

# GeneLab URR analysis notebook

This notebook contains analyses of RNA-seq RSEM unnormalized gene counts, and DESeq2 normalized gene counts generated from Universal Reference RNA.

## Table of Content

- [Total Expressed Genes \(Annotated Genes\)](#)
- [Comparative analyses from different NovaSeq runs](#)

## GeneLab RNA-seq pipeline

Explore the [RNA-seq pipeline](#).

## Setting up the notebook

```
In [1]: # Import python packages
import os
import pandas as pd
import numpy as np
import plotly.graph_objects as go
from plotly.subplots import make_subplots
```

## Total Expressed Genes (Annotated Genes)

Unnormalized Counts

```
In [2]: os.chdir("URR_Compare_Analysis")
os.listdir()
```

```
Out[2]: ['.DS_Store',
'LPkit_SampleTable.csv',
'NS_SampleTable.csv',
'NumNonZeroGenes.csv',
'star_alignment.csv',
'URR_Compare_Analysis.html',
'RSEM_Unnormalized_Counts.csv',
'Normalized_Counts.csv',
'.ipynb_checkpoints']
```

```
In [3]: # Get NumNonZeroGenes.csv
totgenes_file = os.listdir()[3]
totgenes_table = pd.read_csv(totgenes_file, index_col=0)
totgenes_table.index.rename("Sample", inplace=True)
pd.set_option("max_columns", None)
totgenes_table.head()
```

Out[3]: **Number of genes with non-zero counts**

Sample	
FS_20190404_HRep1	25217
FS_20190404_HRep2	24548

## Number of genes with non-zero counts

Sample	
FS_20190404_HRep3	24595
FS_20190404_Rep10	28473
FS_20190404_Rep11	25218

In [4]:

```
# Get RSEM_Unnormalized_Counts.csv
unnorm_file = os.listdir()[6]
unnorm_cutoff = pd.read_csv(unnorm_file).rename(columns={"Unnamed: 0": "Genes"})
unnorm_cutoff.head()
```

Out [4]:

	Genes	FS_20190404_HRep1	FS_20190404_HRep2	FS_20190404_HRep3	FS_20190404_Rep10	FS_20190404_Rep11
0	ENSMUSG000000000001	5373.0	4574.0	4647.0	24595	25218
1	ENSMUSG000000000003	0.0	0.0	0.0	28473	25218
2	ENSMUSG000000000028	1755.0	1376.0	1434.0	24595	25218
3	ENSMUSG000000000031	3181.0	2704.0	2581.0	28473	25218
4	ENSMUSG000000000037	68.0	72.0	63.0	24595	25218

In [5]:

```
# Unnormalized counts cutoff > 10
unnorm_cutoff = unnorm_cutoff.set_index(keys="Genes")
unnorm_cutoff = unnorm_cutoff[unnorm_cutoff.index.str.contains("ENSMUSG")]
unnorm_cutoff = unnorm_cutoff[unnorm_cutoff > 10]
unnorm_10 = unnorm_cutoff.count().to_frame(name="Number of genes with more than 10 counts")
unnorm_10.index.rename("Sample", inplace=True)
unnorm_10.head()
```

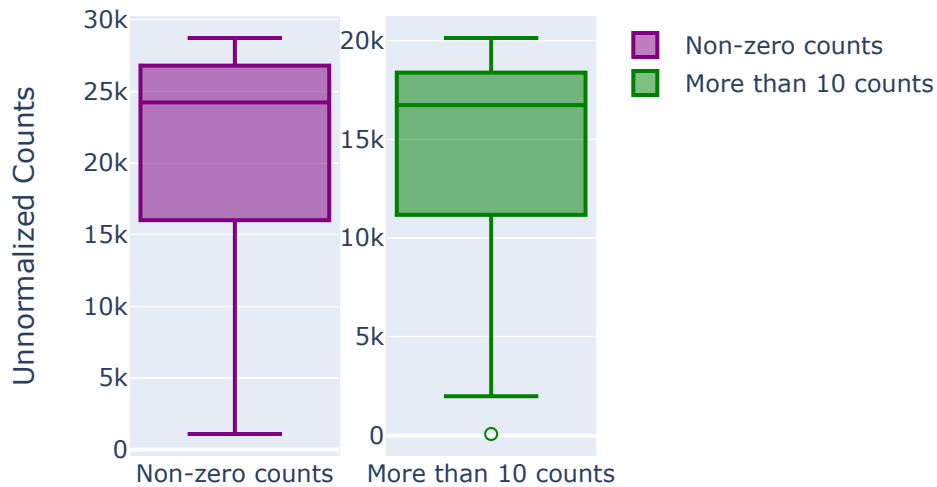
Out [5]:

Sample	Number of genes with more than 10 counts
FS_20190404_HRep1	17074
FS_20190404_HRep2	16730
FS_20190404_HRep3	16669
FS_20190404_Rep10	19635
FS_20190404_Rep11	16985

In [6]:

```
# Box plot of unnormalized counts >0 and >10 for all samples
data = totgenes_table.melt()
data10 = unnorm_10.melt()
fig = go.Figure()
fig = make_subplots(rows=1, cols=2)
fig.add_trace(go.Box(y=data["value"], quartilemethod="inclusive", name="Non-zero counts",
                    row=1, col=1)# "exclusive", "inclusive", or "linear" by default
fig.add_trace(go.Box(y=data10["value"], quartilemethod="inclusive", name="More than 10 counts",
                    row=1, col=2)
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.5)
fig.update_layout(title_text="RSEM Unnormalized Counts", title_x=0.4, yaxis_title="Unnormalized counts",
                    boxmode="overlay", hovermode="x unified", width=500, height=400)
fig.show()
```

## RSEM Unnormalized Counts



## Comparative analyses from different NovaSeq runs

Total Expressed Genes (Annotated Genes)

```
In [7]: # Go to URR_Compare_Analysis
os.listdir()
```

```
Out[7]: ['.DS_Store',
'LPkit_SampleTable.csv',
'NS_SampleTable.csv',
'NumNonZeroGenes.csv',
'star_alignment.csv',
'URR_Compare_Analysis.html',
'RSEM_Unnormalized_Counts.csv',
'Normalized_Counts.csv',
'.ipynb_checkpoints']
```

```
In [8]: # Get NumNonZeroGenes.csv
totgenes_file = os.listdir()[3]
totgenes_table = pd.read_csv(totgenes_file, index_col=0)
totgenes_table.index.rename("Sample", inplace=True)
pd.set_option("max_columns", None)
totgenes_table.head()
```

