

Restriction Endonucleases: The Molecular Scalpel

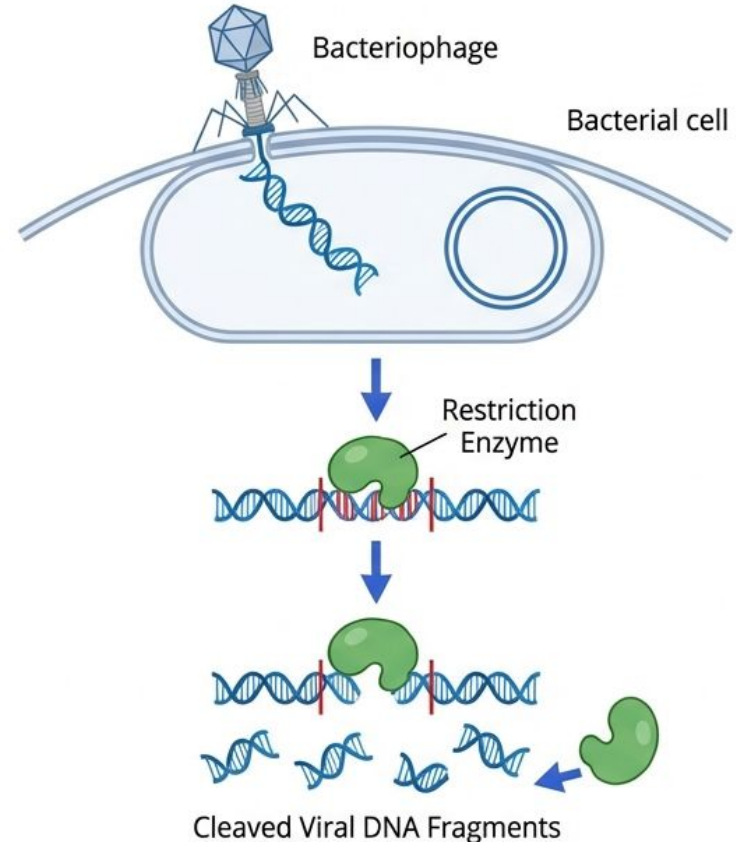
Principles, Mechanisms, and Clinical Applications



Comprehensive Review of Molecular Scissors

Definition & Biological Origin

- **Core Definition:** Restriction enzymes are site-specific endonucleases produced by bacteria and archaea that cleave the phosphodiester bond of DNA at specific recognition sequences.
- **Evolutionary Purpose:** A primitive bacterial immune system.
- **Target:** Bacteriophages (invading viruses).
- **Mechanism:** The enzyme “restricts” viral replication by degrading foreign DNA upon entry.

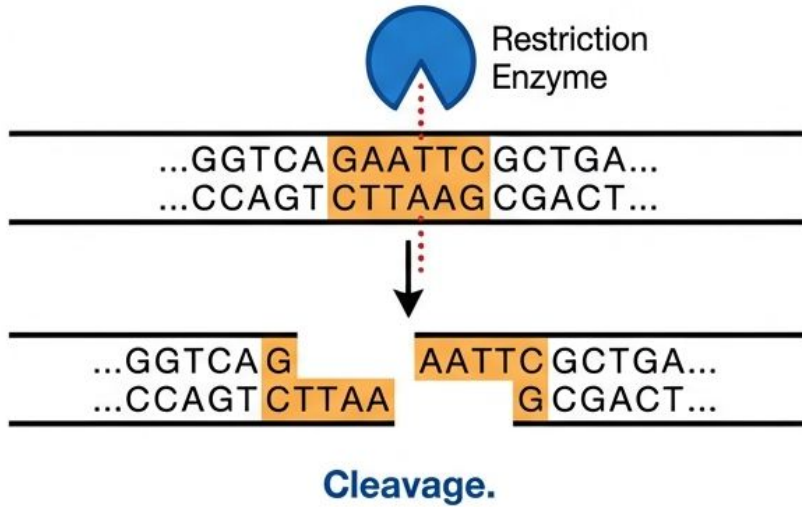


The Restriction-Modification (R-M) System

Sword and Shield

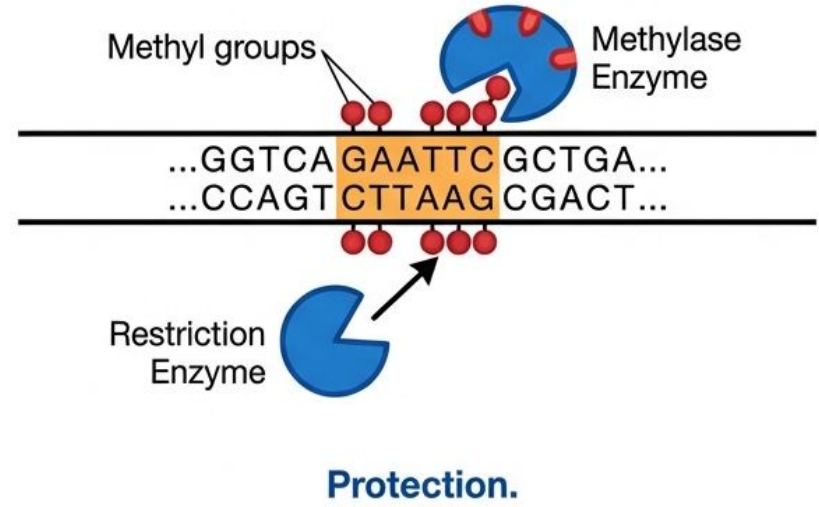
The Sword (Restriction)

Foreign DNA (Unmethylated):
Recognized as a threat.



