Natural variation in teosinte at the domestication locus *teosinte branched1* (*tb1*)

Laura Vann[1], Thomas Kono[1,2], Tanja Pyhäjärvi[1,3], Matthew B. Hufford[*1,4], and Jeffrey Ross-Ibarra[†1,5]

[1]Department of Plant Sciences, University of California Davis

[2]Department of Agronomy and Plant Genetics, University of Minnesota Twin Cities

[3]Department of Biology, University of Oulu

[4]Department of Ecology, Evolution, and Organismal Biology, Iowa State University

[5]Center for Population Biology and Genome Center, University of California Davis

May 16, 2014

[*]mhufford@iastate.edu

[†]rossibarra@ucdavis.edu

1

1 Abstract

2 The *teosinte branched1* (*tb1*) gene, a repressor of lateral organ growth, is a major

3 QTL involved in branching differences between maize and its wild progenitor,

4 teosinte. Further studies have shown that the insertion of a transposable element

5 (*Hopscotch*) upstream of *tb1* enhances its expression, causing the reduction in

6 branching observed in domesticated maize. Observations of the maize *tb1* allele in

7 teosinte individuals, coupled with estimates of the age of insertion of the *Hopscotch*

8 element, led us to investigate the prevalence and potential role of *tb1* in teosinte.

9 Results from genotyping across many natural populations suggest that the

10 *Hopscotch* element is segregating at a higher than expected frequency in a number

11 of populations of two subspecies of teosinte, *Zea mays* ssp. *parviglumis* and *Zea*

12 *mays* ssp. *mexicana*. Analysis of linkage disequilibrium between the *Hopscotch*

13 element and variation in surrounding regions does not support a hypothesis of

14 recent introgression from maize into teosinte, and we find no evidence of

15 environmental correlations that might suggest recent selection. Finally, two

16 greenhouse experiments with *Zea mays* ssp. *parviglumis* do not suggest an

17 important role for *tb1* in controlling tillering in natural populations of this

18 subspecies. Our findings suggest that the role of the *Hopscotch* in tillering in

19 teosinte is not as straightforward as is in domesticated maize, and that other loci

20 may play a role in observed variation in this trait.

1 Introduction

2 Domesticated crops and their wild progenitors provide an excellent system in which
3 to study adaptation and genomic changes associated with human-mediated
4 selection (Ross-Ibarra et al., 2007). Perhaps the central focus of the study of
5 domestication has been the identification of genetic variation underlying
6 agronomically important traits such as fruit size and plant architecture (Olsen and
7 Gross, 2010). Additionally, many domesticates show reduced genetic diversity when
8 compared to their wild progenitors, and an understanding of the distribution of
9 diversity in the wild and its phenotypic effects has become increasingly useful to
10 crop improvement (Kovach and McCouch, 2008). But while some effort has been
11 invested into understanding how wild alleles behave in their domesticated relatives
12 (Bai and Lindhout, 2007), very little is known about the role that alleles found
13 most commonly in domesticates play in natural populations of their wild
14 progenitors. (Whitton J, 1997).

15   Maize (*Zea mays* ssp. *mays*) was domesticated from the teosinte *Zea mays* ssp.
16 *parviglumis* (hereafter, *parviglumis*) roughly 9,000 B.P. in southwest Mexico
17 (Piperno et al., 2009; Matsuoka et al., 2002). Domesticated maize and the teosintes
18 are an attractive system in which to study domestication due to the abundance of
19 genetic tools developed for maize and well-characterized domestication loci (Hufford
20 et al., 2012a; Doebley, 2004; Hufford et al., 2012b). Additionally, large naturally
21 occurring populations of both *Zea mays* ssp. *parviglumis* (the wild progenitor of
22 maize) and *Zea mays* ssp. *mexicana* (highland teosinte; hereafter *mexicana*) can be
23 found throughout Mexico (Wilkes, 1977; Hufford et al., 2013), and genetic diversity
24 of these taxa is estimated to be high (Ross-Ibarra et al., 2009).

25   Many morphological changes are associated with maize domestication, and

3

understanding the genetic basis of these changes has been a focus of maize research for a number of years (Doebley, 2004). One of the most dramatic changes is found in plant architecture: domesticated maize is characterized by a central stalk with few tillers and lateral branches terminating in a female inflorescence, while teosinte is highly tillered and bears tassels (male inflorescences) at the end of its lateral branches. The *teosinte branched1* (*tb1*) gene, a repressor of organ growth, was identified as a major QTL involved in domestication branching differences achieved through higher expression of the maize allele than the teosinte allele (Doebley et al., 1995, 1997). Further work showed that the insertion of a 4.9 kb retrotransposon (*Hopscotch*) in the upstream control region of *tb1* caused the increased expression of this gene reduction in branching observed in domesticated maize Studer et al. (2011). The effects of this insertion have been observed in tiller number in maize, but little is known about its role, if any, in natural populations of teosinte (Studer et al., 2011). Dating of this element has suggested that its insertion predates the domestication of maize, leading to the hypothesis that it was segregating as standing variation in ancient populations of teosinte and increased to high frequency in maize due to selection during domestication (Studer et al., 2011). Furthermore, Studer and Doebley (2012) investigated the phenotypic effects of nine teosinte *tb1* alleles in an isogenic maize background and found that the introgressions sort into three distinct phenotypic classes, suggesting that variation at the *tb1* locus may play a functional role in teosinte.

In high-density species such as teosinte, plants can detect impending competition from their neighbors through detection of the ratio of red to far-red light. An increase in far-red relative to red light accompanies shading and triggers physiological and morphological changes such as reduced tillering, increased plant height and early flowering collectively known as the shade avoidance syndrome

4

1 (Kebrom and Brutnell, 2007). The *tb1* locus appears to play an important role in

2 the shade avoidance pathway in *Zea mays* and other grasses and may therefore be

3 crucial to the ecology of teosinte (Kebrom and Brutnell, 2007; Lukens and Doebley,

4 1999). In this study we aim to characterize the distribution of the *Hopscotch*

5 insertion in *parviglumis*, *mexicana*, and landrace maize, and to examine the

6 phenotypic effects of the insertion in *parviglumis*. We use a combination of PCR

7 genotyping for the *Hopscotch* element in our full panel and sequencing of two small

8 regions upstream of *tb1* in a subset of teosinte populations to explore patterns of

9 genetic variation at this locus. Finally, we test for an association between the

10 *Hopscotch* element and tillering phenotypes in a population of *parviglumis*.

Methods

Sampling and Genotyping

13 We sampled 1,110 individuals from 350 accessions (247 maize landraces, 17

14 *mexicana* populations, and 86 *parviglumis* populations) and assessed the presence

15 or absence of the *Hopscotch* insertion (Table S1, Table S2). DNA was extracted

16 from leaf tissue using a modified CTAB approach (Doyle and Doyle, 1990; Maloof

17 et al., 1984). We designed primers using PRIMER3 (Rozen and Skaletsky, 2000)

18 implemented in Geneious (Kearse et al., 2012) to amplify the entire *Hopscotch*

19 element, as well as an internal primer allowing us to simultaneously check for

20 possible PCR bias between presence and absence of the *Hopscotch* insertion. Two

21 PCRs were performed for each individual, one with primers flanking the *Hopscotch*

22 (HopF/HopR) and one with a flanking primer and an internal primer

23 (HopF/HopIntR). Primer sequences are HopF,

1 5'-TCGTTGATGCTTTGATGGATGG-3'; Hop R,

2 5'-AACAGTATGATTTCATGGGACCG-3'; and HopIntR,

3 5'-CCTCCACCTCTCATGAGATCC-3' (Figure S1, Figure S2) *Primers in Figure S1 should be*

4 *labeled* . Homozygotes show a single band for absence of the element ($\sim$300bp) and

5 two bands for presence of the element ($\sim$5kb and XX *LV, please add the size of the second*

6 *band*), whereas heterozygotes are three-banded (Figure S2). When only one PCR

7 resolved well, we scored one allele for the individual. We used Phusion High

8 Fidelity Enzyme (Finnzymes, Inc.) and the following conditions for amplifications:

9 98°C for 3 min, 30 cycles of 98°C for 15 s, 65°C for 30 s, and 72°C for 3 min 30 s,

10 with a final extension of 72°C for 10 min. PCR products were visualized on a 1%

11 agarose gel and scored for presence/absence of the *Hopscotch* based on band size.

12 Sequencing

13 In addition to genotyping, we chose a subset of *parviglumis* individuals for

14 sequencing. We chose twelve individuals from each of four populations from Jalisco

15 state, Mexico (San Lorenzo, La Mesa, Ejutla A, and Ejutla B). For amplification

16 and sequencing, we selected two regions approximately 600bp in size from within

17 the 5' UTR of *tb1* (sequenced region 1) and from 1,235 bp upstream of the start of

18 the *Hopscotch* and 66,169 bp upstream from the start of the *tb1* ORF (sequenced

19 region 2). We designed the following primers using PRIMER3 (Rozen and

20 Skaletsky, 2000): for the 5' UTR, 5' GGATAATGTGCACCAGGTGT 3' and 5'

21 GCGTGCTAGAGACACYTGTTGCT 3'; for the 50 kb upstream region, 5'

22 TGTCCTCGCCGCAACTC 3' and 5' TGTACGCCCGCCCCTCATCA 3' (). We

23 used Taq Polymerase (New England Biolabs) and the following thermal cycler

24 conditions to amplify fragments: 94°C for 3 min, 30 cycles of 92°C for 40 s,

6

annealing for 1 min, 72°C for 40 s, and a final 10 min extension at 72°C. Annealing

temperatures for sequenced region 1 and sequenced region 2 were 59.7°C and

58.8°C, respectively. To clean excess primer and dNTPs we added two units of

Exonuclease1 and 2.5 units of Antarctic Phosphatase to 8.0 μ L of amplification

product. This mix was placed on a thermal cycler with the following program:

37°C for 30 min, 80°C for 15 min, and a final cool-down step to 4°C.

