunization.

Our dataset includes measurements of antibodies measured in 20 rhesus macaques after they were given the same HIV vaccine at three different time-points and one of three randomly selected anti-Treg treatments(drugs). Blood samples were collected two weeks after vaccine dosing, and antibodies were isolated

from those samples. A different number of antibodies were collected from each blood sample, limited by assay yield. Each observation contains information about the antibody isolated post the glycoprotein immunization.

A human antibody is formed by a heavy chain and light chain. For heavy chain, a human has about 51 V-gene segments, 25 D-gene segments and 6 J-gene segment. For light chain there are 71 V-gene segments and 9 J-gene segments[ref.5]. Any heavy chain V-D-J combination and light chain V-J combinations can randomly happen in germline center. Theoretically, there can be $51 * 25 * 6 * 71 * 9 = 4.88835 \times 10^6$ combinations of gene segments. Considering the frequently happened mutation and other factors, each individual can have over **10 billion** different antibodies. Thus, we decided to follow the convention of vaccine studies and treat each antibody as independent. [Kan – can you identify a reference article or journal here that uses this convention? We don't need to quote it / change the answer, it's just defending our claim that it's a standard practice]

Below is the list of variables with a brief description from our dataset. Please note that in each antibody, there are two sets of heavy chain and light chain, all of which forming a Y-shape immunoglobulin. Thus many of the variables start with H or L, indicating which chain the information comes from.

List of variables

- Monkey_id: Lists the identity of monkey
- Treatment(Drug): Treatment A is the mock control, and treatment B and C are two different kinds of Treg inhibitor treatments.
- Time_Points: 0 represents before immunization; 1 represents 2 weeks post 1st immunization; 2 represents 2 weeks post 2nd immunization; and 3 represents 2 weeks post 3rd immunization, respectively.
- Isotype: The category of antibody type; there are 5 kinds of immunoglobulin isotypes: IgG, IgA, IgM, IgE, IgD. The two most important kinds are IgG and IgM. IgM occurs in the acute stage of infection and perform an role of primary response. The secondary response IgG appears later in serum with higher binding affinity, and neutralizing potentials against toxins and virus. IgA mostly found in mucosal tissues such as Nasal mucosa. Non-dominant IgD and IgE are typically lower than 1% in blood.
- H_ID and L_ID: heavy chain and light chain IDs for the particular observation
- H_VBase: the number of nucleotide of the heavy chain variable region
- H_Substitutions: the number of relative nucleotide mutations in heavy chain.
- H_MuFreq: calculated by $H_Substitutions / H_VBase$
- H_CDR3: the number of amino acid of the heavy chain's third complementarity determining region
- L_VBase: the number of nucleotide of the light chain variable region
- L_Substitutions: the number of relative nucleotide mutations in light chain.
- L_MuFreq: calculated by $L_Substitutions / L_VBase$
- L_CDR3: the number of amino acid of the light chain's third complementarity determining region. **H_CDR3** and **L_CDR3** indicates the length of the third complementarity-determining region on the variable heavy chain and light chain. The longer they are, the more potential there is to produce diverse antibodies. [Kan, could you check to see if this is correct?] In other words, we want the values to be higher.
- Binding: affinity of antibodies against a selected HIV glycoprotein. The larger value indicates stronger binding. **Binding** indicates the rate of neutralizing, meaning how much the antibodies bind with the virus and thus make the virus ineffective. This is the most important measure of the study.

In the current report, we test if the different Treg treatments and number of vaccine injections(? which variables indicates this) cause changes in the antibody characteristics and if the changes are related to immunization/treatment time points.

Specifically, we evaluate:

- Q1. Do the paired antibody heavy chain and light chain differ with respect to mutation and CDR3?
- Q2. Do treatments, and/or isotypes have effects on the mutation frequency and/or the amino acid count in the third complementarity determining region (CDR3)?

Q3. How does the binding strength of the antibodies develop in response to the number of vaccine dosages by treatment? This will be evaluated with a longitudinal analysis by test subject.[NEEDS TO BE UPDATED]

Methods

During the experiment, 20 rhesus macaques were given glycoprotein immunization and supplemental antibody doses, as well as one of three treatments (two experimental regulatory T-cell suppression treatments and one control). For the analysis of mutation frequency and CDR3 count, each antibody within the same treatment was treated as an independent observation.

For exploratory data analysis we used histograms and Q-Q plots. We also computed correlations among variables. For inferential analysis we used Dependent Hotellings T^2 test using the ICSNP package [xxx] for Q1. To answer Q2 *i.e.*, to evaluate the mutation frequency and CDR count vs. Treatments and Isotypes we used two-way MANOVA. we ran the [xxx] function from the [xxx] package [package citation]. It was followed by further pair-wise analysis.

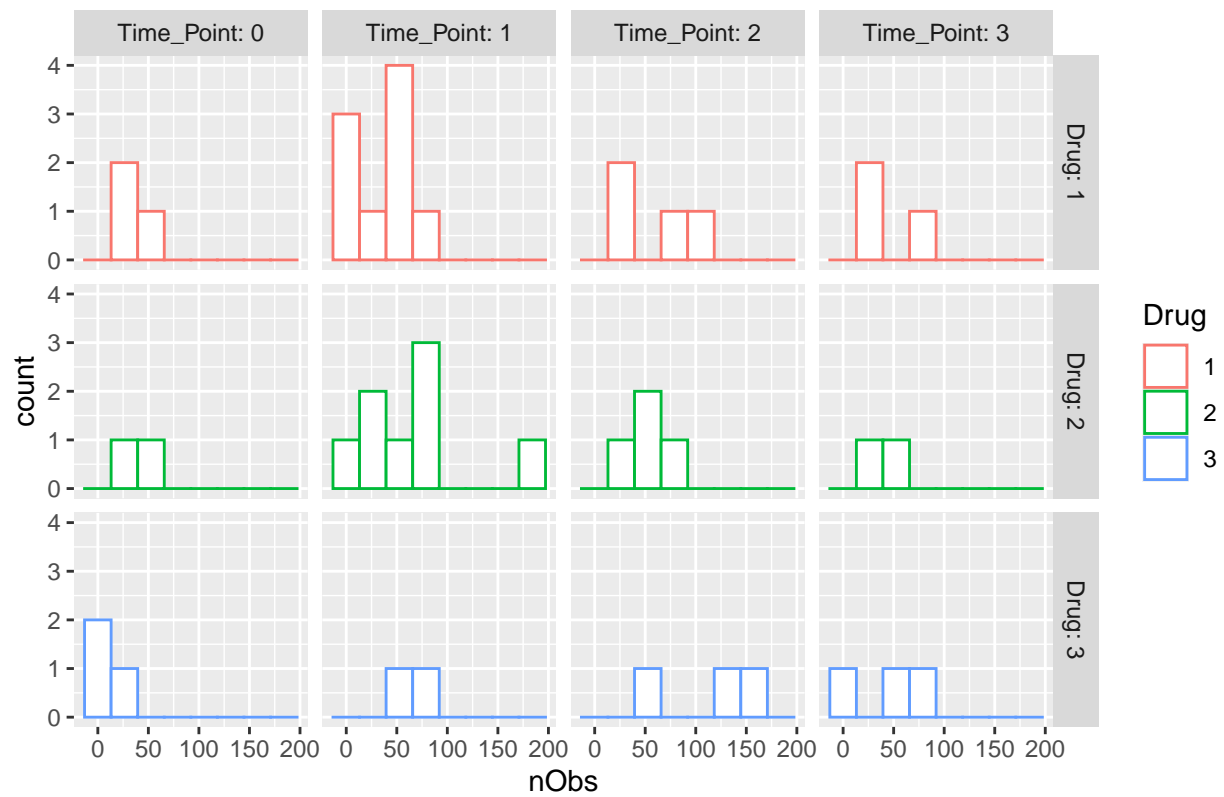
For the longitudinal analysis of binding strength vs number of vaccine doses, we will be using the gls function from the nlme package^[7]. (NEEDS TO UP DATED)

Results

Data Summaries

The dataset has 2465 antibodies collected from 20 rhesus monkeys. We first present our exploratory data analysis and summaries. An overview of the number of antibodies collected per treatment at each time-point is given below (Figure 1) along with tables for treatment counts per time-point and Isootype per treatment for individual time-points.

