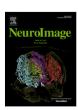


Contents lists available at ScienceDirect

NeuroImage

journal homepage: http://ees.elsevier.com



Core and matrix thalamic sub-populations relate to spatio-temporal cortical connectivity gradients

Eli J. Müller ^{a,1}, Brandon Munn ^{a,1}, Luke J. Hearne ^b, Jared B. Smith ^c, Ben Fulcher ^d, Aurina Arnatkevičiūtė ^e, Daniel J. Lurie ^f, Luca Cocchi ^g, James M. Shine ^{a,d},*

- ^a Brain and Mind Centre, The University of Sydney, Sydney, NSW, Australia
- ^b Center for Molecular and Behavioral Neuroscience, Rutgers University, Newark, USA
- ^c Molecular Neurobiology Laboratory, Salk Institute for Biological Studies, La Jolla, CA, USA
- ^d Complex Systems Research Group, The University of Sydney, Sydney, NSW, Australia
- e Brain and Mental Health Research Hub, Monash Institute of Cognitive and Clinical Neurosciences, School of Psychological Sciences, Monash University, Clayton, VIC, Australia
- f Department of Psychology, University of California, Berkeley, Berkeley, CA, USA
- g QIMR Berghofer, Brisbane, QLD, Australia

ABSTRACT

Recent neuroimaging experiments have defined low-dimensional gradients of functional connectivity in the cerebral cortex that subserve a spectrum of capacities that span from sensation to cognition. Despite well-known anatomical connections to the cortex, the subcortical areas that support cortical functional organization have been relatively overlooked. One such structure is the thalamus, which maintains extensive anatomical and functional connections with the cerebral cortex across the cortical mantle. The thalamus has a heterogeneous cytoarchitecture, with at least two distinct cell classes that send differential projections to the cortex: granular-projecting 'Core' cells and supragranular-projecting 'Matrix' cells. Here we use high-resolution 7T resting-state fMRI data and the relative amount of two calcium-binding proteins, parvalbumin and calbindin, to infer the relative distribution of these two cell-types (Core and Matrix, respectively) in the thalamus. First, we demonstrate that thalamocortical connectivity recapitulates large-scale, low-dimensional connectivity gradients within the cerebral cortex. Next, we show that diffusely-projecting Matrix regions preferentially correlate with cortical regions with longer intrinsic fMRI timescales. We then show that the Core–Matrix architecture of the thalamus is important for understanding network topology in a manner that supports dynamic integration of signals distributed across the brain. Finally, we replicate our main results in a distinct 3T resting-state fMRI dataset. Linking molecular and functional neuroimaging data, our findings highlight the importance of the thalamic organization for understanding low-dimensional gradients of cortical connectivity.

1. Introduction

Advances in whole-brain neuroimaging have identified low-dimensional gradients in the cerebral cortex that interconnect sensorimotor regions with limbic structures (Margulies et al., 2016; Mesulam, 1998). Recent work has shown that these same gradients are underpinned by structural differences that have long been known to comprise the cerebral cortex (García-Cabezas et al., 2019), including circuit complexity (Paquola et al., 2019; Vázquez-Rodríguez et al., 2019) and the extent of cortical myelination (Burt et al., 2018; Demirtaş et al., 2019). Cytoarchitecture and myelination are presumed to differentially enforce coupling between structure and function (Fallon et al., 2020), with granular sensory regions more

closely tethered to their structural constraints than associative agranular regions (Paquola et al., 2019; Vázquez-Rodríguez et al., 2019). The complexity of cortical circuitry has also been shown to underpin a gradient of temporal scales across the brain, with associative regions fluctuating across relatively longer time scales than sensory regions (Murray et al., 2014). Comparable low-dimensional gradients have also been observed in task contexts (Shine et al., 2019a) and share similarities with meta-analytic studies of task fMRI (Margulies and Smallwood, 2017), suggesting that the low-dimensional gradients in the cerebral cortex may be a crucial factor underlying whole-brain functional organization.

A phylogenetic perspective on the brain (Cisek, 2019) suggests that the cortex is supported by several subcortical structures that

Corresponding author at: Brain and Mind Centre, The University of Sydney, Sydney, NSW, Australia.
 E-mail address: mac.shine@sydney.edu.au (J.M. Shine)

¹ These authors contributed equally to this work.

shape, constrain, and augment its activity on a moment-to-moment basis. One such structure that is crucial for shaping whole brain dynamics is the thalamus (Halassa and Sherman, 2019; Jones, 2009; 2001; Shine et al., 2019b) (Fig. 1). Located in the diencephalon, the thalamus is reciprocally connected with the entire cerebral cortex, along with primary sensory receptors (such as the retina and dorsal column tract) and numerous other subcortical systems (such as the basal ganglia, superior colliculus and cerebellum). These inputs innervate distinct, anatomically-segregated sub-nuclei within the thalamus (Fig. 1) that are surrounded by the shell-like GABAergic reticular nucleus (Jones, 2001). Through activity-dependent GABAergic inhibition of cortically-and subcortically-driven activity, the thalamus likely plays a crucial role in shaping and constraining patterns of whole-brain dynamics (Halassa and Sherman, 2019; Jones, 2009, 2001).

Individual sub-nuclei within the thalamus innervate distinct regions of the cerebral cortex (Fig. 1A) (Jones, 2009; 2001). In addition, each thalamic sub-nucleus is comprised of a mixture of excitatory neurons that project in unique ways to the cerebral cortex (Clascá et al., 2012; Jones, 2009). Specifically, Parvalbumin-rich 'Core' cells (Fig. 1B; green), typically dense within sensory thalamic nuclei such as the lateral geniculate nucleus, send axonal projections to Layers III and IV of a specific subset of cortical areas, such as primary visual, auditory and somatosensory cortices. These cells are thought to act as 'drivers' of feed-forward activity (Jones, 2009, 2001). In contrast, Calbindin-rich 'Matrix' cells (Fig. 1; blue), which are more prevalent in higher-order thalamic nuclei (Herkenham, 1979; Jones, 2001), preferentially target relatively agranular cortices in the supragranular layers (I-III), and in a more distributed fashion that crosses typical cortical receptive fields (Jones, 2009, 2001). Individual thalamic nuclei contain a blend of both cell-types. Of note, other schemes have been devised to parse the thalamus (Phillips et al., 2019), including the class of cortical region proposed to 'drive' activity in the glutamatergic relay nuclei (Sherman, 2007). However, these proposals are not considered in detail here.

The densities of Parvalbumin- (Core) and Calbindin- (Matrix) neurons have been used to classify the relative proportion of cells within each thalamic nucleus that project to either granular or supragranular layers, respectively (Fig. 1A) (Jones, 2009, 2001; Münkle et al., 2000). These studies have provided anatomical evidence for the presence of an admixture of core and matrix cell populations within the thalamus. The presence of calcium-binding proteins has also been shown to delineate further levels of anatomical specificity in the thalamus, such as the extent and location of laminar projections to the cortex (Clascá et al., 2012). However, the importance of this anatomi-

cal organizing principle for shaping patterns of whole-brain activity and connectivity has been less well characterized.

In this work, we leveraged the high spatiotemporal resolution of a 60-subject, 7T resting-state fMRI dataset to examine the relationship between thalamic cell-population densities and whole-brain functional connectivity. We first calculated the time-averaged functional connectivity between each thalamic voxel and 400 cortical parcels from the Schaefer atlas (Schaefer et al., 2018). We then used data from the Allen Human Brain Atlas (Arnatkevičiūtė et al., 2019; Gryglewski et al., 2018; Hawrylycz et al., 2012) to estimate the expression of two calcium-binding proteins – PVALB and CALB1 – in each thalamic voxel. This allowed us to determine the relative weighting of Core versus Matrix cells, respectively (Jones, 2009, 2001; Münkle et al., 2000; Phillips et al., 2019). The thalamic patterns were then compared to fluctuations in the activity of parcels within the cerebral cortex, allowing us to quantify the relationship between patterns of thalamic activity and emergent patterns of whole-brain functional connectivity.

