diagnostic graphics

Packages

```
library(tidyverse)
theme_set(theme_bw())
library(car)
library(broom)
library(broom.mixed)
library(magrittr)
## modeling
library(lme4)
library(MCMCglmm)
library(glmmTMB)
library(coda) ## Bayesian methods (trace plots etc.)
library(cowplot)
library(nullabor) ## visual inference
```

Diagnostics: goals and ideas

- detect model failure
- display badness of fit
- fast/convenient
- residuals emphasize deviation, hide fitted pattern

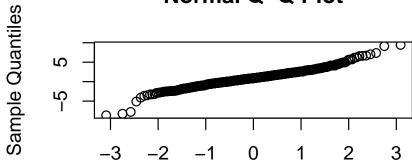
Diagnostics: principles

- diagnosis is exploration
- avoid making decisions based on p-values
- judge by eye (?¿?¿?¿?¿)
- **not** "are my data (linear | normal | heteroscedastic)?"; rather, "how much do the violations change my conclusions?"

diagnostics after fitting model

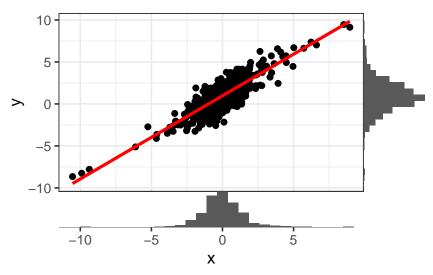
- Interested in conditional, not marginal values
- What does this mean?

Normal Q-Q Plot



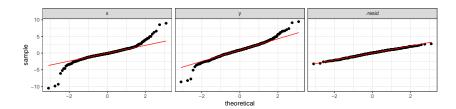
Theoretical Quantiles

marginal distributions



quantile plots

```
m\theta \leftarrow lm(y \sim x, dd)
a0 <- (augment(m0) %>% select(x, y, .resid) %>%
    gather(type, value) %>% mutate(type = factor(type,
    levels = c("x", "y", ".resid"))))
(ggplot(a0, aes(sample = value)) + stat_qq() +
    facet_wrap(~type) + stat_qq_line(colour = "red"))
```



model diagnosis

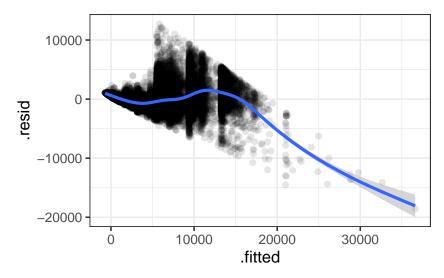
look for *mis-specification* (in order!):

- mean model (bias)
- variance model (heteroscedasticity)
- distributional model (e.g. non-normality)

influential points/groups (leverage/outliers/etc.) upstream problems affect downstream diagnostics

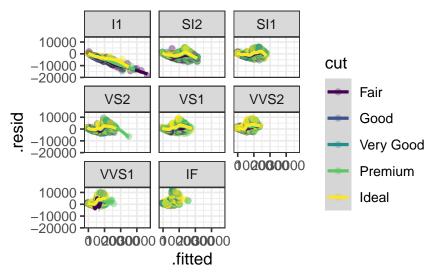
bias

```
m1 <- lm(price ~ carat, diamonds)</pre>
a1 <- augment(m1, data = diamonds) ## include original data</pre>
ggplot(a1, aes(.fitted, .resid)) + geom_point(alpha = 0.1) +
    geom_smooth()
```



bias 2: faceting/colouring

```
ggplot(a1, aes(.fitted, .resid, colour = cut)) +
    facet_wrap(~clarity) + geom_point(alpha = 0.4) +
    geom_smooth()
```



useful to use dynamic graphics ggmap::gglocator (may need devtools::install_github("dkahle/ggmap"))

solutions to bias

- fix the model
- · add covariates and interactions
- transform predictors and/or responses (acepack::avas, Tibshirani (1987))
- nonlinear terms
 - polynomials
 - splines (regular or penalized)
 - 'real' nonlinearity

heteroscedasticity

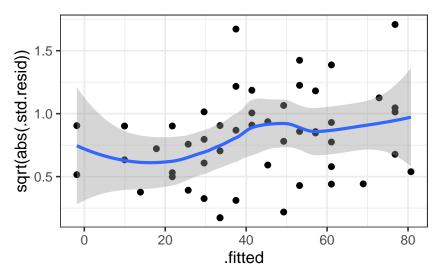
- linear models
 - loss of efficiency (linear fit is still MVUE)
 - inferential problems (Quinn and Keough (2002) p. 193)
- nonlinear models
 - bias

heteroscedasticity diagnostics

- scale-location plot
- use $\sqrt{|r_i|}$:
 - absolute value shows trend
 - square root decreases skewness

