

The effect of a brood-size manipulation and GPS tag on *Hirundo rustica* blood glucose levels

Background:

Life history of an organism gives insights to its quality of life. A way to examine an organism's health is by looking at its stress levels. The best way to quantify stress is through measuring blood glucose, which can be an indicator of stress. In avian species, stress causes an elevation in corticosterone as well as blood glucose (Blas, 2015; Braun & Sweazea, 2008).

One of the most costly endeavors an avian will pursue in its life is reproduction. Reproduction is a personal cost, since it requires so much energy and investment. A way to quantify the cost of reproduction, can be examined through blood glucose. Avians in the genus *Hirundo*, exhibit an exceedingly high blood glucose concentration relative to their body mass, and yet they do not exhibit oxidative stress damage (Braun & Sweazea, 2008; Sweazea et al., 2008). Blood glucose regulation in avian populations is regulated by endocrine hormones that uphold homeostatic control (Dawson, 2000; Vágási et al., 2020). Small avian species, like *Hirundo rustica*, have increased metabolic demand, which in turn leads them to exhibit an increased glucose concentration in blood plasma which is likely due to diet composition and avian physiology (Basile et al., 2018; Braun & Sweazea, 2008) The aim of this study is to look at possible elevation of blood glucose due to the stress of reproduction.

This study examines *Hirundo rustica* blood glucose by adding experimental measures. It is hypothesized that both a brood size-manipulation and addition of a GPS tag will have an effect on the blood glucose levels in Barn Swallows due to possible added stress.

Methods:

Study System

Hirundo rustica, commonly known as Barn Swallows, are small colorful passerines that migrate each year from S. America to N. America in order to nest and brood young (Brown & Brown, 2020). Nest-building occurs from May to mid-June, and adults breed throughout the summer into the early fall months. Barn Swallows create a distinctive mud-cup nest, commonly seen in agricultural barns and across cityscapes. Their preferred habitat is near agricultural fields, which provide a food source for these aerial insectivores (Scordato & Safran, 2014).

Field methods

In this study, we collected data from adult barn swallows during the breeding season in Boulder County, CO between the months of May-September (2019-2020). We collected physiological samples from a population of n=37 adult female barn swallows. The sites selected

are local agricultural barns, horse ranches, and underpasses. The different study sites reflect both rural and urban habitats with 16 sites total. 6 sites were underpasses in urbanized areas, and the other 10 were barns in agricultural areas mostly near livestock.

This study is a part of an ongoing research study in which nesting individuals are studied and catalogued each year in Boulder County by *Dr. Rebecca Safran's* lab at the *University of Colorado Boulder*. The design of the long-term study is that study sites among Boulder County are long-term nesting sites of Barn Swallows in which data from this specific population is collected annually. The population examined migrates annually to these long-term sites in which they are monitored and tagged between the months of May-September.

In the study, individual birds were identified at a nest at each site through distinctive color bands, as well as USGS tag number. Brooding females were of key interest, so each capture physiological measures were assessed as well as adding a color and USGS metal band (if not already present). The same was done for male birds, but since brooding females were the main study for this experiment, throughout the whole population females had physiological measurements taken throughout the study. This included measures such as wing length, tail streamer length, mass, USGS tag number, color bands, and feathers (which were collected from both the body and tail). Brooding females that successfully incubated eggs into nestlings were able to be candidates for the experiment.

All of the brooding females were randomly placed into one of four groups: experimental and control; experimental females were subjected to both a brood size manipulation and the placement of a GPS tag on their back using a Rappole leg-loop harness. We tagged females when chicks were 4-6 days old, and we manipulated brood size when chicks were 8 days old. Nests were either enlarged (E) or reduced (R) through randomized selection. Each female served as her own control, since measurements were taken before and after tag addition and brood size manipulation. We collected blood from the brachial vein using a capillary tube. After blood was extracted, a cotton ball was applied with pressure to the prick site to prevent further bleeding. Blood glucose was measured in the field using a Freestyle Lite® Glucose Monitor and strips. The Blood glucose reading was immediately recorded in $\mu\text{g}/\text{dL}$ and sampled once the bird was out of the cloth bag. Blood glucose was recorded before the tag was placed on and after the tag was removed. The bird's physiological data was also collected when blood glucose was measured, which included: mass, feather samples, measurements, and wing faults.

