

Characterization of the transcriptome, nucleotide sequence polymorphism, and natural selection in the desert adapted mouse *Peromyscus eremicus*

Matthew D. MacManes¹, Michael B. Eisen²,

¹ Department of Molecular, Cellular and Biomedical Sciences, University of New Hampshire, Durham NH, USA

² HHMI and University of California, Berkeley, Berkeley, CA, USA

* E-mail: macmanes@gmail.com, @PeroMHC

1 Abstract

2 Introduction

3 Deserts are widely considered one of Earth's harshest environments. Animals living in desert en-
 4 vironments are forced to endure intense heat and drought, and in turn, species having evolved in
 5 these environments are likely to have evolved specialised mechanisms that may enhance fitness.
 6 While living in deserts likely involves a large number of adaptive phenotypes, the ability to os-
 7 moregulate – to maintain the proper water and electrolyte balance – appears to be paramount [1].
 8 Indeed, the maintenance of water balance in animals is one of the most important physiologic
 9 processes for all animals, desert inhabitants or not. Most animals are exquisitely sensitive to
 10 changes in osmolality, with slight derangement eliciting physiologic compromise. When the loss
 11 of water exceeds dietary intake, dehydration - and in extreme cases, death - can occur, which
 12 suggests that there is strong selection for mechanisms supporting osmoregulation. Understand-
 13 ing these mechanisms will significantly enhance our understanding of the physiologic processes
 14 underlying osmoregulation in extreme environments, having implications for studies of human
 15 health, conservation, and climate change.

16
 17 The genes and structures responsible for the maintenance of water and electrolyte balance
 18 are well characterized in model organisms such as mice [2], rats [3–5], and humans [6–8]. These
 19 studies, many of which have been enabled by newer sequencing technologies, serve as a founda-
 20 tion for studies of renal genomics in non-model organisms. In particular, because researchers
 21 have long been interested in desert adaptation, a number of studies have looked at the mor-
 22 phology or expression of single genes in the renal tissues of desert adapted rodents *Phyllotis*
 23 *darwini* [9], *Psammomys obesus* [10], and *Perognathus penicillatus* [11]. More recently, full re-
 24 nal transcriptomes have been generated for *Dipodomys spectabilis* and *Chaetodipus baileyi* [12]
 25 as well as *Abrothrix olivacea* [13].

These studies provide a rich context for the current and future work, aimed at developing a synthetic understanding of the the genetic and genomic underpinnings of desert adaptation in rodents. As a first step, we have sequenced, assembled, and characterized the transcriptome (using four tissue types - liver, kidney, testes, brain), of a desert adapted cricetid rodent endemic to the Southwest United States [14], *Peromyscus eremicus*. These animals have a lifespan typical of small mammals, and therefore an individual may live it's entire life without ever drinking water. These rodents have distinct advantage over other desert animals (e.g. *Dipodomys*) in that they breed readily in captivity, which enables future laboratory studies of the phenotype of interest. In addition the focal species is positioned in a clade of well known animals (e.g. *P. californicus*, *P. maniculatus* and *P. polionotus*) [15] with growing genetic and genomic resources [16–18] which together suggest that future comparative studies are possible.

While the elucidation of the mechanisms underlying adaptation to desert survival is beyond the scope of this manuscript, we aim here to lay the groundwork by characterizing the transcriptome from four distinct tissues (brain, liver, kidney, testes). These data will be included in current larger effort aimed at sequencing the entire genome. Further, via sequencing the renal tissue of a total of 15 additional animals, we characterize nucleotide polymorphism and genome wide patterns of natural selection. Together, these investigations will aid in our overarching goal – to understand the genetic bases of adaptation in *P. eremicus*.