Figure1: Histograms of Antibodies Collected per Treatment and Timepoint



```
##      TimePoint
## Drug    0    1    2    3
##    1 129 346 251 122
##    2   90 533 225 101
##    3   54 125 347 142
```

```
## , , Isotype = A
```

```
##
```

```
##      TimePoint
```

```
## Drug    0    1    2    3
##    1     4    11     8     1
##    2     6    10     4     1
##    3     1     1     4     0
```

```
##
```

```
## , , Isotype = D
```

```
##
```

```
##      TimePoint
```

```
## Drug    0    1    2    3
##    1     4    22    15    16
##    2     4    45    19     7
##    3     1    26    14     6
```

```
##
```

```
## , , Isotype = E
```

```
##
```

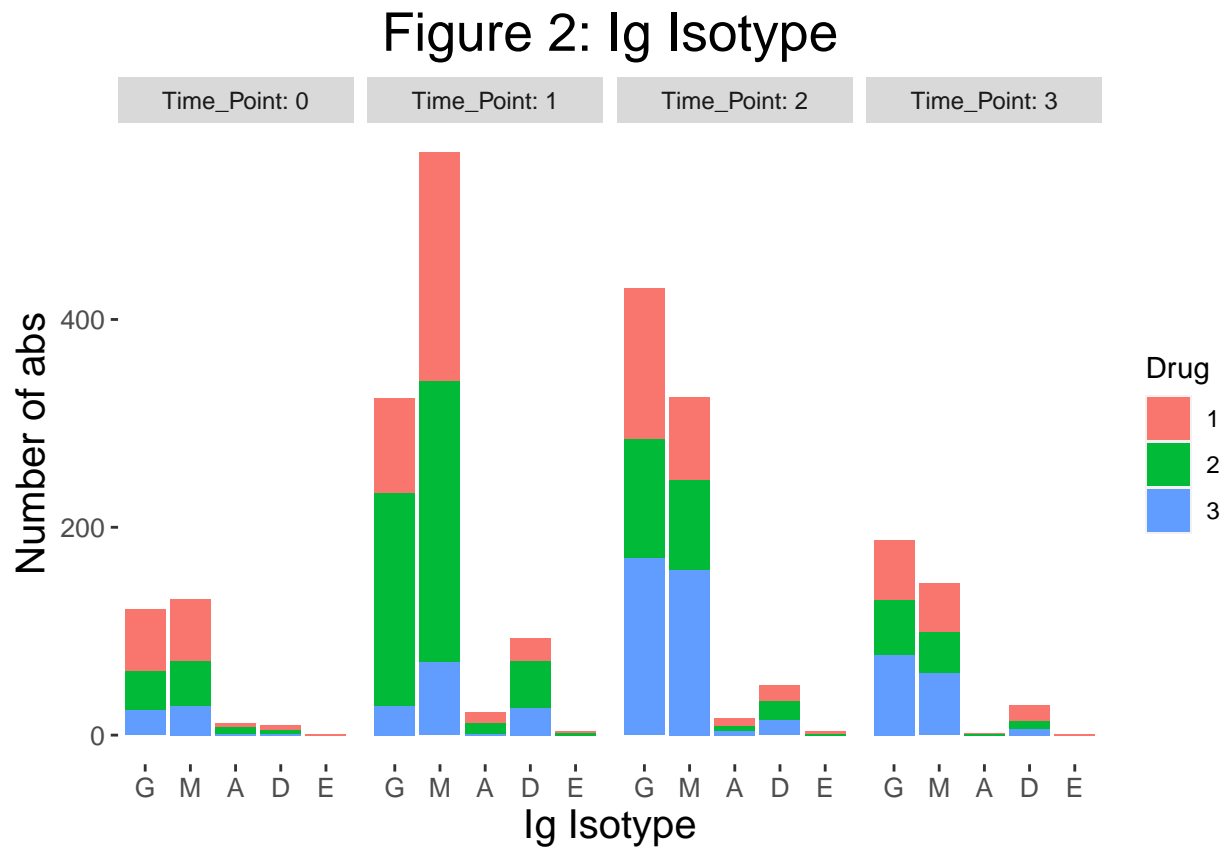
```
##      TimePoint
```

```

## Drug    0    1    2    3
##      1    1    2    3    1
##      2    0    2    1    0
##      3    0    0    0    0
##
## , , Isotype = G
##
##      TimePoint
## Drug    0    1    2    3
##      1  60  91 145  57
##      2  37 205 115  53
##      3  24  28 170  77
##
## , , Isotype = M
##
##      TimePoint
## Drug    0    1    2    3
##      1  60 220  80  47
##      2  43 271  86  40
##      3  28  70 159  59

```

Next, the histogram of `Isotype` is presented in Figure 2.



As expected, we observed that IgG and IgM occupied the biggest proportion of all antibodies in all time points. Before immunization (time point 0), there were similar weight of IgG and IgM found in blood. After the 1st immunization (time point 1), primary immune response resulted an increase of IgM, followed with IgG increase at later time point 2 and 3.

For the response variables, we began with an outlier check, and then reviewed their distributions across treatment and time-points.

Figure 3: Histogram of Response Variables

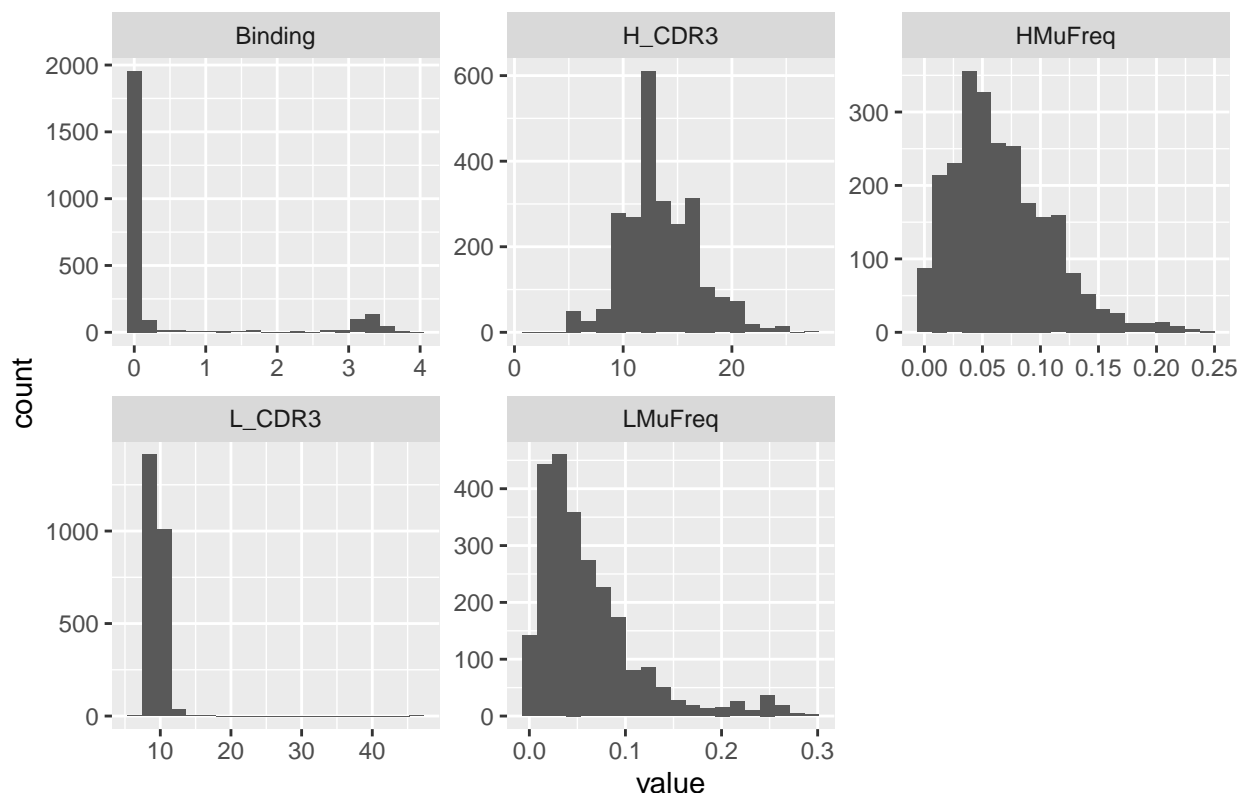


Table 1: Normal Standardized L_CDR3 Statistics

V1	Min. :-2.1860	1st Qu.:-0.5361	Median :-0.5361	Mean : 0.0000	3rd Qu.: 0.2888	Max. :30.8110
----	---------------	-----------------	-----------------	---------------	-----------------	---------------

L_CDR3 appeared to have an outlier several standard (?distribution). We believed that this most likely was caused by a laboratory error, so we excluded this observation and continued the analysis with the remaining data of 2464 observations.

We examined our responses: H_CDR3, HMuFreq, L_CDR3, LMuFreq and Binding.

We observed that for H_CDR3 the distributions were roughly normal with the center around 13 at different time-points (Figure 4) without taking into account different treatments. Figure 5 represents the distribution of H_CDR3 with respect to treatments at different time-points. , and slightly centered around 9 for L_CDR3 at different time- points (Figure 5).

