R codes

17 00000
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Step 1: Install the R packages

```
if(!require("gwasrapidd")) install.packages("gwasrapidd",update = F,ask = F)
if(!require("stringr")) install.packages("stringr",update = F,ask = F)
if(!require("BiocManager")) install.packages("BiocManager",update = F,ask = F)
if(!require("data.table")) install.packages("data.table",update = F,ask = F)
if (!require("devtools")) install.packages("devtools",update = F,ask = F)
if (!require("MRPRESSO")) devtools::install_github("rondolab/MR-PRESSO")
if(!require("ggplot2")) install.packages("ggplot2",update = F,ask = F)
if(!require("ggsci")) install.packages("ggsci",update = F,ask = F)
if(!require("ggpubr")) install.packages("ggpubr",update = F,ask = F)
if(!require("circlize")) install.packages("circlize",update = F,ask = F)
if(!require("RColorBrewer")) install.packages("RColorBrewer",update = F,ask = F)
if (!require("TwoSampleMR")) devtools::install_github("MRCIEU/TwoSampleMR")
if (!require("TSMRhelper")) devtools::install_github("RightSZ/TSMRhelper")
if (!require("ComplexHeatmap")) BiocManager::install("ComplexHeatmap")
if(!require("dplyr")) install.packages("dplyr",update = F,ask = F)
if(!require("data.table")) install.packages("data.table",update = F,ask = F)
if(!require("forestploter")) install.packages("forestploter",update = F,ask = F)
devtools::install github("explodecomputer/plinkbinr")
```

Step 2: Gut microbiota on Osteoporosis

```
# Load R packages
rm(list = ls())
library(stringr)
library(data.table)
library(TwoSampleMR)
library(MRPRESSO)
library(dplyr)
library(ggsci)
library(ggplot2)
library(TSMRhelper)
library(forestploter)
library(grid)
library(stringr)
library(grDevices)
# Read the local outcome (Osteoporosis)
id<-""
outcome all<-fread("./finngen R10 M13 OSTEOPOROSIS.gz",data.table = F)
# Get full Gut microbiota IEU exposure id data
if(file.exists("ao.csv")){ao<-read.csv(file = "ao.csv")}else{
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(ao <- available outcomes(),</pre>
                                message
                                                                 function(c)
                                                                                       if
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
      error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
  write.csv(ao,file = "ao.csv")
}
ao<-read.csv("ao.csv")
id exp<-subset(ao,pmid==33462485)
id exp<-id exp[!str detect(id exp$trait,"unknown"),]
exp<-list()
i <- 1
for (ids in id exp[1:nrow(id exp),]$id) {
  print(ids)
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp clumped<-extract instruments(outcomes = ids,</pre>
```

```
p1 = 1e-05,
                                                                    clump = TRUE,
                                                                    r2 = 0.001,
                                                                    kb = 10000,
                                                                    access token
NULL),
                                                                function(c)
                                                                                      if
                                message
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
      error_to_next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
  if(is.null(exp clumped))next
  exp_clumped$exposure<-id_exp[id_exp$id==exp_clumped$id.exposure[1],]$trait
  exp[[i]]<-exp clumped
  i<-i+1
}
exp_clumped<-do.call(rbind,exp)</pre>
write.csv(exp clumped,file = "exp clumped.csv",row.names = F)
# Calculating the F-statistic
exp_clumped<-read.csv("exp_clumped.csv")
F cal<-function(beta,se){F.Statistics<-beta^2/se^2}
F.Statistics<-F cal(beta =exp clumped$beta.exposure,se =exp clumped$se.exposure)
exp clumped$F.Statistics <- F.Statistics
# Get outcome
if(id==""){
  snp<-unique(exp clumped$SNP)</pre>
  outcome<-outcome all[outcome all$rsids%in%snp,]
  outcome_dat<-format_data(outcome,
                         type = "outcome",
                         beta col = "beta",
                         se col = "sebeta",
                         effect allele col = "alt",
                         other_allele_col = "ref",
                         snp col = "rsids",
                         chr_col = "#chrom",
                         pos col = "pos",
                         pval col = "pval",
                         eaf col = "maf")
}else{
  while(TRUE){
```

```
message_to_next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(outcome dat
                                                    extract outcome data(snps
                                             <-
exp clumped$SNP,
                                                                      outcomes = id,
                                                                      proxies = F,
                                                                      access token =
NULL
    ),
    message =
                    function(c)
                                  if
                                      (stringr::str_detect(as.character(c),"Failed
                                                                                  to"))
message to next <<- FALSE)
      error to next <<- TRUE})
    if(message_to_next == TRUE&error_to_next == TRUE) { break }
  }
}