• use standardized residuals (adjust variance for position)

```
m2 <- lm(dist ~ speed, data = cars)</pre>
ggplot(augment(m2), aes(.fitted, sqrt(abs(.std.resid)))) +
    geom_point() + geom_smooth()
```



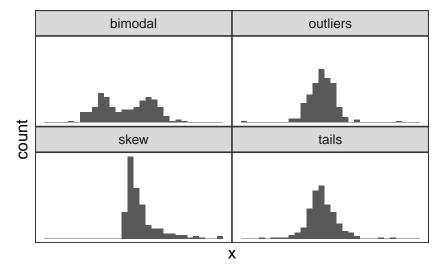
heteroscedasticity solutions

- transformation (Tibshirani 1987)
- explicitly model heteroscedasticity e.g. generalized least squares, GLMs
- robust variance-covariance estimation (e.g. sandwich package: Zeileis (2006))

distributional assumptions

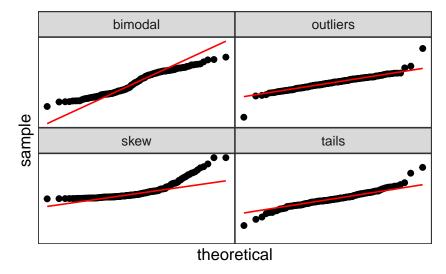
- least important
- quantile-quantile plots

histograms



quantile plots

- ggplot: stat_qq(), stat_qq_line()
- base R: plot.lm(.,which=3); qqnorm()
- car::qqPlot (adds confidence envelope)



distributional solutions

- transformation (avas, Box-Cox (MASS:boxcox), Yeo-Johnson etc. [?car::bcPower])
- GLMs
- maximum likelihood estimation

correlation

rarely tested! can't detect without some kind of structure in data

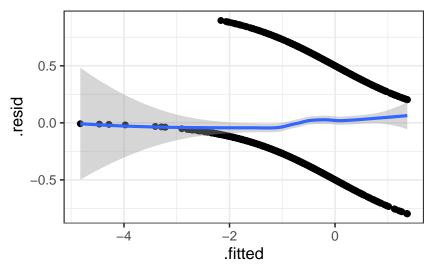
- autocorrelation plots from residuals
- grouped autocorrelation: use gls() on residuals
- spatial autocorrelation: semivariance plot
- or look at maps of residuals with size=abs(.resid), colour=sign(.resid) (or colour ramp)

binary data

- residuals for count data only \approx Normal for large counts
- add smooths or average of grouped data

Fit:

```
library(lme4)
data(Contraception, package = "mlmRev")
Contraception <- Contraception %>% mutate(ch = factor(livch !=
    0, labels = c("N", "Y")))
m3 \leftarrow glmer(use \sim age * ch + I(age^2) + urban +
    (1 | urban:district), data = Contraception,
    family = binomial)
plot
a3 <- augment(m3, data = Contraception, type.residuals = "response")
gg_bin1 <- (ggplot(a3, aes(.fitted, .resid)) +</pre>
    geom_point() + geom_smooth(method = "loess"))
print(gg_bin1)
```

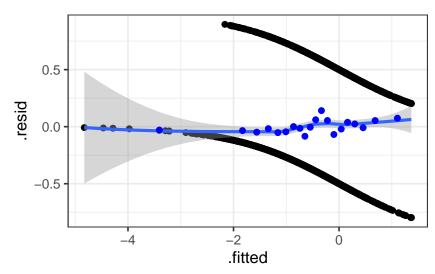


```
grouping
```

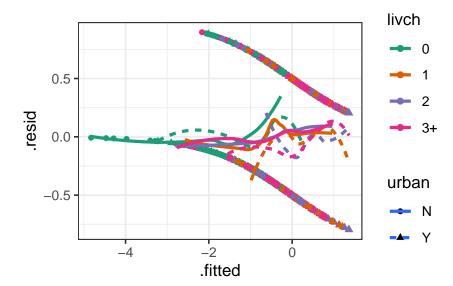
```
get_mid <- function(x) {</pre>
    cc <- as.character(x)</pre>
    lo <- as.numeric(gsub("[\\((\\[])([[:digit:].-]+).*",</pre>
        "\\1", cc))
    hi <- as.numeric(gsub(".*,([[:digit:].-]+)[])]",
        "\\1", cc))
    return((lo + hi)/2)
}
a3_sum <- (a3 %>% mutate(.fit_cut = cut_number(.fitted,
    20)) %% group_by(.fit_cut) %% summarise(.resid = mean(.resid)) %%
    ungroup %>% mutate(.fitted = get_mid(.fit_cut)))
```