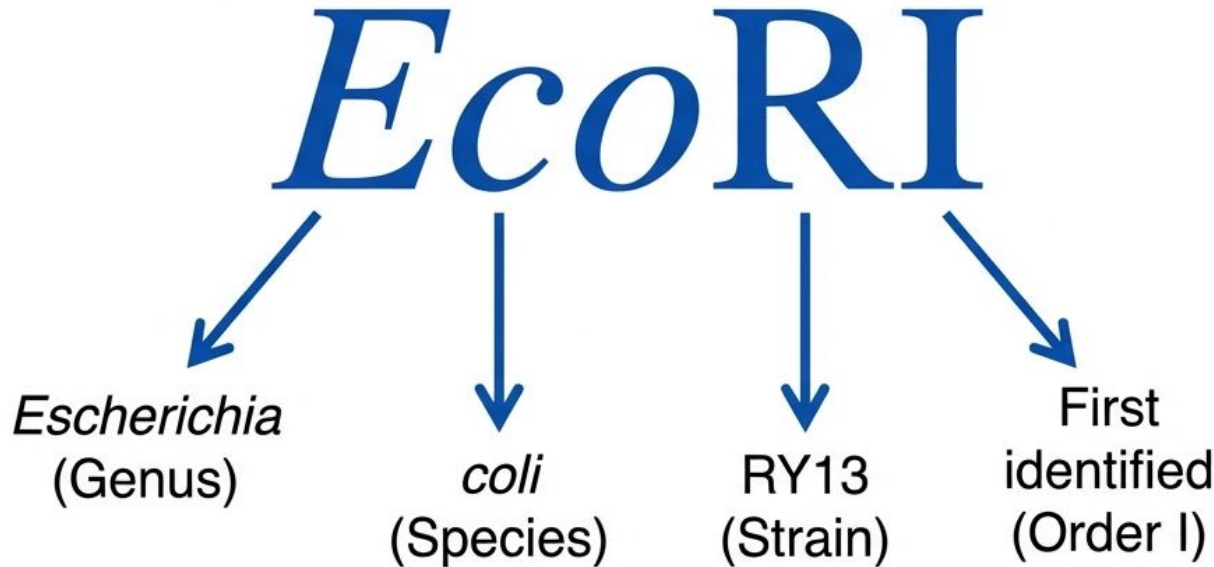
The Shield (Modification)

Host DNA (Methylated):
Protected from cleavage.



Key Concept: Methylation masks the recognition site, preventing 'suicide' by the bacterium's own enzymes.

Nomenclature Rules



**Other Examples:

HindIII: *Haemophilus influenzae* (strain Rd), 3rd enzyme.

BamHI: *Bacillus amyloliquefaciens* (strain H), 1st enzyme.

Classes of Restriction Enzymes

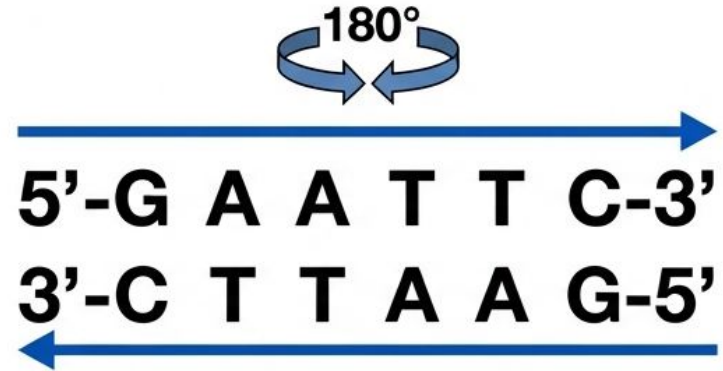
| Type I | Type II (The Lab Standard) | Type III |
|--|---|---|
| Cut Site: Random (>1000bp away) | Cut Site: At specific recognition site | Cut Site: 25-27bp downstream |
| Cofactors: ATP, Mg ²⁺ , SAM | Cofactors: Mg ²⁺ Only (No ATP) | Cofactors: ATP, Mg ²⁺ |
| Utility: Low (Non-specific) | Utility: High (Precise & Reproducible) | Utility: Low |

Type II enzymes are the workhorses of molecular biology because they cut exactly at the recognition sequence.

Recognition Sites & Palindromes

The Concept

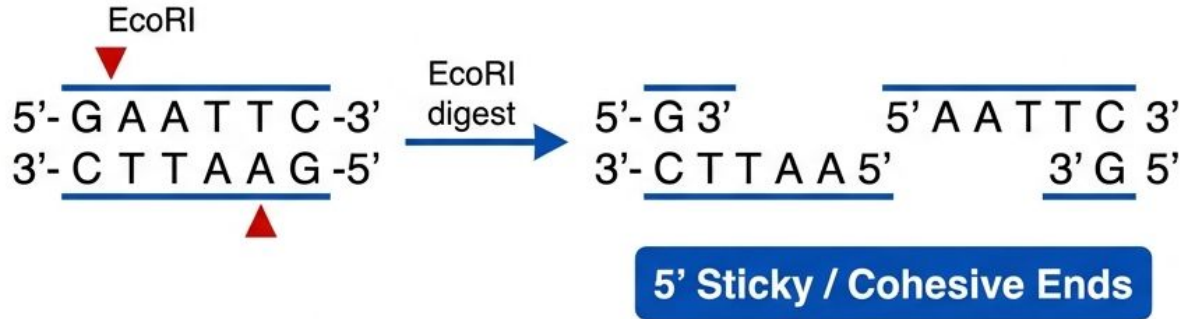
Restriction sites typically display Dyad Symmetry (Palindromes). They read the same 5' → 3' on both strands.



Frequency

- **4bp Cutter (*AluI*):** AGCT (Frequent, ~256bp)
- **6bp Cutter (*EcoRI*):** GAATTC (Standard, ~4,096bp)
- **8bp Cutter (*NotI*):** GCGGCCGC (Rare, Genome Mapping)

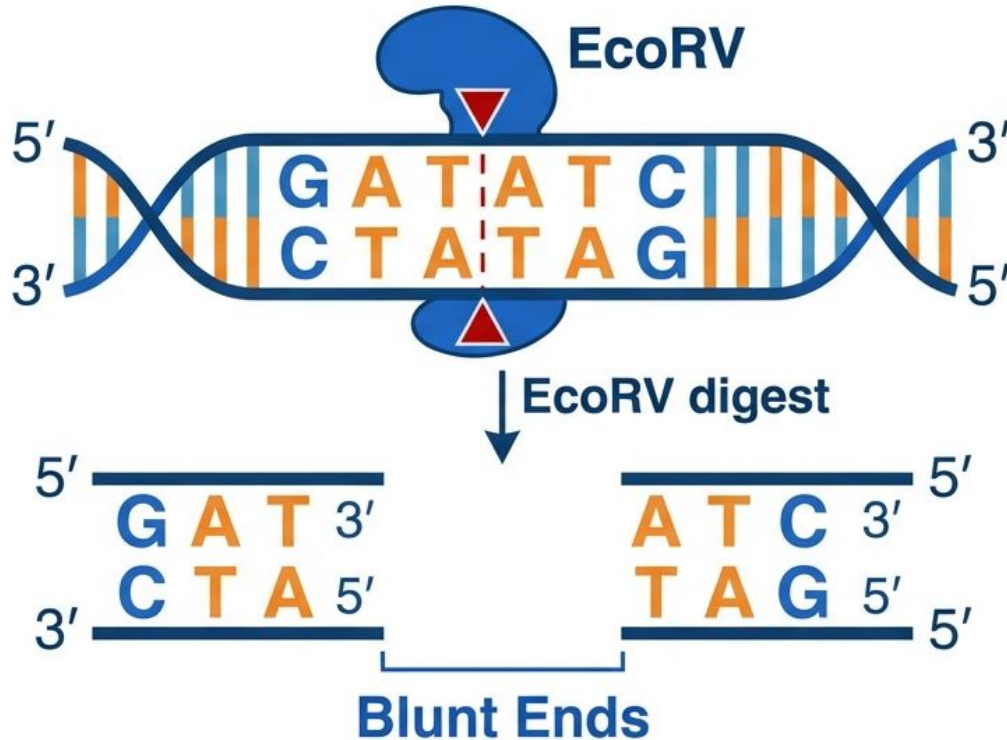
Cleavage Pattern 1: Sticky Ends



Mechanism: Staggered cleavage creates overhangs.