Materials and Methods

Animal Collection and Study Design

To begin to understand how genes may underlie desert adaptation, I collected 16 individuals from a single population *P. eremicus* over a two year time period (2012-2013). These individuals were captured in live traps, then euthanized using isoflurane overdose and decapitation. Immediately post-mortem, the abdominal and pelvic organs were removed, cut in half (in the case of kidney), placed in RNAlater and flash frozen in liquid Nitrogen. Removal of the brain, with similar preservation techniques, followed that. Time from euthanasia to removal of all organs never exceeded five minutes. Samples were transferred to a -80C freezer at a later date. These procedures were approved by the University of California Berkeley Animal Care and Use Committee and follow guidelines established by the American Society of Mammalogy for the use of wild animals in research [19].

RNA extraction and Sequencing

Total RNA was extracted from each tissue using a TRIzol extraction (Invitrogen) following the manufacturers instructions. Because preparation of an RNA library suitable for sequencing is dependent on having high quality, intact RNA, a small aliquot of each total RNA extract was analyzed on a Bioanalyzer 2100 (Agilent, Palo Alto, CA, USA). Following confirmation of sample quality, the reference sequencing libraries were made using the TruSeq stranded RNA prep kit (Illumina), while an unstranded TruSeq kit was used to construct the other sequencing libraries. A unique index was ligated to each sample to allow for multiplexed sequencing. Reference libraries (n=4 tissue types) were then pooled to contain equimolar quantities of each individual library and submitted for Illumina sequencing using two lanes of 150nt paired end sequencing using the rapid-mode of the HiSeq 2500 sequencer at The Hubbard Center for Genome Sciences (University of New Hampshire). The remaining 14 libraries were similarly multiplexed and sequenced in a mixture of 100nt paired and single end across two lanes of an Illumina HiSeq 2000 at the Vincent G. Coates Gnome Center (University of California, Berkeley).

Sequence Data Preprocessing and Assembly

The raw sequence reads were error corrected using the software *bleed* [20], using *kmer*=25, based on the developers default recommendations. The error corrected adapter and quality trimmed following recommendations from MacManes [21] and Mbandi [22]. Specifically, adapter sequence contamination was removed, and low quality nucleotides (defined as PHRED <2) were removed using the program Trimmomatic version 0.32 [23]. Reads from each tissue were assembled using Trinity version released 17 July 2014 [24]. We used flags indicating the stranded nature of sequencing reads and set maximum allowable physical distance between read pairs to 999nt. The assembly was conducted on a linux workstation with 64 cores and 512Gb RAM. To filter the raw sequence assembly, I downloaded *Mus musculus* cDNA and ncRNA datasets from Ensembl (ftp://ftp.ensembl.org/pub/release-75/fasta/mus_musculus/), and the *Peromyscus maniculatus* reference transcriptome from NCBI (ftp://ftp.ncbi.nlm.nih.gov/genomes/Peromyscus_maniculatus_bairdii/RNA/). I used a blastN procedure (default settings, *evalue* set to 10^{-10}) to identify contigs in the *P. eremicus* dataset that are likely biological in origin. This procedure, when a reference dataset is available, retains more putative transcripts than a strategy employing expression-based filtering (remove if *TMP* <1) of the raw assembly. I then concatenated the filtered assemblies from each tissue into a single file, reducing redundancy using the software *cd-hit-est* [25] using default setting except that sequences were clustered based on 95% sequence similarity. The resulting assembly file was characterized using the software package *transrate* (<https://github.com/Blahah/transrate>).

Assembled Sequence Annotation

The filtered assemblies were annotated using default settings of the blastN algorithm [26] against the Ensembl cDNA and ncRNA datasets described above, downloaded on 1 August 2014. Amongst other things, the Ensemble transcript identifiers were used in the analysis of gene ontology, conducted in the PANTHER package [?]. Next, because rapidly evolving nucleotide sequences may evade detection by blast algorithms, we used HMMER3 [27] to search for conserved protein domains contained in the dataset using the Pfam database [28]. Lastly, I extracted putative coding sequences using Transdecoder version 4Jul2014 (<http://transdecoder.sourceforge.net/>)

To identify patterns of gene expression unique to each tissue type, I mapped sequence reads from each tissue type to the reference assembly using bwa-mem. We estimated expression individually for the four tissues using default settings of the software eXpress [29]. Interesting patterns of expression, including instances where expression was limited to a single tissue type were identified and visualized.

