

GeneLab URR analysis notebook

This notebook contains analyses of RNA-seq RSEM unnormalized gene counts, and DESeq2 normalized gene counts generated from Universal Reference RNA.

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- Comparative analyses from different NovaSeq runs

GeneLab RNA-seq pipeline

[Explore the RNA-seq pipeline.](#)

Setting up the notebook

In [1]:

```
# Import python packages
import os
import pandas as pd
import numpy as np
import plotly.graph_objects as go
from plotly.subplots import make_subplots
```

Total Expressed Genes (Annotated Genes)

Unnormalized Counts

In [2]:

```
os.chdir("URR_Compare_Analysis")
os.listdir()
```

Out[2]:

```
['.DS_Store',
'LPkit_SampleTable.csv',
'NS_SampleTable.csv',
'NumNonZeroGenes.csv',
'star_alignment.csv',
'URR_Compare_Analysis.html',
'RSEM_Unnormalized_Counts.csv',
'Normalized_Counts.csv',
'.ipynb_checkpoints']
```

In [3]:

```
# Get NumNonZeroGenes.csv
totgenes_file = os.listdir()[3]
totgenes_table = pd.read_csv(totgenes_file, index_col=0)
totgenes_table.index.rename("Sample", inplace=True)
pd.set_option("max_columns", None)
totgenes_table.head()
```

Out[3]:

Number of genes with non-zero counts

Sample	Count
FS_20190404_HRep1	25217
FS_20190404_HRep2	24548

Number of genes with non-zero counts

Sample	
FS_20190404_HRep3	24595
FS_20190404_Rep10	28473
FS_20190404_Rep11	25218

In [4]:

```
# Get RSEM_Unnormalized_Counts.csv
unnorm_file = os.listdir() [6]
unnorm_cutoff = pd.read_csv(unnorm_file).rename(columns={"Unnamed: 0": "Genes"})
unnorm_cutoff.head()
```

Out [4]:

	Genes	FS_20190404_HRep1	FS_20190404_HRep2	FS_20190404_HRep3	FS_20190404_	1
0	ENSMUSG000000000001	5373.0	4574.0	4647.0		
1	ENSMUSG000000000003	0.0	0.0	0.0		
2	ENSMUSG000000000028	1755.0	1376.0	1434.0		
3	ENSMUSG000000000031	3181.0	2704.0	2581.0		
4	ENSMUSG000000000037	68.0	72.0	63.0		

In [5]:

```
# Unnormalized counts cutoff > 10
unnorm_cutoff = unnorm_cutoff.set_index(keys="Genes")
unnorm_cutoff = unnorm_cutoff[unnorm_cutoff.index.str.contains("ENSMUSG")]
unnorm_cutoff = unnorm_cutoff[unnorm_cutoff > 10]
unnorm_10 = unnorm_cutoff.count().to_frame(name="Number of genes with more than 10 counts")
unnorm_10.index.rename("Sample", inplace=True)
unnorm_10.head()
```

Out [5]:

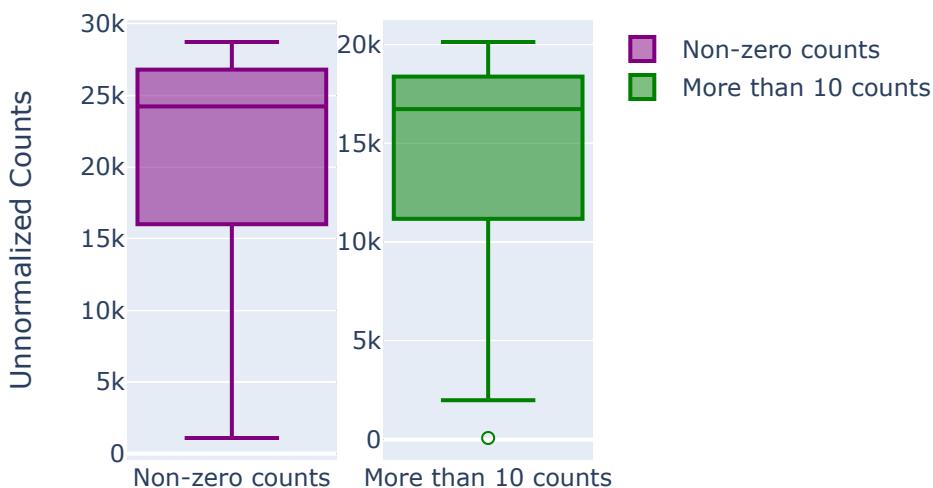
Number of genes with more than 10 counts

Sample	
FS_20190404_HRep1	17074
FS_20190404_HRep2	16730
FS_20190404_HRep3	16669
FS_20190404_Rep10	19635
FS_20190404_Rep11	16985

In [6]:

```
# Box plot of unnormalized counts >0 and >10 for all samples
data = totgenes_table.melt()
data10 = unnorm_10.melt()
fig = go.Figure()
fig = make_subplots(rows=1, cols=2)
fig.add_trace(go.Box(y=data["value"], quartilemethod="inclusive", name="Non-zero counts",
                     row=1, col=1) # "exclusive", "inclusive", or "linear" by default
fig.add_trace(go.Box(y=data10["value"], quartilemethod="inclusive", name="More than 10 cou
                     row=1, col=2)
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.5)
fig.update_layout(title_text="RSEM Unnormalized Counts", title_x=0.4, yaxis_title="Unnorma
                                boxmode="overlay", hovermode="x unified", width=500, height=400)
fig.show()
```