write.csv(outcome dat,file = "outcome.csv",row.names = F)
outcome dat<-read.csv("outcome.csv")
# Harmonise data
dat<-harmonise_data(exposure_dat = exp_clumped,outcome_dat = outcome_dat)
dat<-dat[dat$mr keep,]
write.csv(dat,file = "dat.csv",row.names = F)
# MR analysis
dat<-split(dat,dat$id.exposure)
library(parallel)
cl <- makeCluster(detectCores())</pre>
res<-parLapply(cl,dat,mr)
stopCluster(cl)
res<-do.call(rbind,res)
res<-generate_odds_ratios(res)
p<-p.adjust(res$pval, #
             method ="BH")
res$padjust<-p
write.csv(res,file = "res.csv",row.names = F)
res<-read.csv("res.csv")
dat<-do.call(rbind,dat)
# Heterogeneity analysis
heterogeneity<-mr heterogeneity(dat)
write.csv(heterogeneity,file = "heterogeneity.csv",row.names = F)
# Pleiotropy test
pleiotropy test<-mr pleiotropy test(dat)
```

```
write.csv(pleiotropy_test,file = "pleiotropy_test.csv",row.names = F)
# MR-PRESSO analysis
mp<-list()
mps<-unique(dat$id.exposure)
i=1
for (mpi in mps) {
  data<-dat[dat$id.exposure==mpi,]
  if(nrow(data)<4){next}
  # MRPRESSO
  mp[[i]]<-mr presso(BetaOutcome = "beta.outcome",
                       BetaExposure = "beta.exposure",
                       SdOutcome = "se.outcome",
                       SdExposure = "se.exposure",
                       OUTLIERtest = TRUE,
                       DISTORTIONtest = TRUE,
                       data = data,
                       NbDistribution = 1000,
                       SignifThreshold = 0.05)
  mp[[i]]$phe <- data$exposure[1]
  i=i+1
}
result mp all <- data.frame()
for (i in 1:length(mp)) {
  print(mp[[i]]$phe)
  result_mp <- mp[[i]]$`Main MR results`</pre>
  result_mp$RSSobs <- mp[[i]]$`MR-PRESSO results`$`Global Test`$RSSobs
  result mp$Pvalue <- mp[[i]]$`MR-PRESSO results`$`Global Test`$Pvalue
  result mp$phe <- mp[[i]]$phe
  result mp all <- rbind(result_mp_all,result_mp)</pre>
}
table(is.na(result mp all$`Causal Estimate`))
write.csv(result_mp_all,file = "MRPRESSO.csv",row.names = F)
# Reverse MR analysis
if(id==""){
  exp<-outcome all
  exp$pval<-as.numeric(exp$pval)
  exp<-subset(exp,pval<1e-05)
  exp<-format data(exp,
```

snp_col = "rsids",
chr_col = "#chrom",
pos col = "pos",

effect allele col = "alt",

```
other_allele_col = "ref",
                    eaf col = "maf",
                    pval_col = "pval",
                    beta col = "beta",
                    se col = "sebeta")
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp clumped<-clump data(exp),
                                                                                    if
                               message
                                                               function(c)
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
      error to next <<- TRUE})
    if(message_to_next == TRUE&error_to_next == TRUE) { break }
  }
}else{
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp_clumped<-extract_instruments(outcomes = id,p1 = 1e-
05,access token = NULL),
                                                                                    if
                               message
                                                               function(c)
(stringr::str_detect(as.character(c),"Failed to")) message to next <<- FALSE)
      error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
}
table(!((((exp_clumped$beta.exposure)^2)/((exp_clumped$se.exposure)^2))<10))
exp clumped
exp_clumped[!((((exp_clumped$beta.exposure)^2)/((exp_clumped$se.exposure)^2))<10)
,]
dat<-list()
for (i in 1:length(id exp$id)) {
  print(id_exp$id[i])
  while(TRUE){
    message_to_next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(outcome<-extract outcome data(exp clumped$SNP,
                                                               outcomes
                                                                                    =
id_exp$id[i],
                                                               proxies = F,
                                                               access token = NULL),
```

```
if
                                                                 function(c)
                                message
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
      error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  if(is.null(outcome))next
  dat[[i]]<-harmonise_data(exposure_dat = exp_clumped,outcome_dat = outcome)
}
dat<-do.call(rbind,dat)
dat<-dat[dat$mr keep,]
write.csv(dat,file = "re dat.csv",row.names = F)
re res<-mr(dat)
write.csv(re res,file = "re res.csv",row.names = F)
# Plotting
res id<-read.csv("./res.csv")%>%get sbeta res()%>%dplyr::filter(method=="Inverse
variance weighted",pval<0.05)%>%pull(id.exposure)
dat_total<-read.csv("./dat.csv")
for (idr in res id) {
  dat<-subset(dat_total,id.exposure==idr)
  scatter plot<-mr scatter plot(mr(dat),dat)[[1]]+
    scale color lancet()+
    scale fill lancet()+
    theme(axis.title.y = element text(size = 20))+
    theme bw(base size = 16)+ theme(
      plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm")
    )
  ggsave(plot=scatter plot,filename = paste0(idr," scatter plot plot.pdf"),device
"pdf",width = 10,height = 10)
  funnel plot<-
mr funnel plot(mr singlesnp(dat,all method=c("mr egger regression","mr weighted m
edian","mr_ivw","mr_simple_mode","mr_weighted_mode")))[[1]]+
    scale color lancet()+
    scale fill lancet()+
    theme(axis.title.y = element text(size = 20))+