2. Methods

2.1. Participants

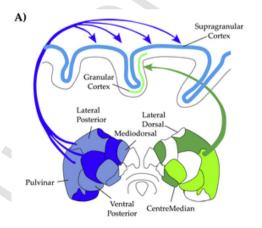
Sixty-five healthy, right-handed adult participants (18–33 years) were recruited, of whom 60 were included in the final analysis (28 females). Participants provided informed written consent to participate in the study. The research was approved by The University of Queensland Human Research Ethics Committee. These data were originally described in Hearne et al. (2017).

2.2. Neuroimaging acquisition

Ten minutes (1050 TRs) of whole-brain 7T resting state fMRI echo planar images were acquired using a multiband sequence (acceleration factor = 5; 2 mm³ voxels; 586 ms TR; 23 ms TE; 40° flip angle; 208 mm FOV; 55 slices). High resolution anatomical images were also collected to assist functional data pre-processing (MP2RAGE sequence – 0.75 mm³ voxels 4300 ms TR; 3.44 ms TE; 256 slices).

2.3. Data pre-processing

DICOM images were first converted to NIfTI format. T1 images were reoriented, skull-stripped (FSL BET), and co-registered to the NIfTI functional images using affine. Segmentation and the DARTEL algorithm were used to improve the estimation of non-neural signal in sub-



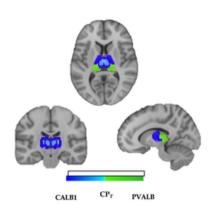


Fig. 1. The Cytoarchitecture of the Thalamus. A) Matrix (blue) and Core (green) thalamic populations have distinct projection patterns: Core cells innervate granular layers of the cerebral cortex in a specific fashion, whereas Matrix cells innervate supragranular cortex in a relatively diffuse manner. Color intensity denotes the relative expression of CALB1 and PVALB in different thalamic subnuclei (Jones, 2009; 2001); B) Axial (upper), coronal (lower left) and sagittal (lower right) loading of calbindin values (CALB1; blue) and parvalbumin values (PVALB; green) within the thalamus.

ject space and the spatial normalization. The aCompCor method (Behzadi et al., 2007) was used to regress out residual signal unrelated to neural activity (i.e., five principal components derived from noise regions- of-interest in which the time series data were unlikely to be modulated by neural activity). Participants with head displacement > 3 mm in > 5% of volumes in any one scan were excluded (n=5). A temporal band pass filter (0.071 < f < 0.125 Hz) was applied to the data. Following pre-processing, the mean time series was extracted from 400 pre-defined cortical parcels using the Schaefer atlas (Schaefer et al., 2018).

2.4. Relative expression of Calbindin and Parvalbumin in the thalamus

We co-registered the thalamic Morel atlas (Niemann et al., 2000) to MNI152 space and then identified 2305 voxels across both hemispheres that were inclusive to the thalamic mask. We then used this volume as a mask to extract the mRNA expression levels for PVALB and CALB1 provided by the Allen Human Brain Atlas (Hawrylycz et al., 2012). Prior to extraction, data were corrected for potential sources of bias (such as subject-level outliers) using variogram modeling (Gryglewski et al., 2018). We extracted spatial maps of estimated mRNA levels for two genes that are known to express distinct calcium-binding proteins: Calbindin (CALB1) and Parvalbumin (PVALB). These proteins have been previously shown to delineate two distinct sub-populations of thalamic projection cells (Matrix and Core, respectively) with distinct anatomical connectivity profiles. From the 58,692 probes in the original AHBA analysis (Hawrylycz et al., 2012), two were used to estimate the expression of PVALB (CUST 11451 PI416261804 and A 23 P17844) and three were used to estimate the expression of CALB1 (CUST_140_PI416408490, CUST_16773_PI416261804 A_23_P43197). Note that there are other calcium binding proteins (notably, Calretinin) with substantial expression in the thalamus (Münkle et al., 2000; Phillips et al., 2019), however these patterns are not considered here.

To create an unbiased estimate of the relative weighting of each thalamic sub-population, mRNA levels were first normalized (z-score) across all voxels within the thalamic mask before creating the differ-

Table 1
Thalamic morel atlas and calcium binding protein values. Samples and % of subjects with data, along with average mRNA loading of Calbindin (CALB1) and Parvalbumin (PVALB) for each of 6 pre-identified thalamic sub-groups.

Region	# Samples	% Subjects	CALB1	PVALB	CP_T
Lateral Geniculate	17	100	0.23	0.25	-0.02
Medial Geniculate	11	100	0.31	0.40	-0.09
Anterior/Ventral	41	100	0.51	0.62	-0.11
Posterior	27	100	0.40	0.57	-0.17
Medial/Dorsal	26	100	0.45	0.41	0.04
Intralaminar	33	100	0.38	0.29	0.09

ence between the normalized Calbindin and Parvalbumin levels (Fig. 1B; Table 1). We denote this measure CP_T : positive levels reflect voxels with higher CALB1 levels, negative levels reflect voxels with higher PVALB levels and values of zero denote a balance between the two populations. To validate the measure, the mean CP_T value was calculated for each of 31 pre-defined thalamic sub-nuclei that fell into 6 distinct groups (Table 1) and compared quantitatively to results obtained from direct histological (Münkle et al., 2000) and immunohistochemistry (Arai et al., 1994) analyses (data were extracted from tables within each of the referenced documents). We observed a positive correlation between these direct measurements and our gene-expression based proxy, CP_T (r=0.546; p=0.003; Fig. S1).

2.5. Thalamocortical connectivity patterns

Following the creation of the CP_T spatial map, we estimated the correlation between thalamocortical functional connectivity strength (Pearson's correlation between each cortical parcel and each voxel in the thalamic mask) and the CP_T value of each thalamic voxel. This allowed us to quantify, with a single value (CP_C), the functional connectivity between each of the 400 cortical parcels and the Matrix and Core thalamic populations. As CP_T represents the relative expression of CALB1 (Matrix-rich) versus PVALB (Core-rich), positive correlations (i.e., $\mathrm{CP}_C > 0$) were interpreted as preferential functional coupling with Matrix thalamic populations and negative correlations (i.e., $\mathrm{CP}_C < 0$) were taken to be indicative of preferential coupling to Core populations. To aid interpretation, we calculated the mean correlation between thalamocortical FC and CP_T within each of 17 pre-defined resting-state networks (Yeo et al., 2011; Schaefer et al., 2018), and projected CP_T onto a thalamic volume (Fig. 1) and CP_C onto the cortical surface (Fig. 2).

2.6. Relationship with low-dimensional cortical gradients

Given the known diverse connectivity of the thalamus with the cerebral cortex, we predicted that known anatomical gradients in the thalamus should relate to low-dimensional patterns of functional connectivity in the cortex. To test this hypothesis, we compared the Core–Matrix organization with the low-dimensional cortical gradients obtained by applying diffusion embedding analyses to resting-state fMRI data from a large cohort of subjects (Margulies et al., 2016). Un-thresholded volumetric maps representing the top 5 low-dimensional cortical gradients were downloaded from NeuroVault (Gorgolewski et al., 2015). These maps were then down sampled into the 400 cortical parcel space by calculating the mean expression of each gradient within each cortical parcel. We then compared the spatial topography of the ${\rm CP}_C$ map to each of the five cortical FC gradients using Pearson's correlations. These comparisons utilized parametric, one-sided 'spin' tests to control for non-linearity and spatial autocorrelation, respectively; see below for details.

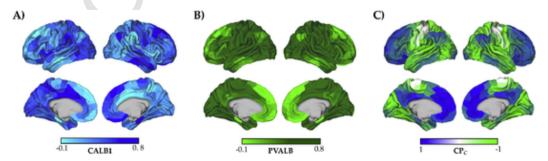


Fig. 2. Correlations between Matrix-Core thalamic regions and cortical parcels, CP_C. A) standardized correlation between Matrix thalamus (CALB1 levels) and cortical parcels; B) standardized correlation between Core thalamus (PVALB levels) and cortical parcels; C) relative difference between Matrix and Core populations relationship with cortical parcels. Color bars depict the relative strength of correlation between each thalamic population at the individual cortical parcels (standardized between 0 and 1).