We cloned cleaned fragments into a TOPO-TA vector (Invitrogen, Carlsbad)

using OneShot TOP10 chemically competent *E. coli* cells, with an extended

ligation time of 30 min for a complex target fragment. We plated cells on LB agar

plates containing kanamycin, and screened colonies using vector primers M13

Forward and M13 Reverse under the following conditions: 96°C for 5 min; then 35

cycles at 96°C for 30 s, 53°C for 30 s, 72°C for two min; and a final extension at

72°C for 4 min. We visualized amplification products for incorporation of our insert

on a 1% agarose TAE gel.

Amplification products with successful incorporation of our insert were cleaned

using Exonuclease 1 and Antarctic Phosphatase following the procedures detailed

above, and sequenced with vector primers M13 Forward and M13 Reverse using

Sanger sequencing at the College of Agriculture and Environmental Sciences

(CAES) sequencing center at UC Davis. We aligned and trimmed primer sequences

from resulting sequences using the software Geneious (Kearse et al., 2012).

Following alignment, we verified singleton SNPs by sequencing an additional one to

four colonies from each clone. If the singleton was not present in these additional

sequences it was considered an amplification or cloning error, and we replaced the

base with the base of the additional sequences. If the singleton appeared in at least

one of the additional sequences we considered it a real variant and kept it for

further analyses.

1 Genotyping Analysis

We examined discrepancies between observed and expected genotype frequencies by calculating Hardy-Weinberg Equilibrium (HWE). To calculate differentiation between populations (F_{ST}) and subspecies (F_{CT}) we used HierFstat (Goudet, 2005). These analyses only included populations in which 8 or more individuals were sampled. To test the hypothesis that the *Hopscotch* insertion may be adaptive under certain environmental conditions, we looked for significant associations between the *Hopscotch* frequency and environmental variables using BayEnv (Coop et al., 2010). BayEnv creates a covariance matrix of relatedness between populations, and then tests a null model that allele frequencies in populations are determined by the covariance matrix of relatedness alone against the alternative model that allele frequencies are determined by a combination of the covariance matrix and an environmental variable, producing a posterior probability (Bayes Factor)(Coop et al., 2010). We used genotyping and covariance data from Pyhäjärvi et al. (2013) for BayEnv, with the *Hopscotch* insertion coded as an additional SNP (). Environmental data were obtained from `www.worldclim.org`, the Harmonized World Soil Database and `www.harvestchoice.org`, and summarized by principle component analysis (Pyhäjärvi et al., 2013).

8

1 Sequence Analysis

2 For population genetic analyses of sequenced region 1 and sequenced region 2 we

3 used the analysis package of Libsequence (Thornton, 2003) to calculate pairwise

4 F_{ST} between populations, and to calculate standard diversity statistics (number of

5 haplotypes; haplotype diversity; Watterson's estimator $\hat{\theta}_W$; pairwise nucleotide

6 diversity $\hat{\theta}_\pi$; and Tajima's D). To produce a visual representation of differentiation

7 between sequences and to examine patterns in sequence clustering by *Hopscotch*

8 genotype we used Phylip

9 (`http://evolution.genetics.washington.edu/phylip.html`) to create

10 neighbor-joining trees with bootstrapping (100 repetitions) to examine the support

11 of nodes in our trees. For creation of trees we also included homologous sequence

12 data from teosinte inbred lines (TILs), some of which are known to be homozygous

13 for the *Hopscotch* insertion (TIL03, TIL17, TIL09), as well as 59 lines of

14 domesticated maize and landraces (data from Maize HapMapV2, (Chia et al.,

15 2012)).

16 Introgression Analysis

17 In order to assess patterns of linkage disequilibrium (LD) around the *Hopscotch*

18 element in the context of chromosomal patterns of LD we used Tassel (Bradbury

19 et al., 2007) and calculated LD between SNPs across chromosome 1 using

20 previously published data from twelve plants each of the Ejutla A (EjuA), Ejutla B

21 (EjuB), San Lorenzo (SLO), and La Mesa (MSA) populations (Pyhäjärvi et al.,

22 2013). We chose these populations because we had both genotyping data for the

23 *Hopscotch* as well as chromosome-wide SNP data for chromosome 1. For each

24 population we filtered the initial set of 5,897 SNPs on chromosome 1 to accept only

SNPs with a minor allele frequency of at least 0.1, resulting in 1,671, 3,023, 3,122, and 2,167 SNPs for SLO, EjuB, EjuA, and MSA, respectively. We then used Tassel (Bradbury et al., 2007) to calculate linkage disequilibrium (r^2) across chromosome 1 for each population.

We examined evidence of introgression on chromosome 1 in these same four populations (EjuA, EjuB, MSA, SLO) using STRUCTURE (Falush et al., 2003) and the same phased 55K SNP data from (Pyhäjärvi et al., 2013) that we used for LD analysis, combined with the corresponding SNP data from a diverse panel of 282 maize lines (Cook et al., 2012). SNPs were anchored in a modified version of the IBM genetic map ((Gerke et al., 2013), http://arxiv.org/abs/1307.7313). We created haplotype blocks using a custom Perl script that grouped SNPs separated by less than 5kb into haplotypes. We ran STRUCTURE at K=2 under the linkage model, performing 3 replicates with an MCMC burn-in of 10,000 steps and 50,000 steps post burn-in. *i'd like this perl script on github, maybe in this repo or as a gist. also structure input file too. all the stuff we'd need to redo this.* *See above note..not all of this or the BayEnv was script'ified* *sure, even command line info should be included where possible. idea is to maximize reproducibility – either other people or subsequent students. for example, matt has a student who wants to work on tb1 in natural pops, and she might want to try/redo some of these analyses with the same or new data.* *okay so I should just put my command line stuff with good commenting as to what is what in repository?*

Phenotyping of *Zea mays.* ssp. *parviglumis*

To investigate the phenotypic effects of the *Hopscotch* insertion in teosinte, we conducted an initial phenotyping trial (Phenotyping 1). We germinated 250 seeds of *parviglumis* collected in Jalisco state, Mexico (population San Lorenzo) (Hufford, 2010) where the *Hopscotch* is segregating at highest frequency (0.44) in our initial

genotyping sample set. In order to maximize the likelihood of finding the *Hopscotch* in our association population we selected seeds from sites where genotyped individuals were homozygous or heterozygous for the insertion. We chose between 10-13 seeds from each of 23 sampling sites. We treated seeds with fungicide and germinated them in petri dishes with filter paper. Following germination, 206 successful germinations were then planted into one gallon size pots with potting soil and randomly spaced one foot apart on greenhouse benches. Plants were watered three times a day with an automatic drip containing 10-20-10 fertilizer. *it ended up being a combination of drip and hand watering because they dried out so much and did better when they had water on the leaves as well as in the soil*

To investigate the phenotypic effects of the *Hopscotch* insertion in teosinte, we conducted an initial phenotyping trial (Phenotyping 1). We germinated 250 seeds of *parviglumis* collected in Jalisco state, Mexico (population San Lorenzo) (Hufford, 2010) where the *Hopscotch* is segregating at highest frequency (0.44) in our initial genotyping sample set. In order to maximize the likelihood of finding the *Hopscotch* in our association population we selected seeds from sites where genotyped individuals were homozygous or heterozygous for the insertion. We chose between 10-13 seeds from each of 23 sampling sites. We treated seeds with fungicide and germinated them in petri dishes with filter paper. Successful germinations (206 individuals) were then planted into one gallon size pots with potting soil and randomly spaced one foot apart on greenhouse benches. Plants were watered three times a day.