    theme_bw(base_size = 16)+ theme(
      plot.margin = margin(0.5, 0.5, 0.5, 0.5, unit = "cm")
  ggsave(plot=funnel_plot,filename
                                    =
                                           paste0(idr," funnel plot plot.pdf"),device
"pdf", width = 10, height = 10)
  forest_plot<-mr_forest_plot(mr_singlesnp(dat))[[1]]+
    scale color lancet()+
```

```
scale fill lancet()+
    theme bw()+
    theme(legend.position = 'none',plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm"))
  ggsave(plot=forest_plot,filename =
                                           paste0(idr," forest plot plot.pdf"),device
"pdf", width = 10, height = 10)
  leaveoneout plot<-mr leaveoneout plot(mr leaveoneout(dat))[[1]]+
    scale color lancet()+
    scale fill lancet()+
    theme bw()+
    theme(legend.position = 'none',plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm"),
           axis.title.x = element text(size = 12))
  ggsave(plot=leaveoneout_plot,filename
paste0(idr," leaveoneout plot plot.pdf"),device = "pdf",width = 10,height = 10)
}
# Forest plot
res sign<-read.csv("./res.csv")%>%dplyr::filter(id.exposure%in%res id)
res sign$exposure<-str split(str split(res sign$exposure,regex("\\(|\\)"),simplify = T)[,2],"
id.", simplify = T)[,1]
res sign$`OR (95% CI)` <- sprintf("%.3f (%.3f - %.3f)",res sign$or, res sign$or lci95,
res_sign$or_uci95)
res sign$outcome[duplicated(res sign$outcome)]<-""
dt < -res sign[,c(3:6,9,12:14,15)]
dt$` ` <- paste(rep(" ", 30), collapse = " ")
dt$pval<-round(dt$pval,digits = 3)
dt$pval<-ifelse(dt$pval<0.001,"<0.001",dt$pval)
colnames(dt)[5]<-"italic(P)*-Value"
dt<-dt%>%dplyr::rename(Outcome=outcome,
                          `Trait`=exposure,
                          Method=method,
                          nSNP=nsnp,)
dt$Method<-ifelse(dt$Method=="Inverse variance weighted","IVW",dt$Method)
tm <- forest theme(base size = 8,colhead=list(fg params = list(parse=TRUE)))
p=forest(dt[,c(2:5,10,9)],
          est = dt or,
```

```
lower = dt\$or\_lci95, \\ upper = dt\$or\_uci95, \\ sizes = 0.6, \\ ci\_column = 5, \\ ref\_line = 1, \\ xlim = c(0,3), \\ ticks\_at = c(0,0.5,1,2,3), \\ theme = tm) \\ ggsave("GM forest plot.pdf",plot=p,device = "pdf",width = 8,height = 8)
```

```
### Step 3: Cyclic heat map
```

```
if(!require("gwasrapidd")) install.packages("gwasrapidd",update = F,ask = F)
if(!require("stringr")) install.packages("stringr",update = F,ask = F)
if(!require("data.table")) install.packages("data.table",update = F,ask = F)
if (!require("devtools")) { install.packages("devtools") } else {}
# devtools::install_github("rondolab/MR-PRESSO",force = TRUE)
library(TwoSampleMR)
library(MRPRESSO)
library(circlize)
library(stringr)
library(ComplexHeatmap)
library(RColorBrewer)
dat_total<-read.csv("./dat.csv")
i=1
result<-list()
for (file in unique(dat total$id.exposure)) {
  print(which(file==unique(dat total$id.exposure)))
  print(file)
  dat <- subset(dat total,id.exposure==file)
  dat <- dat[dat$mr_keep,]
  if(nrow(dat)<2)next
  res
           <-
                   generate odds ratios(mr res
                                                              mr(dat,method list
c("mr_ivw","mr_egger_regression","mr_weighted_median")))
  res hete <- mr heterogeneity(dat)
  res t <- cbind(merge(x
                               =
                                     res,y
                                                  res hete,by =
                                                                     "method",all.x
T),mr pleiotropy test(dat)[,c("egger intercept","pval")])
  if(length(dat$SNP)<4){res t$mp<-NA}else{
  res t$mp<-mr presso(BetaOutcome = "beta.outcome",
                       BetaExposure = "beta.exposure",
                       SdOutcome = "se.outcome",
                       SdExposure = "se.exposure",
                       OUTLIERtest = TRUE,
                       DISTORTIONtest = TRUE,
                       data = dat,
                       NbDistribution = 1000,
                       SignifThreshold
                                               0.05)$`MR-PRESSO
                                                                        results`$`Global
Test`$Pvalue
  result[[i]]<-res t
```

```
i=i+1
}
result all <- do.call(rbind,result)
write.csv(result all,file = "result all.csv")
result_all<-read.csv("result_all.csv")
IVW<-result all[result all$method=="Inverse variance weighted",][,c(6,10,13,22,24,25)]
colnames(IVW)<-c("bac","IVW P-Value","Odd Ratio","Q P-Value","Egger intercept P-
Value", "MRPRESSO Global Test P-Value")
IVW$`MRPRESSO Global Test P-Value`<-ifelse(IVW$`MRPRESSO Global Test P-
Value`=="<0.001",0.001,IVW$`MRPRESSO Global Test P-Value`)
IVW$`MRPRESSO Global Test P-Value`<-as.numeric(IVW$`MRPRESSO Global Test P-
Value')
MREGGER<-result all[result all$method=="MR Egger",][,c(6,10)]
colnames(MREGGER)<-c("bac","MR Egger P-Value")
WM<-result all[result all$method=="Weighted median",][,c(6,10)]
colnames(WM)<-c("bac","WM P-Value")
data<-merge(x=IVW,y=MREGGER,by = "bac")
data<-merge(x=data,y=WM,by = "bac")
rownames(data) <- data[,1]
data<-data[,-1]
data<-as.matrix(data)
rownames(data)<-str split(str split(rownames(data),regex("\\(|\\)"),simplify
                                                                                T)[,2],"
id.", simplify = T)[,1]
pdf(file = "1.pdf", width = 15, height = 15)