Out[8]: **Number of genes with non-zero counts**

Sample	
FS_20190404_HRep1	25217
FS_20190404_HRep2	24548
FS_20190404_HRep3	24595
FS_20190404_Rep10	28473
FS_20190404_Rep11	25218

```
In [9]: # Get LPkit_SampleTable.csv
kit_file = os.listdir()[1]
kit_table = pd.read_csv(kit_file).rename(columns={"Unnamed: 0": "Sample", "condition": "Library kit"})
kit_table = kit_table.sort_values(by="Sample", ascending=True)
kit_table.head()
```

```
Out[9]:
```

	Sample	Library kit
0	FS_20190404_HRep1	Illumina_TrueSeq_Stranded_totRNA_Gold
1	FS_20190404_HRep2	Illumina_TrueSeq_Stranded_totRNA_Gold
2	FS_20190404_HRep3	Illumina_TrueSeq_Stranded_totRNA_Gold
3	FS_20190404_Rep1	Illumina_TrueSeq_Stranded_totRNA_Gold
4	FS_20190404_Rep10	Illumina_TrueSeq_Stranded_totRNA_Gold

```
In [10]: # Get NS_SampleTable.csv
ns_file = os.listdir()[2]
ns_table = pd.read_csv(ns_file).rename(columns={"Unnamed: 0": "Sample", "condition": "NovaSeq"})
ns_table.head()
```

```
Out[10]:
```

	Sample	NovaSeqRun
0	FS_20190404_HRep1	FS
1	FS_20190404_HRep2	FS
2	FS_20190404_HRep3	FS
3	FS_20190404_Rep1	FS
4	FS_20190404_Rep2	FS

```
In [11]: # Get Uniquely mapped reads from star_alignment.csv
aligned_file = os.listdir()[4]
aligned_table = pd.read_csv(aligned_file).rename(columns={"Category": "Sample"})
depth_table = aligned_table.filter(["Sample", "Uniquely mapped"]).rename(columns={"Uniquely mapped": "Uniquely mapped reads"})
depth_table["Uniquely mapped reads"] = depth_table["Uniquely mapped reads"] / 1000000
depth_table = depth_table.rename(columns={"Uniquely mapped reads": "Uniquely mapped reads (M)"})
depth_table.head()
```

```
Out[11]:
```

	Sample	Uniquely mapped reads (M)
0	FS_20190404_HRep1	40.272953
1	FS_20190404_HRep2	34.034887
2	FS_20190404_HRep3	34.851778
3	FS_20190404_Rep1	110.888770
4	FS_20190404_Rep10	115.690366

```
In [12]: # Non-zero unnormalized counts of different library kits used
kit_summary = kit_table.merge(totgenes_table, on="Sample")
kit_summary = kit_summary.set_index(keys="Sample")
kit_summary.head()
```

```
Out[12]:
```

	Library kit	Number of genes with non-zero counts
--	-------------	--------------------------------------

	Sample	Library kit	Number of genes with non-zero counts
	Sample		
	FS_20190404_HRep1	Illumina_TrueSeq_Stranded_totRNA_Gold	25217
	FS_20190404_HRep2	Illumina_TrueSeq_Stranded_totRNA_Gold	24548
	FS_20190404_HRep3	Illumina_TrueSeq_Stranded_totRNA_Gold	24595
	FS_20190404_Rep1	Illumina_TrueSeq_Stranded_totRNA_Gold	28129
	FS_20190404_Rep10	Illumina_TrueSeq_Stranded_totRNA_Gold	28473

In [13]:

```
# Non-zero unnormalized counts of different NovaSeq runs
ns_summary = ns_table.merge(kit_summary, on="Sample")
ns_summary
```

Out[13]:

	Sample	NovaSeqRun	Library kit	Number of genes with non-zero counts
0	FS_20190404_HRep1	FS	Illumina_TrueSeq_Stranded_totRNA_Gold	25217
1	FS_20190404_HRep2	FS	Illumina_TrueSeq_Stranded_totRNA_Gold	24548
2	FS_20190404_HRep3	FS	Illumina_TrueSeq_Stranded_totRNA_Gold	24595
3	FS_20190404_Rep1	FS	Illumina_TrueSeq_Stranded_totRNA_Gold	28129
4	FS_20190404_Rep2	FS	Illumina_TrueSeq_Stranded_totRNA_Gold	26941
...	...	...	...	...
149	RR10_KDN_UPX_20220104_7	RR10_KDN_UPX	QIAseq_UPX_mRNA	14842
150	RR10_KDN_UPX_20220104_8	RR10_KDN_UPX	QIAseq_UPX_mRNA	14774
151	RR23_LVR_LNG_20220112_2_Xp	RR23_LVR_LNG	Illumina_TrueSeq_Stranded_totRNA_Gold	28463
152	RR23_LVR_LNG_20220112_3_Xp	RR23_LVR_LNG	Illumina_TrueSeq_Stranded_totRNA_Gold	26158
153	PI_HRT_20220112_Xp	PI_HRT	Illumina_TrueSeq_Stranded_totRNA_Gold	24734

154 rows x 4 columns

In [14]:

```
# Non-zero unnormalized counts of different NovaSeq runs
depth_summary = ns_summary.merge(depth_table, on="Sample")
depth_summary = depth_summary.filter(["Sample", "NovaSeqRun", "Number of genes with non-zero counts"])
depth_summary.head()
```

Out[14]:

	Sample	NovaSeqRun	Number of genes with non-zero counts	Uniquely mapped reads (M)
0	FS_20190404_HRep1	FS	25217	40.272953
1	FS_20190404_HRep2	FS	24548	34.034887
2	FS_20190404_HRep3	FS	24595	34.851778
3	FS_20190404_Rep1	FS	28129	110.888770
4	FS_20190404_Rep2	FS	26941	72.993442

