



THE 3<sup>rd</sup> VIETNAM SCHOOL OF BIOLOGY (VSOB-3)

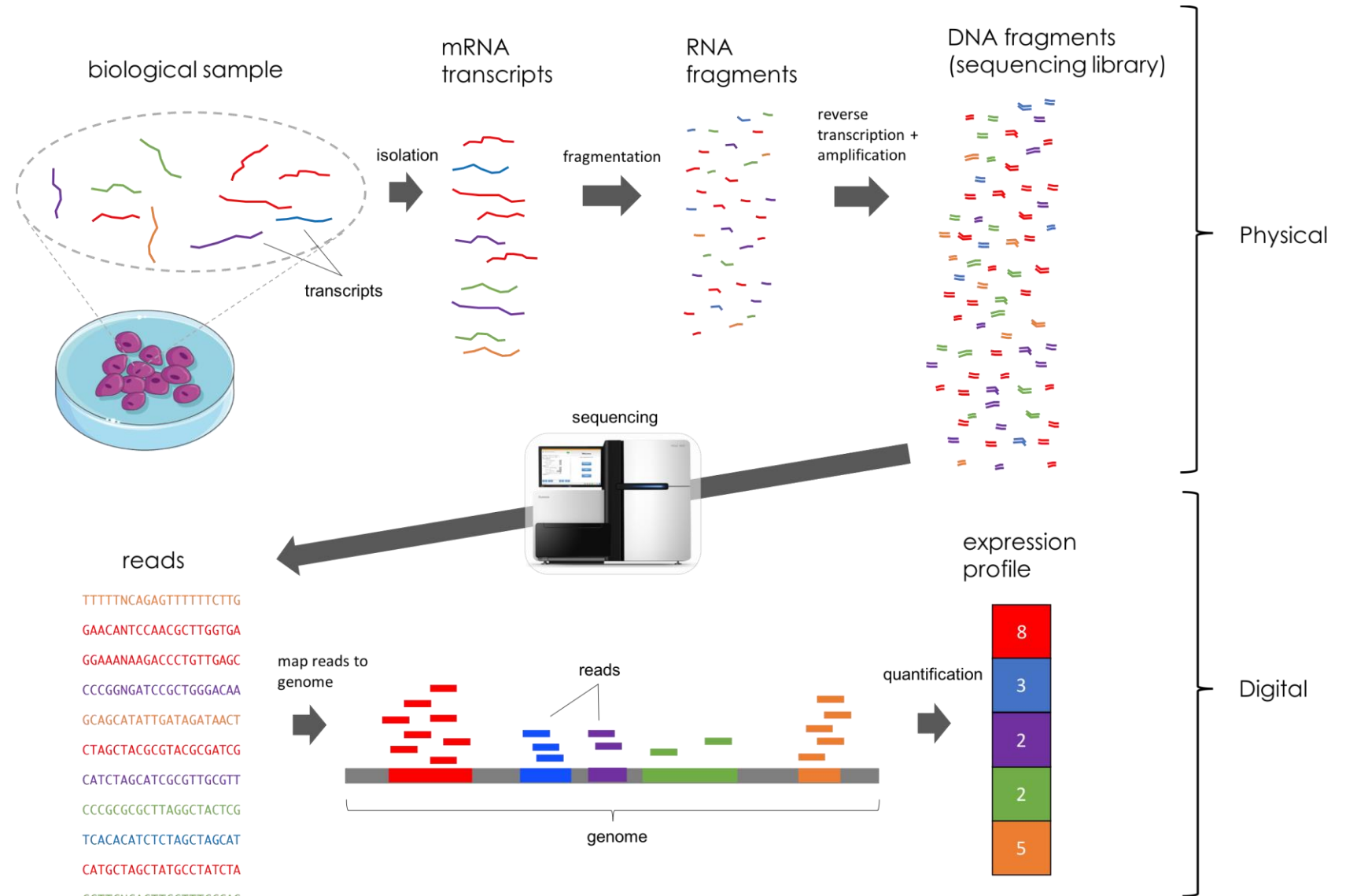
# Bioinformatic Analysis For Bulk RNAseq Data