RSEM Unnormalized Counts



Comparative analyses from different NovaSeq runs

Total Expressed Genes (Annotated Genes)

```
In [7]: # Go to URR_Compare_Analysis  
os.listdir()
```

```
Out[7]: ['.DS_Store',  
'LPkit_SampleTable.csv',  
'NS_SampleTable.csv',  
'NumNonZeroGenes.csv',  
'star_alignment.csv',  
'URR_Compare_Analysis.html',  
'RSEM_Unnormalized_Counts.csv',  
'Normalized_Counts.csv',  
.ipynb_checkpoints]
```

```
In [8]: # Get NumNonZeroGenes.csv  
totgenes_file = os.listdir()[3]  
totgenes_table = pd.read_csv(totgenes_file, index_col=0)  
totgenes_table.index.rename("Sample", inplace=True)  
pd.set_option("max_columns", None)  
totgenes_table.head()
```

```
Out[8]: Number of genes with non-zero counts
```

Sample	Count
FS_20190404_HRep1	25217
FS_20190404_HRep2	24548
FS_20190404_HRep3	24595
FS_20190404_Rep10	28473
FS_20190404_Rep11	25218

In [9]:

```
# Get LPkit_SampleTable.csv
kit_file = os.listdir()[1]
kit_table = pd.read_csv(kit_file).rename(columns={"Unnamed: 0": "Sample", "condition": "Library kit"})
kit_table.sort_values(by="Sample", ascending=True)
kit_table.head()
```

Out[9]:

	Sample	Library kit
0	FS_20190404_HRep1	Illumina_TruSeq_Stranded_totRNA_Gold
1	FS_20190404_HRep2	Illumina_TruSeq_Stranded_totRNA_Gold
2	FS_20190404_HRep3	Illumina_TruSeq_Stranded_totRNA_Gold
3	FS_20190404_Rep1	Illumina_TruSeq_Stranded_totRNA_Gold
4	FS_20190404_Rep10	Illumina_TruSeq_Stranded_totRNA_Gold

In [10]:

```
# Get NS_SampleTable.csv
ns_file = os.listdir()[2]
ns_table = pd.read_csv(ns_file).rename(columns={"Unnamed: 0": "Sample", "condition": "NovaSeqRun"})
ns_table.head()
```

Out[10]:

	Sample	NovaSeqRun
0	FS_20190404_HRep1	FS
1	FS_20190404_HRep2	FS
2	FS_20190404_HRep3	FS
3	FS_20190404_Rep1	FS
4	FS_20190404_Rep2	FS

In [11]:

```
# Get Uniquely mapped reads from star_alignment.csv
aligned_file = os.listdir()[4]
aligned_table = pd.read_csv(aligned_file).rename(columns={"Category": "Sample"})
depth_table = aligned_table.filter(["Sample", "Uniquely mapped"]).rename(columns={"Unique": "Uniquely mapped reads"})
depth_table["Uniquely mapped reads"] = depth_table["Uniquely mapped reads"] / 1000000
depth_table = depth_table.rename(columns={"Uniquely mapped reads": "Uniquely mapped reads"})
depth_table.head()
```

Out[11]:

	Sample	Uniquely mapped reads (M)
0	FS_20190404_HRep1	40.272953
1	FS_20190404_HRep2	34.034887
2	FS_20190404_HRep3	34.851778
3	FS_20190404_Rep1	110.888770
4	FS_20190404_Rep10	115.690366

In [12]:

```
# Non-zero unnormalized counts of different library kits used
kit_summary = kit_table.merge(totgenes_table, on="Sample")
kit_summary = kit_summary.set_index(keys="Sample")
kit_summary.head()
```

Out[12]:

Library kit	Number of genes with non-zero counts
-------------	--------------------------------------

Sample	Library kit	Number of genes with non-zero counts
Sample		
FS_20190404_HRep1	Illumina_TruSeq_Stranded_totRNA_Gold	25217
FS_20190404_HRep2	Illumina_TruSeq_Stranded_totRNA_Gold	24548
FS_20190404_HRep3	Illumina_TruSeq_Stranded_totRNA_Gold	24595
FS_20190404_Rep1	Illumina_TruSeq_Stranded_totRNA_Gold	28129
FS_20190404_Rep10	Illumina_TruSeq_Stranded_totRNA_Gold	28473

In [13]:

```
# Non-zero unnormalized counts of different NovaSeq runs
ns_summary = ns_table.merge(kit_summary, on="Sample")
ns_summary
```

Out[13]:

	Sample	NovaSeqRun	Library kit	Number of genes with non-zero counts
0	FS_20190404_HRep1	FS	Illumina_TruSeq_Stranded_totRNA_Gold	25217
1	FS_20190404_HRep2	FS	Illumina_TruSeq_Stranded_totRNA_Gold	24548
2	FS_20190404_HRep3	FS	Illumina_TruSeq_Stranded_totRNA_Gold	24595
3	FS_20190404_Rep1	FS	Illumina_TruSeq_Stranded_totRNA_Gold	28129
4	FS_20190404_Rep2	FS	Illumina_TruSeq_Stranded_totRNA_Gold	26941
...
149	RR10_KDN_UPX_20220104_7	RR10_KDN_UPX	QIAseq_UPX_mRNA	14842
150	RR10_KDN_UPX_20220104_8	RR10_KDN_UPX	QIAseq_UPX_mRNA	14774
151	RR23_LVR_LNG_20220112_2_Xp	RR23_LVR_LNG	Illumina_TruSeq_Stranded_totRNA_Gold	28463
152	RR23_LVR_LNG_20220112_3_Xp	RR23_LVR_LNG	Illumina_TruSeq_Stranded_totRNA_Gold	26158
153	PI_HRT_20220112_Xp	PI_HRT	Illumina_TruSeq_Stranded_totRNA_Gold	24734

