

Northern Pikeminnow Abundance in Deadwater Slough, Salmon River, Idaho, and Potential Impacts to Local Chinook Salmon

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Impact Statement

- 18 A large population of Northern Pikeminnow in a slow-water reach of the Salmon River, Idaho
- can potentially consume large numbers of juvenile Chinook Salmon, hindering ongoing recovery
- 20 efforts.

Abstract

22 Predation on emigrating juvenile salmonids by piscivorous fishes is a widely-studied source of 23 mortality within reservoirs and dam tailraces in the Columbia River basin. Native Northern 24 Pikeminnow have been estimated to consume ~8% of the approximately 200 million juvenile 25 salmonids emigrating through the lower Snake and Columbia rivers, annually. Less is known 26 about the interaction between upstream, river-dwelling Northern Pikeminnow and their impacts 27 on salmonid recovery above the reservoir systems. In this study, we examine the abundance of 28 Northern Pikeminnow in a slow-water reach of the Salmon River, Idaho, known as Deadwater 29 Slough, and consider their potential impacts on local Chinook Salmon populations. The 30 Deadwater Slough is within a migration corridor for several Chinook Salmon populations as well 31 as other Endangered Species Act-listings including steelhead and Sockeye Salmon. We estimated 32 the abundance of Northern Pikeminnow in Deadwater Slough, an approximately 1.9 kilometer 33 reach of the Salmon River, to be 19,499 in the fall and 10,352 in the spring, corresponding with 34 the peak emigration windows of juvenile Chinook Salmon. Using these abundance values, we 35 estimated Northern Pikeminnow consumption of juvenile Chinook Salmon. Assuming 60% of the Northern Pikeminnow diet is fish, of which 50% is juvenile Chinook Salmon, we estimated 36 37 the Northern Pikeminnow population can consume 61,409 juvenile Chinook Salmon, annually. 38 After performing a sensitivity analysis, we estimated Northern Pikeminnow predation on

- 39 juvenile Chinook Salmon was equivalent to 377 (95% CI: 161 - 935) returning adults. Given the
- 40 relatively small size of the Salmon River, the high densities and potential consumption rates of
- 41 Northern Pikeminnow in Deadwater Slough suggest predation likely has a consequential impact
- 42 on Chinook Salmon recovery in the Upper Salmon River.



INTRODUCTION

The Snake and Columbia rivers drain a network of inland tributaries that provide essential
spawning and rearing habitat for anadromous Pacific salmon <i>Oncorhynchus</i> spp. and steelhead
O. mykiss. These fishes were historically abundant throughout the basin and consequently have
tremendous ecological, cultural, and economic value (Lewis et al. 2019; Atlas et al. 2021). Over
the last century, salmonid stocks throughout the Snake and Columbia rivers have undergone
significant declines related to habitat modification in the freshwater environment (e.g., removal
of beavers, river channel simplification, water withdrawals, hydrosystem development) and poor
marine conditions, affecting their survival and recruitment at multiple life stages (Justice et al.
2017; Clark et al. 2020; Crozier et al. 2020). Consequently, action agencies have made
considerable investments in the rehabilitation of tributary ecosystems and evaluation of factors
attributed to the species decline (Roni et al. 2018; White et al. 2021).
Predation on emigrating juveniles by piscivorous fishes is another important, and potentially
under-estimated, source of salmonid mortality. In the lower mainstem Snake and Columbia
rivers it is estimated that predation on out-migrating salmonids during peak emigration has a
significant negative impact on the overall population and success of recovery efforts (Tabor et al
1993; Beamesderfer et al. 1996; Friesen and Ward 1999). Dams and reservoirs in the Columbia
River are the primary locations associated with high rates of piscine predation on salmonids
(Petersen 1994; Ward et al. 1995). There are generally two mechanisms that explain these high
predation zones. First, migration (movement) rates of juvenile salmon are reduced during
reservoir passage (Venditti et al. 2000), thereby increasing the time migrating smolts are
vulnerable to predation. Second, reservoirs and downstream tailraces associated with dams on

66	the Snake and Columbia rivers create favorable slow-water habitat for predatory fishes known to
67	consume juvenile salmonids, such as the Northern Pikeminnow Ptychocheilus oregonsis.
68	Northern Pikeminnow tolerate and thrive in relatively warm, slow-water habitats (Wydoski and
69	Whitney 2003) and thus have benefited from dams on the Columbia River, becoming abundant
70	predators of salmonid outmigrants (Knutsen and Ward 1999). Northern Pikeminnow are
71	estimated to consume ~8% (16.4 million) of the approximately 200 million juvenile salmonids
72	emigrating through the lower Snake and Columbia rivers, annually (Beamesderfer et al. 1996).
73	Most predation studies have focused on reservoirs (Murphy et al. 2021) and mainstem reaches
74	(Tabor et al. 1993; Ward et al. 1995; Shively et al. 1996; Zimmerman and Ward 1999) of the
75	Columbia River basin whereas considerably less is known about the interaction between
76	upstream, river-dwelling piscivorous fishes and their impacts on salmonid recovery above the
77	reservoir systems (Rubenson et al. 2020). Upstream habitats containing slower water velocities
78	and other attributes that support piscivorous predators may overlap with essential habitat for
79	some salmonid species and life stages. One such example is Deadwater Slough, an
80	approximately 1.9 km long reach of unnaturally slow and deep water in the Salmon River, Idaho.
81	The Deadwater Slough is within a section of the Salmon River containing historically important
82	overwinter rearing habitat for juvenile Chinook Salmon O. tshawytscha and is part of the
83	migratory pathway for upstream Chinook Salmon populations, the endangered Snake River
84	Sockeye Salmon O. nerka population (Axel et al. 2015), and several populations of threatened
85	Snake River steelhead. The slough is also inhabited by piscivorous predators, including Northern
86	Pikeminnow and Smallmouth Bass Micropterus dolomieu, that are potential sources of mortality
87	for rearing and migrating salmonids.