*December 06th-08th, 2024, ICISE, Quy Nhon, Vietnam*

# A Guide for Designing and Analyzing RNAseq Data

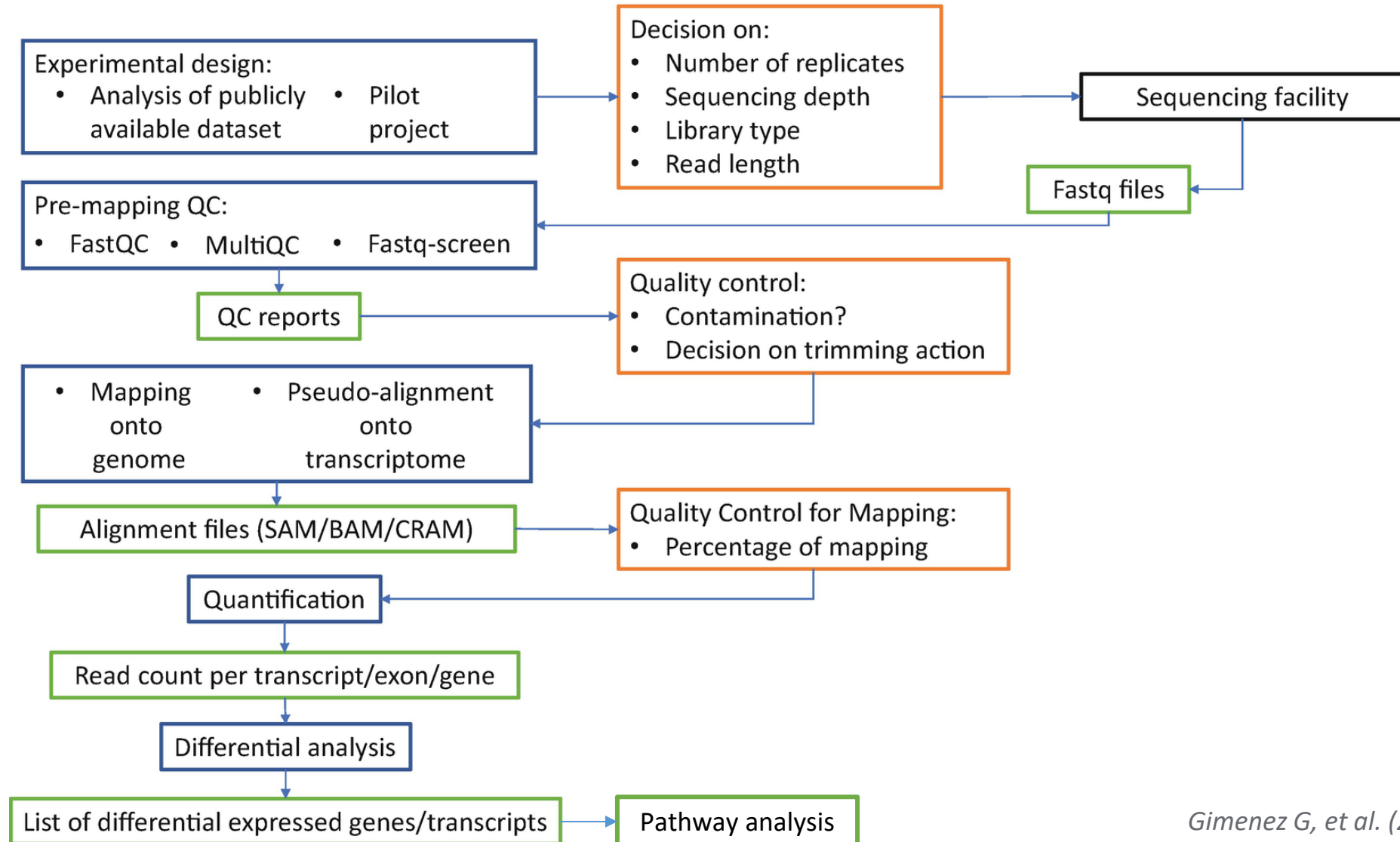
NGUYEN Thuy Vy

# General steps of an RNAseq experiment



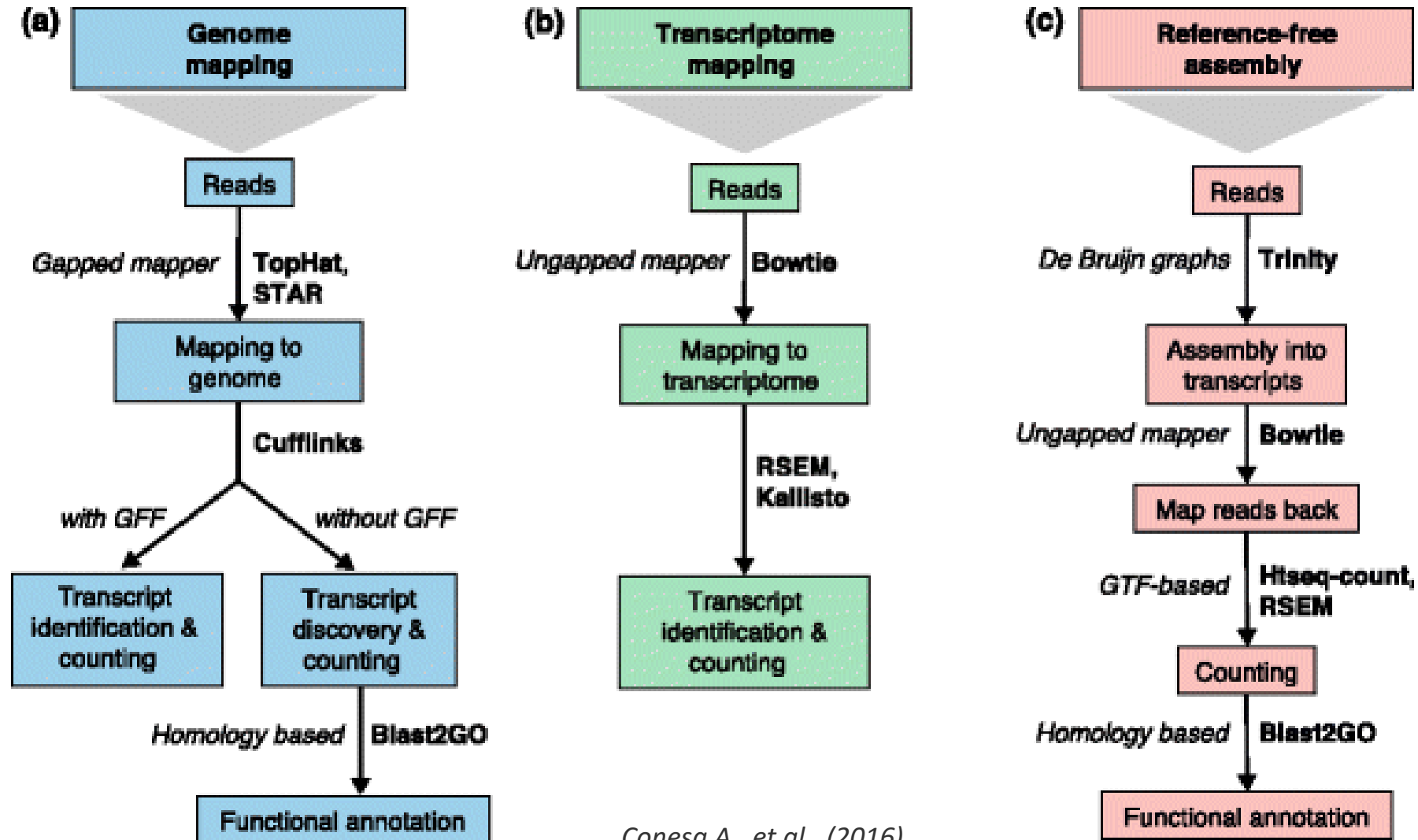
[https://mbernste.github.io/posts/rna\\_seq\\_basics/](https://mbernste.github.io/posts/rna_seq_basics/)