Benefit: Complementary overhangs hydrogen bond easily, increasing ligation efficiency.

Cleavage Pattern 2: Blunt Ends

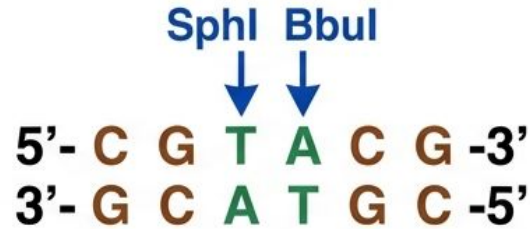


- **Mechanism:** Cleavage at the axis of symmetry.
- **Pros:** Universally compatible (Blunt-to-Blunt).
- **Cons:** Lower ligation efficiency (no hydrogen bonding to stabilize).

Isoschizomers & Neoschizomers

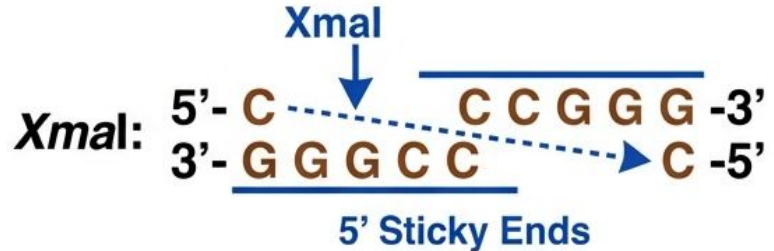
Isoschizomers (Same Site, Same Cut)

SphI and **BbuI**



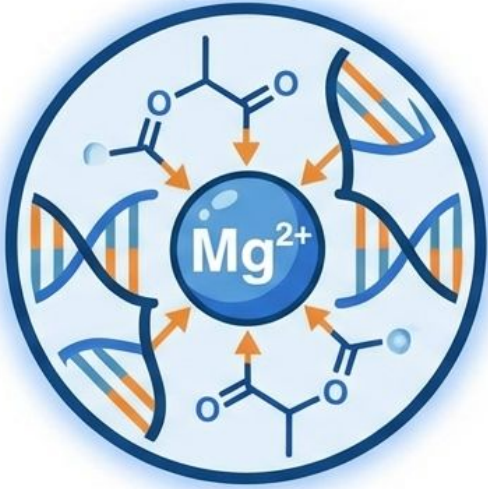
Neoschizomers (Same Site, Different Cut)

SmaI vs **XmaI** (Target: CCCGGG)



Neoschizomers allow researchers to generate different ends (sticky vs blunt) from the exact same DNA sequence.

Reaction Conditions



Cofactor

Magnesium (Mg^{2+}) is essential for Type II catalytic activity.



Buffer

pH 7.0 - 8.0 and specific ionic strength (NaCl/KCl) required for specificity.



Temperature

37°C (Standard for *E. coli* enzymes). **Exception:** TaqI requires **65°C**.

Star Activity

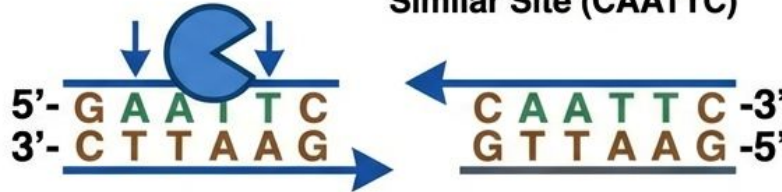
Star Activity: When restriction enzymes lose specificity and cleave sequences *similar* but *not identical* to their recognition site.

Correct Site (GAATTC)

5'- G A A T T C ... C A A T T C - 3'
3'- C T T A A G ... G T T A A G - 5'

Similar Site (CAATTC)

Normal
Conditions



Star
Conditions



Causes



High Glycerol (>5%)



High Enzyme
Concentration
(>100 U/μg)



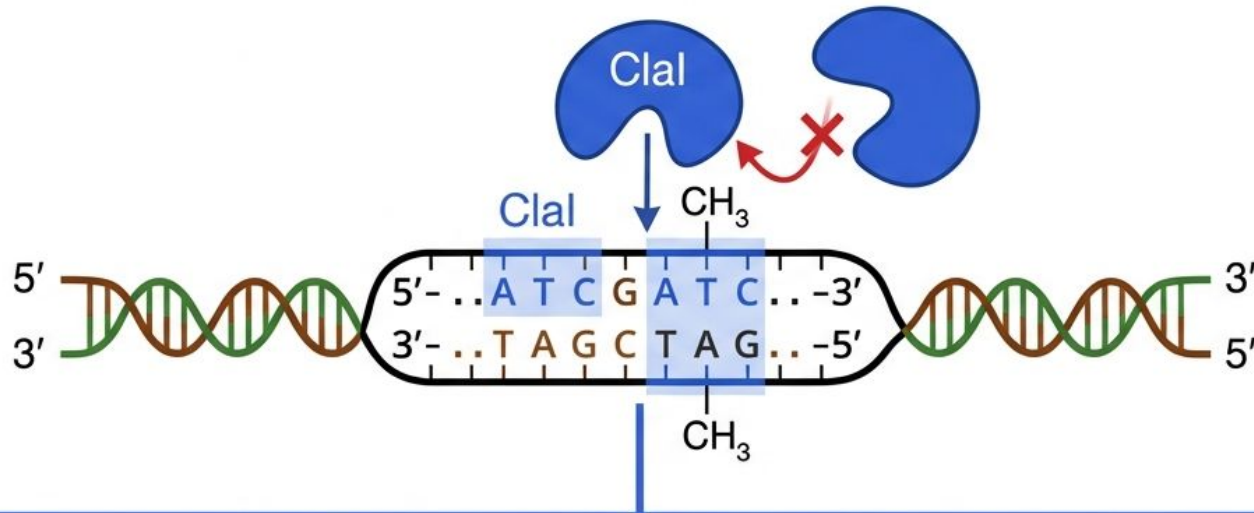
Low Ionic Strength



Prolonged
Incubation time

Methylation Sensitivity

Standard *E. coli* strains express Dam and Dcm methylases. If a restriction site overlaps with a methylation site, the enzyme may be blocked.



