



MODULE OVERVIEW

Objective: Provide an introduction to the practice of bioinformatics as well as a practical guide to using common bioinformatics databases and algorithms

1.1 ▶ *Introduction to Bioinformatics*

1.2 ▶ **Sequence Alignment and Database Searching**

1.3 ▶ *Structural Bioinformatics*

1.4 ▶ *Genome Informatics: High Throughput Sequencing Applications and Analytical Methods*

WEEK ONE REVIEW

- Answers to last weeks homework (19/20):

[Answers week 1](#)

- Muddy Point Assessment (14/20):

[Responses](#)

- Need for FASTA header lines ">example1"
- More on protein structure viewing and NGL...
- "what does the AU assembly mean?
- "Great first lab!" ... Nice Assignment".

THIS WEEK'S HOMEWORK

- Check out the "Background Reading" material online:

[Dynamic Programming](#)

[Database Searching](#)

- Complete the **lecture 1.2 homework questions**:

<http://tinyurl.com/bioinf525-quiz2>

TODAYS MENU

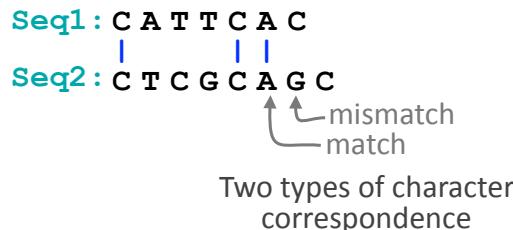
- Alignment basics
 - Why compare biological sequences?
- Homologue detection
 - Orthologs, paralogs, similarity and identity
 - Sequence changes during evolution
 - Alignment view: matches, mismatches and gaps
- Pairwise sequence alignment methods
 - Brute force alignment
 - Dot matrices
 - Dynamic programming (global vs local alignment)
- Rapid heuristic approaches
 - BLAST
- Practical database searching
 - PSI-BLAST and HMM approaches

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.

Seq1: C A T T C A C

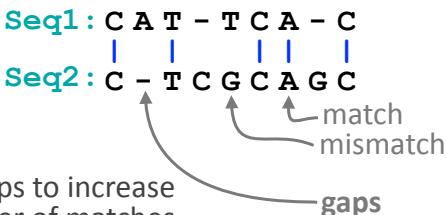
Seq2: C T C G C A G C

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.



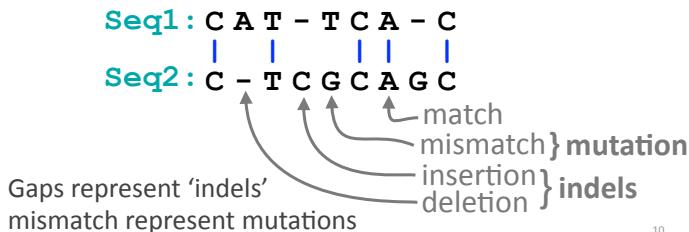
8

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.



9

Basic Idea: Display one sequence above another with spaces (termed **gaps**) inserted in both to reveal **similarity** of nucleotides or amino acids.



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Why compare biological sequences?

- To obtain **functional or mechanistic insight** about a sequence by inference from another potentially better characterized sequence
- To find whether two (or more) genes or proteins are **evolutionarily related**
- To find **structurally or functionally similar regions** within sequences (e.g. catalytic sites, binding sites for other molecules, etc.)
- Many practical bioinformatics applications...

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Practical applications of sequence alignment include...

- **Similarity searching of databases**
 - Protein structure prediction, annotation, etc...
- **Assembly of sequence reads** into a longer construct such as a genomic sequence
- **Mapping sequencing reads to a known genome**
 - "Resequencing", looking for differences from reference genome - SNPs, indels (insertions or deletions)
 - Mapping transcription factor binding sites via ChIP-Seq (chromatin immuno-precipitation sequencing)
 - Pretty much all next-gen sequencing data analysis

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N.B. Pairwise sequence alignment is arguably the most fundamental operation of bioinformatics!

Outline for today

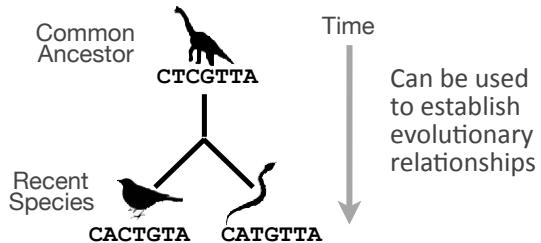
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Sequence comparison is most informative when it detects **homologs**

Homologs are sequences that have common origins i.e. they share a **common ancestor**

- They may or may not have common activity



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Key terms

When we talk about related sequences we use specific terminology.

Homologous sequences may be either:

- **Orthologs or Paralogs**

(Note. these are all or nothing relationships!)

Any pair of sequences may share a certain level of:

- **Identity and/or Similarity**

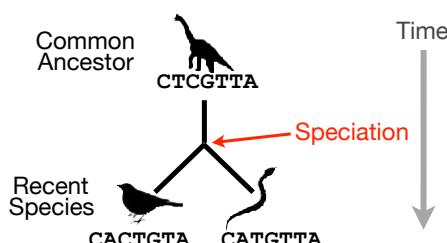
(Note. if these metrics are above a certain level we often infer homology)

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Orthologs tend to have similar function

Orthologs: are homologs produced by speciation that have diverged due to divergence of the organisms they are associated with.

- Ortho = [greek: straight] ... implies direct descent

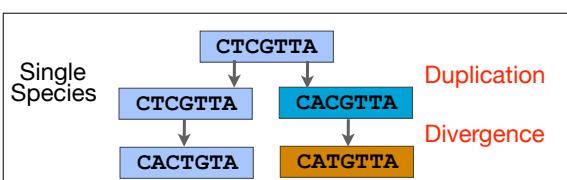


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Paralogs tend to have slightly different functions

Paralogs: are homologs produced by **gene duplication**. They represent genes derived from a common ancestral gene that duplicated within an organism and then subsequently diverged by accumulated mutation.

– Para = [greek: along side of]



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Orthologs vs Paralogs

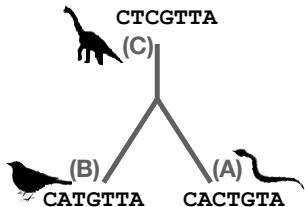
- In practice, determining ortholog vs paralog can be a complex problem:
 - gene loss after duplication,
 - lack of knowledge of evolutionary history,
 - weak similarity because of evolutionary distance
- Homology does not necessarily imply exact same function
 - may have similar function at very crude level but play a different physiological role

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Sequence changes during evolution

There are three major types of sequence change that can occur during evolution.

- Mutations/Substitutions
- Deletions
- Insertions

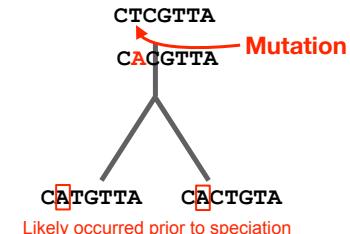


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Mutations, deletions and insertions

There are three major types of sequence change that can occur during evolution.

- **Mutations/Substitutions** $\text{CTCGTTA} \rightarrow \text{CAGTTA}$
- Deletions
- Insertions

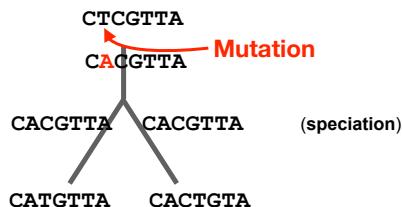


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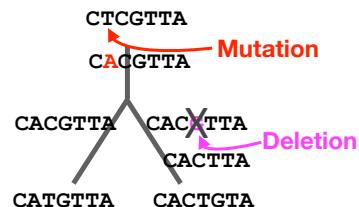


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- Deletions $\text{CACGTTA} \rightarrow \text{CACTTA}$
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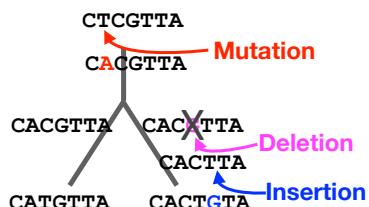


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- Mutations/Substitutions $\text{CTCGTTA} \rightarrow \text{CAGTTA}$
- Deletions $\text{CACGTTA} \rightarrow \text{CACTTA}$
- Insertions $\text{CACTTA} \rightarrow \text{CACTGTA}$

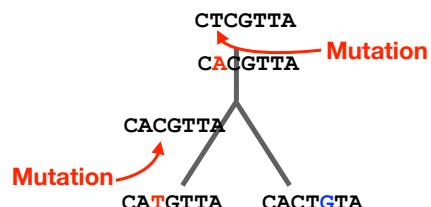


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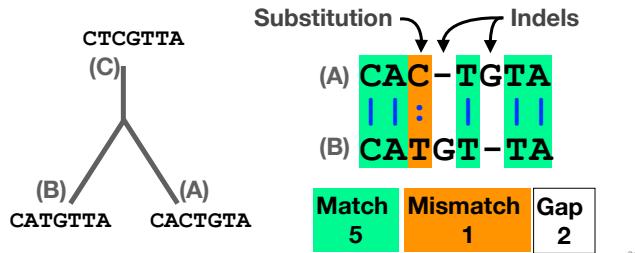


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Alignment view

Alignments are great tools to visualize sequence similarity and evolutionary changes in homologous sequences.

- **Mismatches** represent mutations/substitutions
- **Gaps** represent insertions and deletions (indels)



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Alternative alignments

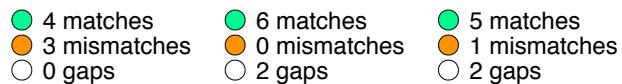
- Unfortunately, finding the correct alignment is difficult if we do not know the evolutionary history of the two sequences
 - There are many possible alignments
 - Which alignment is best?