# RNAseq Experimental Design and Data Analysis



*Gimenez G, et al. (2023)*

# RNAseq Data Analysis



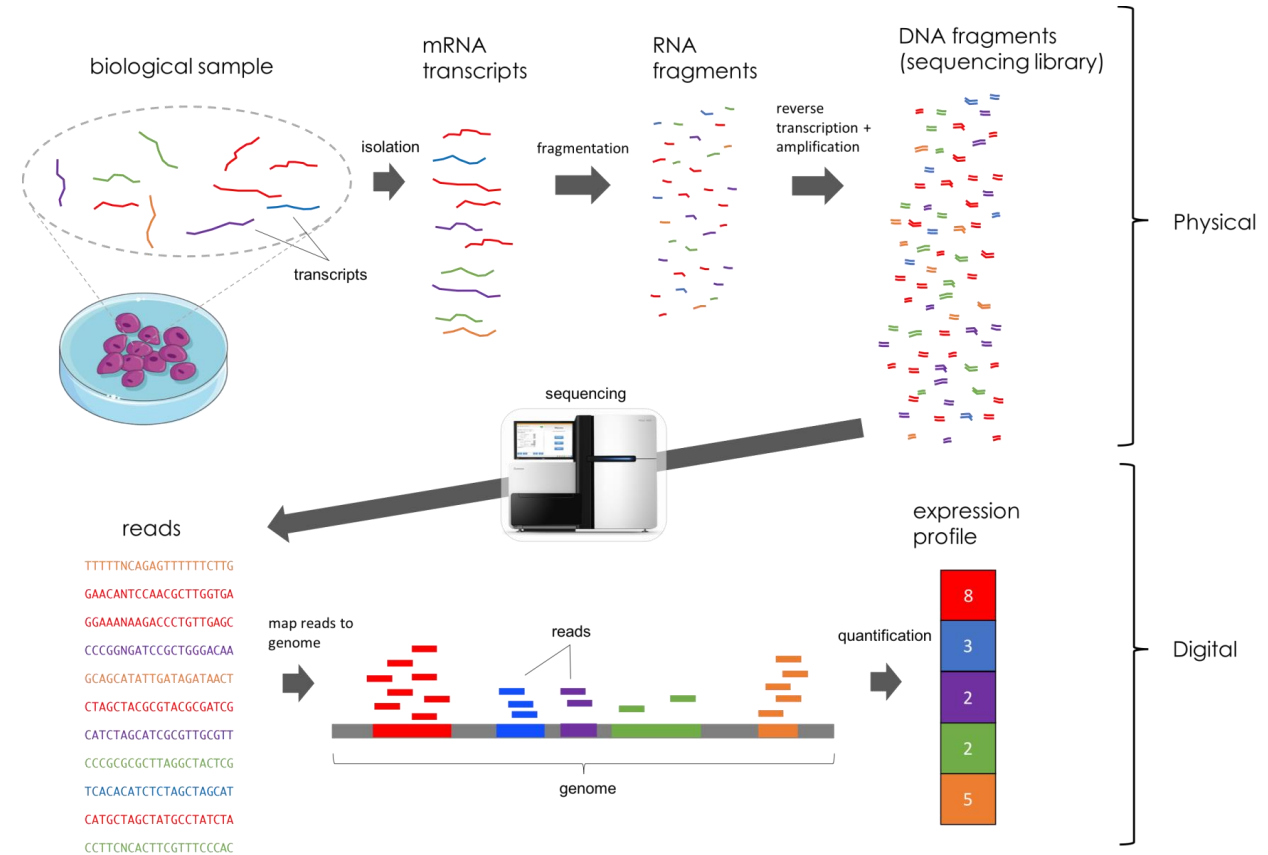
Conesa A., et al., (2016)

# Replicates

- Compared to arrays, RNA-Seq is a very clean technical measure of expression
  - Generally, don't run **technical** replicates
  - Must run **biological** replicates
- For clean systems (eg cell lines) 3x3 or 4x4 is common
- Higher numbers required as the system gets more variable
- Always plan for at least one sample to fail
- Randomise across sample groups

# How many reads

- Typically aim for 20 million reads for human/mouse sized genome
- More reads:
  - De-novo discovery
  - Low expressed transcripts
- More replicates more useful than more reads