Females that did not receive a GPS tag were captured and handled to be subjected under the same stress conditions administered to experimental females with a GPS tag. Females that had a GPS tag were also captured and handled in a similar manner, except the GPS tag was placed on. In order to measure the blood glucose level of a bird, birds were placed in a cloth bag for 15 minutes (Montoya et al., 2020). Other covariates measured included mass (g) that was collected using a scale, time the bird was captured was recorded, type brood treatment (enlarged or reduced), year, and site were recorded. These variables are used to isolate the effect of treatment on blood glucose, since a bird's mass, the time of day the measurement was taken, year, and brood treatment may all have an effect on the blood glucose measure.

Statistical methods

The data was first cleaned in Excel. The filtering of the data was done in RStudio and birds that had two paired glucose measurements before and after treatment were only used. The data was analyzed by first assessing the univariate distributions, bivariate descriptive statistics and associations, and finally the full generalized mixed model. The mixed model included change in blood glucose (treatment 1 - treatment 2), tag, brood treatment, mass (before treatment), year, and site. The independent variables are tag and brood treatment. One of the dependent variables is blood glucose, specifically the change in blood glucose. A residual model was created for change in blood glucose to account for time of day. Time of day was recorded in 24 hour format, which allowed for transformation of this data. The sine of the time data was taken, which is a method used to transform circular data using the R package 'circular' (Cremers and Klugkist, 2018). The other dependent variable is mass. The random effects are site and year. The software used was RStudio (version 1.2.1335) and the packages used were broom, dotwhisker, dplyr, emmeans, car, circular, ggplot2, lme4, lmerTest, lubridate, and tidyverse.

Results:

In this study, we found that there is no effect of brood size manipulation or the addition of GPS tag on blood glucose levels in Barn Swallows. The brood size manipulation showed no significant effect ($p = 0.54$) on within-individual change in blood glucose. The addition of a GPS tag showed no significant effect ($p = 0.58$) on within-individual change in blood glucose. Therefore, the null hypothesis cannot be rejected. The implications of these findings is that this research provides valuable insights for future studies in which stressed blood glucose is measured in a way to control for external factors such as foraging schedule, time of day, and activity levels. For future research, creating a standardized measure for blood glucose (such as measuring fasted blood glucose in the birds) may help control for variation.

Tables and Figures: (on next page)

Link to GitHub page with code: https://github.com/maay21/EBIO_4410

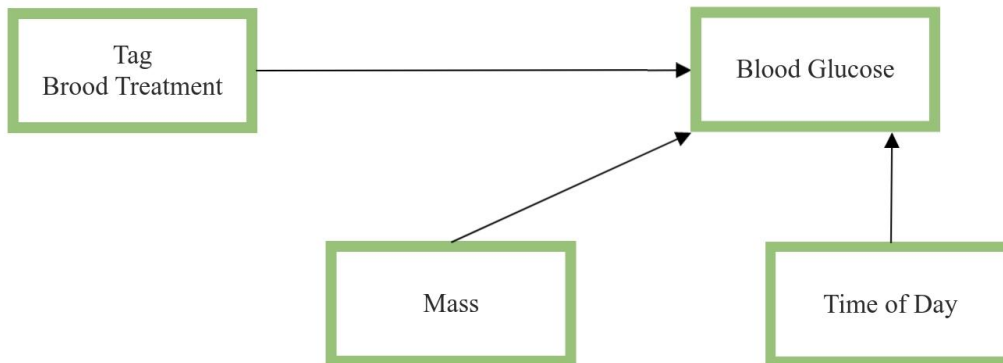


Figure 1. A DAG Directed-acyclic graph (DAG) showing the relationship between variables and blood glucose (Laubach et al. 2021). Blood glucose (specifically the change) is the dependent variable. Mass and time of day are precision covariates. The independent variables are tag and brood treatment.

$$\Delta \text{Blood Glucose} \sim \text{tag} + \text{broodSize} + \text{tag} * \text{broodSize} + \text{mass1} + \text{year} + \text{time} + (1 | \text{site})$$

Figure 2. Equation used to model the change in blood glucose.