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Alternative alignments

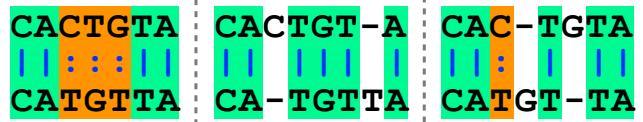
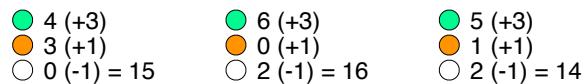
- One way to judge alignments is to compare their number of matches, insertions, deletions and mutations



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Scoring alignments

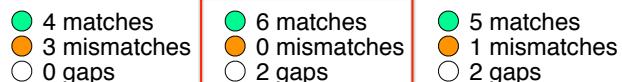
- We can assign a score for each match (+3), mismatch (+1) and indel (-1) to identify the **optimal alignment for this scoring scheme**



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Optimal alignments

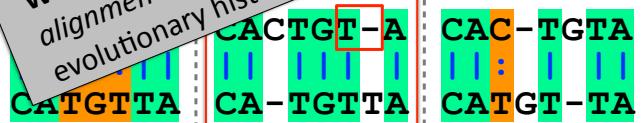
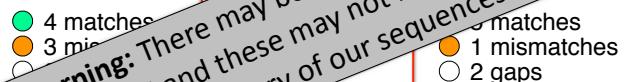
- Biologists often prefer **parsimonious alignments**, where the number of postulated sequence changes is minimized.



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Optimal alignments

- Biologists often prefer **parsimonious alignments**, where the number of postulated sequence changes is minimized.



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Side note: sequence *identity* and *similarity*

- Two commonly quoted metrics for pairs of aligned sequences.
 - **Sequence identity:** typically quotes the percent of identical characters in the aligned region of two sequences
 - **Sequence similarity:** typically the score resulting from optimal pair-wise alignment (**note dependence on parameters used: i.e. scoring scheme**)
- N.B. In contrast, **homology is an all or nothing relationship, you can not have a percent homology!**

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Side note: sequence identity and similarity

- High sequence similarity is frequently used as an indicator of homology
 - Use to find genes and/or proteins with potentially similar or identical function
 - Can query a database of sequences by performing a series of pair-wise alignments
- Knowledge of the difference between sequences can also yield valuable functional and mechanistic insights
 - A gene from a normal and an affected subject – possible cause of a heritable disease
 - Similar proteins with different substrate specificities – what amino acid changes might be responsible for this?

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 - BLAST **Quiz questions:**
- Practical database searching
 - PSI-BLAST

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Pair-wise Sequence Alignment

- **Objective:** arrange two sequences in such a fashion that pairs of matching characters between the two sequences are maximized
 - Match does not have to be identity, can be defined by a function that ranks or scores the characters being compared (often termed a **substitution matrix**)
 - Ungapped alignment example – bars indicate matching characters

Seq1: GTAATCTG-
 | | | | |
Seq2: -TAAGCTGA

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Simplest case – brute force alignments

- In the simplest case we can simply slide one sequence across the other and count matching characters for each possible alignment
 - Choose a scoring scheme and do not allow internal gaps within sequences
 - Algorithmic complexity is linear
- $N + M$ alignments to consider
(where N and M are the length of each sequence)

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GTAATCTG
TTAAGCTGA

GTAATCTG
| |
TTAAGCTGA

Brute Force
Alignment,
No Gaps

GTAATCTG
|
TTAAGCTGA

GTAATCTG
| | | |
TTAAGCTGA

GTAATCTG
|
TTAAGCTGA

GTAATCTG
| |
TTAAGCTGA

GTAATCTG
TTAAGCTGA

GTAATCTG
|
TTAAGCTGA

GTAATCTG
| |
TTAAGCTGA

GTAATCTG
|
TTAAGCTGA

GTAATCTG
TTAAGCTGA

Etc...

Slide from Jeffery de Wet

Gaps make the brute force method unusable for all but the shortest sequences

- Pairs of related sequences often have insertions or deletions relative to one-another, we therefore require **gapped pair-wise alignment**
 - Need to generate all the possible gap lengths and combinations of gaps at all possible positions in both sequences
 - For two sequences of equal length, the formula is:

$$\binom{2N}{N} = \frac{(2N)!}{(N!)^2} \cong \frac{2^{2N}}{\sqrt{\pi N}}$$

N = 10: 184756
N = 50: ~1.00E29
N = 250: ~1.17E149

Slide from Jeffery de Wet

Three general solutions to the alignment problem

- The **dot plot or dot matrix** approach
 - A simple graphical method for pair-wise alignment
 - No scoring, so difficult to compare alternative alignments
 - Can give visual clues to sequence structure but requires human interaction
- Dynamic programming** algorithms
 - Provides Optimal solutions (but not necessarily unique solutions)
- Heuristic **word** or **k-tuple** approaches
 - Much faster (e.g. **BLAST** and **FASTA**)
 - Widely used for database searches
 - May miss some pairs with low similarity

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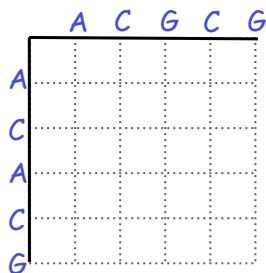
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Dot plots: simple graphical approach

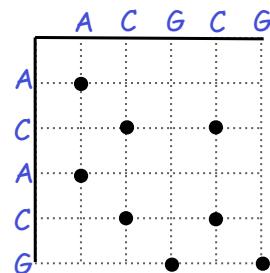
- Place one sequence on the vertical axis of a 2D grid (or matrix) and the other on the horizontal



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Dot plots: simple graphical approach

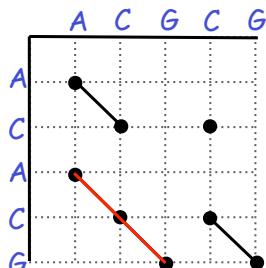
- Now simply put dots where the horizontal and vertical sequence values match



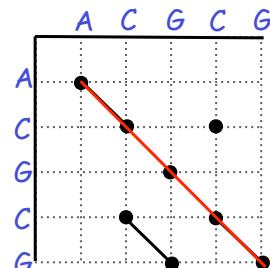
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Dot plots: simple graphical approach

- Diagonal runs of dots indicate matched segments of sequence



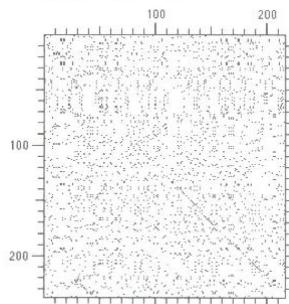
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Dot plots: simple graphical approach

- Dot matrices for long sequences can be noisy



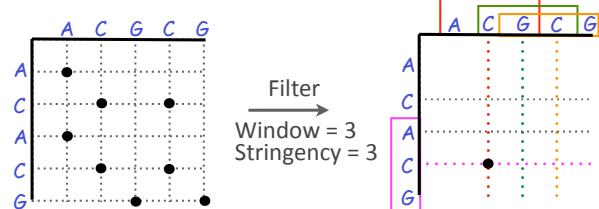
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Dot plots: window size and match stringency

Solution: use a window and a threshold

- compare character by character within a window
- require certain fraction of matches within window in order to display it with a dot.

- You have to choose window size and stringency



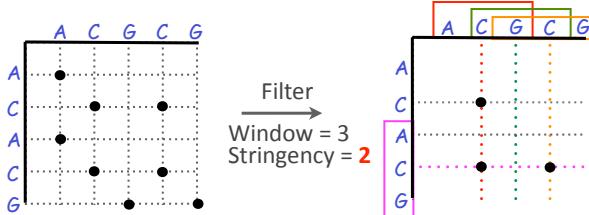
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Dot plots: window size and match stringency

Solution: use a window and a threshold

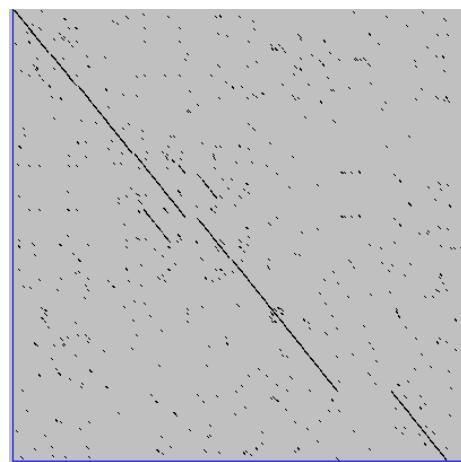
- compare character by character within a window
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Window size = 5 bases

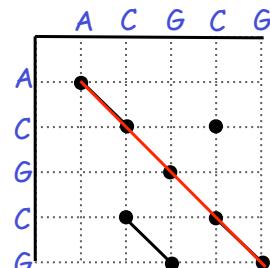


A dot plot simply puts a dot where two sequences match. In this example, dots are placed in the plot if 5 bases in a row match perfectly. Requiring a 5 base perfect match is a **heuristic** – only look at regions that have a certain degree of identity.

Do you expect evolutionarily related sequences to have more word matches (matches in a row over a certain length) than random or unrelated sequences?

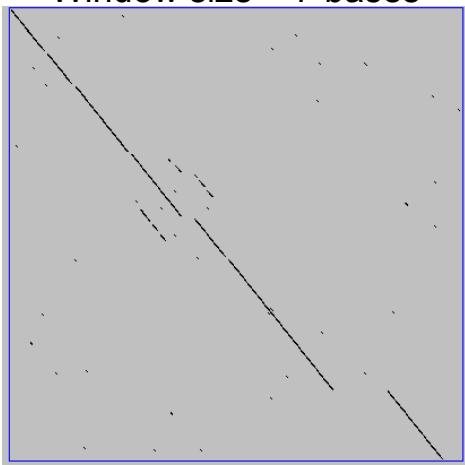
Dot plots: simple graphical approach

- What would the dot matrix of a two identical sequences look like?



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Window size = 7 bases



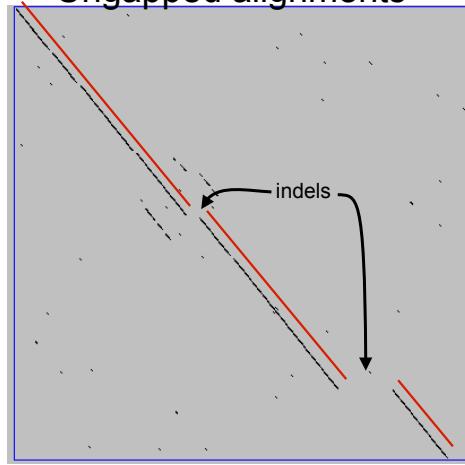
Web site used: <http://www.vivo.colostate.edu/molkit/dnadot/>

This is a dot plot of the same sequence pair. Now 7 bases in a row must match for a dot to be placed. Noise is reduced.