In [15]:

```
# Group by NS run
```

```
ind_run = dict(list(depth_summary.groupby("NovaSeqRun")))
ind_run
```

```
Out[15]: {'FS':
          Sample NovaSeqRun  Number of genes with non-zero counts \
0    FS_20190404_HRep1      FS                                25217
1    FS_20190404_HRep2      FS                                24548
2    FS_20190404_HRep3      FS                                24595
3    FS_20190404_Rep1       FS                                28129
4    FS_20190404_Rep2       FS                                26941
5    FS_20190404_Rep3       FS                                27068
6    FS_20190404_Rep4       FS                                27455
7    FS_20190404_Rep5       FS                                26860
8    FS_20190404_Rep6       FS                                27144
9    FS_20190404_Rep7       FS                                28065
10   FS_20190404_Rep8       FS                                28421
11   FS_20190404_Rep9       FS                                27759
12   FS_20190404_Rep10      FS                                28473
13   FS_20190404_Rep11      FS                                25218
14   FS_20190404_Rep12      FS                                25485
15   FS_20190404_Rep13      FS                                27567
16   FS_20190404_Rep14      FS                                28147
17   FS_20190404_Rep15      FS                                28424
18   FS_20190404_Rep16      FS                                26099
19   FS_20190404_Rep17      FS                                23202
20   FS_20190404_Rep18      FS                                26323
21   FS_20190404_Rep19      FS                                27128
22   FS_20190404_Rep20      FS                                26497

          Uniquely mapped reads (M)
0          40.272953
1          34.034887
2          34.851778
3         110.888770
4          72.993442
5          76.767071
6          89.686997
7          79.059419
8          84.585888
9          88.462019
10         95.498190
11         87.566747
12        115.690366
13         45.072266
14         47.183193
15         76.889819
16        141.445416
17        121.429975
18         67.321401
19         95.642894
20         78.873987
21         84.886688
22        120.532743 ,

'PI_FF_20211026':
Sample      NovaSeqRun  Number of genes with non-zero co
unts \
138  PI_FF_20211026_1  PI_FF_20211026                24574
139  PI_FF_20211026_2  PI_FF_20211026                24160
140  PI_FF_20211026_3  PI_FF_20211026                24188

          Uniquely mapped reads (M)
138          37.173529
139          31.926943
140          33.138743 ,

'PI_FF_20211124':
Sample      NovaSeqRun  Number of genes with non-zero co
unts \
141  PI_FF_20211124_1  PI_FF_20211124                21979
```

Uniquely mapped reads (M)

141	23.961512
142	26.511764 ,

'PI\_GS':

	Sample	NovaSeqRun	Number of genes with non-zero counts \
23	PI_GS_20190422_Rep8	PI_GS	28545
24	PI_GS_20190422_Rep9	PI_GS	27504
25	PI_GS_20190422_Rep10	PI_GS	27400

Uniquely mapped reads (M)

23	112.353609
24	89.063295
25	87.982191 ,

'PI\_HRT':

	Sample	NovaSeqRun	Number of genes with non-zero counts \
153	PI_HRT_20220112_Xp	PI_HRT	24734

Uniquely mapped reads (M)

153	65.852981 ,
-----	-------------

'PI\_Rad\_HPC':

	Sample	NovaSeqRun \
97	PI_Rad_HPC_20210225_Rep1	PI_Rad_HPC
98	PI_Rad_HPC_20210225_Rep2	PI_Rad_HPC

	Number of genes with non-zero counts	Uniquely mapped reads (M)
97	27226	80.995246
98	26550	71.985778 ,

'RF\_LVR\_SLS':

	Sample	NovaSeqRun \
60	RF_LVR_SLS_20200528_Rep3	RF_LVR_SLS
61	RF_LVR_SLS_20200528_Rep4	RF_LVR_SLS
62	RF_LVR_SLS_20200528_Rep5	RF_LVR_SLS
63	RF_LVR_SLS_20200528_Rep6	RF_LVR_SLS
64	RF_LVR_SLS_20200528_Rep7	RF_LVR_SLS
65	RF_LVR_SLS_20200528_Rep8	RF_LVR_SLS
66	RF_LVR_SLS_20200528_Rep9	RF_LVR_SLS

	Number of genes with non-zero counts	Uniquely mapped reads (M)
60	27863	112.960576
61	26606	76.112090
62	27355	73.127886
63	27368	85.572623
64	28308	77.245352
65	27865	70.986349
66	27706	73.385536 ,

'RF\_SPL':

	Sample	NovaSeqRun	Number of genes with non-zero counts \
57	RF_SPL_20200213_Rep1	RF_SPL	25292
58	RF_SPL_20200213_Rep2	RF_SPL	27234
59	RF_SPL_20200213_Rep6	RF_SPL	26231

Uniquely mapped reads (M)

57	60.263281
58	60.070304
59	59.919477 ,

'RR10\_KDN\_UPX':

	Sample	NovaSeqRun \
143	RR10_KDN_UPX_20220104_1	RR10_KDN_UPX
144	RR10_KDN_UPX_20220104_2	RR10_KDN_UPX
145	RR10_KDN_UPX_20220104_3	RR10_KDN_UPX
146	RR10_KDN_UPX_20220104_4	RR10_KDN_UPX
147	RR10_KDN_UPX_20220104_5	RR10_KDN_UPX
148	RR10_KDN_UPX_20220104_6	RR10_KDN_UPX
149	RR10_KDN_UPX_20220104_7	RR10_KDN_UPX
150	RR10_KDN_UPX_20220104_8	RR10_KDN_UPX

	Number of genes with non-zero counts	Uniquely mapped reads (M)
143	16439	14.041522
144	13554	6.058263
145	16012	21.257030