Steric Hindrance: Methylation prevents enzyme binding.

Solution: Use *dam*-/*dcm*- competent cells to propagate DNA for these sensitive enzymes.

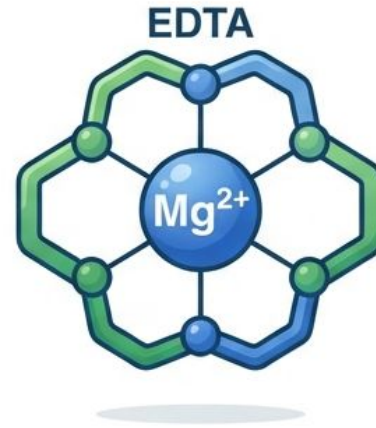
Thermal Inactivation

Heat (The Standard)



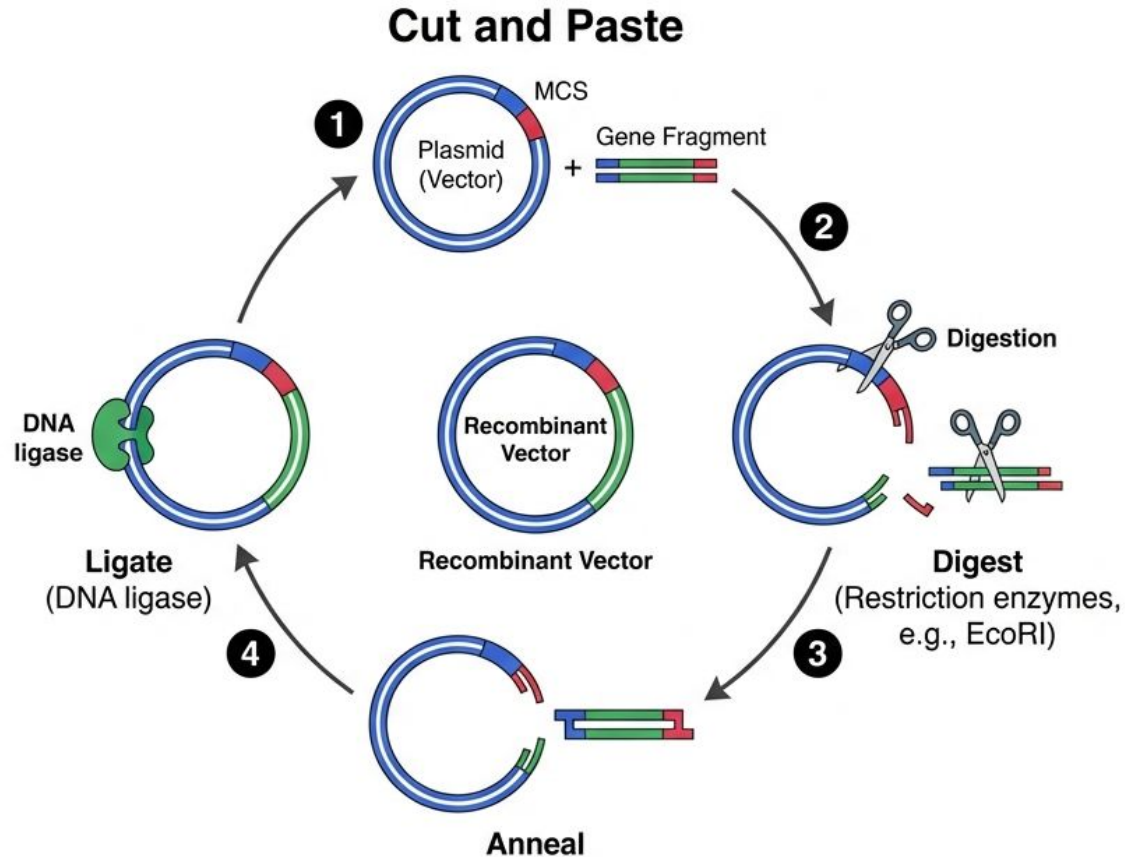
Heat Inactivation: 65°C for 20 minutes denatures most enzymes (e.g., EcoRI). Essential before ligation.

Chelation (The Alternative)



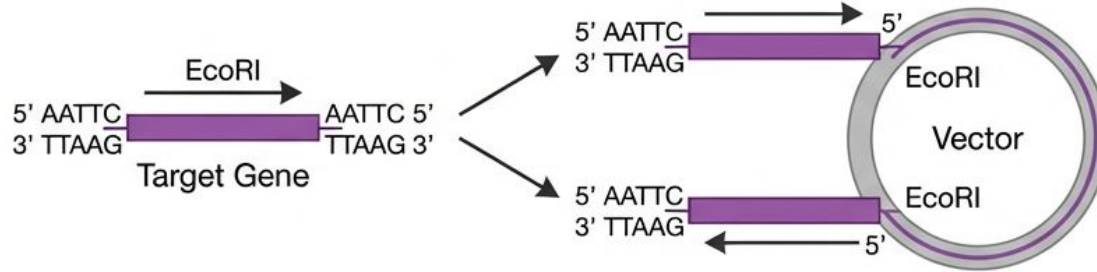
Chelation: For heat-resistant enzymes (e.g., BamHI, TaqI), add **EDTA**. EDTA binds Mg²⁺, starving the enzyme of its cofactor.

