

# Assignment 3: Data Exploration

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## OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

## Directions

1. Change “Student Name, Section #” on line 3 (above) with your name and section number.
2. Work through the steps, **creating code and output** that fulfill each instruction.
3. Be sure to **answer the questions** in this assignment document.
4. When you have completed the assignment, **Knit** the text and code into a single PDF file.
5. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai. Add your last name into the file name (e.g., “FirstLast\_A03\_DataExploration.Rmd”) prior to submission.

The completed exercise is due on <>.

## Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX\_Neonicotinoids\_Insects\_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON\_NIWO\_Litter\_massdata\_2018-08\_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. **Be sure to add the `stringsAsFactors = TRUE` parameter to the function when reading in the CSV files.**

```
# Check working directory
getwd()
```

```
## [1] "/Users/lydiecostes/Documents/Duke/DataAnalytics/GithubRepos/Environmental_Data_Analytics_2022/A
```

```
# Import packages
library(tidyverse)
# Read in the data
Neonics <- read.csv("../Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv",
                    stringsAsFactors = T)
Litter <- read.csv("../Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv",
                   stringsAsFactors = T)
```

## Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency’s ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely

in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer:

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: It could help inform knowledge of changes in nutrient cycling (nitrogen, carbon), biomass, and biodiversity.

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: \* Plant debris samples are sorted into functional groups, such as leaves, needles, seeds, woody material, and other (lichens, mosses, unidentified). \* Samples are collected in ground traps and elevated traps. \* Sampling interval depends on multiple factors including vegetation type, access, and trap type, ranging from once per year to every two weeks.

## Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
# Check data dimensions
dim(Neonics)
```

```
## [1] 4623  30
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
# summarize effects
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##              12              102              360              11
##      Cell(s)      Development      Enzyme(s)      Feeding behavior
##              9              136              62              255
##      Genetics      Growth      Histology      Hormone(s)
##              82              38              5              1
##      Immunological      Intoxication      Morphology      Mortality
##              16              12              22              1493
##      Physiology      Population      Reproduction
##              7              1803              197
```

Answer: The most commonly studied effects are population and mortality. Both of these variables are important to track because changes can indicate the health and success of a species.

- Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

```
# Summarize species (by common name)
summary(Neonics$Species.Common.Name)
```

```
## Honey Bee Parasitic Wasp
## 667 285
## Buff Tailed Bumblebee Carniolan Honey Bee
## 183 152
## Bumble Bee Italian Honeybee
## 140 113
## Japanese Beetle Asian Lady Beetle
## 94 76
## Euonymus Scale Wireworm
## 75 69
## European Dark Bee Minute Pirate Bug
## 66 62
## Asian Citrus Psyllid Parastic Wasp
## 60 58
## Colorado Potato Beetle Parasitoid Wasp
## 57 51
## Erythrina Gall Wasp Beetle Order
## 49 47
## Snout Beetle Family, Weevil Sevenspotted Lady Beetle
## 47 46
## True Bug Order Buff-tailed Bumblebee
## 45 39
## Aphid Family Cabbage Looper
## 38 38
## Sweetpotato Whitefly Braconid Wasp
## 37 33
## Cotton Aphid Predatory Mite
## 33 33
## Ladybird Beetle Family Parasitoid
## 30 30
## Scarab Beetle Spring Tiphia
## 29 29
## Thrip Order Ground Beetle Family
## 29 27
## Rove Beetle Family Tobacco Aphid
## 27 27
## Chalcid Wasp Convergent Lady Beetle
## 25 25
## Stingless Bee Spider/Mite Class
## 25 24
## Tobacco Flea Beetle Citrus Leafminer
## 24 23
## Ladybird Beetle Mason Bee
```

##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17
##	Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
##	16	16
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
##	13	13
##	Yellow Fever Mosquito	Braconid Parasitoid
##	13	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Spotless Ladybird Beetle	Glasshouse Potato Wasp
##	11	10
##	Lacewing	Southern House Mosquito
##	10	10
##	Two Spotted Lady Beetle	Ant Family

```
##                               10                               9
##                               Apple Maggot                     (Other)
##                               9                               670
```

Answer: 1. Honey Bee 2. Parasitic Wasp 3. Buff Tailed Bumblebee 4. Carniolan Honey Bee 5. Bumble Bee 6. Italian Honey Bee

Bees and wasps are important pollinators and some of these species may be endangered. Plus, species like non-native (i.e., European) honey bees have posed a threat to native bees because they outcompete them, even though they continue to be propped up by the honey industry.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` in the dataset, and why is it not numeric?