146		14724	7.484766
147		14189	12.430669
148		14143	6.223526
149		14842	9.257538
150		14774	8.399899
'RR23_LVR_LNG': Sample NovaSeqRun \			
151	RR23_LVR_LNG_20220112_2_Xp	RR23_LVR_LNG	
152	RR23_LVR_LNG_20220112_3_Xp	RR23_LVR_LNG	
Number of genes with non-zero counts Uniquely mapped reads (M)			
151		28463	171.437695
152		26158	74.171619
'RR6_CLN_LNG': Sample NovaSeqRun \			
38	RR6_CLN_LNG_20190718_Rep1	RR6_CLN_LNG	
39	RR6_CLN_LNG_20190718_Rep2	RR6_CLN_LNG	
40	RR6_CLN_LNG_20190718_Rep3	RR6_CLN_LNG	
Number of genes with non-zero counts Uniquely mapped reads (M)			
38		27484	82.853300
39		27002	68.550399
40		26636	73.374007
'RR6_LVR_SPL': Sample NovaSeqRun \			
41	RR6_LVR_SPL_20190805_Rep3	RR6_LVR_SPL	
42	RR6_LVR_SPL_20190805_Rep4	RR6_LVR_SPL	
43	RR6_LVR_SPL_20190805_Rep9	RR6_LVR_SPL	
44	RR6_LVR_SPL_20190805_Rep10	RR6_LVR_SPL	
45	RR6_LVR_SPL_20190805_Rep13	RR6_LVR_SPL	
46	RR6_LVR_SPL_20190805_Rep14	RR6_LVR_SPL	
Number of genes with non-zero counts Uniquely mapped reads (M)			
41		28731	95.641397
42		28133	104.343357
43		27875	93.640154
44		27517	90.053019
45		27392	85.881207
46		27002	86.680351
'RR6_TMS_DSKN': Sample NovaSeqRun \			
35	RR6_TMS_DSKN_20190628_Rep1	RR6_TMS_DSKN	
36	RR6_TMS_DSKN_20190628_Rep2	RR6_TMS_DSKN	
37	RR6_TMS_DSKN_20190628_Rep3	RR6_TMS_DSKN	
Number of genes with non-zero counts Uniquely mapped reads (M)			
35		27178	87.336601
36		26803	79.650733
37		27614	93.389118
'RR7_KDN_SKN': Sample NovaSeqRun \			
47	RR7_KDN_SKN_20190909_Rep1	RR7_KDN_SKN	
48	RR7_KDN_SKN_20190909_Rep2	RR7_KDN_SKN	
49	RR7_KDN_SKN_20190909_Rep3	RR7_KDN_SKN	
50	RR7_KDN_SKN_20190909_Rep4	RR7_KDN_SKN	
51	RR7_KDN_SKN_20190909_Rep5	RR7_KDN_SKN	
52	RR7_KDN_SKN_20190909_Rep6	RR7_KDN_SKN	
53	RR7_KDN_SKN_20190909_Rep7	RR7_KDN_SKN	
54	RR7_KDN_SKN_20190909_Rep8	RR7_KDN_SKN	
55	RR7_KDN_SKN_20190909_Rep9	RR7_KDN_SKN	
56	RR7_KDN_SKN_20190909_Rep10	RR7_KDN_SKN	
Number of genes with non-zero counts Uniquely mapped reads (M)			
47		26669	58.771134
48		26778	58.922385
49		26928	65.582509
50		24600	61.024954
51		25151	59.019873
52		25111	55.183371
53		24950	61.833689
54		24961	58.939524

55		25282	59.381925
56		7785	3.839760 ,
	'RR9_RR5_MHU2':	Sample	NovaSeqRun \
26	RR9_RR5_MHU2_20190522_Rep1	RR9_RR5_MHU2	
27	RR9_RR5_MHU2_20190522_Rep2	RR9_RR5_MHU2	
28	RR9_RR5_MHU2_20190522_Rep3	RR9_RR5_MHU2	
29	RR9_RR5_MHU2_20190522_Rep4	RR9_RR5_MHU2	
30	RR9_RR5_MHU2_20190522_Rep5	RR9_RR5_MHU2	
31	RR9_RR5_MHU2_20190522_Rep6	RR9_RR5_MHU2	
32	RR9_RR5_MHU2_20190522_Rep7	RR9_RR5_MHU2	
33	RR9_RR5_MHU2_20190522_Rep8	RR9_RR5_MHU2	
34	RR9_RR5_MHU2_20190522_Rep9	RR9_RR5_MHU2	
	Number of genes with non-zero counts	Uniquely mapped reads (M)	
26		26331	54.975307
27		25778	52.599008
28		25680	60.889496
29		25965	55.978324
30		26014	58.339829
31		25100	55.400545
32		25742	52.250636
33		27184	55.831453
34		26164	49.281373 ,
	'RRRM1_LVR':	Sample	NovaSeqRun \
99	RRRM1_LVR_RR6_20210318_Rep1	RRRM1_LVR	
100	RRRM1_LVR_RR6_20210318_Rep2	RRRM1_LVR	
101	RRRM1_LVR_20210318_Rep1	RRRM1_LVR	
102	RRRM1_LVR_20210318_Rep2	RRRM1_LVR	
103	RRRM1_LVR_20210318_Rep3	RRRM1_LVR	
104	RRRM1_LVR_20210318_Rep4	RRRM1_LVR	
105	RRRM1_LVR_20210318_Rep5	RRRM1_LVR	
106	RRRM1_LVR_20210318_Rep6	RRRM1_LVR	
107	RRRM1_LVR_20210318_Rep7	RRRM1_LVR	
	Number of genes with non-zero counts	Uniquely mapped reads (M)	
99		24254	47.040706
100		23912	48.326822
101		24672	48.409954
102		25135	38.875188
103		24071	44.860812
104		25177	51.146926
105		24190	43.066764
106		24209	41.793269
107		25073	49.907092 ,
	'RiboTest_RiboZeroGold':	Sample	Nova
	SeqRun \		
112	RiboTest_RiboZeroGold_20210402_100ng_RNA_1	RiboTest_RiboZeroGold	
113	RiboTest_RiboZeroGold_20210402_100ng_RNA_2	RiboTest_RiboZeroGold	
114	RiboTest_RiboZeroGold_20210402_500ng_RNA_1	RiboTest_RiboZeroGold	
115	RiboTest_RiboZeroGold_20210402_500ng_RNA_2	RiboTest_RiboZeroGold	
	Number of genes with non-zero counts	Uniquely mapped reads (M)	
112		24214	49.649754
113		24372	47.243508
114		23027	43.804980
115		23320	48.423480 ,
	'RiboTest_RiboZeroPlus':	Sample	Nova
	SeqRun \		
108	RiboTest_RiboZeroPlus_20210402_100ng_RNA_1	RiboTest_RiboZeroPlus	
109	RiboTest_RiboZeroPlus_20210402_100ng_RNA_2	RiboTest_RiboZeroPlus	
110	RiboTest_RiboZeroPlus_20210402_500ng_RNA_1	RiboTest_RiboZeroPlus	
111	RiboTest_RiboZeroPlus_20210402_500ng_RNA_2	RiboTest_RiboZeroPlus	
	Number of genes with non-zero counts	Uniquely mapped reads (M)	
108		16832	19.791358
109		18834	32.755994

