### Predicting Intracranial Pressure Monitor Placement in Children with Traumatic Brain Injury

Seth Russell, MS Peter E. DeWitt, Ph.D.

Data Science & Artificial Intelligence Symposium University of Colorado Denver

1 November 2024





#### This work was supported by:

- Mindsource Brain Injury Network of the Colorado Department of Human Services, PI Tell Bennett
- Eunice Kennedy Shriver National Institute for Child Health and Human Development, grant R03 HD094912, PI Tell Bennett

This work is currently under peer review for publication

#### Team of people contributed to this work:

Laura Helmkamp, MS; Kathryn Colborn PhD, MSPH; Charlotte Gray, MPH; Margaret Rebull, MA; Yamila L. Sierra, MPH; Rachel Greer, BA; Lexi Petruccelli, BA; Sara Shankman, CPNP-AC, DNP; Todd C. Hankinson, MD, MBA; Fuyong Xing, PhD; David J. Albers, PhD; and Tellen D. Bennett, MD, MS

#### Motivation

Traumatic Brain Injury - A violent blow or jolt to the head or body that can cause temporary or permanent complications up to death. Swelling is a common serious complication.

Annually in the United States, traumatic brain injury causes:

- 2,200 pediatric deaths
- 35,000 pediatric hospitalizations

To monitor and track brain swelling, an invasive procedure is performed to install a device called an Intercranial Pressure Monitor (ICP) into the skull. Clinicians currently make decisions about ICP placement with traumatic brain injury without the benefit of an accurate clinical decision support tool

#### Study

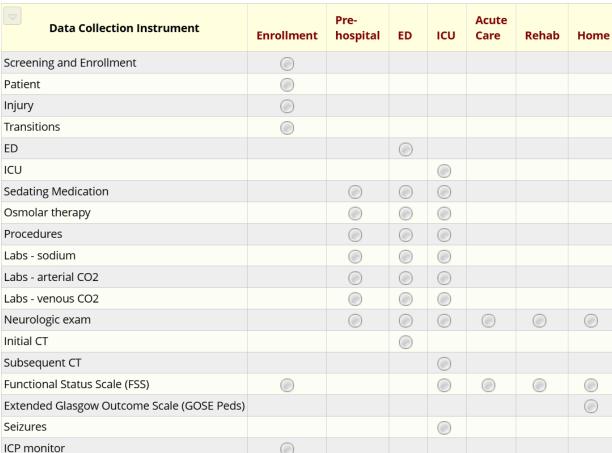
**Aim** - Develop and validate model(s) that predict placement of an ICP monitor. Models should update individual predictions as new information becomes available during an Intensive Care Unit (ICU) stay.

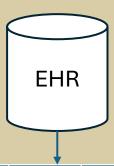
#### Study

- Prospective observational cohort study: September 2014 January 2024
- 389 children with acute traumatic brain injury admitted to the ICU
- 138 (35%) of the 389 participants received ICP monitoring.
- Approved and granted a waiver of consent by the Colorado Multiple
   Institutional Review Board
   https://github.com/magic-lantern/2024-dsai

#### Data

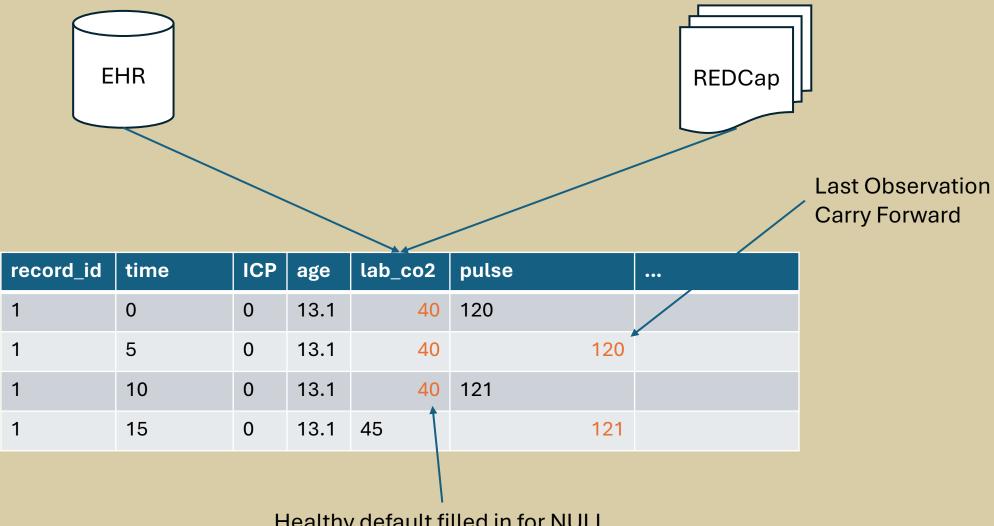






record_id	vital	Units	Value	Datetime
1	pulse	bpm	90	2027-01-01 13:01
1	pulse oximetry	%	98	2027-01-01 13:01
1	respirations		24	2027-01-01 13:01
1	pulse	bpm	91	2027-01-01 14:01