Using windows of a certain length is very similar to using words (kmers) of N characters in the heuristic alignment search tools

Bigger window (kmer) fewer matches to consider

Ungapped alignments

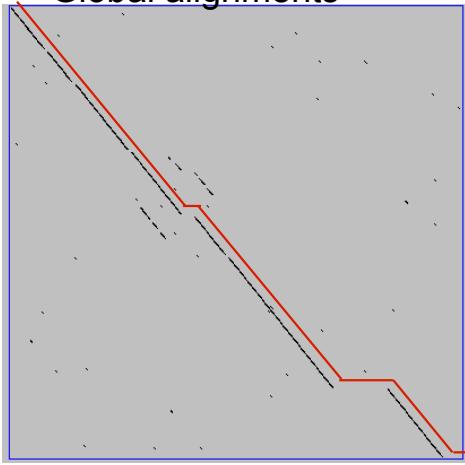


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Only **diagonals** can be followed.

Downward or rightward paths represent **insertion** or **deletions** (gaps in one sequence or the other).

Global alignments



Web site used: <http://www.vivo.colostate.edu/molkit/dnadot/>

Global alignments go from end to end, i.e. from the upper left corner to the lower right corner.

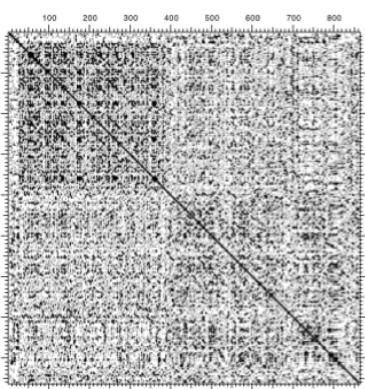
Global alignments do not have good statistical characterization and are **not used for database searches**.

Uses for dot matrices

- Visually assessing the similarity of two protein or two nucleic acid sequences
- Finding local repeat sequences within a larger sequence by comparing a sequence to itself
 - Repeats appear as a set of diagonal runs stacked vertically and/or horizontally

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Repeats



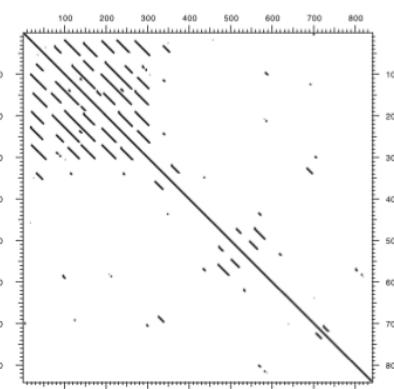
Human LDL receptor protein sequence (Genbank P01130)

W = 1
S = 1

(Figure from Mount, "Bioinformatics sequence and genome analysis")

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Repeats



Human LDL receptor protein sequence (Genbank P01130)

W = 23
S = 7

(Figure from Mount, "Bioinformatics sequence and genome analysis")

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Side note: dots can have “weights”

- Some matches can be rewarded more than others, depending on likelihood
- Use PAM or BLOSUM **substitution matrix**
– (more on these later)
- Put a dot only if a minimum total or average weight is achieved
– See chapter 3 in Mount, “*Bioinformatics sequence and genome analysis*”.

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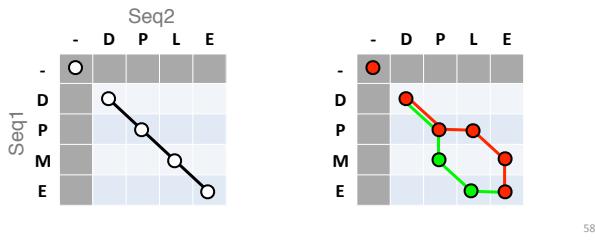
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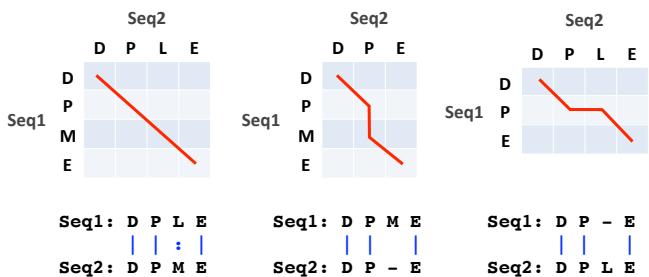
The Dynamic Programming Algorithm

- The dynamic programming algorithm can be thought of an extension to the dot plot approach
 - One sequence is placed down the side of a grid and another across the top
 - Instead of placing a dot in the grid, we **compute a score** for each position
 - Finding the optimal alignment corresponds to finding the path through the grid with the **highest possible score**



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Different paths represent different alignments



Matches are represented by diagonal paths and indels with horizontal or vertical path segments

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Algorithm of Needleman and Wunsch

- The Needleman–Wunsch approach to global sequence alignment has three basic steps:
 - (1) setting up a 2D-grid (or **alignment matrix**),
 - (2) **scoring the matrix**, and
 - (3) identifying the **optimal path** through the matrix



Needleman, S.B. & Wunsch, C.D. (1970) “A general method applicable to the search for similarities in the amino acid sequences of two proteins.” J. Mol. Biol. 48:443-453.

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Scoring the alignment matrix

- Start by filling in the first row and column – these are all indels (gaps).
 - Each step you take you will add the **gap penalty** to the score ($S_{i,j}$) accumulated in the previous cell

j	Sequence 2									
-	D	P	L	E						
-	0	-2	-4	-6	-8					
D	-2									
P	-4									
M	-6									
E	-8									

Scores: match = +1, mismatch = -1, gap = -2

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- Start by filling in the first row and column – these are all indels (gaps).
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		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2				
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

$S_{i+4} = (-2) + (-2) + (-2) + (-2)$

Seq1: DPME
Seq2: -----

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Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which of the three directions gives the highest score?
 - keep track of this score and direction

		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2	?			
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

$i-1 \quad j-1 \quad i \quad j$

$S(i-1, j-1) \xrightarrow{(1)} S(i, j-1) \xrightarrow{(2)} S(i-1, j) \xrightarrow{(3)} S(i, j)$

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Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which of the three directions gives the highest score?
 - keep track of this score and direction

		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2	?			
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

$S(i, j) = \text{Max} \begin{cases} S(i-1, j-1) + (\text{mis})\text{match} \\ S(i-1, j) - \text{gap penalty} \\ S(i, j-1) - \text{gap penalty} \end{cases}$

64

Scoring the alignment matrix

- Then go to the empty corner cell (upper left). It has filled in values in up, left and diagonal directions
 - Now can ask which direction gives the highest score
 - keep track of direction and score

		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2	1			
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

Alignment

① (0)+(+1) = +1 <= (D-D) match!

② (-2)+(-2) = -4 D D

③ (-2)+(-2) = -4

65

Scoring the alignment matrix

- At each step, the score in the current cell is determined by the scores in the neighboring cells
 - The maximal score and the direction that gave that score is stored (we will use these later to determine the optimal alignment)

		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2	1	1		
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

Alignment

① (-2)+(-1) = -3 <= (D-P) mismatch!

② (-4)+(-2) = -6 D P

③ (1)+(-2) = -1

66

Scoring the alignment matrix

- We will continue to store the alignment score ($S_{i,j}$) for all possible alignments in the alignment matrix.

		Sequence 2				
		-	D	P	L	E
Sequence 1	-	0	-2	-4	-6	-8
	D	-2	1	1	1	3
	P	-4				
	M	-6				
	E	-8				
	j					

Scores: match = +1, mismatch = -1, gap = -2

Alignment

① (-4)+(-1) = -5 <= (D-L) mismatch!

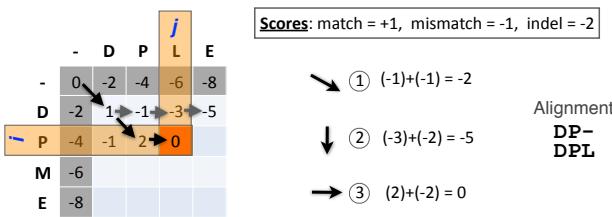
② (-6)+(-2) = -8 D-- DPL

③ (-1)+(-2) = -3

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Scoring the alignment matrix

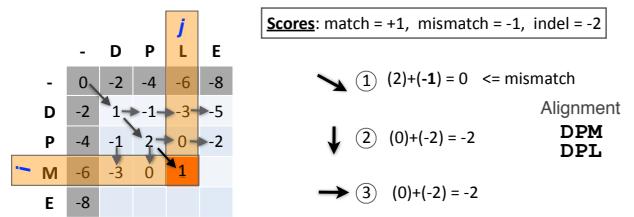
- For the highlighted cell, the corresponding score ($S_{i,j}$) refers to the score of the optimal alignment of the first i characters from sequence1, and the first j characters from sequence2.