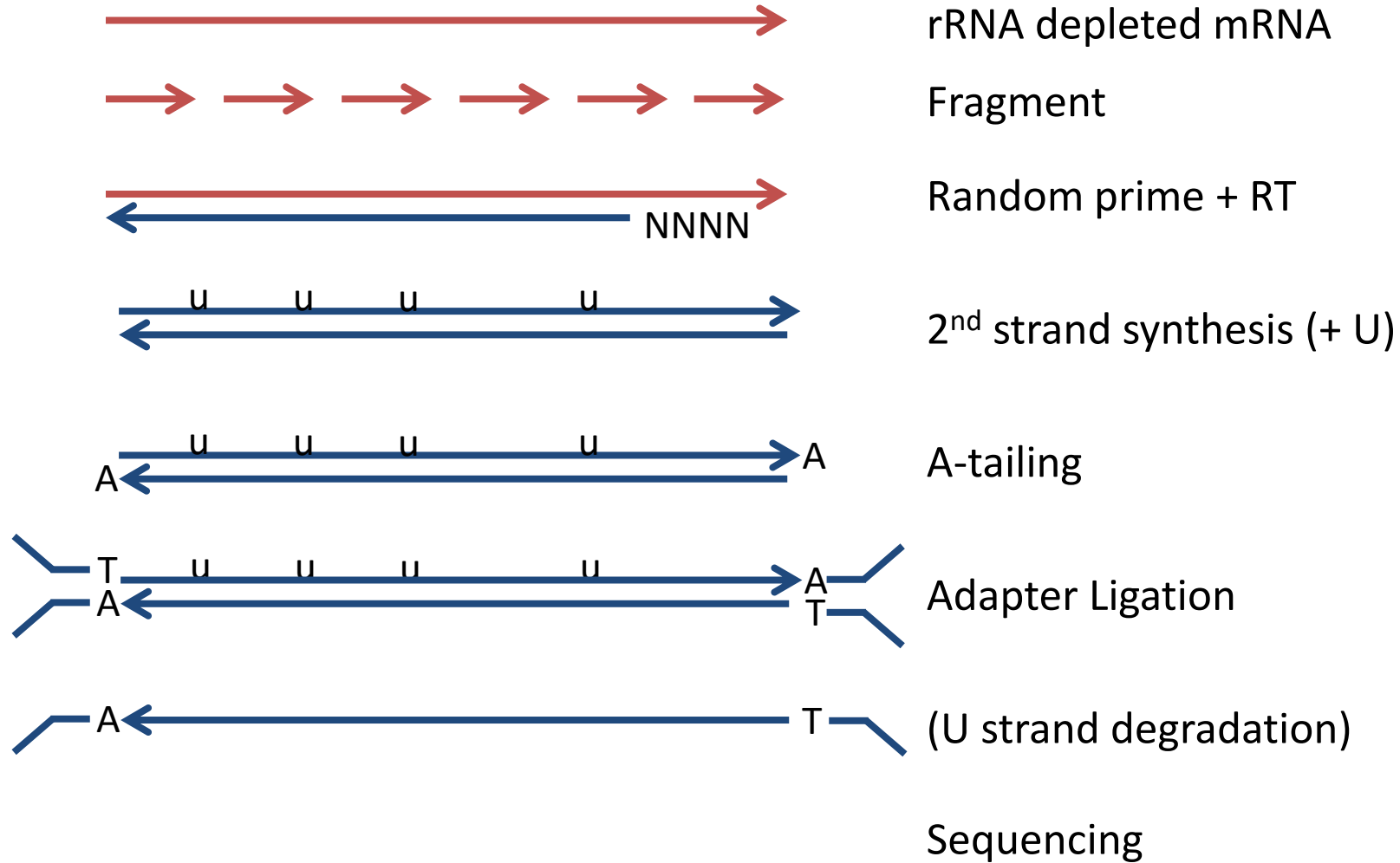


# What type of library?

- Directional libraries if possible
  - Easier to spot contamination
  - No mixed signals from antisense transcription
  - May be difficult for low input samples
- mRNA vs total vs depletion etc.
  - Down to experimental questions
  - Remember LINC RNA may not have polyA tail
  - Active transcription vs standing mRNA pool



# RNAseq Libraries

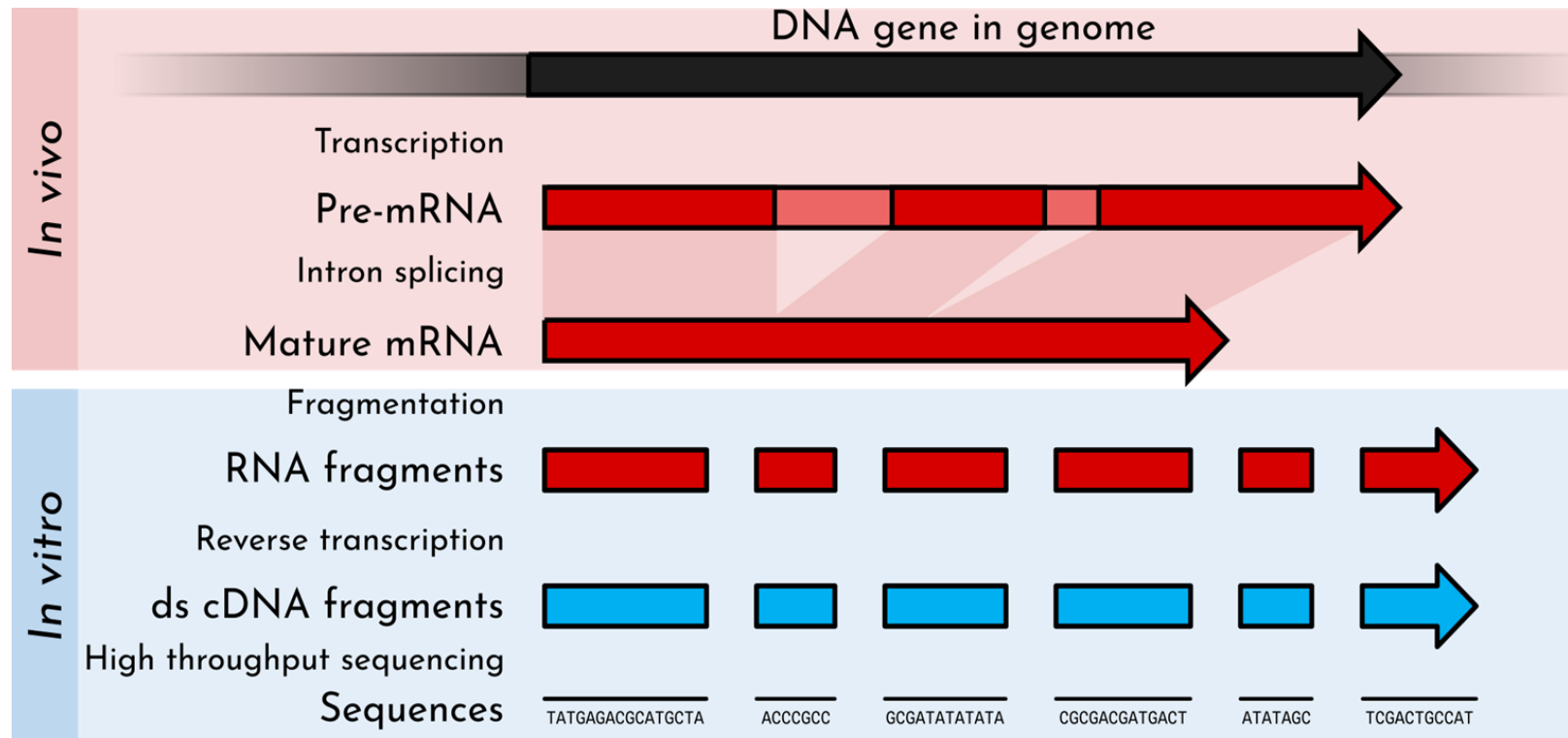


# What type of sequencing

- Depends on your interest
  - Expression quantitation of known genes
    - 50 bp single end is fine
  - Expression plus splice junction usage
    - 100 bp (or longer if possible) single end
  - Novel transcript discovery or per transcript expression
    - > 100 bp paired end

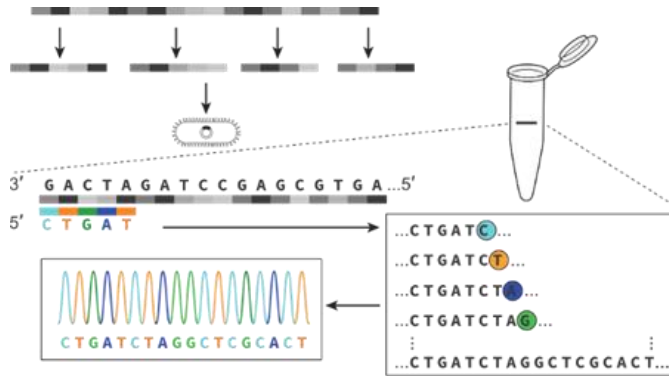
# Available platforms for RNAseq

The basic principle behind RNAseq is the construction of a library, from there, we can choose different platforms to sequence our library