Starting on day 15, we measured tillering index, the ratio of the sum of tiller lengths to the height of the plant (Briggs et al., 2007). Tillering index has been shown to be the most effective way to observe the phenotypic effects of the *Hopscotch* insertion on plant architecture in maize (Clark et al., 2006). Following

1 initial measurements, we phenotyped plants for tillering index every 5 days through
2 day 40, and then on day 50 and day 60. On day 65 we measured culm diameter
3 between the third and fourth nodes of each plant. Culm diameter is not believed to
4 be correlated with tillering index, or variation at *tb1* (e.g. *Hopscotch* genotype).
5 Following phenotyping we extracted DNA from all plants using a modified SDS
6 extraction protocol (http://www.ars.usda.gov). We genotyped individuals for the
7 *Hopscotch* insertion following the protocols listed above. Based on these initial
8 data, we conducted a post hoc power analysis using data from day 40 of
9 phenotyping 1, indicating that a minimum of 71 individuals in each genotype class
10 are needed to detect the observed effect of the *Hopscotch* on tillering index. *do you*
11 *still have these posthoc calculations?   I believe I do, otherwise they are likely in a lab meeting slide on Dropbox –*
12 *do you want them in here?   would be good to include in a github, again so we could go back and reassess how we*
13 *do things in case, for example, we decide to give the greenhouse experiment a 3rd try (yes, i'm a masochist)   I*
14 *would do it a 3rd time, I'm convinced something went wrong....third time with expression analyses :)*
15   We performed a second phenotyping experiment (phenotyping 2) in which we
16 germinated 372 seeds of *parviglumis*, choosing equally between sites previously
17 determined to have or not have the *Hopscotch* insertion. Seeds were germinated
18 and planted on day 7 post fruit-case removal into 2 gallon pots. Plants were
19 watered twice daily, alternating between fertilized and non-fertilized water. We
20 began phenotyping successful germinations (302) for tillering index on day 15 post
21 fruit case removal, and phenotyped every five days until day 50. At day 50 we
22 measured culm diameter between the third and fourth nodes. We extracted DNA
23 and genotyped plants following the same guidelines as in phenotyping 1.
24   Resulting tillering index data for each genotype class did not meet the criteria
25 for a repeated measures ANOVA, so we transformed the data using a Box-Cox
26 transformation ($\alpha = 0$) implemented in the car package in R (Fox and Weisberg,

12

1 2011) to improve the normality and homogeneity of variance among genotype

2 classes. We analyzed relationships between genotype and tillering index and tiller

3 number using a repeated measures ANOVA through a general linear model

4 function implemented in SAS v.9.3 (SAS Institute Inc., Cary, NC, USA).

5 Additionally, in order to compare any association between *Hopscotch* genotype and

6 tillering and associations at other presumably unrelated traits, we performed an

7 ANOVA between culm diameter and genotype using the same general linear model

8 in SAS. *please add SAS scripts/code to a gist or something.*

9 Results

10 Genotyping

11 Genotype of the *Hopscotch* insertion was confirmed with two PCRs for 837

12 individuals. Among the 247 maize landrace accessions genotyped, all but 8 were

13 homozygous for the presence of the insertion (**??**, **??**). *please fix table/figure references to say*

14 *table/figure* Within our *parviglumis* and *mexicana* samples we found the *Hopscotch*

15 insertion segregating in 37 and 4 populations *this is confusing as the map shows ¡37 populations.*

16 *we should be consistent about reporting for all pops, or all pops with n¿8, etc. or at least be explicit what cutoff we*

17 *are using for each result* , respectively, and at highest frequency in the states of Jalisco,

18 Colima, and Michoacán in central-western Mexico in both subspecies (1) *the map only*

19 *shows parviglumis. can we add mexicana? the text makes it seem as if both should be on the map* . We

20 examined Hardy-Weinberg equilibrium in a total of 14 populations (10 *parviglumis*

21 and 4 *mexicana*) with more than 8 individuals sampled per population. Three

22 populations (RIMPA0073, RIMPA0093, and RIMPA0158) show evidence of

23 deviations from expected genotype frequencies under the assumptions of HWE
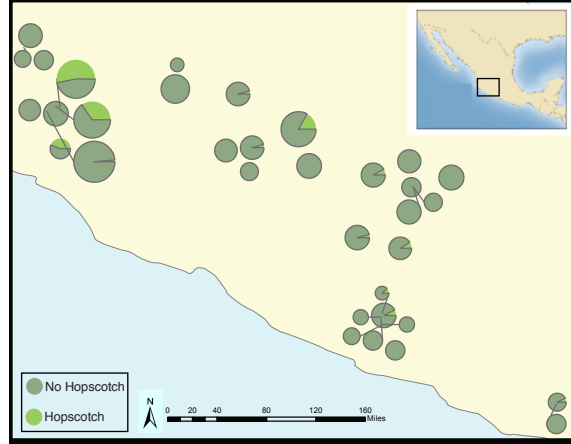
13

Figure 1: Map showing the frequency of the *Hopscotch* allele in populations of *Zea mays* ssp. *parviglumis* where we sampled more than 6 individuals. Size of circles reflects number of alleles sampled.

1 ($p<0.05$). *in what direction? too many hets? what's the F?   they weren't all in the same direction, do you still*

2 *want me to list out?   no need to list all, but if there were sig. deviations in multiple directions in different pops,*

3 *that is worth saying. maybe could list them all out in supp. table?*

4 *please fix whitespace and black border on figure*

5   Using our *Hopscotch* genotyping data, we calculated differentiation between

6 populations (F_{ST}) and subspecies (F_{CT}) for populations in which we sampled 8 or

7 more alleles. F_{CT} is 0 within our dataset, and we found similar levels of F_{ST} among

8 populations within each subspecies (0.22) *is this an average? this comes from libsequence?*   and

9 among all populations (0.23), to those reported in genome-wide estimates from

10 previous studies Pyhäjärvi et al. (2013) (1).

11   Although we found large variation in *Hopscotch* allele frequency among our

12 populations, BayEnv analysis did not indicate a correlation between the *Hopscotch*

13 insertion and environmental variables (all Bayes Factors < 1;).

14

Table 1: Pairwise F_{CT} values from sequence and *Hopscotch* genotyping data

Comparison	Seq. Region 2	Seq. Region 1	*Hopscotch*
EjuA & EJuB	0	0	0
EjuA & MSA	0.328	0.326	0.186
EjuA & SLO	0.258	0.416	0.28
EjuB & MSA	0.365	0.397	0.188
EjuB & SLO	0.29	0.512	0.28
MSA & SLO	0	0.007	0.016

1 Sequencing

To investigate patterns of sequence diversity and linkage disequilibrium (LD) in the *tb1* region, we sequenced two small (<1kb) regions upstream of the tb1 ORF in four populations. After alignment and singleton checking we recovered 40 and 48 segregating sites for the 50kb upstream region and the 5' UTR region, respectively. For region 1, Ejutla A has the highest values of haplotype diversity, and θ_π, while Ejutla B and La Mesa have comparable values of these summary statistics, and San Lorenzo has much lower values. Additionally, Tajima's D is strongly negative in the two Ejutla populations and La Mesa, but is more positive in San Lorenzo (2). *can drop theta W from table and text. we show pi and D, which is sufficient. fix caption in table. also add* Hopscotch *allele frequencies to table or list somewhere – i think it's useful for comparison of seq stats* *the frequencies are in supplemental table 1, do you want them put in this table too, or just me to refer people to supp table 1?* *yeah refer to table is fine*

For region 2, haplotype diversity, θ_W, and θ_π, are similar for Ejutla A and Ejutla B, while La Mesa and San Lorenzo have slightly lower values for these statistics (2). Tajima's D is positive in all populations except San Lorenzo,

15

Table 2: Add caption

Population	# Haplotypes	Hap. Diversity	$\hat{\theta}_W$	$\hat{\theta}_\pi$	Tajima's D
Seq. region 2 (50kb upstream)					
EJUA	8	0.89394	0.01548	0.01763	0.6231
EJUB	8	0.89394	0.01493	0.01591	0.29504
MSA	3	0.68182	0.01111	0.01055	-0.22212
SLO	4	0.74242	0.01167	0.01413	0.93185
Seq. region 1(5? UTR)					
EJUA	8	0.85897	0.00874	0.00527	-1.64955
EJUB	5	0.70909	0.00663	0.00378	-1.83123
MSA	6	0.68182	0.00646	0.00373	-1.75506
SLO	3	0.31818	0.00176	0.00137	-0.72873

1 indicating an excess of low frequency variants in this population (2). Pairwise

2 values of F_{ST} within population pairs Ejutla A/Ejutla B and San Lorenzo/La Mesa

3 are 0 for both sequenced regions as well as for the *Hopscotch*, while they are high

4 for other population pairs (1). Neighbor joining trees of our sequence data and

5 data from the teosinte inbred lines (TILs; data from Maize HapMapV2, (Chia

6 et al., 2012)) do not reveal any clear clustering pattern with respect to population

7 or *Hopscotch* genotype (??); individuals within our sample that have the *Hopscotch*

8 insertion do not group with the teosinte inbred lines or the lines of domesticated

9 maize that have the *Hopscotch* insertion.

1 Evidence of introgression

2 The teosinte populations *which?* with the highest frequency of the *Hopscotch*

3 insertion in this study were sympatric with cultivated maize. Our initial hypothesis

4 was that the high frequency of the *Hopscotch* element in these populations could be

5 attributed to introgression from maize into teosinte. To investigate this possibility

6 we examined overall patterns of linkage disequilibrium across chromosome one, and

7 specifically in the *tb1* region. If the *Hopscotch* is found in these populations due to

8 recent introgression we would expect to find large blocks of linked markers near this

9 element. We find no evidence of elevated linkage disequilibrium between the

10 *Hopscotch* and SNPs surrounding the *tb1* region in our resequenced populations

11 (2), and r^2 in the *tb1* region does not differ significantly between populations with

12 (average r^2 of 0.085) and without the *Hopscotch* genotype (average $r^2 = 0.082$). In

13 fact, average r^2 is lower in the *tb1* region ($r^2 = 0.056$) than across the rest of

14 chromosome 1 ($r^2 = 0.083$) (3). *table is too wide, need to round numbers, and column headers are*

15 *messed up.*

16 The lack of clustering of *Hopscotch* genotypes in our NJ tree as well as the lack

17 of LD around *tb1* does not support the hypothesis that the *Hopscotch* insertion in

18 these populations of *parviglumis* is the result of recent introgression. However, to

19 further explore this hypothesis we performed a STRUCTURE analysis using

20 Illumina MaizeSNP50 data from four of our *parviglumis* populations (EjuA, EjuB,

21 MSA, and SLO) and the maize 282 diversity panel (Cook et al., 2012; Pyhäjärvi

22 et al., 2013). The linkage model implemented in STRUCTURE can be used to

23 identify ancestry of blocks of linked variants, which would arise as a result of recent

24 admixture between populations. If the *Hopscotch* insertion is present in

25 populations of *parviglumis* as a result of recent admixture with domesticated maize,

Table 3: r^2 values between SNPs in the *tb1* region (positions 264,596,664-265,891,456 on chromosome 1 of the maize AGPv2 genome) and the rest of chromosome 1, within the 5' UTR (Sequenced region 1), and within the 66,169 bp upstream region (Sequenced region 2).