Population Genomics

In addition to the reference individual sequenced at four different tissue types, we sequenced 15 other conspecific individuals from the same population, located in Palm Desert, California. Sequence data were mapped to the reference transcriptome using bwa-mem. The alignments were sorted and converted to BAM format, then passed to the program ANGSD version 0.610, which was used for calculating the folded site frequency spectrum (SFS) and Tajima's D [30].

Natural Selection

To characterize natural selection on several genes related to water and ion homeostasis, we identified several of the transcripts identified as experiencing positive selection in a recent work on desert-adapted *Dipodomys* rodents. The coding sequence corresponding to these genes, Solute Carrier family 2 member 9 (Slc2a9) and the Vitamin D3 receptor (Vdr), were extracted from the dataset, aligned using the software MACSE [31] to homologous sequences in *Mus musculus*, *Rattus norvegicus*, *Peromyscus maniculatus*, and *Homo sapiens* identified by the conditional reciprocal best blast procedure (CRBB, [32]). An unrooted gene tree was constructed using the online resource Clustal-Omega, and together the tree and alignment were analyzed using the branch-site model (model=2, nsSites=2, fix_omega=0 versus model=2, nsSites=2, fix_omega=1,

126 omega=1) implemented in PAML version 4.8 [33,34].

127 Results

128 RNA extraction, Sequencing, Assembly, Mapping

129 RNA was extracted from the hypothalamus, renal medulla, testes, or liver from each individual
 130 using sterile technique. TRIzol extraction resulted in a large amount of high quality ($RIN \geq 8$)
 131 total RNA, which was used as input. Libraries were constructed as per the standard Illumina
 132 protocol, and were sequenced as described above. The number of reads per library varied from
 133 56 million strand-specific paired-end reads in Peer360 kidney, to 9 million single-end reads in
 134 Peer321. Adapter sequence contamination and low-quality nucleotides were eliminated, which
 135 resulted in a loss of <2% of reads.

136
 137 Transcriptome assembly for each tissue type was accomplished using the program Trin-
 138 ity [24]. The raw assembly for brain, liver, testes and kidney contained 185425, 222096, 180233,
 139 and 514091 assembled sequences respectively. This assembly was filtered using a blastN pro-
 140 cedure against the *Mus* cDNA and ncRNA which resulted in a final dataset containing 68331
 141 brain-specific transcripts, 71041 liver-specific transcripts, 67340 testes-specific transcripts, and
 142 113050 kidney-specific transcripts. Mapping the error-corrected adapter/quality trimmed reads
 143 to these datasets resulted in mapping 94.98% (87.01% properly paired) of brain-derived reads
 144 to the brain transcriptome, 96.07% (88.13% properly paired) of liver-derived reads to the liver
 145 transcriptome, 96.81% (85.10% properly paired) of testes-derived reads to the testes transcrip-
 146 tome, and 91.87% (83.77% properly paired) of kidney-derived reads to the kidney transcriptome.
 147 Together, these statistics suggest that the tissue-specific transcriptomes are of extremely high
 148 quality.

149

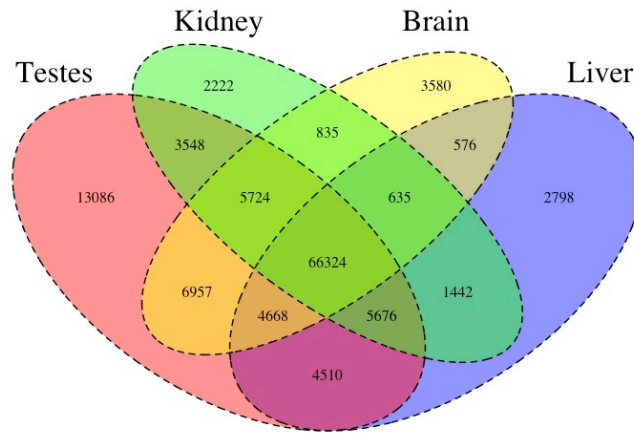


Figure 1. The Venn Diagram.