Table 1. Mean and Standard Deviation for Blood Glucose and Mass	Mean	Standard Deviation
Change in Blood Glucose	19.10811	42.90868
Initial Blood Glucose (Before Treatment)	217.4595	31.91969
Final Blood Glucose (After Treatment)	236.5676	39.28566
Change in Mass	-0.7683784	0.8600146
Initial Mass (Before Treatment)	17.96108	0.9339694
Final Mass (After Treatment)	17.1927	1.07599
Time of Day (Radian Hours, Before Treatment)	0.255361	0.7089235
Time of Day (Radian Hours, After Treatment)	0.00649219	0.7089235

Table 1. Mean and standard deviation for blood glucose, mass, and time of day as radian hours.

Table 2. Brood Treatment and Tag	Sample Size	% of Individuals
Brood Treatment (Enlarged)	20	54%
Brood Treatment (Reduced)	17	46%
Tag	20	54%
No Tag	17	46%

Table 2. Brood treatment vs. tag. The sample size is indicated for each variable, as well as the percentage of individuals in the sample population that received each treatment.

Table 3. Estimate and Confidence Intervals Categorical	Estimate	Standard Error	Degrees of Freedom	P-value	Lower CI	Upper CI
Change in Blood Glucose and Tag	12.61	14.2	35	0.3806	-16.214881	41.42665
Change Blood Glucose and Brood Treatment	10.9	14.24	35	0.449	-18.00256	39.80256

Table 3. Estimate and confidence intervals for the categorical variables: brood treatment and tag. Both p-values >0.05, so the relation to blood glucose is not significant. 95% confidence intervals used.

Table 4. Change in Blood Glucose and Tag	SE	df	lower CI	upper CI
No Tag	10.44	35	-8.9	33.5
Tag	9.62	35	5.36	44.4

Table 4. Tag vs. no tag. 95% confidence intervals used.

Table 5. Change in Blood Glucose and Brood Treatment	SE	df	lower CI	upper CI
Enlarged	9.65	35	-5.49	33.7
Reduced	10.47	35	3.75	46.2

Table 5. Change in blood glucose vs. brood treatment. 95% confidence intervals used.

Time of Day That Data Samples Were Collected Before Treatment

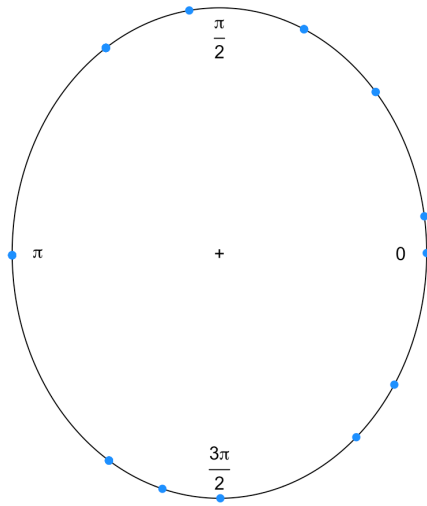


Figure 3. Time of day as radian hours before treatment (initial measure of blood glucose). Time was converted to radian hours by first converting time to 24-hour time, then taking the sin of each hour. $\frac{\pi}{2}$ represents noon/midnight (Cremers and Klugkist, 2018). Each point represents one measure of blood glucose.

Time of Day That Data Samples Were Collected After Treatment

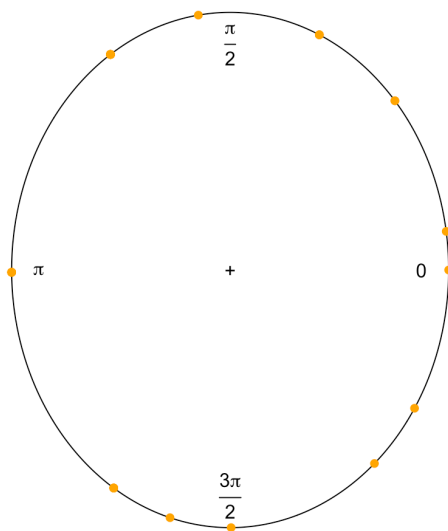


Figure 4. Time of day as radian hours after treatment (final measure of blood glucose). Time was converted to radian hours by first converting time to 24-hour time, then taking the sin of each hour. $\frac{\pi}{2}$

represents noon/midnight (Cremers and Klugkist, 2018). Each point represents one measure of blood glucose.