154 rows × 4 columns

In [14]:

```
# Non-zero unnormalized counts of different NovaSeq runs
depth_summary = ns_summary.merge(depth_table, on="Sample")
depth_summary = depth_summary.filter(["Sample", "NovaSeqRun", "Number of genes with non-zero counts"])
depth_summary.head()
```

Out[14]:

	Sample	NovaSeqRun	Number of genes with non-zero counts	Uniquely mapped reads (M)
0	FS_20190404_HRep1	FS	25217	40.272953
1	FS_20190404_HRep2	FS	24548	34.034887
2	FS_20190404_HRep3	FS	24595	34.851778
3	FS_20190404_Rep1	FS	28129	110.888770
4	FS_20190404_Rep2	FS	26941	72.993442

In [15]:

```
# Group by NS run
```

```
ind_run = dict(list(depth_summary.groupby("NovaSeqRun")))
ind_run
```

Out[15]:

'FS':	Sample	NovaSeqRun	Number of genes with non-zero counts	\
0	FS_20190404_HRep1	FS	25217	
1	FS_20190404_HRep2	FS	24548	
2	FS_20190404_HRep3	FS	24595	
3	FS_20190404_Rep1	FS	28129	
4	FS_20190404_Rep2	FS	26941	
5	FS_20190404_Rep3	FS	27068	
6	FS_20190404_Rep4	FS	27455	
7	FS_20190404_Rep5	FS	26860	
8	FS_20190404_Rep6	FS	27144	
9	FS_20190404_Rep7	FS	28065	
10	FS_20190404_Rep8	FS	28421	
11	FS_20190404_Rep9	FS	27759	
12	FS_20190404_Rep10	FS	28473	
13	FS_20190404_Rep11	FS	25218	
14	FS_20190404_Rep12	FS	25485	
15	FS_20190404_Rep13	FS	27567	
16	FS_20190404_Rep14	FS	28147	
17	FS_20190404_Rep15	FS	28424	
18	FS_20190404_Rep16	FS	26099	
19	FS_20190404_Rep17	FS	23202	
20	FS_20190404_Rep18	FS	26323	
21	FS_20190404_Rep19	FS	27128	
22	FS_20190404_Rep20	FS	26497	
Uniquely mapped reads (M)				
0		40.272953		
1		34.034887		
2		34.851778		
3		110.888770		
4		72.993442		
5		76.767071		
6		89.686997		
7		79.059419		
8		84.585888		
9		88.462019		
10		95.498190		
11		87.566747		
12		115.690366		
13		45.072266		
14		47.183193		
15		76.889819		
16		141.445416		
17		121.429975		
18		67.321401		
19		95.642894		
20		78.873987		
21		84.886688		
22		120.532743 ,		
'PI_FF_20211026':	Sample	NovaSeqRun	Number of genes with non-zero co	unts \
138	PI_FF_20211026_1	PI_FF_20211026	24574	
139	PI_FF_20211026_2	PI_FF_20211026	24160	
140	PI_FF_20211026_3	PI_FF_20211026	24188	
Uniquely mapped reads (M)				
138		37.173529		
139		31.926943		
140		33.138743 ,		
'PI_FF_20211124':	Sample	NovaSeqRun	Number of genes with non-zero co	unts \
141	PI_FF_20211124_1	PI_FF_20211124	21979	