Gene Cloning (Recombinant DNA)



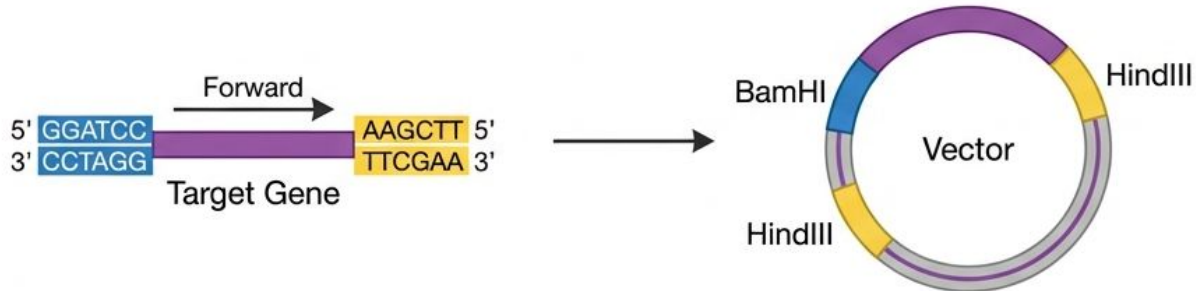
Directional Cloning

Single Enzyme (EcoRI)



50% Wrong Orientation

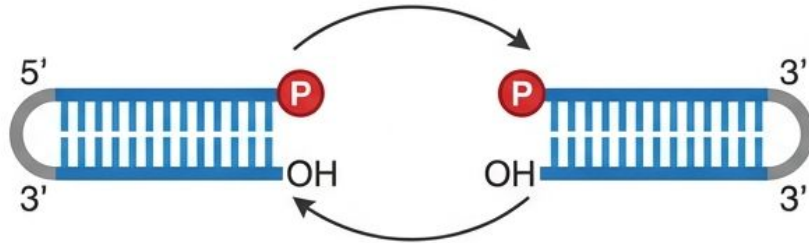
Double Enzyme (BamHI + HindIII)



100% Correct Orientation

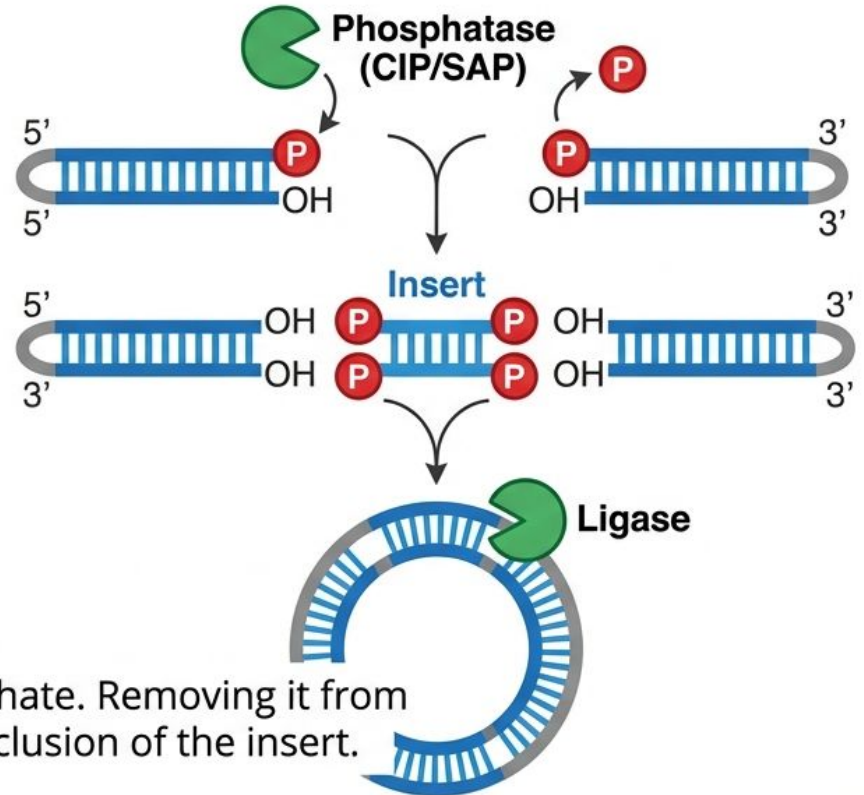
Preventing Self-Ligation

The Problem



Vectors tend to re-ligate to themselves (empty vector).

The Solution



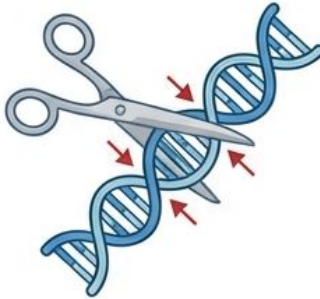
Ligase requires a 5' Phosphate. Removing it from the vector forces the inclusion of the insert.

RFLP Analysis

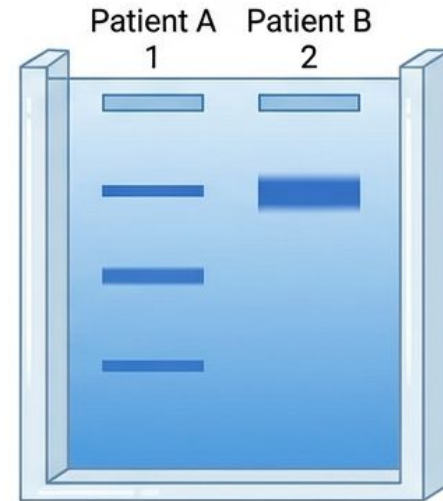
Step 1:
DNA Extraction
from two patients.



Step 2:
Digestion with
Restriction Enzyme.



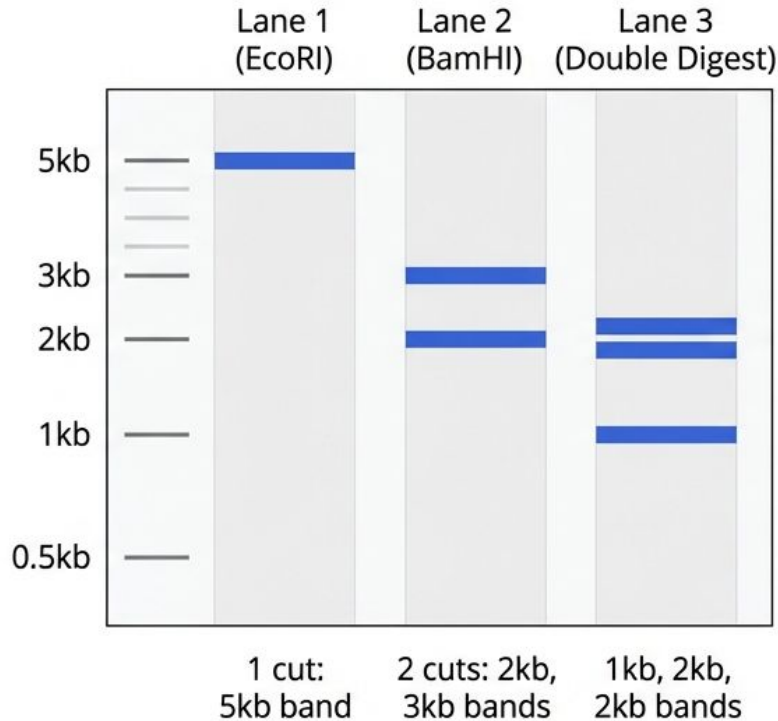
Step 3:
Electrophoresis.