2.7. Relationship between CP_c and intrinsic timescale

Different areas of the cerebral cortex are characterized by an intrinsic timescale that reflects the length of the time window over which the signals entering each brain region are integrated (Mesulam, 1998; Murray et al., 2014). There are two predominant methods utilised in fMRI studies for investigating the intrinsic timescale of activity: autocorrelation (Murray et al., 2014; Watanabe et al., 2019) and fractal based (He, 2011; Churchill et al., 2016) techniques. While both measures identify temporal self-similarity, autocorrelation techniques typically focus on linear correlations with past values of a particular region, whereas in contrast, fractal analysis is used as a measure of complexity (Dong et al., 2018) and can be used to discern the significance of differing time-scale frequencies. For instance, a purely fractal signal suggests all frequencies contain information (Churchill et al., 2016).

To examine the relationship between Core–Matrix architecture and the intrinsic BOLD timescale of cortical parcels, we first performed an autocorrelation analysis (Murray et al., 2014). Specifically, to estimate the extent of the autocorrelation, ac, in each region, r, we calculate the correlation coefficient between the BOLD activity, $X_r(t)$, at times t and t+k with different time lags $k=0,\ 1,\ ...,\ k_{\max}$. For our analysis $k_{\max}=50$ TR, however our findings were not sensitive for differences in k_{\max} (from 15 – 1000). The autocorrelation for each region is given by

$$ac_r(k) = \int_0^{T-k} X_r(t+k)X_r(t) dt,$$
 (1)

where T is the duration of the BOLD activity and the autocorrelation is normalised such that the autocorrelation at a lag of zero is unity. As the time lag increases, the autocorrelation decays according to the regions' intrinsic timescale, which is well fit to an exponential decay, N_r , with an offset given by

$$N_r = a \left(e^{-\frac{k\Delta T}{\tau_r}} + b \right), \tag{2}$$

where τ_r is the intrinsic timescale, a and b are the scaling and offset constants, respectively, k is the lag, and ΔT is the time resolution (i.e., the length of the TR). We averaged the region-wise auto-correlation across participants and fit the exponential decay to the population level auto-correlation, in order to obtain a population level statistic for each region in our dataset. The curve parameters were estimated using a Levenberg-Marquardt nonlinear least-squares fitting algorithm. It should be noted that our method is identical to the method adopted by Murray et al. (2014), who also fit an exponential decay with a baseline constant. Another proportional method is to calculate the area under the normalized autocorrelation curve (Gollo, 2019; Watanabe et al., 2019). In practice, we found our method was more flexible for fitting rapidly decaying BOLD signals.

To further relate the thalamocortical circuit properties with cortical temporal dynamics, we calculated the fractal nature of the BOLD timeseries of each cortical region in all subjects. To do so, we employed a detrended fluctuation analysis (DFA) to BOLD activity at each region (Peng et al., 1995). DFA is an efficient fractal measure to estimate the Hurst exponent of a signal that is intrinsically robust to signal non-stationarity, which may be present in BOLD activity. To calculate the DFA, we first computed the integrated time-series $y_r(t) = \sum_{i=1}^t \left(X_r(i) - \langle X_r \rangle \right)$, where $\langle X_r \rangle$ is the average BOLD activity. We then subdivided $y_r(t)$ into windows of equal length n and estimated a linear regression per subdivision, $w_n(t)$. We then computed the root-

mean-square magnitude of fluctuations, F(n), on the detrended data:

$$F(n) = \sqrt{\frac{1}{T} \sum_{t=1}^{T} (y_r(t) - w_n(t))^2}$$
 (3)

Finally, we calculated the RMS fluctuation for varying window sizes and calculated the slope of F(n) vs n plotted on a logarithmic scale, again using a Levenberg-Marquardt nonlinear least-squares fitting algorithm. The slope of log-log plot corresponds to the Hurst exponent, H, where H = 0.5 corresponds to Brownian motion (Beran, 1992; Munn et al., 2020). As H increases, the level of positive long-range temporal correlation in the signal increases. That is, an increase in BOLD activity is typically followed by another increase, and a reduction is typically followed by further reduction. As such, larger values of H indicate longer time scales of activity within individual regions. In our analysis, we calculated the Hurst exponent over window ranges from 5 to 60 TRs, sampled in logarithmically uniformly window sizes. The window range avoids zero residuals across the shortest intervals and low-frequency confounds identified in the power-spectral density across the long intervals. Consistent with (Churchill et al., 2016; He, 2011), we observed a crossover between two fractal ranges, consisting of an initial large increase of F(n) and large H for small window lengths due to serial autocorrelations which decay rapidly transitioning to smaller H values.

2.8. Network topology

To embed the CP_C results within the context of broader cortical network organization, we estimated time-averaged functional connectivity between each pair of cortical parcels in all 60 subjects and then subjected the weighted, unthresholded connectivity matrices to topological analysis. This involved using a weighted- and signed- version of the Louvain modularity algorithm from the Brain Connectivity Toolbox (Rubinov and Sporns, 2010). The Louvain algorithm iteratively maximizes the modularity statistic, Q, for different community assignments until the maximum possible score of Q has been obtained (Eq. (4)). The modularity estimate for a given network is, therefore, quantification of the extent to which the network may be subdivided into communities with stronger within-module than between-module connections.

$$Q_{T} = \frac{1}{\mathbf{b}^{+}} \sum_{ij} \left(w_{ij}^{+} - e_{ij}^{+} \right) \delta_{M_{i}M_{j}} - \frac{1}{\mathbf{b}^{+} + \mathbf{b}^{-}} \sum_{ii} \left(w_{ij}^{-} - e_{ij}^{-} \right) \delta_{M_{i}M_{j}}$$
(4)

where ν is the total weight of the network (sum of all negative and positive connections), w_{ij} is the weighted and signed connection between regions i and j, e_{ij} is the strength of a connection divided by the total weight of the network, and δ_{MiMj} is set to 1 when regions are in the same community and 0 otherwise. '+' and '–' superscripts denote all positive and negative connections, respectively. In our experiment, the γ parameter was set to 1.1. To determine this parameter, we ran the Louvain algorithm within a range of gamma = 0.5–2.0 across 100 iterations and then determined the value of gamma for which Q was most stable (i.e., had the lowest variance).

The diversity inherent within functional connectivity networks is such that different networks can play different topological roles in the network. The Participation Coefficient (PC) quantifies the extent to which a region connects across all modules. This measure has previously been used to characterize diversely connected hub regions within cortical brain networks (e.g., see Power et al., 2013). The PC was calculated for each of the 400 cortical parcels for each subject, where $\kappa_{\rm isT}$ is the strength of the positive connections of region i to regions in mod-

ule s, and κ_{iT} is the sum of strengths of all positive connections of region i. The participation coefficient of a region is therefore close to 1 if its connections are uniformly distributed among all the modules and 0 if all of its links are within its own module:

$$PC = 1 - \sum_{s=1}^{n_M} \left(\frac{\kappa_{isT}}{\kappa_{iT}}\right)^2 \tag{5}$$

To investigate the topological relationships between the Core and Matrix populations and network topology, we classified cortical regions into three distinct groups: Core (CP $_C < -0.1$), Matrix (CP $_C > 0.1$) and connector regions (i.e., parcels with PC greater than 0.5). We observed no overlap between these three groups and tested the significance of this null relationship using a permutation test (5000 permutations, p < 0.001). For interpretation, we also plotted these three populations onto a diffusion embedding graph (Fig. 5).

2.9. Time-varying topological dynamics

To quantify the relationship between CP_C and time-varying network topology, we used a windowed Pearson's correlation design to estimate patterns of undirected functional connectivity between all 400 cortical parcels. In contrast to typical sliding-window approaches (Lurie et al., 2020), we split regional time series data into 20 non-overlapping windows (each of 50 TRs). This approach was adopted so as to avoid artificial smoothing that occurs with strongly overlapping windows. The Pearson's correlation of each of these windows defined a functional connectivity matrix and we calculated the standard deviation of FC edges across all windows. The regional mean of this variability is then calculated and provides a measure of the stability of a given regions' functional connectivity signature. To mitigate against a bias resulting from windowing the time series, this analysis was run for five different initial window positions spanning the window size (10, 20, 30, 40, and 50 TRs), and the resulting FC variability for each region was averaged across these. We then correlated this measure with the CPc vector to test the hypothesis that Matrix-supported cortical regions demonstrated a greater temporal diversity of functional connectivity patterns than Core-supported cortical regions.