pal<-brewer.pal(11,'Spectral')
col_pval = colorRamp2(c(0, 0.05, 1), c("#9E0142", "#FFFFBF", "#5E4FA2"))
split bac<-factor(str split(rownames(data),"
                                               ",simplify
                                                                    T)[,1],levels
unique(str_split(rownames(data)," ",simplify = T)[,1]))
circos.clear()
circos.par(start.degree = 90, gap.degree = 5,gap.after = c(2,2,2,2,60),track.height =
0.1,clock.wise=F,circle.margin=0.75)
circos.heatmap.initialize(data,split = split bac, cluster = F)
circos.heatmap(data[,1], col = col_pval,track.height = 0.05)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[1]
    circos.text(CELL META$cell.xlim[2] + convert x(2, "mm"),
```

```
CELL META$ycenter, cn,
                 cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
  }
}, bg.border = NA)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  name <- rownames(data)[CELL META$subset]
  name <- name[CELL META$row order]
  circos.text(seg_along(name)-
0.5,CELL META$cell.ylim[2]+(convert_y(strwidth(name,units
                                                                               "inches"),
"inches"))/4+convert y(2,"mm"),name,cex = 0.5,facing = "reverse.clockwise")
, cell.padding = c(0.02, 0, 0.02, 0))
circos.heatmap(data[,6], col = col pval,bq.border = "black",bq.lwd = 1, bq.lty =
1, track.height = 0.05)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[6]
    circos.text(CELL_META$cell.xlim[2] + convert_x(2, "mm"),
                 CELL META$ycenter, cn,
                 cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
  }
}, bg.border = NA)
circos.heatmap(data[,7], col = col pval,bq.border = "black",bq.lwd = 1, bq.lty =
1,\text{track.height} = 0.05
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[7]
    circos.text(CELL META$cell.xlim[2] + convert x(2, "mm"),
                 CELL META$ycenter, cn,
                 cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
}, bg.border = NA)
circos.heatmap(data[,4], col = col pval,bg.border = "black",bg.lwd = 1, bg.lty =
1, track.height = 0.05)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[4]
    circos.text(CELL META$cell.xlim[2] + convert x(2, "mm"),
                 CELL META$ycenter, cn,
                  cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
  }
```

```
}, bg.border = NA)
circos.heatmap(data[,5], col = col pval,bg.border = "black",bg.lwd = 1, bg.lty =
1, track.height = 0.05)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL_META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[5]
    circos.text(CELL_META$cell.xlim[2] + convert_x(2, "mm"),
                  CELL META$ycenter, cn,
                  cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
  }
}, bg.border = NA)
circos.heatmap(data[,3], col = col pval,bq.border = "black",bq.lwd = 1, bq.lty =
1, track.height = 0.05)
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[3]
    circos.text(CELL META$cell.xlim[2] + convert x(2, "mm"),
                  CELL META$ycenter, cn,
                  cex = 0.5, adj = c(0, 0.5), facing = "inside", niceFacing = F)
  }
}, bg.border = NA)
row or = data[,2]
circos.track(ylim = range(row_or), panel.fun = function(x, y) {
  y = row or[CELL META$subset]
  y = y[CELL META$row order]
  circos.lines(CELL META$cell.xlim, c(1,1), lty = 2, col = "#7570B3")
  circos.points(seq along(y)-0.5, y, col = ifelse(y < 1, "#1B9E77", "#D95F02"),cex =
0.5, pch = 16)
, cell.padding = c(0.02, 0, 0.02, 0))
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  if(CELL META$sector.numeric.index == 1) { # the last sector
    cn = colnames(data)[2]
    circos.text(CELL_META$cell.xlim[2] + convert_x(2, "mm"),
                  CELL META$ycenter, cn,
                  cex = 0.5, adj = c(0, 0.5), facing = "inside",niceFacing = F)
  }
}, bg.border = NA)
circos.yaxis( side = "left",labels.cex = 0.5,at = c(0.7,1,1.3))
circos.track(track.index = get.current.track.index(), panel.fun = function(x, y) {
  circos.text(CELL META$xcenter, CELL META$cell.ylim[1] - convert y(3, "mm"),
```

```
CELL_META$sector.index,
facing = "bending.inside", cex = 0.5,
adj = c(0.5, 0), niceFacing = TRUE)
}, bg.border = NA)

pval = Legend(title = "P.Value", col_fun = col_pval,at = c(0,0.05,0.5,1))
or = Legend(labels = c("Risk factor","Protective factor"), title = "Odd Ratio", legend_gp = gpar(fill=c("#D95F02", "#1B9E77")))
all<-packLegend(pval,or)
grid.draw(all)
dev.off()
```

Step 4: Plasma metabolites on Osteoporosis

```
# Load R packages
rm(list = ls())
library(dplyr)
library(MRPRESSO)
library(TwoSampleMR)
library(dplyr)
library(ggsci)
library(data.table)
library(ggplot2)
library(ggpubr)
library(ggview)
library(plotly)
library(TSMRhelper)
library(forestploter)
library(grid)
library(stringr)
library(grDevices)
library(parallel)
# Read the local outcome (Osteoporosis)
id<-""
outcome all<-fread("./finngen R10 M13 OSTEOPOROSIS.gz ",data.table = F)
# Get full Plasma metabolites IEU exposure id data
ao<-read.csv("ao.csv")
id exp <-subset(ao,pmid== 36635386)
id exp<-id exp[!str detect(id exp$trait,"unknown"),]
exp<-list()
i <- 1
for (ids in id_exp[1:nrow(id_exp),]$id) {
  print(ids)
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp_clumped<-extract_instruments(outcomes = ids,</pre>
                                                                     p1 = 1e-05.