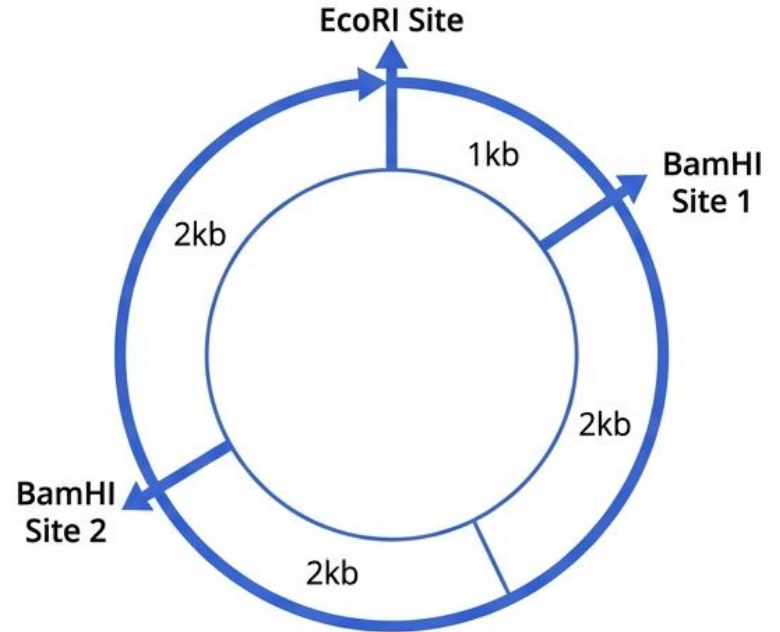
Differences in DNA sequence create different fragment lengths, visible as distinct banding patterns.

Restriction Mapping

The Gel Data



The Map Construction

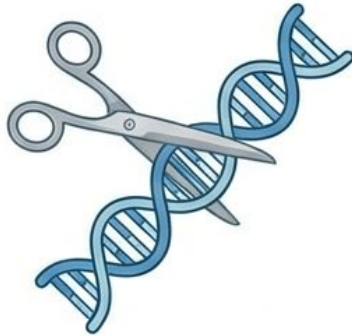


Logic puzzle visualization: Map construction based on fragment sizes and digest patterns.

Summary & Evolution: The Next Generation

Restriction Enzymes

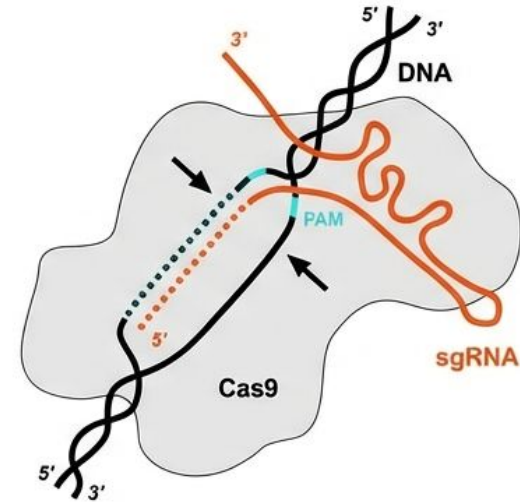
- Fixed recognition (4-8bp).
- Dependent on natural palindromes.
- The Foundation of Cloning.



Molecular Scissors

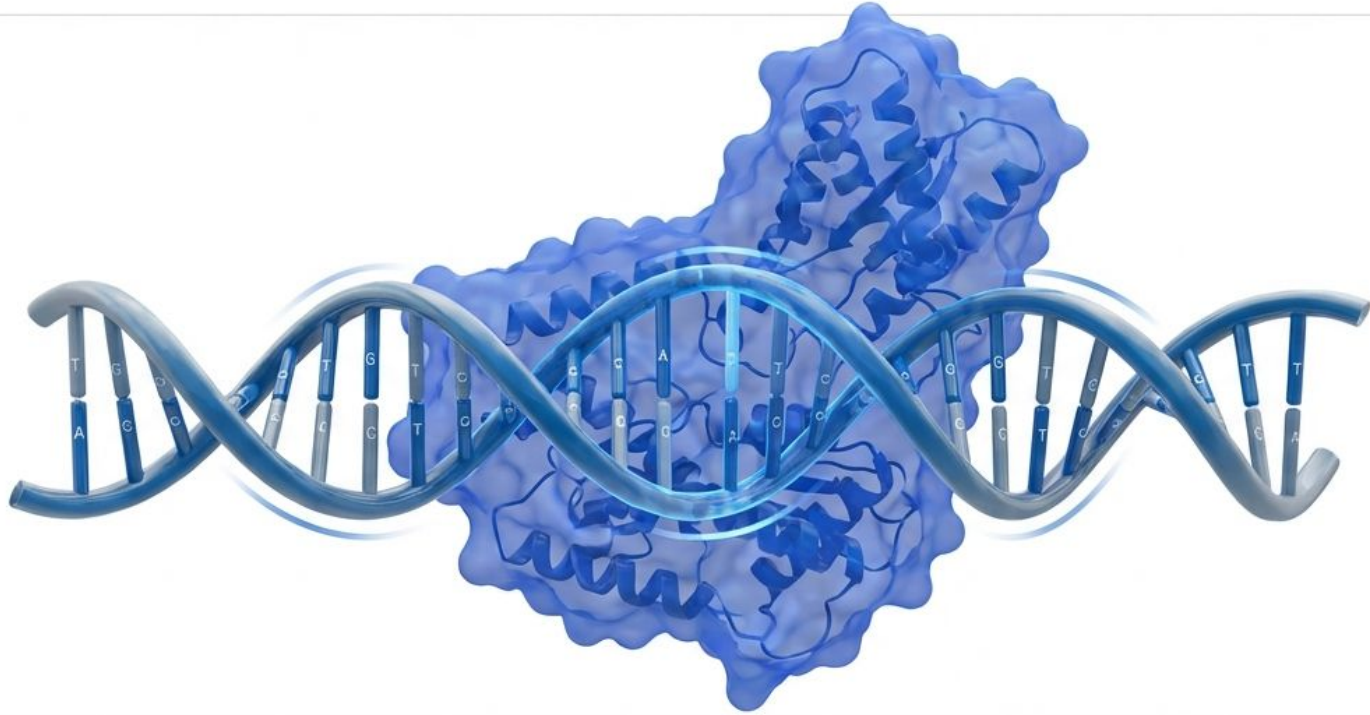
CRISPR-Cas9

- Programmable targeting (Guide RNA).
- High Specificity (~20bp).
- Genome Editing in living cells.