Figure 4: Histogram H_CDR3

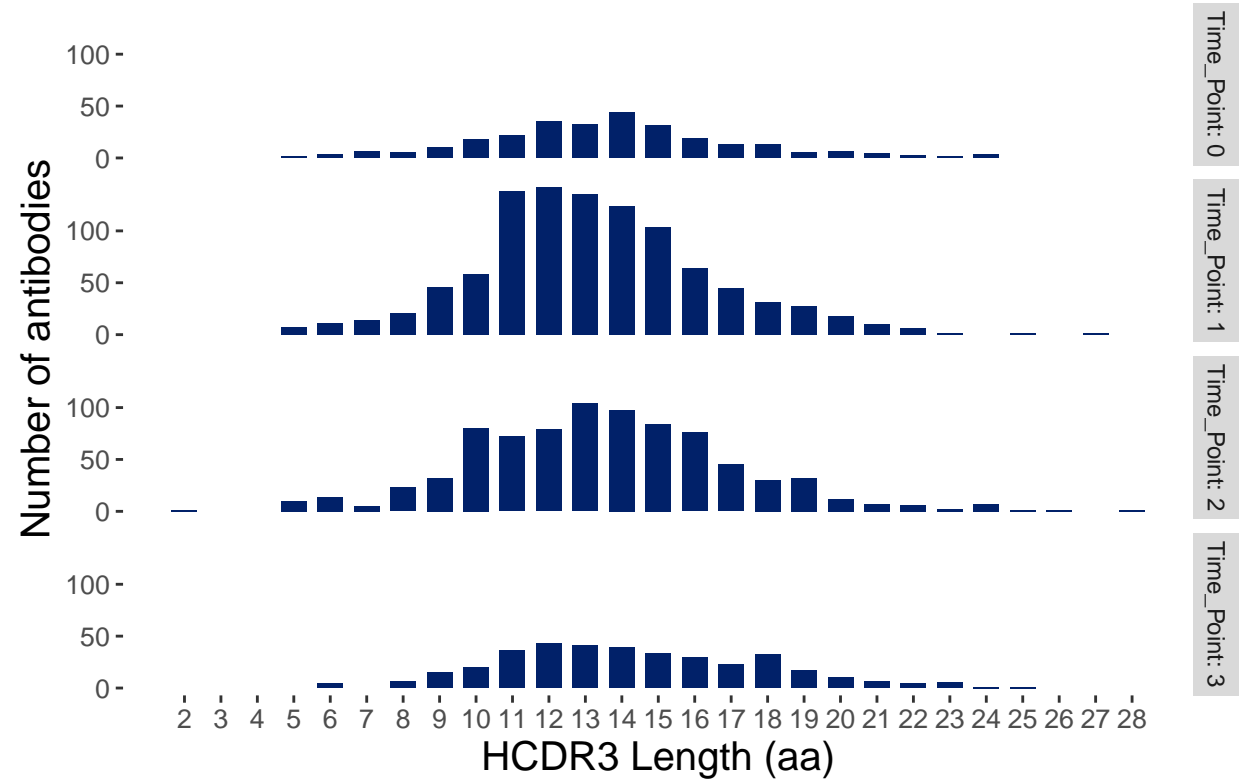
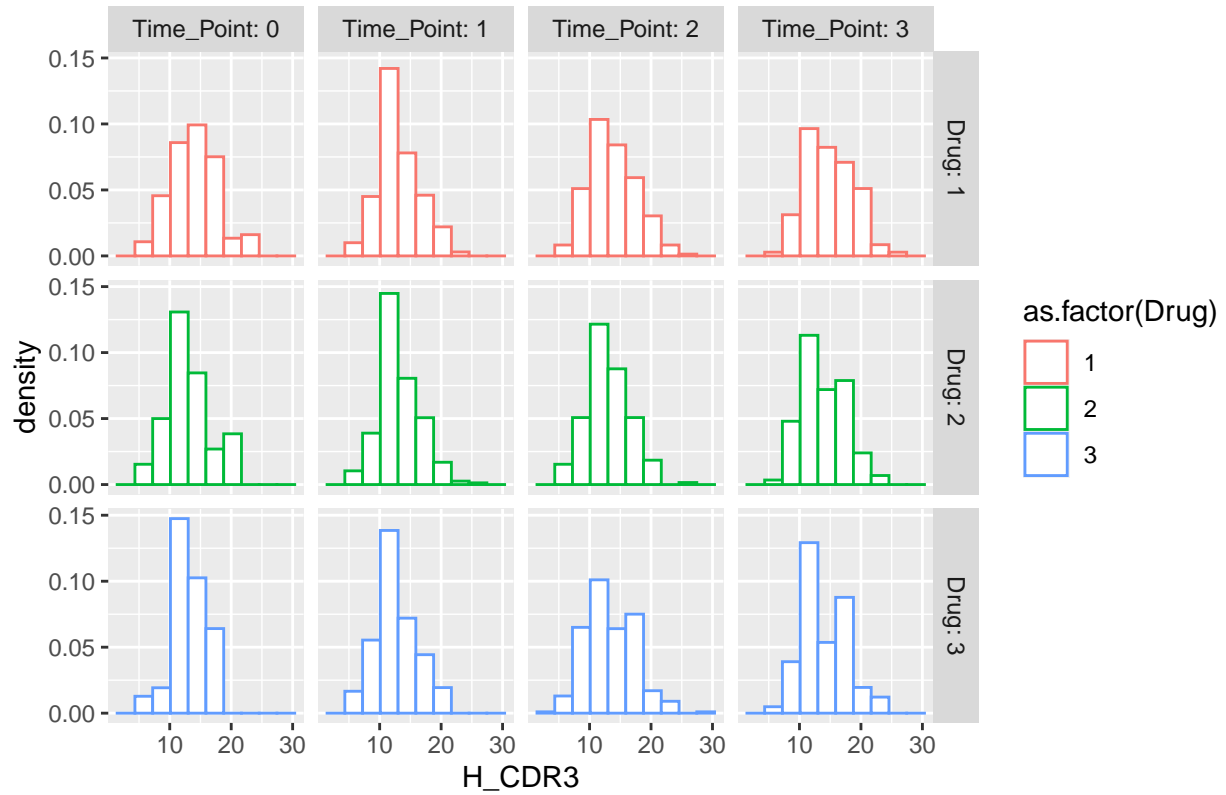


Figure 5: Histograms of H_CDR3 vs. Treatment and Timepoint



Figures for L_CDR3 are shown next.

Figure 6: Histogram L_CDR3

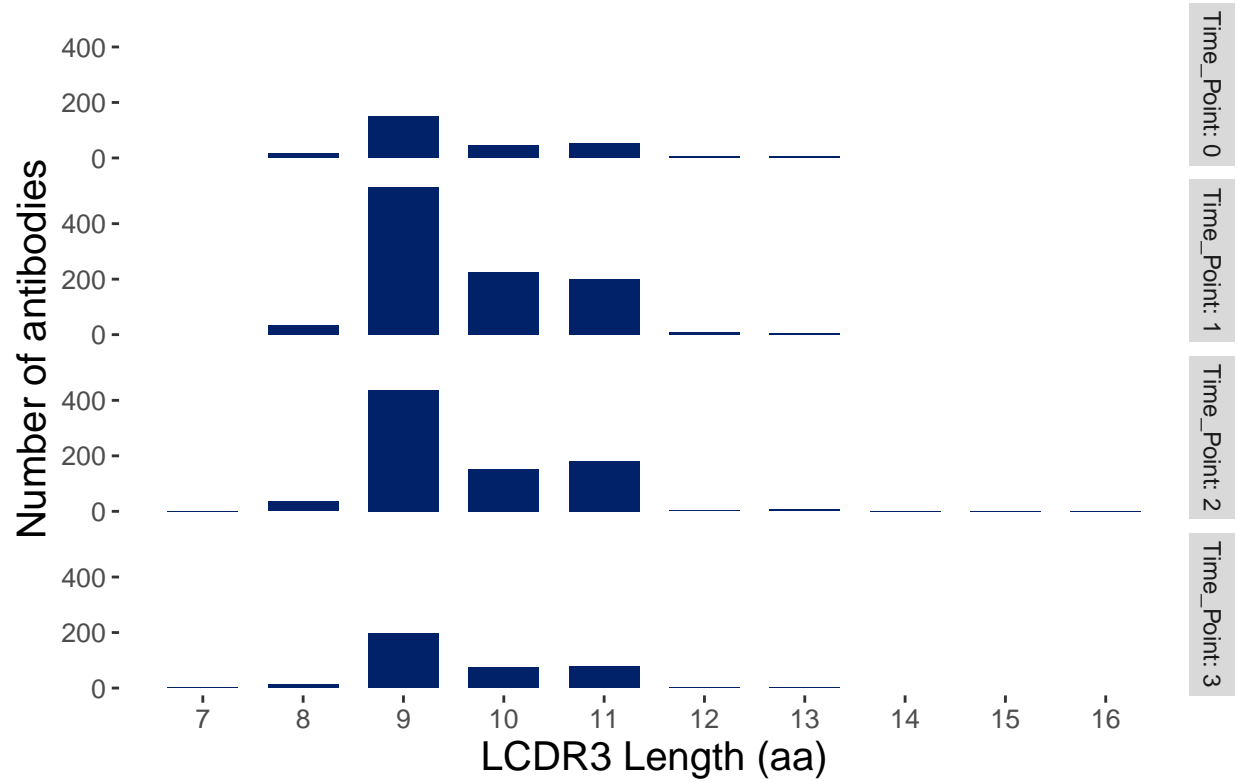
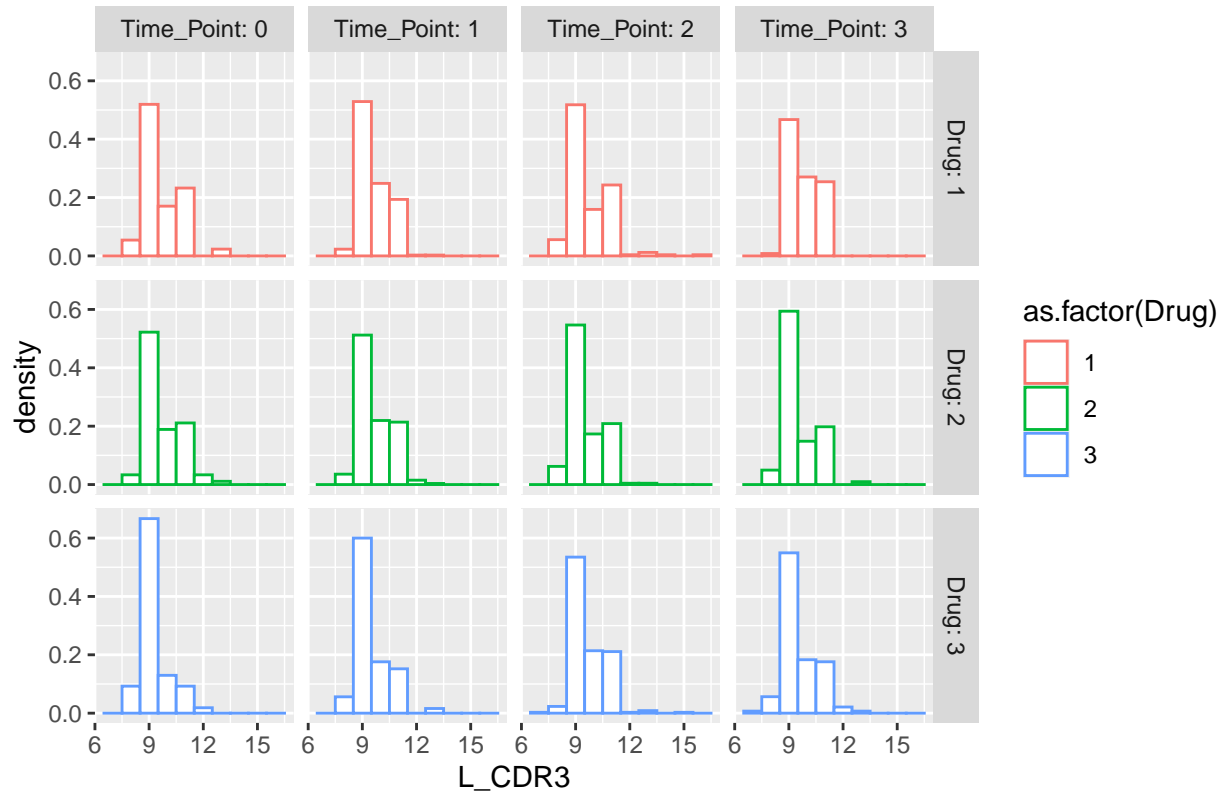


