Homework 3

Insert your name here

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! Important

Please read the instructions carefully before submitting your assignment.

- 1. This assignment requires you to only upload a PDF file on Canvas
- 2. Don't collapse any code cells before submitting.
- 3. Remember to make sure all your code output is rendered properly before uploading your submission.

Please add your name to the author information in the frontmatter before submitting your assignment

For this assignment, we will be using the Wine Quality dataset from the UCI Machine Learning Repository. The dataset consists of red and white *vinho verde* wine samples, from the north of Portugal. The goal is to model wine quality based on physicochemical tests

We will be using the following libraries:

```
library(readr)
  library(tidyr)
  library(dplyr)
Attaching package: 'dplyr'
The following objects are masked from 'package:stats':
    filter, lag
The following objects are masked from 'package:base':
    intersect, setdiff, setequal, union
  library(purrr)
  library(car)
Loading required package: carData
Attaching package: 'car'
The following object is masked from 'package:purrr':
    some
The following object is masked from 'package:dplyr':
    recode
  library(glmnet)
Loading required package: Matrix
```

```
Attaching package: 'Matrix'

The following objects are masked from 'package:tidyr':

expand, pack, unpack

Loaded glmnet 4.1-8

library(broom)
library(corrplot)

corrplot 0.92 loaded
```

Question 1



Regression with categorical covariate and t-Test

1.1 (5 points)

Read the wine quality datasets from the specified URLs and store them in data frames df1 and df2.

```
url1 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequalit
url2 <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequalit
df1 <- read.csv(url1, sep = ";")
df2 <- read.csv(url2, sep = ";")</pre>
```

1.2 (5 points)

Perform the following tasks to prepare the data frame df for analysis:

- 1. Combine the two data frames into a single data frame df, adding a new column called type to indicate whether each row corresponds to white or red wine.
- 2. Rename the columns of df to replace spaces with underscores
- 3. Remove the columns fixed_acidity and free_sulfur_dioxide
- 4. Convert the type column to a factor
- 5. Remove rows (if any) with missing values.

```
#1
  df1$type <- "white"
  df2$type <- "red"
  df <- rbind(df1, df2)</pre>
  #2
  colnames(df) <- gsub("\\.", "_", colnames(df))</pre>
  #3
  df <- df %>%
    select(-fixed_acidity, -free_sulfur_dioxide)
  df$type <- as.factor(df$type)</pre>
  #5
  nrow(df)
[1] 6497
  df <- na.omit(df)</pre>
  dim(df) #none were removed
[1] 6497
            11
Your output to R dim(df) should be
[1] 6497
            11
  head(df)
```

```
volatile_acidity citric_acid residual_sugar chlorides total_sulfur_dioxide
              0.27
                           0.36
1
                                            20.7
                                                     0.045
                                                                              170
              0.30
2
                           0.34
                                             1.6
                                                     0.049
                                                                              132
3
              0.28
                           0.40
                                             6.9
                                                     0.050
                                                                               97
4
                                                     0.058
              0.23
                           0.32
                                             8.5
                                                                              186
5
              0.23
                           0.32
                                             8.5
                                                     0.058
                                                                              186
6
              0.28
                           0.40
                                             6.9
                                                     0.050
                                                                               97
 density
            pH sulphates alcohol quality type
  1.0010 3.00
                     0.45
                              8.8
                                         6 white
1
2 0.9940 3.30
                     0.49
                               9.5
                                         6 white
3 0.9951 3.26
                     0.44
                             10.1
                                         6 white
4 0.9956 3.19
                     0.40
                              9.9
                                         6 white
5 0.9956 3.19
                     0.40
                              9.9
                                         6 white
6 0.9951 3.26
                     0.44
                             10.1
                                         6 white
```

1.3 (20 points)

Recall from STAT 200, the method to compute the t statistic for the difference in means (with the equal variance assumption)

- 1. Using df compute the mean of quality for red and white wine separately, and then store the difference in means as a variable called diff_mean.
- 2. Compute the pooled sample variance and store the value as a variable called sp_squared.
- 3. Using sp_squared and diff_mean, compute the t Statistic, and store its value in a variable called t1.

```
# 1
mean_red <- mean(df$quality[df$type == "red"])
mean_white <- mean(df$quality[df$type == "white"])
diff_mean <- mean_red - mean_white

# 2
n_red <- sum(df$type == "red")
n_white <- sum(df$type == "white")
var_red <- var(df[df$type == "red", "quality"])
var_white <- var(df[df$type == "white", "quality"])
sp_squared <- ((n_red - 1) * var_red + (n_white - 1) * var_white) / (n_red + n_white - 2)
sp = sqrt(sp_squared)
# 3</pre>
```

```
t1 <- diff_mean / sqrt(sp_squared * (1/n_red + 1/n_white))
```

1.4 (10 points)

Equivalently, R has a function called t.test() which enables you to perform a two-sample t-Test without having to compute the pooled variance and difference in means.