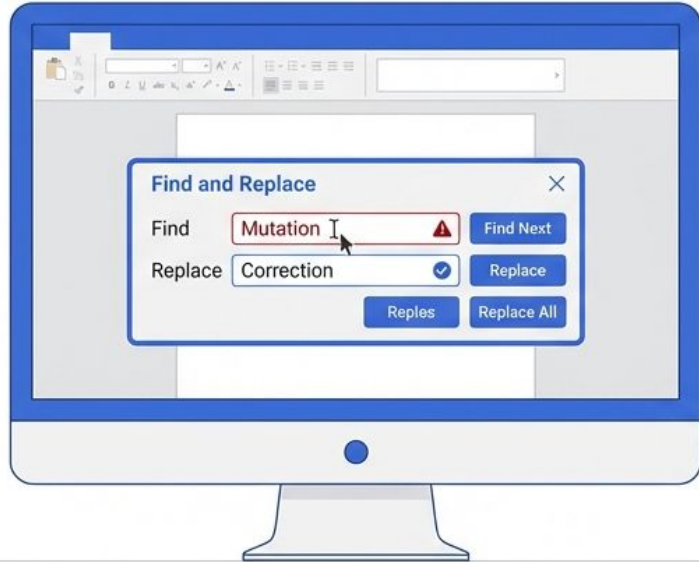
CRISPR-Cas9: The Genetic Scissors

A Revolution in Genome Editing



The power to rewrite the code of life.

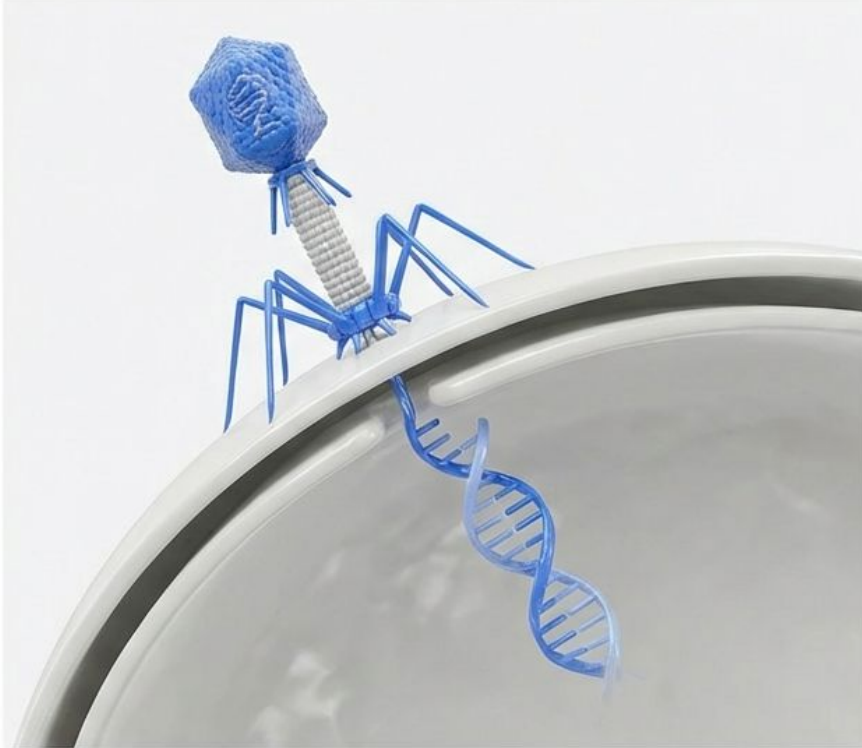
The 'Find and Replace' for DNA



Imagine the genome as a document with 3 billion letters.

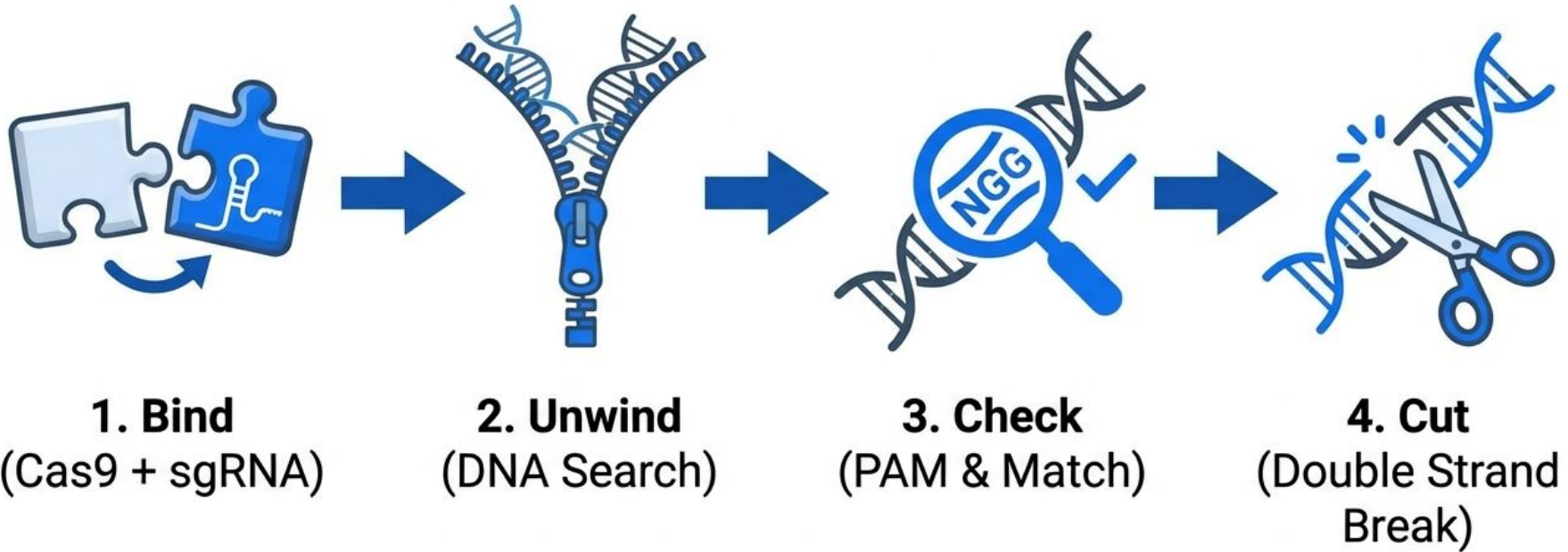
- **The Problem:** A 'typo' (mutation) causes disease.
- **The Solution:** CRISPR acts like the Ctrl+F tool.
- **The Function:** It searches for a specific sequence and edits it with precision.