Population	Chromosome 1	tb1 region	Seq. region 1	Seq. region 1
Ejutla A	0.095426101	0.050304	0.747295	0.214933
Ejutla B	0.068681837	0.051295	0.660354	0.186395
La Mesa	0.069500533	0.053306	0.914286	0.766234
San Lorenzo	0.100536784	0.067251	0.912281	0.636364

1 we would expect the insertion and linked variants in surrounding sites to be

2 assigned to the "maize" cluster in our STRUCTURE runs, not the "teosinte"

3 cluster. In all runs, assignment to maize in the *tb1* region across all four

4 *parviglumis* populations is low (average 0.017) *is this really 0.017 or 0.17?* *Yes really 0.017*

5 *assignment to maize in the tb1 region, and avg assignment across chr1 is 0.2* *I also have a table of assignment*

6 *values for SLO individuals based on genotype. Though we had decided this wasn't super informative because sample*

7 *size was low* and much below the chromosome-wide average (0.20; 3).

8 *please put figures in the text rather than at the end.* *I can't figure out why the figures are going at the end.*

9 *I did them following the format in Sofiane's 282 paper*

10 Phenotyping

11 Phenotyping

12 To assess the contribution of *tb1* to phenotypic variation in tillering in a natural

13 population, we grew plants from seed sampled from the San Lorenzo population of
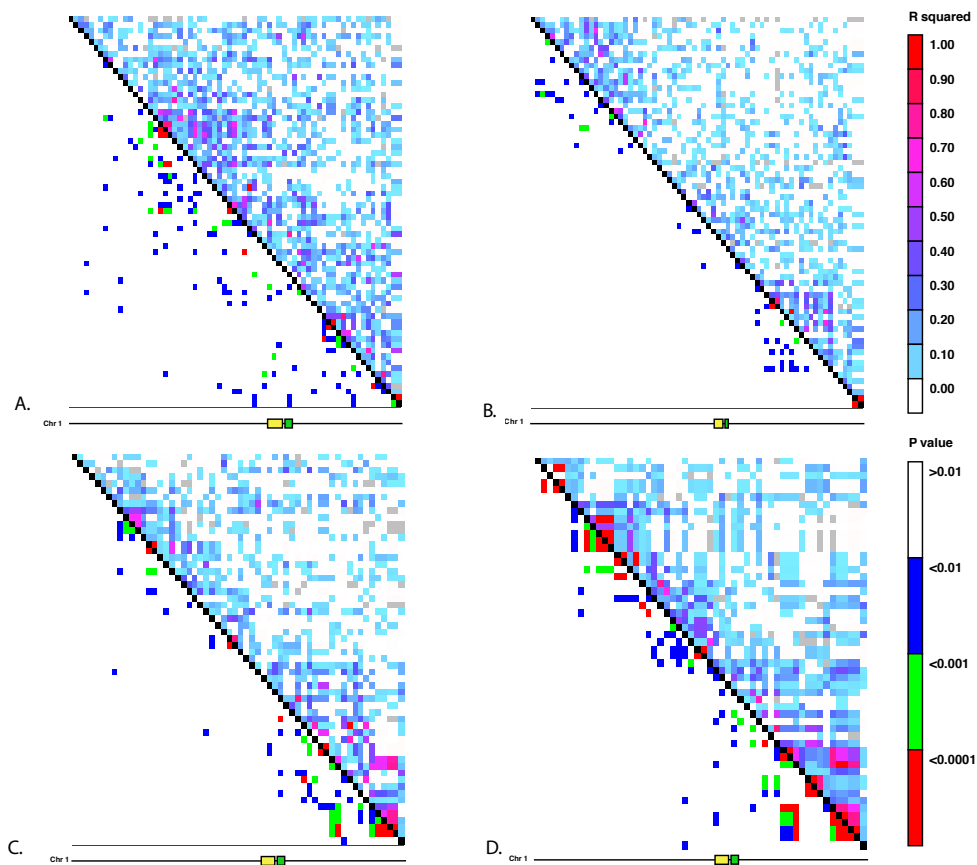
Figure 2: Linkage disequilibrium for SNPs in Mb 261-268 on chromosome 1. The yellow rectangle indicates the location of the *Hopscotch* insertion and the green represents the *tb1* ORF. A) Ejutla A; B) Ejutla B; C) La Mesa; D). San Lorenzo

1 *parviglumis*, which had a high mean frequency (0.44) of the *Hopscotch* insertion

2 from our initial genotyping. We measured tillering index (TI), the ratio of the sum

3 of tiller lengths to plant height, for 216 plants from within the San Lorenzo

4 population, and genotyped plants for the *Hopscotch* insertion. We found the

5 *Hopscotch* segregating at a frequency of 0.65 with no significant deviations from
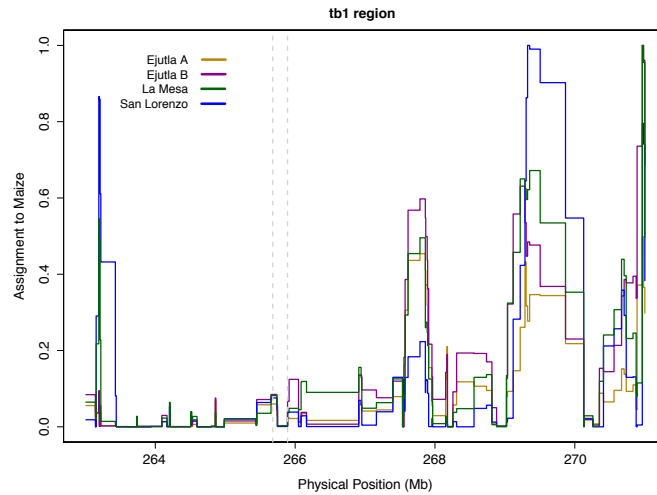
19

Figure 3: STRUCTURE assignment to maize across a section of chromosome 1. The dotted lines mark the beginning of the sequenced region 50kb upstream (Sequenced region 2) and the end of the *tb1* ORF.

1 expected frequencies under Hardy-Weinberg equilibrium. After performing a

2 repeated measures ANOVA between our transformed tillering index data and

3 *Hopscotch* genotype we find a weak positive correlation between presence of the

4 *Hopscotch* and tillering index on day 40 (p=0.0848), but no correlation between

5 tillering index and genotype on any other day (4). Additionally we find no

6 significant correlation between tiller number and *Hopscotch* genotype, or culm

7 diameter and *Hopscotch* genotype in phenotyping 1. *shouldn't we expect a negative correlation*

8 *betweeh Hop and TI on day 40? need to have an A and B in the figure and explain one is for pheno1 and one is*

9 *for pheno2. please explain whiskers and dots on figure too.   sure, I mean, presumably we would expect things with*

10 *Hop to have a smaller TI   yup, but we should mention that the expectation is negative/*

11   *lots of white space in fig 4 and fig. s1 too.*   We performed a second grow-out of teosinte to

20

1 assess whether lighting conditions or sample size may have affected our ability to

2 detect and effect of *tb1*. For the second grow-out we measured tillering index every

3 five days through day 50 for 302 plants. We found the *Hopscotch* allele segregating

4 at a frequency of 0.69, *is it in HWE in this pop?* with a 0.6 frequency of *Hopscotch*

5 homozygotes, and a 0.2 frequency of both heterozygotes and homozygotes for the

6 teosinte allele. We found similar patterns, with a weak positive correlation between

7 tillering index and *Hopscotch* genotype at day 40 (p¡0.0611), with no significant

8 correlation on any day. Similarly, relationships between *Hopscotch* genotype and

9 tiller number, and *Hopscotch* genotype and culm diameter are not significant.

10 Discussion

11 Adaptation occurs either due to selection on standing variation or on *de novo*

12 mutations. Adaptation as a result of selection on standing variation has been

13 well-described in a number of systems, for example, selection for lactose tolerance

14 in humans (Plantinga et al., 2012; Tishkoff et al., 2007); variation at the *Eda* locus

15 in three-spined stickleback (Kitano et al., 2008; Colosimo et al., 2005); and pupal

16 diapause in the Apple Maggot fly (Feder et al., 2003). Although the role of

17 standing variation with respect to adaptation has been described in many systems,

18 its importance to domestication is not as well studied.

19   In maize, alleles at important domestication loci (*RAMOSA1*, (**?**); *barren*

20 *stalk1*, (Gallavotti et al., 2004); and *grassy tillers1*, (Whipple et al., 2011)) have

21 been shown to have been selected from standing variation, suggesting that diversity

22 already present in teosinte may have played an important role in the domestication

23 of maize. The *teosinte branched1* gene has long been a central focus of research

24 concerning maize domestication, and, while previous studies have suggested that

1 differences in plant architecture between domesticated maize and teosinte are a

2 result of selection on standing variation, little is known about variation at this locus

3 in teosinte (Clark et al., 2006; Studer et al., 2011). Studer et al. (2011) genotyped

4 90 accessions of teosinte (inbred and outbred), providing the first evidence that the

5 *Hopscotch* insertion is segregating in teosinte (Studer et al., 2011).