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Scoring the alignment matrix

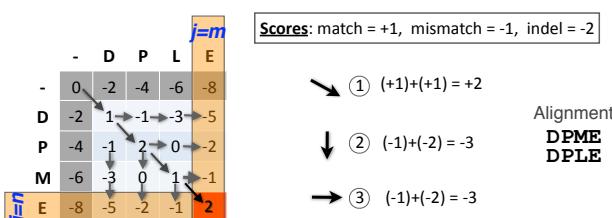
- At each step, the score in the current cell is determined by the scores in the neighboring cells
 - The maximal score and the direction that gave that score is stored



69

Scoring the alignment matrix

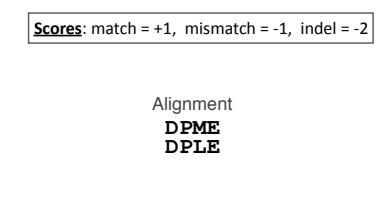
- The score of the best alignment of the entire sequences corresponds to $S_{n,m}$
 - (where n and m are the length of the sequences)



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Scoring the alignment matrix

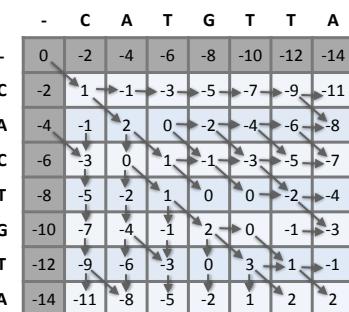
- To find the best alignment, we retrace the arrows starting from the bottom right cell
 - N.B. The optimal alignment score and alignment are dependent on the chosen scoring system



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Questions:

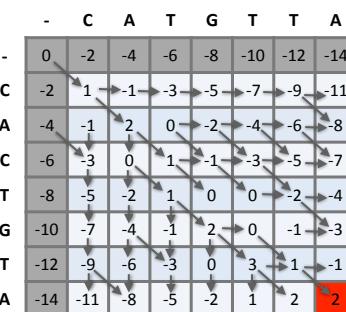
- What is the optimal score for the alignment of these sequences and how do we find the optimal alignment?



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Questions:

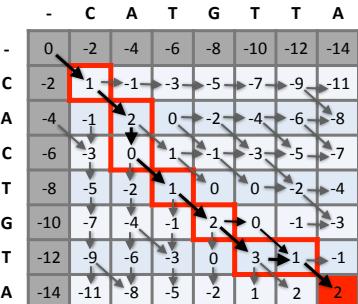
- What is the optimal score for the alignment of these sequences and how do we find the optimal alignment?



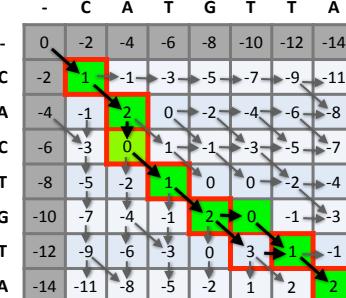
73

Questions:

- To find the best alignment we retrace the arrows starting from the bottom right cell



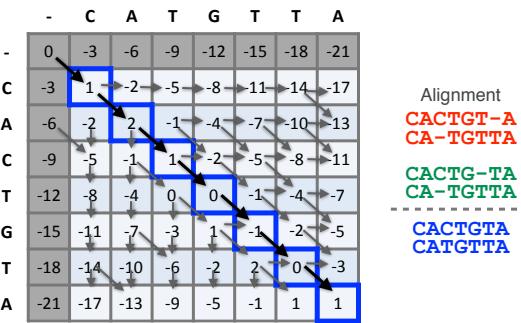
74



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The alignment and score are dependent on the scoring system

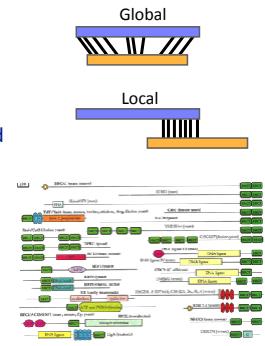
- Here we increase the gap penalty from -2 to -3



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Global vs local alignments

- Needleman-Wunsch is a **global alignment** algorithm
 - Resulting alignment spans the complete sequences end to end
 - This is appropriate for closely related sequences that are similar in length
- For many practical applications we require **local alignments**
 - Local alignments highlight sub-regions (e.g. protein domains) in the two sequences that align well



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Local alignment: Definition

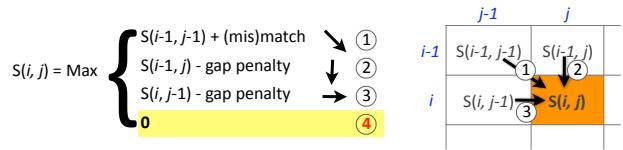
- Smith & Waterman proposed simply that a local alignment of two sequences allow arbitrary-length segments of each sequence to be aligned, with no penalty for the unaligned portions of the sequences. Otherwise, the score for a local alignment is calculated the same way as that for a global alignment

Smith, T.F. & Waterman, M.S. (1981) "Identification of common molecular subsequences." J. Mol. Biol. 147:195-197.

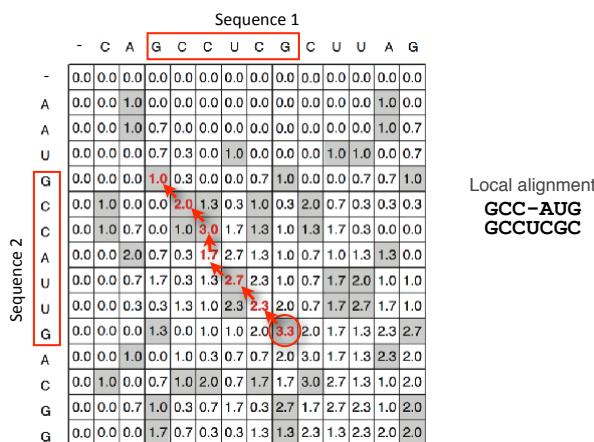
78

The Smith-Waterman algorithm

- Three main modifications to Needleman-Wunsch:
 - Allow a node to start at 0
 - The score for a particular cell cannot be negative
 - if all other score options produce a negative value, then a zero must be inserted in the cell
 - Record the highest-scoring node, and trace back from there



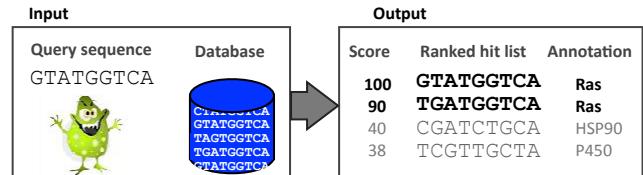
79



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Local alignments can be used for database searching

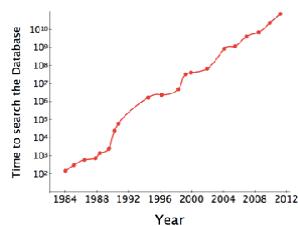
- **Goal:** Given a query sequence (Q) and a sequence database (D), find a list of sequences from D that are most similar to Q
- **Input:** Q, D and scoring scheme
- **Output:** Ranked list of hits



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The database search problem

- Due to the rapid growth of sequence databases, search algorithms have to be both efficient and sensitive
 - Time to search with SW is proportional to $m \times n$ (m is length of query, n is length of database), **too slow for large databases!**



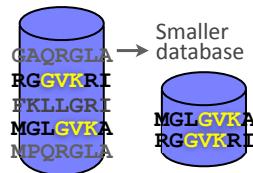
To reduce search time **heuristic algorithms**, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

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The database search problem

- Due to the rapid growth of sequence databases, search algorithms have to be both efficient and sensitive
 - Time to search with SW is proportional to $m \times n$ (m is length of query, n is length of database), **too slow for large databases!**

Query RGGVKRIKLMR



To reduce search time **heuristic algorithms**, such as BLAST, first remove database sequences without a strong local similarity to the query sequence in a quick initial scan.

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Outline for today

- Alignment basics
 - Why compare biological sequences?
- Homologue detection
 - Orthologs, paralogs, similarity and identity
 - Sequence changes during evolution
 - Alignment view: matches, mismatches and gaps
- Pairwise sequence alignment methods
 - Brute force alignment
 - Dot matrices
 - Dynamic programming (global vs local alignment)
- Rapid heuristic approaches
 - BLAST
- Practical database searching
 - PSI-BLAST and HMM approaches

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Rapid, heuristic versions of Smith-Waterman: **BLAST**

- **BLAST** (Basic Local Alignment Search Tool) is a simplified form of Smith-Waterman (SW) alignment that is popular because it is **fast** and **easily accessible**
 - BLAST is a heuristic approximation to SW - It examines only part of the search space
 - BLAST saves time by restricting the search by scanning database sequences for likely matches before performing more rigorous alignments
 - Sacrifices some sensitivity in exchange for speed
 - In contrast to SW, BLAST is not guaranteed to find optimal alignments

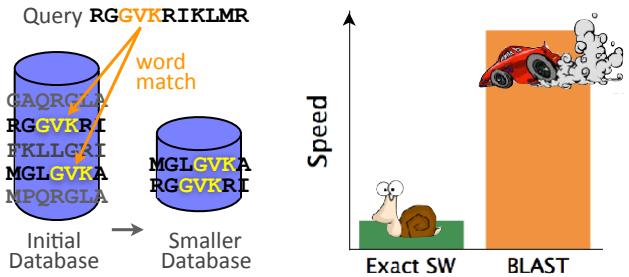
85

Rapid, heuristic versions of Smith–Waterman: BLAST

- BLAST (Basic Local Alignment Search Tool) is a simplified form of Smith-Waterman (SW) alignment algorithm. It is faster because it is **fast** and **easily parallelizable**.
 - BLAST finds regions of local sequence similarity.
 - BLAST uses a “**word pair**” match to sequence pairs that contain an initial **word pair** match”
- “The central idea of the BLAST algorithm is to confine attention to sequence pairs that contain an initial word pair match” Altschul et al. (1990)*
- Altschul et al. (1990) suggests some sensitivity in exchange for speed
- In contrast to SW, BLAST is not guaranteed to find optimal alignments

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- BLAST uses this pre-screening heuristic approximation resulting in an approach that is about 50 times faster than the Smith-Waterman algorithm



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How BLAST works

- Four basic phases
 - **Phase 1:** compile a list of query word pairs ($w=3$)

RGGVKRI Query sequence
RGG
GGV
GVK
VKR
KRI

generate list of **w=3 words** for query

88

- **Phase 2:** expand word pairs to include those similar to query (defined as those above a similarity threshold to original word, i.e. match scores in substitution matrix)

RGGVKRI Query sequence
RGG RAG RIG RLG ...
GGV GAV GTV GCV ...
GVK GAK GIK GGK ...
VKR VRR VHR VER ...
KRI KKI KHI KDI ...

extend list of words similar to query

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Blast

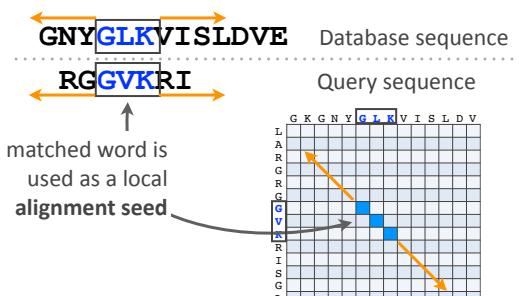
- **Phase 3:** a database is scanned to find sequence entries that match the compiled word list

GNYGLKVISLDVE Database sequence
RGGVKRI Query sequence
RGG RAG RIG RLG ...
GGV GAV GTV GCV ...
GVK GLK GIK GGK ...
VKR VRR VHR VER ...
KRI KKI KHI KDI ...

