

Michael Jahn

Curriculum vitae

Personal information

Name Dr. rer. nat. Michael Jahn

Date of birth Dec 31, 1985

Born in Dresden, Germany

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Current position

Post-doctoral fellow, group of Prof Paul Hudson, Science For Life Laboratory, Stockholm. Topic: Systems biology and metabolic engineering of photoautotrophic bacteria.

Education

July 10, 2015 **Graduated Doctor rerum naturalium**, at the University of Leipzig, Germany, with summa cum laude (1.0).

2011–2015 **PhD studies**, Group of Prof Susann Müller, Dept. Environmental Microbiology, Helmholtz-Centre for Environmental Research, Leipzig.

Thesis: 'Characterization of population heterogeneity in a model biotechnological process using *Pseudomonas putida*'.

2005–2011 **Diploma studies, biology**, Dresden University of Technology (TUD), main subjects: genetics, biochemistry, immunology.

Diploma thesis: 'Dynamic mating pheromone gradients for induction of mating projection and fluorescence in yeast', group of Prof Gerhard Rödel, grade 1.2.

1996–2004 **High school**, Romain-Rolland-Gymnasium, Dresden, Abitur with grade 1.6.

1992–1996 **Elementary school**, Dresden.

Work experience

5/2016-now **Post-doctoral researcher**, Systems biology analysis of cyanobacteria at the group of Prof Paul Hudson, Science For Life Laboratory, Stockholm (belongs to KTH).

8–12/2009 **Erasmus internship**, Center of Excellence in Evolutionary Genetics and Physiology at the group of Prof Mikko Nikinmaa, Dept. of Biology, University of Turku, Finland.

2008–2009 **Student assistant**, Institute of Genetics, group of Prof Gerhard Rödel, Dept. of Biology, Dresden University of Technology (TUD).

2007–2008 **Student assistant**, Mitteldeutscher Praxisverbund Humangenetik, Dresden.

2004–2005 **Voluntary service 'Freiwilliges Ökologisches Jahr'**, national park 'Sächsische Schweiz', tour guide and education.

Leaves

11/2019-3/2020 Parental leave with 2nd child Sophie 9/2015-4/2016 Parental leave with 1st child Simon

Awards

Nov 25, 2015 **PhD-Award** 2015 by the Helmholtz-Centre for Environmental Research, Leipzig. Awarded annually for excellence of the dissertation, prize money 1,000 Euro.

Awarded grants

Nov 11, 2019 **Investigation of protein resource allocation in the model CO₂ fixing bacterium** *Ralstonia eutropha* (Swedish: Undersökning av proteiners resursfördelning i en modell för CO₂-fixerande bakterier, Ralstonia eutropha).

Funding agency: FORMAS. Project number: 2019-01491. Project leader: Dr. Michael Jahn. Awarded funding for period 2020-01-01-2021-12-31. Total funds: 2,000,000 SEK (200,000 Euro).

Grant proposals

Jan 26, 2016 **Development of single cell screening tools to optimize N-glycosylation efficiency in Saccharomyces cerevisiae** (German: Entwicklung von Einzelzell-Analyseverfahren zur Optimierung der N-Glykosylierung in Saccharomyces cerevisiae).

Funding agency: Deutsche Forschungsgemeinschaft (DFG). Research Fellowship, two year funding period. Project number: JA 2529/1-1. Positively evaluated but turned down due to competing job offer.

Jun, 2015 **Analysis of plasmid and chromosomal copy number variation in heterotrophic bacteria**. Co-author/mentor for PhD research proposal.

Funding agency: Deutsche Bundesstiftung Umwelt (DBU). Declined in second stage of evalutation.

Teaching and supervision

Teaching

lecture for course KE2130 'Renewable fuels', at KTH Stockholm. Winter semester 2020/2021. Topics: Metabolic engineering, whole cell biocatalysts, genome-scale models, flux balance analysis.

lecture for course CB2030 'Systems biology', at KTH Stockholm. Winter semester 2019/2020, *postponed* due to parental leave.

Introduction to the programming language R and its application for flow cytometry data, at the Helmholtz Centre for Environmental Research, Leipzig, 2015.

PhD council

Member of the PhD council and the Environmental Board of the Helmholtz Centre for Environmental Research, Leipzig, 2011-2015. Among other tasks providing guidance for starting PhD students.