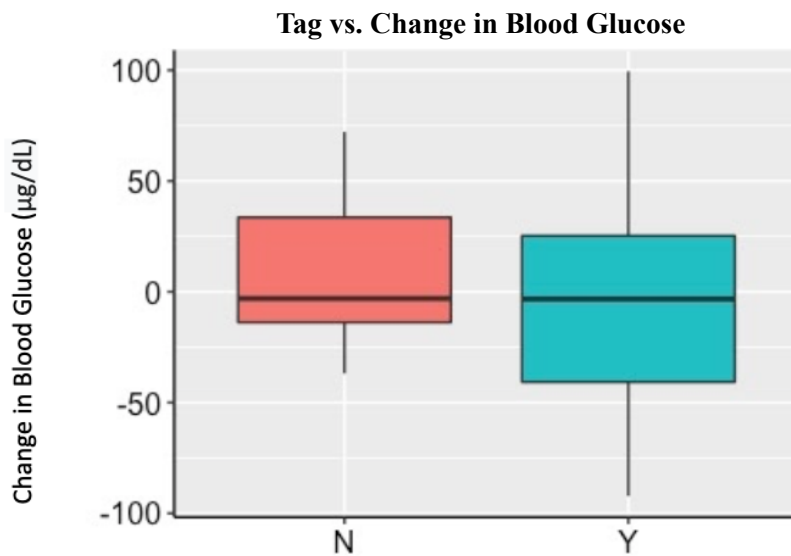


Figure 5. Tag (N- no tag, Y- tag) vs. change in blood glucose (measured in $\mu\text{g/dL}$). Middle line indicates median value, top and bottom represent 75th and 25th percentile, respectively.

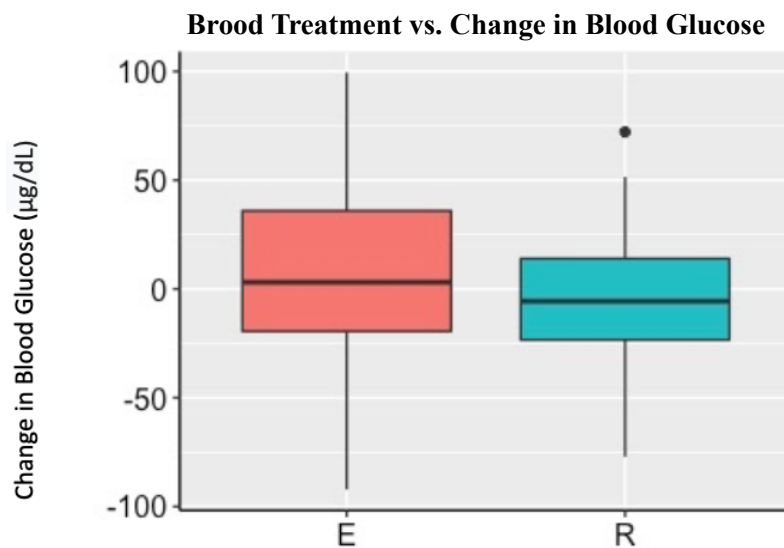


Figure 6. Brood treatment (E- enlarged, R- reduced) vs. change in blood glucose (measured in $\mu\text{g/dL}$). Middle line indicates median value, top and bottom represent 75th and 25th percentile, respectively.