I then estimated gene expression on each of these tissue-specific datasets, which allowed me to understand expression patterns in the multiple tissues (Figure 1). After expression estimation, the filtered assemblies were concatenated together, and after removal of redundancy with cd-hit-est, 123,123 putative transcripts remained. From this filtered concatenated dataset, I extracted 71626 putative coding sequences (72Mb). Of these 71626 sequences, 38221 were complete exons (containing both start and stop codons), while other were either truncated at the 5-prime end (20239 sequences), 3-prime end (6445 sequences), or were internal (6721 sequencing having neither stop nor start codon).

Population Genomics

As detailed above, the RNAseq data from 15 individuals were mapped to the reference transcriptome with the resulting BAM files being used as input to the software package ANGSD. The Tajima's D statistic was calculated for all transcripts covered by at least 14 of the 15 individuals. The distribution of the results, shown in Figure 2, suggest that the vast majority of the transcriptome is under purifying selection ($D < 0$), with a much smaller fraction being subject to neutral or positive selection.

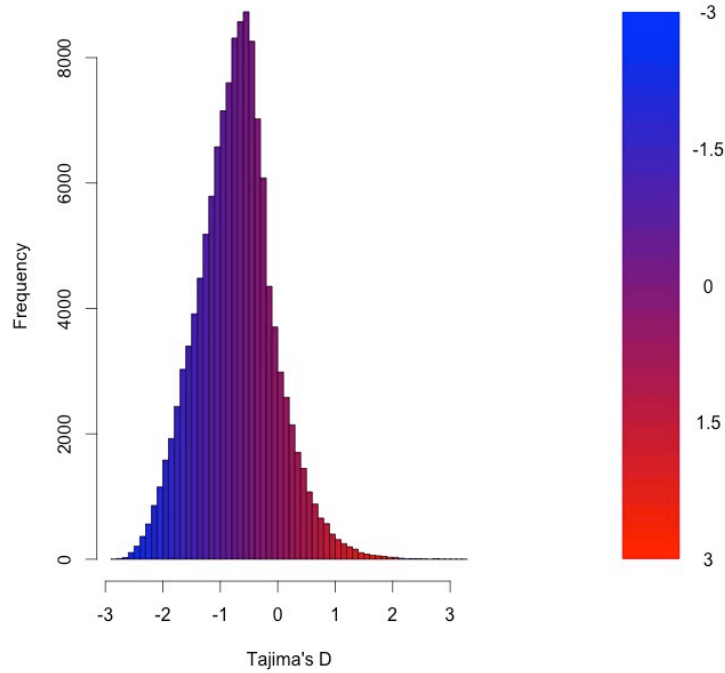


Figure 2. Tajima's D

Natural Selection

To test the hypothesis that selection on transcripts related to osmoregulation is enhanced in the desert adapted *P. eremicus*, I implemented the branch-site test as described above, setting the sequence corresponding to *P. eremicus* for both *Slc2a9* and *Vdr* as the foreground lineages in 2 distinct program executions. The test for *Slc2a9* was highly significant ($2\Delta\text{Lnl}=51.4$, $\text{df}=1$, $p=0$), indicating enhanced selection in *P. eremicus* relative to the other lineages. The branch site test for positive selection conducted on the *Vdr* gene was non-significant ($2\Delta\text{Lnl}=0.68$, $\text{df}=1$, $p=1$).