Supervision

Supervised master and intern students during my PhD and postdoctoral studies:

- o Alexander Mattausch, intern biotechnology, University of Heidelberg, 2019
- o Timothy Bergmann, master student biotechnology (KTH, Stockholm), 2019
- o Raquel Perruca, master student, Lund Technical University, 2018
- o Carsten Vorpahl, master student University of Leipzig, 2015
- o Dominique Türkowsky, master student University of Leipzig, 2014

Scientific interests and expertise

Sustainability

Current overuse of (fossil) planetary resources and disposal of non-degradable waste into the environement makes the shift towards a sustainable economy mandatory. The concept of the **bioeconomy** proposes to replace conventional chemical processes with sustainable ones. Autotrophic microbes can play a roll as versatile, efficient and biodegradable catalysts.

Metabolic modeling, data science High-throughput methods generate larger amounts of biological data than ever before. This poses problems, but also opportunities. I envision a close integration of experiments and data with **predictive mathematical models**, such as genome scale metabolic models.

Metabolic engineering

Microbial cells are 'programmed' for production of biomass, not fine chemicals. **Rational design and engineering of pathways** is necessary to turn photo-autotrophic bacteria into 'solar power plants'.

Proteomics & other 'omics

Proteomics, transcriptomics and metabolomics are probing the whole cell instead of isolated parts. This allows to discover biological relationships that were not considered in the original hypothesis, and gives an unbiased picture.

Single cell variability

Previous work regarding **single cell variability** of *S. cerevisiae*, *E. coli*, and *Pseudomonas putida* showed me that a population is never uniform but varies extensively in size, shape, productivity, genetic inventory. Single cell variability must be considered in microbial research.

Reproducibility and Transparency

Adopting **open standards, sharing protocols and resources**, and depositing data and apps on publicly available sites has great advantages. It increases reach, trust of the scientific community, and reproducibility of results.

Qualifications and skills

Courses Approved project manager for genetic works, according to German laws (§ 14,15

(selection) GenTSV), 2015.

Laser scanning microscopy, 1 week course, 2013.

Scientific writing and project proposal writing, 2 week course, 2013.

Model Cultivation and molecular biology of cyanobacteria (e.g. Synechocystis sp.,

organisms Synechococcus), Cupriavidus necator, Pseudomonas putida, E. coli

Techniques Bioreactor cultivation, chemostat, turbidostat

Mass spectrometry, proteomics

Flow cytometry, cell sorting, automated imaging

Data analysis and visualization

Molecular biology

Programming R (advanced), Python (moderate)

lanuguages
Authored R

packages

SysbioTreemaps – visualizing gene expression with treemaps ShinyMC – interactive app to monitor bioreactor cultivations

ShinyProt - interactive app to visualize proteomics data

ShinyLib - interactive app to visualize gene knockout library data

Metabolic Improved *Synechocystis* coarse grained (simplified) resource allocation model, models for GAMS, and ported to python.

Improved and updated genome scale model (GEM) for *Cupriavidus necator* a.k.a. *Ralstonia eutropha*.

Constructed new genome scale resource balance (RBA) model for *Cupriavidus* necator a.k.a. *Ralstonia eutropha*

Peer review and outreach

Peer review Active reviewer for several journals including Microbial Cell Factories, Frontiers

in Microbiology, Photosynthesis Research, and others. The full list of my review

contributions is available via Publons.org

Platforms My home page ↗

My github page ↗

My ORCID page ↗

My ResearchGate page ↗

Languages

German Native language.

English **Proficient user**, CEF level C1.

Swedish Vantage user, CEF level B2.