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The Salmon River was historically the most productive tributary for Chinook Salmon in the Columbia River basin (Nemeth and Kiefer 1999), but those populations have become depleted in recent decades. Therefore, quantifying the mortality of spring/summer-run Chinook Salmon (hereafter Chinook Salmon) in the Upper Salmon River associated with piscine predation is of particular interest. Importantly, the Salmon River supports eight extant populations of Chinook Salmon upstream of Deadwater Slough (National Oceanic and Atmospheric Administration 2017). This includes the Lemhi River, which was historically the largest population, and is a prioritized candidate for restoration of natural processes to increase production of juvenile Chinook Salmon (Zimmerman et al. 2012). Chinook Salmon in the Upper Salmon River are stream-type and exhibit two distinct migration tactics; downstream rearing (DSR) and natal reach rearing (NRR) (Copeland et al. 2014). The DSR migrants leave their natal area as subyearlings between June and November and typically overwinter in downstream, mainstem habitats until the following spring when they emigrate to the ocean as smolts. Alternatively, NRR migrants remain in their natal areas for approximately one year after emergence until emigration to the ocean as smolts. Diversity of migration tactics provides a mechanism for coping with adverse conditions in freshwater rearing and migration environments and buffers against catastrophic events, thereby increasing population resiliency (Dodson et al. 2013). Deadwater Slough represents an important habitat for Chinook Salmon in the Upper Salmon River as it supports rearing and migration of juveniles from all upstream populations including fall DSR and spring NRR migrants. Recent studies have examined the downstream movement, distribution, and apparent survival (hereafter 'survival') of juvenile salmonid emigrants through the Salmon River, including the Deadwater Slough reach. Sockeye Salmon migrating through the mainstem Salmon River during

- 111 spring were estimated to have 32% lower survival in the Deadwater Slough reach compared to 112 directly adjacent reaches (Axel et al. 2015). Similarly, emigrating DSR Chinook Salmon had an 113 approximate 10% reduction in transition probability through the Deadwater Slough compared to 114 surrounding reaches during fall and early winter months (Ackerman et al. 2018; Porter et al. 115 2019). Low survival was attributed to lack of fish cover and low-velocity water delaying 116 movement rates, thereby increasing predation risk. 117 In this study, we estimated the abundance of Northern Pikeminnow in Deadwater Slough and 118 evaluated potential impacts to juvenile salmon emigrants, focusing on DSR and NRR Chinook 119 Salmon. We hypothesized that high densities of piscivorous predators in the Deadwater Slough 120 might explain the reduced survival observed for juvenile Chinook Salmon. To test this, our 121 objectives were four-fold: 122 Estimate the abundance of Northern Pikeminnow in Deadwater Slough during the peaks 1. 123 of fall DSR and spring NRR juvenile emigrations;
- Document predation on juvenile Chinook Salmon during the DSR and NRR emigration
 periods using gastric lavage;

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- Use an established bioenergetics approach to estimate consumption potential (grams) of the Northern Pikeminnow population on DSR and NRR Chinook Salmon emigrants at Deadwater Slough;
- 4. Estimate how consumption of juvenile Chinook Salmon emigrants at Deadwater Slough
 by Northern Pikeminnow might impact adult returns to the Upper Salmon River.

METHODS

Study Site

The Deadwater Slough is an approximately 1.9 km section of the mainstem Salmon River located roughly 6 river kilometers (rkm) downstream from the town of North Fork, Idaho (Figure 1). The downstream end of the slough is located at the confluence of Dump Creek and the Salmon River. A large alluvial fan at the mouth of Dump Creek has, at least partially, created a hydraulic control in the Salmon River resulting in the formation of Deadwater Slough. Although the origin and timing of the alluvial fan and Deadwater Slough is somewhat ambiguous (Reichmuth et al. 1985; USACE 1986), some believe they coincided with the failure of a small mining reservoir in the Dump Creek drainage around 1897 that deposited substantial amounts of sediment at its confluence with the Salmon River (Emerson 1973). The slough is currently a slow, deep section in the river, spanning approximately 12 hectares, averaging 68 m wide, and has characteristics resembling a small reservoir.

Northern Pikeminnow Demographics

We estimated the population size of piscine predators in the Deadwater Slough using a mark-recapture survey design and a catch per unit effort (CPUE) approach. Predators were sampled near the peaks of the fall DSR and spring NRR emigrations. During our initial survey in 2019, Northern Pikeminnow were the most prevalent piscine predator in Deadwater Slough and consequently became our focal taxa. Our intent was to survey during the emigration of the 2018 Chinook Salmon brood year that occurred fall 2019 and spring 2020, however logistical constraints during the onset of the COVID-19 pandemic delayed the spring 2020 survey until

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2021. Over this period, an additional fall survey occurred during the 2020 DSR emigration. Fall surveys were constrained to two weeks to minimize Northern Pikeminnow immigration and emigration. Multiple capture methods were employed during the fall 2019 survey to reduce selectivity and bias for species and size classes. Methods included raft electrofishing, fyke netting, snorkeling, and angling. After evaluating all methods, angling was the most effective method for capturing Northern Pikeminnow while also minimizing potential impacts to Endangered Species Act (ESA)-listed adult steelhead that were present during our surveys. Less effective methods were abandoned in subsequent years and the following analyses will focus on fish captured by angling, unless otherwise noted. Our study relied heavily on volunteer anglers who were permitted to fish anywhere within Deadwater Slough. Each survey day, anglers boated or hiked their catch (periodically or upon filling a livewell) to a processing station at the boat ramp, approximately 500 m downstream from the top of the slough. For each fish, we recorded the date of capture, species, total length (TL; mm), and whether the fish was previously marked. Unmarked fish were given a physical mark (e.g., hole punch of lower caudal, upper caudal, left pelvic, right pelvic) unique to each day and then were released. Fish release sites were distributed throughout Deadwater Slough to facilitate mixing back into the population. Fish that died prior to release were included in that day's total capture (or recapture) count, but not included in the number of marked fish available for recapture after that date. Finally, we recorded the angling start and end time for each crew (person or combinations of persons) to calculate CPUE as the number of Northern Pikeminnow caught per angler hour.

Two types of mark-recapture estimators were used to explore variation in abundance estimates: a single census and a multiple census estimator. For the single census estimator, we treated the first week of the survey as the mark event and the following week as the recapture event, pooling data within each week. Alternatively, the multiple census estimator treated each day as a sampling event and used information about the total number of marked fish from all previous events to calculate the total abundance. For the single census estimator, we used the Chapman-modified Lincoln-Peterson estimator:

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$$\hat{N} = \frac{(M+1)(n+1)}{(m+1)} - 1$$

where M is the total number of fish marked and returned to the population during the first event (week), n is the total number of fish caught in the second event (week), and m is the number of marked fish caught during the second week. For a multiple census estimator, we used the Chapman-modified Schnabel estimator:

$$\hat{N} = \frac{\sum_{i=1}^{k} n_i M_i}{\left(\sum_{i=1}^{k} m_i\right) + 1}$$

where *M*, *n*, and *m* are indexed by each survey (day), *i*. The 95% confidence intervals of the Schnabel estimator were calculated using a Poisson approximation (Krebs 1999).