Discussion

Acknowledgments

References

1. Walsberg G (2000) Small mammals in hot deserts: Some generalizations revisited. *Bio-science* 50: 109–120.
2. Tatum R, Zhang Y, Salleng K, Lu Z, Lin JJ, et al. (2009) Renal salt wasting and chronic dehydration in claudin-7-deficient mice. *AJP: Renal Physiology* 298: F24–F34.
3. Romero DG, Plonczynski MW, Welsh BL, Gomez-Sanchez CE, Zhou MY, et al. (2007) Gene expression profile in rat adrenal zona glomerulosa cells stimulated with aldosterone secretagogues. *Physiological Genomics* 32: 117–127.
4. Rojek A, Rojek A, Fuchtbauer E, Fuchtbauer E, Kwon T, et al. (2006) Severe urinary concentrating defect in renal collecting duct-selective AQP2 conditional-knockout mice. *Proceedings of The National Academy of Sciences of The United States of America* 103: 6037–6042.
5. Nielsen S, CHOU C, MARPLES D, CHRISTENSEN E, KISHORE B, et al. (1995) Vasopressin Increases Water Permeability of Kidney Collecting Duct by Inducing Translocation of Aquaporin-Cd Water Channels to Plasma-Membrane. *Proceedings of The National Academy of Sciences of The United States of America* 92: 1013–1017.
6. Mobasheri A, Marples D, Young IS, Floyd RV, Moskaluk CA, et al. (2007) Distribution of the AQP4 Water Channel in Normal Human Tissues: Protein and Tissue Microarrays Reveal Expression in Several New Anatomical Locations, including the Prostate Gland Seminal Vesicles. *Channels* 1: 30–39.
7. Bedford JJ, Bedford JJ, Leader JP, Leader JP, Walker RJ, et al. (2003) Aquaporin expression in normal human kidney and in renal disease. *Journal of the American Society of Nephrology : JASN* 14: 2581–2587.
8. Nielsen S, KWON T (1999) Physiology and Pathophysiology of Renal Aquaporins. *Journal of the ...*
9. Gallardo PA, Cortés A, Bozinovic F (2005) Phenotypic flexibility at the molecular and organismal level allows desert-dwelling rodents to cope with seasonal water availability. *Physiological and Biochemical Zoology* 78: 145–152.

- 205 10. Kaissling B, De Rouffignac C, Barrett JM, Kriz W (1975) The structural organization
206 of the kidney of the desert rodent *Psammomys obesus*. *Anatomy and embryology* 148:
207 121–143.
- 208 11. Altschuler EM, Altschuler EM, Nagle RB, Nagle RB, Braun EJ, et al. (1979) Morpholog-
209 ical study of the desert heteromyid kidney with emphasis on the genus *Perognathus*. *The*
210 *Anatomical record* 194: 461–468.
- 211 12. Marra NJ, Romero a, DeWoody Ja (2014) Natural selection and the genetic basis of
212 osmoregulation in heteromyid rodents as revealed by RNA-seq. *Molecular Ecology* 23:
213 2699–2711.
- 214 13. Giorello FM, Feijoo M, a GD, Valdez L, Opazo JC, et al. (2014) Characterization of the
215 kidney transcriptome of the South American olive mouse *Abrothrix olivacea* 15: 1–10.
- 216 14. Veal R, Caire W (2001) *Peromyscus eremicus*. *Mammalian Species* 118: 1–6.
- 217 15. Feng BJ, Sun LD, Soltani-Arabshahi R, Bowcock AM, Nair RP, et al. (2007) Toward
218 a Molecular Phylogeny for *Peromyscus*: Evidence from Mitochondrial Cytochrome- b
219 Sequences. *Journal of Mammalogy* 88: 1146–1159.
- 220 16. Shorter KR, Owen A, anderson V, Hall-South AC, Hayford S, et al. (2014) Natural Ge-
221 netic Variation Underlying Differences in *Peromyscus* Repetitive and Social/Aggressive
222 Behaviors. *Behavior genetics* .
- 223 17. Panhuis TM, Panhuis TM, Broitman-Maduro G, Broitman-Maduro G, Uhrig J, et al.
224 (2011) Analysis of Expressed Sequence Tags from the Placenta of the Live-Bearing Fish
225 *Poeciliopsis* (Poeciliidae). *Journal of Heredity* 102: 352–361.
- 226 18. Shorter KR, Crossland JP, Webb D, Szalai G, Felder MR, et al. (2012) *Peromyscus* as a
227 Mammalian Epigenetic Model. *Genetics Research International* 2012: 1–11.
- 228 19. Sikes RS, Gannon WL, Animal Care and Use Committee of the American Society of
229 Mammalogists (2011) Guidelines of the American Society of Mammalogists for the use of
230 wild mammals in research. *Journal of Mammalogy* 92: 235–253.
- 231 20. Heo Y, Wu XL, Chen D, Ma J, Hwu WM (2014) BLESS: bloom filter-based error cor-
232 rection solution for high-throughput sequencing reads. *Bioinformatics* (Oxford, England)
233 30: 1354–1362.
- 234 21. MacManes MD (2014) On the optimal trimming of high-throughput mRNA sequence
235 data. *Frontiers in Genetics* 5.