142 PI FF 20211124 2 PI FF 20211124

22183

	Sample	NovaSeqRun	\
146	14724	7.484766	,
147	14189	12.430669	,
148	14143	6.223526	,
149	14842	9.257538	,
150	14774	8.399899	,
'RR23_LVR_LNG':	Sample	NovaSeqRun	\
151 RR23_LVR_LNG_20220112_2_Xp	RR23_LVR_LNG		,
152 RR23_LVR_LNG_20220112_3_Xp	RR23_LVR_LNG		,
Number of genes with non-zero counts Uniquely mapped reads (M)			
151	28463	171.437695	,
152	26158	74.171619	,
'RR6_CLN_LNG':	Sample	NovaSeqRun	\
38 RR6_CLN_LNG_20190718_Rep1	RR6_CLN_LNG		,
39 RR6_CLN_LNG_20190718_Rep2	RR6_CLN_LNG		,
40 RR6_CLN_LNG_20190718_Rep3	RR6_CLN_LNG		,
Number of genes with non-zero counts Uniquely mapped reads (M)			
38	27484	82.853300	,
39	27002	68.550399	,
40	26636	73.374007	,
'RR6_LVR_SPL':	Sample	NovaSeqRun	\
41 RR6_LVR_SPL_20190805_Rep3	RR6_LVR_SPL		,
42 RR6_LVR_SPL_20190805_Rep4	RR6_LVR_SPL		,
43 RR6_LVR_SPL_20190805_Rep9	RR6_LVR_SPL		,
44 RR6_LVR_SPL_20190805_Rep10	RR6_LVR_SPL		,
45 RR6_LVR_SPL_20190805_Rep13	RR6_LVR_SPL		,
46 RR6_LVR_SPL_20190805_Rep14	RR6_LVR_SPL		,
Number of genes with non-zero counts Uniquely mapped reads (M)			
41	28731	95.641397	,
42	28133	104.343357	,
43	27875	93.640154	,
44	27517	90.053019	,
45	27392	85.881207	,
46	27002	86.680351	,
'RR6_TMS_DSKN':	Sample	NovaSeqRun	\
35 RR6_TMS_DSKN_20190628_Rep1	RR6_TMS_DSKN		,
36 RR6_TMS_DSKN_20190628_Rep2	RR6_TMS_DSKN		,
37 RR6_TMS_DSKN_20190628_Rep3	RR6_TMS_DSKN		,
Number of genes with non-zero counts Uniquely mapped reads (M)			
35	27178	87.336601	,
36	26803	79.650733	,
37	27614	93.389118	,
'RR7_KDN_SKN':	Sample	NovaSeqRun	\
47 RR7_KDN_SKN_20190909_Rep1	RR7_KDN_SKN		,
48 RR7_KDN_SKN_20190909_Rep2	RR7_KDN_SKN		,
49 RR7_KDN_SKN_20190909_Rep3	RR7_KDN_SKN		,
50 RR7_KDN_SKN_20190909_Rep4	RR7_KDN_SKN		,
51 RR7_KDN_SKN_20190909_Rep5	RR7_KDN_SKN		,
52 RR7_KDN_SKN_20190909_Rep6	RR7_KDN_SKN		,
53 RR7_KDN_SKN_20190909_Rep7	RR7_KDN_SKN		,
54 RR7_KDN_SKN_20190909_Rep8	RR7_KDN_SKN		,
55 RR7_KDN_SKN_20190909_Rep9	RR7_KDN_SKN		,
56 RR7_KDN_SKN_20190909_Rep10	RR7_KDN_SKN		,
Number of genes with non-zero counts Uniquely mapped reads (M)			
47	26669	58.771134	,
48	26778	58.922385	,
49	26928	65.582509	,
50	24600	61.024954	,
51	25151	59.019873	,
52	25111	55.183371	,
53	24950	61.833689	,
54	24961	58.939524	,

110	22855	57.839715
111	22741	54.113288
'UPX_ALSDA100':	Sample	NovaSeqRun \
134	UPX_ALSDA100_20210806_TecRep1	UPX_ALSDA100
135	UPX_ALSDA100_20210806_TecRep2	UPX_ALSDA100
136	UPX_ALSDA100_20210806_TecRep3	UPX_ALSDA100
137	UPX_ALSDA100_20210806_TecRep4	UPX_ALSDA100

	Number of genes with non-zero counts	Uniquely mapped reads (M)
134	23799	97.064195
135	21729	76.338304
136	21411	71.400526
137	21744	75.452050

'UPX_test':	Sample	NovaSeqRun \
67	UPX_test_10_ng_ERCC_20201020_tRep1	UPX_test
68	UPX_test_10_ng_ERCC_20201020_tRep2	UPX_test
69	UPX_test_10_ng_ERCC_20201020_tRep3	UPX_test
70	UPX_test_10_ng_no_ERCC_20201020_tRep1	UPX_test
71	UPX_test_10_ng_no_ERCC_20201020_tRep2	UPX_test
72	UPX_test_10_ng_no_ERCC_20201020_tRep3	UPX_test
73	UPX_test_1_ng_ERCC_20201020_tRep1	UPX_test
74	UPX_test_1_ng_ERCC_20201020_tRep2	UPX_test
75	UPX_test_1_ng_ERCC_20201020_tRep3	UPX_test
76	UPX_test_1_ng_no_ERCC_20201020_tRep1	UPX_test
77	UPX_test_1_ng_no_ERCC_20201020_tRep2	UPX_test
78	UPX_test_1_ng_no_ERCC_20201020_tRep3	UPX_test
79	UPX_test_20_ng_ERCC_20201020_tRep1	UPX_test
80	UPX_test_20_ng_ERCC_20201020_tRep2	UPX_test
81	UPX_test_20_ng_ERCC_20201020_tRep3	UPX_test
82	UPX_test_20_ng_no_ERCC_20201020_tRep1	UPX_test
83	UPX_test_20_ng_no_ERCC_20201020_tRep2	UPX_test
84	UPX_test_20_ng_no_ERCC_20201020_tRep3	UPX_test
85	UPX_test_500_pg_ERCC_20201020_tRep1	UPX_test
86	UPX_test_500_pg_ERCC_20201020_tRep2	UPX_test
87	UPX_test_500_pg_ERCC_20201020_tRep3	UPX_test
88	UPX_test_500_pg_no_ERCC_20201020_tRep1	UPX_test
89	UPX_test_500_pg_no_ERCC_20201020_tRep2	UPX_test
90	UPX_test_500_pg_no_ERCC_20201020_tRep3	UPX_test
91	UPX_test_50_pg_ERCC_20201020_tRep1	UPX_test
92	UPX_test_50_pg_ERCC_20201020_tRep2	UPX_test
93	UPX_test_50_pg_ERCC_20201020_tRep3	UPX_test
94	UPX_test_50_pg_no_ERCC_20201020_tRep1	UPX_test
95	UPX_test_50_pg_no_ERCC_20201020_tRep2	UPX_test
96	UPX_test_50_pg_no_ERCC_20201020_tRep3	UPX_test