```
# Check class of "Conc.1..Author"
class(Neonics$Conc.1..Author.)
```

```
## [1] "factor"
```

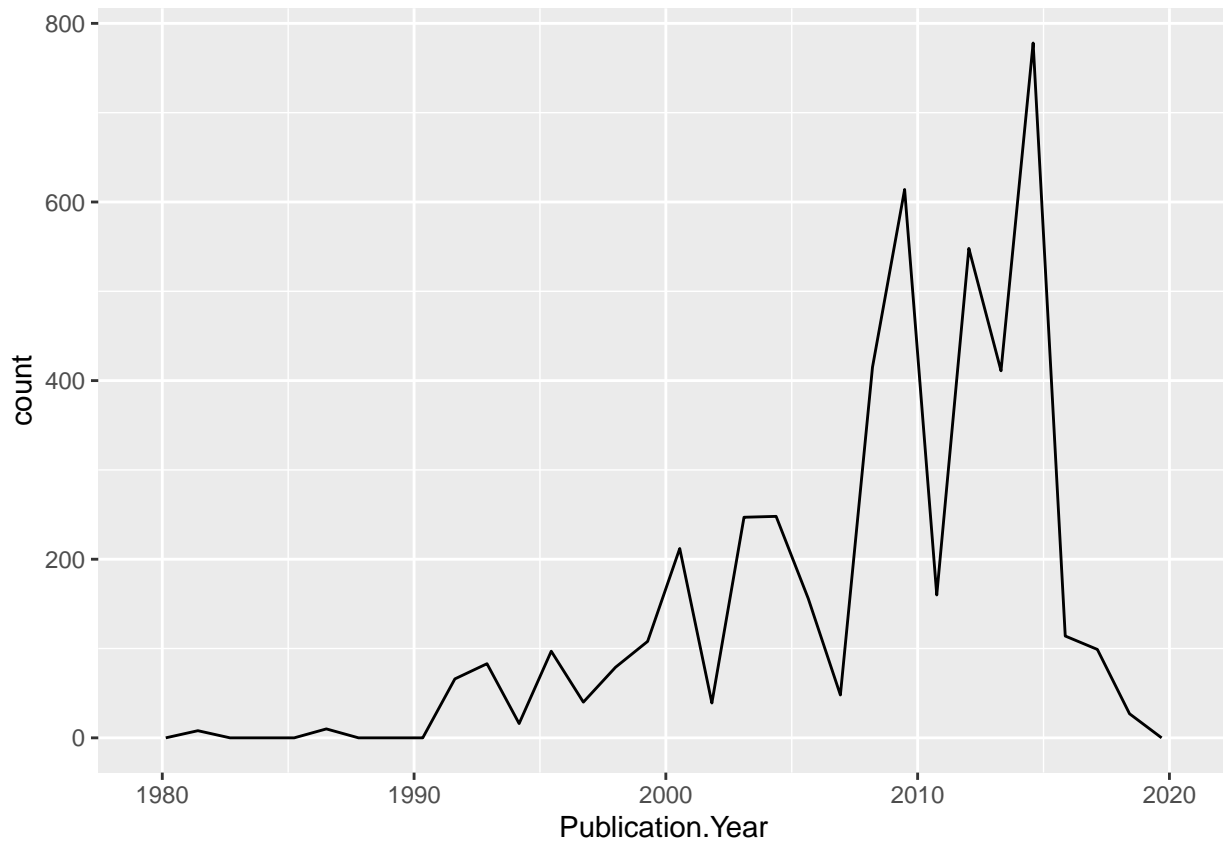
Answer: It is a factor because some of the values are “NR” or end with a slash (e.g., “144.0/”), so R doesn’t read it as numeric.

## Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
# Graph number of studies by publication year
ggplot(Neonics) +
  geom_freqpoly(aes(x=Publication.Year))
```