2.10. Statistical approach

We used non-parametric testing to determine statistical significance of the relationships identified across our study (Nichols and Holmes, 2002). A distribution of 5000 null Pearson's correlations was calculated for each comparison, against which the observed correlation was compared. Using this approach, p_{RAW} was calculated as the proportion of the null distribution that was less extreme than the observed correlation value.

We also conducted a separate set of statistical tests in an attempt to control for the prominent spatial autocorrelation associated with low-dimensional gradients in the brain. Specifically, for each spatial correlation assessed, gradients were projected back into FreeSurfer space. The cortical parcel locations were then rotated around the sphere (i.e., preserving spatial autocorrelation) by creating a random starting point (i.e., a value between 1-32,492) and re-orienting the data to this location (Reardon et al., 2018). The mean value for each parcel was then used to estimate a null distribution of correlations. A distribution of 1000 of these null Pearson's correlations was calculated for each comparison, against which the observed correlation was compared. This statistical test was denoted as p_{SPIN} , and was calculated as the proportion of the null distribution that was less extreme than the observed correlation value.

2.11. Reproducibility

To test the reproducibility of our results, we performed a separate replication analysis on 3T resting state fMRI data from the Human Connectome Project (100 subjects). Information regarding the specific imaging parameters from this data is presented elsewhere (Glasser et al., 2013). Data were pre-processed in the same fashion as the above analysis. Due to the fact that the comparisons were purely for replication purposes, statistical analyses were limited to traditional permutation tests (i.e., p_{RAW}).

2.12. Data and code availability

Data and code are openly available at https://github.com/macshine/

3. Results

3.1. Thalamic sub-populations are differentially correlated with the cerebral cortex

We observed differential functional coupling between thalamic populations, dissociated based on gene expression, and the cortex, CP_C (Fig. 2). Matrix regions of thalamus, which have higher expression of CALB1 (Fig. 1B), showed a strong preference for functional connectivity within trans-modal cortices, incorporating Default, Control, Limbic, and Ventral Attention sub-networks (Fig. 2A, blue). In contrast, the PVALB-rich Core cells showed preferential coupling to primary somatosensory cortices, as evidenced by their stronger correlations with parcels from the Visual, Somatomotor, Dorsal-Attention and Temporo-Parietal networks (Fig. 2B, green). Fig. 2C highlights the difference between Matrix (blue) and Core (green) functional connectivity patterns.

3.2. Matrix-core thalamus recapitulates primary cortical gradient

We observed a strong positive correlation between the first diffusion embedding gradient estimated from resting state fMRI data (Margulies and Smallwood, 2017) and the extent with which cortical parcels were preferentially connected with Matrix vs. Core thalamic voxels (r=0.772, $p_{RAW}=2.48\times10^{-80}$, $p_{SPIN}=1.0\times10^{-3}$; Fig. 3; see Fig. S2 for higher gradients). Consistent with these patterns, we observed a moderate negative correlation with t1w:t2w ratio from the HCP S1200 data release (r=-0.319; $p_{RAW}=6.6\times10^{-11}$, $p_{SPIN}=0.025$; Fig. 3B), which is an indirect proxy for cortical myelination (Glasser et al., 2016) that is known to be negatively correlated with the hierarchical level of the cerebral cortex (Burt et al., 2018).

3.3. Matrix thalamus preferentially couples to regions with slower timescales

We observed a significant positive correlation between CP_C and both: the intrinsic timescale, τ (mean = 3.52 \pm 0.3, r = 0.419, $p_{RAW}=1.9\times10^{-18}; p_{SPIN}=1.0\times10^{-3}; Fig~4A/B)$ and regional time fractality, Η $(mean = 0.78 \pm 0.03,$ series r = 0.245, $p_{RAW} = 7.2 \times 10^{-7}$, $p_{SPIN} = 0.045$; Fig 4C/D). We found that regional H and τ values were strongly positively correlated (r = 0.622, $p_{RAW}=4.0\times10^{-44},\,p_{SPIN}=1.0\times10^{-3}),$ suggesting that regions the longer integration windows in associative regions of the cerebral cortex (Murray et al., 2014) may be due to an increase in regional fractality (i.e., larger H value; Dong et al., 2018) and the presence of long-range correlations. Both timescale measures were also positively correlated with the first gradient of functional connectivity present within the cerebral cortex (τ : r=0.598, $p_{RAW}=3.61\times 10^{-40}$, $p_{SPIN}=1.0\times 10^{-3}$; H: r=0.272, $p_{RAW}=3\times 10^{-8}$; $p_{SPIN}=0.005$).

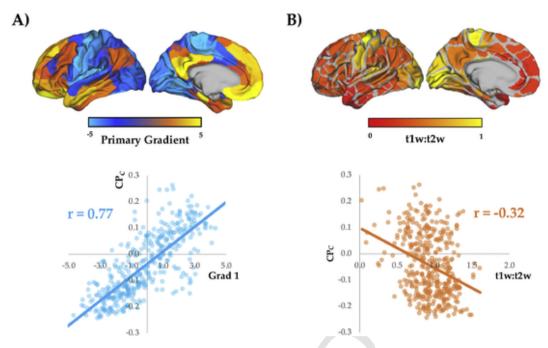


Fig. 3. Relationship between CP_C and low-dimensional cortical gradients. A) Surface plot of the first low-dimensional gradient (Margulies and Smallwood, 2017). We observed a positive Pearson's correlation with the CP_C (r = 0.772; $p = 2.48 \times 10^{-80}$); B) Surface plot of the t1w:t2w ratio on the cerebral cortex. We observed an inverse correlation with the CP_C (r = -0.320; $p = 5.8 \times 10^{-11}$).

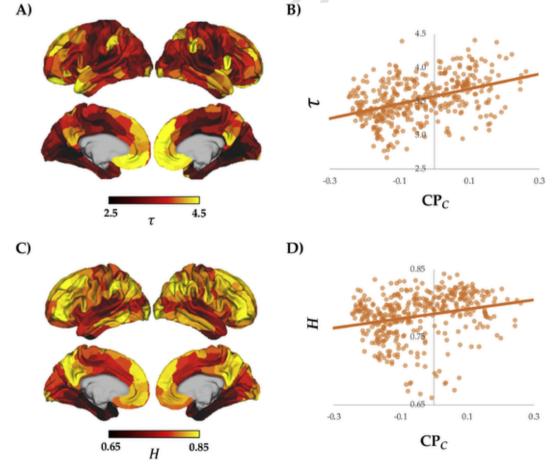


Fig. 4. Relationship between Thalamic Functional Connectivity and Cortical Intrinsic Timescale. A) The autocorrelation parameter, τ , in seconds, plotted onto the cerebral cortex; B) a scatter plot between τ and CP_C (r=0.419, p_{RAW} = 1.9 × 10⁻¹⁸; p_{SPIN} = 1.0 × 10⁻³); C) the Hurst exponent, H, plotted onto the cerebral cortex; D) a scatter plot between τ and CP_C (r=0.245, p_{RAW} = 7.2 × 10⁻⁷; p_{SPIN} = 0.045).

Together, these results suggest that the associative cortical regions preferentially supplied by the Matrix thalamus were associated with a longer intrinsic timescale when compared to the regions supported by the Core thalamus, at least during the resting state, and may support the notion of quasi-criticality in the resting brain (Moretti and Munoz, 2013). To test this hypothesis, we related CP_T (i.e., CALB1:PVALB ratio in thalamic nuclei) to the Hurst exponent calculated on thalamic timeseries. The resultant correlation was positive (r=0.366, p=0.042), suggesting that Matrix thalamic nuclei were active over longer timescales, and hence, were related to longer-timescale interactions in associative regions of the cerebral cortex.