Perform a two-sample t-test to compare the quality of white and red wines using the t.test() function with the setting var.equal=TRUE. Store the t-statistic in t2.

```
t_test <- t.test(quality ~ type, data = df, var.equal = TRUE)
t2 <- t_test$statistic</pre>
```

1.5 (5 points)

Fit a linear regression model to predict quality from type using the lm() function, and extract the t-statistic for the type coefficient from the model summary. Store this t-statistic in t3.

```
fit <- lm(quality ~ type, data = df)
summary_coef <- summary(fit)$coefficients
t_stat <- summary_coef["typewhite", "t value"]
t3 <- t_stat</pre>
```

1.6 (5 points)

Print a vector containing the values of t1, t2, and t3. What can you conclude from this? Why?

```
c(t1, t2, t3) # Insert your code here
```

```
-9.68565 -9.68565 9.68565
```

Since t1 and t2 are postive while t3 is negative we see that there is a significant difference in mean quality between red and white wines.

Also, since the magnitive of each is the same we know that Welch's t-test and the two-sample t-test yield similar results.

Question 2



Collinearity

2.1 (5 points)

Fit a linear regression model with all predictors against the response variable quality. Use the broom::tidy() function to print a summary of the fitted model. What can we conclude from the model summary?

```
lm_model <- lm(quality ~ ., data = df)
summary(lm_model)</pre>
```

Call:

```
lm(formula = quality ~ ., data = df)
```

Residuals:

```
Min 1Q Median 3Q Max -3.3415 -0.4725 -0.0405 0.4573 3.1140
```

Coefficients:

```
Estimate Std. Error t value Pr(>|t|)

(Intercept) 5.753e+01 9.331e+00 6.166 7.44e-10 ***

volatile_acidity -1.609e+00 8.057e-02 -19.965 < 2e-16 ***

citric_acid 2.721e-02 7.833e-02 0.347 0.72827

residual_sugar 4.509e-02 4.158e-03 10.844 < 2e-16 ***

chlorides -9.639e-01 3.328e-01 -2.897 0.00378 **
```

Residual standard error: 0.7371 on 6486 degrees of freedom Multiple R-squared: 0.2887, Adjusted R-squared: 0.2876 F-statistic: 263.3 on 10 and 6486 DF, p-value: < 2.2e-16

tidy(lm_model)

A tibble: 11 x 5

	term	estimate	std.error	statistic	p.value
	<chr></chr>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
1	(Intercept)	57.5	9.33	6.17	7.44e-10
2	volatile_acidity	-1.61	0.0806	-20.0	4.07e-86
3	citric_acid	0.0272	0.0783	0.347	7.28e- 1
4	residual_sugar	0.0451	0.00416	10.8	3.64e-27
5	chlorides	-0.964	0.333	-2.90	3.78e- 3
6	total_sulfur_dioxide	-0.000329	0.000262	-1.25	2.10e- 1
7	density	-55.2	9.32	-5.92	3.34e- 9
8	рН	0.188	0.0661	2.85	4.38e- 3
9	sulphates	0.662	0.0758	8.73	3.21e-18
10	alcohol	0.277	0.0142	19.5	1.87e-82
11	typewhite	-0.386	0.0549	-7.02	2.39e-12

I found that volatile acidity, residual sugar, chlorides, density, sulphates, alcohol, and the type of wine are significant predictors of wine quality. On the other hand, variables like citric acid and total sulfur dioxide do not seem to have a significant effect on quality.