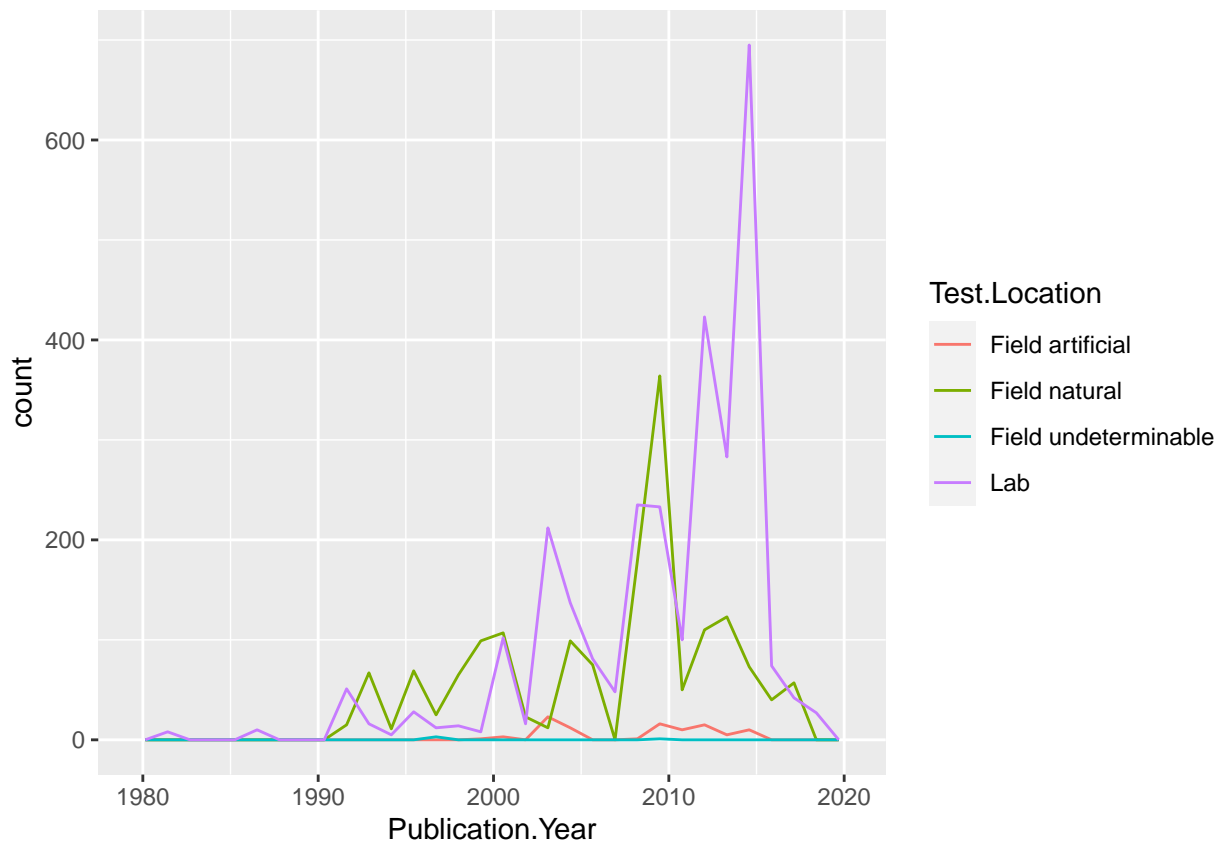
```
## ‘stat_bin()’ using ‘bins = 30’. Pick better value with ‘binwidth’.
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
# Graph number of studies by publication year, colored by location
ggplot(Neonics) +
  geom_freqpoly(aes(x=Publication.Year, color = Test.Location))
```

```
## 'stat_bin()' using 'bins = 30'. Pick better value with 'binwidth'.
```