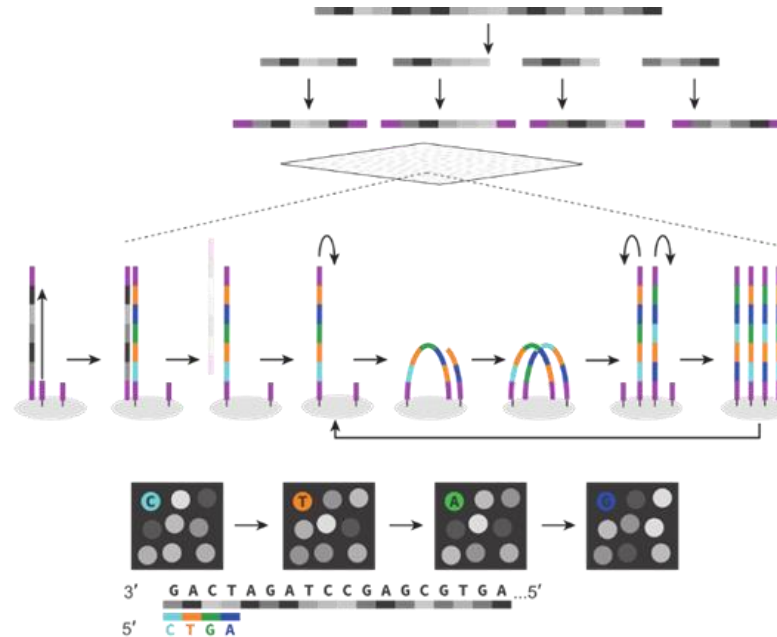
Lowe. 2017. doi: 10.1371/journal.pcbi.1005457