Figure 7: Histograms of L_CDR3 vs. Treatment and Timepoint



Q-Q plots of H_CDR3 and L_CDR3 are shown below.

Figure 8a: Q-Q Plot of H_CDR3

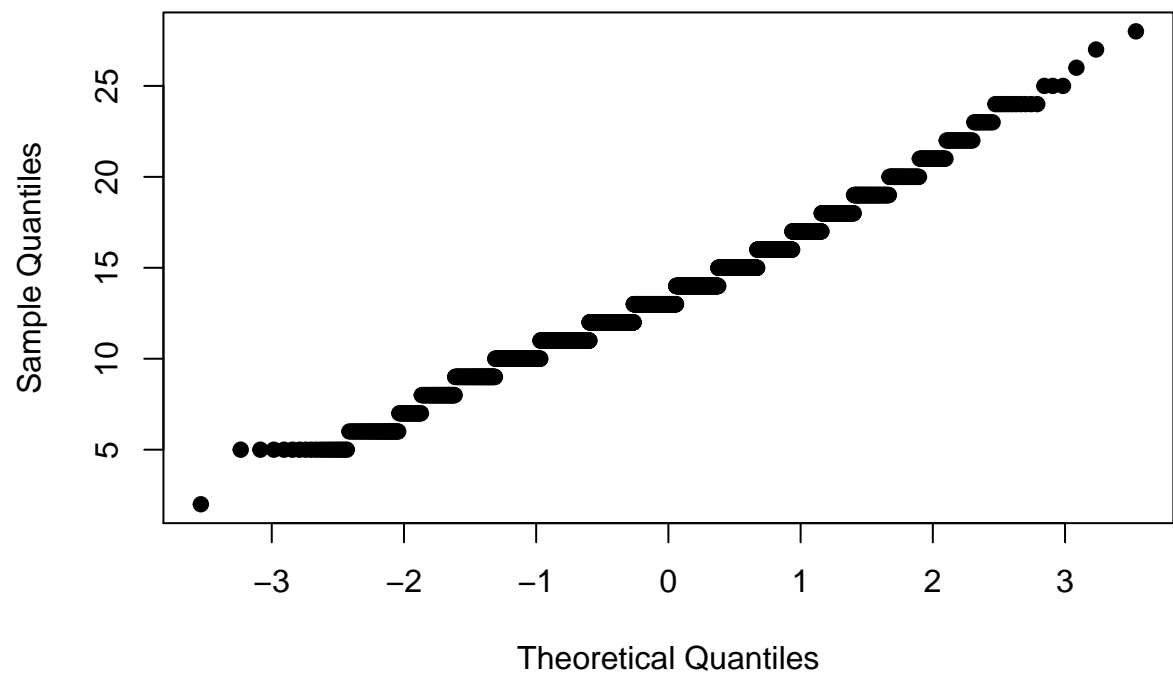


Figure 8b: Q-Q Plot of L_CDR3

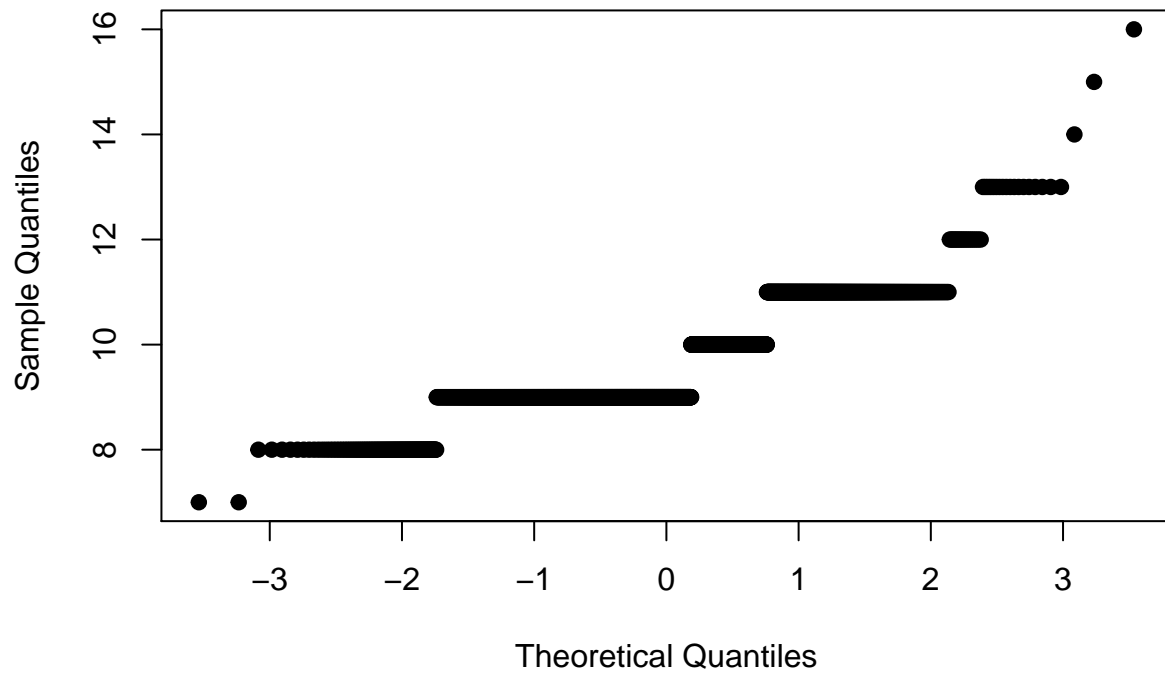


Figure 9: Histogram HMuFreq

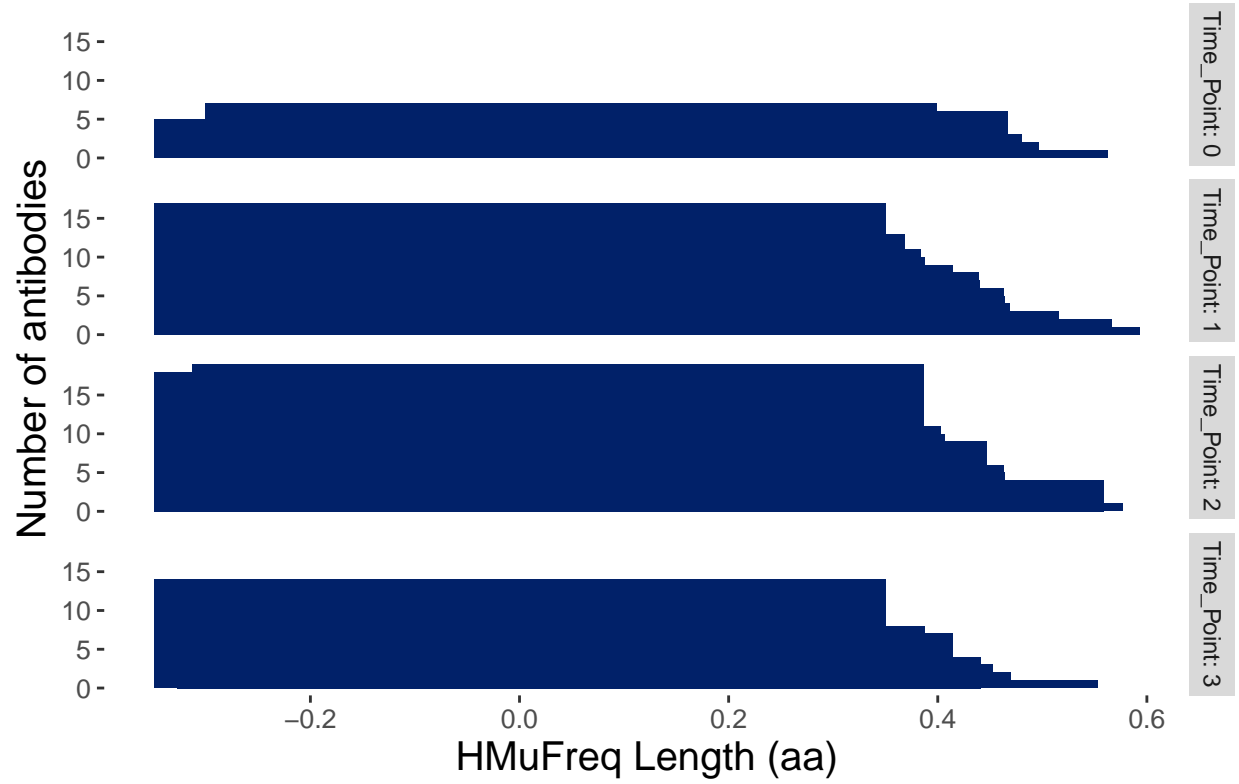