6   Given that the *Hopscotch* insertion has been estimated to predate the

7 domestication of maize, it is not surprising that it can be found segregating in

8 populations of teosinte. However, in sampling numerous individuals from many

9 teosinte populations our study provides greater insight into the distribution and

10 prevalence of the *Hopscotch* in teosinte. While our findings are consistent with a

11 previous study by Studer et al. (2011) in that we identified the *Hopscotch* allele

12 segregating in teosinte, we find it at higher frequency than previously suggested

13 (Studer et al., 2011). Many of our populations with high frequency of the

14 *Hopscotch* allele fall in the Jalisco cluster identified by Fukunaga (????), possibly

15 suggesting a different history of the *tb1* locus than in the Balsas region where maize

16 was domesticated (Matsuoka et al., 2002). While gene flow from crops into their

17 wild relatives is well-known, ((Ellstrand et al., 1999; Zhang et al., 2009; Thurber

18 et al., 2010; Baack et al., 2008; Hubner et al., 2012; Wilkes, 1977; van Heerwaarden

19 et al., 2011; Barrett, 1983)), our results are more consistent with Hufford et al.

20 (2013) who found resistance to introgression from maize into teosinte (Hufford

21 et al., 2013). Furthermore, Hufford et al. (2013) showed that domestication loci,

22 such as *tb1*, are particularly resistant to introgression in both directions of gene

23 flow (i.e., maize to teosinte and teosinte to maize) (Hufford et al., 2013).

24   We find no evidence of recent introgression in our analyses. Clustering patterns

25 in our NJ trees do not reflect a pattern expected if maize alleles at the *tb1* locus

26 had introgressed into populations of teosinte. Moreover, analysis of linkage in the

1 *tb1* region does not reveal patterns of high LD relative to the rest of chromosome 1,

2 and assignment to maize in this region in our STRUCTURE analysis is lower than

3 the average across chromosome 1 (3, 4). Together, these data point to an

4 explanation other than recent introgression for the high observed frequency of

5 *Hopscotch* in some of our *parviglumis* populations.

Table 4: Assignments to maize and teosinte in the *tb1* and chromosome 1 regions from STRUCTURE

	tb1 region		Chr 1	
Population	Maize assignment	Teosinte assignment	Maize assignment	Teosinte assignment
Ejutla A	0.02158681	0.9784132	0.2026814	0.7973186
Ejutla B	0.01888194	0.9811181	0.1872131	0.8127869
La Mesa	0.0118675	0.9881333	0.8068998	0.1931017
San Lorenzo	0.01551389	0.9844861	0.2048252	0.7951748

6   Although recent introgression seems unlikely, we cannot rule out ancient

7 introgression as an explanation for the presence of the *Hopscotch* in these

8 populations. If the *Hopscotch* allele was introgressed in the distant past, they could

9 have been sufficient recombination to break up any initial LD, leading to

10 observations similar to those obtained here. We find this scenario less plausible,

11 however, as there is no reason why gene flow should have been high in the past but

12 absent in present-day sympatric populations. In fact, early generation

13 maize-teosinte hybrids are easy to find in these populations today (MB Hufford,

14 pers. observation), and genetic data support ongoing gene flow between

15 domesticated maize and both *Zea mays* ssp. *mexicana* and *Zea mays* ssp.

16 *parviglumis* in a number of sympatric populations (Hufford et al., 2013; Ellstrand

17 et al., 2007; **?**).

1   Other explanation for differential frequencies of the *Hopscotch* among teosinte

2   populations include both drift and natural selection. Previous studies using both

3   SSRs and genome-wide SNP data have found evidence for a population bottleneck

4   in the San Lorenzo population (Hufford, 2010; Pyhäjärvi et al., 2013), and the

5   lower levels of sequence diversity in the 5' UTR region and the more positive values

6   of Tajima's D we present here are consistent with these findings. *deviations from HWE*

7   *may be consistent too if we see excess of homozygotes. do we?* . This bottleneck, however, does not

8   explain differences in *Hopscotch* allele frequency among populations, and the

9   available information on diversity and population structure among these

10   populations (Hufford, 2010; Pyhäjärvi et al., 2013) is not suggestive of colonization

11   or other demographic events that might predict a high frequency of the allele in

12   multiple populations. *here we need a few sentences on selection. the 5' UTR has much more negative D*

13   *than the upstream. do we know the Hop genotype for sequenced lines? can we separate the sequences into hop/no*

14   *hop and look for differences? it wasn't until we did this that gt1 stuff really popped out.*   *we should know for some*

15   *of them, i will check*

16   The phenotypic effects of the *Hopscotch* insertion in domesticated maize have

17   been well documented (Clark et al., 2006; Studer et al., 2011), and Weber et al.

18   (2007) have described its effects in partially inbred lines of teosinte (Weber et al.,

19   2007) *i don't think these were inbred.please doublecheck.* . Our study is the first to explicitly

20   examine the phenotypic effects of the *Hopscotch* insertion in individuals sampled

21   from a natural population of teosinte. *isn't this what weber did?? for 70+ populations!?*

22   However, we found no significant effect of the *Hopscotch* on tillering index or tiller

23   number in our phenotyping experiments, and the effect of the *Hopscotch* insertion

24   in teosinte is discordant with that of maize. The lack of correlation between

25   *Hopscotch* genotype and tillering index or tiller number is surprising given its

26   effects in maize. It is certainly possible that even though previous data

24

demonstrate an effect of the *Hopscotch* on tillering in maize (Studer et al., 2011),
that the effect of the *Hopscotch* in teosinte is more complicated and may be more
difficult to observe. Moreover, *tb1* is a single gene in a complex pathway that
affects branching and tillering traits, and perhaps in combination with alleles at
other loci the phenotypic effects of the *Hopscotch* on tillering may not be
consistent. *thsi section still needs work. i think we have to do more here. weber shows an association between*
SNPs in tb1 and branch length. we need to discuss that!

   MBH todo Variation at *tb1* has also been shown to contribute to phenotypes
other than tillering (Clark et al., 2006), and a recent study by Studer and Doebley
(2012) examined the possibility of an allelic series at the *tb1* locus in teosinte.
Studer and Doebley (2012) introgressed 9 separate teosinte segments (one from *Zea*
diploperennis, and four from both *Zea mays* ssp. *mexicana* and *Zea mays* ssp.
parviglumis) spanning the *tb1* locus into an isogenic maize background and
investigated their effects on previously associated phenotypes. They found that
plants with teosinte chromosomal segments had greater tillering than their maize
isogenic lines, and that different chromosomal segments of *tb1* confer different
amounts of tillering, suggesting that there are multiple genetic factors in this region
that affect tillering. However, in addition to elucidating variance in tillering among
tb1 teosinte segments, Studer and Doebley (2012) found significant variance among
W22 control lines, suggesting that there are other genetic factors aside from alleles
at the *tb1* locus that affect tillering in maize. Doebley and Stec (1991, 1993) first
attempted to map QTL controlling many of the phenotypic differences between
domesticated maize and teosinte, and demonstrated the existence of numerous
QTL that contribute to the differences in branching architecture between the two.
Many of these loci (*grassy tillers*, *gt1*; *tassel-replaces-upper-ears1*, *tru1*; *terminal*
ear1, *ter1*) have been shown to interact with *tb1* (Whipple et al., 2011; Li, 2012),

and both *tru1* and *ter1* have been shown to affect the same phenotypic traits as *tb1* (Doebley et al., 1995). *tassel-replaces-upper-ears1* (*tru1*), for example, has been shown to act either epistatically or downstream of *tb1*, affecting both branching architecture (decreased apical dominance) and tassel phenotypes (shortened tassel and shank length and reduced tassel number) (Li, 2012). It seems plausible that variation in some of these other loci could have affected tillering in our greenhouse population, and contributed to the lack of correlation we see between *Hopscotch* genotype and tillering.