# Available platforms for RNAseq, an overview



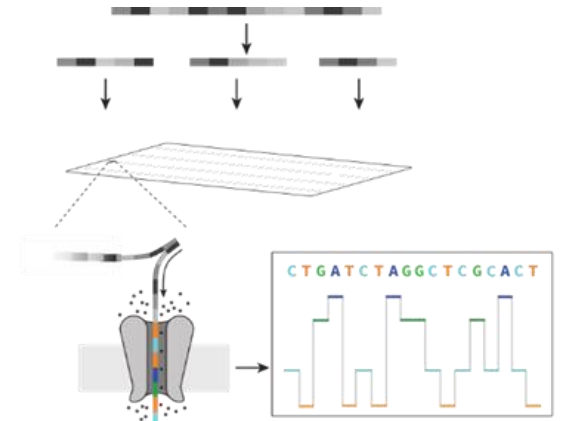
## 1st Generation

- Up to 1000 bp/fragment
- Up to 96 frags/run



## 2nd Generation

- Up to 600 bp/frag (2 x 300)
- Billions of frags/run

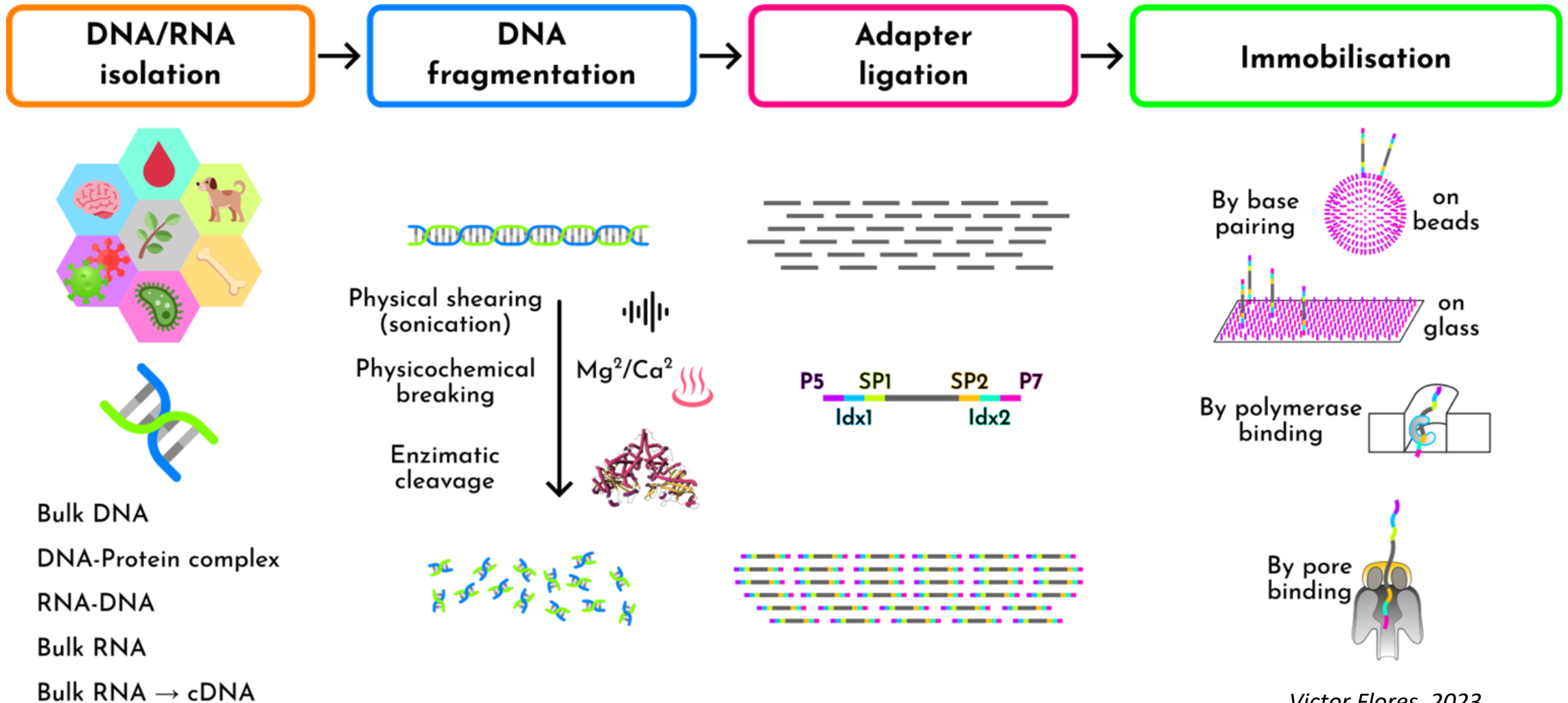


## 3rd Generation

- Up to **4.4 Mbp/frag**