Figure 10: Histograms of HMuFreq vs. Treatment and Timepoint

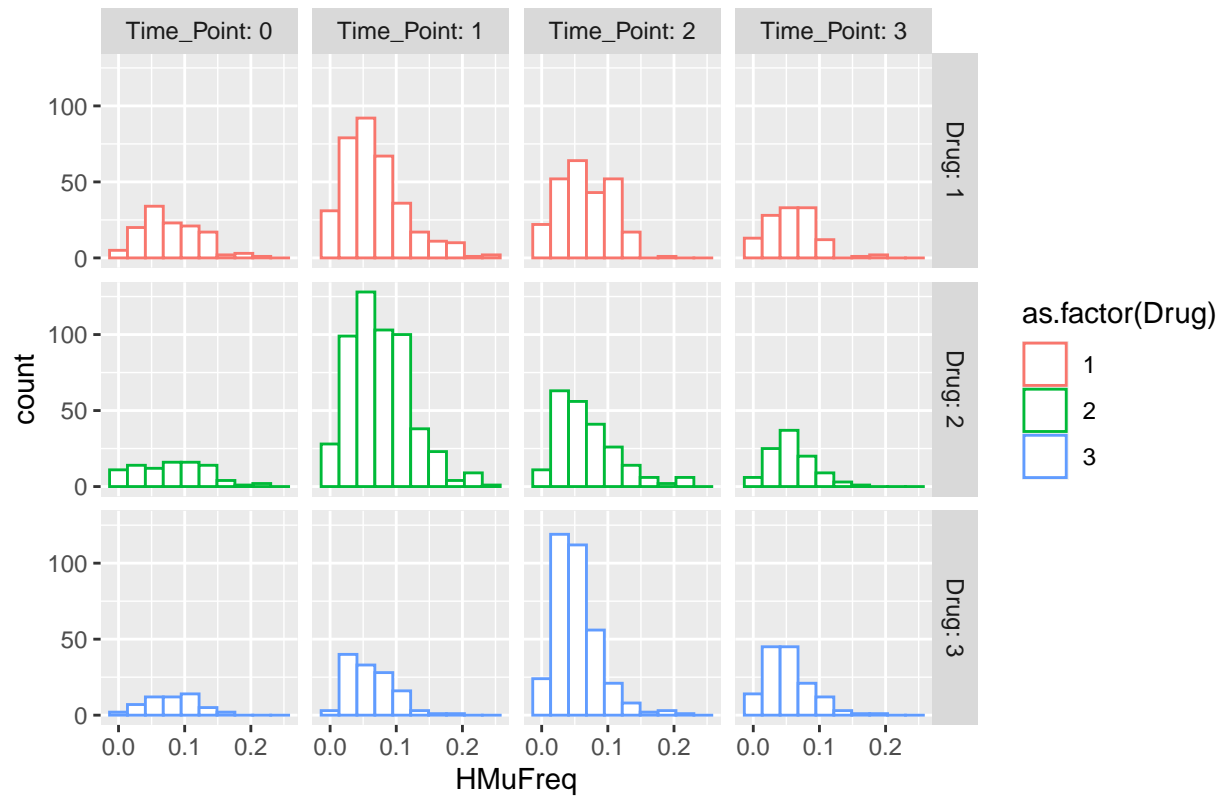


Figure 11: Histogram LMuFreq

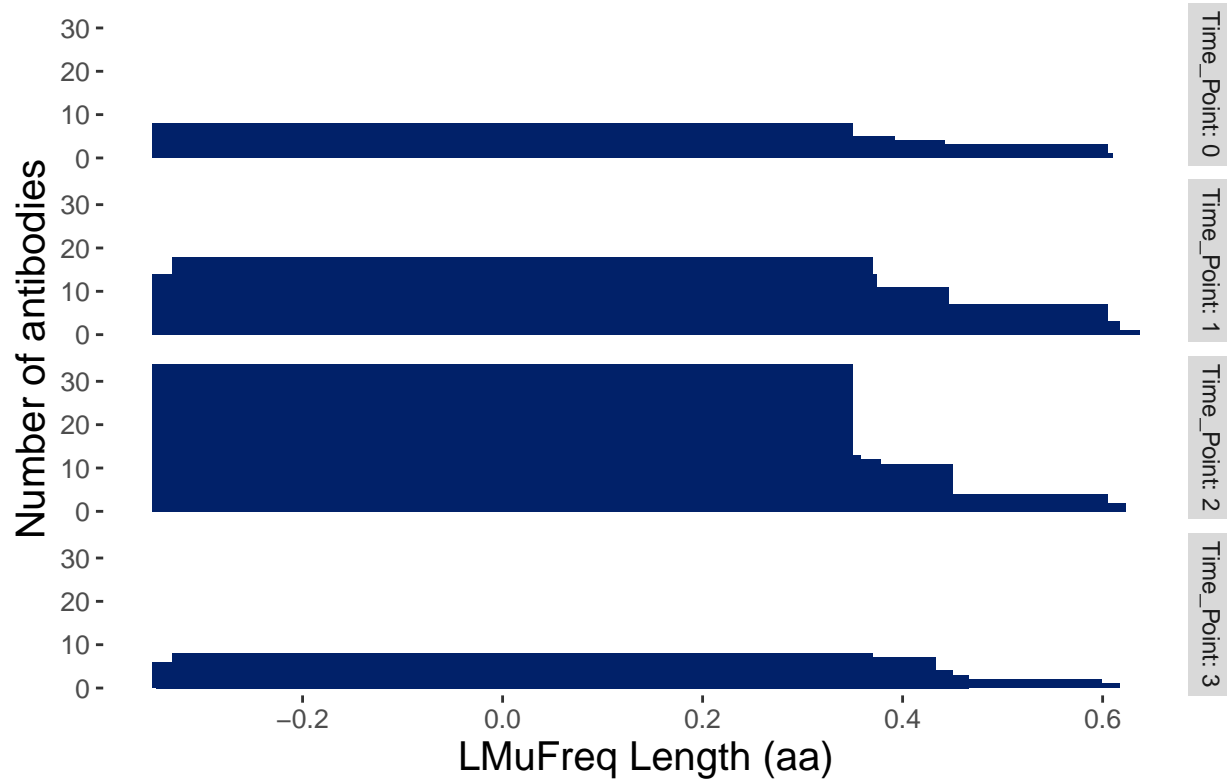
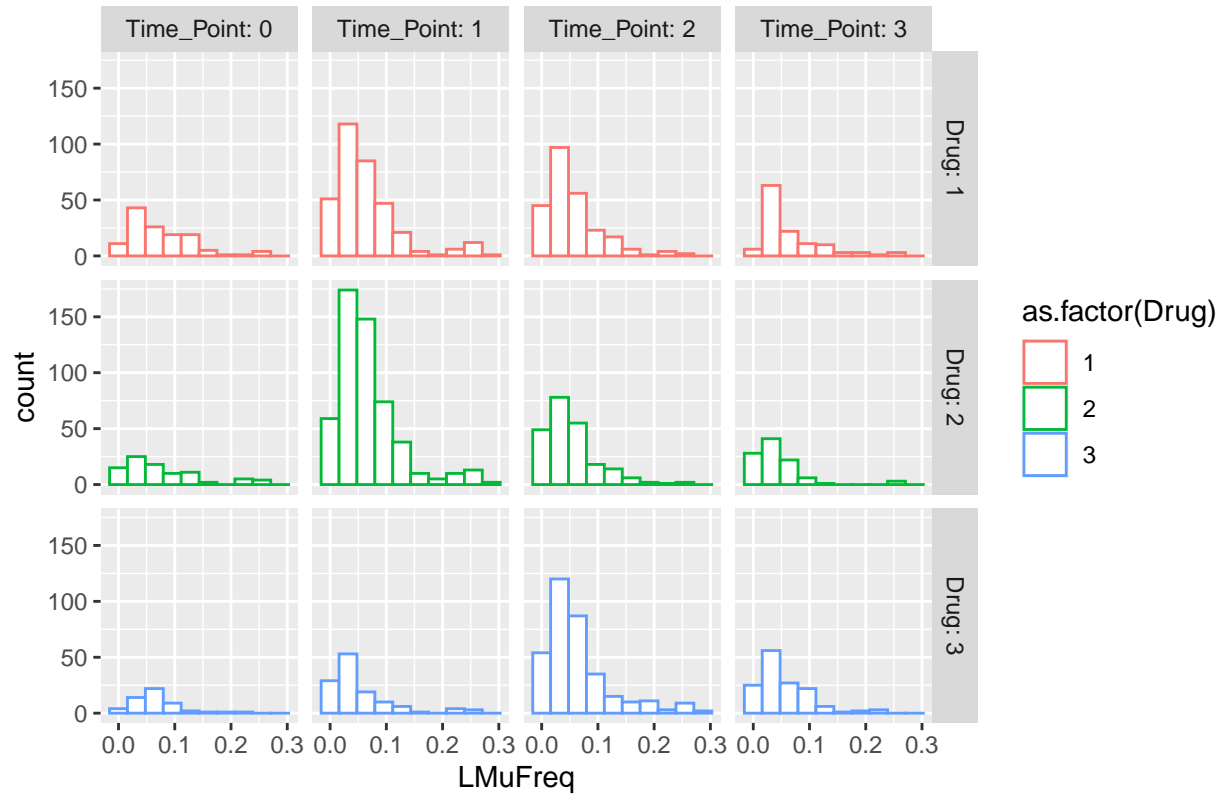


Figure 12: Histograms of LMuFreq vs Treatment and Timepoint



Next, the Q-Q plots of HMuFreq and LMuFreq are shown.