110		22855	57.839715
111		22741	54.113288 ,
	'UPX_ALSDA100':	Sample	NovaSeqRun \
134	UPX_ALSDA100_20210806_TecRep1	UPX_ALSDA100	
135	UPX_ALSDA100_20210806_TecRep2	UPX_ALSDA100	
136	UPX_ALSDA100_20210806_TecRep3	UPX_ALSDA100	
137	UPX_ALSDA100_20210806_TecRep4	UPX_ALSDA100	

	Number of genes with non-zero counts	Uniquely mapped reads (M)
134	23799	97.064195
135	21729	76.338304
136	21411	71.400526
137	21744	75.452050 ,

	'UPX_test':	Sample	NovaSeqRun \
67	UPX_test_10_ng_ERCC_20201020_tRep1	UPX_test	
68	UPX_test_10_ng_ERCC_20201020_tRep2	UPX_test	
69	UPX_test_10_ng_ERCC_20201020_tRep3	UPX_test	
70	UPX_test_10_ng_no_ERCC_20201020_tRep1	UPX_test	
71	UPX_test_10_ng_no_ERCC_20201020_tRep2	UPX_test	
72	UPX_test_10_ng_no_ERCC_20201020_tRep3	UPX_test	
73	UPX_test_1_ng_ERCC_20201020_tRep1	UPX_test	
74	UPX_test_1_ng_ERCC_20201020_tRep2	UPX_test	
75	UPX_test_1_ng_ERCC_20201020_tRep3	UPX_test	
76	UPX_test_1_ng_no_ERCC_20201020_tRep1	UPX_test	
77	UPX_test_1_ng_no_ERCC_20201020_tRep2	UPX_test	
78	UPX_test_1_ng_no_ERCC_20201020_tRep3	UPX_test	
79	UPX_test_20_ng_ERCC_20201020_tRep1	UPX_test	
80	UPX_test_20_ng_ERCC_20201020_tRep2	UPX_test	
81	UPX_test_20_ng_ERCC_20201020_tRep3	UPX_test	
82	UPX_test_20_ng_no_ERCC_20201020_tRep1	UPX_test	
83	UPX_test_20_ng_no_ERCC_20201020_tRep2	UPX_test	
84	UPX_test_20_ng_no_ERCC_20201020_tRep3	UPX_test	
85	UPX_test_500_pg_ERCC_20201020_tRep1	UPX_test	
86	UPX_test_500_pg_ERCC_20201020_tRep2	UPX_test	
87	UPX_test_500_pg_ERCC_20201020_tRep3	UPX_test	
88	UPX_test_500_pg_no_ERCC_20201020_tRep1	UPX_test	
89	UPX_test_500_pg_no_ERCC_20201020_tRep2	UPX_test	
90	UPX_test_500_pg_no_ERCC_20201020_tRep3	UPX_test	
91	UPX_test_50_pg_ERCC_20201020_tRep1	UPX_test	
92	UPX_test_50_pg_ERCC_20201020_tRep2	UPX_test	
93	UPX_test_50_pg_ERCC_20201020_tRep3	UPX_test	
94	UPX_test_50_pg_no_ERCC_20201020_tRep1	UPX_test	
95	UPX_test_50_pg_no_ERCC_20201020_tRep2	UPX_test	
96	UPX_test_50_pg_no_ERCC_20201020_tRep3	UPX_test	

	Number of genes with non-zero counts	Uniquely mapped reads (M)
67	17111	3.894863
68	17089	3.943846
69	17100	3.808051
70	17421	5.345093
71	1089	0.025915
72	17377	5.002224
73	12244	3.329272
74	12829	4.375843
75	13711	5.861074
76	12188	4.492884
77	12705	4.673743
78	12356	4.122470
79	17230	3.217031
80	18107	4.148696
81	18361	4.340296
82	17631	3.599179
83	18233	4.156169
84	17363	3.116021
85	11000	4.803449
86	10053	3.897370

87	10174	3.945274
88	10097	3.183167
89	11091	4.639503
90	10954	4.200025
91	5256	3.746737
92	5181	3.883702
93	5990	5.213186
94	4664	4.175198
95	4378	3.799869
96	3924	3.199142

	Sample	NovaSeqRun
'UPX_test2_L001':		
116	UPX_test2_L001_S1_Qiagen_008X_20210415_1_10ng	UPX_test2_L001
117	UPX_test2_L001_S1_Qiagen_008X_20210415_2_10ng	UPX_test2_L001
118	UPX_test2_L001_S1_Qiagen_008X_20210415_3_10ng	UPX_test2_L001
119	UPX_test2_L001_S3_Illumina_RZP_20210415_1_10ng	UPX_test2_L001
120	UPX_test2_L001_S3_Illumina_RZP_20210415_2_10ng	UPX_test2_L001
121	UPX_test2_L001_S3_Illumina_RZP_20210415_3_10ng	UPX_test2_L001
122	UPX_test2_L001_S5_No_Ribo_Dep_20210415_1_10ng	UPX_test2_L001
123	UPX_test2_L001_S5_No_Ribo_Dep_20210415_2_10ng	UPX_test2_L001
124	UPX_test2_L001_S5_No_Ribo_Dep_20210415_3_10ng	UPX_test2_L001

	Number of genes with non-zero counts	Uniquely mapped reads (M)
116	16682	14.087898
117	16385	14.548883
118	16996	17.584845
119	7489	13.358559
120	9188	29.790044
121	8203	15.754161
122	16654	15.155427
123	17427	17.572372
124	16653	13.089206