- Millions of frags/run

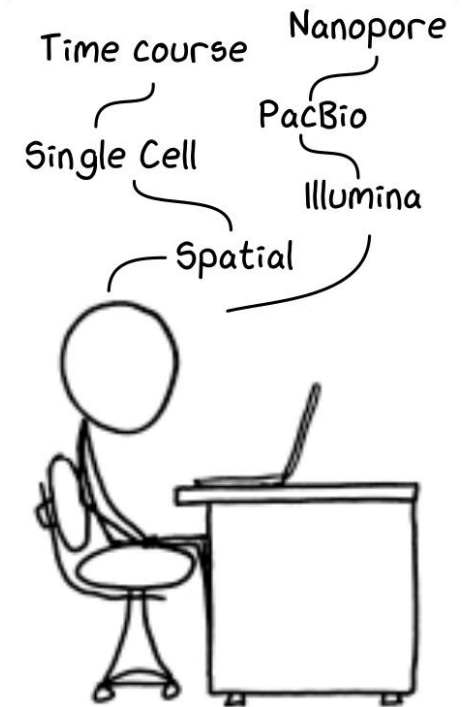
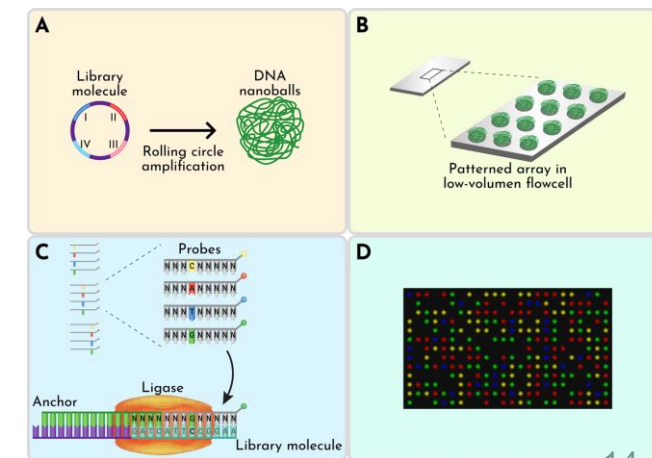
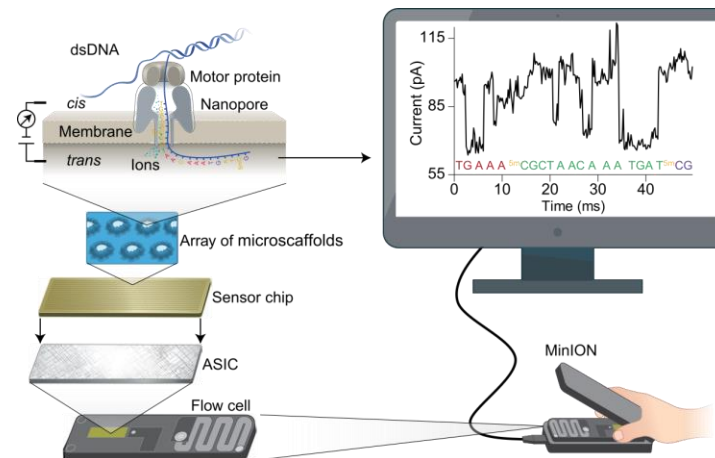
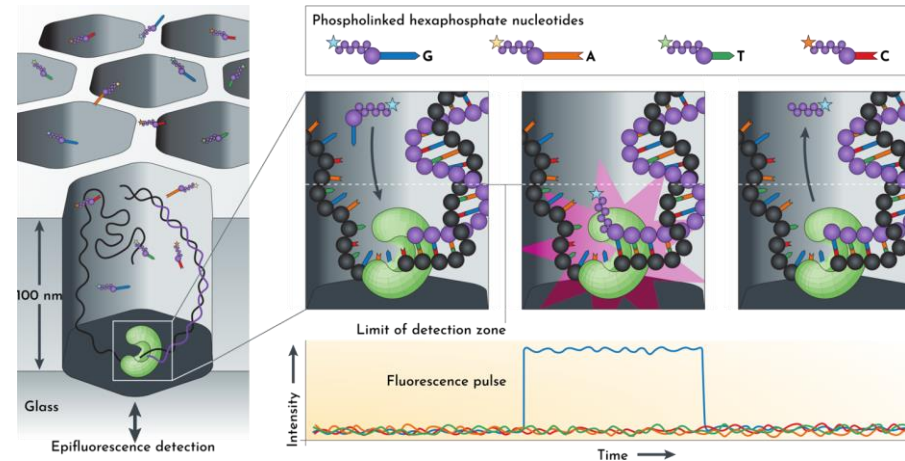
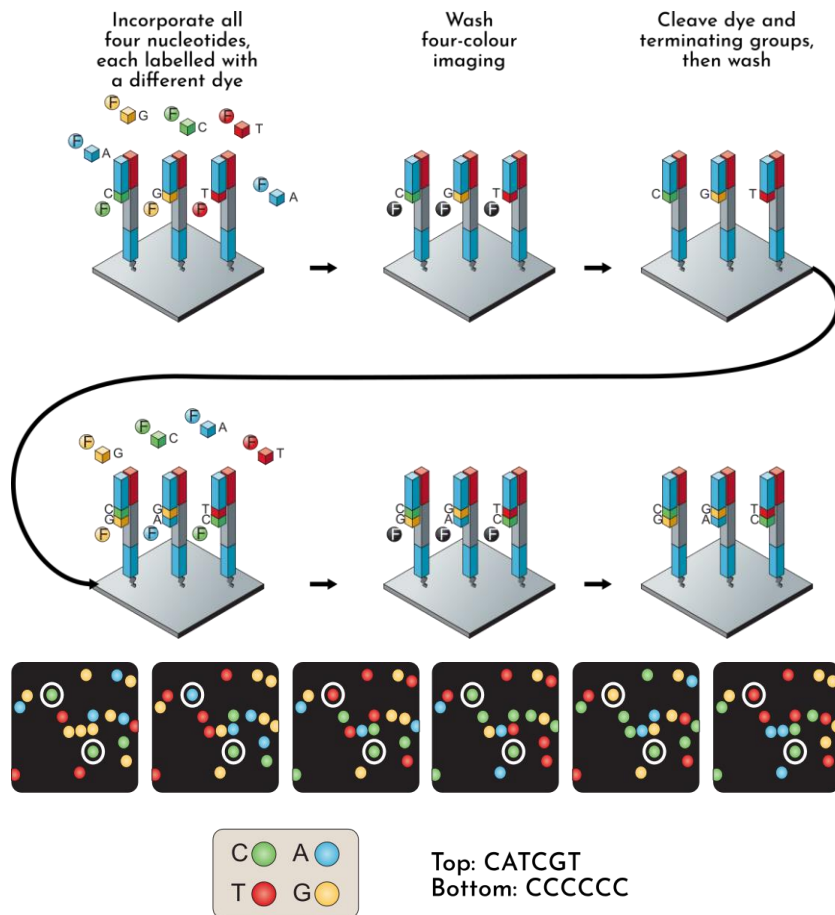
*Adapted from Shendure. 2017. doi: 10.1038/nature24286. Numbers updated from 2021 data.*

