bigWig

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Contents

troduction	_
tup]
Install package	
From Github	
From local directory	
sage	6
bigWig utilities	. :
load.bigWig	
unload.bigWig	
$\operatorname{query.bigWig}^{\circ} \ldots \ldots$	
print.bigWig	
Region	
By region	
Region by Bed	
Step	
Step through region	
Step through region by Bed	
Bed6 files	. 10

Introduction

bigWig is a R package that has utility function for analyzing and manipulating bigWig files.

Setup

Install package

Since bigWig is not available on CRAN, we can not use the basic install.packages('bigWig'). However, there are several ways to install packages. Here I will explain 2 of them.

From Github

The most up to date version of the bigWig pkg is located at bigWig. Using devtools, you can download and install bigWig from github directly.

```
#install devtools if necessary
install.packages("devtools")
library('devtools')
#location of bigWig package and subfolder
pkgLoc='andrelmartins/bigWig'
subFld='bigWig'
devtools::install_github(pkgLoc, subdir=subFld)
```

From local directory

If you don't have an internet connection or don't use github, you can build from the sources file.

```
#install devtools if necessary
install.packages("devtools")
library('devtools')
#Set the working directory to the directory where the source files are located
setwd('~/Dir')
build()
```

Usage

After installation like any other package bigwig needs to be loaded with

```
library(bigWig)
```

For these examples, we will use PRO-seq data from GSE126919_RAW.

Download the tar ball and unpack it in a local directory of your choice. Then in R make sure that you define the directory and filename.

```
#directory where data is stored
dtDir='/home/directory'
# specific bigWig file being used
dtFn='GSM3618124_HEK293T_TIR1_C14_3hrDMS0_rep1_minus_body_0-mer.bigWig'
```

bigWig utilities

These a functions that load, unload, query and print the information that is in each bigWig.

load.bigWig

```
load.bigWig(filename, udcDir = NULL)
```

- arguments
 - filename [required] is a string, which is either the local file directory or URL.
 - udcDir is a string which is the location for storing cached copies of remote files locally, while in
 use. Theseare destroyed when you unload the bigWig. If left as the default udcDir = NULL, then
 it uses /tmp/udcCache.

load.bigWig creates a list in R. This list contains relevant informasyion about the bigWig file and serves as a pointer to the underlying C object of the entire bigWig file. The only parameter required for this is a string of the location and filename.

```
bw=load.bigWig(paste0(dtDir, dtFn))
```

unload.bigWig

```
unload.bigWig(bw)
```

- arguements
 - bw is the R pointer created in load.bigWig

Use unload.bigWig(bw) to destroy the C object and remove it from memory. This does not clear the R list. To do that use rm() or remove()

```
unload.bigWig(bw)
ls()
#> [1] "bw" "dtDir" "dtFn"
remove(bw)
ls()
#> [1] "dtDir" "dtFn"
```

query.bigWig

```
query.bigWig(bw, chrom, start, end, clip = TRUE)
```

- arguments
 - bw is the R pointer created in load.bigWig
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - clip is a logical value; if TRUE bigWig regions are clipped to the query interval.

query.bigWig allows you to search the bigWig files for specific chromosomes (chrom='chr1', a string representative of the desired chromosome within a defined window (start=1, end = 12000 both are integers and end is inclusive meaning it searches up to and including end). It then prints query results to the command line.

```
query.bigWig(bw, chrom='chr1', start=1, end=12000)
#>
        start
                end value
#> [1,] 10496 10497
#> [2,] 10500 10501
                        1
#> [3,] 10518 10519
                        1
#> [4,] 10521 10522
                        2
#> [5,] 10527 10528
                        1
#> [6,] 10535 10536
                        1
#> [7,] 10541 10542
                        1
#> [8,] 10554 10555
                        1
#> [9,] 10933 10934
                        1
#> [10,] 11081 11082
                        2
#> [11,] 11165 11166
#> [12,] 11456 11457
                        1
#> [13,] 11584 11585
```

You can set the query to a variable for storage

```
bwQ=query.bigWig(bw, chrom='chr1', start=1, end=20000)
bwQ[3]
#> [1] 10518
```

Then access the array like any other indexed array. This returns the entire row.

```
bwQ[1,]
#> start end value
#> 10496 10497 1
```

This returns the specific row and column.

```
bwQ[1,2]
#> end
#> 10497
```

It can be accessed by keyword too.

```
bwQ[1,'start']
#> start
#> 10496
```

print.bigWig

print.bigWig(bw) is used to print all of the attributes contained within the object.

```
print.bigWig(bw)
```