	Sample	NovaSeqRun
'UPX_test2_L002':		
125	UPX_test2_L002_S1_Qiagen_05X_20210415_1_10ng	UPX_test2_L002
126	UPX_test2_L002_S1_Qiagen_05X_20210415_2_10ng	UPX_test2_L002
127	UPX_test2_L002_S1_Qiagen_05X_20210415_3_10ng	UPX_test2_L002
128	UPX_test2_L002_S2_Qiagen_1X_20210415_1_10ng	UPX_test2_L002
129	UPX_test2_L002_S2_Qiagen_1X_20210415_2_10ng	UPX_test2_L002
130	UPX_test2_L002_S2_Qiagen_1X_20210415_3_10ng	UPX_test2_L002
131	UPX_test2_L002_S3_No_Ribo_Dep_20210415_1_10ng	UPX_test2_L002
132	UPX_test2_L002_S3_No_Ribo_Dep_20210415_2_10ng	UPX_test2_L002
133	UPX_test2_L002_S3_No_Ribo_Dep_20210415_3_10ng	UPX_test2_L002

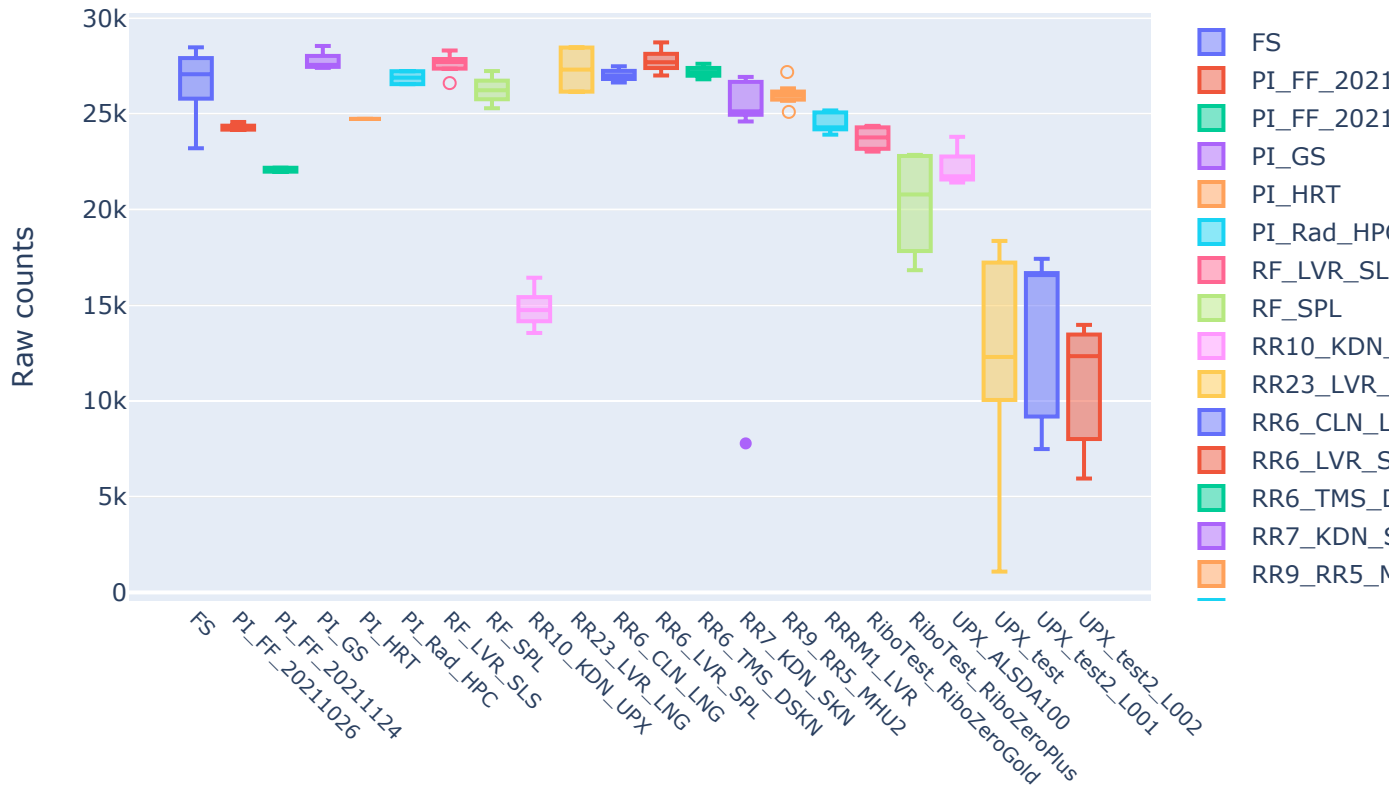
  

	Number of genes with non-zero counts	Uniquely mapped reads (M)
125	12342	6.287190
126	13472	9.021345
127	13976	11.122008
128	8013	15.886517
129	6126	9.574508
130	5950	6.696182
131	13410	4.821391
132	8655	1.303264
133	13629	4.788189

In [16]:

```
# Box plot grouped by uniquely mapped reads for each NS run on same graph
fig = go.Figure()
fig = make_subplots(rows=1, cols=1)
for i in ind_run:
    fig.add_trace(go.Box(y=ind_run[i]["Number of genes with non-zero counts"], quartilemet
        name=i), row=1, col=1)
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.75)
fig.update_layout(title_text="Raw Counts by NovaSeq Run", title_x=0.4, yaxis_title="Raw co
    boxmode="group", hovermode="x unified", width=800, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()
```