                                                                     clump = TRUE,
                                                                     r2 = 0.1
                                                                     kb = 500,
                                                                     access token
NULL),
                                                                                       if
                                message
(stringr::str_detect(as.character(c),"Failed to")) message to next <<- FALSE)
```

```
error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
  if(is.null(exp_clumped))next
  exp clumped$exposure<-id exp[id exp$id==exp clumped$id.exposure[1],]$trait
  exp[[i]]<-exp_clumped
  i<-i+1
}
exp clumped<-do.call(rbind,exp)</pre>
write.csv(exp clumped,file = 1400 exp clumped 1e05 0.1 500.csv ",row.names = F)
# Calculating the F-statistic
exp clumped<-read.csv("1400 exp clumped 1e05 0.1 500.csv")
F cal<-function(beta,se){F.Statistics<-beta^2/se^2}
F.Statistics<-F cal(beta =exp clumped$beta.exposure,se =exp clumped$se.exposure)
exp clumped$F.Statistics <- F.Statistics
# Get outcome
if(id==""){
  snp<-unique(exp clumped$SNP)</pre>
  outcome<-outcome_all[outcome_all$rsids%in%snp,]
  outcome dat<-format data(outcome,
                             type = "outcome",
                             beta col = "beta",
                             se col = "sebeta",
                             effect allele col = "alt",
                             other allele col = "ref",
                             snp_col = "rsids",
                             chr col = "#chrom",
                             pos_col = "pos",
                             pval col = "pval",
                             eaf col = "af alt")
}else{
  #
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(outcome dat <-
                                                    extract outcome data(snps
unique(exp_clumped$SNP),
                                                                      outcomes = id,
                                                                      proxies = F,
                                                                      access token =
NULL,
```

```
splitsize
                                                                                     =
50000
    ),
                    function(c)
                                       (stringr::str_detect(as.character(c),"Failed
    message =
                                  if
                                                                                  to"))
message_to_next <<- FALSE)
      error_to_next <<- TRUE})
    if(message_to_next == TRUE&error_to_next == TRUE) { break }
  }
}
# Harmonise data
dat <- harmonise data(exposure dat = exp clumped,outcome dat = outcome dat)
dat<-dat[dat$mr_keep,]
write.csv(dat,file = "1400_dat.csv",row.names = F)
# MR analysis
dat<-split(dat,dat$id.exposure)
library(parallel)
cl <- makeCluster(detectCores())</pre>
res<-parLapply(cl,dat,mr)
stopCluster(cl)
res<-do.call(rbind,res)
res<-generate odds ratios(res)
write.csv(res,file = "1400_res.csv",row.names = F)
dat<-do.call(rbind,dat)
# Heterogeneity analysis
heterogeneity<-mr heterogeneity(dat)
write.csv(heterogeneity,file = "1400_heterogeneity.csv",row.names = F)
# Pleiotropy test
pleiotropy test<-mr pleiotropy test(dat)
write.csv(pleiotropy_test,file = "1400_pleiotropy_test.csv",row.names = F)
# MR-PRESSO analysis
mps<-res%>%get sbeta res()%>%dplyr::filter(method=="Inverse
                                                                              variance
weighted",pval<0.05)%>%pull(id.exposure)
mp<-list()
i=1
for (mpi in mps) {
  data<-dat[dat$id.exposure==mpi,]
  if(nrow(data)<4){next}
  # MRPRESSO
```

```
mp[[i]]<-mr_presso(BetaOutcome = "beta.outcome",
                       BetaExposure = "beta.exposure",
                       SdOutcome = "se.outcome",
                       SdExposure = "se.exposure",
                       OUTLIERtest = TRUE,
                       DISTORTIONtest = TRUE,
                       data = data,
                       NbDistribution = 1000,
                       SignifThreshold = 0.05)
  mp[[i]]$phe <- data$exposure[1]
  i=i+1
}
result_mp_all<- data.frame()
for (i in 1:length(mp)) {
  print(mp[[i]]$phe)
  result mp <- mp[[i]]$`Main MR results`
  result_mp$RSSobs <- mp[[i]]$`MR-PRESSO results`$`Global Test`$RSSobs
  result mp$Pvalue <- mp[[i]]$`MR-PRESSO results`$`Global Test`$Pvalue
  result mp$phe <- mp[[i]]$phe
  result_mp_all <- rbind(result_mp_all,result_mp)</pre>
}
table(is.na(result_mp_all$`Causal Estimate`))
write.csv(result mp all,file = "1400 MRPRESSO.csv")
# Reverse MR analysis
if(id==""){
  exp<-outcome_all
  exp$pval<-as.numeric(exp$pval)
  exp<-subset(exp,pval<1e-05)
  exp<-format data(exp,
                    snp_col = "rsids",
                     chr col = "#chrom",
                    pos col = "pos",
                     effect allele col = "alt",
                    other allele col = "ref",
                    eaf_col = "af_alt",
                     pval col = "pval",
                    beta_col = "beta",
                    se col = "sebeta")
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp_clumped<-clump_data(exp),</pre>
                                                                                      if
                                message
                                                                function(c)
```

```
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)</pre>
      error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
}else{
  while(TRUE){
    message to next <<- TRUE