Because fish capture was dependent on volunteer anglers, there was concern that sampling effort was biased to locations preferred by anglers. If release sites for marked fish were not proximal to preferred angling locales, recently released fish may have been less susceptible to immediate capture than unmarked fish. To account for this potential bias, we included an adjusted "delayed-

mixing" Schnabel estimator that assumed fish marked and released on a given survey day were not available for recapture for 48 hours. The delayed-mixing Schnabel estimator is given by:

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$$\hat{N} = \frac{\sum_{i=1}^{k} n_i M_{i-1}}{\left(\sum_{i=1}^{k} m_i\right) + 1}$$

All mark-recapture abundance estimators include a Chapman correction for small sample sizes (Chapman 1951) and assumed: (1) the population is closed (no immigration, emigration, births or deaths during the survey period), (2) all fish have equal chance of being caught in subsequent survey days, (3) marking a fish does not affect its chances of recapture, (4) no loss of marks, and (5) marks are not missed or misidentified.

Surveys conducted in spring 2021 did not follow a mark-recapture study design so Northern Pikeminnow abundance was estimated as the ratio of mean total CPUE in the fall surveys to the total CPUE in the spring survey, then multiplied that ratio by the mean abundance from fall surveys:

$$\hat{N}_{spring} = \overline{N}_{fall} * \frac{CPUE_{spring}}{CPUE_{fall}}$$

This approach assumes equal capture probabilities for fall and spring surveys. Fall and spring population estimates were converted into linear and areal densities by dividing by the length and area of Deadwater Slough, respectively. Length and area were measured in QGIS software (QGIS Development Team 2022) and using drone-generated orthoimagery.

Anglers reported that Northern Pikeminnow in Deadwater Slough were large-bodied. To describe the population size structure, we calculated the proportional stock density (PSD) for

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Northern Pikeminnow in Deadwater Slough. PSD is the percentage of all fish that meet criteria for "quality-length" (larger) individuals and "stock-length" individuals:

$$PSD = 100 * \frac{FQ}{FS}$$

- where FQ is the number of fish \geq quality-length, and FS is the number of fish \geq stock-length.
- For Northern Pikeminnow in Deadwater Slough, we used 380 mm TL for quality-length and 250
- 217 mm TL for stock-length (Winther et al. 2020).

Stomach Contents

Gastric lavage (Foster 1977) was used to examine the stomach contents of Northern Pikeminnow for the presence of juvenile Chinook Salmon and other fishes occupying Deadwater Slough. Immediately following lavage, stomach contents of individuals were preserved with 99% isopropyl alcohol in whirl-paks to be analyzed in a controlled environment. For each stomach sample, total wet weight (grams) was recorded, including all fish and non-fish items (e.g., macroinvertebrates, organic matter). Fish and fish remnants were identified to the lowest taxonomic unit using diagnostic bones (Hansel et al. 1988; Frost 2000) or were categorized as unknown. Approximately 5% of Northern Pikeminnow were euthanized for dissection (n = 75) after gastric lavage to validate the efficacy of the methodology. Contents of livewells were examined during fish processing to ensure food items were not regurgitated.

Fish Consumption Potential

To estimate the total consumption potential of Northern Pikeminnow in Deadwater Slough during the peaks of fall DSR and spring NRR emigrations, we used the Fish Bioenergetics v4.0 application developed by Deslauriers et al. (2017) applied in R statistical software (R Core Team 2021). The daily rate of consumption in grams for an individual Northern Pikeminnow was estimated based on predator and prey energy densities, predator start and end weights, and water temperatures. Separate models were run for the fall period (September 15 - November 30) and the spring period (March 1 - May 31) to coincide with peak emigrations of DSR and NRR juveniles from the Lemhi River, the largest Chinook Salmon population in the Upper Salmon River.

Predator energy density for Northern Pikeminnow was fixed at 6,703 Joules (J)/g (Deslauriers et al. 2017). Prey energy densities were fixed at 3,000 J/g for invertebrates and 21,500 J/g for juvenile Chinook Salmon (Moss et al. 2016). Because we were unable to differentiate juvenile Chinook Salmon from other fish prey in model runs, we assume all fish prey have the same energy densities as juvenile Chinook Salmon. The average TL of Northern Pikeminnow caught in Deadwater Slough during our study was converted to fork length (FL) and then to weight (grams) using a weight-length formula from Parker et al. (1995).

$$log_{10}W_s = -4.886 + 2.986[log_{10}(FL)]$$

This resulted in average fall DSR and spring NRR starting weights of 598.7 and 430.7 g for Northern Pikeminnow with average TLs of 394.1 and 352.9 mm, respectively. We assumed no growth in individual Northern Pikeminnow, resulting in equal start and end weights. Mean daily water temperatures were summarized from 15-minute interval temperature readings between March 3, 2013 and June 14, 2021 from USGS gage station 13307000, approximately 22 rkm downstream of Deadwater Slough.

The proportion of fish in the Northern Pikeminnow's diet relative to non-fish prey items is unknown in Deadwater Slough. Therefore, we conducted a series of model runs with varying proportions of fish in the diet ranging from 30 - 90%, in 10% increments. These values were supported by observations elsewhere in the Columbia River basin (54-86%: Shively et al. (1996); 48-86%: Zimmerman and Ward (1999); 37%: Gray and Dauble (2001)) that suggest fish are the majority of Northern Pikeminnow prey by volume. A model run was conducted for each combination of diet scenario and season, resulting in fourteen estimates of the total grams of fish consumed by an individual Northern Pikeminnow. To estimate the total biomass of fish consumed by an individual Northern Pikeminnow by the estimated Northern Pikeminnow population sizes during fall DSR and spring NRR emigrations.