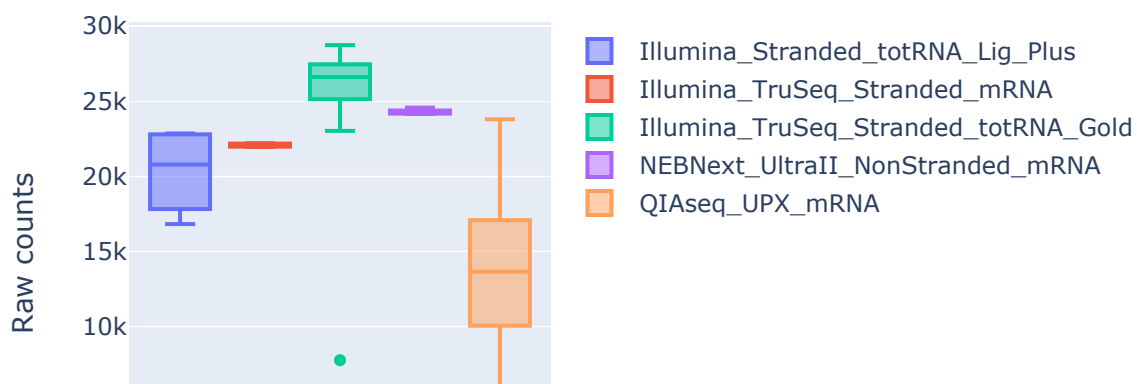
## Raw Counts by NovaSeq Run

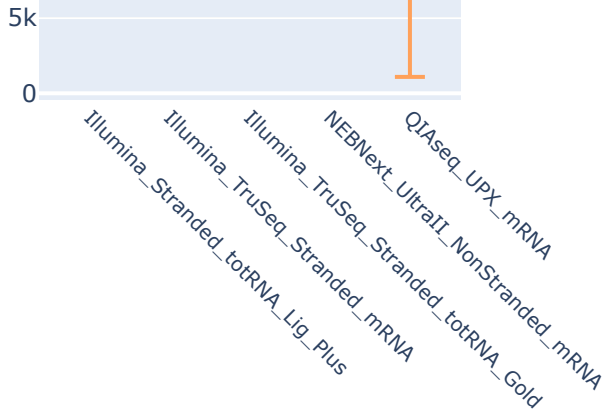


```
In [17]: # Group by library kits
kit_run = dict(list(ns_summary.groupby("Library kit")))
```

```
In [18]: # Box plot grouped by library preparation for each NS run on same graph
fig = go.Figure()
for i in kit_run:
    fig.add_trace(go.Box(y=kit_run[i]["Number of genes with non-zero counts"], quartilemet
        name=i))
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.75)
fig.update_layout(title_text="Raw Counts by Library Kit", title_x=0.12, yaxis_title="Raw c
    boxmode="group", hovermode="x unified", width=600, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()
```

## Raw Counts by Library Kit





In [19]:

```
# Box plot grouped by uniquely mapped reads for samples from all NS runs on same graph
fig = go.Figure()
fig.add_trace(go.Box(x=depth_summary["Uniquely mapped reads (M)"], y=depth_summary["Number of uniquely mapped reads"],
                    name="Non-zero counts", marker_color="purple"))
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.3, textsrc="inside", width=1)
fig.update_layout(title_text="Raw Counts by Aligned Uniquely Mapped Reads", title_x=0.5, title_y=0.5,
                  boxmode="group", hovermode="x unified", width=600, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()
```

## DESeq2 Normalized Data

In [ ]:

```
# Change to R kernel
getwd()
```

```
In [1]: # Go to Normalized_Counts.csv and SampleTable.csv directory
work_dir="URR_Compare_Analysis"
setwd(file.path(work_dir))
```

```
In [2]: ## Install and load ggfortify and ggplot if not already installed
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")

BiocManager::install("tidyverse")
BiocManager::install("ggfortify")
BiocManager::install("plotly")
library(tidyverse)
library(ggfortify)
library(plotly)
```

'getOption("repos")' replaces Bioconductor standard repositories, see  
'?repositories' for details

replacement repositories:  
CRAN: <https://cran.r-project.org>

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:

"package(s) not installed when version(s) same as current; use `force = TRUE` to  
re-install: 'tidyverse'"

Old packages: 'BiocManager', 'blob', 'brew', 'callr', 'class', 'cli', 'clipr',  
'cluster', 'colorspace', 'commonmark', 'cpp11', 'crayon', 'curl',  
'data.table', 'DBI', 'desc', 'devtools', 'digest', 'evaluate', 'fansi',  
'farver', 'foreign', 'formatR', 'GenomeInfoDb', 'gert', 'gh', 'gitcreds',  
'glue', 'gtable', 'htmltools', 'httr', 'IRkernel', 'isoband', 'jsonlite',  
'knitr', 'locfit', 'magrittr', 'markdown', 'MASS', 'Matrix', 'matrixStats',  
'mgcv', 'nlme', 'nnet', 'openssl', 'packrat', 'pbdZMQ', 'pillar', 'pkgload',  
'processx', 'ps', 'purrr', 'RColorBrewer', 'Rcpp', 'RcppArmadillo', 'RCurl',  
'readr', 'rmarkdown', 'roxygen2', 'rpart', 'rprojroot', 'rsconnect',  
'RSQLite', 'rstudioapi', 'rversions', 'S4Vectors', 'spatial', 'stringi',  
'stringr', 'survival', 'sys', 'testthat', 'tibble', 'tidyselect', 'tinytex',  
'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',  
'zip'

'getOption("repos")' replaces Bioconductor standard repositories, see  
'?repositories' for details

replacement repositories:  
CRAN: <https://cran.r-project.org>

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:

"package(s) not installed when version(s) same as current; use `force = TRUE` to  
re-install: 'ggfortify'"