    error to next <<- FALSE
    try({withCallingHandlers(exp clumped<-extract instruments(outcomes = id,p1 = 1e-
05,access token = NULL),
                                                                 function(c)
                                                                                       if
                                message
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)</pre>
      error to next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
}
table(!((((exp_clumped$beta.exposure)^2)/((exp_clumped$se.exposure)^2))<10))
exp clumped
exp clumped[!((((exp clumped$beta.exposure)^2)/((exp clumped$se.exposure)^2))<10)
,]
idTotrait<-read.csv("./idTotrait.csv")
total data<-dir("./data1400/")
dat<-list()
for (i in 1:length(total_data)) {
  print(total data[i])
  outcome dat<-fread(paste0("./data1400/",total data[i]),data.table = F)
  outcome dat$p value<-as.numeric(outcome dat$p value)
  outcome dat<-subset(outcome dat, `variant id` %in%unique(exp clumped$SNP))
  outcome_dat$id <- str_split(total_data[i],"_buildGRCh38.tsv.gz",simplify = T)[1]
  outcome dat$phe<-idTotrait[idTotrait$ID==outcome dat$id[1],]$Trait
  outcome dat<-format data(outcome dat,
                              type = "outcome",
                              snp col = "variant id",
                              beta col = "beta",
                              se col = "standard error",
                              eaf_col = "effect_allele_frequency",
                              effect allele col = "effect allele",
                              other allele col = "other allele",
                              pval_col = "p_value",
                              chr col = "chromosome",
```

```
pos_col = "base_pair_location",
                              id col = "id",
                              phenotype col = "phe")
  dat[[i]]<-harmonise data(exposure dat = exp clumped,outcome dat = outcome dat)
}
dat<-do.call(rbind,dat)
dat<-dat[dat$mr_keep,]
write.csv(dat,file = "C 1400 re dat.csv",row.names = F)
re res<-mr(dat)
write.csv(re res,file = "C 1400 re res.csv",row.names = F)
# Plotting
res id<-
read.csv("./1400 res.csv")%>%get sbeta res()%>%dplyr::filter(method=="Inverse
variance weighted",pval<0.05)%>%pull(id.exposure)
dat total<-read.csv("./1400 dat.csv")
for (idr in res id) {
  dat<-subset(dat total,id.exposure==idr)
  scatter plot<-mr scatter plot(mr(dat),dat)[[1]]+
    scale_color_lancet()+
    scale fill lancet()+
    theme(axis.title.y = element text(size = 20))+
    theme bw(base size = 16)+ theme(
      plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm")
  ggsave(plot=scatter plot,filename = paste0(idr," scatter plot plot.pdf"),device
"pdf",width = 10,height = 10)
  funnel plot<-
mr_funnel_plot(mr_singlesnp(dat,all_method=c("mr_egger_regression","mr_weighted_m
edian","mr ivw","mr simple mode","mr weighted mode")))[[1]]+
    scale color lancet()+
    scale fill lancet()+
    theme(axis.title.y = element text(size = 20))+
    theme_bw(base_size = 16)+ theme(
      plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm")
    )
                                          paste0(idr," funnel plot plot.pdf"),device
  ggsave(plot=funnel_plot,filename
                                    =
"pdf", width = 10, height = 10)
  forest plot<-mr forest plot(mr singlesnp(dat))[[1]]+
    scale color lancet()+
    scale fill lancet()+
    theme bw()+
```

```
theme(legend.position = 'none',plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm"))
  ggsave(plot=forest_plot,filename
                                           paste0(idr,"_forest_plot_plot.pdf"),device
"pdf", width = 10, height = 10)
  leaveoneout plot<-mr leaveoneout plot(mr leaveoneout(dat))[[1]]+
    scale_color_lancet()+
    scale fill lancet()+
    theme bw()+
    theme(legend.position = 'none',plot.margin = margin(0.5,0.5,0.5,0.5, unit = "cm"),
           axis.title.x = element_text(size = 12))
  ggsave(plot=leaveoneout plot,filename
                                                                                       =
paste0(idr," leaveoneout plot plot.pdf"),device = "pdf",width = 10,height = 10)
}
# Forest plot
res sign<-read.csv("./1400 res.csv")%>%dplyr::filter(id.exposure%in%res id)
res sign$`OR (95% CI)` <- sprintf("%.3f (%.3f - %.3f)",res sign$or, res sign$or lci95,
res sign$or uci95)
res sign$outcome[duplicated(res sign$outcome)]<-""
dt < -res sign[,c(3:6,9,12:14,15)]
dt$` ` <- paste(rep(" ", 30), collapse = " ")
dt$pval<-round(dt$pval,digits = 3)
dt$pval<-ifelse(dt$pval<0.001,"<0.001",dt$pval)
colnames(dt)[5]<-"italic(P)*-Value"
dt<-dt%>%dplyr::rename(Outcome=outcome,
                          `Trait`=exposure,
                         Method=method,
                         nSNP=nsnp,)
dt$Method<-ifelse(dt$Method=="Inverse variance weighted","IVW",dt$Method)
dt<-dt[dt$Method=="IVW", ]
tm <- forest theme(base size = 8,colhead=list(fg params = list(parse=TRUE)))
p=forest(dt[,c(2:5,10,9)],
          est = dt or,
          lower = dt$or lci95,
          upper = dt$or uci95,
```

```
sizes = 0.6,
ci\_column = 5,
ref\_line = 1,
xlim = c(0.5,1.5),
ticks\_at = c(0.5,0.75,1,1.25,1.5),
theme = tm)
ggsave("1400 forest plot.pdf",plot=p,device = "pdf",width = 20,height = 20)
```

```
### Step 5: GM on Plasma metabolites
```

```
# Load R packages
rm(list = ls())
library(TSMRhelper)
library(dplyr)
# Read data
gut res<-read.csv("./res.csv")%>%get sbeta res()%>%dplyr::filter(method=="Inverse
variance weighted",pval<0.05)
res1400<-
read.csv("./1400_res.csv")%>%get_sbeta_res()%>%dplyr::filter(method=="Inverse
variance weighted",pval<0.05)
gut exp<-