Impacts to Chinook Salmon Populations

The proportion of Chinook Salmon prey relative to other fish prey in the Northern Pikeminnow diet is also unknown. However, there is some evidence that juvenile salmonids are by far the most consumed fish prey (Shively et al. 1996; Zimmerman and Ward 1999). Moreover, it is estimated that Chinook Salmon make up 64.2%, 29.3%, and 49.3% of the fish prey consumed by Northern Pikeminnow in the Columbia River below Bonneville Dam, in Columbia River reservoirs, and in the lower Snake River, respectively (Zimmerman and Ward 1999). Because the bioenergetics model does not differentiate juvenile Chinook Salmon from other fish prey items, we performed a sensitivity analysis to assess potential impacts of Northern Pikeminnow predation on local Chinook Salmon populations. Using values similar to Zimmerman and Ward (1999), we modeled three diet scenarios where Chinook Salmon comprised 30%, 50%, and 65%

of the fish prey, by weight, in the Northern Pikeminnow diet during the fall DSR and spring
NRR emigration periods. In this analysis, we assume 60% of the Northern Pikeminnow's total
diet is comprised of fish, representing the median scenario in our range of modeled diets.
We obtained an estimate of the number of juvenile Chinook Salmon consumed by dividing total
biomass consumed by the average weight of DSR (10.3 g) and NRR (10.9 g) emigrants in the
Upper Salmon River. The average weights of DSR and NRR emigrants were calculated from fish
captured at seven rotary screw traps upstream of Deadwater Slough during the fall and spring
periods. Although the primary impact to Chinook Salmon in Deadwater Slough by Northern
Pikeminnow is juvenile predation, the common metric to evaluate salmon recovery is adult
returns. We quantified the potential impact of Northern Pikeminnow predation on adult returns
by estimating the number of equivalent adults expected to return to Lower Granite Dam if
predation by Pikeminnow was eliminated in Deadwater Slough. To accomplish this, we
multiplied the estimated total juvenile Chinook Salmon consumed by the median Granite-to-
Granite smolt-to-adult return rate (SAR) of 0.00614 (SD = 0.00051) from McCann et al. (2019)
for Chinook Salmon in the Upper Salmon River.
All data and code for the analyses presented here can be found in a GitHub repository release
(https://doi.org/10.5281/zenodo.6458124).

RESULTS

Northern Pikeminnow Demographics

Using hook-and-line angling, we caught a total of 1,663 Northern Pikeminnow over the course of the study including 14 recaptures; the overall CPUE was 1.32 Northern Pikeminnow per angler

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hour (Table 1). Mark-recapture abundance estimates of Northern Pikeminnow in Deadwater Slough ranged from 12,480 to 18,732 in fall 2019 and from 24,381 to 37,016 in fall 2020 (Table 3). We estimated larger populations of Northern Pikeminnow using the unadjusted multiple census estimator compared to the single census and adjusted delayed-mixing multiple census estimators (Figure 2). Our sampling design most closely matched a multiple census estimator; therefore, the Schnabel estimates were considered most appropriate. Accordingly, the mean Northern Pikeminnow abundance for the two fall sampling events was 27,874 (95% CI: 14,244 -59,388) using the unadjusted Schabel estimator. Using the delayed-mixing Schnabel estimator the mean fall abundance estimate was 19,499 (95% CI: 9,952 - 41,597). All subsequent analyses use results from the adjusted delayed-mixing Schnabel estimator. For spring 2021, we estimated 10,352 (95% CI: 5,284 - 22,084) Northern Pikeminnow in Deadwater Slough. Those estimates translate to linear densities of 10,422 and 5,533 Northern Pikeminnow per rkm and areal densities (fish per 100 m²) of 16.3 and 8.7 for fall and spring, respectively. Lengths of Northern Pikeminnow ranged from 176 to 639 mm TL with an average of 389 mm (Figure 3). The PSD for Northern Pikeminnow in Deadwater Slough across all three surveys was 50%.

Stomach Contents

We found that gastric lavage successfully removed food items in all dissected individuals, indicating the method was effective. Of the 1,558 Northern Pikeminnow sampled using gastric lavage, we found contents in 350 stomachs and confirmed fish or fish parts in 44 of those.

Northern Pikeminnow captured during spring 2021 had a higher proportion of fish content in their diet compared to individuals captured in the fall surveys (Table 4). The mean wet weight of

total contents for an individual was 0.98 g (median = 0.25 g; SD = 2.16 g). Overall, fish or fish remnants made up 11.7% of all stomach contents examined.

Fish Consumption Potential

During the fall DSR emigration, we estimated an average-size Northern Pikeminnow (394.1 TL mm) to consume 43.55 g of fish to maintain their body size, assuming 60% of their diet consisted of fish prey (Figure 4). Fall consumption ranged from 35.74 g for a diet of 30% fish to 46.91 g for a diet of 90% fish. During the spring NRR emigration, we estimated an average-size Northern Pikeminnow (352.9 TL mm) to consume 42.51 g of fish, assuming 60% of their diet was fish prey (Figure 4). Spring consumption ranged from 34.01 for a diet of 30% fish to 44.33 for a diet of 90% fish. Daily consumption rates were higher early in the fall and late in the spring and corresponded with higher water temperatures, especially above 8 degrees Celsius (Figure 4). Given estimated mean Northern Pikeminnow population sizes of 19,499 in the fall and 10,352 in the spring 2021, and a diet consisting of 60% fish, the Northern Pikeminnow population was estimated to consume 0.8 metric tons of fish during the fall DSR emigration and 0.4 metric tons during the spring NRR emigration in Deadwater Slough.

Impacts to Chinook Salmon Populations

Given yearly fish consumption estimates during the fall DSR and spring NRR periods, we produced a sensitivity analysis to calculate the consumption of juvenile Chinook Salmon where 30%, 50%, and 65% of total fish prey consumed were juvenile Chinook Salmon (Figure 5). At the median value of 50%, we estimated that 61,409 (95% CI: 31,342 - 131,004) juvenile Chinook Salmon would be consumed. Using the median Granite-to-Granite SAR for Chinook

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Salmon in the Upper Salmon River, we estimated the "adult equivalents" of juveniles consumed to be 377 (95% CI: 161 - 935) adults (Figure 6).

DISCUSSION

All mark-recapture abundance estimators suggest a large population of Northern Pikeminnow occupies Deadwater Slough. The mean population size of Northern Pikeminnow was estimated to be 19,499 during the fall DSR emigration and 10,352 during the spring NRR emigration even after accounting for the potential of delayed mixing of marked and released fish. Those estimates translate to linear densities (fish per rkm) that are 15 and 8 times greater than densities reported by Beamesderfer and Rieman (1991) for the John Day Reservoir and approximately 4 and 2 times greater than projections for the lower Columbia River (Beamesderfer et al. 1996). Beamesderfer and Rieman (1991) acknowledged that their methods were unsuitable for sampling offshore, noting that water velocity, depth, and irregular bottom contours, and barge traffic made sampling offshore ineffective; therefore, it is possible that their estimates may have been low. Conversely, the maximum depth at Deadwater Slough was approximately 6 m with a relatively homogenous bottom contour, making angling an effective method throughout the entire reach. Our estimated densities of Northern Pikeminnow suggest that slow-water reaches outside of reservoir complexes on the Snake and Columbia rivers may support exceptionally high predator densities, consistent with findings of Harnish et al. (2014), Gray and Dauble (2001), and Zimmerman and Ward (1999). Given the dearth of current data available throughout the Columbia River basin, it is unclear how Northern Pikeminnow abundance estimates in this study compare to elsewhere in the basin; additional information on contemporary piscine predator abundances may be needed (Widener et al. 2021).