	Number of genes with non-zero counts	Uniquely mapped reads (M)
67	17111	3.894863
68	17089	3.943846
69	17100	3.808051
70	17421	5.345093
71	1089	0.025915
72	17377	5.002224
73	12244	3.329272
74	12829	4.375843
75	13711	5.861074
76	12188	4.492884
77	12705	4.673743
78	12356	4.122470
79	17230	3.217031
80	18107	4.148696
81	18361	4.340296
82	17631	3.599179
83	18233	4.156169
84	17363	3.116021
85	11000	4.803449
86	10053	3.897370

```

87          10174            3.945274
88          10097            3.183167
89          11091            4.639503
90          10954            4.200025
91          5256             3.746737
92          5181             3.883702
93          5990             5.213186
94          4664             4.175198
95          4378             3.799869
96          3924             3.199142 , Sample NovaSeqRun \
'UPX_test2_L001':
116  UPX_test2_L001_S1_Qiagen_008X_20210415_1_10ng UPX_test2_L001
117  UPX_test2_L001_S1_Qiagen_008X_20210415_2_10ng UPX_test2_L001
118  UPX_test2_L001_S1_Qiagen_008X_20210415_3_10ng UPX_test2_L001
119  UPX_test2_L001_S3_Illumina_RZP_20210415_1_10ng UPX_test2_L001
120  UPX_test2_L001_S3_Illumina_RZP_20210415_2_10ng UPX_test2_L001
121  UPX_test2_L001_S3_Illumina_RZP_20210415_3_10ng UPX_test2_L001
122  UPX_test2_L001_S5_No_Ribo_Dep_20210415_1_10ng UPX_test2_L001
123  UPX_test2_L001_S5_No_Ribo_Dep_20210415_2_10ng UPX_test2_L001
124  UPX_test2_L001_S5_No_Ribo_Dep_20210415_3_10ng UPX_test2_L001

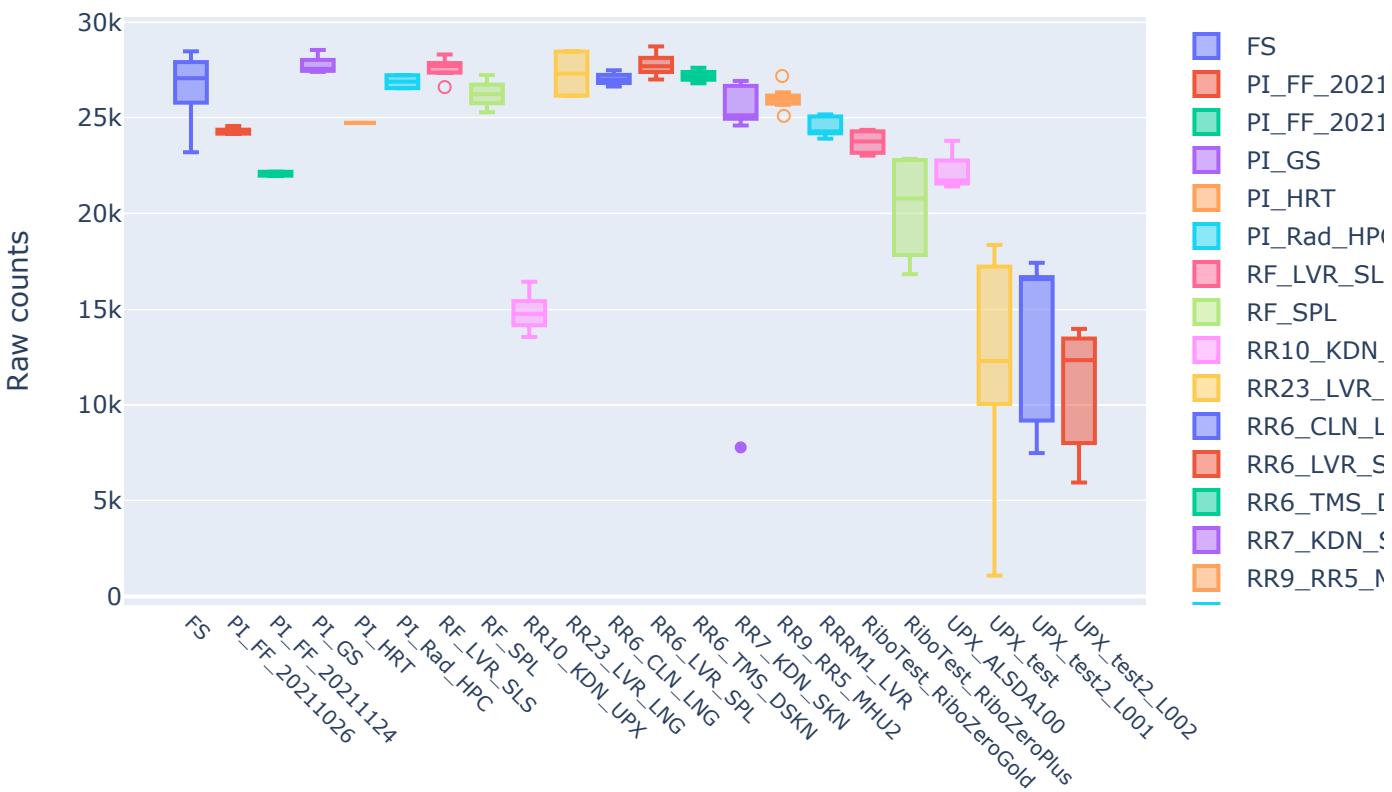
Number of genes with non-zero counts  Uniquely mapped reads (M)
116                      16682            14.087898
117                      16385            14.548883
118                      16996            17.584845
119                      7489             13.358559
120                      9188             29.790044
121                      8203             15.754161
122                      16654            15.155427
123                      17427            17.572372
124                      16653            13.089206 , Sample NovaSeqRun \
'UPX_test2_L002':
125  UPX_test2_L002_S1_Qiagen_05X_20210415_1_10ng UPX_test2_L002
126  UPX_test2_L002_S1_Qiagen_05X_20210415_2_10ng UPX_test2_L002
127  UPX_test2_L002_S1_Qiagen_05X_20210415_3_10ng UPX_test2_L002
128  UPX_test2_L002_S2_Qiagen_1X_20210415_1_10ng UPX_test2_L002
129  UPX_test2_L002_S2_Qiagen_1X_20210415_2_10ng UPX_test2_L002
130  UPX_test2_L002_S2_Qiagen_1X_20210415_3_10ng UPX_test2_L002
131  UPX_test2_L002_S3_No_Ribo_Dep_20210415_1_10ng UPX_test2_L002
132  UPX_test2_L002_S3_No_Ribo_Dep_20210415_2_10ng UPX_test2_L002
133  UPX_test2_L002_S3_No_Ribo_Dep_20210415_3_10ng UPX_test2_L002

Number of genes with non-zero counts  Uniquely mapped reads (M)
125                      12342            6.287190
126                      13472            9.021345
127                      13976            11.122008
128                      8013             15.886517
129                      6126             9.574508
130                      5950             6.696182
131                      13410            4.821391
132                      8655             1.303264
133                      13629            4.788189 }

In [16]: # Box plot grouped by uniquely mapped reads for each NS run on same graph
fig = go.Figure()
fig = make_subplots(rows=1, cols=1)
for i in ind_run:
    fig.add_trace(go.Box(y=ind_run[i]["Number of genes with non-zero counts"], quartilemethod="range", name=i), row=1, col=1)
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.75)
fig.update_layout(title_text="Raw Counts by NovaSeq Run", title_x=0.4, yaxis_title="Raw counts", boxmode="group", hovermode="x unified", width=800, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()