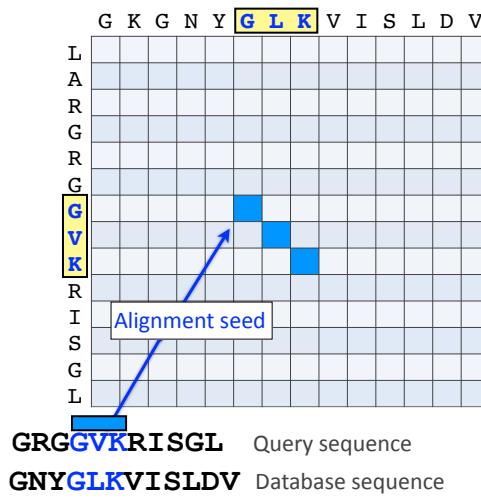
search for perfect matches in the database sequence

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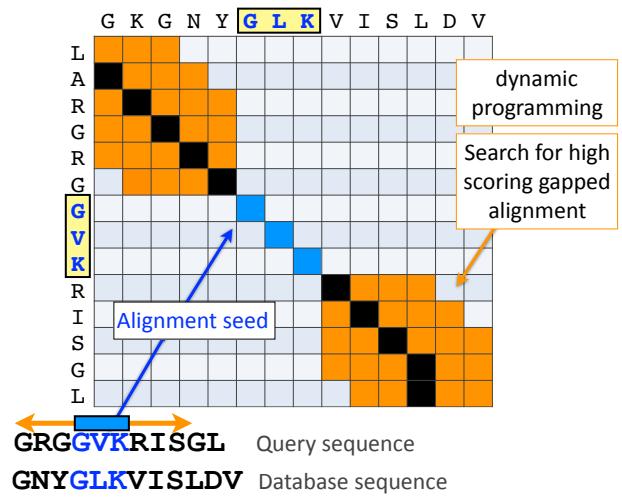
- **Phase 4:** the initial database hits are extended in both directions using dynamic programming



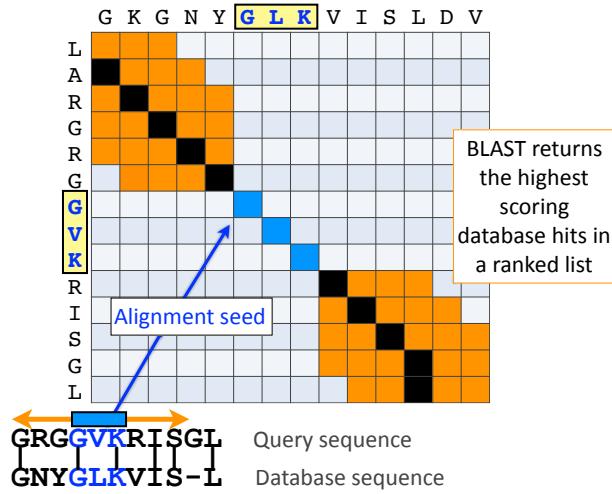
91



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BLAST output

- BLAST returns the highest scoring database hits in a ranked list along with details about the target sequence and alignment statistics

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
Kinesin-14 heavy chain [Danio rerio]	595	88%	0	78%	XP_00320703
hypothetical protein EGK_18589	48.2	40%	0.03	32%	ELK35081.1
mKIAA4102 protein [Mus musculus]	42.7	38%	3.02	24%	EHH28205.1

95

Statistical significance of results

- An important feature of BLAST is the computation of statistical significance for each hit. This is described by the **E value** (expect value)

Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
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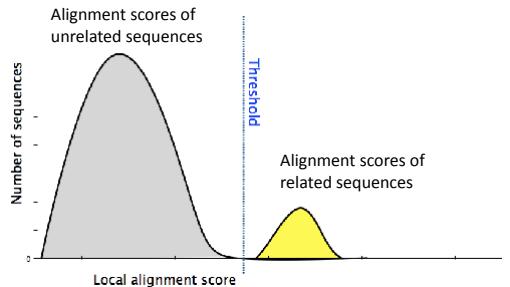
96

BLAST scores and E-values

- The **E value** is the **expected** number of hits that are as good or better than the observed local alignment score (with this score or better) if the query and database are **random** with respect to each other
 - i.e. the number of alignments expected to occur by chance with equivalent or better scores
- Typically, only hits with E value **below** a significance threshold are reported
 - This is equivalent to selecting alignments with score above a certain score threshold

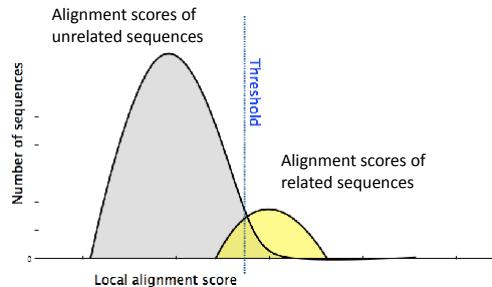
97

- Ideally, a threshold separates all query related sequences (yellow) from all unrelated sequences (gray)



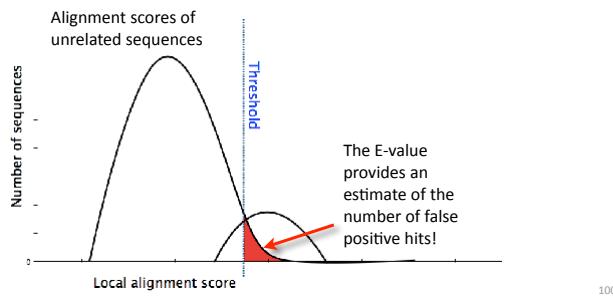
98

- Unfortunately, often both score distributions overlap
 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



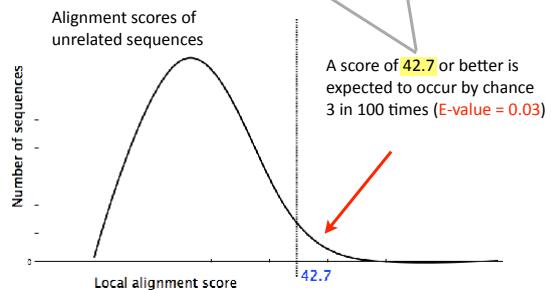
99

- Unfortunately, often both score distributions overlap
 - The E value describes the expected number of hits with a score above the threshold if the query and database are unrelated



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Description	Max score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	100%	0	100%	NP_004512.1
Kif5b protein [Mus musculus]	676	100%	0	98%	AAA20133.1
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hypothetical protein EGK_18589	42.7	40%	0.03	32%	ELK35081.1



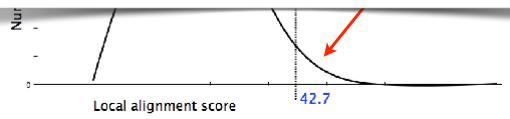
101

Description	Max score	Total score	Query cover	E value	Max ident	Accession
kinesin-1 heavy chain [Homo sapiens]	677	677	100%	0	100%	NP_004512.1

In general E values < 0.005 are usually significant.

To find out more about E values see: "The Statistics of Sequence Similarity Scores" available in the help section of the NCBI BLAST site:

<http://www.ncbi.nlm.nih.gov/blast/tutorial/Altschul-1.html>



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Practical database searching with BLAST

The screenshot shows the NCBI BLAST Home Page with several search options:

- Basic BLAST**: Search a nucleotide database using a nucleotide query.
- blastn**: Search a nucleotide database using a nucleotide query.
- blastp**: Search protein database using a protein query.
- blastx**: Search protein database using a translated nucleotide query.
- tblastn**: Search nucleotide database using a protein query.
- tblastx**: Search translated nucleotide database using a translated nucleotide query.

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Practical database searching with BLAST

- There are four basic components to a traditional BLAST search
 - (1) Choose the sequence (query)
 - (2) Select the BLAST program
 - (3) Choose the database to search
 - (4) Choose optional parameters
- Then click “BLAST”

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Step 1: Choose your sequence

- Sequence can be input in FASTA format or as accession number

The screenshot shows the NCBI Protein page with a sequence entry for hemoglobin subunit beta [Homo sapiens]. The sequence is:

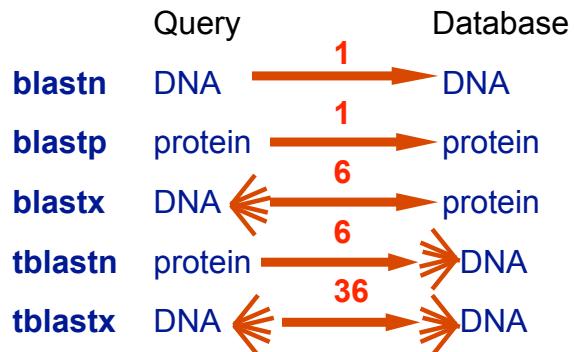
```

>NP_0010091 hemoglobin subunit beta [Homo sapiens]
MGI:1404349I refNP_0010091
Signal: 1
Analyze this sequence
Run BLAST
Identify Conserved Domains
Find this sequence

```

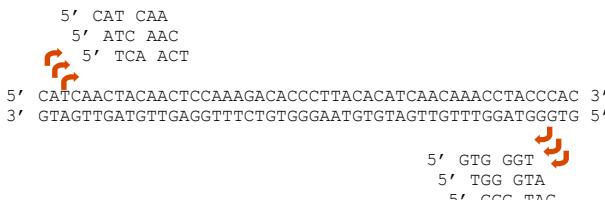
106

Step 2: Choose the BLAST program



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DNA potentially encodes six proteins