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Several assumptions in our estimators may have influenced the magnitude of our abundance results. First, all models assumed a closed population; however, it is possible that immigration and emigration occurred. Nevertheless, our estimators still provide unbiased estimates of abundance assuming the immigration and emigration rates were equal between marked and unmarked fish. Emigration of marked individuals could reduce the marking fraction in the population leading to an upward bias in abundance estimates, but emigration rates would have needed to be substantial. We also captured two fish with marks from previous years, indicating seasonal residency of some fish. The multiple census estimators, which we report, are more robust to this assumption because the marking fraction was estimated daily. We additionally have little reason to believe that emigration occurs during the two-week survey window owing to favorable habitat for Northern Pikeminnow in Deadwater Slough relative to adjacent reaches. Given the size of the sampling area and the short duration of our surveys, the closed population assumption was likely met. This assumption could likewise have affected results if mortality occurred for some marked fish released back to the population. During field processing, no mortalities or injuries post release were observed and fish injured prior to release were retained. Nevertheless, the potential for mortality amongst marked and released fish cannot be discounted. Next, our abundance estimates assumed that capture and recapture events are random samples of the population. Angling methods often have a size selection bias, thereby limiting abundance estimates to a size range susceptible to angling, resulting in a conservative estimate of the total population. The estimators also assumed equal catchability of individual fish between sampling events; two fish were recaptured during multiple days within a survey, indicating that marked fish were continually susceptible to angling. It is also possible that marked and released fish may not have immediately mixed thoroughly back into the population. Although we attempted to

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Northern Pikeminnow.

release marked fish evenly throughout Deadwater Slough, we anecdotally observed anglers congregating in particular areas of the slough to socialize or exploit "good" fishing locales. Violations of the assumption of equal catchability between marked and unmarked fish may have led to an overestimate of abundance of Northern Pikeminnow in our study if marked fish have lower catchability. This bias will be proportional to the difference in catchability of marked and unmarked fish. As an example, if marked fish were only 70% as likely to be caught as unmarked fish, the true abundance is closer to 70% of our abundance estimate. This would additionally account for marked and released fish being "hook shy" for a period of time. The adjusted delayed-mixing Schnabel estimator may account for these biases; however, the magnitude of bias is unknown. The spring abundance estimate was smaller than fall abundance estimates due to the spring CPUE being approximately half of the mean fall CPUE (Table 1). Although our approach assumed equal capture probabilities between the fall and spring surveys, we believe that higher spring flows may result in lower capture probabilities of Northern Pikeminnow. Therefore, we believe our spring abundance estimate to be conservative. In addition to the large population abundance, the observed (50%) PSD in this study was greater than observations of 41% below Bonneville Dam and 18% in Bonneville Reservoir (Winther et al. 2020). This suggests that a larger fraction of Northern Pikeminnow in Deadwater Slough are of a quality size relative to populations reported elsewhere in the Columbia River. Notably, the Idaho state catch-and-release record Northern Pikeminnow, measuring 639 mm TL, was caught in Deadwater Slough during the fall 2020 survey alluding to the favorable conditions for

We performed gastric lavage on nearly all Northern Pikeminnow collected during this study.
Observed prey species included juvenile Chinook Salmon as well as Redside Shiner
Richardsonius balteatus, Largescale Sucker Catostomus macrocheilus, sculpin Cottus spp., and
Mountain Whitefish Prosopium williamsoni, though decomposition from digestion rendered
most fish contents unidentifiable. Although our approach provided useful information on prey
species, the use of angling and gastric lavage to quantify diet composition in Northern
Pikeminnow can be problematic. First, diets of angled fish can differ from those collected by
other methods such as electrofishing (Hodgson and Cochran 1988). For instance, angled fish are
more likely to have empty stomachs (Jurajda et al. 2016); possibly because they are captured
while actively searching for food, suggesting they are hungry. Second, unlike other predators
such as Smallmouth Bass and Walleye, that are commonly sampled using gastric lavage (Kamler
and Pope 2001), Northern Pikeminnow are cyprinids and therefore lack a true stomach.
Consequently, performing gastric lavage on cyprinid species can be ineffective for accurately
quantifying diet composition (Hartleb and Moring 1995), or alternately, requires a modification
in methodology to flush food items out the vent of the fish (Wasowicz and Valdez 1994).
Therefore, we consider our diet composition estimates from gastric lavage to be conservative.
Our bioenergetics approach assumed that all available prey consumed by Northern Pikeminnow
have energy densities equal to juvenile Chinook Salmon. Generalizing energy densities using a
single species is a common approach (Petersen and Ward 1999). Other salmonids including
juvenile steelhead and Sockeye Salmon are likely also available prey in Deadwater Slough,
especially during the spring migration, and so generalizing energy densities among salmonids
may not be problematic. Additionally, hatchery smolt releases (Chinook Salmon, steelhead, and
Sockeye Salmon) are also prevalent in the mainstem Salmon River during the spring migration.