Old packages: 'BiocManager', 'blob', 'brew', 'callr', 'class', 'cli', 'clipr',  
'cluster', 'colorspace', 'commonmark', 'cpp11', 'crayon', 'curl',  
'data.table', 'DBI', 'desc', 'devtools', 'digest', 'evaluate', 'fansi',  
'farver', 'foreign', 'formatR', 'GenomeInfoDb', 'gert', 'gh', 'gitcreds',  
'glue', 'gtable', 'htmltools', 'httr', 'IRkernel', 'isoband', 'jsonlite',  
'knitr', 'locfit', 'magrittr', 'markdown', 'MASS', 'Matrix', 'matrixStats',  
'mgcv', 'nlme', 'nnet', 'openssl', 'packrat', 'pbdZMQ', 'pillar', 'pkgload',  
'processx', 'ps', 'purrr', 'RColorBrewer', 'Rcpp', 'RcppArmadillo', 'RCurl',  
'readr', 'rmarkdown', 'roxygen2', 'rpart', 'rprojroot', 'rsconnect',  
'RSQLite', 'rstudioapi', 'rversions', 'S4Vectors', 'spatial', 'stringi',  
'stringr', 'survival', 'sys', 'testthat', 'tibble', 'tidyselect', 'tinytex',

```

'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',
'zip'

'getOption("repos")' replaces Bioconductor standard repositories, see
'?repositories' for details

replacement repositories:
  CRAN: https://cran.r-project.org

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:
"package(s) not installed when version(s) same as current; use `force = TRUE` to
  re-install: 'plotly'"
Old packages: 'BiocManager', 'blob', 'brew', 'callr', 'class', 'cli', 'clipr',
  'cluster', 'colorspace', 'commonmark', 'cpp11', 'crayon', 'curl',
  'data.table', 'DBI', 'desc', 'devtools', 'digest', 'evaluate', 'fansi',
  'farver', 'foreign', 'formatR', 'GenomeInfoDb', 'gert', 'gh', 'gitcreds',
  'glue', 'gtable', 'htmltools', 'httr', 'IRkernel', 'isoband', 'jsonlite',
  'knitr', 'locfit', 'magrittr', 'markdown', 'MASS', 'Matrix', 'matrixStats',
  'mgcv', 'nlme', 'nnet', 'openssl', 'packrat', 'pbdZMQ', 'pillar', 'pkgload',
  'processx', 'ps', 'purrr', 'RColorBrewer', 'Rcpp', 'RcppArmadillo', 'RCurl',
  'readr', 'rmarkdown', 'roxygen2', 'rpart', 'rprojroot', 'rsconnect',
  'RSQLite', 'rstudioapi', 'rversions', 'S4Vectors', 'spatial', 'stringi',
  'stringr', 'survival', 'sys', 'testthat', 'tibble', 'tidyselect', 'tinytex',
  'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',
  'zip'

```

```

— Attaching packages — tidyverse 1.3.2 —
✓ ggplot2 3.4.0      ✓ purrr 0.3.4
✓ tibble 3.1.6       ✓ dplyr 1.0.10
✓ tidyr 1.2.1        ✓ stringr 1.4.0
✓ readr 2.1.2        ✓ forcats 0.5.2

— Conflicts — tidyverse_conflicts() —
✖ dplyr::filter() masks stats::filter()
✖ dplyr::lag() masks stats::lag()

```

Attaching package: 'plotly'

The following object is masked from 'package:ggplot2':

last\_plot

The following object is masked from 'package:stats':

filter

The following object is masked from 'package:graphics':

layout

In [3]:

```

# Import table with samples and respective groups and column numbers
samp_group <- read.csv(Sys.glob("LPkit_SampleTable.csv"), header = TRUE, row.names = 1, st
samp_group1 <- read.csv(Sys.glob("NS_SampleTable.csv"), header = TRUE, row.names = 1, stri
colnames(samp_group)[1] <- "LibraryKit"
colnames(samp_group1)[1] <- "NovaSeqRun"
samp_group2 <- merge(samp_group, samp_group1, by.x=0, by.y=0)

```

In [4]: `head(samp_group1)`

A data.frame: 6 × 1

NovaSeqRun	
	<fct>
FS_20190404_HRep1	FS
FS_20190404_HRep2	FS
FS_20190404_HRep3	FS
FS_20190404_Rep1	FS
FS_20190404_Rep2	FS
FS_20190404_Rep3	FS

In [5]: `head(samp_group2)`

A data.frame: 6 × 3

Row.names		LibraryKit	NovaSeqRun
	<l<chr>>	<fct>	<fct>
1	FS_20190404_HRep1	Illumina_TrueSeq_Stranded_totRNA_Gold	FS
2	FS_20190404_HRep2	Illumina_TrueSeq_Stranded_totRNA_Gold	FS
3	FS_20190404_HRep3	Illumina_TrueSeq_Stranded_totRNA_Gold	FS
4	FS_20190404_Rep1	Illumina_TrueSeq_Stranded_totRNA_Gold	FS
5	FS_20190404_Rep10	Illumina_TrueSeq_Stranded_totRNA_Gold	FS
6	FS_20190404_Rep11	Illumina_TrueSeq_Stranded_totRNA_Gold	FS

In [6]: 

```
# Import normalized counts table
normCounts <- read.csv(Sys.glob("Normalized_Counts.csv"), header = TRUE, row.names = 1, st
normCounts <- normCounts +1
```

In [7]: 

```
# PCA plot, all samples grouped by NS run
## Indicate PCA plot size
size_var <- 3
alpha_var <- 0.3
exp_raw <- log2(normCounts)
PCA_raw <- prcomp(t(exp_raw), scale = FALSE)

NS <- autoplot(PCA_raw, data = samp_group2, colour = 'NovaSeqRun', shape = 'NovaSeqRun', s
  theme_bw() + theme(axis.text.x = element_text(size=12), axis.text.y = element_text(si
  theme(legend.title = element_text(size=8), legend.text = element_text(size=6)) +
  guides(alpha = guide_legend(order = 1), size = guide_legend(order = 2)) +
  ggtitle("Grouped by NovaSeq Run") + theme(plot.title = element_text(hjust = 0.5)) +
  scale_shape_manual(values = rep(17:22, len = 22))
ggplotly(NS, tooltip = c("text", "size"), width = 700, height = 750)
```

In [8]:

```
# PCA plot, all samples grouped by LP kit
## Indicate PCA plot size
size_var <- 3
alpha_var <- 0.3
exp_raw <- log2(normCounts)
PCA_raw <- prcomp(t(exp_raw), scale = FALSE)

kit <- autoplot(PCA_raw, data = samp_group2, colour = 'LibraryKit', shape = 'LibraryKit',
  theme_bw() + theme(axis.text.x = element_text(size=12), axis.text.y = element_text(s
  theme(legend.title = element_text(size=8), legend.text = element_text(size=8)) +
  guides(alpha = guide_legend(order = 1), size = guide_legend(order = 2)) +
  ggtitle("Grouped by Library Kit") + theme(plot.title = element_text(hjust = 0.5)) +
  scale_shape_manual(values = rep(17:22, len = 22))
ggplotly(kit, tooltip = c("text", "size"), width = 700, height = 400)
```