plot with grouping

```
gg_bin1 + geom_point(data = a3_sum, colour = "blue")
```

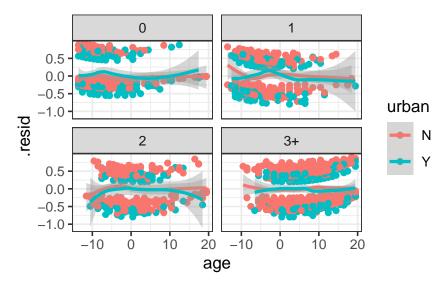


```
ggplot(a3, aes(.fitted, .resid, colour = livch,
    shape = urban, linetype = urban)) + geom_point() +
    geom_smooth(se = FALSE) + scale_colour_brewer(palette = "Dark2")
## 'geom_smooth()' using method = 'loess' and formula 'y \sim x'
```



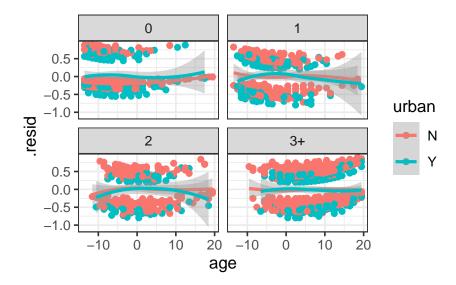
keep trying ...

```
ggplot(a3, aes(age, .resid, colour = urban)) +
    geom_point() + geom_smooth(method = "loess") +
    facet_wrap(~livch)
```



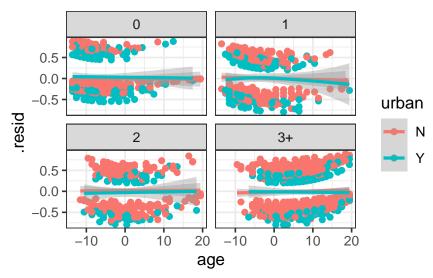
• loess too bumpy?

```
ggplot(a3, aes(age, .resid, colour = urban)) +
    geom_point() + geom_smooth(method = "loess",
    method.args = list(family = "symmetric"),
    span = 1) + facet_wrap(~livch)
```



• try method="gam" ?

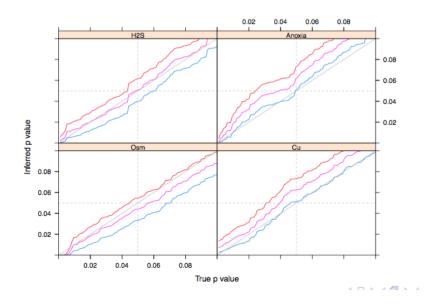
```
ggplot(a3, aes(age, .resid, colour = urban)) +
    geom_point() + geom_smooth(method = "gam",
    formula = y \sim s(x, k = 25)) + facet_wrap(~livch)
```



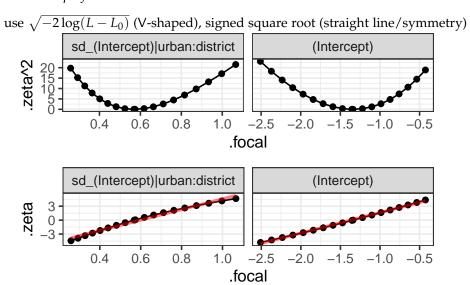
- note: qq.gam() (Augustin, Sauleau, and Wood 2012)
- improved QQ for GLMs (only for fits via mgcv::gam()?)
- binary data still problematic: from ?qq.gam,

Note that plots for raw residuals from fits to binary data contain almost no useful information about model fit. Whether the residual is negative or positive is decided by whether the response is zero or one. The magnitude of the residual, given its sign, is determined entirely by the fitted values. In consequence only the most gross violations of the model are detectable from QQ-plots of residuals for binary data. To really check distributional assumptions from residuals for binary data you have to be able to group the data somehow. Binomial models other than binary are ok.