To address the prevalence of other fish in the Northern Pikeminnow's diet, we considered
scenarios where DSR and NRR Chinook Salmon represent only 30%, 50%, and 65% of total fish
prey consumed. These values may be considered conservative during the peak emigration
periods when Chinook Salmon are abundant and the Northern Pikeminnow diet shifts almost
entirely to piscivory, presumably on juvenile Chinook Salmon (Poe et al. 1991; Shively et al.
1996). This is likely the case during fall months when DSR Chinook Salmon are the dominant
prey species available within the Salmon River and Deadwater Slough as 1) few to no hatchery
releases are present in the river and 2) fall emigrations of steelhead and Sockeye Salmon are less
prominent than Chinook Salmon. During spring when natural-origin steelhead and Sockeye
Salmon are also actively emigrating from the Upper Salmon River and hatchery-origin releases
of all three species (Chinook Salmon, steelhead, Sockeye Salmon) are present in the river, less
than 50% Chinook Salmon in the Northern Pikeminnow's diet may be more likely. Future work
to quantify and identify juvenile salmonids in Deadwater Slough during their seasonal migration
would be useful to validate the diet composition assumptions used in our model and to
understand impacts to local populations.
Chinook Salmon populations above Deadwater Slough are within the Upper Salmon major
population group (MPG) which supports eight independent, extant populations including Salmon
River (above Redfish Lake Creek), Valley Creek, Yankee Fork Salmon River, East Fork Salmon
River, Salmon River (mainstem below Redfish Lake Creek), Pahsimeroi River, Lemhi River, and
North Fork Salmon River (National Oceanic and Atmospheric Administration 2017). Recovery
of the MPG is desired to support local fisheries and economies. At least five of the eight
populations must meet criteria set forth by McElhany et al. (2000) and the Interior Columbia
Technical Recovery Team (2007) for the MPG to be considered viable and for recovery of the

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Snake River Evolutionary Significant Unit. We estimated that 61,409 juvenile Chinook Salmon may be consumed by Northern Pikeminnow assuming that 60% of their diet is fish and 50% of fish prey are Chinook Salmon. For context, this is approximately 1.35 times the average annual combined DSR and NRR emigration estimates for brood years 2011 - 2018 at a rotary screw trap located in the lower Lemhi River (Poole et al. 2019; Feeken et al. 2020; McClure et al. 2021), the largest population in the Upper Salmon MPG. Even under the most conservative scenario where 30% of the Northern Pikeminnow diet is fish and 30% of fish prey are Chinook Salmon, we estimate that 29,988 juvenile Chinook Salmon may be consumed which is 66% of the total DSR and NRR emigration for brood years 2011 - 2018. Considering that diet scenarios used in our study are likely conservative assumptions, especially during the fall DSR emigration, the estimated number of Chinook Salmon consumed is substantial. Spawner abundance is perhaps the most important metric considered in determining a population's viability and productivity. We estimated that consumption of juvenile Chinook Salmon by Northern Pikeminnow in the Deadwater Slough is equivalent to 377 adults, annually, which is 68% of the total mean spawner escapement for the Upper Salmon MPG in 2017 – 2019 (Kinzer et al. 2020). In the most conservative scenario, we estimated juvenile consumption equivalent to 184 adults, which is 33% of recent natural-origin adult Chinook Salmon returns. In the least conservative scenarios, adult equivalents approached recent escapements for the entire Upper Salmon MPG. The sensitivity analysis used some simplifying assumptions; for example, it assumed no juvenile mortality between Deadwater Slough and Lower Granite Dam which is an unrealistic assumption. Nevertheless, we found converting juveniles consumed to "adult equivalents" a useful exercise to place results in context with a metric commonly used for recovery.

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In addition to juvenile Chinook Salmon, juvenile steelhead and Sockeye Salmon (both naturalorigin and hatchery releases) are also likely prey items for Northern Pikeminnow in Deadwater Slough. Accordingly, we surmise that juvenile emigration and adult returns of steelhead and Sockeye Salmon are also affected by predation, including hatchery populations, which provide for recreational fishing opportunities in the Upper Salmon River. Consequently, reducing predation mortality in Deadwater Slough could potentially benefit multiple upriver natural and hatchery populations, including other ESA-listed species. Because Deadwater Slough is part of the migratory pathway for multiple species and populations of emigrating salmonids, the positive impact could be greater than individual tributary rehabilitation actions which typically benefit a single population. Predation on juveniles from any of the ESA-listed salmonid species is likely detrimental to their recovery. Deadwater Slough is a favorable candidate for management or restoration actions to benefit local Chinook Salmon populations. Three potential management actions could reduce predation at Deadwater Slough: 1) removing or reducing the Dump Creek alluvial fan, 2) a local Northern Pikeminnow bounty program to encourage harvest in Deadwater Slough aimed at reducing the predator population size, and 3) adding structure or cover within Deadwater Slough to provide refuge for juvenile salmonids to reduce predation rates. Reducing or removing the Dump Creek alluvial fan has the benefit of restoring natural fluvial processes in the Salmon River that likely existed in the reach prior to the formation or increase in size of the alluvial fan. Restoring natural processes could reduce Northern Pikeminnow densities to levels similar to upstream and downstream reaches where higher survival and transition probabilities for juvenile salmon have been observed relative to Deadwater Slough (Axel et al. 2015; Ackerman et al. 2018; Porter et al. 2019). Managers ought also to consider the feasibility and net benefit of restoring fluvial

498 processes relative to potential losses for recreational fishing and bird watching opportunities 499 (https://www.audobon.org/important-bird-areas/deadwater-slough). 500 Northern Pikeminnow bounty programs have proven successful at reducing population sizes and 501 impacts on emigrating salmonids in the Columbia River (Winther et al. 2020). A local Northern 502 Pikeminnow bounty program could provide monetary incentive for sportfisherman, in addition to 503 boosting the local economy from lost revenues. Bounty programs could be conducted year-round 504 or seasonally to coincide with peak juvenile outmigration(s). Each approach would require 505 continual/annual effort and harvest to suppress the population size and as such, appropriate 506 funding would be needed to support such a program. Lastly, a more passive approach could be to 507 improve cover or structure within Deadwater Slough that provides refugia for juvenile salmonids 508 during their emigration. Cover would need to be appropriately sized to provide concealment to 509 juvenile fishes while reducing access by larger fishes like Northern Pikeminnow. Adding cover 510 could be a cost-effective approach; however, its potential effectiveness is unclear. 511 Although the origins of Deadwater Slough are somewhat ambiguous (Reichmuth et al. 1985; 512 USACE 1986), it appears likely that human activities in Dump Creek have either exacerbated or 513 contributed to its formation, and in turn, created favorable conditions for Northern Pikeminnow. 514 Among the three surveys, our lowest population abundance estimate was greater than 10,000 515 Northern Pikeminnow occupying Deadwater Slough, suggesting a remarkably high density given 516 the size of the area. The slow water velocity and lack of cover for fish also create conditions 517 where juvenile salmonids that are rearing in or emigrating through Deadwater Slough, including 518 Chinook Salmon, are susceptible to predation. We estimate that, at a minimum, Northern 519 Pikeminnow in Deadwater Slough consume greater than 15,000 juvenile Chinook Salmon (the 520 lower endpoint of the confidence interval for the most conservative scenario); however, it is

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likely that this value is closer to 61,000, annually. The magnitude of predation by Northern Pikeminnow is equivalent to 33% and 94% of recent adult escapements. Deadwater Slough predates the ESA-listing of Chinook Salmon populations in the Upper Salmon MPG in the 1990s and is therefore unlikely to be the primary cause for the population's decline. However, predation by Northern Pikeminnow in Deadwater Slough and elsewhere should be considered ing recov among limiting factors hindering recovery efforts of ESA-listed Chinook Salmon populations in the Upper Salmon MPG.