#### Data



Healthy default filled in for NULL

#### Data

#### Exclude CT extracted features and exclude Invasive BP

record_id	time	ICP	age	lab_co2	pulse	
1	0	0	13.1	40	120	
1	5	0	13.1	40	120	
1	10	0	13.1	40	121	
1	15	0	13.1	45	121	

#### Include CT extracted features and exclude Invasive BP

record_id	time	ICP	age	lab_co2	ct_fracture.factor	ct	ct	 
1	0	0	13.1	40	0			
1	5	0	13.1	40	0			
1	10	0	13.1	40	0			
1	15	0	13.1	45	0			

#### Exclude CT extracted features and include Invasive BP

record_id	time	ICP	age	lab_co2	inv_bp_available	•••
1	0	0	13.1	40	0	
1	5	0	13.1	40	0	
1	10	0	13.1	40	0	
1	15	0	13.1	45	1	

#### Include CT extracted features and include Invasive BP

record_id	time	ICP	age	ct_fracture.factor	ct	ct	inv_bp_available	•••
1	0	0	13.1	0			0	
1	5	0	13.1	0			0	
1	10	0	13.1	0			0	
1	15	0	13.1	0			1	

#### Additionally split data temporally into training and testing datasets

Moons KGM, Altman DG, Reitsma JB, *et al.* Transparent Reporting of a multivariable prediction model for Individual Prognosis or Diagnosis (TRIPOD): explanation and elaboration. *Ann Intern Med.* 2015;162:W1-73. doi: 10.7326/M14-0698

#### Modeling

- Standard logistic regression
- Penalized logistic regression (lasso & ridge)
- Support vector machines
- Generalized estimating equations
- Generalized additive models
- Neural Network with Long Short-Term Memory layers



https://clipground.com/images/food-buffet-clipart-3.jpg

#### Modeling – Regression p1

```
2 #' Define several formulae for the glm type models. Same baseline inputs with
 3 #' inclusion and exclusion of ct and invasive bp.
   glm_formula <- list()</pre>
   glm_formula$noct_noinvbp <-</pre>
   icpever ~
   timeint +
   age +
10 - female +
11 inj_mvc +
12 | inj_abuse +
   # inj_fall + # REFERENCE LEVEL for injury mechanism
14 | · · · · inj_other · +
15 | · · · · ni_sbp · +
16 | · · · · ni_dbp · +
17 | · · · etco2 · +
18 | heartrate +
19 · · · spo2 · +
   resprate +
21 | · · · · temp · +
22 ···total_score ·+
23 ···motor_score +
   # pupils_bothreactive + # REFERENCE LEVEL for pupils
   pupils_onefixed +
26 pupils_bothfixed +
27 pupils_unknown +
28 ····lab_na·+
29 |----lab_co2-+
30 ····lab_co2venous
```