read.csv("exp clumped.csv")%>%dplyr::filter(id.exposure%in%gut res$id.exposure)
idTotrait<-read.csv("./idTotrait.csv")
ids<-paste0(res1400$id.exposure,"_buildGRCh38.tsv.gz")
dat<-list()
for (i in 1:length(ids)) {
  print(ids[i])
  outcome dat<-fread(paste0("./data1400/",ids[i]),data.table = F)
  outcome dat$p value<-as.numeric(outcome dat$p value)
# Screening for SNPs
outcome dat<-subset(outcome dat, variant id '%in%unique(gut exp$SNP))
  outcome dat$id <- str split(ids[i]," buildGRCh38.tsv.gz",simplify = T)[1]
  outcome dat$phe<-idTotrait[idTotrait$ID==outcome dat$id[1],]$Trait
  outcome dat<-format data(outcome dat,
                              type = "outcome",
                              snp_col = "variant_id",
                              beta col = "beta",
                              se col = "standard error",
                              eaf col = "effect allele frequency",
                              effect allele col = "effect allele",
                              other allele col = "other allele",
                              pval col = "p value",
                              chr col = "chromosome",
                              pos col = "base pair location",
                              id col = "id",
                              phenotype_col = "phe")
  dat[[i]]<-harmonise data(exposure dat = gut exp,outcome dat = outcome dat)
}
dat<-do.call(rbind,dat)
dat<-dat[dat$mr keep,]
write.csv(dat,file = "gutto1400_dat.csv",row.names = F)
```

```
dat<-split(dat,dat$id.exposure)
library(parallel)
cl <- makeCluster(detectCores())
res<-parLapply(cl,dat,mr)
stopCluster(cl)
res<-do.call(rbind,res)
write.csv(res,file = "gutto1400_res.csv",row.names = F)

# Possible mediation
id_final<-res%>%dplyr::filter(method=="Inverse variance weighted",pval<0.05)

for (a in 1:nrow(id_final)) {
    print(paste0("exposure: ",id_final[a,]$id.exposure," mediation: ",id_final[a,]$id.outcome))
}</pre>
```

Step 6: Multivariate MR analysis

Load R packages

```
library(TwoSampleMR)
library(data.table)
library(dplyr)
library(RMediation)
library(stringr)
```

To run multiple variables separately, please fill in the corresponding id numbers separately

GM id example

ids<-"ebi-a-GCST90016925"

Plasma metabolite id example

id1400<-"GCST90200756"

```
# outcome
out id<-""
out dat<-fread("./finngen R10 M13 OSTEOPOROSIS.gz",data.table = F)
idTotrait<-read.csv("./idTotrait.csv")</pre>
res_all<-list()
i=1
for (id in ids) {
exp1<-fread(paste0("./data1400/",id1400,"_buildGRCh38.tsv.gz"),data.table = F)
exp1$`p value`<-as.numeric(exp1$`p value`)</pre>
exp1$id <- id1400
exp1$phe<-idTotrait[idTotrait$ID==exp1$id[1],]$Trait
exp1<-format data(exp1,
             type = "outcome",
             snp col = "variant id",
             beta col = "beta",
             se col = "standard error",
             eaf col = "effect allele frequency",
             effect allele col = "effect allele",
             other_allele_col = "other_allele",
              pval col = "p value",
             chr_col = "chromosome",
              pos col = "base pair location",
             id col = "id",
              phenotype col = "phe")
```

```
exp1 1 <- subset(exp1, pval.outcome < 1e-05)
out <- ieugwasr::ld clump(dplyr::tibble(rsid=exp1 1$SNP, pval=exp1 1$pval.outcome,id
= exp1_1$id.outcome),
                           clump kb = 10000,
                            clump r2 = 0.001,
                            clump_p = 1e-05,
                            plink bin = plinkbinr::get plink exe(),
                            bfile = "F:/1kg.v3/EUR",
                            pop = "EUR")
keep <- paste(exp1 1$SNP,exp1 1$id.outcome) %in% paste(out$rsid,out$id)
exp1 1 <- exp1 1[keep,]
exp1 1<-format data(exp1 1,
                   type = "exposure",
                   snp col = "SNP",
                   chr_col = "chr.outcome",
                   pos col = "pos.outcome",
                   beta col = "beta.outcome",
                   se col = "se.outcome",
                   pval_col = "pval.outcome",
                   samplesize_col = "samplesize.outcome",
                   id col = "id.outcome",
                   effect_allele_col = "effect_allele.outcome",
                   other allele col = "other allele.outcome",
                   eaf_col = "eaf.outcome",
                   phenotype col = "outcome")
while(TRUE){
  message_to_next <<- TRUE
  error to next <<- FALSE
  try({withCallingHandlers(exp2<-extract instruments(outcomes
                                                                     id,p1
                                                                                  1e-
05,access token = NULL),
                                                              function(c)
                                                                                    if
                             message
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
    error to next <<- TRUE})
  if(message to next == TRUE&error to next == TRUE) { break }
}
exp2$chr.exposure<-as.numeric(exp2$chr.exposure)
exposure dat <- dplyr::bind rows(list(exp1 1,exp2))
id exposure <- unique(exposure dat$id.exposure)
temp <- exposure dat
temp$id.exposure <- 1
temp <- temp[order(temp$pval.exposure, decreasing = FALSE), ]
temp <- subset(temp, !duplicated(SNP))
out <- ieugwasr::ld clump(dplyr::tibble(rsid=temp$SNP, pval=temp$pval.exposure,id =
temp$id.exposure),
```

```
clump kb = 10000,
                           clump r2 = 0.001,
                           clump p = 1e-05,
                           plink_bin = plinkbinr::get_plink_exe(),
                            bfile = "F:/1kg.v3/EUR",
                            pop = "EUR")