In summary, our findings demonstrate that the *Hopscotch* allele is more widespread in populations of *parviglumis* and *mexicana* than previously thought. Analysis of linkage using SNPs from across chromosome 1 does not suggest that the *Hopscotch* allele is present in these populations due to recent introgression; however, it seems unlikely that it would have drifted to high frequency in multiple populations and there may be another explanation for the high frequency we observe in some of our populations. The *Hopscotch* does not appear to have a strong effect reducing tillering in teosinte as it does in maize, and other loci involved in branching architecture may play also play roles in the regulation of tillering in teosinte. Finally, although we see no clear evidence of recent strong selection, the high frequency of the *Hopscotch* insertion in a number of populations continues to suggest to us that it plays an ecological role in teosinte. In the future, additional experiments will be needed to examine expression levels of *tb1* and additional loci involved in branching architecture (e.g. *gt1*, *tru1*, and *ter1*) in conjunction with a more exhaustive phenotyping and genotyping assay. *why not Phyb and phya?* *Are they necessary to include? I'd had them in before in a paragraph but had been voted out* *I'd ditch gt1 tru1 ter1 and maybe just cite some people including phyb etc.*

please check format of supp figs and tables; some are running off the page. you can use "longtable" to fix that

1 *(ask Paul for example). check fig/table references, bibliography, etc. what does "rotation" mean in supp. table 3?*

1 *it isn't mentioned in methods. please check that all the tables and figs (including supplement) are referenced in the*
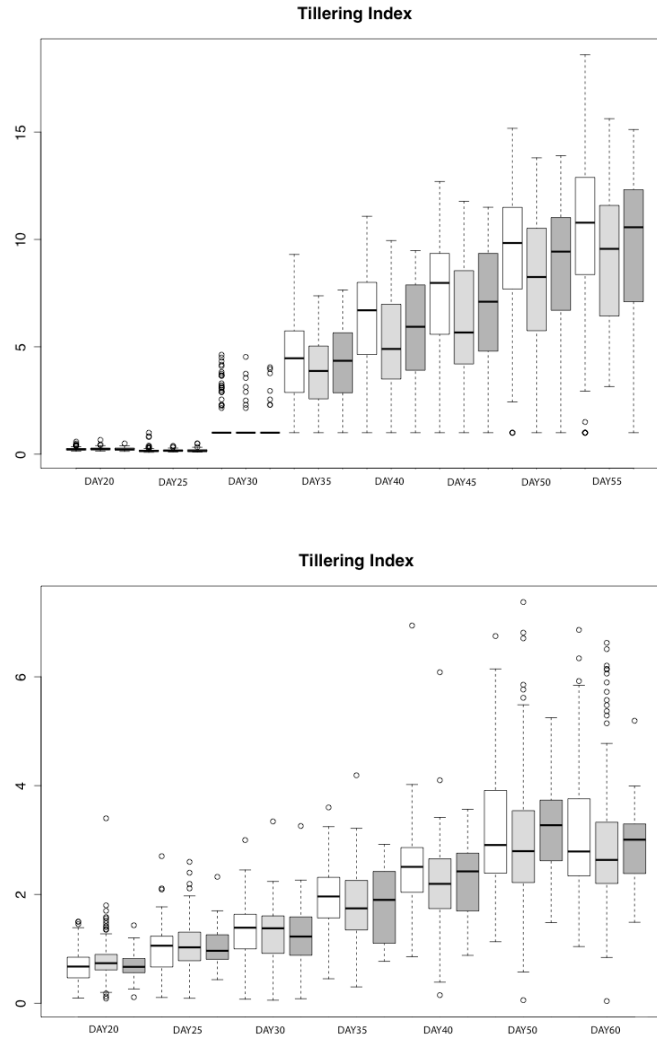
2 *text.*

Figure 4: Box-plot showing tillering index in our greenhouse population from day 20-60. White indicates individuals homozygous for the *Hopscotch*, light grey represents heterozygotes, and dark grey represents homozygotes for the teosinte (No *Hopscotch*) allele. Within boxes, dark black lines represent the median, and the edges of the boxes are the first and third quartiles.

References

BAACK, E., Y. SAPIR, M. CHAPMAN, J. BURKE, AND L. RIESEBERG. 2008.
Selection on domestication traits and quantitative trait loci in crop-wild
sunflower hybrids. *Mol Ecol* 17: 666–677.

BAI, Y. AND P. LINDHOUT. 2007. Domestication and breeding of tomatoes: What
have we gained and what can we gain in the future? *Annals of Botany* 100:
1085–1094.

BARRETT, S. 1983. Crop mimicry in weeds. *Econ Bot* 37: 255–282.

BRADBURY, P., Z. ZHANG, D. KROON, T. CASSTEVENS, Y. RAMDOSS, AND
E. BUCKLER. 2007. Tassel: software for association mapping of complex traits in
diverse samples. *Bioinformatics* 23: 2633–2635.

BRIGGS, W., M. MCMULLEN, B. GAUT, AND J. DOEBLEY. 2007. Linkage
mapping of domestication loci in a large maize-teosinte backcross resource.
Genetics 177: 1915–1928.

CHIA, J., C. SONG, P. BRADBURY, D. COSTICH, N. DE, LEON, J. DOEBLEY,
R. ELSHIRE, B. GAUT, L. GELLER, J. GLAUBITZ, M. GORE, K. GUILL,
J. HOLLAND, M. HUFFORD, J. LAI, M. LI, X. LIU, Y. LU, R. MCCOMBIE,
R. NELSON, J. POLAND, B. PRASANNA, T. PYHÄJÄRVI, T. RONG,
R. SEKHON, Q. SUN, M. TENAILLON, F. TIAN, J. WANG, X. XU,
Z. ZHANG, S. KAEPPLER, J. ROSS-IBARRA, M. MCMULLEN, E. BUCKLER,
G. ZHANG, Y. XU, AND D. WARE. 2012. Maize hapmap2 identifies extant
variation from a genome in flux. *Nat Genet* 44: 803–U238.

CLARK, R., T. WAGLER, P. QUIJADA, AND J. DOEBLEY. 2006. A distant
upstream enhancer at the maize domestication gene tb1 has pleiotropic effects on
plant and inflorescent architecture. *Nat Genet* 38: 594–597.

COLOSIMO, P., K. HOSEMANN, S. BALABHADRA, G. VILLARREAL,
M. DICKSON, J. GRIMWOOD, J. SCHMUTZ, R. MYERS, D. SCHLUTER, AND
D. KINGSLEY. 2005. Widespread parallel evolution in sticklebacks by repeated
fixation of ectodysplasin alleles. *Science* 307: 1928–1933.

COOK, J., M. MCMULLEN, J. HOLLAND, F. TIAN, P. BRADBURY,
J. ROSS-IBARRA, E. BUCKLER, AND S. FLINT-GARCIA. 2012. Genetic
architecture of maize kernel composition in the nested association mapping and
inbred association panels. *Plant Physiol* 158: 824–834.

COOP, G., D. WITONSKY, A. DI, RIENZO, AND J. PRITCHARD. 2010. Using
environmental correlations to identify loci underlying local adaptation. *Genetics*
185: 1411–1423.

DOEBLEY, J. 2004. The genetics of maize evolution. *Annu Rev Genet* 38: 37–59.

DOEBLEY, J. AND A. STEC. 1991. Genetic-analysis of the morphological
differences between maize and teosinte. *Genetics* 129: 285–295.

DOEBLEY, J. AND A. STEC. 1993. Inheritance of the morphological differences
between maize and teosinte: Comparison of results for two f_2 populations.
Genetics 134: 559–570.

DOEBLEY, J., A. STEC, AND C. GUSTUS. 1995. *teosinte branched1* and the origin
of maize: Evidence for epistasis and the evolution of dominance. *Genetics* 141:
333–346.

3 DOEBLEY, J., A. STEC, AND L. HUBBARD. 1997. The evolution of apical
4   dominance in maize. *Nature* 386: 485–488.

5 DOYLE, J. AND J. DOYLE. 1990. A rapid total dna preparation procedure for
6   small quantities of fresh tissue. *Phytochemical Bulletin* 19: 11–15.

7 ELLSTRAND, N., L. GARNER, S. HEDGE, R. GUADAGNUOLO, AND
8   L. BLANCAS. 2007. Spontaneous hybridization between maize and teosinte.
9   *Journal of Heredity* 98: 183–187.

10 ELLSTRAND, N., H. PRENTICE, AND J. HANCOCK. 1999. Gene flow and
11   introgression from domesticated plants into their wild relatives. *Annu Rev Ecol*
12   *Syst* 30: 539–563.

13 FALUSH, D., M. STEPHENS, AND J. PRITCHARD. 2003. Inference of population
14   structure using multilocus genotype data: Linked loci and correlated allele
15   frequencies. *Genetics* 164: 1567–1587.

16 FEDER, J., S. BERLOCHER, J. ROETHELE, H. DAMBROSKI, J. SMITH,
17   W. PERRY, V. GAVRILOVIC, K. FILCHAK, J. RULL, AND M. ALUJA. 2003.
18   Allopatric genetic origins for sympatric host-plant shifts and race formation in
19   rhagoletis. *P Natl Acad Sci Usa* 100: 10314–10319.

20 FOX, J. AND S. WEISBERG. 2011. An R Companion to Applied Regression, vol.
21   Second Edition. Sage, Thousand Oaks, CA.

1 FUKUNAGA, N.-W. T. L. B. Z. Q. V. Y. F. M. B. K. L. L. D. J., K. ????
2   Genetic diversity and population structure of teosinte. *Genetics* .

3 GALLAVOTTI, A., Q. ZHAO, J. KYOZUKA, R. MEELEY, M. RITTER,
4   J. DOEBLEY, M. PE, AND R. SCHMIDT. 2004. The role of barren stalk1 in the
5   architecture of maize. *Nature* 432: 630–635.

6 GERKE, J., J. EDWARDS, G. KE, J. ROSS-IBARRA, AND M. MCMULLEN. 2013.
7   The genomic impacts of drift and selection for hybrid performance in maize.
8   *arXiv* 1307.7313.

9 GOUDET, J. 2005. Hierfstat, a package for r to compute and test hierarchical
10   f-statistics. *Mol Ecol Notes* 5: 184–186.

11 HUBNER, S., T. GUNTHER, A. FLAVELL, E. FRIDMAN, A. GRANER,
12   A. KOROL, AND K. SCHMID. 2012. Islands and streams: clusters and gene flow
13   in wild barley populations from the levant. *Mol Ecol* 21: 1115–1129.