Publications

- Yao L, Shabestary K, Björk SM, Asplund-Samuelsson J, Joensson HN, Jahn M, Hudson EP. Pooled CRISPRi screening of the cyanobacterium Synechocystis sp PCC 6803 for enhanced industrial phenotypes. Nature Communications, 2019.
- Karlsen J, Asplund-Samuelsson J, Thomas Q, Jahn M, Hudson EP. Ribosome Profiling of Synechocystis Reveals Altered Ribosome Allocation at Carbon Starvation. MSystems 3, e00126-18, 2018.
- Jahn M, Vialas V, Karlsen J, Maddalo G, Edfors F, Forsström B, Uhlén M, Käll L, Hudson EP. Growth of Cyanobacteria Is Constrained by the Abundance of Light and Carbon Assimilation Proteins. Cell Reports 25, 478–486.e8., 2018.
- Shabestary K, Anfelt J, Ljungqvist E, Jahn M, Yao L, Hudson EP. Targeted Repression of Essential Genes To Arrest Growth and Increase Carbon Partitioning and Biofuel Titers in Cyanobacteria. ACS Synthetic Biology, 7, 1669–1675, 2018.
- Jahn M, Vorpahl C, Hübschmann T, Harms H, Müller S. Copy number variability of expression plasmids determined by cell sorting and Droplet Digital PCR. Microbial Cell Factories, 2016.
- Lindmeyer M, Jahn M, Vorpahl C, Müller S, Schmid A, Bühler B. Variability in subpopulation formation propagates into biocatalytic variability of engineered Pseudomonas putida strains. Frontiers in microbiology 6, 2015.
- Lieder S, Jahn M, Koepff J, Müller S, Takors Ralf. Environmental stress speeds up DNA replication in Pseudomonas putida in chemostat cultivations. Biotechnology journal, 2015.
- Jahn M, Günther S, Müller S. Non-random distribution of macromolecules as driving forces for phenotypic variation. Current Opinion in Microbiology. 25:49-55,
 2015.
- Rödiger S, Burdukiewicz M, Blagodatskikh K, Jahn M, Schierack P. R as an environment for reproducible analysis of DNA amplification experiments. R Journal 7/1:127-150, 2015.
- Jahn M, Vorpahl C, Türkowsky D, Lindmeyer M, Bühler B, Harms H, Müller S. Accurate determination of plasmid copy number of flow-sorted cells using droplet digital PCR. Analytical Chemistry 86:5969-76, 2014.
- Lieder S, Jahn M, Seifert J, von Bergen M, Müller S, Takors R. Subpopulationproteomics reveal growth rate, but not cell cycling, as a major impact on protein composition in Pseudomonas putida KT2440. AMB Express 4:71, 2014.
- Jahn M, Seifert J, von Bergen M, Schmid A, Bühler B, Müller S. Subpopulationproteomics in prokaryotic populations. Current Opinion in Biotechnology 24:79-87, 2013.
- o **Jahn M**, Seifert J, Hübschmann T, von Bergen M, Harms H, Müller S. *Comparison of preservation methods for bacterial cells in cytomics and proteomics*. Journal Of Integrated Omics 3:1-9, **2013**.
- Jahn M, Mölle A, Rödel G, Ostermann K. Temporal and spatial properties of a yeast multi-cellular amplification system based on signal molecule diffusion. Sensors 13:14511-22, 2013.

Conference contributions	Confer	ence co	ontribu	utions	
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Sep 07-09, 2020	11EWBC - 11th European workshop on the biology of cyanobacteria, Porto, Portugal (changed to virtual format). Oral presentation.
May 22-24, 2019	NPC 14 - Nordic congress on photosynthesis, Turku, Finland. Short oral presentation.
Oct 24-26, 2018	4th Applied Synthetic Biology meeting - Toulouse, France. Oral presentation.
Jul 20-22, 2015	Single Cell VI – 6 th International Conference on Analysis of Microbial Cells at the Single Cell Level, Retz (Austria). Oral presentation.
Oct 15-17, 2014	DGFZ – 24 th Annual Conference of the German Society for Cytometry, Dresden. Oral presentation.
Jul 13-16, 2014	ECB16 – 16 th European Congress on Biotechnology, Edinburgh. Oral presentation.
May 26-28, 2014	DECHEMA – Biomaterials Made in Bioreactors, Dresden. Poster.
Mar 06-08, 2013	RPP7 – 7 th European Conference on Recombinant Protein Production, Ulm. Oral presentation.
Jul 21-25, 2013	FEMS – 5 th Congress of European Microbiologists, Leipzig. Poster.
Oct 10-12, 2012	DGFZ – 22 nd Annual Conference of the German Society for Cytometry, Bonn. Oral presentation.
Jun 23-27, 2012	CYTO 2012 – 27 th Congress of the International Society for Advancement of Cytometry, Leipzig. Oral presentation.