 \bullet arguments

```
- bw is the R pointer created in load.bigWig
```

```
#> bigWig
#> version: 4
#> isCompressed TRUE
#> isSwapped FALSE
#> primaryDataSize: 11,243,315
#> primaryIndexSize: 176,800
#> zoomLevels: 7
#>
   chromCount: 455
#>
       chr1 248956422
#>
        chr10 133797422
#>
       chr10_GL383545v1_alt 179254
        chr10_GL383546v1_alt 309802
#>
       chr10_KI270824v1_alt 181496
#>
#>
#>
       chrX 156040895
#>
       chrX_KI270880v1_alt 284869
#>
       chrX_KI270881v1_alt 144206
       chrX_KI270913v1_alt 274009
#>
       chrY 57227415
#>
#>
        chrY KI270740v1 random 37240
   basesCovered: 5,010,318
#>
#>
   mean: 1.187913
#> min: 1
#> max: 3034
#> std: 5.843396
```

Region

The following sections group bpQuery and probeQuery functions together because they operate the same except on how they calculate the average.

By region

This set of functions takes a region defined by chrom, start and end and returns the result of the operation on the counts.

- arguments
 - bw is the R pointer created in load.bigWig
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
 - abs.value is a logical argument which determines if the absolute value of the input is performed before the op.
 - bwMap is a bigWig file of areas that cannot be mapped for a reason

This allows you to find out basic information on a specific query. Starting with a specific query,

Operations

Sum op='sum'

To find how many instances there are where there is a 'chr2' in the bigWig

```
region.bpQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='sum')
#> [1] 9
region.probeQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='sum')
#> [1] 9
```

Maximum op='max'

If you want to find the highest number of instances of chr2

```
region.bpQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='max')
#> [1] 2
region.probeQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='max')
#> [1] 2
```

Minimum op='min'

To find the lowest number of instances

```
region.bpQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='min')
#> [1] 1
region.probeQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='min')
#> [1] 1
```

Average op='avg'

To find the average

```
region.bpQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='avg')
#> [1] 0.03673469
region.probeQuery.bigWig(bw,chrom='chr2',start=229990, end=230235, op='avg')
#> [1] 1.285714
```

Notice the difference in the return of the average. This is because bpQuery counts the number of base pairs to use as the denominator of the average. This is the difference of the end value and the start value.

```
230235-229990
#> [1] 245
```

probeQuery counts the number of probes. Essentially, this is the number of rows returned by the query. In this example, it is 7.

abs.value = FALSE

Sometimes, the bigWig will have negative values. To keep these values in the counts the abs.value=TRUE option can be used. For this example, you'll need a different data set. negative bigWig files. Download both

- $GSM3452725_K562_Nuc_NoRNase_minus.bw$
- GSM3452725_K562_Nuc_NoRNase_plus.bw

and store them in thier own directory. Then using the load.bigWig and store them as bw.plus and bw.minus, respectively. We will use them in later examples.

When we run a query on bw.minus, you can see that it returns negative counts. You can check out the appendix to see how to search and find negative values.

```
query.bigWig(bw.minus, chrom='chr1', start=10140, end=10190)
#> start end value
#> [1,] 10151 10153 -1
#> [2,] 10153 10154 -2
#> [3,] 10154 10155 -1
#> [4,] 10158 10160 -1
```

There is a reason the bigWig file returns negative values. We only care about the case that there is a recorded event [+/-]. In this case, we apply the abs.value=TRUE which takes the absolute value of each count before applying the operation. Remember the default is abs.value=FALSE

```
region.probeQuery.bigWig(bw.minus,chrom='chr1',start=10140, end=10190, op='avg')
#> [1] -1.25
region.probeQuery.bigWig(bw.minus,chrom='chr1',start=10140, end=10190, op='avg', abs.value=TRUE)
#> [1] 1.25
```

Region by Bed

- arguments
 - bw is the R pointer created in load.bigWig
 - bed is a dataframe structured like a bed file with columns for chrom, start and end
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position

- end is an integer value designation the ending position
- op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
- abs.value is a logical argument which determines if the absolute value of the input is performed before the op.

This function is similar to region.bpQuery.bigWig except that when defining the areas we want to examine is defined in a bed file rather than chrom, start, and end.