3.4. Relationship between core and matrix thalamic populations and network topology

Based on the known importance of the thalamus for defining whole-brain network topological integration (Bell and Shine, 2015; Hwang et al., 2017), we next compared CP_C levels with measures that quantify the topology of resting functional connectivity. We did not observe a significant correlation between CP_C and PC (r=-0.022; $\mathrm{p}_{RAW}=0.663$; $\mathrm{p}_{SPIN}=0.603$). Despite the lack of strong linear relationships between CP_C and network topology, viewing the data within a force-directed plot highlighted a key topological relationship between CP_C and PC (Fig. 5A). Namely, highly integrative regions (PC > 50th percentile; Fig. 5A; red) were found to be topologically located between the populations of cortical parcels that were preferentially connected to either Matrix (Fig. 5A; blue) or Core (Fig. 5A; green) parcels when using a force-directed layout at 10% density. This suggests that highly integrative regions of the cerebral cortex topologically inter-connect regions on either end of the Core-Matrix thalamocortical gradient.

Based on the observation that Matrix thalamic nuclei and associative cerebral cortical regions were associated with fluctuations at longer time scales than Core thalamic or sensory cortical regions (i.e., regions with high $\mathrm{CP}_T/\mathrm{CP}_C$ had higher τ/H and vice versa), we predicted that regions with high CP_C should be associated with more variable functional connectivity dynamics. In essence, the ability to hold information over longer delays should afford more opportunities for fostering system-level variability. To test this hypothesis, we calcu-

lated the variability of windowed time-varying functional connectivity in the cerebral cortex, and then related this variability to the CP_C vector. As predicted, we observed a significant, positive correlation between functional connectivity variability and CP_C (r = 0.409; $p_{RAW} = 1.6 \times 10^{-17}$; $p_{SPIN} = 1.0 \times 10^{-3}$; Fig. 5B), suggesting that Matrix-supported cortical regions were associated with more variable functional connectivity dynamics than the Core-supported cortical regions that are more tethered to sensory inputs (Buckner and Krienen, 2015). Functional connectivity variability was also positively correlated with the first gradient of whole-brain time-averaged functional connectivity (r = 0.583, $p_{RAW} = 1.0 \times 10^{-37}$; $p_{SPIN} = 1.0 \times 10^{-3}$) as well as both timescale measures (τ : r = 0.657, $p_{RAW} = 5.8 \times 10^{-49}$; $p_{RAW} = 7.5 \times 10^{-42}$: $p_{SPIN} = 1.0 \times 10^{-3}$: H: r = 0.608 $p_{SPIN} = 1.0 \times 10^{-3}$) providing further evidence that these features all reflect a low-dimensional organizing principle for the thalamocortical system.

3.5. Reproducibility

The major results were replicated in data from the HCP. The CP_C maps calculated from the two datasets were positively correlated $(r=0.445,\,p_{RAW}=1.3\times10^{-17};\,\text{Fig. S3}),$ and the replicated CP_C map also correlated with the first diffusion embedding gradient $(r=0.334,\,p_{RAW}=4.0\times10^{-10})$ and $\tau(r=0.315,\,p_{RAW}=4.2\times10^{-9}),$ and inversely correlated with the t1w:t2w map $(r=-0.251,\,p_{RAW}=3.5\times10^{-6}).$ These results suggest that the thalamocortical measures identified in this study were reliable across independent datasets collected using different scanners and imaging protocols.

4. Discussion

Here, we analyze multi-modal neuroimaging and gene expression data to reveal how a core meso-scale organizing principle of thalamocortical interactions manifests in and shapes low dimensional gradients of time-averaged functional connectivity, temporal properties of resting brain dynamics, and whole-brain functional network topology. This relationship between the cortex and thalamus is consistent with a substantial wealth of both empirical (Garrett et al., 2018; Hwang et al., 2017; Olsen et al., 2012; Rikhye et al., 2018; Schmitt et

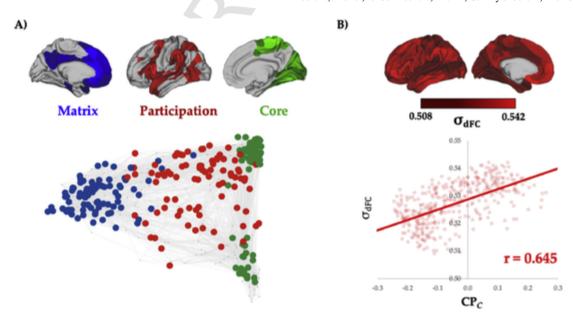


Fig. 5. Network Topology. A) Top: parcels within the top 100 ranked regions for each of: Matrix (blue), Participation (PC; red) and Core (green) populations; Bottom: an embedding plot, in which the top 270 nodes are depicted according to their association with either the Matrix (blue), high Participation regions (red) or Core (green) populations (the remaining 100 regions were plotted as small gray nodes), and edges represent the top 5% of network connections (resting-state functional connectivity); B) scatter plot between the standard deviation of dynamic functional connectivity (σ_{dFC}) and CP_C (r = 0.409, $p_{RAW} = 1.6 \times 10^{-17}$; $p_{SPIN} = 1.0 \times 10^{-3}$).

al., 2017; Shine et al., 2019b) and theoretical (Bell and Shine, 2016; Halassa and Sherman, 2019; Jones, 2009; Sherman, 2007) research, and suggests that an appreciation of the dynamics of subcortical–cortical interactions is crucial to understanding the organization of the human brain and the way it supports cognition and behavior.

Our results highlight a number of empirical signatures of cerebral cortical dynamics that relate directly to the mesoscale organization of the thalamus. There are a number of anatomical explanations for these patterns. For one, the thalamus has a vastly lower number of cells than the cortex, suggesting that its engagement would likely foster a relatively low-dimensional cortical architecture (Jones, 2001). Secondly, the different input patterns of the Core and Matrix to the cortex, which project to the granular and supragranular regions, would lead to a relatively feed-forward and feed-back mode of processing (García-Cabezas et al., 2019). This pattern can potentially explain the differences observed in the intrinsic timescales (Fig. 4), as feedback processing (via infragranular pyramidal cells inputs into supragranular layers of cortex) is known to occur on slower timescales than feed-forward processing (Bastos et al., 2012; Fries, 2005; Chaudhuri et al., 2015). Interestingly, there is also evidence that low-dimensional gradients in the cerebral cortex are associated with distinct electrophysiological signals (Hunt et al., 2016), suggesting that the patterns observed in this experiment may also exist at faster time scales than can be accurately measured with BOLD data. Finally, the thalamus represents a major synaptic input to the cortex, and thus likely plays a major role in shaping its functional network organization (Bell and Shine, 2016; Hwang et al., 2017; Shine et al., 2019b), both in BOLD data (as presented here) but also in electrophysiological data (Gollo, 2019; Watanabe et al., 2019). Together, these lines of reasoning suggest a putative explanation for how functional dynamics within the cerebral cortex may be shaped and constrained by activity patterns in the thalamus.

The thalamus has been argued to play a crucial role in shaping brain network topology (Bell and Shine, 2016; Hwang et al., 2017; Shine et al., 2019b). By interconnecting the cerebral cortex with key structures in the subcortex and brainstem, the thalamus likely plays an integrative, hub-like role (Bell and Shine, 2016; Hwang et al., 2017; Shine et al., 2019b). However, a closer inspection of thalamic circuitry suggests that the topological role of the thalamus may be qualitatively distinct from the hubs that exist within the cerebral cortex. For instance, although the thalamus is often identified as a hub in topological analyses of functional brain networks, its intrinsic activity is highly dampened by pervasive GABAergic input, both from the reticular nucleus, as well as GABAergic interneurons (Jones, 2001). As such, an increase in thalamic activity could promote both integration (by interconnecting otherwise distributed regions of the network) and segregation (by only allowing certain channels of activity to be on-line at any one point in time). Our results support this notion, as distinct populations of cells within the thalamus were found to interact with cortical regions organized into distinct topological zones (Fig. 5). Further work, particularly in the context of different cognitive tasks (Rikhye et al., 2018; Schmitt et al., 2017; Shine et al., 2019b), is undoubtedly required before the relationships between thalamic sub-populations and cortical network topology can be parsed at the functional level.

