





PixelMap: An Application for Flexible Electrode Selection on Neuropixels Probes

Maxime Beau ^{1,2}, Christian Tabedzki ¹, and Carlos D. Brody ^{1,2}

¹ Princeton Neuroscience Institute, Princeton University, USA ² Howard Hughes Medical Institute, USA
 Corresponding author

DOI: [10.xxxxxx/draft](https://doi.org/10.xxxxxx/draft)

Software

- [Review](#) 
- [Repository](#) 
- [Archive](#) 

Editor: [Open Journals](#) 

Reviewers:

- [@openjournals](#)

Submitted: 01 January 1970

Published: unpublished

License

Authors of papers retain copyright and release the work under a Creative Commons Attribution 4.0 International License ([CC BY 4.0](#)).

Abstract

PixelMap is a browser-based application for creating custom channelmaps for Neuropixels probes that respects electrode wiring constraints. Neuropixels probes, widely used for high-density neural recordings, have more physical electrodes than can be used for simultaneous recording because they contain fewer analogue-to-digital converters (ADCs) than data lines. Each ADC is hard-wired to several electrodes, creating complex interdependencies where selecting one electrode makes others unavailable. PixelMap provides an installation-free, browser-based interface for researchers to design arbitrary recording configurations that meet their experimental requirements while satisfying these hardware constraints. The tool generates IMRO (IMec Read Out) files compatible with SpikeGLX, the most common acquisition software for Neuropixels recordings.

Statement of need

Neuropixels probes have revolutionised systems neuroscience by enabling simultaneous recordings from hundreds of neurons across multiple brain regions at any depth (Beau et al., 2021, 2025; Bondy et al., 2024; Jun et al., 2017; Steinmetz et al., 2021; Ye et al., 2025). However, configuring these probes presents challenges. Limited by the number of integrated analogue-to-digital converters (ADCs), Neuropixels probes contain 960–5120 electrodes but can only record from 384–1536 channels simultaneously (Table 1). Users must therefore select a subset of electrodes to activate for each recording, a “channelmap”. Researchers often need to create custom channelmaps to target specific brain regions, and sometimes must adjust them rapidly based on feedback from ongoing recordings. Because the electrode-to-ADC wiring follows complex, probe version-dependent patterns, manual channelmap design is error-prone and time-consuming.

SpikeGLX is the most common acquisition software for Neuropixels recordings and uses the .imro file format to encode channelmaps. While SpikeGLX provides tools to edit channelmaps, it requires a desktop app, comes with limited preset channelmaps, and does not easily allow selection of fully arbitrary electrode geometries.

PixelMap addresses these needs by:

1. **Being available on any machine installation-free:** The tool is available both as a web application at <https://pixelmap.pni.princeton.edu> and as a Python package.
2. **Visualising wiring constraints interactively:** When users select electrodes, the interface immediately shows which other electrodes become unavailable (marked in black) due to shared ADC lines, preventing invalid configurations.
3. **Supporting arbitrary electrode geometries:** Users can select electrodes by choosing from common preset geometries, entering electrode ranges as text for reproducibility, directly

clicking or dragging on the probe visualization, or loading pre-existing .imro files. These four selection methods are fully interoperable and can be combined. For instance, a SpikeGLX .imro file can be loaded as a starting point, and selection boxes used to further refine the channelmap geometry.

Probe Version	Physical Channels	Simultaneously Recordable Channels
Neuropixels 1.0	960	384
Neuropixels 2.0 (single shank)	1,280	384
Neuropixels 2.0 (4-shank)	5,120 (1,280 per shank)	384
Neuropixels 2.0 Quad Base	5,120 (1,280 per shank)	1,536

Table 1: Number of physical and simultaneously addressable electrodes across Neuropixels probe versions.

Software Design

PixelMap is implemented in Python using HoloViz Panel (Yang et al., 2022) for the web interface, providing an interactive and responsive user experience. The software architecture consists of three main components.

First, the **wiring maps** at ./wiring_maps/*.csv are hand-built CSV files describing the electrode-to-ADC mappings for each supported probe type. They were adapted from files provided by IMEC (Neuropixels manufacturer - downloadable [here](#)).

Second, the **core logic** at ./backend.py implements the constraint-checking algorithms that validate electrode selections against probe-specific wiring maps. This handles the complex mapping between physical electrodes and ADC channels for different probe types (Neuropixels 1.0, 2.0 single-shank, and 2.0 four-shank so far). Hash tables (Python dictionaries) are used to query incompatible electrode pairs with O(1) complexity and improve performance.

Finally, the **graphical user interface** at ./gui/gui.py was built with HoloViz Panel. The interface provides real-time visualisation of the probe layout with electrode colour-coded based on their selection state (available in grey, selected in red, or unavailable in black). The interface supports the abovementioned four selection modes, including Bokeh-based interactive click-selection and box-selection to select or deselect electrodes. User interactions trigger immediate recalculation of available electrodes based on the current selection state. This design ensures users receive instant feedback about constraint violations, preventing invalid configurations before file generation.