The source of the bed file can be something created by hand or previous identified regions from other experiments. The basics of the bed is that it's in a R data frame.

```
bed=data.frame('chr1',10496,10497)
#set column headers
colnames(bed)=c('chrom','start', 'end')
```

Now this is for a single factor in R. When creating a dataframe in R, it automatically turns strings into factors. This limits the ability to add different chrom designations. Meaning that when created the original bed file, chr1 was the only level created. It will return an error if you just try to add

```
rbind(bed, c('chr2', 10000, 20000))
#> Warning in `[<-.factor`(`*tmp*`, ri, value = "chr2"): invalid factor level,
#> NA generated
#> chrom start end
#> 1 chr1 10496 10497
#> 2 <NA> 10000 20000
```

If you ever want to add different factors, you'll need to use levels()

```
levels(bed$chrom)=c('chr1', 'chr2')
```

Take a look at how the data.frame is structured

```
dim(bed)
#> [1] 1 3
attributes(bed)
#> $names
#> [1] "chrom" "start" "end"
#>
#> $row.names
#> [1] 1
#>
#> $class
#> [1] "data.frame"
bed
#> chrom start end
#> 1 chr1 10496 10497
```

dim returns the size of the matrix [1 row, 3 columns]. while attributes returns information on column names, row names and class type.

You can take this bed file and run it through the bigWig file to see what regions overlap

```
# note: If you leave out op='', it will default to op='sum'
bed.region.bpQuery.bigWig(bw, bed)
#> [1] 1
```

Now adding a few other regions to the data frame

```
bed=rbind(bed, c('chr2', 10500,10501))
```

In the original query, this region is occupied by a chr1 and since the bed file refers to a chr2 the sum should be the same because there is no overlap. Then if you rerun

```
bed.region.bpQuery.bigWig(bw, bed)
#> [1] 1 0
```

We see that the then returned values are 1 and 0. This is because the first region of the bed file overlaps regions of the bigWig, but the second bed region does not overlap any regions of the bigWig.

Now adding a third row to the bed file that will overlap a larger range of the bigWig and rerun

```
bed2=rbind(bed, c('chr1', 13000,14001))
bed.region.bpQuery.bigWig(bw, bed2)
#> [1] 1 0 11
```

The returned values are the sums of the counts in those regions.

Step

The following functions operate over defined steps and is described by step= argument. This means in a given region [start=1 and end=10] and a step=5, the function will create subregions of 5. In this example, it will run on [start=1, end=5] and [start=6, end=10]. Again, probeQuery and bpQuery functions are the same, exce;pt when calculating op=avg.

Step through region

- arguments
 - bw is the R pointer created in load.bigWig
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
 - abs.value is a logical argument which determines if the absolute value of the input is performed before the op.
 - gap.value is an integer value that replaces areas that have no overlaps
 - with.attributes is a logical argument that determines if the results are returned annotated with their source components and/or step size.

The Step function will run through the range provide breaking it up into equal size steps as defined by step =. The key here is that the length of the range [end-start] has to be a multiple of the step. For example if end=21 and start=1, The length of the range is 20. This allows for step = [1,2,4,5,10,20]. The return

is the value of the operation over that step. So if step =1 and op = 'min', then the return would be 20 minimums.

Now if step = 5 and op = 'max', the return will be a 4 element array of the maximum value in the step.

Let's take a look over a 20000 interval start=1, end=20001 and a step=1000.

```
step.bpQuery.bigWig(bw,chrom='chr1',start=1, end=20001, op='sum', step=1000)
#> [1] 0 0 0 0 0 0 0 0 0 0 10 5 11 11 1 1 2 0 4 3
#> attr(,"chrom")
#> [1] "chr1"
#> attr(,"start")
#> [1] 1
#> attr(,"end")
#> [1] 20001
#> attr(,"step")
#> [1] 1000
```

The result is a 20 element array of the sum of all the counts in the interval. Notice that the steps that have no counts are zero. If we needed to fill these values in with a specific number like 10, we use gap.value=10

Step through region by Bed

- arguments
 - bw is the R pointer created in load.bigWig
 - bed is a dataframe structured like a bed file with columns for chrom, start and end
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
 - abs.value is a logical argument which determines if the absolute value of the input is performed before the op.
 - gap.value is an integer value that replaces areas that have no overlaps
 - with.attributes is a logical argument that determines if the results are returned annotated with

their source components and/or step size.

This is similar to bed.region.bigWig(), where you can add a bed of regions that you are interested in.

```
#Create bed dataframe
bed3 = data.frame('chr1', 15000, 25000)
colnames(bed3)=c('chrom', 'start', 'end')
bed3=rbind(bed3, c("chr1", 30000, 35000))
bed.step.bpQuery.bigWig(bw, bed3, step=1000, op='avg', with.attributes=FALSE)
#> [[1]]
#> [1] 0.001 0.002 0.000 0.004 0.003 0.002 0.007 0.010 0.003 0.004
#>
#> [[2]]
#> [1] 0 0 0 0 0 0
```

Notice that the defined regions in the bed file are exact multiples of the step. This is explained in the bed.bpQuery.bigWig example. The other attribute of this bed file is the regions defined do not need to be the same size. row 1 in the bed files contains 10 steps, while Row 2 has 5 steps. the final aspect of this example is that bpQuery version uses the step size as the denominator in the average. While probeQuery will use the number of rows in the query

In the probe version, we end up with where there are no overlapping regions. This is because dividing by zero is not possible. Instead the function returns a NA.