# Available platforms for RNAseq: common themes



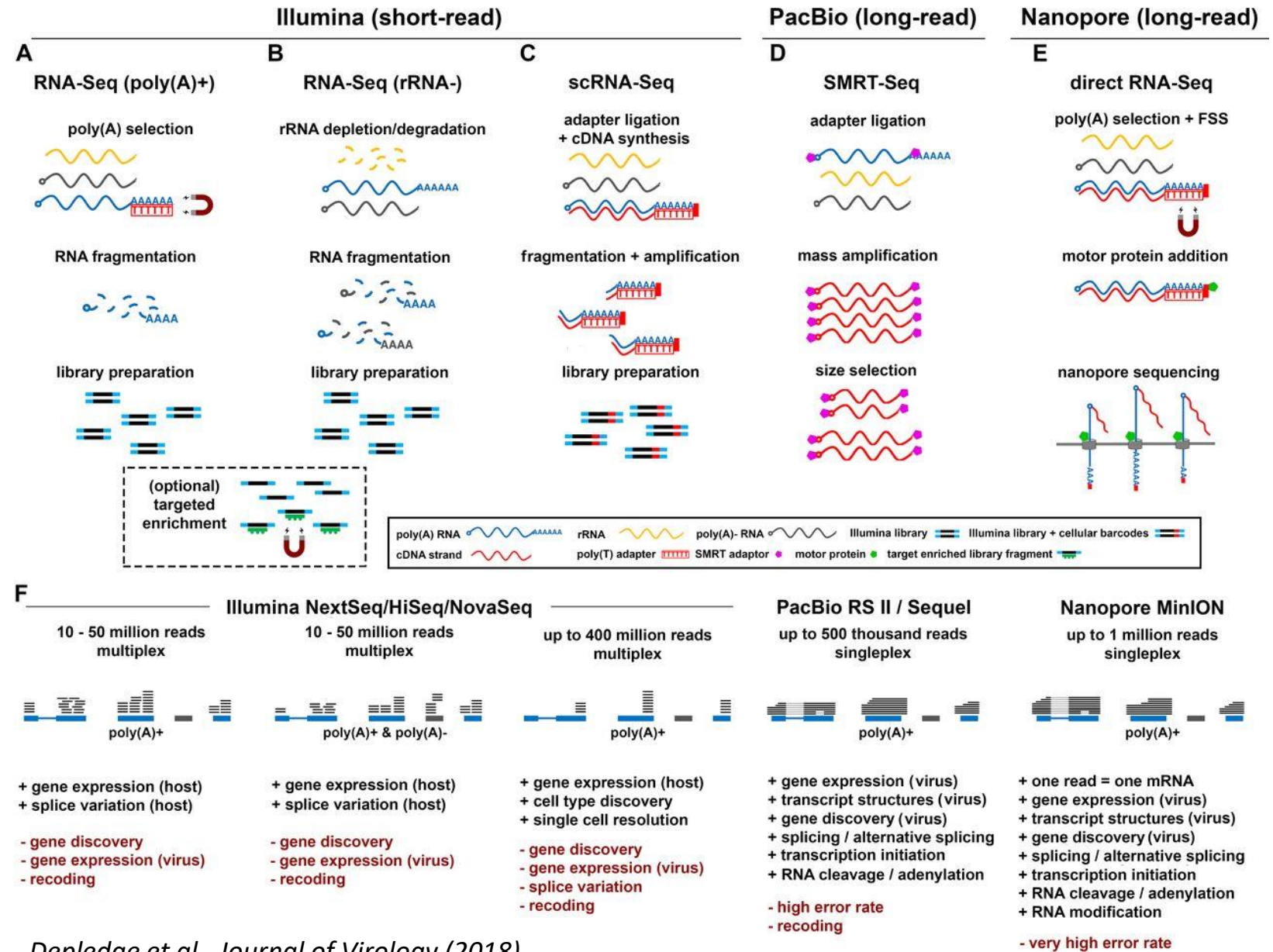
Victor Flores. 2023

# Which platform should I use?



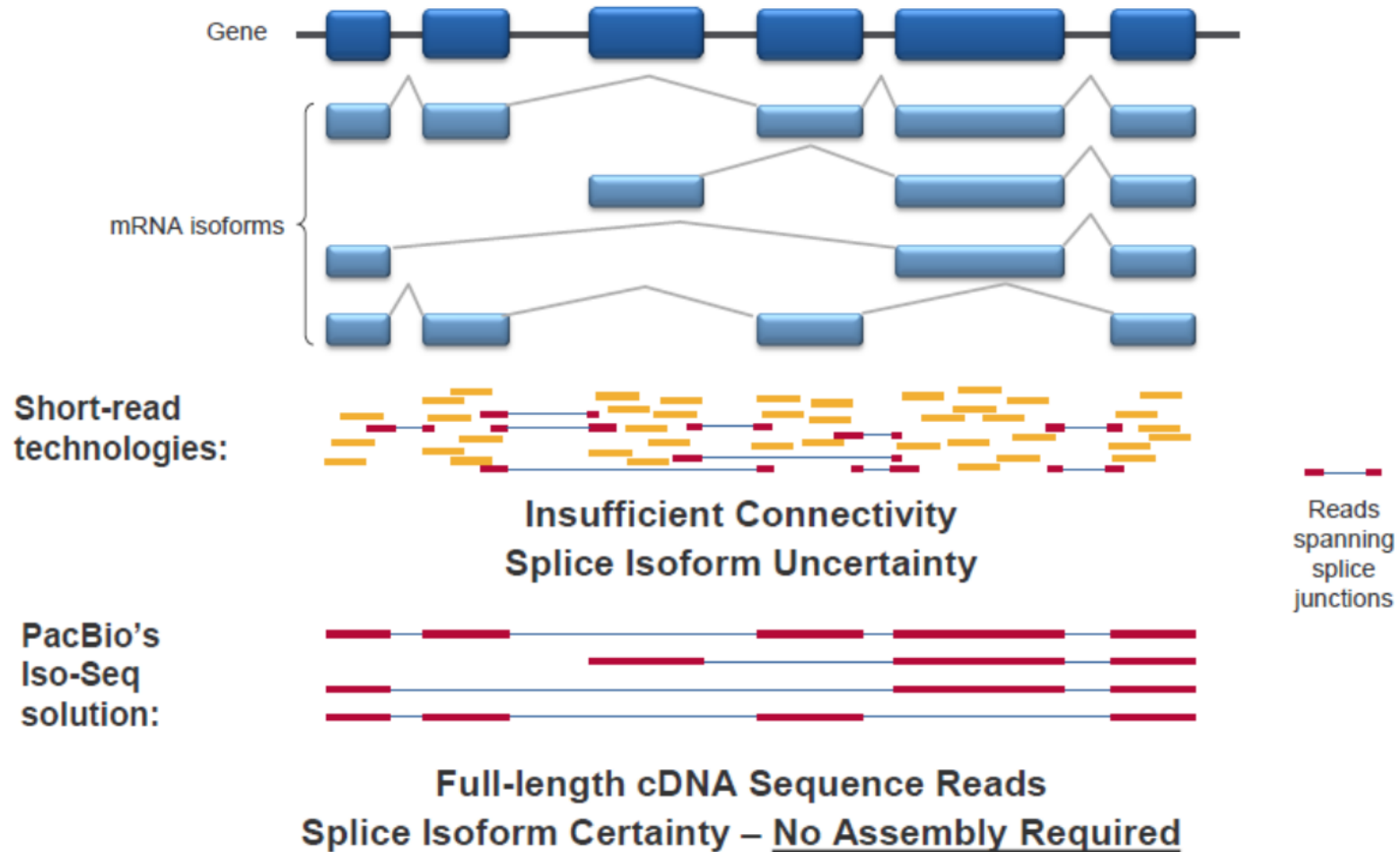


# Major RNA sequencing methodologies



# Iso-Seq by PacBio

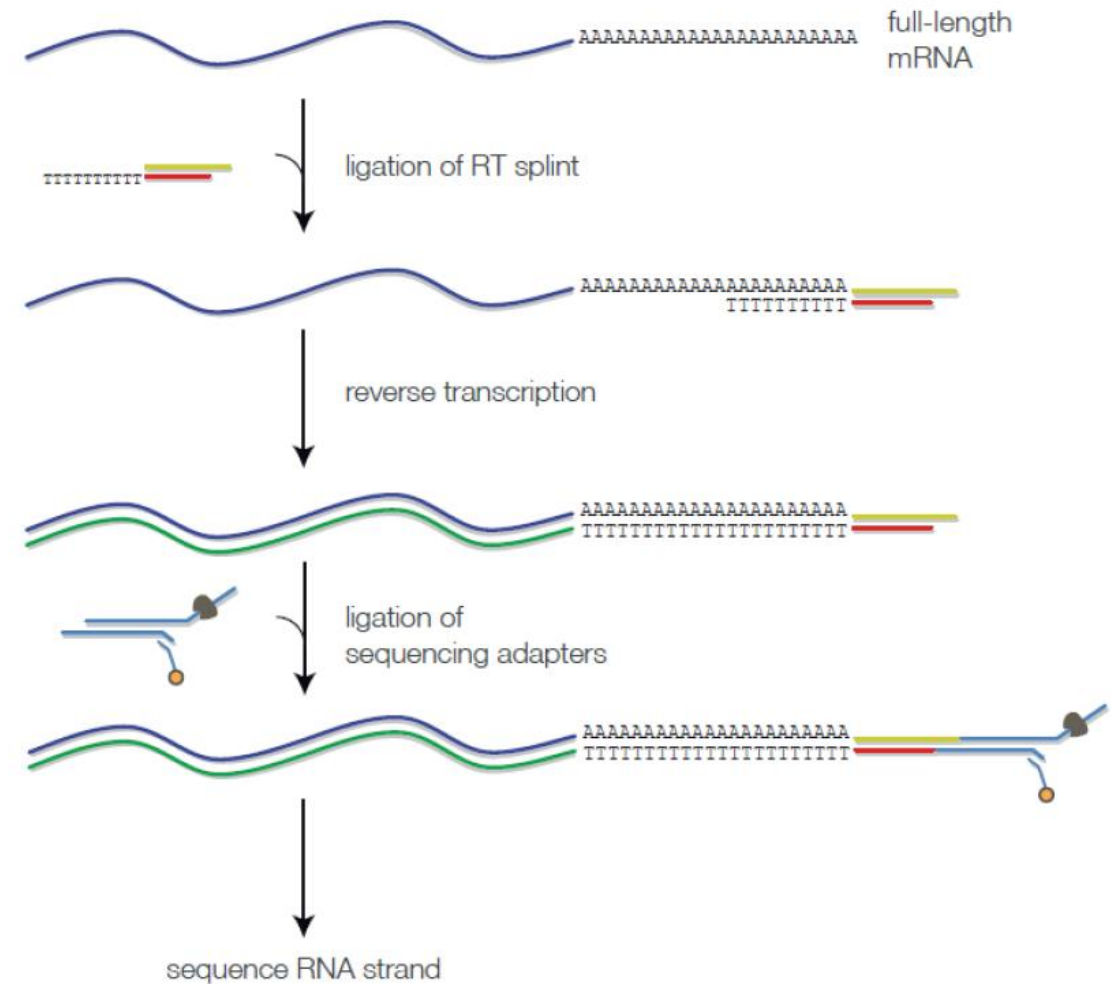
## DETERMINATION OF TRANSCRIPT ISOFORMS





# Direct Nanopore sequencing of RNA and its modifications

- “3-prime down” reading
- RNA-specific motor protein
- RNA directly read in nanopore
- Modified bases distinguishable
- Long read lengths
- cDNA is not sequenced
- Reverse transcription bias (?)
- PCR-free
- More quantitative (?)



*Garalde et al., Nature Methods (2018)*



**We are happy to help you!**