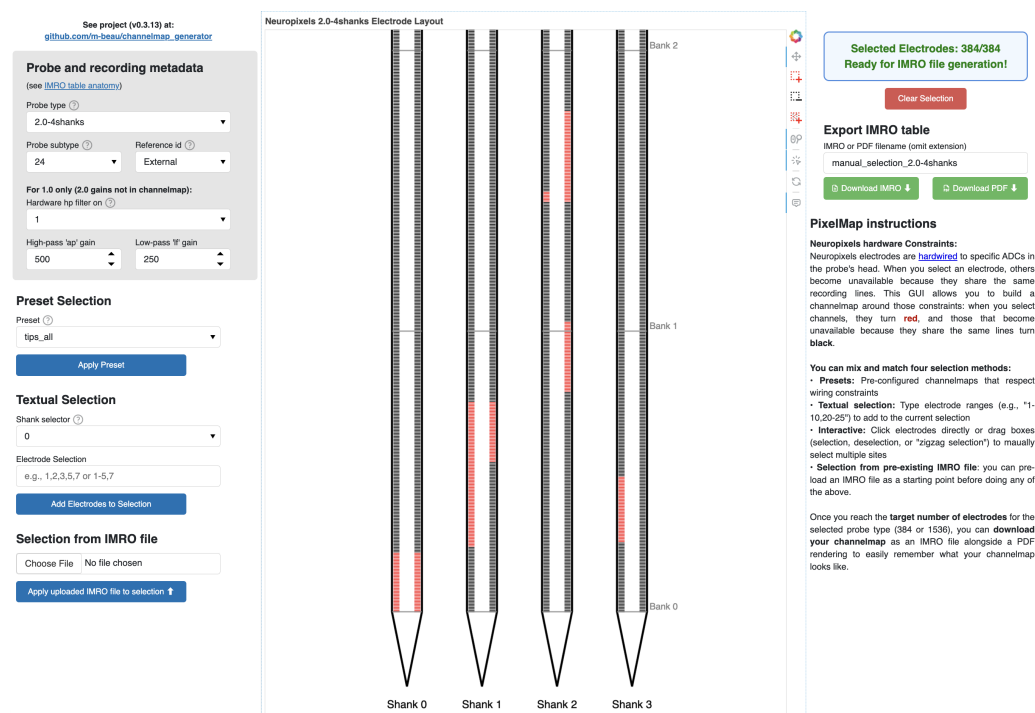


Figure 1: PixelMap's browser-based graphical user interface.

Center: Main panel featuring the probe's physical layout with one or four shanks that exhibit the 960 (Neuropixels 1.0) or 1,280 (Neuropixels 2.0) physical electrodes/shank to be selected. Electrodes available for selection are light grey, selected electrodes turn red, and electrodes that become unavailable due to hardware wiring constraints turn black. In this example, 384 electrodes have been selected (matching the maximum simultaneous recording capacity), with a distributed pattern across multiple banks, illustrating that PixelMap allows selection of arbitrary channelmap geometries.

Left: panel to input probe metadata (also part of .imro files) as well as three methods of electrode selection: preset geometries, manual textual input of electrode ranges, and pre-loading an existing .imro file. These three methods of electrode selection can be mixed together with an interactive click-and-drag box selector and deselector.

Right: electrode status indicator that turns green to confirm the selection is complete and is ready for IMRO file generation. Users can export their configuration via the "Download IMRO" button for direct use in SpikeGLX or save a PDF visualisation to easily remember the geometry of the corresponding .imro file in the future. Below the status indicator are PixelMap's instructions.

Installation and Usage

PixelMap can be used through:

1. **Web application:** Available at <https://pixelmap.pni.princeton.edu> for immediate use without installation.
2. **Local installation:** Via pip (pip install .) or uv (uv run cmap_gui) from the cloned GitHub repository.
3. **Docker container:** Users can download the image used for the website and run the container locally.
4. **Programmatic API:** Python scripts can directly call generate_imro_channelmap() for batch processing or integration into analysis pipelines.

For more details, see the project repository at https://github.com/m-beau/channelmap_generator.

The software includes an automated test suite with 41 tests covering hardware constraint validation, all preset configurations, IMRO file generation for all supported probe types, and

end-to-end workflows. Tests run automatically via GitHub Actions continuous integration on every code change, ensuring software reliability. See the repository's tests/ directory for details.

Research Impact Statement

PixelMap addresses a practical bottleneck in Neuropixels experimental workflows. Neuropixels have become the dominant technology for large-scale electrophysiology, with exponential growth in publications using the technology (PubMed). Yet no existing tool provided installation-free channelmap design with support for arbitrary electrode geometries (see Statement of Need).

PixelMap demonstrates community-readiness through comprehensive documentation and a permissive open-source license (GPL3). The tool is immediately accessible via web application (https://neuropixels-channelmap-generator.pni.princeton.edu), Python package, Docker container, or programmatic API. The tool builds on the authors' established track record using Neuropixels probes in their research (Beau et al., 2025; Bondy et al., 2024; Kostadinov et al., 2019; Steinmetz et al., 2021) and developing Neuropixels software (Beau et al., 2021).