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The screenshot shows the NCBI Protein BLAST search interface with the following parameters:

- Enter accession number(s), file(s), or FASTA sequence(s):** NP_0010091
- Or, upload file:** Choose file / net bio extracted
- Job Title:** Enter a descriptive title for your BLAST search.
- Choose Search Set:** Non-redundant protein sequences (nr)
- Database:** Enter organism common name, binomial, or tax id. Only 2D top taxa will be shown.
- Exclude:** Mito (XMPP) Unaligned/environmental sample sequences.
- Entrez Query:** Enter an Entrez query to limit search.
- Program Selection:** blasp (protein-protein BLAST)
- Algorithm parameters:** Search database Non-redundant protein sequences (nr) using Blasp (protein-protein BLAST)

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Step 3: Choose the database

nr = non-redundant (most general database)
 dbest = database of expressed sequence tags
 dbsts = database of sequence tag sites
 gss = genomic survey sequences

Genomic plus Transcript

- Non-redundant protein sequences (nr)
- Expressed sequence tags (dbEST)
- Sequence tag sites (dbSTS)
- Genomic survey sequences (gss)
- Protein Data Bank (PDB)
- Environmental samples (env_nr)

nucleotide databases

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Non-redundant protein sequences (nr)

- Non-redundant protein sequences (nr)
- RefSeq protein sequences (refseq_protein)
- SwissProt protein sequences (swissprot)
- Published protein sequences (pub)
- Protein Data Bank (PDB)
- Environmental samples (env_nr)

protein databases

Organism

Entrez

Settings!

BLAST

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Step 4a: Select optional search parameters

Expect threshold: 100

Word size: 3

Scoring matrix: BLOSUM62

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Step 4: Optional parameters

You can...

- choose the organism to search
- change the substitution matrix
- change the expect (E) value
- change the word size
- change the output format

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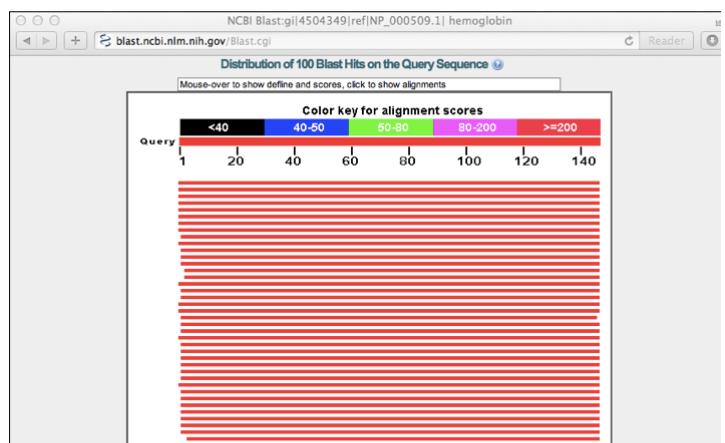
Results page

Query ID: g14504349[refNP_000509.1] hemoglobin

Database Name: nr

Program: BLASTP 2.2.27+ > Citation

Further down the results page...



Further down the results page...

NCBI Blast:gi|4504349|ref|NP_000509.1| hemoglobin

blast.ncbi.nlm.nih.gov | Blast.cgi | Reader

Sequences producing significant alignments:

Select: All None Selected: 0

All Alignments Download v GenPept Graphics Distance tree of results Multiple alignment

	Description	Max score	Total score	Query cover	E value	Max ident	Accession
hemoglobin beta [synthetic construct]	301	301	100%	9e-103	100%	AAX37051.1	
hemoglobin beta [synthetic construct]	301	301	100%	1e-102	100%	AAX95567.1	
hemoglobin subunit beta [Homo sapiens] >ref XP_508242.1 PREDICTED: hemoglobin s	301	301	100%	1e-102	100%	NP_000509.1	
RedName: Full=Hemoglobin subunit beta; AltName: Full=Beta-globin; AltName: Full=Hemoglobin beta	300	300	100%	4e-102	99%	P02042.2	
beta globin chain variant [Homo sapiens]	299	299	100%	5e-102	99%	AAN84548.1	
beta globin [Homo sapiens] >gb AAZ39781.1 beta globin [Homo sapiens] >gb AAZ39781.1	299	299	100%	5e-102	99%	AAZ39780.1	
beta globin [Homo sapiens]	299	299	100%	5e-102	99%	ACU56984.1	
hemoglobin beta chain [Homo sapiens]	299	299	100%	6e-102	99%	AAD19896.1	
Chain B. Structure Of Hemoglobin In The Deoxy Quaternary State With Ligand Bound At	298	298	99%	9e-102	100%	1COH_B	
hemoglobin beta subunit variant [Homo sapiens] >gb AAA88054.1 beta-globin [Homo sapiens]	298	298	100%	1e-101	99%	AOA04889.1	
Chain B. Human Hemoglobin D Los Angeles: Crystal Structure >pdBfYRSQI Chain D_H	298	298	99%	2e-101	99%	ZYRS_B	
Chain B. High-Resolution X-Ray Study Of Deoxy Recombinant Human Hemoglobins Syn	297	297	99%	3e-101	99%	1DXU_B	
Chain B. Analysis Of The Crystal Structure, Molecular Modeling And Infrared Spectroscop	297	297	99%	3e-101	99%	1HDB_B	

Further down the results page...

Different output formats are available

NCBI Blast:gi|4504349[ref|NP_000509.1] hemoglobin

blast.ncbi.nlm.nih.gov/Blast.cgi Reader

BLAST® Basic Local Alignment Search Tool My NCBI [Sign In | Register]

Home Recent Results Saved Strategies Help

► NCBI BLAST! blast suite/ Formatting Results - FVGUTMP219

Edit and Resubmit Save Search Strategies **Formatting options** > Download Change the result display back

You Tube Learn about the enhanced report

Formatting options

Show Alignment as **HTML** Old View Reset form to defaults

Alignment View Query-anchored with letters for identities

Display Graphical Overview Sequence Retrieval NCBI-gi

Masking Character: Lower Case Color: Grey

Limit results Descriptions: 50 Graphical overview: 50 Alignments: 50

Organism Type common name, binomial, taxid, or group name. Only 20 top taxa will be shown.

[Enter organism name or ID—completions will be suggested] Exclude

Entrez query:

Expect Min: Expect Max:

Percent Identity Min: Percent Identity Max:

Format for PSI-BLAST with inclusion threshold:

E.g. Query anchored alignments

NCBI Blast:gi 4504349 [ref NP_000509.1] hemoglobin	
Query	MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
RAAX37051	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAX29557	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
NP_000509	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
P02242	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAQ41848	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAI32945	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
ACU56984	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAO19566	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
ICON_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAF00489	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
ZYRS_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
PHBZ_HUMAN	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
I1H0P_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
IDXV_B	2 HLTLEEKESATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
J3KMF_C	2 HLTLEEKESATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
AAI68978	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
IN0P_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
I1K1K_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
QAN3230	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
XP_002822173	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
LYB5_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
LYE0_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
O1010_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
CAA23752	1 MVHLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
LYE1_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
LYE2_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
JAO0_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
I1HBS_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
JABY_B	1 MHLTTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK
ICMY_B	1 VHLLTEEEKSATAVLGKVNKVDEVGEGALGRLLVYFWPTQRFFSFSGDLSTPDAVGNFK

... and alignments with dots for identities

Query	1	MVIVLPEEKSAAVTLAHOVYRVEVKGALGRLVLYVPPWGRPFPSQDLSPTPDAVHGNFK	60
JAX37051	1	60
JAX29557	1	60
NP_000509	1	60
P02024	1	60
JANH84548	1	60
JAX36780	1X.....	60
ACUS56188	1X.....	60
AN019626	1L.....	60
JCOB_B	1	59
AAF004189	1	60
ZYRS_B	1	59
JDXU_B	1	M.....	59
JDZB_B	1	59
JDXV_B	2	59
JKMF_C	2	59
AAL68378	1	60
JNQP_B	1X.....	59
JKIK_B	1X.....	59
RAAS4320	1V.....	60
XP_002822173	1	59
LYBS_	1	59
LYBD_	1	M.....	59
IOLO_B	1	M.....	59
CAA23759	1V.....X.....	60
LYE2_B	1	M.....	59
LYSF_B	1	M.....	59
IA00_B	1	M.....Y.....	58

Common problems

- Selecting the wrong version of BLAST
 - Selecting the wrong database
 - Too many hits returned
 - Too few hits returned
 - Unclear about the significance of a particular result - are these sequences homologous?

How to handle too many results

- Focus on the question you are trying to answer
 - select “refseq” database to eliminate redundant matches from “nr”
 - Limit hits by organism
 - Use just a portion of the query sequence, when appropriate
 - Adjust the expect value; lowering E will reduce the number of matches returned

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How to handle too few results

- Many genes and proteins have no significant database matches
 - remove Entrez limits
 - raise E-value threshold
 - search different databases
 - try scoring matrices with lower BLOSUM values (or higher PAM values)
 - use a search algorithm that is more sensitive than BLAST (*e.g.* PSI-BLAST or HMMer)

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Side note: Scoring matrices

- A substitution matrix contains values proportional to the probability that amino acid i mutates into amino acid j for all pairs of amino acids
- Substitution matrices are constructed by assembling a large and diverse sample of verified pairwise alignments (or multiple sequence alignments) of amino acids.
- Substitution matrices should reflect the probabilities of mutations occurring through a period of evolution
- The two major types of substitution matrices are **PAM** and **BLOSUM**

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BLOSUM62 is the default BLASTp scoring matrix

- BLOSUM matrices are based on short, ungapped blocks of conserved amino acid sequences from multiple alignments
 - members of a block that have a most X percent sequence identity to each other are used to generate a BLOSUMX matrix
 - For example, using a cutoff of 62% identity will generate the BLOSUM62 matrix
- PAM matrices are similar but built from multiple alignments where amino acid substitutions are at rate of 1% (PAM 1)
 - Matrix multiplication is used generate higher PAM matrices
 - PAM3 = (PAM1 x PAM1 x PAM1) etc...