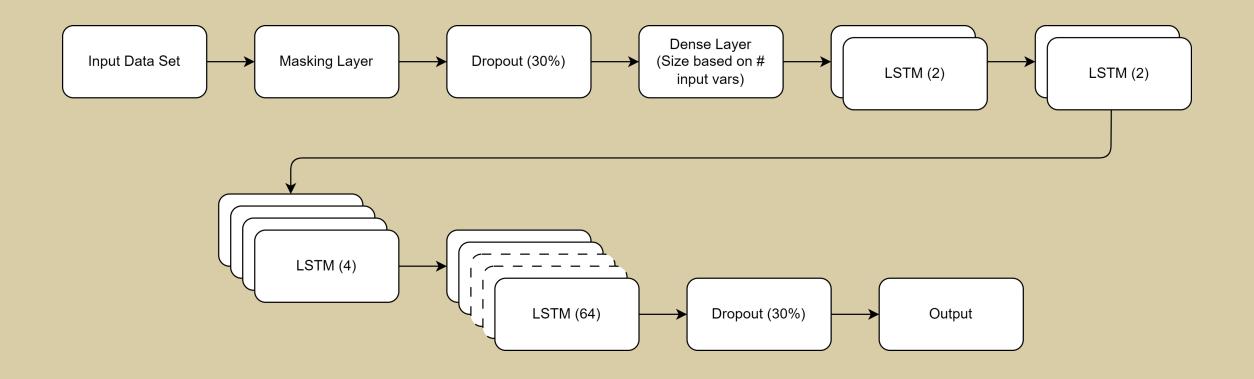
#### Modeling – Regression p2

```
33 if (n_distinct(icp_data_3$noniny_ncnt_vit) > 1) {
34 glm_formula$noct_noinvbp <- update(glm_formula$noct_noinvbp, . ~ . + noninv_ncnt_vit)
  glm_formula$noct_invbp <- update(glm_formula$noct_noinvbp, . ~ . + inv_bp_available)</pre>
39 glm_formula$ct_noinvbp <--
40 | update(glm_formula$noct_noinvbp,
42 ct_compression.factor + ct_intraparhem.factor + ct_subarachem.factor
43 + ct_intravenhem.factor + ct_subduralhem.factor +
44 ct_epiduralhem.factor)
46 glm_formula$ct_invbp <- update(glm_formula$ct_noinvbp, . ~ . + inv_bp_available)
47
     ### Training GLMs
51
     We will fit several models on the training data.
53
     #### Standard logistic regression models.
56 #+ label = "standard glm training fit"
57 tic("Fitting GLM Models")
58 icpever_glm <-
59 lapply(glm_formula,
60 function(x) {
61 | glm(x,
62 | data = icp_data_3,
64 family = binomial())
65 | . . | . . | . . | . . | . })
66 names(icpever_glm) <- paste0("glm_", names(icpever_glm))
```

```
function(x_train,
   ....x_test,
   v_train,
   v_test,
   num_epochs = 10,
   batch_size = 10,
   validation_split = 0.2,
  dropout = 0.35,
random_seed = RANDOM_SEED,
tensorboard_label = 'run',
11 opt = NULL,
12 | layers = c(128, 32, 16, 0),
   checkpoint_file = "model_checkpoint.keras") {
   * # reset random seed and keras states so results are reproducible
15
    reset_state(random_seed)
16
17
    *### Network architecture definition
18
    segmodel <- keras_model_sequential(input_shape = c(dim(x_train)[2], dim(x_train)[3]))</pre>
19
    ··#·without·input·shape
    # building a model with no input shape results in exact same accuracy as with input shape
20
    # however, models without an `input_shape` passed to the first layer cannot reload their
    # optimizer state (cannot continue training)
23
    segmodel %>%
    ···# note - keras allows specifying of some or none of the input_shape size
25
    ···# have to use list(NULL) as c(NULL) automatically discards the NULL entry
    # only specify predictors size for input_shape
    layer_masking(mask_value = 0) %>%
28
   layer_dropout(rate = dropout) %>%
   layer_dense(units = dim(x_train)[3], activation = "relu")
    # could try bidirectional | CTM with code like
```

```
33
   # can specify layers as (layer1, layer2, ..., layern)
34
   # once a layer with 0 is encounted all subsequent values should be 0
   # e.g. (128, 64, 0, 0, 0) is ok
36
   ··#····(128, 0, 64, 0)····is·invalid
   for (i in seq_along(layers)) {
37
38
  if (layers[i] > 0) {
  # if there is a next layer that is not empty, need to include return sequences for LSTM to work
  | | if((i + 1) <= length(layers) & layers[i + 1] > 0) {
40
   ···· # print(paste("i:", i, "adding layer (" , layers[i], ") with return_sequences"))
41
  | | | segmodel %>%
43
  layer_lstm(units = layers[i], return_sequences = TRUE)
  # tried `bias_initializer = "ones" `option, but made results worse given other
45
  # hyperparameters that already worked... Might need to start all over for good
  # results
46
  } else {
47
   segmodel %>%
50
   layer_lstm(units = layers[i])
51
```

```
55
     segmodel %>%
56
    layer_dropout(rate = dropout) %>%
    layer_dense(units = 1, activation = "sigmoid")
57
58
    *# with relu and 80 epochs validation accuracy 0.5492
    *# with sigmoid and 80 epochs validation accuracy 0.
59
60
    # 88% end accuracy before adding dropout layer
61
62
    if (is.null(opt)) {
    seqmodel %>% compile(
    loss = 'binary_crossentropy',
    optimizer = 'rmsprop'
        metrics = c('f1_score', 'binary_accuracy', tf$keras$metrics$Precision(), tf$keras$metrics$Recall()))
67
    } else {
    segmodel %>% compile(
68
   loss = 'binary_crossentropy',
    optimizer = opt,
70
71
        metrics = c('f1_score', 'binary_accuracy', tf$keras$metrics$Precision(), tf$keras$metrics$Recall()))
72
73
     print('Model summary:')
75
     summary(segmodel)
76
77
     checkpoint_file_path <- paste0(save_dir, "checkpoint/", checkpoint_file)</pre>
78
     print(paste("Best model checkpoint saved to:", checkpoint_file_path))
```



```
history <- segmodel %>% fit(
81
    · x_train,
82
     . y_train,
83
    epochs = num_epochs,
84
     batch_size = batch_size,
   validation_split = validation_split,
   callbacks = list(
86
   callback_tensorboard(paste0(save_dir, "tensorboard/", tensorboard_label)),
87
   callback_reduce_lr_on_plateau(monitor = "val_loss",
    factor = 0.5,
       patience = 20,
90
    verbose = 1,
   min_lr = 0.0000001),
92
   callback_early_stopping(monitor = "val_loss",
93
       min delta = 0.0001,
94
      patience = 40,
    verbose = 1,
    mode = c("min")),
97
   ····#·since we have early stopping, this shouldn't be necessary
   ····# also checkpoint can only be every epoch, so will greatly increase disk usage
   callback_model_checkpoint(checkpoint_file_path,
      monitor = "val_loss",
                         save best_only = TRUE,
                          save_weights_only = FALSE,
                         mode = "min",
   save_freq = "epoch")
105
106
   # , shuffle = FALSE # this option results in > 2x more epochs to get to same accuracy
107
   # include this to adjust model weights based on class
   #class weight = list("0"=1,"1"=length(which(y_train==0)) / length(which(y_train==1))),
109
110
    return(list('model' = segmodel, 'history' = history))
112 }
```