2.2 (10 points)

Fit two **simple** linear regression models using lm(): one with only citric_acid as the predictor, and another with only total_sulfur_dioxide as the predictor. In both models, use quality as the response variable. How does your model summary compare to the summary from the previous question?

```
model_citric <- lm(quality ~ citric_acid, data = df)</pre>
  model_sulfur <- lm(quality ~ total_sulfur_dioxide, data = df)</pre>
  summary(model_citric)
Call:
lm(formula = quality ~ citric_acid, data = df)
Residuals:
    Min
             1Q Median
                             3Q
                                    Max
-2.9938 -0.7831 0.1552 0.2426 3.1963
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) 5.65461
                       0.02602 217.343
                                          <2e-16 ***
citric_acid 0.51398
                        0.07429
                                           5e-12 ***
                                  6.918
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 0.8701 on 6495 degrees of freedom
Multiple R-squared: 0.007316, Adjusted R-squared: 0.007163
F-statistic: 47.87 on 1 and 6495 DF, p-value: 5.002e-12
  summary(model_sulfur)
Call:
lm(formula = quality ~ total_sulfur_dioxide, data = df)
Residuals:
             1Q Median
    Min
                             3Q
-2.8866 -0.7971 0.1658 0.2227 3.1965
Coefficients:
                       Estimate Std. Error t value Pr(>|t|)
                      5.8923848 0.0246717 238.831 < 2e-16 ***
(Intercept)
total_sulfur_dioxide -0.0006394  0.0001915  -3.338  0.000848 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
Residual standard error: 0.8726 on 6495 degrees of freedom Multiple R-squared: 0.001713, Adjusted R-squared: 0.001559 F-statistic: 11.14 on 1 and 6495 DF, p-value: 0.000848
```

The reuslts are very comparable to the previous one. They show that, variables like citric acid and total sulfur dioxide do not seem to have a significant effect on quality.

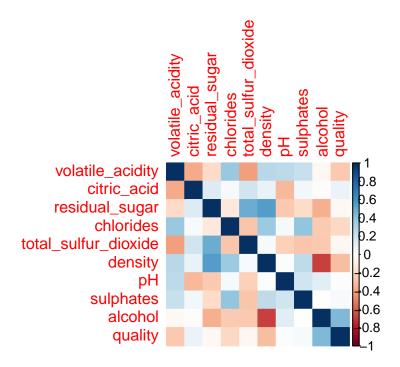
2.3 (5 points)

Visualize the correlation matrix of all numeric columns in df using corrplot()

```
numeric_df <- df %>% select_if(is.numeric)

correlation_matrix <- cor(numeric_df)

corrplot(correlation_matrix, method = "color")</pre>
```



2.4 (5 points)

Compute the variance inflation factor (VIF) for each predictor in the full model using vif() function. What can we conclude from this?

```
lm_model <- lm(quality ~ ., data = df)</pre>
vif_scores <- vif(lm_model)</pre>
vif_scores
 volatile_acidity
                             citric_acid
                                                 residual_sugar
          2.103853
                                 1.549248
                                                        4.680035
         chlorides total_sulfur_dioxide
                                                         density
          1.625065
                                 2.628534
                                                        9.339357
                                sulphates
                                                         alcohol
                рΗ
```

6.694679

1.352005

type

The VIF scores indicate that there is potential multicollinearity in density and type, where VIF values exceeded the common threshold of 10.

1.522809

3.419849

Question 3



• 40 points

Variable selection

3.1 (5 points)