Figure 13a: Q-Q Plot of HMuFreq

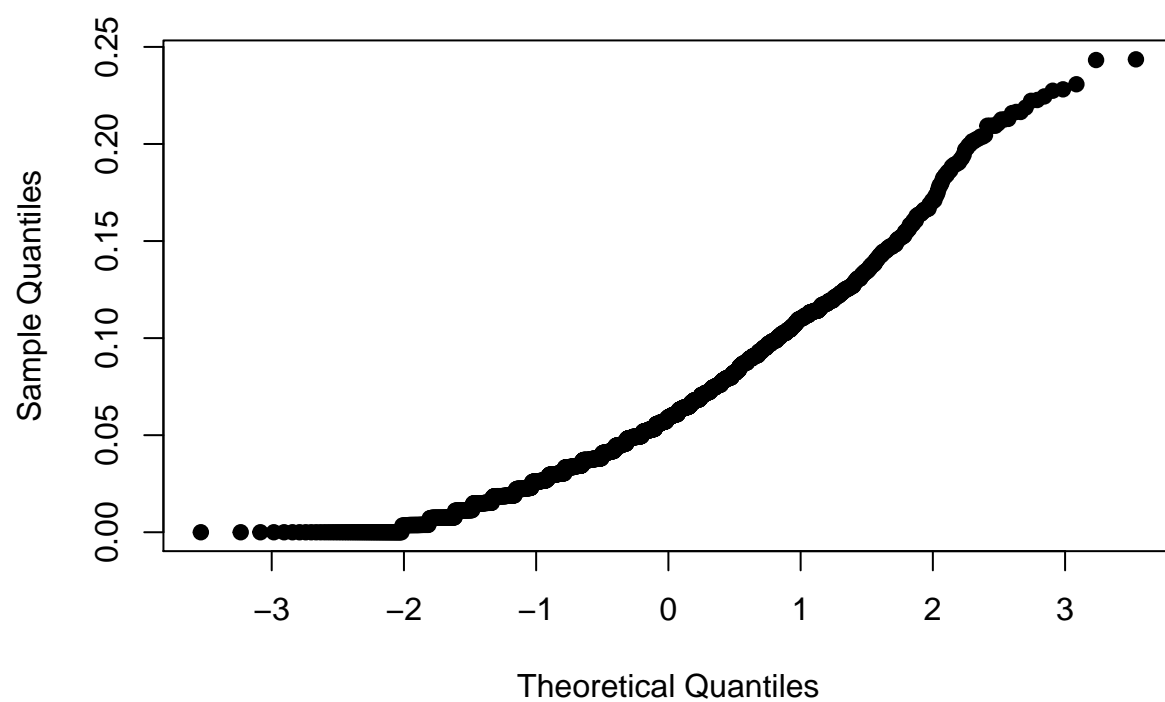
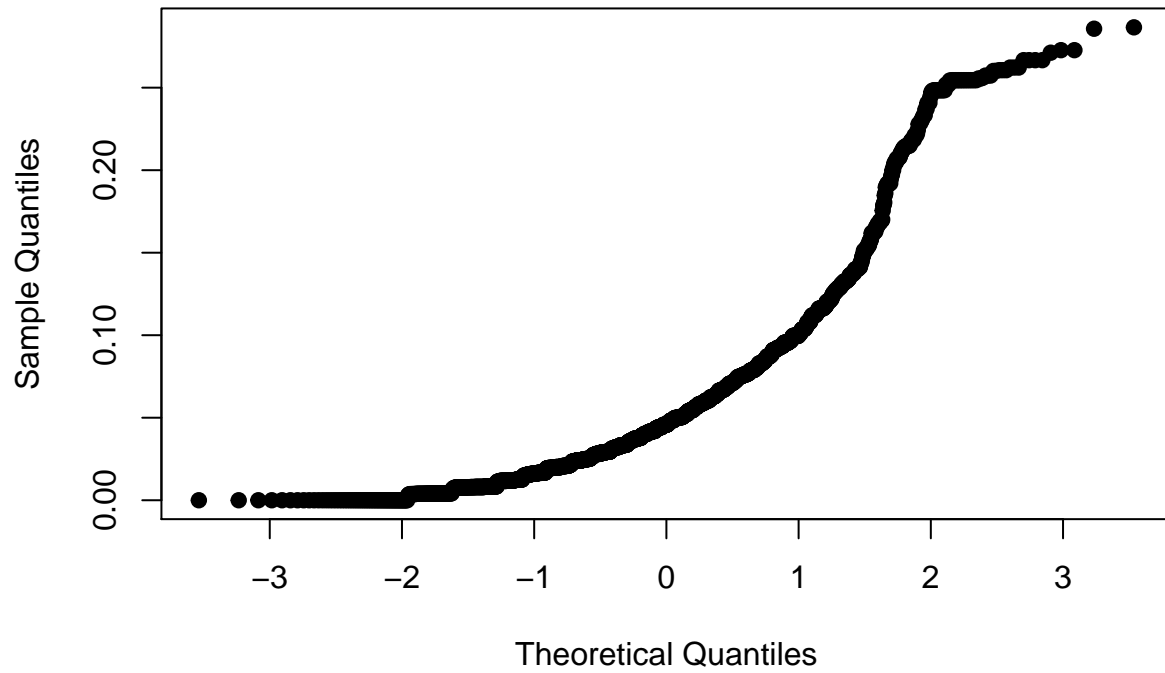


Figure 13b: Q-Q Plot of LMuFreq



Lastly, Histogram of `Binding` with respect to treatments at different time points and Q-Q plot are shown. We observed that `Binding` was not normally distributed. However, since our sample size is larger than 2000, we can use the Central Limit Theorem and assume normality.