Bed6 files

bed.region.bpQuery.bigWig() and bed.step.bpQuery.bigWig() have counterparts that can take a bed6 file. The bed6 file is similar to a bed file except it has 3 more columns of data.

Remember the standard bed file has chrom, start and end. The bed6 adds name, score, strand columns to its structure. For these functions, we only need the added strand column. However this column needs to be in the 6th position. Meaning even though name and score columns exist in the dataframe, they can be populated with nulls. You could populate it with identifying information, but the function essentially ignores them. The strand column requires either a + or - to denote the plus or minus strand.

Here is an example

```
bed6=data.frame('chr1',1,100000,'','','+')
colnames(bed6)=c('chrom', 'start', 'end', 'name', 'score', 'strand')
```

This introduces the biological concept of plus and minus strands. This refers to the situation where the biological experiment returns the sense [plus strand] and it's complimentary RNA strand, antisense [minus strand]

bed6.region

```
with.attributes = TRUE, as.matrix = FALSE,
follow.strand = FALSE)
```

- arguments
 - bw.plus is the R pointer created in load.bigWig and refers to the plus strand
 - bw.minus is the R pointer created in load.bigWig and refers to the minus strand
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
 - abs.value is a logical argument which determines if the absolute value of the input is performed before the op.
 - gap.value is an integer value that replaces areas that have no overlaps
 - with.attributes is a logical argument that determines if the results are returned annotated with their source components and/or step size.

Let's look at an example. We will use data from the negative values used with abs.value = TRUE In the Region section.

```
dtDir = '/home/directory'
dtFnPlus='GSM3452725_K562_Nuc_NoRNase_plus.bw'
dtFnMinus='GSM3452725_K562_Nuc_NoRNase_minus.bw'
bw.plus=load.bigWig(paste0(dtDirNeg, dtFnPlus))
bw.minus=load.bigWig(paste0(dtDirNeg, dtFnMinus))
```

Using the bw.plus and bw.minus strands, we can evaluate a bed6.region function. First, take a look at the query for each strand.

```
query.bigWig(bw.minus, chrom='chr1', start=25000, end=50000)
#> start end value
#> [1,] 28567 28568 -1
#> [2,] 28570 28571 -1
#> [3,] 46605 46606 -1
#> [4,] 47071 47072 -1
#> [5,] 49218 49219 -1
query.bigWig(bw.plus, chrom='chr1', start=25000, end=50000)
#> start end value
#> [1,] 29459 29460 1
```

These will be used as reference for when we use the function.

```
bed6=data.frame('chr1',25000,50000,'','','+')
colnames(bed6)=c('chrom', 'start', 'end', 'name', 'score', 'strand')
```

This particular bed file defines a region between start = 25000 and end = 50000 on the + strand.

The query of the plus strand shows only one overlapping region. The average of 1 region with 1 count is 1.

Now add another row to our bed6 file and rerun the previous bed6.region.probeQuery.bigWig function.

```
levels(bed6$strand)=c('+', '-')
bed6=rbind(bed6, c('chr1', 25000, 50000, '', '', '-'))
bed6.region.probeQuery.bigWig(bw.plus, bw.minus, bed6, op='sum', abs.value = FALSE, gap.value=0)
#> [1] 1 -5
```

Similarly to the bed.region function the return is 2 values one for each overlapping region.

We can invoke ab.value = TRUE argument and our second result change to a positive value.

bed6.step

- arguments
 - bw.plus is the R pointer created in load.bigWig and refers to the plus strand
 - bw.minus is the R pointer created in load.bigWig and refers to the minus strand
 - chrom is a string referring to what chromosome is referenced
 - start is an integer value designation the starting position
 - end is an integer value designation the ending position
 - op is a string representing the operation to perform on the step.
 - * sum adds all the counts
 - * avg averages the counts
 - * min finds the smallest count
 - * max finds the largest count
 - abs.value is a logical argument which determines if the absolute value of the input is performed before the op.
 - gap.value is an integer value that replaces areas that have no overlaps
 - with.attributes is a logical argument that determines if the results are returned annotated with their source components and/or step size.

This is just like step.bed.xxx functions.

as.matrix

Here the attribute as.matrix will be introduced. This attribute causes the output to be a matrix.

follow.strand

follow.strand is an attribute that will switch the direction of how it reads the - strand. This allows you to read both strands + and - from the 3' end. To show this we can see that the results are mirrors of each other.

Mirrors