Run a backward stepwise regression using a full_model object as the starting model. Store the final formula in an object called backward_formula using the built-in formula() function

```
full_model <- lm(quality ~ ., data = df)</pre>
```

```
backward_model <- step(full_model, direction = "backward")</pre>
Start: AIC=-3953.43
quality ~ volatile acidity + citric acid + residual sugar + chlorides +
    total_sulfur_dioxide + density + pH + sulphates + alcohol +
    type
                      Df Sum of Sq
                                      RSS
                                              AIC
                              0.066 3523.6 -3955.3
- citric_acid
                        1
- total_sulfur_dioxide 1
                              0.854 3524.4 -3953.9
                                    3523.5 -3953.4
<none>
                        1
                             4.413 3527.9 -3947.3
- pH
- chlorides
                            4.559 3528.1 -3947.0
                       1 19.054 3542.6 -3920.4
- density
                       1 26.794 3550.3 -3906.2
- type
                       1 41.399 3564.9 -3879.5
- sulphates
                       1 63.881 3587.4 -3838.7
- residual_sugar
- alcohol
                       1 206.860 3730.4 -3584.8
- volatile acidity
                       1 216.549 3740.0 -3567.9
Step: AIC=-3955.3
quality ~ volatile_acidity + residual_sugar + chlorides + total_sulfur_dioxide +
    density + pH + sulphates + alcohol + type
                      Df Sum of Sq
                                               AIC
                                       RSS
                             0.818 3524.4 -3955.8
- total_sulfur_dioxide 1
<none>
                                    3523.6 -3955.3
                       1
                             4.495 3528.1 -3949.0
- chlorides
                            4.536 3528.1 -3948.9
- pH
                          20.794 3544.4 -3919.1
- density
                       1
- type
                       1 26.943 3550.5 -3907.8
- sulphates
                       1 41.491 3565.1 -3881.2
- residual_sugar
                       1
                           67.371 3590.9 -3834.3
- alcohol
                       1
                           235.151 3758.7 -3537.6
- volatile_acidity
                       1
                           252.565 3776.1 -3507.5
Step: AIC=-3955.8
quality ~ volatile_acidity + residual_sugar + chlorides + density +
   pH + sulphates + alcohol + type
                   Df Sum of Sq
                                   RSS
                                           AIC
                                3524.4 -3955.8
<none>
```

```
1
                        4.295 3528.7 -3949.9
- pH
- chlorides
                        4.523 3528.9 -3949.5
                 1
                  1 21.540 3545.9 -3918.2
- density
- sulphates
                 1 40.711 3565.1 -3883.2
- type
                  1 43.664 3568.0 -3877.8
- alcohol 1 244.545 3768.9 -3521.9
- volatile_acidity 1 256.695 3781.1 -3501.0
  backward_formula <- formula(backward_model)</pre>
  backward_formula
quality ~ volatile_acidity + residual_sugar + chlorides + density +
   pH + sulphates + alcohol + type
3.2 (5 points)
Run a forward stepwise regression using a null_model object as the starting model. Store the
final formula in an object called forward_formula using the built-in formula() function in
R
  null_model <- lm(quality ~ 1, data = df)</pre>
  forward_model <- step(null_model, direction = "forward", scope = formula(~ .), data = df)</pre>
Start: AIC=-1760.04
quality ~ 1
  forward_formula <- formula(forward_model)</pre>
  forward_formula
quality ~ 1
```

3.3 (10 points)

- 1. Create a y vector that contains the response variable (quality) from the df dataframe.
- 2. Create a design matrix X for the full_model object using the make_model_matrix() function provided in the Appendix.
- 3. Then, use the cv.glmnet() function to perform LASSO and Ridge regression with X and y.

```
... # Insert your code here.
```

Create side-by-side plots of the ridge and LASSO regression results. Interpret your main findings.

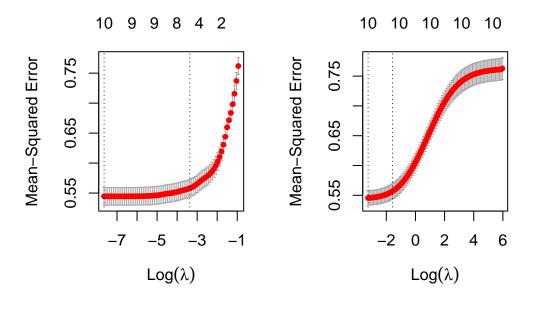
```
# 1
y <- df$quality

# 2
X <- as.matrix(model.matrix(full_model))

# 3
lasso_model <- cv.glmnet(X, y, alpha = 1)

ridge_model <- cv.glmnet(X, y, alpha = 0)

par(mfrow=c(1, 2))
plot(lasso_model)
plot(ridge_model)</pre>
```



3.4 (5 points)

Print the coefficient values for LASSO regression at the lambda.1se value? What are the variables selected by LASSO?

Store the variable names with non-zero coefficients in lasso_vars, and create a formula object called lasso_formula using the make_formula() function provided in the Appendix.

```
# Extract coefficients at lambda.1se for LASSO model
lasso_coef <- coef(lasso_model, s = "lambda.1se")

# Print coefficient values
print(lasso_coef)

# Identify variables with non-zero coefficients
lasso_vars <- names(lasso_coef)[lasso_coef != 0]

# Remove the response variable "quality"
lasso_vars <- lasso_vars[!lasso_vars %in% "quality"]

# Create the LASSO formula</pre>
```

```
lasso_formula <- as.formula(paste("quality ~", paste(lasso_vars, collapse = " + ")))
# Print LASSO formula
print(lasso_formula)</pre>
```

3.5 (5 points)

Print the coefficient values for ridge regression at the lambda.1se value? What are the variables selected here?

Store the variable names with non-zero coefficients in ridge_vars, and create a formula object called ridge_formula using the make_formula() function provided in the Appendix.