14 HUFFORD, M. 2010. Genetic and ecological approaches to guide conservation of
15   teosinte (zea mays ssp. parviglumis), the wild progenitor of maize. *PhD*
16   *Dissertation* : 130pp.

17 HUFFORD, M., P. BILINSKI, T. PYHÄJÄRVI, AND J. ROSS-IBARRA. 2012a.
18   Teosinte as a model system for population and ecological genomics. *Trends in*
19   *Genetics* 12: 606–615.

20 HUFFORD, M., P. LUBINSKY, T. PYHÄJÄRVI, M. DEVENGENZO,
21   N. ELLSTRAND, AND J. ROSS-IBARRA. 2013. The genomic signature of
22   crop-wild introgression in maize. *PLoS Genetics* 9: e1003477.

23 HUFFORD, M., X. XU, J. VAN, HEERWAARDEN, T. PYHÄJÄRVI, J. CHIA,
1   R. CARTWRIGHT, R. ELSHIRE, J. GLAUBITZ, K. GUILL, S. KAEPPLER,
2   J. LAI, P. MORRELL, L. SHANNON, C. SONG, N. SPRINGER,

R. Swanson-Wagner, P. Tiffin, J. Wang, G. Zhang, J. Doebley, M. McMullen, D. Ware, E. Buckler, S. Yang, and J. Ross-Ibarra. 2012b. Comparative population genomics of maize domestication and improvement. *Nat Genet* 44: 808–U118.

Kearse, M., R. Moir, A. Wilson, S. Stones-Havas, M. Cheung, S. Sturrock, S. Buxton, A. Cooper, S. Markowitz, C. Duran, T. Thierer, B. Ashton, P. Meintjes, and A. Drummond. 2012. Geneious basic: An integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28: 1647–1649.

Kebrom, T. and T. Brutnell. 2007. The molecular analysis of the shade avoidance syndrome in the grasses has begun. *Journal of Experimental Botany* 58: 3079–3089.

Kitano, J., D. Bolnick, D. Beauchamp, M. Mazur, S. Mori, T. Nakano, and C. Peichel. 2008. Reverse evolution of armor plates in the threespine stickleback. *Curr Biol* 18: 769–774.

Kovach, M. and S. McCouch. 2008. Leveraging natural diversity: back through the bottleneck. *Genome studies and Molecular Genetics* 11: 193–200.

Li, W. 2012. Tassels replace upper ears1 encodes a putative transcription factor that regulates maize shoot architecture by multiple pathways. *PhD Dissertation* : 122.

Lukens, L. and J. Doebley. 1999. Epistatic and environmental interactions for quantitative trait loci involved in maize evolution. *Genet Res* 74: 291–302.

3 MALOOF, M., K. SOLIMAN, R. JORGENSEN, AND R. ALLARD. 1984. Ribosomal
   dna spacer length polymorphisms in barley - mendelian inheritance, chromosomal
   location, and population dynamics. *P Natl Acad Sci Usa* 81: 8014–8018.

6 MATSUOKA, Y., Y. VIGOUROUX, M. GOODMAN, G. SANCHEZ, E. BUCKLER,
   AND J. DOEBLEY. 2002. A single domestication for maize shown by multilocus
   microsatellite genotyping. *P Natl Acad Sci Usa* 99: 6080–6084.

9 OLSEN, K. AND B. GROSS. 2010. Genetic perspectives on crop domestication.
   Trends in Plant Science 15: 529–537.

11 PIPERNO, D., A. RANERE, I. HOLST, J. IRIARTE, AND R. DICKAU. 2009.
   Starch grain and phytolith evidence for early ninth millennium bp maize from
   the central balsas river valley, mexico. *P Natl Acad Sci Usa* 106: 5019–5024.

14 PLANTINGA, T., S. ALONSO, N. IZAGIRRE, M. HERVELLA, R. FREGEL,
   J. VAN DER MEER, M. NETEA, AND C. DE LA RUA. 2012. Low prevalence of
   lactase persistence in neolithic south-west europe. *Eur J Hum Genet* 20:
   778–782.

18 PYHÄJÄRVI, T., M. HUFFORD, AND J. ROSS-IBARRA. 2013. Complex patterns of
   local adaptation in the wild relatives of maize. *Genome Biology and Evolution* 5:
   1594–1609.

21 ROSS-IBARRA, J., P. MORRELL, AND B. GAUT. 2007. Plant domestication, a
   unique opportunity to identify the genetic basis of adaptation. *P Natl Acad Sci
   Usa* 104: 8641–8648.

1 ROSS-IBARRA, J., M. TENAILLON, AND B. GAUT. 2009. Historical divergence
   and gene flow in the genus zea. *Genetics* 181: 1399–1413.

3 ROZEN, S. AND H. SKALETSKY. 2000. Primer3 on the www for general users and
4   for biologist programmers. *Methods in Molecular Biology* : 365–386.

5 STUDER, A. AND J. DOEBLEY. 2012. Evidence for a natural allelic series at the
6   maize domestication locus teosinte branched1. *Genetics* 19: 951–958.

7 STUDER, A., Q. ZHAO, J. ROSS-IBARRA, AND J. DOEBLEY. 2011. Identification
8   of a functional transposon insertion in the maize domestication gene tb1. *Nat*
9   *Genet* 43: 1160–U164.

10 THORNTON, K. 2003. libsequence: a c++ class library for evolutionary genetic
11   analysis. *Bioinformatics* 19: 2325–2327.

12 THURBER, C., M. REAGON, B. GROSS, K. OLSEN, Y. JIA, AND A. CAICEDO.
13   2010. Molecular evolution of shattering loci in us weedy rice. *Mol Ecol* 19:
14   3271–3284.

15 TISHKOFF, S., F. REED, A. RANCIARO, B. VOIGHT, C. BABBITT,
16   J. SILVERMAN, K. POWELL, H. MORTENSEN, J. HIRBO, M. OSMAN,
17   M. IBRAHIM, S. OMAR, G. LEMA, T. NYAMBO, J. GHORI, S. BUMPSTEAD,
18   J. PRITCHARD, G. WRAY, AND P. DELOUKAS. 2007. Convergent adaptation of
19   human lactase persistence in africa and europe. *Nat Genet* 39: 31–40.

20 VAN HEERWAARDEN, J., J. DOEBLEY, W. BRIGGS, J. GLAUBITZ,
21   M. GOODMAN, J. GONZALEZ, AND J. ROSS-IBARRA. 2011. Genetic signals of
22   origin, spread, and introgression in a large sample of maize landraces. *P Natl*
23   *Acad Sci Usa* 108: 1088–1092.

1 WEBER, A., R. CLARK, L. VAUGHN, J. SANCHEZ-GONZALEZ, J. YU,
2   B. YANDELL, P. BRADBURY, AND J. DOEBLEY. 2007. Major regulatory genes

in maize contribute to standing variation in teosinte (zea mays ssp parviglumis). *Genetics* 177: 2349–2359.

WHIPPLE, C., T. KEBROM, A. WEBER, F. YANG, D. HALL, R. MEELEY, R. SCHMIDT, J. DOEBLEY, T. BRUTNELL, AND D. JACKSON. 2011. grassy tillers1 promotes apical dominance in maize and responds to shade signals in the grasses. *P Natl Acad Sci Usa* 108: E506–E512.

WHITTON J, A. D. S. A. R. L., WOLF DE. 1997. The persistence of cultivar alleles in wild populations of sunflowers fiver generations after hybridization. *Theoretical and Applied Genetcs* 95: 33–40.

WILKES, H. 1977. Hybridization of maize and teosinte, in mexico and guatemala and the improvement of maize. *Economic Botany* 31: 254–293.

ZHANG, L., Q. ZHU, Z. WU, J. ROSS-IBARRA, B. GAUT, S. GE, AND T. SANG. 2009. Selection on grain shattering genes and rates of rice domestication. *New Phytol* 184: 708–720.

Supplementary Materials

5' Hopscotch tbl 3'