With these results in mind, an appreciation of the factors that bias activity within the thalamus become of prime importance. One major difference between the Core and Matrix populations is the extent to which they are driven by glutamatergic inputs from the sensory apparatuses, with Core neurons receiving a higher proportion of inputs when compared to Matrix neurons (Jones, 2009, 2001). Inputs from prominent subcortical structures are also known to disproportionately contact the Core and Matrix nuclei, with the deep cerebellar nuclei synapsing with the former, and the basal ganglia with the latter (Ku-

ramoto et al., 2009). Another factor that is of great importance is the neurochemical tone imposed by the ascending arousal system (Shine et al., 2016, 2018). It is well known that different neurotransmitters differentially effect cell populations in the thalamus, with some boosting and others silencing activity within the different nuclei (Llinás and Steriade, 2006; McCormick et al., 2015, 1991; Varela, 2014). Future work that maps the relationships between these neuromodulatory chemicals and cortico-thalamic functional connectivity will help to clarify the relationships described in this study. The importance of this work in understanding higher brain function is underscored by the fact that arousal, matrix thalamus and supragranular cortex are all causally involved in the processes underling consciousness (Redinbaugh et al., 2019; Suzuki and Larkum, 2020).

There are a number of limitations to our study that deserve consideration. For one, the granularity of the approach was such that the genetic expression for small thalamic regions could have been blended across multiple regions. Although the primary data used in our study was of high spatial resolution relative to many other whole-brain fMRI analyses in the field, the subtle relationships between the thalamus and cortex will undoubtedly benefit from higher resolution data. Secondly, the AHBA genetic expression was estimated from a small number of subjects (N = 6), which limits the ability to extrapolate these results to the broader population level, where individual differences might be found to represent important sources of variance (Arnatkevičiūtė et al., 2019). Finally, the Core vs. Matrix distinction represents two ends of a spectrum of different functional connectivity profiles (Jones, 2001; Clasca et al., 2012), and hence the proper delineation of thalamocortical functional connectivity patterns will likely require a more high-dimensional categorization.

5. Conclusion

Our results describe relationships between the relative weighting of distinct cell populations in the thalamus and low-dimensional spatial, temporal, and topological gradients in the cerebral cortex. It is important to note that many other elements within the central nervous system are also organized along spatial gradients, including the striatum (Stanley et al., 2019), cerebellum (Guell et al., 2018), colliculi (Cooper and Rakic, 1981) and hippocampus (Przeździk et al., 2019), suggesting that the interaction between the gradient structure across these diverse systems may represent a key organizing principle for the nervous system (Cisek, 2019; Nieuwenhuys, 1999). In addition, our work highlights the value of linking information from the cellular scale with whole-brain data to understand fundamental principles of brain organization. Future work incorporating high-resolution structural connectivity (Lambert et al., 2017; Iglehart et al., 2020) will also help to refine our understanding of how interactions between the thalamus and cerebral cortex help to shape the patterns that underlie cognitive func-

Uncited references

Morel et al., 1998, Redinbaugh et al., 2020, Buckner and Krienen, 2013, Wilting and Priesemann, 2018.

Acknowledgments

JMS was supported by the University of Sydney Robinson Fellowship and NHMRC GNT1156536.

Credit author statement

EJM, BM, JBS, DJL and JMS conceptualized the study LJH and LC collected and preprocessed the imaging data JMS wrote the initial draft JMS, BF and AA conducted the genetic analysis

All co-authors edited the final manuscript

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.neuroimage.2020.117224.

References

- Arnatkevičiūtė, A, Fulcher, B D, Fornito, A, 2019. A practical guide to linking brain-wide gene expression and neuroimaging data. Neuroimage 189, 353–367.
- Arai, R, Jacobowitz, D M, Deura, S, 1994. Distribution of calretinin, calbindin-D28k, and parvalbumin in the rat thalamus. Brain Res. Bull. 33, 595–614. doi:10.1016/0361-9230(94)90086-8.
- Bastos, A M, Usrey, W M, Adams, R A, Mangun, G R, Fries, P, Friston, K J, 2012. Canonical microcircuits for predictive coding. Neuron 76, 695–711. doi:10.1016/j.neuron.2012.10.038.
- Behzadi, Y, Restom, K, Liau, J, Liu, T T, 2007. A component based noise correction method (CompCor) for BOLD and perfusion based fMRI. Neuroimage 37, 90–101. doi:10.1016/j.neuroimage.2007.04.042.
- Bell, P T, Shine, J M, 2016. Subcortical contributions to large-scale network communication. Neurosci. Biobehav. Rev. 71, 313–322. doi:10.1016/ j.neubiorev.2016.08.036.
- Beran, J, 1992. Statistical methods for data with long-range dependence. Stat. Sci. 7, 404–427. doi:10.1214/ss/1177011123.
- Buckner, R L, Krienen, F M, 2013. The evolution of distributed association networks in the human brain. Trends Cogn. Sci. 17, 648–665. doi:10.1016/j.tics.2013.09.017.
- Burt, J B, Demirtaş, M, Eckner, W J, Navejar, N M, Ji, J L, Martin, W J, Bernacchia, A, Anticevic, A, Murray, J D, 2018. Hierarchy of transcriptomic specialization across human cortex captured by structural neuroimaging topography. Nat. Neurosci. 21, 1251–1259. doi:10.1038/s41593-018-0195-0.
- Chaudhuri, R, Knoblauch, K, Gariel, M-A, Kennedy, H, Wang, X-J, 2015. A large-scale circuit mechanism for hierarchical dynamical processing in the primate cortex. Neuron 88 (2), 419–431. doi:10.1016/j.neuron.2015.09.008.
- Churchill, N W, Spring, R, Grady, C, Cimprich, B, Askren, M K, Reuter-Lorenz, P A, Jung, M S, Peltier, S, Strother, S C, Berman, M G, 2016. The suppression of scale-free fMRI brain dynamics across three different sources of effort: aging, task novelty and task difficulty. Sci. Rep. 6, 30895. doi:10.1038/srep30895.
- Cisek, P, 2019. Resynthesizing behavior through phylogenetic refinement. Atten. Percept. Psychophys. 26, 535. doi:10.3758/s13414-019-01760-1.
- Clascá, F, Rubio-Garrido, P, Jabaudon, D, 2012. Unveiling the diversity of thalamocortical neuron subtypes. Eur. J. Neurosci. 35, 1524–1532. doi:10.1111/ j.1460-9568.2012.08033.x.
- Cooper, M L, Rakic, P, 1981. Neurogenetic gradients in the superior and inferior colliculi of the rhesus monkey. J. Comp. Neurol. 202, 309–334. doi:10.1002/cne.902020303.
- Demirtaş, M, Burt, J B, Helmer, M, Ji, J L, Adkinson, B D, Glasser, M F, Van Essen, D C, Sotiropoulos, S N, Anticevic, A, Murray, J D, 2019. Hierarchical heterogeneity across human cortex shapes large-scale neural dynamics. Neuron 101, 1181–1194. doi:10.1016/j.neuron.2019.01.017.
- Dong, J, Jing, B, Ma., X, Liu, H, Mo, X, Li, H, 2018. Hurst exponent analysis of resting-state fMRI signal complexity across the adult lifespan. Front. Neurosci. 12, 34. doi:10.3389/fnins.2018.00034.
- Fallon, J, Ward, P, Parkes, L, Oldham, S, Arnatkeviciūte, A, Fornito, A, Fulcher, B, 2020. Timescales of spontaneous activity fluctuations relate to structural connectivity in the brain. bioRxiv 1 (4), 655050. doi:10.1101/655050.
- Fries, P, 2005. A mechanism for cognitive dynamics: neuronal communication through neuronal coherence. Trends Cogn. Sci. Regul. Ed. 9, 474–480. doi:10.1016/ j.tics.2005.08.011.
- García-Cabezas, MÁ, Zikopoulos, B, Barbas, H, 2019. The structural model: a theory linking connections, plasticity, pathology, development and evolution of the cerebral cortex. Brain Struct. Funct. 72, 429. doi:10.1007/s00429-019-01841-9.
- Garrett, D D, Epp, S M, Perry, A, Lindenberger, U, 2018. Local temporal variability reflects functional integration in the human brain. Neuroimage 183, 776–787. doi:10.1016/ j.neuroimage.2018.08.019.
- Glasser, M F, Coalson, T S, Robinson, E C, Hacker, C D, 2016. A multi-modal parcellation of human cerebral cortex. Nature 536, 171–178. doi:10.1038/nature18933.
- Glasser, M F, Sotiropoulos, S N, Wilson, J A, Coalson, T S, Fischl, B, Andersson, J L, Xu, J, Jbabdi, S, Webster, M, Polimeni, J R, Van Essen, D C, Jenkinson, M, 2013. The minimal preprocessing pipelines for the human connectome project. Neuroimage 80, 105–124. doi:10.1016/j.neuroimage.2013.04.127.
- Gollo, L L, 2019. Exploring atypical timescales in the brain. Elife 8, e15252. doi:10.7554/ eLife.45089.
- Gorgolewski, K J, Varoquaux, G, Rivera, G, Schwartz, Y, Sochat, V V, Ghosh, S S, Maumet, C, Nichols, T E, Poline, J-B, Yarkoni, T, Margulies, D S, Poldrack, R A, 2015. NeuroVault.org: a repository for sharing unthresholded statistical maps, parcellations, and atlases of the human brain. NeuroImage.. doi:10.1016/j.neuroimage.2015.04.016.
- Gryglewski, G, Seiger, R, James, G M, Godbersen, G M, Komorowski, A, Unterholzner, J, Michenthaler, P, Hahn, A, Wadsak, W, Mitterhauser, M, Kasper, S, Lanzenberger, R, 2018. Spatial analysis and high resolution mapping of the human whole-brain transcriptome for integrative analysis in neuroimaging. Neuroimage 176, 259–267. doi:10.1016/j.neuroimage.2018.04.068.
- Guell, X, Schmahmann, J D, Gabrieli, J D, Ghosh, S S, 2018. Functional gradients of the cerebellum. Elife 7, 568. doi:10.7554/eLife.36652.
- Halassa, M M, Sherman, S M, 2019. Thalamocortical circuit motifs: a general framework. Neuron 103, 762–770. doi:10.1016/j.neuron.2019.06.005.