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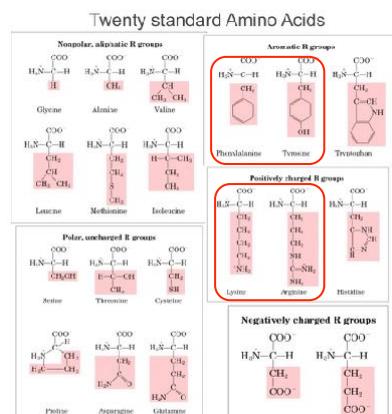
By default BLASTp Match scores come from the BLOSUM62 matrix

C	S
S	-1 4
T	-1 1 5
P	-3 -1 -1 7
A	0 1 0 -1 4
G	-3 0 -2 -2 0 6
N	-3 0 0 -2 0 4
D	-3 0 -1 -1 -2 1 6
E	-4 0 -1 -1 -1 -2 0 5
Q	-3 0 -1 -1 -1 -2 0 0 2 5
H	-3 1 -2 -2 -2 2 1 0 8
R	-3 -1 -1 -2 -1 -2 0 -2 0 1 0 5
K	-3 0 -1 -1 -1 -2 0 -1 0 1 -1 3
M	-1 -1 -1 -2 -1 -3 -2 -3 -2 0 -2 -1 -1 5
I	-1 -2 -1 -3 -1 -4 -3 -3 -3 -3 -3 -3 -3 1 4
L	-1 -2 -1 -3 -1 -4 -3 -4 -3 -2 -3 -2 -2 2 2 4
V	-1 -2 0 -2 0 -3 -3 -2 -2 -3 -3 -2 1 3 1 1
F	-2 -2 -2 -4 -2 -3 -3 -3 -3 -3 -1 -3 -3 0 0 0 1 6
Y	-2 -2 -2 -3 -2 -3 -2 -1 1 2 -2 -1 -1 -1 3 7
W	-2 -3 -2 -4 -3 -2 -4 -3 -2 -2 -3 -3 -1 -3 -2 -3 1 2 1 1
C	S T P A G N D E Q H R K M I L V F Y W

Note. Some amino acid mismatches have positive scores – highlighted in red

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Protein scoring matrices reflect the properties of amino acids



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Two problems standard BLAST cannot solve

- Use human beta globin as a query against human RefSeq proteins, and blastp does not “find” human myoglobin
 - This is because the two proteins are too distantly related
 - PSI-BLAST at NCBI as well as hidden Markov models (HMMs) easily solve this problem
- How can we search using 10,000 base pairs as a query, or even millions of base pairs?
 - Many BLAST-like tools for genomic DNA are now available such as Megablast

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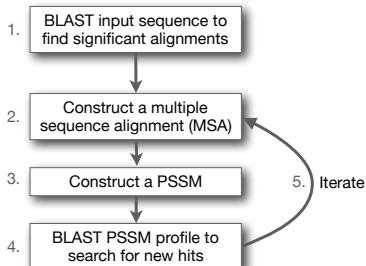
PSI-BLAST: Position specific iterated BLAST

- The purpose of PSI-BLAST is to look deeper into the database for matches to your query protein sequence by employing a scoring matrix that is customized to your query
- PSI-BLAST constructs a multiple sequence alignment from the results of a first round BLAST search and then creates a “profile” or specialized **position-specific scoring matrix (PSSM)** for subsequent search rounds

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PSI-BLAST: Position-Specific Iterated BLAST

- Many proteins in a database are too distantly related to a query to be detected using standard BLAST. In many other cases matches are detected but are so distant that the inference of homology is unclear. Enter the more sensitive PSI-BLAST



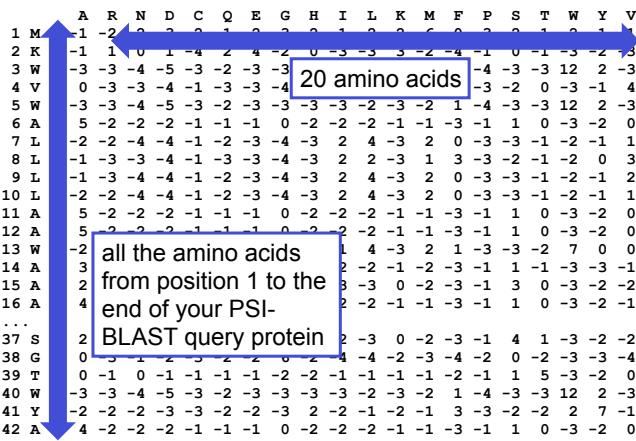
130

Inspect the blastp output to identify empirical “rules” regarding amino acids tolerated at each position

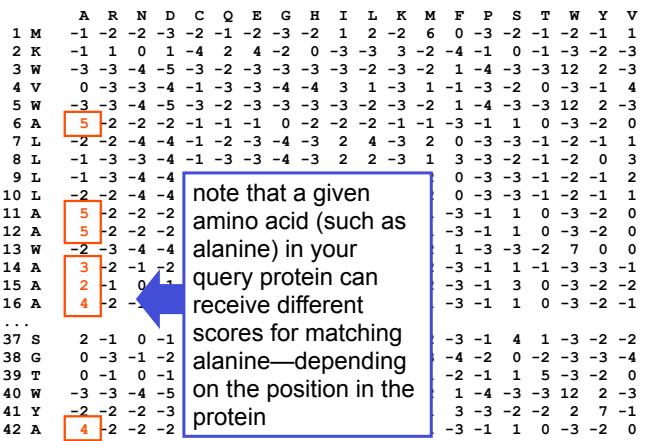
730496	66	F TVDENQHSATAKGRVRLFLNNWDVCADMGSTDTTEDP AKFKMKYVGAVASFLQKGNDHH
203679	63	FSVDEKGHMSATAKGRVRLLSNWEVCADMVGFTDTEDP AKFKMKYVGAVASFLQKGNDHH
206589	34	FSVDEKGHMSATAKGRVRLLSNWEVCADMVGFDTDTEDP AKFKMKYVGAVASFLQKGNDHH
2136812	2	MSATAKGRVRLLNWUDVCADMVGFTDTEDP AKFKMKYVGAVASFLQKGNDHH
132408	65	FKIEDNGKTTATAKGRVFLDKLELCANHVGIFTETNDPAKF KMKYVGAVASFLQKGNDHH
267584	44	FSVDESGKVTATAKGRVILLNWEMCANHFGTETDTPD AKFKMKYVGAVASFLQKGNDHH
267585	44	FSVDGSRKVTATAKGRVILLNWEMCANHFGTETDTPD AKFKMKYVGAVASFLQKGNDHH
8777608	63	FTIHEDGANTATAKGRVILLNWEMCADHMATEFTTPD AKFKMKYVGAVASFLQKGNDHH
6687453	60	FKVEEDGTMATAKGRVILLNWEMCANHFGTFETDTPD AKFKMKYVGAVASFLQKGNDHH
10697027	61	FKVQEDGTMATAKGRVILLNWEMCANHFGTETDPEP ARFKMKYVGAAAYLQTGYDDH
13645517	1	IVGTTDTEDP AKFKMKYVGAVASFLQKGNDHH
13925316	38	FSVDSGSRKMTATAQGRVILLNWEMCANHFGTETDTPD AKFKMKYVGAVASFLQKGNDHH
131649	65	TVVEEDGTMATAKGRVFLFGFWVICANDAAQYTDPTTP AKWVITYQGLASVYLSSGGDNUT

R,I,K C D,E,T K,R,T N,L,Y,G

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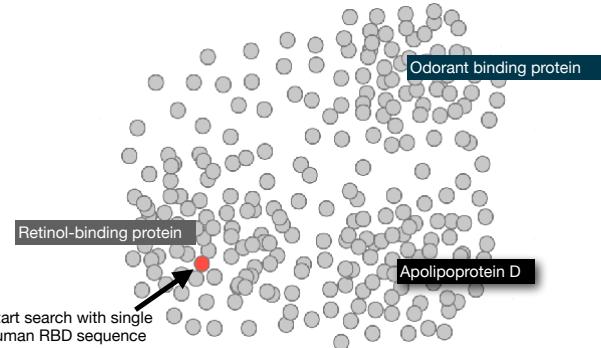
133

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	
1	M	-1	-2	-2	-3	-2	-1	-2	-3	-2	1	2	-2	6	0	-3	-2	-1	-2	-1	1
2	K	5	-2	-2	-2	-1	-1	0	-2	-2	-2	-1	-1	-3	-1	1	0	-3	-2	0	
3	W	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
4	V	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
5	W	-1	-3	-3	-4	5	-2	-2	-4	-4	1	2	-2	6	0	-3	-3	-1	-2	-1	1
6	A	-2	-2	-4	-4	5	-2	-2	-2	-2	-1	-1	-1	-3	-1	1	0	-3	-2	0	
7	L	-2	-2	-4	-4	-1	-2	-3	-4	-3	2	4	-3	2	0	-3	-3	-1	-2	-1	1
8	L	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
9	L	-1	-3	-3	-4	-1	-3	-3	-4	-3	2	2	-3	1	3	-3	-2	-1	-2	0	3
10	I	-2	-2	-4	-4	-1	-2	-2	-2	-2	-1	-1	-1	-3	-1	-2	-1	-1	-2	-1	1
11	A	5	-2	-2	-2	-1	-2	-2	-2	-2	-1	-1	-1	-3	-1	1	0	-3	-2	0	
12	A	5	-2	-2	-2	-1	-2	-2	-2	-2	-1	-1	-1	-3	-1	1	0	-3	-2	0	
13	W	-2	-3	-4	-4	-1	-2	-3	-4	-3	2	2	-3	1	3	-3	-2	7	0	0	
14	A	3	-2	-1	-2	-1	-2	-2	-2	-2	-1	-1	-1	-3	-1	1	-1	-3	-3	-1	
15	A	2	-1	0	-1	-1	-2	-2	-2	-2	-1	-1	-1	-3	-1	3	0	-3	-2	-2	
16	A	4	-2	-2	-2	-2	-2	-2	-2	-2	-1	-1	-1	-3	-1	1	0	-3	-2	0	
...																					
37	S	2	-1	0	-1	-1	-2	-2	-2	-2	-1	-1	-1	-3	-2	-2	-2	-2	-2	0	
38	G	0	-3	-1	-2	-1	-2	-2	-2	-2	-1	-1	-1	-3	-2	-3	-3	-4	-4	0	
39	T	0	-1	0	-1	-1	-2	-2	-2	-2	-1	-1	-1	-3	-2	-2	-2	-2	-2	0	
40	W	-3	-3	-4	-5	-1	-2	-3	-4	-5	1	4	-3	-3	12	2	2	-3	-3	-2	0
41	Y	-2	-2	-2	-2	-3	-2	-2	-2	-2	3	-3	-2	-2	2	7	-1	-3	-2	-2	
42	A	4	-2	-2	-2	-2	-2	-2	-2	-2	-3	-1	-1	-3	-1	1	0	-3	-2	0	