- 236 22. Christoffels A (2014) A glance at quality score: implication for *de novo* transcriptome
237 reconstruction of Illumina reads : 1–5.
- 238 23. Lohse M, Bolger AM, Nagel A, Fernie AR, Lunn JE, et al. (2012) RobiNA: a user-friendly,
239 integrated software solution for RNA-Seq-based transcriptomics. *Nucleic Acids Research*
240 40: W622–7.
- 241 24. Haas BJ, Papanicolaou A, Yassour M, Grabherr M, Blood PD, et al. (2013) *De novo*
242 transcript sequence reconstruction from RNA-seq using the Trinity platform for reference
243 generation and analysis. *Nature protocols* 8: 1494–1512.
- 244 25. Li W, Li W, Godzik A, Godzik A (2006) Cd-hit: a fast program for clustering and
245 comparing large sets of protein or nucleotide sequences. *Bioinformatics (Oxford, England)*
246 22: 1658–1659.
- 247 26. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, et al. (2009) BLAST+:
248 architecture and applications. *BMC Bioinformatics* 10: 421.
- 249 27. Wheeler TJ, Wheeler TJ, Eddy SR, Eddy SR (2013) nhmmer: DNA homology search
250 with profile HMMs. *Bioinformatics (Oxford, England)* 29: 2487–2489.
- 251 28. Punta M, Coghill PC, Eberhardt RY, Mistry J, Tate J, et al. (2012) The Pfam protein
252 families database. *Nucleic Acids Research* 40: D290–301.
- 253 29. Roberts A, Pachter L (2013) Streaming fragment assignment for real-time analysis of
254 sequencing experiments. *Nature Methods* 10: 71–73.
- 255 30. Korneliussen TS, Moltke I, Albrechtsen A (2013) Calculation of Tajima’s D and other
256 neutrality test statistics from low depth next-generation sequencing data. *BMC*
- 257 31. Ranwez V, Harispe S, Delsuc F, Douzery EJP (2011) MACSE: Multiple Alignment of
258 Coding SEquences Accounting for Frameshifts and Stop Codons. *PLOS ONE* 6: e22594.
- 259 32. Aubry S, Kelly S, Kumpers BMC, Smith-Unna RD, Hibberd JM (2014) Deep Evolutionary
260 Comparison of Gene Expression Identifies Parallel Recruitment of Trans-Factors in Two
261 Independent Origins of C4 Photosynthesis. *PLOS Genetics* 10: e1004365.
- 262 33. Yang Z, dos Reis M (2011) Statistical Properties of the Branch-Site Test of Positive
263 Selection. *Molecular Biology and Evolution* 28: 1217–1228.
- 264 34. Yang Z (2007) PAML 4: Phylogenetic Analysis by Maximum Likelihood. *Molecular*
265 *Biology and Evolution* 24: 1586–1591.

266 **Figure Legends**

267 **Tables**

268 **Table 1**

269

DATASET	NUM. RAW READS
PEER360 TESTES	32M PE
PEER360 LIVER	53M PE
PEER360 KIDNEY	56M PE
PEER360 BRAIN	23M PE
PEER305	19M PE
PEER308	15M PE
PEER319	14M PE
PEER321	9M SE
PEER340	16M PE
PEER352	14M PE
PEER354	9M SE
PEER359	14M PE
PEER365	16M PE
PEER366	16M PE
PEER368	14M PE
PEER369	14M PE
PEER372	17M SE
PEER373	23M SE
PEER380	16M SE
PEER382	14M SE

270

271 Table 1. The number of sequencing reads per sample