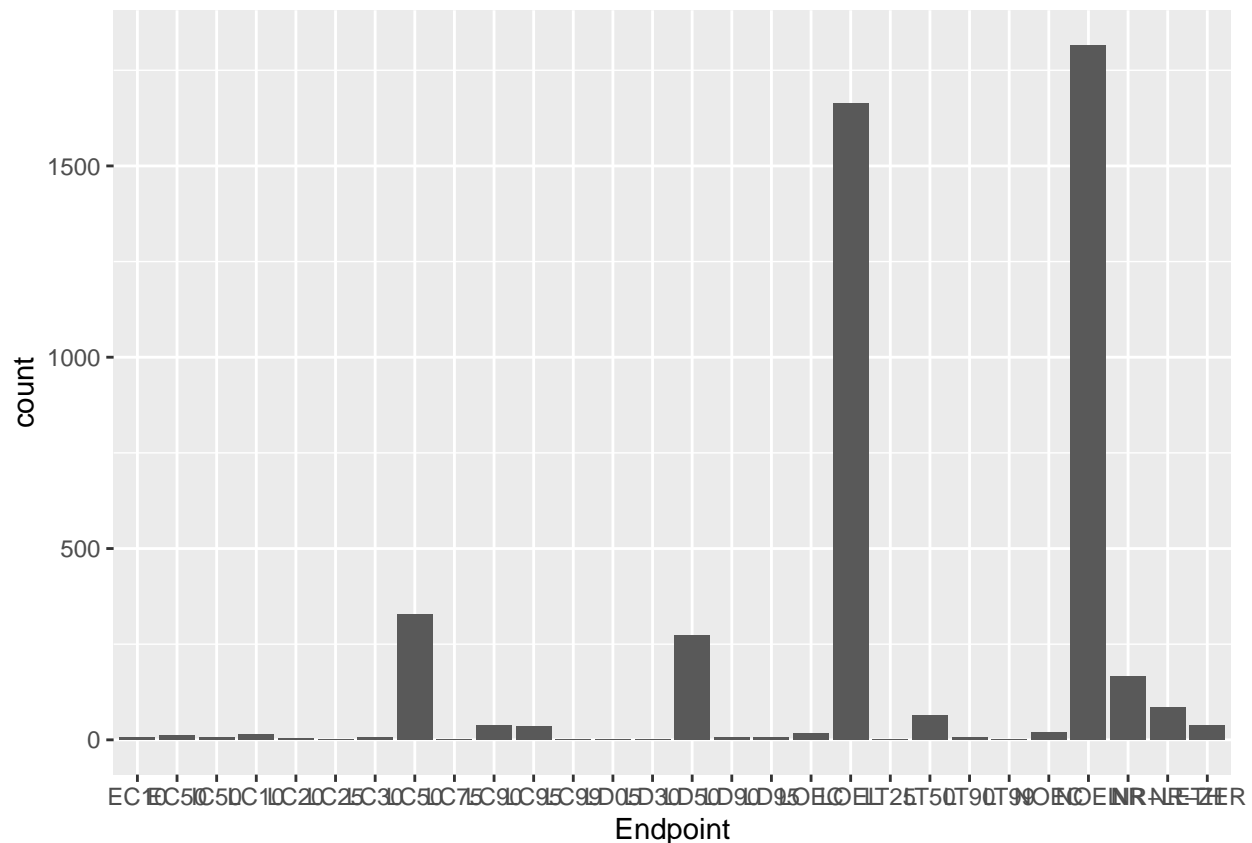


Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are natural field and lab, and in the past decade, lab research has really taken over compared with other types.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX\_CodeAppendix for more information.

```
# Graph endpoint counts
ggplot(Neonics) +
  geom_bar(aes(x=Endpoint))
```