Evidence of adoption includes community engagement on the project repository (22 GitHub stars as of January 2025) and deployment at Princeton Neuroscience Institute's public server.

AI usage Disclosure

AI-assisted technologies used: Claude Sonnet 4.1, Sonnet 4.5, and Opus 4.5 (Anthropic). AI assistance was used for (1) optimization suggestions and documentation improvements (docstrings, code comments) in backend.py, (2) initial scaffolding of the HoloViz Panel GUI architecture in gui/gui.py, (3) manuscript grammatical and syntactical review. AI was not used for project conceptualization, core algorithm design, electrode wiring map construction. App hosting infrastructure was designed independently of AI assistance.

Author Contributions

Table with 4 columns: Contribution, Maxime Beau, Christian Tabedzki, Carlos D. Brody. Rows include Conceptualisation, Backend and GUI, App hosting, and Supervision and funding.

Acknowledgements

We thank Julie Fabre for discussions and designing PixelMap's logo. We also thank Jesse C. Kaminsky, Jorge Yanar, and members of the Brody laboratory for testing and feedback during development, and PNI IT members Garrett McGrath and Gary Lyons for their advice concerning hosting. Finally, we thank the Princeton Neuroscience Institute for hosting the web application. This work was supported by Howard Hughes Medical Institute and the National Institutes of Health.

References

Beau, M., D'Agostino, F., Lajko, A., Martínez, G., Häusser, M., & Kostadinov, D. (2021). NeuroPyxels: Loading, processing and plotting Neuropixels data in Python. Zenodo.

- 115 <https://doi.org/10.5281/zenodo.5509733>
- 116 Beau, M., Herzfeld, D. J., Naveros, F., Hemelt, M. E., D'Agostino, F., Oostland, M.,
117 Sánchez-López, A., Chung, Y. Y., Maibach, M., Kyranakis, S., Stabb, H. N., Martínez
118 Lopera, M. G., Lajko, A., Zedler, M., Ohmae, S., Hall, N. J., Clark, B. A., Cohen,
119 D., Lisberger, S. G., & others. (2025). A deep learning strategy to identify cell types
120 across species from high-density extracellular recordings. *Cell*, 188(8), 2218–2234.e22.
121 <https://doi.org/10.1016/j.cell.2025.01.041>
- 122 Bondy, A. G., Charlton, J. A., Luo, T. Z., Kopec, C. D., Stagnaro, W. M., Venditto, S. J. C.,
123 Lynch, L., Janarthanan, S., Oline, S. N., Harris, T. D., & Brody, C. D. (2024). Coordinated
124 cross-brain activity during accumulation of sensory evidence and decision commitment.
125 *bioRxiv*. <https://doi.org/10.1101/2024.08.21.609044>
- 126 Jun, J. J., Steinmetz, N. A., Siegle, J. H., Denman, D. J., Bauza, M., Barbarits, B., Lee, A. K.,
127 Anastassiou, C. A., Andrei, A., Aydin, Ç., Barbic, M., Blanche, T. J., Bonin, V., Couto, J.,
128 Dutta, B., Gratiy, S. L., Gutnisky, D. A., Häusser, M., Karsh, B., & others. (2017). Fully
129 integrated silicon probes for high-density recording of neural activity. *Nature*, 551(7679),
130 232–236. <https://doi.org/10.1038/nature24636>
- 131 Kostadinov, D., Beau, M., Blanco-Pozo, M., & Häusser, M. (2019). Predictive and reactive
132 reward signals conveyed by climbing fiber inputs to cerebellar purkinje cells. *Nature*
133 *Neuroscience*, 22(6), 950–962.
- 134 Steinmetz, N. A., Aydin, C., Lebedeva, A., Okun, M., Pachitariu, M., Bauza, M., Beau,
135 M., Bhagat, J., Böhm, C., Broux, M., Chen, S., Colonell, J., Gardner, R. J., Karsh, B.,
136 Kloosterman, F., Kostadinov, D., Mora-Lopez, C., O'Callaghan, J., Park, J., & others.
137 (2021). Neuropixels 2.0: A miniaturized high-density probe for stable, long-term brain
138 recordings. *Science*, 372(6539), eabf4588. <https://doi.org/10.1126/science.abf4588>
- 139 Yang, S., Madsen, M. S., & Bednar, J. A. (2022). HoloViz: Visualization and Interactive
140 Dashboards in Python. *Proceedings of the 28th ACM SIGKDD Conference on Knowledge*
141 *Discovery and Data Mining*, 4846–4847. <https://doi.org/10.1145/3534678.3542621>
- 142 Ye, Z., Shelton, A. M., Shaker, J. R., Boussard, J., Colonell, J., Birman, D., Manavi, S., Chen,
143 S., Windolf, C., Hurwitz, C., Yu, H., Namima, T., Pedraja, F., Weiss, S., Raducanu, B. C.,
144 Ness, T. V., Jia, X., Mastroberardino, G., Rossi, L. F., & others. (2025). Ultra-high-density
145 Neuropixels probes improve detection and identification in neuronal recordings. *Neuron*.
146 <https://doi.org/10.1016/j.neuron.2025.08.030>