## Modeling LSTM Hyperparameters

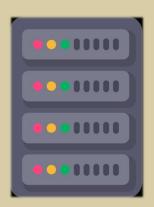
```
23 num_simultaneous_jobs <- 7
24 EPOCH_RUN_TIME <- 40 # time in seconds based on single simultaneous job
25 job_file <- "jul10best.txt"
26 k_folds <- 5 # normally use 5 when finding best hyperparameters
27 # at least with other chosen hyperparameters, 200 epochs gives the best result
28 # doing early stopping if no improvements, so will just use 500 for everything
29 epochs <-c(500)
30 batch_size <- c(1024) # seems to have little effect on accuracy; big difference
31 learn_rate <- c(0.1, 0.05, 0.01, 0.005)
32 validation_split <- c(0.2, 0.3, 0.4)
33 dropout \leftarrow c(0.05, 0.1, 0.2, 0.3, 0.4)
34 run_final <- TRUE
35 # which optimizers to use? see:
36 # http://cs231n.github.io/neural-networks-3/
37 # https://ruder.io/optimizing-gradient-descent/index.html
38 optimizer <- c(
39 ""msprop=optimizer_rmsprop",
40 adam=optimizer_adam",
   "sgd=optimizer_sgd",
42 "adadelta=optimizer_adadelta"
43 |-----
44 predict_advance_times <- c(0)
45 time_block_size <- 5
46 icpever_pred <- TRUE
48 # create grid of network architecture options from these variables
49 num_layers <- seq(4)
50 layer_size <- c(2, 4, 8, 16, 32, 48, 64)
51
52 random_seed <- c(42, 634906796, 1977035194, 1699415226, 1673646895, 1393266542,
                  1083032605, 136228259, 1361975978, 1627064900, 43586684)
54
55 # primarily for final run, want to compare model against restricted input model
56 exclude_vars <- c('"inv","ct"', '"ct"', '"inv"', "")
```