Answer: NOEL and LOEL are the two most common endpoints. NOEL: No Observable Effect Level - highest dose producing effects not significantly different from controls LOEL: Lowest Observable Effect Level - lowest dose producing effects significantly different from controls

## Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
# Check "collectDate" class
```

```
class(Litter$collectDate)
```

```
## [1] "factor"
```

```
# Change "collectDate" class to date
```

```
Litter$collectDate <- as.Date(Litter$collectDate, format="%Y-%m-%d")
```

```
# Check "collectDate" class
```

```
class(Litter$collectDate)
```

```
## [1] "Date"
```

```
# Check unique sample dates
```

```
unique(Litter$collectDate)
```



```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
# Check number of plots sampled  
unique(Litter$plotID)
```

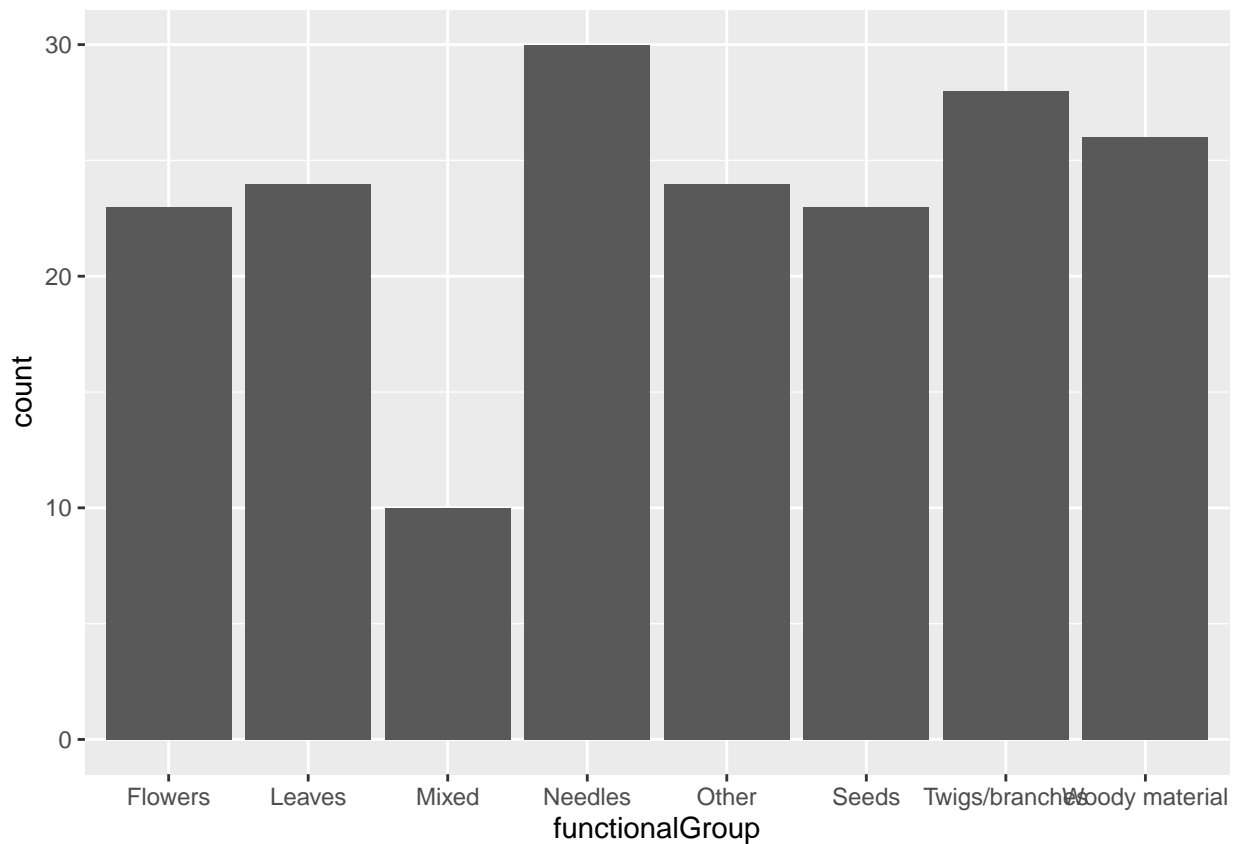
```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051  
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057  
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

```
# 12 plots
```