coverage



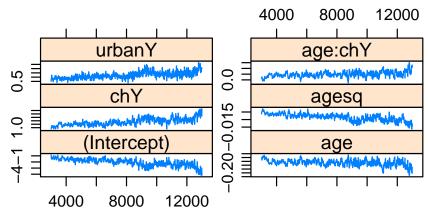
likelihood profiles



MCMC

• trace plots - should look like white noise, with no trend ...

```
lattice::xyplot(m4$Sol, aspect = "fill", layout = c(2,
    3))
```

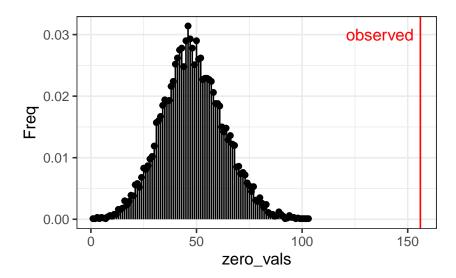


Iteration number

• trace plots

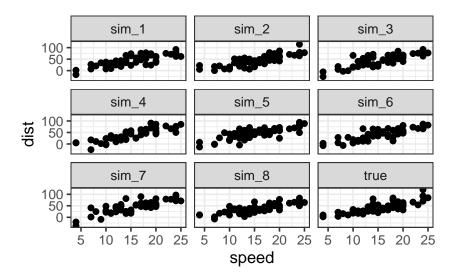
posterior predictive plots

```
set.seed(101)
nsim <- 10000
owls_sim <- simulate(owls_nb1, nsim)</pre>
sumfun < -function(x) sum(x == 0)
zero_vals <- apply(owls_sim, 2, sumfun)</pre>
dz <- as.data.frame(table(zero_vals)/nsim) %>%
    mutate(zero_vals = as.numeric(zero_vals))
obsval <- sumfun(Owls$SiblingNegotiation)</pre>
gsim <- ggplot(dz, aes(zero_vals, Freq)) + geom_point() +</pre>
    geom_segment(aes(xend = zero_vals, yend = 0)) +
    geom_vline(xintercept = obsval, col = "red") +
    annotate(geom = "text", x = obsval, y = 0.03,
        label = "observed", col = "red", hjust = 1.1)
```



complex models (Wickham et al. 2010; Gelman 2004; A. Buja et al. 2009)

```
simdat <- (simulate(m2, 8) %>% data.frame(speed = cars$speed) %>%
    gather(sample, dist, -speed))
ddsim <- (cars %>% select(dist, speed) %>% mutate(sample = "true") %>%
    bind_rows(simdat))
ddsimplot <- ggplot(ddsim, aes(speed, dist)) +</pre>
    geom_point() + facet_wrap(~sample)
```



references

Augustin, Nicole H., Erik-André Sauleau, and Simon N. Wood. 2012. "On Quantile Quantile Plots for Generalized Linear Models." Computational Statistics & Data Analysis 56 (8): 2404-9. doi:10.1016/j.csda.2012.01.026.

Buja, A., D. Cook, H. Hofmann, M. Lawrence, E.-K. Lee, D. F. Swayne, and H. Wickham. 2009. "Statistical Inference for Exploratory Data Analysis and Model Diagnostics." Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences 367 (1906): 4361-83. doi:10.1098/rsta.2009.0120.

Gelman, Andrew. 2004. "Exploratory Data Analysis for Complex Models." Journal of Computational and Graphical Statistics 13 (4): 755-79. doi:10.1198/106186004X11435.

Quinn, Gerry P., and Michael J. Keough. 2002. Experimental Design and Data Analysis for Biologists. Cambridge, England: Cambridge University Press.

Tibshirani, Rob. 1987. "Estimating Optimal Transformations for Regression." Journal of the American Statistical Association 83: 394. Wickham, H., D. Cook, H. Hofmann, and Andreas Buja. 2010.

"Graphical Inference for Infovis." IEEE Transactions on Visualization and Computer Graphics 16 (6): 973-79. doi:10.1109/TVCG.2010.161.

Zeileis, Achim. 2006. "Object-Oriented Computation of Sandwich Estimators." Journal of Statistical Software 16 (9): 1–16. http://www. jstatsoft.org/v16/i09/.