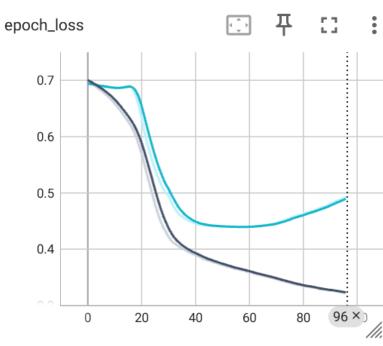
```
81 base_q <- expand.grid(k_folds = k_folds,</pre>
     epochs = epochs,
   batch_size = batch_size,
    learn_rate = learn_rate,
   validation_split = validation_split,
   dropout = dropout,
   random_seed = random_seed,
   optimizer = optimizer,
   predict_advance_times = predict_advance_times,
   run_final = run_final,
   time_block_size = time_block_size,
   icpever_pred = icpever_pred,
   exclude vars = exclude vars)
95 if (!is.na(layers)) {
   base_g$layers <- layers</pre>
96
   g <- as.data.table(base_q)
98 } else {
99
    layer_g <- expand.grid(num_layers = num_layers,</pre>
100
    layer1_size = layer_size,
101
    layer2_size = layer_size,
102
    ·····layer3_size·=·layer_size,
103
   layer4_size = layer_size)
104
105
    *# not sure how to get expand.grid to do what I want, so cleanup and remove extra stuff
106
    q <- layer_q %>%
107
   dplyr::mutate(layer2_size = ifelse(num_layers <= 1, NA, layer2_size)) %>%
   dplyr::mutate(layer3_size = ifelse(num_layers <= 2, NA, layer3_size)) %>%
108
      dplyr::mutate(layer4_size = ifelse(num_layers <= 3, NA, layer4_size)) %>%
109
   tidyr::unite("layers", layer1_size, layer2_size, layer3_size, layer4_size, sep = ",", na.rm = TRUE) %>%
110
111
      dplyr::select(-num_layers) %>%
     unique
112
113
114
    qdt <- as.data.table(q)</pre>
115
    base_gdt <- as.data.table(base_g)
116
    g <- unique(CJ.dt_2(gdt, base_gdt))</pre>
117 }
112
```

#### Modeling – LSTM training

```
cd ~/aim2data_long_job
# run these jobs on GPU0
CUDA_VISIBLE_DEVICES=0 parallel --jobs 4 --delay 120 --timeout=14400 --resume-failed --bar --joblog rnd_gpu0.log < rnd_gpu0_jobs.txt
# Run these jobs on GPU1
CUDA_VISIBLE_DEVICES=1 parallel --jobs 4 --delay 120 --timeout=14400 --resume-failed --bar --joblog rnd_gpu1.log < rnd_gpu1_jobs.txt
# run these jobs on CPU
CUDA_VISIBLE_DEVICES=-1 parallel --jobs 28 --delay 120 --timeout=86400 --memfree 150G --resume-failed --bar --joblog rnd_cpu.log < rnd_cpu_jobs.txt</pre>
```

- (4) Intel Xeon Gold 6416H 'Sapphire Rapids-SP' 2.2 GHz 18-core CPU
- (32) 64GB DDR5 4800 MHz ECC/Registered Memory
- (2) NVIDIA "Ampere" A100 PCI-E+NVLink 80GB
- (4) 7.68TB Intel SSD D3-S4520 2.5" SATA 6Gbps
- (1) 15.36TB Micron 9400 PRO 2.5" U.3 NVMe SSD