```

Raw Counts by NovaSeq Run



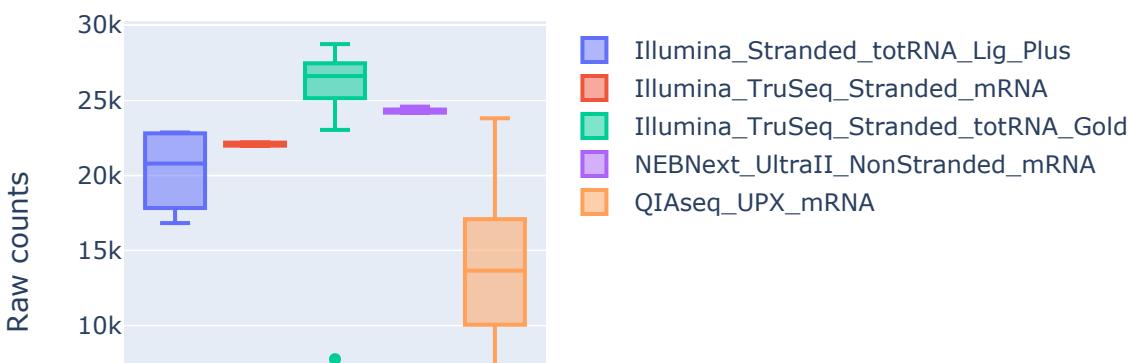
In [17]:

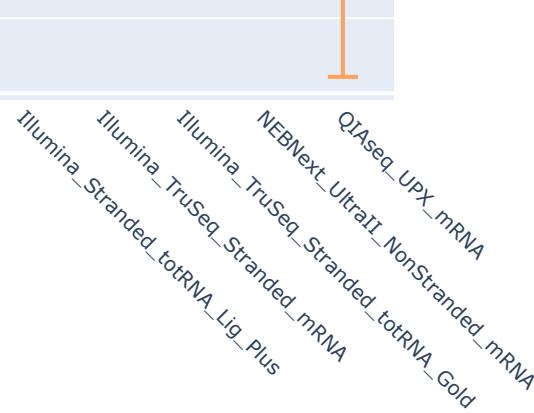
```
# Group by library kits
kit_run = dict(list(ns_summary.groupby("Library kit")))
```

In [18]:

```
# Box plot grouped by library preparation for each NS run on same graph
fig = go.Figure()
for i in kit_run:
    fig.add_trace(go.Box(y=kit_run[i] ["Number of genes with non-zero counts"], quartilemethod="exclusive", name=i))
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.1, textsrc="inside", width=0.75)
fig.update_layout(title_text="Raw Counts by Library Kit", title_x=0.12, yaxis_title="Raw counts", boxmode="group", hovermode="x unified", width=600, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()
```