Answer:

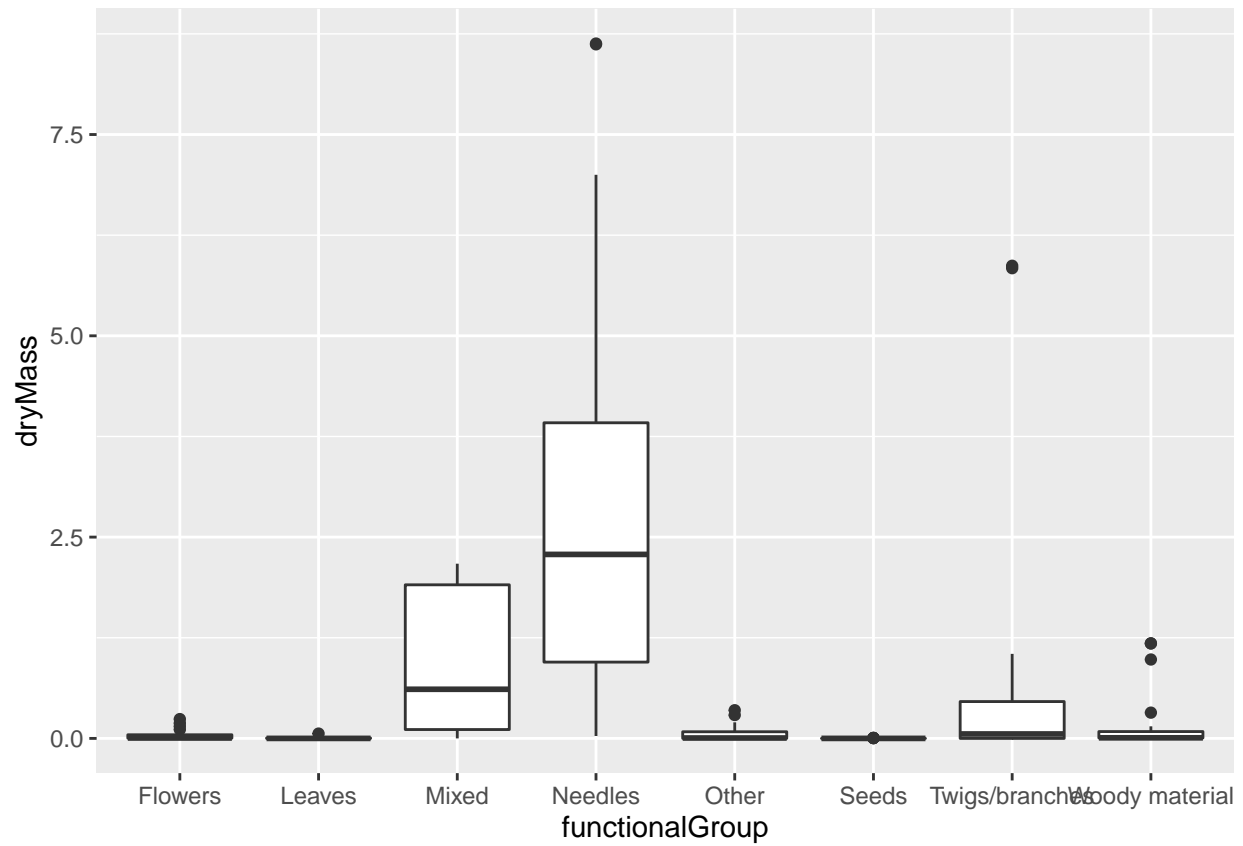
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
# Graph functional group counts  
ggplot(Litter) +  
  geom_bar(aes(x=functionalGroup))
```

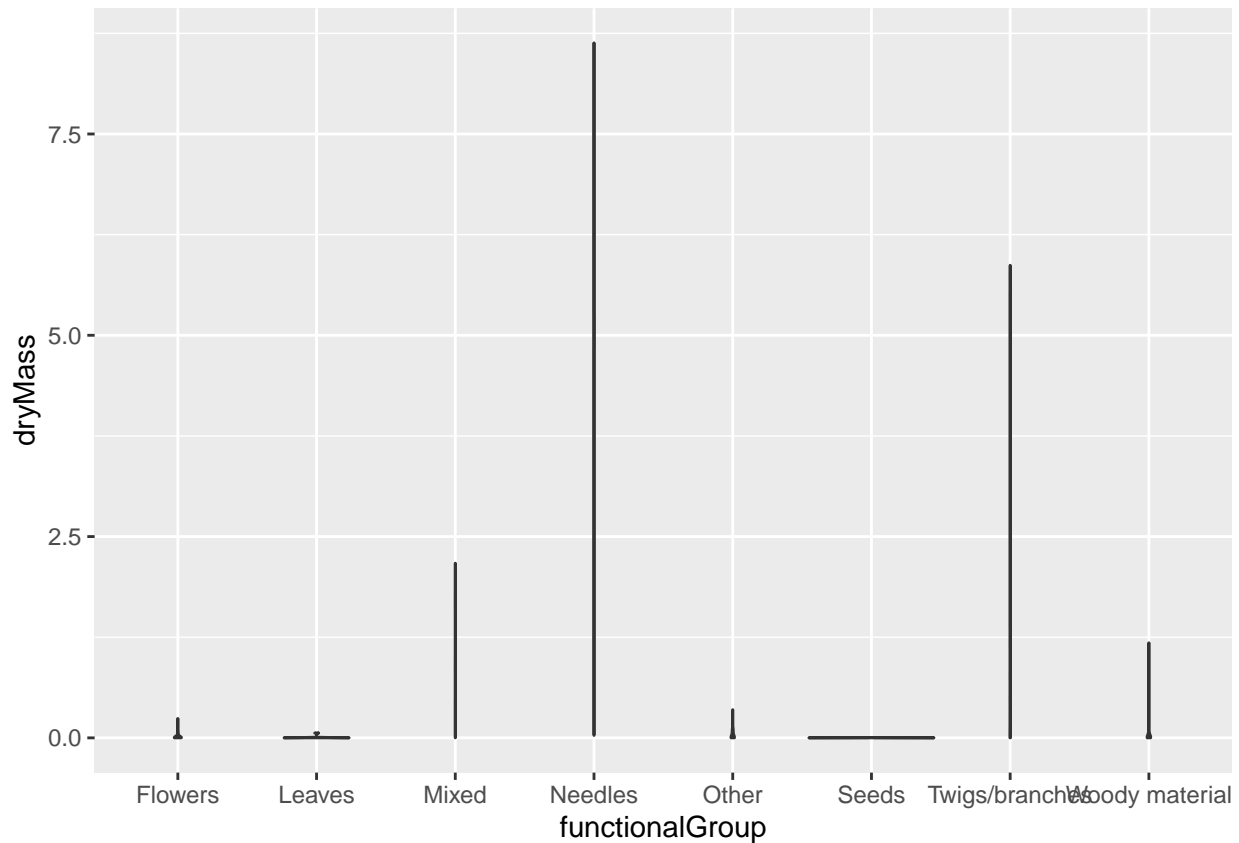


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
# Graph dry mass by functional group (boxplot)
ggplot(Litter) +
  geom_boxplot(aes(x=functionalGroup, y=dryMass))
```



```
# Graph dry mass by functional group (violin)
ggplot(Litter) +
  geom_violin(aes(x=functionalGroup, y=dryMass))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot more effectively shows the spread and density of each distribution. In this case, the violin's shape is flattened into 2 dimensions, making it very difficult to interpret or compare. I'm not sure why this occurred, beyond that perhaps eight groups was too many for the violin visualization to display properly.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles tend to have the highest amount of biomass, with "mixed" as a second. There appear to be a couple outliers with high amounts of trigs/branches.