Figure 14: Histograms of Binding Strength vs Treatment and Timepoint

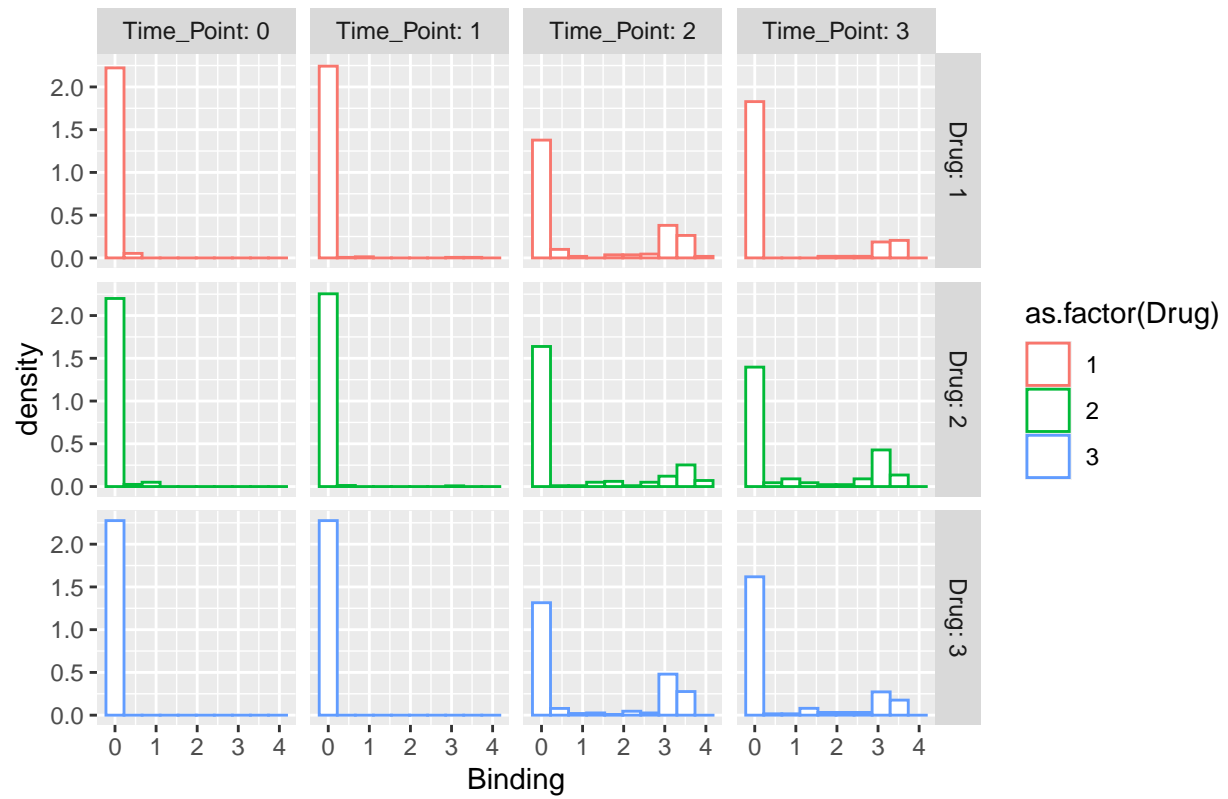
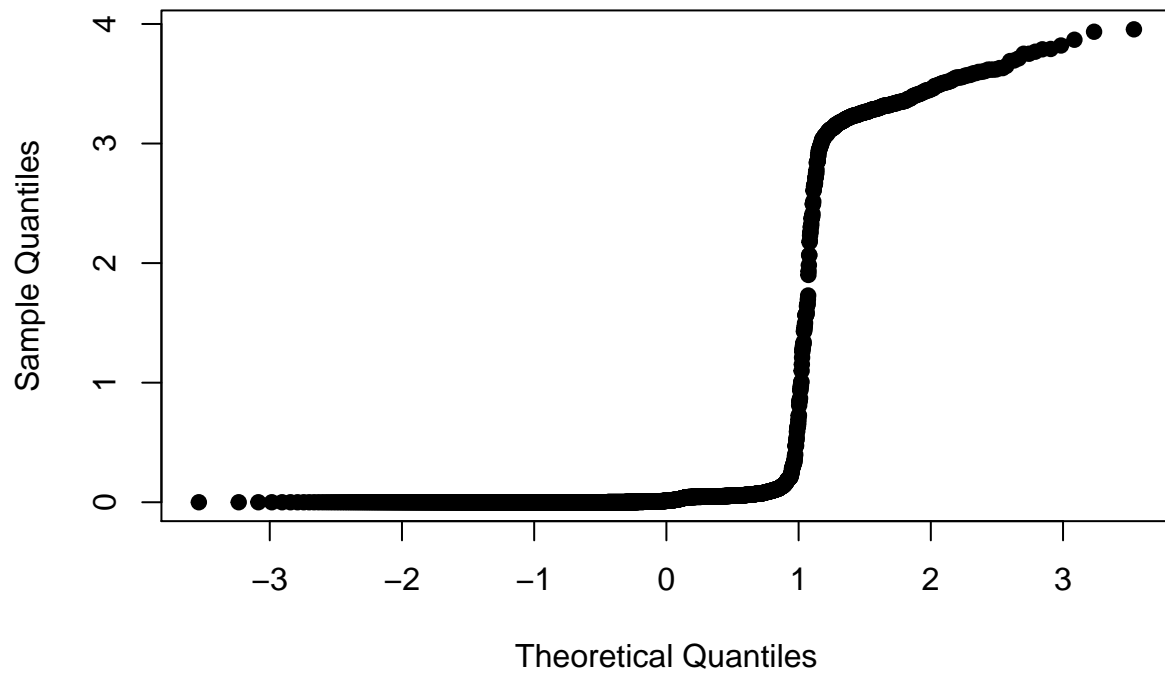
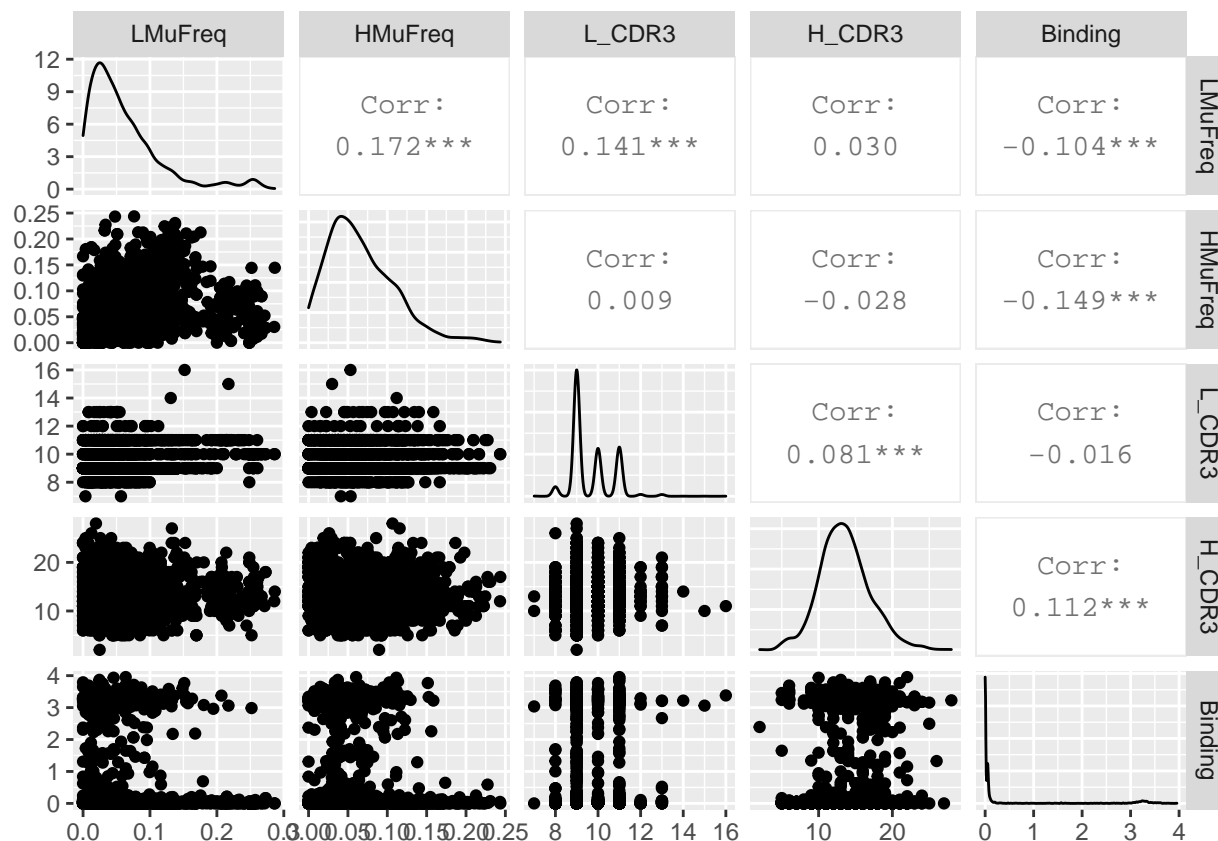


Figure 15: Q-Q Plot of Binding



We checked whether response variables could be correlated. In the plot below, we observed that none of the response variables were highly correlated.



inferential Data Analysis

To answer $Q1$ we conducted Hotellings T^2 test for a pairwise comparison between respective heavy chain and light chain variable pairs L_CDR3, H_CDR3 and HMuFreq, LMuFreq.

```
##
## Hotelling's one sample T2-test
##
## data: D
## T.2 = 1585.6, df1 = 2, df2 = 2459, p-value < 2.2e-16
## alternative hypothesis: true location is not equal to c(0,0)
```

For $Q2$ We wanted to test whether predictors Drug and Isotype had effects on the five responses: H_CDR3, HMuFreq, L_CDR3, LMuFreq, and Binding. We used *MANOVA* due to large sample size ($n = 2464$) without worrying about the normality assumption. In the output, we noticed that the main effects of Drug and Isotype and the interaction effects were all significant.

```
##          Df  Pillai approx F num Df den Df    Pr(>F)
## drug      2 0.058434   14.711    10  4888 < 2.2e-16 ***
## it        4 0.300048   39.672    20  9784 < 2.2e-16 ***
## drug:it   7 0.069958    4.960    35 12235 < 2.2e-16 ***
## Residuals 2447
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Pairwise comparison

To know more details about which groups have different means, we use pairwise comparisons for each treatment group, drug, and Isotype. (We set α as 0.05, and use a Bonferroni correction.)

```
## [1] "L_CDR3 pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## 1 - 2      0.0451 0.0450 2458 -0.0870    0.177
## 1 - 3      0.1018 0.0492 2458 -0.0429    0.246
## 2 - 3      0.0567 0.0480 2458 -0.0844    0.198
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.996666666666667
## [1] "LMuFreq pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## 1 - 2     -0.000176 0.00256 2458 -0.00770    0.00735
## 1 - 3      0.002320 0.00280 2458 -0.00592    0.01056
## 2 - 3      0.002496 0.00273 2458 -0.00554    0.01053
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.996666666666667
## [1] "H_CDR3 pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## 1 - 2      0.4907 0.160 2458  0.0204    0.961
## 1 - 3      0.4438 0.175 2458 -0.0712    0.959
## 2 - 3     -0.0469 0.171 2458 -0.5492    0.456
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.996666666666667
## [1] "HMuFreq pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## 1 - 2     -0.006 0.00199 2458 -0.01186 -0.000145
## 1 - 3      0.011 0.00218 2458  0.00461  0.017432
## 2 - 3      0.017 0.00213 2458  0.01076  0.023276
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.996666666666667
## [1] "Binding pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## 1 - 2      0.118 0.0522 2458 -0.0355    0.271
## 1 - 3     -0.356 0.0571 2458 -0.5235   -0.188
## 2 - 3     -0.473 0.0557 2458 -0.6372   -0.310
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.996666666666667

## [1] "L_CDR3 pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## A - D      0.1136 0.1510 2456 -0.3838    0.611
```