keep <- paste(temp$SNP,temp$id.exposure) %in% paste(out$rsid,out$id)
temp <- temp[keep,]
exposure dat <- subset(exposure dat, SNP %in% temp$SNP)
# Retrieve snp
exp1 2<-subset(exp1, SNP %in% exposure dat$SNP)
while(TRUE){
  message to next <<- TRUE
 error to next <<- FALSE
  try({withCallingHandlers(exp2 2<-extract outcome data(snps
exposure dat$SNP,outcomes = id,proxies = F,access token = NULL),
                            message
                                                             function(c)
                                                                                   if
(stringr::str_detect(as.character(c),"Failed to")) message_to_next <<- FALSE)
    error to next <<- TRUE})
 if(message to next == TRUE&error to next == TRUE) { break }
}
exp1 2<-exp1 2%>%dplyr::rename(
 chr=`chr.outcome`,
 pos='pos.outcome'
)
exp2 2$chr<-as.numeric(exp2 2$chr)
exp2 2$pos<-as.numeric(exp2 2$pos)
d1<-dplyr::bind_rows(list(exp1_2,exp2_2))
d1 <- subset(d1, mr keep.outcome)
d2 <- subset(d1, id.outcome != id exposure[1])
d1 <- convert outcome to exposure(subset(d1, id.outcome ==
                                              id exposure[1]))
d <- harmonise data(d1, d2)
tab <- table(d$SNP)
keepsnps <- names(tab)[tab == length(id exposure) - 1]
d <- subset(d, SNP %in% keepsnps)
dh1 <- subset(d, id.outcome == id.outcome[1], select = c(SNP,
                                                             exposure,
                                                                        id.exposure,
effect allele.exposure, other allele.exposure,
                                                             eaf.exposure,
beta.exposure, se.exposure, pval.exposure))
dh2 <- subset(d, select = c(SNP, outcome, id.outcome, effect allele.outcome,
                              other allele.outcome,
                                                      eaf.outcome,
                                                                       beta.outcome,
```

```
se.outcome,
                               pval.outcome))
names(dh2) <- gsub("outcome", "exposure", names(dh2))</pre>
exp_dat <- rbind(dh1, dh2)
if(out id==""){
  out_dat<-subset(out_dat,rsids%in%exp_dat$SNP)
  out dat<-format data(out dat,
                     type = "outcome",
                     beta_col = "beta",
                     se col = "sebeta",
                     effect_allele_col = "alt",
                     other allele col = "ref",
                     snp col = "rsids",
                     chr_col = "#chrom",
                     pos_col = "pos",
                     pval col = "pval",
                     eaf col = "af alt")
}else{
  while(TRUE){
    message_to_next <<- TRUE
    error_to_next <<- FALSE
    try({withCallingHandlers(out_dat <- extract_outcome_data(snps = exp_dat$SNP,</pre>
                                                                        outcomes
                                                                                      =
out_id,
                                                                        proxies = F,
                                                                        access_token =
NULL
    ),
    message =
                     function(c)
                                  if
                                       (stringr::str_detect(as.character(c),"Failed
                                                                                   to"))
message to next <<- FALSE)
      error_to_next <<- TRUE})
    if(message to next == TRUE&error to next == TRUE) { break }
  }
}
mvdat <- mv_harmonise_data(exposure_dat = exp_dat, outcome_dat = out_dat)
res <- mv_multiple(mvdat,pval_threshold = 1e-05)
res_all[[i]]<-res[["result"]]
i=i+1
```

```
}
res all<-do.call(rbind,res all)
if(res all[!str detect(res all$exposure,"Gut
                                                                           microbiota
abundance"), | $pval<0.05)print("mediated p-value adjusted to be less than 0.05")
write.csv(res all,file = "res all.csv")
# Getting outcome data for the same exposure
gut<-read.csv("./res.csv")%>%dplyr::filter(id.exposure==id,method=="Inverse
                                                                             variance
weighted")
# Getting ending data from the same mediator
res1400<-
read.csv("./1400_res.csv")%>%dplyr::filter(id.exposure==id1400,method=="Inverse
variance weighted")
# Getting the outcome data exposed to the mediator
gutto1400<-
read.csv("./gutto1400 res.csv")%>%dplyr::filter(id.exposure==id,id.outcome==id1400,me
thod=="Inverse variance weighted")
# Calculating the mediating effect
EM beta<- gutto1400$b
EM se<- gutto1400$se
MO_beta<-res_all[!str_detect(res_all$exposure,"Gut microbiota abundance"),]$b
MO_se<-res_all[!str_detect(res_all$exposure,"Gut microbiota abundance"),]$se
# Delta
product method Delta <- function(EM beta, EM se, MO beta, MO se, verbose=F){
  # method 1
  # INDIRECT = TOTAL (exposure -> mediator) x TOTAL (mediator -> outcome)
  # method 2
  # INDIRECT = TOTAL (exposure -> mediator) x DIRECT (of mediator, mvmr)
  EO <- EM beta * MO beta
  if (verbose) {print(paste("Indirect effect = ", round(EM beta, 2)," x ", round(MO beta, 2),
```

```
" = ", round(EO, 3)))}
  Cls = medci(EM_beta, MO_beta, EM_se, MO_se, type="dop")
  df <-data.frame(b = EO,
                   se = CIs$SE,
                   lo_ci = Cls[["95% Cl"]][1],
                   up_ci= Cls[["95% Cl"]][2])
  #OR
  df$or
               <- exp(df$b)
  df$or_lci95 <- exp(df$lo_ci)
  df$or_uci95 <- exp(df$up_ci)
  df<-round(df,3)
  return(df)
}
#INDIRECT = TOTAL (exposure -> mediator) x DIRECT (of mediator, mvmr)
product_method_Delta(EM_beta, EM_se, MO_beta, MO_se)
```