1,235bp

1,335bp

58.5kb
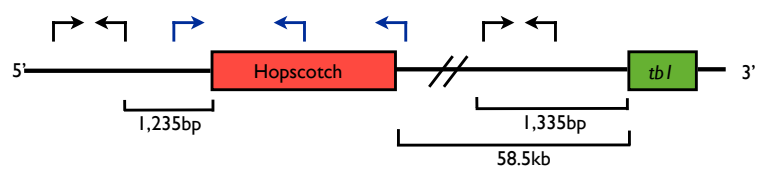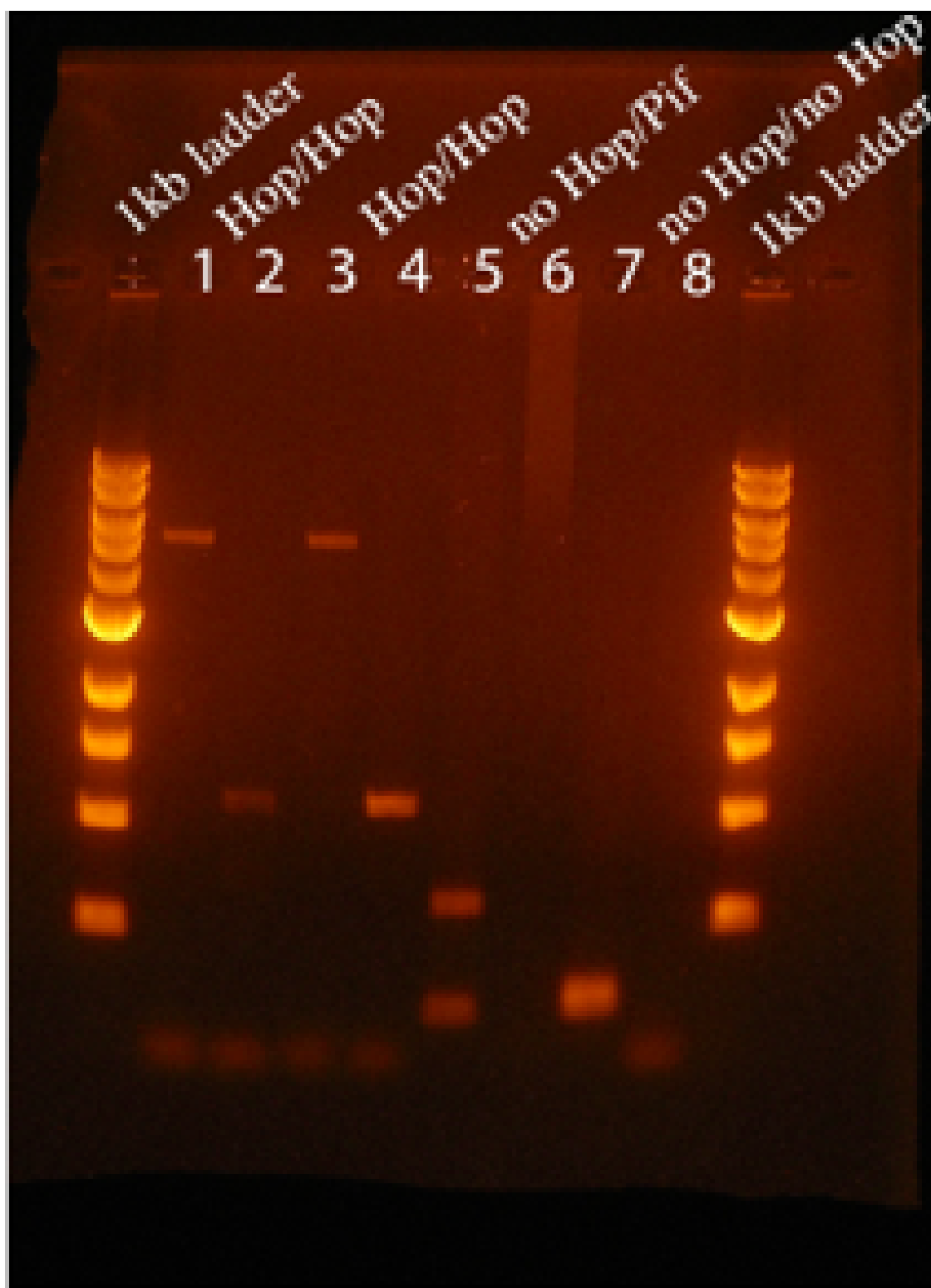
Accession	USDA Accession ID	Locality	Number alleles sampled	*Ho*
RIHY0009	N/A	N/A	2	
RIMME0006	566673	Durango, Mexico	2	
RIMME0007	566680	Guanajuato, Mexico	2	
RIMME0008	566681	Michoacan, Mexico	2	
RIMME0009	566682	Distrito Federal, Mexico	2	
RIMME0011	566685	Mexico, Mexico	2	
RIMME0014	714151	Breeders line; Puga: 11066	6	
RIMME0017	699874	Ayotlan, Mexico	8	
RIMME0021	N/A	El Porvenir, Mexico	69	
RIMME0026	N/A	Opopeo, Mexico	42	
RIMME0028	N/A	Puruandiro, Mexico	28	
RIMME0029	N/A	Ixtlan, Mexico	35	
RIMME0030	N/A	San Pedro, Mexico	27	
RIMME0031	N/A	Tenango del Aire, Mexico	25	
RIMME0032	N/A	Nabogame, Mexico	24	
RIMME0033	N/A	Puerta Encantada, Mexico	25	
RIMME0034	N/A	Santa Clara, Mexico	23	
RIMME0035	N/A	Xochimilco, Mexico	25	
RIMPA0001	87168	El Salado, Mexico	4	
RIMPA0003	87171	Mazatlan, Mexico	8	
RIMPA0017	87200	N/A	4	
RIMPA0019	87213	El Salado, Mexico	2	
RIMPA0029	87244	N/A	2	
RIMPA0031	87249	N/A	2	
RIMPA0035	87288	Jalisco, Mexico	4	
RIMPA0040	288185	Mexico, Mexico	4	
RIMPA0042	288187	Guerrero, Mexico	4	
RIMPA0043	288188	Guerrero, Mexico	4	
RIMPA0045	288193	Guerrero, Mexico	4	
RIMPA0055	714152	Breeders line	2	
RIMPA0056	714153	Breeders line	2	

Accession	Number of alleles sampled	*Hopscotch* Frequency
RIMMA0066	2	1
RIMMA0075	2	1
RIMMA0077	2	1
RIMMA0079	2	1
RIMMA0081	2	1
RIMMA0084	2	1
RIMMA0086	2	1
RIMMA0088	2	1
RIMMA0089	2	1
RIMMA0090	2	1
RIMMA0092	4	1
RIMMA0094	4	1
RIMMA0097	2	1
RIMMA0099	2	1
RIMMA0100	2	1
RIMMA0101	2	1
RIMMA0104	2	1
RIMMA0108	2	1
RIMMA0111	6	1
RIMMA0115	2	1
RIMMA0117	2	1
RIMMA0130	2	1
RIMMA0133	2	1
RIMMA0134	2	1
RIMMA0135	2	1
RIMMA0142	2	0.5
RIMMA0143	4	1
RIMMA0146	4	1
RIMMA0149	2	1
RIMMA0152	2	1
RIMMA0153	2	1

PC1		PC2		PC3		PC4		PC5	
Var	**Rot**	**Var**	**Rot**	**Var**	**Rot**	**Var**	**Rot**	**Var**	**Rot**
bio1	0.146	bio4	0.244	prec7	0.287	ts_clay	0.41	bio2	0.38
tmean11	0.146	bio3	0.241	prec8	0.276	v_mod	0.359	sq4	0.328
tmean12	0.145	bio7	0.241	prec11	0.262	ts_sand	0.329	ts_loam	0.289
bio11	0.145	prec6	0.237	bio13	0.247	bio15	0.272	ts_sand	0.266
tmax12	0.145	sq7	0.218	prec1	0.246	prec4	0.259	sq7	0.231
tmin5	0.145	prec9	0.217	bio16	0.242	x_mod	0.244	bio18	0.213
tmean1	0.145	sq3	0.207	prec12	0.24	prec3	0.226	bio13	0.207
tmean2	0.145	prec12	0.207	bio19	0.238	sq3	0.21	prec11	0.183
tmin4	0.145	bio12	0.204	bio12	0.231	prec5	0.21	bio7	0.17
tmax1	0.145	bio19	0.196	prec2	0.222	prec7	0.19	bio16	0.163
tmean4	0.145	prec2	0.188	bio18	0.221	sq4	0.186	bio4	0.157
tmin11	0.144	prec1	0.185	sq4	0.2	bio3	0.185	bio12	0.156
tmax11	0.144	prec10	0.184	prec9	0.18	bio18	0.178	bio3	0.155
tmin12	0.144	bio16	0.183	prec10	0.171	sq7	0.132	prec6	0.154
tmin2	0.144	prec8	0.17	prec5	0.161	bio14	0.116	x_mod	0.152
tmean5	0.144	prec5	0.165	prec4	0.154	bio13	0.099	prec9	0.144
tmean10	0.144	bio14	0.158	sq3	0.147	bio16	0.095	prec8	0.143
bio6	0.144	bio13	0.151	bio2	0.143	prec8	0.09	v_mod	0.142
tmax2	0.144	bio17	0.149	bio17	0.129	bio7	0.077	bio15	0.136
tmean3	0.144	prec3	0.144	ts_loam	0.127	bio4	0.075	prec7	0.112
tmin1	0.143	ts_clay	0.141	v_mod	0.123	bio2	0.074	prec4	0.108
tmin10	0.143	bio2	0.129	prec3	0.113	prec2	0.074	bio14	0.096
Altitude	0.143	prec7	0.108	x_mod	0.111	bio19	0.068	tmax7	0.093
bio9	0.143	tmax6	0.107	bio14	0.099	prec12	0.056	tmax8	0.092
tmin3	0.143	x_mod	0.106	bio4	0.07	ts_loam	0.053	prec1	0.091
bio10	0.142	bio15	0.098	tmax3	0.067	tmax12	0.047	prec2	0.086
tmax10	0.142	ts_loam	0.088	ts_clay	0.065	bio17	0.047	tmin11	0.086
tmax3	0.142	tmean6	0.085	bio15	0.056	bio9	0.043	prec5	0.082
tmax4	0.142	tmin7	0.082	tmax2	0.055	tmax8	0.042	bio17	0.082
tmin6	0.142	bio5	0.082	tmean3	0.052	tmax1	0.041	tmin12	0.08

Ejutla A	4	0.15217	0.11902	0.76191
Ejutla B	5	0.15258	0.14877	0.07412
La Mesa	3	0.12802	0.08926	1.09209
San Lorenzo	3	0.09098	0.08926	0.04845