```

## A - E      0.0667 0.3289 2456 -1.0170 1.150
## A - G      0.0041 0.1364 2456 -0.4451 0.453
## A - M      0.0441 0.1361 2456 -0.4042 0.492
## D - E     -0.0469 0.3091 2456 -1.0651 0.971
## D - G     -0.1095 0.0769 2456 -0.3627 0.144
## D - M     -0.0695 0.0764 2456 -0.3210 0.182
## E - G     -0.0626 0.3022 2456 -1.0581 0.933
## E - M     -0.0225 0.3021 2456 -1.0177 0.973
## G - M      0.0400 0.0404 2456 -0.0931 0.173
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.999
## [1] "LMuFreq pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## A - D     -0.021206 0.00858 2456 -0.04947 0.00705
## A - E     -0.006829 0.01869 2456 -0.06840 0.05475
## A - G     -0.023476 0.00775 2456 -0.04900 0.00205
## A - M     -0.021669 0.00773 2456 -0.04714 0.00380
## D - E      0.014376 0.01756 2456 -0.04348 0.07223
## D - G     -0.002270 0.00437 2456 -0.01666 0.01212
## D - M     -0.000463 0.00434 2456 -0.01476 0.01383
## E - G     -0.016647 0.01717 2456 -0.07321 0.03992
## E - M     -0.014839 0.01716 2456 -0.07138 0.04170
## G - M      0.001807 0.00230 2456 -0.00576 0.00937
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.999
## [1] "H_CDR3 pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## A - D      0.297 0.530 2456 -1.448 2.043
## A - E     -2.229 1.154 2456 -6.032 1.574
## A - G     -0.728 0.479 2456 -2.304 0.849
## A - M      0.503 0.478 2456 -1.070 2.076
## D - E     -2.527 1.085 2456 -6.100 1.046
## D - G     -1.025 0.270 2456 -1.914 -0.137
## D - M      0.206 0.268 2456 -0.677 1.089
## E - G      1.502 1.060 2456 -1.992 4.995
## E - M      2.733 1.060 2456 -0.760 6.225
## G - M      1.231 0.142 2456 0.764 1.698
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.999
## [1] "HMuFreq pairwise CI's"
## contrast estimate      SE   df lower.CL upper.CL
## A - D      0.00523 0.00659 2456 -0.0165 0.02693
## A - E      0.00848 0.01435 2456 -0.0388 0.05576
## A - G     -0.01628 0.00595 2456 -0.0359 0.00332
## A - M      0.00393 0.00594 2456 -0.0156 0.02349
## D - E      0.00325 0.01349 2456 -0.0412 0.04768
## D - G     -0.02151 0.00335 2456 -0.0326 -0.01046
## D - M     -0.00130 0.00333 2456 -0.0123 0.00968

```

```

## E - G    -0.02476 0.01319 2456  -0.0682  0.01868
## E - M    -0.00455 0.01318 2456  -0.0480  0.03888
## G - M      0.02021 0.00176 2456   0.0144  0.02602
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.999
## [1] "Binding pairwise CI's"
## contrast estimate      SE    df lower.CL upper.CL
## A - D      0.30395 0.1610 2456  -0.227   0.834
## A - E      0.20847 0.3508 2456  -0.947   1.364
## A - G     -0.66527 0.1454 2456  -1.144  -0.186
## A - M      0.30220 0.1451 2456  -0.176   0.780
## D - E     -0.09548 0.3296 2456  -1.181   0.990
## D - G     -0.96922 0.0820 2456  -1.239  -0.699
## D - M     -0.00175 0.0814 2456  -0.270   0.267
## E - G     -0.87374 0.3223 2456  -1.936   0.188
## E - M      0.09373 0.3222 2456  -0.968   1.155
## G - M      0.96747 0.0431 2456   0.825   1.109
##
## Results are averaged over the levels of: rep.meas
## Note: contrasts are still on the [.: scale
## Confidence level used: 0.999

```

Here are the pairs that have significant differences:

- Treatment
 - L_CDR3: none
 - LMuFreq: none
 - H_CDR3:
 - * group 1 > group 4
 - HMuFreq:
 - * group 1 > group 7
 - * group 3 < group 5
 - * group 3 < group 6
 - * group 5 > group 7
 - * group 6 > group 7
 - Binding:
 - * group 1 < group 2
 - * group 1 > group 6
 - * group 2 > group 3
 - * group 2 > group 4
 - * group 2 > group 5
 - * group 2 > group 6
 - * group 3 < group 7
 - * group 4 > group 6
 - * group 4 < group 7
 - * group 5 < group 7
 - * group 6 < group 7
- Drug
 - L_CDR3: none

- LMuFreq: none
- H_CDR3:
 - * $1 > 2$
- HMuFreq:
 - * $1 < 2$
 - * $1 > 3$
 - * $2 > 3$
- Binding:
 - * $1 < 3$
 - * $2 < 3$
- Isotype
 - L_CDR3: none
 - LMuFreq: none
 - H_CDR3:
 - * $\text{IgD} < \text{IgG}$
 - HMuFreq:
 - * $\text{IgD} < \text{IgG}$
 - * $\text{IgG} > \text{IgM}$
 - Binding:
 - * $\text{IgA} < \text{IgG}$
 - * $\text{IgD} < \text{IgG}$
 - * $\text{IgG} > \text{IgM}$

In short, L_CDR3 and LMuFreq do not have significant paired differences.

For H_CDR3, treatment group 1 (drug 1) is higher than treatment group 7 (control), and IgG has a longer H_CDR3 length than IgD.

For HMuFreq, drug 2 has the highest mean, followed by drug 1 and control. More specifically, treatment groups 5 and 6 (two doses in drug 2) have the highest mutation rates. IgG has higher mutation rate than IgD.

For Binding, drug 3 has the highest mean, but drug 1 and drug 2 do not have significant differences. IgG has higher binding rate than IgA, IgD, and IgM.

We could conclude that the drugs/treatment groups do increase mutation rate; however, they do not increase binding rate. That is to say, although the treatments do help increase the diversity of antibodies, they are not specific to the HIV antigens and thus do not increase binding.

Discussion

Test Assumptions Results Implications (Interpretation for practical significance)

Conclusion (we can omit this and merge with Discussion)

Disclosure: The dataset, which can be found here, was provided by Kan Luo, as he was one of authors for the following four publications that used the dataset:

1. Luo K, Liao HX, Zhang R, et al. Tissue memory B cell repertoire analysis after ALVAC/AIDSVAX B/E gp120 immunization of rhesus macaques. *JCI Insight*. 2016;1(20):e88522. Published 2016 Dec 8. doi:10.1172/jci.insight.88522

Reference

The dataset, which can be found here, was provided by Kan Luo, as he was one of authors for the following four publications that used the dataset:

1. Luo K, Liao HX, Zhang R, et al. Tissue memory B cell repertoire analysis after ALVAC/AIDSVAX B/E gp120 immunization of rhesus macaques. *JCI Insight*. 2016;1(20):e88522. Published 2016 Dec 8. doi:10.1172/jci.insight.88522
2. Bradley, T., Kuraoka, M., Yeh, C.-H., Tian, M., Chen, H., Cain, D. W., . . . Haynes, B. F. (2020). Immune checkpoint modulation enhances HIV-1 antibody induction. *Nature Communications*, 11(1), 948. doi:10.1038/s41467-020-14670-w
3. Easterhoff, D., Pollara, J., Luo, K., Tolbert, W. D., Young, B., Mielke, D., . . . Ferrari, G. (2020). Boosting with AIDSVAX B/E Enhances Env Constant Region 1 and 2 Antibody-Dependent Cellular Cytotoxicity Breadth and Potency. *Journal of Virology*, 94(4), e01120-01119. doi:10.1128/jvi.01120-19
4. Wiehe, K., Easterhoff, D., Luo, K., Nicely, N. I., Bradley, T., Jaeger, F. H., Dennison, S. M., Zhang, R., Lloyd, K. E., Stolarchuk, C., Parks, R., Sutherland, L. L., Searce, R. M., Morris, L., Kaewkungwal, J., Nitayaphan, S., Pitisuttithum, P., Rerks-Ngarm, S., Sinangil, F., Phogat, S., . . . Haynes, B. F. (2014). Antibody light-chain-restricted recognition of the site of immune pressure in the RV144 HIV-1 vaccine trial is phylogenetically conserved. *Immunity*, 41(6), 909-918. <https://doi.org/10.1016/j.immuni.2014.11.014>
5. Lefranc MP, Giudicelli V, Ginestoux C, Bodmer J, Muller W, Bontrop R, Lemaitre M, Malik A, Barbie V, Chaume D. IMGT, the international ImMunoGeneTics database. *Nucleic Acids Res*. 1999;27:209-212. doi: 10.1093/nar/27.1.209.
6. Jenny M Woof , Dennis R Burton, Human antibody-Fc receptor interactions illuminated by crystal structures. *Nat Rev Immunol*. 2004 Feb;4(2):89-99. doi: 10.1038/nri1266.
7. Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team (2020). *nlme: Linear and Nonlinear Mixed Effects Models*. R package

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Regulatory T (Treg) cells prevent autoimmune diseases and suppress allergic reactions by inhibiting adaptive antibody immune response in the germinal center. Theoretically, this adaptive response lowers the effectiveness of vaccines. Thus the experiment used T-cell suppression treatments to investigate the effect on immunization. These drugs are widely used in post transplant immuno-suppression treatment to prevent rejection.

While we might expect different variance within subject vs between subjects, the number of potential antibodies observed is much higher than the number sampled in a blood draw.