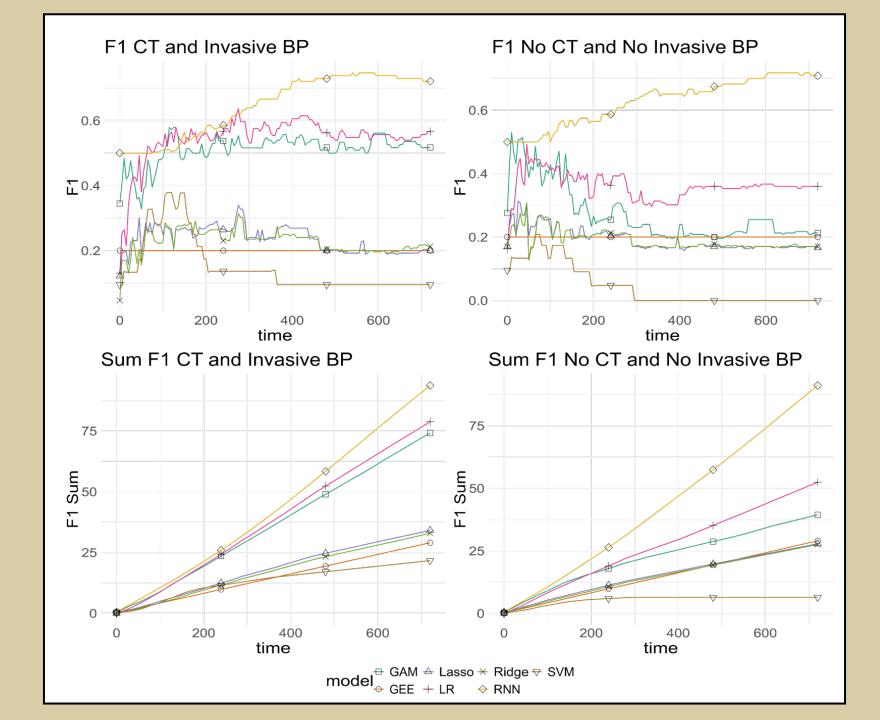




Run ↑

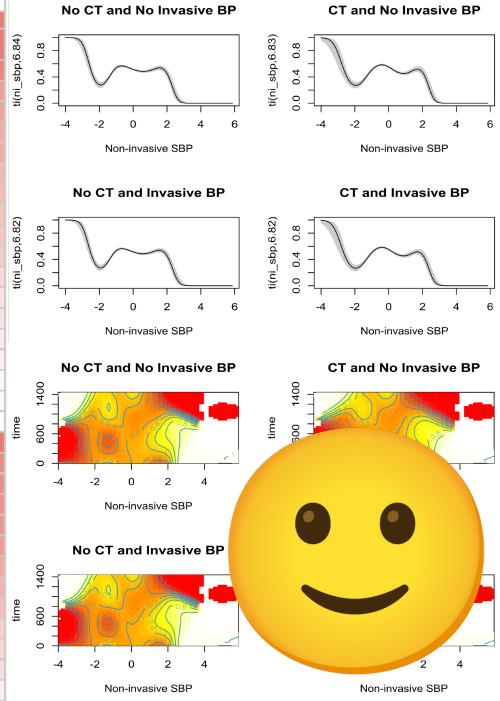
- all\_data\_fold\_1\_epochs\_500\_bs\_1024\_split\_0.2\_drop\_l train
- all\_data\_fold\_1\_epochs\_500\_bs\_1024\_split\_0.2\_drop\_0
- validation

#### Results



# nterpretability

Variable	LR	Lasso	Ridge	Absolute Rank	
CT - Intraparenchymal Hemorrhage	0.88	0.864	0.676	2	
Injury Mechanism - Known or Suspected Abuse	0.903	0.877	0.559	2	6.84)
CT - Cerebral edema	0.742	0.734	0.604	4	sbp.6
CT - Midline shift	0.335	0.327	0.288	10	ti(ni
Pupils - Both Fixed	0.295	0.292	0.266	11	=
Age	0.29	0.284	0.191	13	
Non-invasive DBP	0.279	0.277	0.238	13	
CT - Skull fracture	0.287	0.273	0.174	16	
Pupils - Unknown	0.249	0.243	0.201	17	
Injury Mechanism - Motor Vehicle Crash	0.274	0.255	0.104	19	82)
Arterial pCO2	0.193	0.187	0.195	19	
Injury Mechanism - Other	0.258	0.241	0.111	21	sbp.6
Invasive BP available	0.108	0.105	0.095	24	ti(ni
Female	0.105	0.098	0.072	26	-
Oxygen Saturation (SpO2)	0.097	0.093	0.084	26	
Non-invasive SBP	0.075	0.071	0.079	28	
CT - Basilar cistern compression	0.082	0.075	0.052	29	
Venous pCO2	0.051	0.049	0.061	31	
Number of Vitals Updated	0.028	0.026	0.038	32	
Pupils - One Fixed	0.029	0.026	0.007	33	time
Intercept	-0.938	-2.448	-2.251	1	175
CT - Intraventricular Hemorrhage	-0.62	-0.614	-0.528	5	
CT - Subdural Hematoma	-0.478	-0.467	-0.336	6	
CT - Epidural Hematoma	-0.432	-0.419	-0.337	7	
GCS-Motor	-0.378	-0.382	-0.346	8	
Heart Rate	-0.358	-0.355	-0.295	9	
Time Interval	-0.002	-0.879	-0.717	12	
CT - Subarachnoid Hemorrhage	-0.292	-0.274	-0.154	15	
Respiratory Rate	-0.25	-0.245	-0.177	18	time
End-tidal CO2	-0.147	-0.145	-0.144	22	-
Temperature	-0.128	-0.123	-0.107	23	
Sodium Level	-0.114	-0.106	-0.067	25	
GCS-Total	-0.033	-0.031	-0.092	29	





#### Questions?

Slides available at:

https://github.com/magic-lantern/2024-dsai