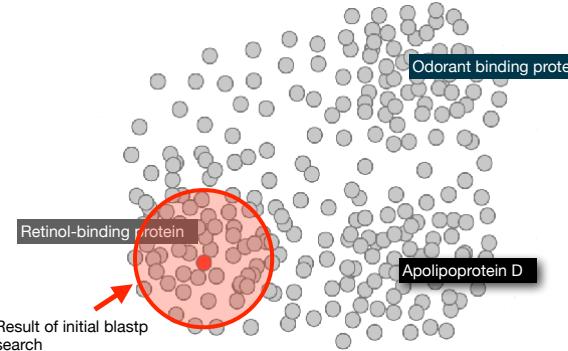
The PSI-BLAST PSSM is essentially a query customized scoring matrix that is more sensitive than PAM or BLOSUM.

note that a given amino acid (such as alanine) in your query protein can receive different scores for matching alanine—depending on the position in the protein

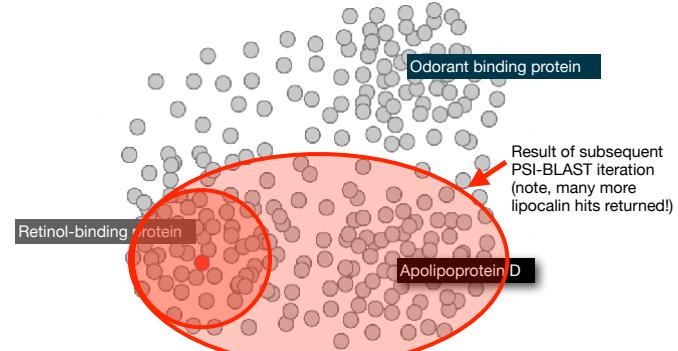
134



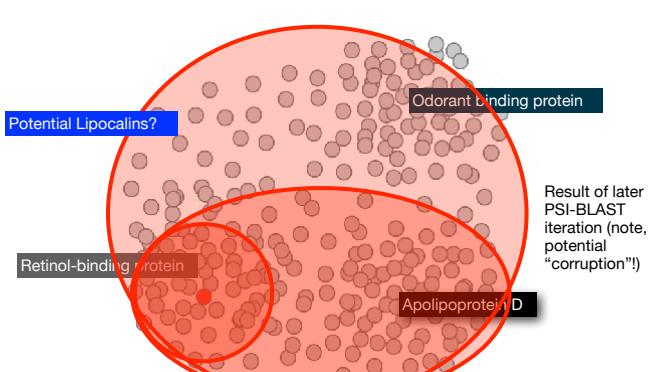
135



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PSI-BLAST returns dramatically more hits

- The search process is continued iteratively, typically about five times, and at each step a new PSSM is built
 - You must decide how many iterations to perform and which sequences to include!
 - You can stop the search process at any point - typically whenever few new results are returned or when no new "sensible" results are found

Iteration	Hits with E < 0.005	Hits with E > 0.005
1	34	61
2	314	79
3	416	57
4	432	50
5	432	50

Human retinol-binding protein 4 (RBP4; P02753) was used as a query in a PSI-BLAST search of the RefSeq database.

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The screenshot shows the HHMMR3 website. At the top, there's a header with the title "HHMMR3" and a sub-header "biosequence analysis using profile hidden Markov models". Below the header is a navigation bar with links for Home, Search, Results, Software, Help, and About. To the right of the header, there's a search bar with the placeholder "Search Google" and a logo for "HHMI janelia farm research campus". The main content area features a large heading "HHMMR3: a new generation of sequence homology search software". Below it, a paragraph explains what HHMMR3 is used for. A section titled "Compared to BLAST, FASTA, and other sequence alignment and database search tools based on older scoring methodology, HHMMR aims to be significantly more accurate and more able to detect remote homologs because of the strength of its underlying mathematical models. In the past, this strength came at significant computational expense, but in the new HHMMR3 project, HHMMR is now essentially as fast as BLAST." Another section discusses the evolution of the software, mentioning that earlier releases were command line only and now offer web-based interfaces. It also notes the current version is 3.0 (28 March 2010). A sidebar on the right is titled "Download HHMMR" and includes a "v3.0" badge, release notes, and download links. It also has sections for "Alternative Download Options" and "Search".

HMMER

<http://hmmer.janelia.org/search>


HMMER
biosequence analysis using profile hidden Markov models
HHMI
janelia farm
research campus

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[Results](#)
[Software](#)
[Help](#)
[About](#)

[HMMER](#)
[hmmer](#)
[hmmscan](#)

protein sequence vs protein sequence database
[Advanced](#)

Paste in your sequence or use the [example](#)

```
>sp|Q14807|NP_212134_HUMAN
MAAGCSTCRRRDAASASAAAGACRLSISCATTRPPARVAVVSLRFYVGCTACA
SDPVCPRHDSCSLLVAVWVHGETLVYGFADYFSTGCGCCTGCGCCTGCGCCTG
QEVVLDLPPGGGVWEDORGANIPCSQPSISSLADMVHIFPASINRIVLCAIRLN
QEVVLDLPPGGGVWEDORGANIPCSQPSISSLADMVHIFPASINRIVLCAIRLN
QEVVLDLPPGGGVWEDORGANIPCSQPSISSLADMVHIFPASINRIVLCAIRLN
FAASKEVIRPFTNESELCHALGPWLSKELLCPEARCAFPEEEICBEPENNAAPAF
AASUNAHFHRVNGEERDQIESTLAEFHFADEWVWVQNTTEAHGEGIDENEXS
AASUNAHFHRVNGEERDQIESTLAEFHFADEWVWVQNTTEAHGEGIDENEXS
AASUNAHFHRVNGEERDQIESTLAEFHFADEWVWVQNTTEAHGEGIDENEXS
```

Submit
Reset

Comments or questions on the site? Send a mail to hmmer@janelia.hhmi.org
 Howard Hughes Medical Institute

[Follow](#) 

The screenshot shows the HMMER web interface with the URL <http://hmmer.janelia.org/results/score/9924f9AC-FE85-11E0-A304-28C998A7913>. The main content area displays a chart titled "Pfam Domains" showing the distribution of significant hits across various domains. Below the chart, a "Show hit details" link is visible. At the bottom, there is a table titled "Query Matches (5109)" with columns for Target, Description, Species, E-value, and Alignments (show all). Several rows of data are listed, including entries for kinase-like protein KIF22 and kinase-like protein KIF22-like.

Target	Description	Species	E-value	Alignments (show all)
I2397973569	kinase family member 22	synthetic construct	0.0e+00	show
D039364621	kinase-like protein KIF22	Homo sapiens	0.0e+00	show
30384621	Homo sapiens kinase-like 4	synthetic construct	0.0e+00	show
I2396511369	kinase family member 22	synthetic construct	0.0e+00	show
I8905331262	unnamerd protein product	synthetic construct	0.0e+00	show
6261984242	kinase family member 22 variant	Homo sapiens	0.0e+00	show
I2394311317	PREDICTED: kinase family member 22 isoform 2	Predictor	0.0e+00	show
76603117	PREDICTED: kinase family member 22 isoform 2	Predictor	0.0e+00	show
3322604889	PREDICTED: kinase-like protein KIF22-like isoform 1	Narracusa luciferans	0.0e+00	show
297263749B	PREDICTED: hputential protein LOC_706491 isoform 3	Macaca fasciata	0.0e+00	show
296219941B	PREDICTED: LOW QUALITY PROTEIN: kinase-like protein KIF22-like	Callithrix jacchus	0.0e+00	show
009130300	PREDICTED: kinase-like protein KIF22-like	Callicebus jacchus	0.0e+00	show
J352640129	PREDICTED: kinase-like protein KIF22-like	Sus scrofa	0.0e+00	show
2212461598	unnamerd protein product	Homo sapiens	0.0e+00	show
3293461089	unnamerd protein product	Mus musculus	0.0e+00	show

HHMMR

biosequence analysis using profile hidden Markov models

HHMI
janelia farm
research campus

Score Taxonomy Domain Download

Job: 9924F9AC-FEB5-11E0-A304-2B0C998A/923
Started: 2011-10-24 23:01:15
Alignments: 1
HMMER Options: -E 1 --domE 1 --incE 0.01 --incmE 0.03 --max BLOSUM62 --pextland 0.4 --ppen 0.02 --seqdb nr

Format

Fasta
Download file containing hits from your search as a gzipped FASTA file.

Full length Fasta
A zipped file containing the full length sequences as aligned search hits.

Aligned Fasta
A gzipped file containing aligned significant search hits in FASTA for nr.

Stockholm
Download an alignment of significant hits as a standard STOCKHOLM file.

Text
A plain text file containing the Fasta alignments and scores.

XML
An XML file to format for machine parsing of the data.

HMM
Profile HMM downloads are not available.

Download Reset

Summary

- Alignment basics
 - Why compare biological sequences?
 - Homologue detection
 - Orthologs, paralogs, similarity and identity
 - Sequence changes during evolution
 - Alignment view: matches, mismatches and gaps
 - Pairwise sequence alignment methods
 - Brute force alignment
 - Dot matrices
 - Dynamic programming
(global vs local alignment)
 - Rapid heuristic approaches
 - BLAST
 - Practical database searching
 - BLAST, PSI-BLAST and HMM approaches