References:

- Basile AJ, Jarrett CL, Sweazea KL. 2018. Relationship between Dietary Profile and Blood Glucose Concentration in Birds. *FASEB J.* 32(S1):602.3-602.3. doi:https://doi.org/10.1096/fasebj.2018.32.1_supplement.602.3.
- Blas J. 2015. Chapter 33 - Stress in Birds. In: Scanes CG, editor. *Sturkie's Avian Physiology* (Sixth Edition). San Diego: Academic Press. p. 769–810. [accessed 2021 Apr 28]. <https://www.sciencedirect.com/science/article/pii/B9780124071605000336>.
- Braun EJ, Sweazea KL. 2008. Glucose regulation in birds. *Comp Biochem Physiol B Biochem Mol Biol.* 151(1):1–9. doi:10.1016/j.cbpb.2008.05.007.
- C. Agostinelli and U. Lund (2017). R package 'circular': Circular Statistics (version 0.4-93). URL <https://r-forge.r-project.org/projects/circular/>
- Cremers J, Klugkist I. 2018. One Direction? A Tutorial for Circular Data Analysis Using R With Examples in Cognitive Psychology. *Front Psychol.* 9. doi:10.3389/fpsyg.2018.02040. [accessed 2020 Nov 30]. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6218623/>.
- David Robinson, Alex Hayes and Simon Couch (2021). broom: Convert Statistical Objects into TidyTibbles. R package version 0.7.6. <https://CRAN.R-project.org/package=broom>
- Dawson A. 2000. Mechanisms of Endocrine Disruption with Particular Reference to Occurrence in Avian Wildlife: A Review. *Ecotoxicology.* 9(1):59–69. doi:10.1023/A:1008964128501.
- Douglas Bates, Martin Maechler, Ben Bolker, Steve Walker (2015). Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software,* 67(1), 1-48. doi:10.18637/jss.v067.i01.
- Frederick Solt and Yue Hu (2021). dotwhisker: Dot-and-Whisker Plots of Regression Results. R package version 0.6.0. <https://CRAN.R-project.org/package=dotwhisker>
- Garrett Grolemond, Hadley Wickham (2011). Dates and Times Made Easy with lubridate. *Journal of Statistical Software,* 40(3), 1-25. URL <https://www.jstatsoft.org/v40/i03/>.
- H. Wickham. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2016.
- Hadley Wickham, Romain François, Lionel Henry and Kirill Müller (2021). dplyr: A Grammar of Data Manipulation. R package version 1.0.5. <https://CRAN.R-project.org/package=dplyr>

John Fox and Sanford Weisberg (2019). An {R} Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage. URL:

<https://socialsciences.mcmaster.ca/jfox/Books/Companion/>

Kuznetsova A, Brockhoff PB, Christensen RHB (2017). “lmerTest Package: Tests in Linear Mixed Effects Models.” *Journal of Statistical Software*, *82*(13), 1-26. doi:10.18637/jss.v082.i13 (URL:<https://doi.org/10.18637/jss.v082.i13>).

Laubach ZM, Murray EJ, Hoke KL, Safran RJ, Perng W. 2021. A biologist’s guide to model selection and causal inference. *Proc R Soc B Biol Sci*. 288(1943):20202815. doi:10.1098/rspb.2020.2815.

Montoya B, Briga M, Jimeno B, Verhulst S. 2020. Glucose regulation is a repeatable trait affected by successive handling in zebra finches. *J Comp Physiol B*. 190(4):455–464. doi:10.1007/s00360-020-01283-4.

Russell V. Lenth (2021). emmeans: Estimated Marginal Means, aka Least-Squares Means. R package version 1.5.5-1. <https://CRAN.R-project.org/package=emmeans>

Sweazea KL, Wolf BO, Braun EJ, Walker BR. 2008. High blood glucose is not associated with oxidative stress or vascular dysfunction in birds. *FASEB J*. 22(S1):1239.24-1239.24. doi:https://doi.org/10.1096/fasebj.22.1_supplement.1239.24.

Umminger BL. 1975. Body size and whole blood sugar concentrations in mammals. *Comp Biochem Physiol A Physiol*. 52(3):455–458. doi:10.1016/S0300-9629(75)80065-X.

Vágási CI, Tóth Z, Péntes J, Pap PL, Ouyang JQ, Lendvai ÁZ. 2020. The Relationship between Hormones, Glucose, and Oxidative Damage Is Condition and Stress Dependent in a Free-Living Passerine Bird. *Physiol Biochem Zool*. 93(6):466–476. doi:10.1086/711957.

Wickham et al., (2019). Welcome to the tidyverse. *Journal of Open Source Software*, 4(43), 1686, <https://doi.org/10.21105/joss.01686>