- Hawrylycz, M J, Lein, E S, Guillozet-Bongaarts, A L, Shen, E H, Ng, L, Miller, J A, van de Lagemaat, L N, Smith, K A, Ebbert, A, Riley, Z L, Abajian, C, Beckmann, C F, Bernard, A, Bertagnolli, D, Boe, A F, Cartagena, P M, Chakravarty, M M, Chapin, M, Chong, J, Dalley, R A, David Daly, B, Dang, C, Datta, S, Dee, N, Dolbeare, T A, Faber, V, Feng, D, Fowler, D R, Goldy, J, Gregor, B W, Haradon, Z, Haynor, D R, Hohmann, J G, Horvath, S, Howard, R E, Jeromin, A, Jochim, J M, Kinnunen, M, Lau, C, Lazarz, E T, Lee, C, Lemon, T A, Li, L, Li, Y, Morris, J A, Overly, C C, Parker, P D, Parry, S E, Reding, M, Royall, J J, Schulkin, J, Sequeira, P A, Slaughterbeck, C R, Smith, S C, Sodt, A J, Sunkin, S M, Swanson, B E, Vawter, M P, Williams, D, Wohnoutka, P, Zielke, H R, Geschwind, D H, Hof, P R, Smith, S M, Koch, C, Grant, S G N, Jones, A R, 2012. An anatomically comprehensive atlas of the adult human brain transcriptome. Nature 489, 391–399. doi:10.1038/nature11405.
- He, J, 2011. A new fractal derivation. Therm. Sci. 15, S145–S147. doi:10.2298/tsci11s1145h.
- Herkenham, M, 1979. The afferent and efferent connections of the ventromedial thalamic nucleus in the rat. J. Comp. Neurol. 183, 487–517. doi:10.1002/cne.901830304.
- Hunt, B A E, Tewarie, P K, Mougin, O E, Geades, N, Jones, D K, Singh, K D, Morris, P G, Gowland, P A, Brookes, M J, 2016. Relationships between cortical myeloarchitecture and electrophysiological networks. PNAS 113, 13510–13515. doi:10.1073/pnas.1608587113.
- Hwang, K, Bertolero, M A, Liu, W B, D'Esposito, M, 2017. The human thalamus is an integrative hub for functional brain networks. J. Neurosci. 37, 5594–5607. doi:10.1523/ineurosci.0067-17.2017.
- Iglehart, C, Monti, M, Cain, J, Tourdias, T, Sranthan, M, 2020. 2020. A Systematic comparison of structural-, structural connectivity-, and functional connectivity-based thalamus parcellation techniques. Brain Struct. Funct..
- Jones, E.G., 2009. Synchrony in the interconnected circuitry of the thalamus and cerebral cortex. Ann. N. Y. Acad. Sci. 1157, 10–23. doi:10.1111/j.1749-6632.2009.04534.x.
- Jones, E G, 2001. The thalamic matrix and thalamocortical synchrony. Trends Neurosci. 24, 595–601. doi:10.1016/s0166-2236(00)01922-6.
- Kuramoto, E, Futura, T, Nakamura, K C, Unzai, T, Kioki, H, Kaneko, T, 2009. Two types of thalamocortical projections from the motor thalamic nuclei of the rat: a single neuron-tracing study using viral vectors. Cereb. Cortex 19, 2065–2077. doi:10.1093/ cercor/bhn231.
- Lambert, C, Simon, H, Colman, J, Barrick, R T, 2017. Defining thalamic nuclei and topographic connectivity gradients in vivo. Neuroimage 158, 466–479.
- Llinás, R R, Steriade, M, 2006. Bursting of thalamic neurons and states of vigilance. J. Neurophysiol. 95, 3297–3308. doi:10.1152/jn.00166.2006.
- Lurie, D, Kessler, D, Bassett, D S, Betzel, R F, Breakspear, M, Kielholz, S, Kuyci, A, Liégeois, R, Lindquist, M A, McIntosh, A R, Poldrack, R A, Shine, J M, Thompson, W H, Bielczyk, N Z, Douw, L, Kraft, D, Miller, R L, Muthuramann, M, Pasquini, L, Razi, A, Viduarre, D, Xie, H, Calhoun, V D, 2020. Questions and controversies in the study of time-varying functional connectivity in resting fMRI. Netw. Neurosci. 4, 30–69. doi:10.1162/netn_a_00116.
- Margulies, D S, Ghosh, S S, Goulas, A, Falkiewicz, M, Huntenburg, J M, Langs, G, Bezgin, G, Eickhoff, S B, Castellanos, F X, Petrides, M, Jefferies, E, Smallwood, J, 2016. Situating the default-mode network along a principal gradient of macroscale cortical organization. Proc. Natl. Acad. Sci. U.S.A. 113, 12574–12579. doi:10.1073/pnas.1608282113.
- Margulies, D S, Smallwood, J, 2017. Converging evidence for the role of transmodal cortex in cognition. Proc. Natl. Acad. Sci. U.S.A. 114, 12641–12643. doi:10.1073/ pnas.1717374114.
- McCormick, D A, McGinley, M J, Salkoff, D B, 2015. Brain state dependent activity in the cortex and thalamus. Curr. Opin. Neurobiol. 31, 133–140. doi:10.1016/ j.conb.2014.10.003.
- McCormick, D A, Pape, H C, Williamson, A, 1991. Actions of norepinephrine in the cerebral cortex and thalamus: implications for function of the central noradrenergic system. Prog. Brain Res. 88, 293–305. doi:10.1016/s0079-6123(08)63817-0.
- Mesulam, M M, 1998. From sensation to cognition. Brain 121 (Pt 6), 1013–1052. doi:10.1093/brain/121.6.1013.
- Morel, A, Magnin, M, Jeanmonod, D, 1998. Multiarchitectonic and stereotactic atlas of the human thalamus. J. Comp. Anat. 387 (4), 588–630.
- Moretti, P, Munoz, M A, 2013. Griffiths phases and the stretching of criticality in brain networks. Nat. Commun. 4, 2521. doi:10.1038/ncomms3521.
- Münkle, M C, Waldvogel, H J, Faull, R L, 2000. The distribution of calbindin, calretinin and parvalbumin immunoreactivity in the human thalamus. J. Chem. Neuroanat. 19, 155–173. doi:10.1016/s0891-0618(00)00060-0.
- Munn, B, Zeater, N, Pietersen, A N, Solomon, S G, Cheong, S K, Martin, P R, Gong, P, 2020.
 Fractal spike dynamics and neuronal coupling in the primate visual system. J. Physiol. In Press. doi:10.1113/jp278935.
- Murray, J D, Benacchia, A, Freedman, D J, Romo, R, Wallis, J D, Cai, X, Padoa-Schioppa, C, Pasternak, T, Seo, H, Lee, D, Wang, X-J, 2014. A hierarchy of intrinsic timescales across primate cortex. Nat. Neurosci. 17, 1661–1663. doi:10.1038/nn.3862.
- Nichols, T E, Holmes, A P, 2002. Nonparametric permutation tests for functional neuroimaging: a primer with examples. Hum. Brain Mapp. 15, 1–25. doi:10.1002/ hbm.1058.
- Niemann, K, Mennicken, V R, Jeanmonod, D, Morel, A, 2000. The Morel stereotactic atlas of the human thalamus: atlas-to-MR registration of internally consistent canonical model. Neuroimage 12, 601–616. doi:10.1006/nimg.2000.0650.
- Nieuwenhuys, R, 1999. The morphological pattern of the vertebrate brain. Eur. J. Morphol. 37, 81–84. doi:10.1076/ejom.37.2.81.4746.
- Olsen, S R, Bortone, D S, Adesnik, H, Scanziani, M, 2012. Gain control by layer six in cortical circuits of vision. Nature 483, 47–52. doi:10.1038/nature10835.
- Paquola, C, Vos De Wael, R, Wagstyl, K, Bethlehem, R A I, Hong, S-J, Seidlitz, J, Bullmore, E T, Evans, A C, Mišić, B, Margulies, D S, Smallwood, J, Bernhardt, B C, 2019. Microstructural and functional gradients are increasingly dissociated in transmodal cortices. Plos Biol. 17, e3000284. doi:10.1371/journal.pbio.3000284.
- Peng, C K, Havlin, S, Stanley, H E, Goldberger, A L, 1995. Quantification of scaling exponents and crossover phenomena in nonstationary heartbeat time series. Chaos 5, 82–87. doi:10.1063/1.166141.