Raw Counts by Library Kit





In [19]:

```
# Box plot grouped by uniquely mapped reads for samples from all NS runs on same graph
fig = go.Figure()
fig.add_trace(go.Box(x=depth_summary["Uniquely mapped reads (M)"], y=depth_summary["Number of samples"],
                     name="Non-zero counts", marker_color="purple"))
fig.update_traces(boxpoints="suspectedoutliers", jitter=0.3, textsrc="inside", width=1)
fig.update_layout(title_text="Raw Counts by Aligned Uniquely Mapped Reads", title_x=0.5,
                  boxmode="group", hovermode="x unified", width=600, height=500)
fig.update_xaxes(tickangle = 45, tickfont = {"size": 10})
fig.show()
```

DESeq2 Normalized Data

In []:

```
# Change to R kernel
getwd()
```

```
In [1]: # Go to Normalized_Counts.csv and SampleTable.csv directory  
work_dir="URR_Compare_Analysis"  
setwd(file.path(work_dir))
```

```
In [2]: ## Install and load ggfortify and ggplot if not already installed  
if (!requireNamespace("BiocManager", quietly = TRUE))  
    install.packages("BiocManager")  
  
BiocManager::install("tidyverse")  
BiocManager::install("ggfortify")  
BiocManager::install("plotly")  
library(tidyverse)  
library(ggfortify)  
library(plotly)
```

'getOption("repos")' replaces Bioconductor standard repositories, see
'?repositories' for details

replacement repositories:
CRAN: <https://cran.r-project.org>

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:

"package(s) not installed when version(s) same as current; use `force = TRUE` to
re-install: 'tidyverse'"

Old packages: 'BiocManager', 'blob', 'brew', 'callr', 'class', 'cli', 'clipr',
'cluster', 'colorspace', 'commonmark', 'cpp11', 'crayon', 'curl',
'data.table', 'DBI', 'desc', 'devtools', 'digest', 'evaluate', 'fansi',
'farver', 'foreign', 'formatR', 'GenomeInfoDb', 'gert', 'gh', 'gitcreds',
'glue', 'gtable', 'htmltools', 'httr', 'IRkernel', 'isoband', 'jsonlite',
'knitr', 'locfit', 'magrittr', 'markdown', 'MASS', 'Matrix', 'matrixStats',
'mgcv', 'nlme', 'nnet', 'openssl', 'packrat', 'pbDZMQ', 'pillar', 'pkgload',
'processx', 'ps', 'purrr', 'RColorBrewer', 'Rcpp', 'RcppArmadillo', 'RCurl',
'readr', 'rmarkdown', 'roxygen2', 'rpart', 'rprojroot', 'rsconnect',
'RSQLite', 'rstudioapi', 'rversions', 'S4Vectors', 'spatial', 'stringi',
'stringr', 'survival', 'sys', 'testthat', 'tibble', 'tidyselect', 'tinytex',
'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',
'zip'

'getOption("repos")' replaces Bioconductor standard repositories, see
'?repositories' for details

replacement repositories:
CRAN: <https://cran.r-project.org>

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:

"package(s) not installed when version(s) same as current; use `force = TRUE` to
re-install: 'ggfortify'"

Old packages: 'BiocManager', 'blob', 'brew', 'callr', 'class', 'cli', 'clipr',
'cluster', 'colorspace', 'commonmark', 'cpp11', 'crayon', 'curl',
'data.table', 'DBI', 'desc', 'devtools', 'digest', 'evaluate', 'fansi',
'farver', 'foreign', 'formatR', 'GenomeInfoDb', 'gert', 'gh', 'gitcreds',
'glue', 'gtable', 'htmltools', 'httr', 'IRkernel', 'isoband', 'jsonlite',
'knitr', 'locfit', 'magrittr', 'markdown', 'MASS', 'Matrix', 'matrixStats',
'mgcv', 'nlme', 'nnet', 'openssl', 'packrat', 'pbDZMQ', 'pillar', 'pkgload',
'processx', 'ps', 'purrr', 'RColorBrewer', 'Rcpp', 'RcppArmadillo', 'RCurl',
'readr', 'rmarkdown', 'roxygen2', 'rpart', 'rprojroot', 'rsconnect',
'RSQLite', 'rstudioapi', 'rversions', 'S4Vectors', 'spatial', 'stringi',
'stringr', 'survival', 'sys', 'testthat', 'tibble', 'tidyselect', 'tinytex',
'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',
'zip'

```

'usethis', 'uuid', 'viridisLite', 'vroom', 'waldo', 'xfun', 'XML', 'yaml',
'zip'

'getOption("repos")' replaces Bioconductor standard repositories, see
'?repositories' for details

replacement repositories:
  CRAN: https://cran.r-project.org

Bioconductor version 3.14 (BiocManager 1.30.16), R 4.1.2 (2021-11-01)

Warning message:
“package(s) not installed when version(s) same as current; use `force = TRUE` to
re-install: ‘plotly’”

Old packages: ‘BiocManager’, ‘blob’, ‘brew’, ‘callr’, ‘class’, ‘cli’, ‘clipr’,
‘cluster’, ‘colorspace’, ‘commonmark’, ‘cpp11’, ‘crayon’, ‘curl’,
‘data.table’, ‘DBI’, ‘desc’, ‘devtools’, ‘digest’, ‘evaluate’, ‘fanansi’,
‘farver’, ‘foreign’, ‘formatR’, ‘GenomeInfoDb’, ‘gert’, ‘gh’, ‘gitcreds’,
‘glue’, ‘gttable’, ‘htmltools’, ‘httr’, ‘IRkernel’, ‘isoband’, ‘jsonlite’,
‘knitr’, ‘locfit’, ‘magrittr’, ‘markdown’, ‘MASS’, ‘Matrix’, ‘matrixStats’,
‘mgcv’, ‘nlme’, ‘nnet’, ‘openssl’, ‘packrat’, ‘pbDZMQ’, ‘pillar’, ‘pkgload’,
‘processx’, ‘ps’, ‘purrr’, ‘RColorBrewer’, ‘Rcpp’, ‘RcppArmadillo’, ‘RCurl’,
‘readr’, ‘rmarkdown’, ‘roxygen2’, ‘rpart’, ‘rprojroot’, ‘rsconnect’,
‘RSQLite’, ‘rstudioapi’, ‘rversions’, ‘S4Vectors’, ‘spatial’, ‘stringi’,
‘stringr’, ‘survival’, ‘sys’, ‘testthat’, ‘tibble’, ‘tidyselect’, ‘tinytex’,
‘usethis’, ‘uuid’, ‘viridisLite’, ‘vroom’, ‘waldo’, ‘xfun’, ‘XML’, ‘yaml’,
‘zip’
```