"'R# Extract coefficients at lambda.1se for Ridge model ridge_coef <- coef(ridge_model, s = "lambda.1se")

Print coefficient values

print(ridge_coef)

Identify variables with non-zero coefficients

ridge_vars <- names(ridge_coef)[ridge_coef != 0]

Create formula object using make_formula() function

ridge_formula <- make_formula(ridge_vars)

Print Ridge formula

print(ridge_formula)

```
---
```

3.6 (10 points)

What is the difference between stepwise selection, LASSO and ridge based on your analyses about

4.2 (20 points)

Store the names of the predictor variables (all columns except `quality`) in an object called

```
::: {.cell}
```{.r .cell-code}
x_vars <- colnames(df %>% select(-quality))
:::
```

- Use:
  - the combn() function (built-in R function) and
  - the make\_formula() (provided in the Appendix)

to generate all possible linear regression formulas using the variables in x\_vars. This is most optimally achieved using the map() function from the purr package.

```
formulas <- map(
 1:length(x_vars),
 \(x){
 vars <- combn(x_vars, x)
 map(vars, \(var_set\)) make_formula(var_set))
}
) %>% unlist()
```

If your code is right the following command should return something along the lines of:

```
sample(formulas, 4) %>% as.character()
Output:
[1] "quality ~ volatile_acidity + residual_sugar + density + pH + alcohol"
[2] "quality ~ citric_acid"
[3] "quality ~ volatile_acidity + citric_acid + residual_sugar + total_sulfur_dioxide +
[4] "quality ~ citric_acid + chlorides + total_sulfur_dioxide + pH + alcohol + type"
```

#### 4.3 (10 points)

Use map() and lm() to fit a linear regression model to each formula in formulas, using df as the data source. Use broom::glance() to extract the model summary statistics, and bind them together into a single tibble of summaries using the bind\_rows() function from dplyr.

```
models <- map(formulas, ~lm(., data = df))
summaries <- map(models, glance)</pre>
```

```
summaries <- bind_rows(summaries, .id = "model")
head(summaries)</pre>
```

4.4 (5 points)

Extract the adj.r.squared values from summaries and use them to identify the formula with the *highest* adjusted R-squared value.

```
adj_r_squared <- summaries$adj.r.squared
best_model_index <- which.max(adj_r_squared)</pre>
```

Store resulting formula as a variable called rsq\_formula.

```
rsq_formula <- formulas[[best_model_index]]</pre>
```

4.5 (5 points)

Extract the AIC values from summaries and use them to identify the formula with the *lowest* AIC value.

```
aic_values <- summaries$AIC
best_model_index <- which.min(aic_values)</pre>
```

Store resulting formula as a variable called aic\_formula.

```
aic_formula <- formulas[[best_model_index]]
print(aic_formula)</pre>
```

4.6 (15 points)

Combine all formulas shortlisted into a single vector called final\_formulas.

```
null_formula <- formula(null_model)
full_formula <- formula(full_model)

final_formulas <- c(
 null_formula,
 full_formula,
 backward_formula,
 forward_formula,
 lasso_formula,
 ridge_formula,
 rsq_formula,
 aic_formula
)</pre>
```

- Are aic\_formula and rsq\_formula the same? How do they differ from the formulas shortlisted in question 3?
- Which of these is more reliable? Why?
- If we had a dataset with 10,000 columns, which of these methods would you consider for your analyses? Why?

#### 4.7 (10 points)

Use map() and glance() to extract the sigma, adj.r.squared, AIC, df, and p.value statistics for each model obtained from final\_formulas. Bind them together into a single data frame summary\_table. Summarize your main findings.

```
summary_table <- map(
 final_formulas,
 \((formula)) {
 model <- lm(formula, data = df)
 glance(model)
 }
) %>%
bind_rows()
summary_table %>% knitr::kable()
```

## **Appendix**

#### Convenience function for creating a formula object

The following function which takes as input a vector of column names x and outputs a formula object with quality as the response variable and the columns of x as the covariates.

```
make_formula <- function(x){
 as.formula(
 paste("quality ~ ", paste(x, collapse = " + "))
)
}

For example the following code will
result in a formula object
"quality ~ a + b + c"
make_formula(c("a", "b", "c"))</pre>
```

### Convenience function for glmnet

The make\_model\_matrix function below takes a formula as input and outputs a rescaled model matrix X in a format amenable for glmnet()

```
make_model_matrix <- function(formula){
 X <- model.matrix(formula, df)[, -1]
 cnames <- colnames(X)
 for(i in 1:ncol(X)){
 if(!cnames[i] == "typewhite"){
 X[, i] <- scale(X[, i])
 } else {
 colnames(X)[i] <- "type"
 }
 }
 return(X)
}</pre>
```

# i Session Information

Print your  ${\tt R}$  session information using the following command

sessionInfo()