Phillips, J W, Schulmann, A, Hara, E, Winnubst, J, Liu, C, Valakh, V, Wang, L, Shields, B C, Korff, W, Chandrashekar, J, Lemire, A L, Mensh, B, Dudman, J T, Nelson, S B, Hantman, A W, 2019. A repeated molecular architecture across thalamic pathways. Nat. Neurosci. 22, 1925–1935. doi:10.1038/s41593-019-0483-3.

- Power, J D, Schlaggar, B L, Lessov-Schlaggar, C N, Petersen, S E, 2013. Evidence for hubs in human functional brain networks. Neuron 79, 798–813. doi:10.1016/ j.neuron.2013.07.035.
- Przeździk, I, Faber, M, Fernández, G, Beckmann, C F, Haak, K V, 2019. The functional organisation of the hippocampus along its long axis is gradual and predicts recollection. Cortex 119, 324–335. doi:10.1016/j.cortex.2019.04.015.
- Reardon, P K, Seidlitz, J, Vandekar, S, Liu, S, Patel, R, Park, M T M, Alexander-Bloch, A, Clasen, L S, Blumenthal, J D, Lalonde, F M, Giedd, J N, Gur, R C, Gur, R E, Lerch, J P, Chakravarty, M M, Satterthwaite, T D, Shinohara, R T, Raznahan, A, 2018. Normative brain size variation and brain shape diversity in humans. Science 360, 1222–1227. doi:10.1126/science.aar.9578
- Redinbaugh, M J, Phillips, J M, Kambi, N A, Mohanta, S, Andryk, S, Dooley, G L, Afrasiabi, M, Raz, A, Saalman, Y, 2020. Thalamus modulates consciousness via layer-specific control of cortex. Neuron In Press. doi:10.1016/j.neuron.2020.01.005.
- Rikhye, R V, Gilra, A, Halassa, M M, 2018. Thalamic regulation of switching between cortical representations enables cognitive flexibility. Nat. Neurosci. 21, 1753–1763. doi:10.1038/s41593-018-0269-z.
- Rubinov, M, Sporns, O, 2010. Complex network measures of brain connectivity: uses and interpretations. Neuroimage 52, 1059–1069. doi:10.1016/j.neuroimage.2009.10.003.
- Schaefer, A, Kong, R, Gordon, E M, Laumann, T O, Zuo, X-N, Holmes, A J, Eickhoff, S B, Yeo, B T T, 2018. Local-global parcellation of the human cerebral cortex from intrinsic functional connectivity MRI. Cereb. Cortex 28, 3095–3114. doi:10.1093/cercor/bbx179.
- Schmitt, L I, Wimmer, R D, Nakajima, M, Happ, M, Mofakham, S, Halassa, M M, 2017. Thalamic amplification of cortical connectivity sustains attentional control. Nature 545, 219–223. doi:10.1038/nature22073.
- Sherman, S M, 2007. The thalamus is more than just a relay. Curr. Opin. Neurobiol. 17, 417–422. doi:10.1016/j.conb.2007.07.003.
- Shine, J M, Aburn, M J, Breakspear, M, Poldrack, R A, 2018. The modulation of neural gain facilitates a transition between functional segregation and integration in the brain. Elife 7, e31130. doi:10.7554/eLife.31130.001.

- Shine, J M, Bissett, P G, Bell, P T, Koyejo, O, Balsters, J H, Gorgolewski, K J, Moodie, C A, Poldrack, R A, 2016. The dynamics of functional brain networks: integrated network states during cognitive task performance. Neuron 92, 544–554. doi:10.1016/j.neuron.2016.09.018.
- Shine, J M, Breakspear, M, Bell, P T, Ehgoetz Martens, K A, Koyejo, O, Shine, R, Sporns, O, Poldrack, R A, 2019. Human cognition involves the dynamic integration of neural activity and neuromodulatory systems. Nat. Neurosci. 28, 289–296. doi:10.1038/s41593-018-0312-0.
- Shine, J M, Hearne, L J, Breakspear, M, Hwang, K, Müller, E J, Sporns, O, Poldrack, R A, Mattingley, J B, Cocchi, L, 2019. The low-dimensional neural architecture of cognitive complexity is related to activity in medial thalamic nuclei. Neuron 104, 849–855. doi:10.1016/j.neuron.2019.09.002.
- Stanley, G, Gokce, O, Malenka, R C, Südhof, T C, Quake, S R, 2019. Continuous and discrete neuron types of the adult murine striatum. Neuron 105, 688–699. doi:10.1016/j.neuron.2019.11.004.
- Suzuki, M, Larkum, M E, 2020. General anesthesia decouples cortical pyramidal neurons. Cell 180, 666–676. doi:10.1016/j.cell.2020.01.024.
- Varela, C, 2014. Thalamic neuromodulation and its implications for executive networks. Front. Neural Circuits 8, 69. doi:10.3389/fncir.2014.00069.
- Vázquez-Rodríguez, B, Suárez, L E, Markello, R D, Shafiei, G, Paquola, C, Hagmann, P, van den Heuvel, M P, Bernhardt, B C, Spreng, R N, Mišić, B, 2019. Gradients of structure-function tethering across neocortex. Proc. Natl. Acad. Sci. U.S.A. 116, 21219–21227. doi:10.1073/pnas.1903403116.
- Watanabe, T, Rees, G, Masuda, N, 2019. Atypical intrinsic neural timescale in autism. Elife 8, 839. doi:10.7554/eLife.42256.
- Wilting, J, Priesemann, V, 2018. Inferring collective dynamical states from widely unobserved systems. Nat. Commun. 9, 2325. doi:10.1038/s41467-018-04725-4.
- Yeo, B T, Krienen, F M, Sepulcre, J, Sabuncu, M R, Lashkari, D, Hollinshead, M, Roffman, J L, Smoller, J W, Zöllei, L, Polimeni, J R, Fischl, B, Liu, H, Buckner, R L, 2011. The organization of the human cerebral cortex estimated by intrinsic functional connectivity. J. Neurophysiol. 106 (3), 1125–1165. doi:10.1152/jn.00338.2011.