— Attaching packages ————— tidyverse 1.3.2 —————

- ✓ ggplot2 3.4.0 ✓ purrr 0.3.4
- ✓ tibble 3.1.6 ✓ dplyr 1.0.10
- ✓ tidyverse 1.3.2 ✓ stringr 1.4.0
- ✓ readr 2.1.2 ✓forcats 0.5.2

— Conflicts ————— tidyverse_conflicts() —————

- ✗ dplyr::filter() masks stats::filter()
- ✗ dplyr::lag() masks stats::lag()

Attaching package: ‘plotly’

The following object is masked from ‘package:ggplot2’:

last_plot

The following object is masked from ‘package:stats’:

filter

The following object is masked from ‘package:graphics’:

layout

In [3]:

```
# Import table with samples and respective groups and column numbers
samp_group <- read.csv(Sys.glob("LPkit_SampleTable.csv"), header = TRUE, row.names = 1, st
samp_group1 <- read.csv(Sys.glob("NS_SampleTable.csv"), header = TRUE, row.names = 1, str
colnames(samp_group)[1] <- "LibraryKit"
colnames(samp_group1)[1] <- "NovaSeqRun"
samp_group2 <- merge(samp_group, samp_group1, by.x=0, by.y=0)
```

```
In [4]: head(samp_group1)
```

A data.frame: 6 × 1

NovaSeqRun

FS_20190404_HRep1 FS
FS_20190404_HRep2 FS
FS_20190404_HRep3 FS
FS_20190404_Rep1 FS
FS_20190404_Rep2 FS
FS_20190404_Rep3 FS

In [5]: head(samp_group2)

A data.frame: 6 × 3

	Row.names	LibraryKit	NovaSeqRun
	<I<chr>>	<fct>	<fct>
1	FS_20190404_HRep1	Illumina_TruSeq_Stranded_totRNA_Gold	FS
2	FS_20190404_HRep2	Illumina_TruSeq_Stranded_totRNA_Gold	FS
3	FS_20190404_HRep3	Illumina_TruSeq_Stranded_totRNA_Gold	FS
4	FS_20190404_Rep1	Illumina_TruSeq_Stranded_totRNA_Gold	FS
5	FS_20190404_Rep10	Illumina_TruSeq_Stranded_totRNA_Gold	FS
6	FS_20190404_Rep11	Illumina_TruSeq_Stranded_totRNA_Gold	FS

In [6]:

```
# Import normalized counts table
normCounts <- read.csv(Sys.glob("Normalized_Counts.csv"), header = TRUE, row.names = 1, st
normCounts <- normCounts +1
```

In [7]:

```

# PCA plot, all samples grouped by NS run
## Indicate PCA plot size
size_var <- 3
alpha_var <- 0.3
exp_raw <- log2(normCounts)
PCA_raw <- prcomp(t(exp_raw), scale = FALSE)

NS <- autoplot(PCA_raw, data = samp_group2, colour = 'NovaSeqRun', shape = 'NovaSeqRun', s
  theme_bw() + theme(axis.text.x = element_text(size=12), axis.text.y = element_text(si
  theme(legend.title = element_text(size=8), legend.text = element_text(size=6)) +
  guides(alpha = guide_legend(order = 1), size = guide_legend(order = 2)) +
  ggtitle("Grouped by NovaSeq Run") + theme(plot.title = element_text(hjust = 0.5)) +
  scale_shape_manual(values = rep(17:22, len = 22))
ggplotly(NS, tooltip = c("text", "size"), width = 700, height = 750)

```

In [8]:

```
# PCA plot, all samples grouped by LP kit
## Indicate PCA plot size
size_var <- 3
alpha_var <- 0.3
exp_raw <- log2(normCounts)
PCA_raw <- prcomp(t(exp_raw), scale = FALSE)

kit <- autoplot(PCA_raw, data = samp_group2, colour = 'LibraryKit', shape = 'LibraryKit',
  theme_bw() + theme(axis.text.x = element_text(size=12), axis.text.y = element_text(
    theme(legend.title = element_text(size=8), legend.text = element_text(size=8)) +
    guides(alpha = guide_legend(order = 1), size = guide_legend(order = 2)) +
    ggtitle("Grouped by Library Kit") + theme(plot.title = element_text(hjust = 0.5)) +
    scale_shape_manual(values = rep(17:22, len = 22)))
ggplotly(kit, tooltip = c("text", "size"), width = 700, height = 400)
```