A Bacterial Shield



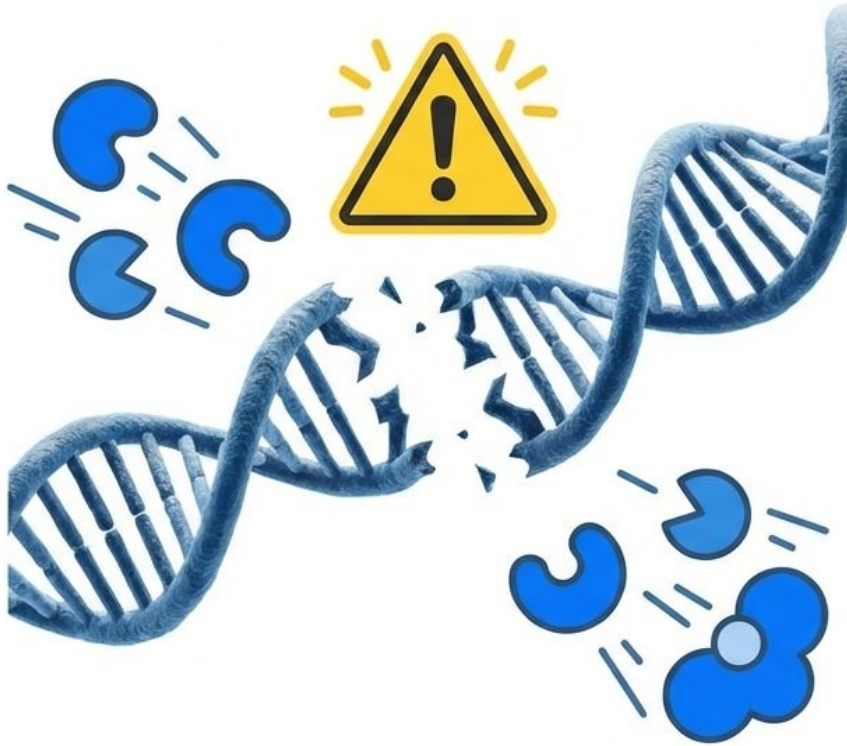
- **Origin:** CRISPR was not invented in a lab; it was discovered in nature.
- **The War:** It is an adaptive immune system found in bacteria and archaea.
- **The Enemy:** Bacteriophages (viruses that attack bacteria).
- **The Defense:** Bacteria use CRISPR to 'remember' and destroy viral invaders.

Mechanism Summary: Step-by-Step



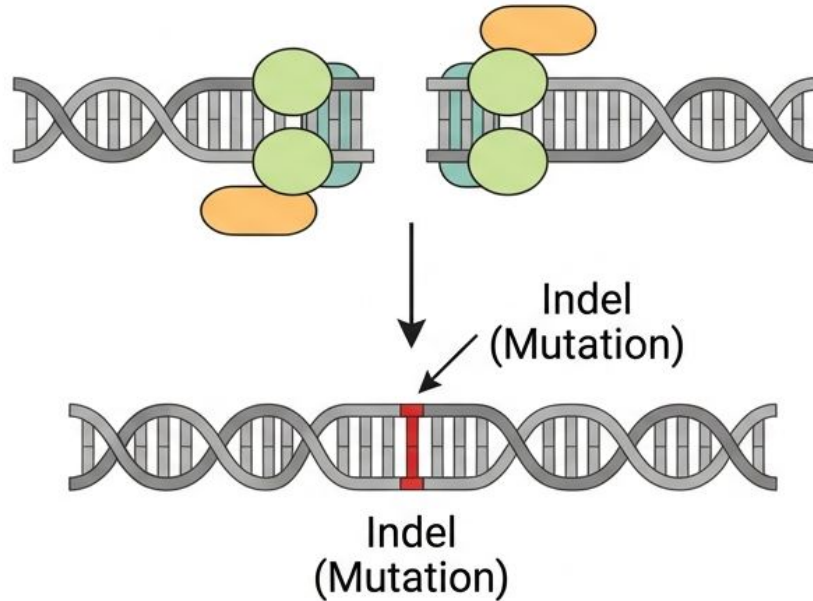
The complex scans the genome, verifies the address, and makes the cut.

The Cut: Cellular Panic Mode



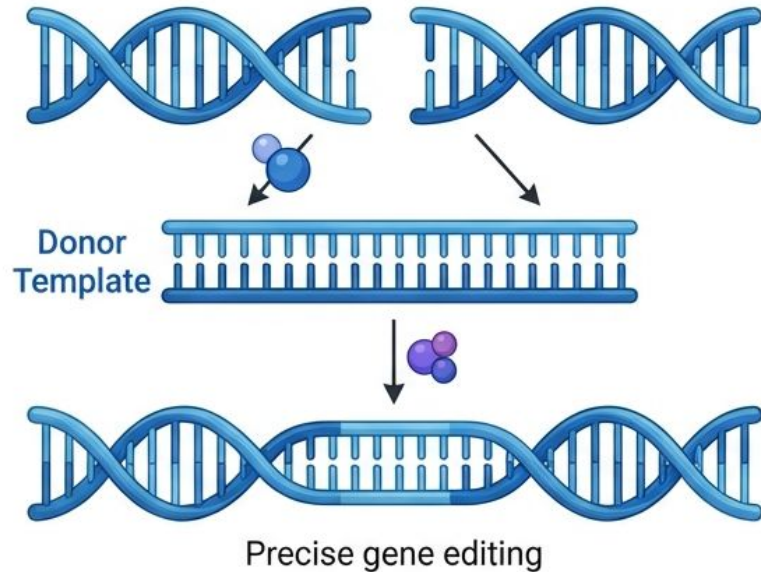
- **The Status:** Cas9 has created a Double-Strand Break (DSB).
- **The Crisis:** To the cell, this is a life-threatening emergency.
- **The Response:** Emergency Repair Crews (cellular enzymes) rush to fix the break.
- **The Outcome:** Depends entirely on HOW the cell fixes this break.

Repair Pathway 1: NHEJ (The Quick Fix)



- **Name:** Non-Homologous End Joining (NHEJ).
- **The Method:** Glue the ends back together immediately.
- **The Result:** Error-prone. Random letters are lost or added (Indels).
- **The Application:** Gene Knockout. The mutation breaks the gene's instructions, effectively turning it off.

Repair Pathway 2: HDR (The Precise Edit)



- **Name:** Homology-Directed Repair (HDR).
- **The Method:** Copy-Paste from a Template.
- **The Requirement:** Scientists inject a 'Donor DNA' template along with Cas9.
- **The Result:** Precise editing. The cell copies the correct sequence from the donor.
- **The Application:** Gene Knock-in. Fixing a typo or inserting a new gene.

**THANKS FOR
LISTENING**