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727 **TABLES**

728

729

TABLE 1. Summary of Northern Pikeminnow captures from angling surveys in Deadwater Slough. Fall surveys included mark and recapture weeks for the single census estimate. Spring surveys included only a single capture week to estimate CPUE.

Survey Week	Date	Fish Caught	Marked Fish Caught	Marked Fish Returned	CPUE
			Fall 2019		
	Nov 12	29	0	28	
1	Nov 13	146	0	146	
	Nov 14	93	1	93	
					1.83
	Nov 19	149	2	132	
2	Nov 20	104	1	77	
	Nov 21	143	4	118	
			Fall 2020		
	Oct 20	173	0	170	
1	Oct 21	188	1	187	
1	Oct 22	104	0	102	
	Oct 23	41	0	41	
					1.23
	Oct 27	42	0	41	
2	Oct 28	47	1	46	
2	Oct 29	157	4	156	
	Oct 30	45	0	45	
			Spring 2021		
	May 18	85	-	-	
1	May 19	64	-	-	
1	May 20	41	-	-	
	May 21	12	-	-	
					0.81

TABLE 2. Estimated fish consumed by an individual Northern Pikeminnow to maintain its body weight during the fall DSR and spring NRR Chinook Salmon emigrations. Bioenergetic model results for each season include seven model runs that assume different percentages of fish in the diet. Northern Pikeminnow start and end weights were 598.7 and 430.7 g for the fall and spring seasons, respectively.

		E: 1
	Fish in Diet	g Fish
Date Range	(%)	Consumed
		(g)
	Fall	
	30	35.74
	40	39.37
	50	41.75
Sep 15 - Nov 30	60	43.55
	70	44.88
	80	45.98
	90	46.91
	Spring	
	30	34.01
	40	39.54
	50	39.54
Mar 1 - May 31	60	42.51
	70	42.51
	80	43.52
	90	44.33

TABLE 3. Estimated Northern Pikeminnow in Deadwater Slough and the associated standard
 errors and 95% confidence intervals from mark-recapture estimators calculated for the fall
 surveys. Standard errors are not available for the multiple census estimators.

Chapman			
Chapman		Fall 20	19
Chapman	13,298	4,322	6,898 - 27,893
Schnabel	18,732	NA	10,057 - 37,851
Schnabel - Delayed Mixing	12,480	NA	6,701 - 25,219
		Fall 202	20
Chapman	24,381	9,066	11,547 - 55,761
Schnabel	37,016	NA	18,430 - 80,924
Schnabel - Delayed Mixing	26,518	NA	13,203 - 57,975

TABLE 4. Total number of Northern Pikeminnow gastric lavaged and the number of those
 individuals with contents (including fish) and with contents that include fish.

Curror	Lavaged	Stomach	Fish
Survey	Lavaged	Contents	Contents
Fall 2019	660	57	12
Fall 2020	793	188	25
Spring 2021	105	105	7
Total	1,558	350	44

FIGURES

746	FIGURE 1. Map of the Deadwater Slough study area within the Upper Salmon River MPG. The
747	Dump Creek alluvial fan is located at the downstream end of the study reach.
748	FIGURE 2. Estimated abundance of Northern Pikeminnow in Deadwater Slough from three
749	mark-recapture estimators for the fall surveys. Error bars indicate 95% confidence intervals.
750	FIGURE 3. Length frequency histogram of Northern Pikeminnow caught using hook-and-line
751	angling during the study.
752	FIGURE 4. The cumulative fish prey consumed (g) by an individual Northern Pikeminnow
753	during the peak Chinook Salmon emigrations for fall DSR (top left) and spring NRR (top right)
754	and corresponding daily mean water temperatures (bottom panels). The black line shows the
755	consumption by Northern Pikeminnow when the diet consists of 60% fish and the gray area
756	shows the range between all model runs in the series, from 30% to 90% fish prey. Daily mean
757	water temperatures were summarized from six years of data available from the Shoup gage
758	station.
759	FIGURE 5. Estimated number of juvenile Chinook Salmon consumed by Northern Pikeminnow
760	from a sensitivity analysis of diet scenarios made up of variable percentages of fish. Diet
761	scenarios include 30%, 50%, and 65% Chinook Salmon in the total fish prey consumed by
762	Northern Pikeminnow, similar to those reported by Zimmerman and Ward 1999.
763	FIGURE 6. Estimated "adult equivalents" of the total juvenile Chinook Salmon consumed by
764	Northern Pikeminnow in Deadwater Slough. Error bars indicate 95% confidence intervals.
765	

Colophon

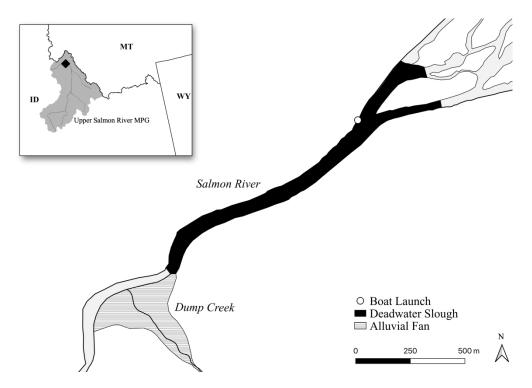
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772
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                Windows 10 x64 (build 22000)
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774
     #> system
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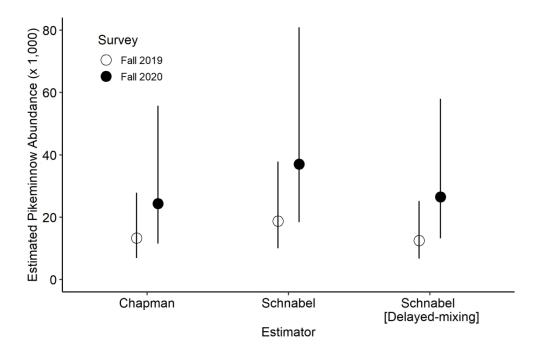
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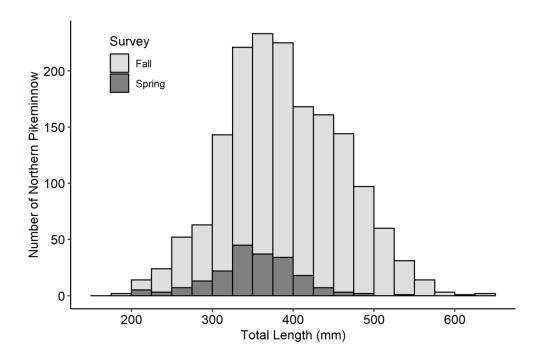
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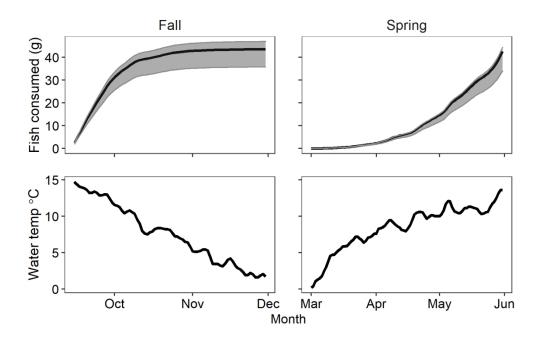
Map of the Deadwater Slough study area within the Upper Salmon River MPG. The Dump Creek alluvial fan is located at the downstream end of the study reach.



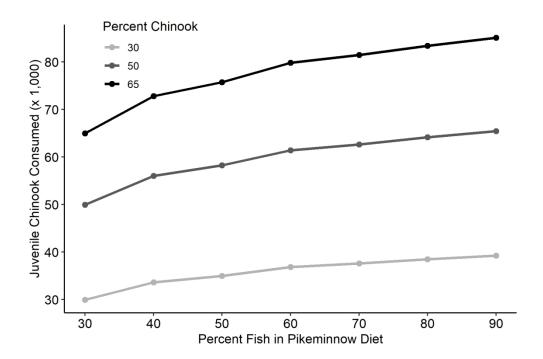
Estimated abundance of Northern Pikeminnow in Deadwater Slough from three mark-recapture estimators for the fall surveys. Error bars indicate 95% confidence intervals.



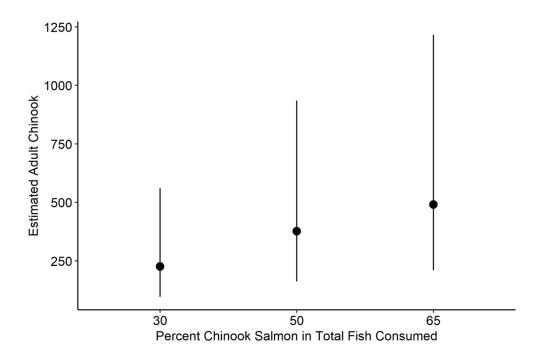
Length frequency histogram of Northern Pikeminnow caught using hook-and-line angling during the study. $387 x 258 mm \ (118 \ x \ 118 \ DPI)$



The cumulative fish prey consumed (g) by an individual Northern Pikeminnow during the peak Chinook Salmon emigrations for fall DSR (top left) and spring NRR (top right) and corresponding daily mean water temperatures (bottom panels). The black line shows the consumption by Northern Pikeminnow when the diet consists of 60% fish and the gray area shows the range between all model runs in the series, from 30% to 90% fish prey. Daily mean water temperatures were summarized from six years of data available from the Shoup gage station.



Estimated number of juvenile Chinook Salmon consumed by Northern Pikeminnow from a sensitivity analysis of diet scenarios made up of variable percentages of fish. Diet scenarios include 30%, 50%, and 65% Chinook Salmon in the total fish prey consumed by Northern Pikeminnow, similar to those reported by Zimmerman and Ward 1999.



Estimated "adult equivalents" of the total juvenile Chinook Salmon consumed by Northern Pikeminnow in Deadwater Slough. Error bars indicate 95% confidence intervals.

Supplement - Avian Predation in Deadwater Slough

Although not formally assessed in this study, avian predators are another potential source of mortality for juvenile Chinook Salmon and other ESA-listed salmonids in the Deadwater Slough. The Deadwater Slough is recognized as an important bird watching and nesting area due to the associated upstream braided channels and riparian and backwater habitats (Deadwater Slough -Audubon Important Bird Areas). Several piscivorous bird species have been documented using Deadwater Slough including the Common Mergus merganser and Hooded Lophodytes cucullatus mergansers, the Great Blue Heron Ardea herodias, the Double-crested Cormorant Phalacrocorax auritus, and the Belted Kingfisher Megaceryle alcyon (eBird 2021). During the initial sampling effort in fall 2019, a two-person crew walked the entire reach including surrounding and upstream areas scanning for passive integrated transponder (PIT) tags that are implanted into thousands of natural- and hatchery-origin salmonids in the Upper Salmon River, annually. During that informal survey, 10 PIT tags were recovered near active bird nests and in an upstream anastomizing reach where herons and eagles are prevalent, suggesting that mortality may have been a result of avian predation. The PIT tag histories in PTAGIS indicate these tags were implanted into a combination of juvenile Chinook Salmon (3), Sockeye Salmon (4), and steelhead (3). Two of the Chinook Salmon and one steelhead were natural-origin, the remaining were of hatchery-origin. Avian predation contributes a major component of the total mortality for yearling Chinook Salmon in some locations in the lower Snake and Columbia rivers, particularly at hydroelectric dams and within reservoirs (Evans et al. 2012; 2016); however, we did not observe large colonies of piscivorous birds within the study area. Although we did observe Belted Kingfishers, mergansers, and Great Blue Herons at Deadwater Slough; Great Blue Herons

were especially prevalent in the anastomizing reach immediately upriver of the slack water reach. Additionally, there is documentation of individual Double-crested Cormorants (eBird 2021) at the Deadwater Slough; however, the site is not within their breeding range, rather, it is part of a migration corridor. Given the current avian species known to occupy Deadwater Slough, it is unlikely that avian predation on juvenile salmonids is comparable to elsewhere in the Columbia River basin with large piscivorous bird colonies. Nevertheless, we hypothesize that the reservoir-like conditions and associated habitat features at the Deadwater Slough may increase the probability of avian predation on juvenile Chinook Salmon and other ESA-listed salmonids from the many piscivorous birds known to use the site. Future studies of predation would benefit from estimates from piscivorous avian predators.

Literature Cited to Supplement

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