

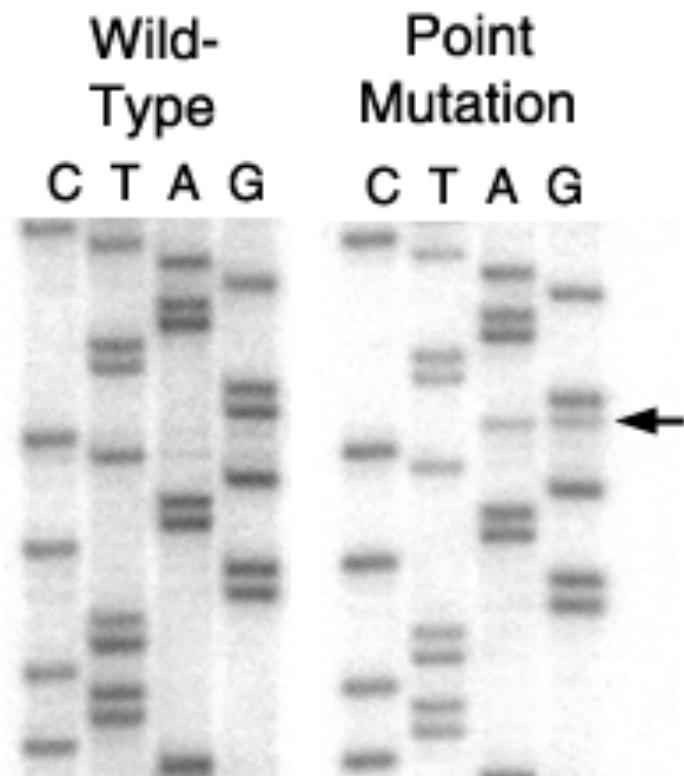
High throughput sequencing methods in systems biology

Bioinformatics 524/525

Module 3, Lecture 2

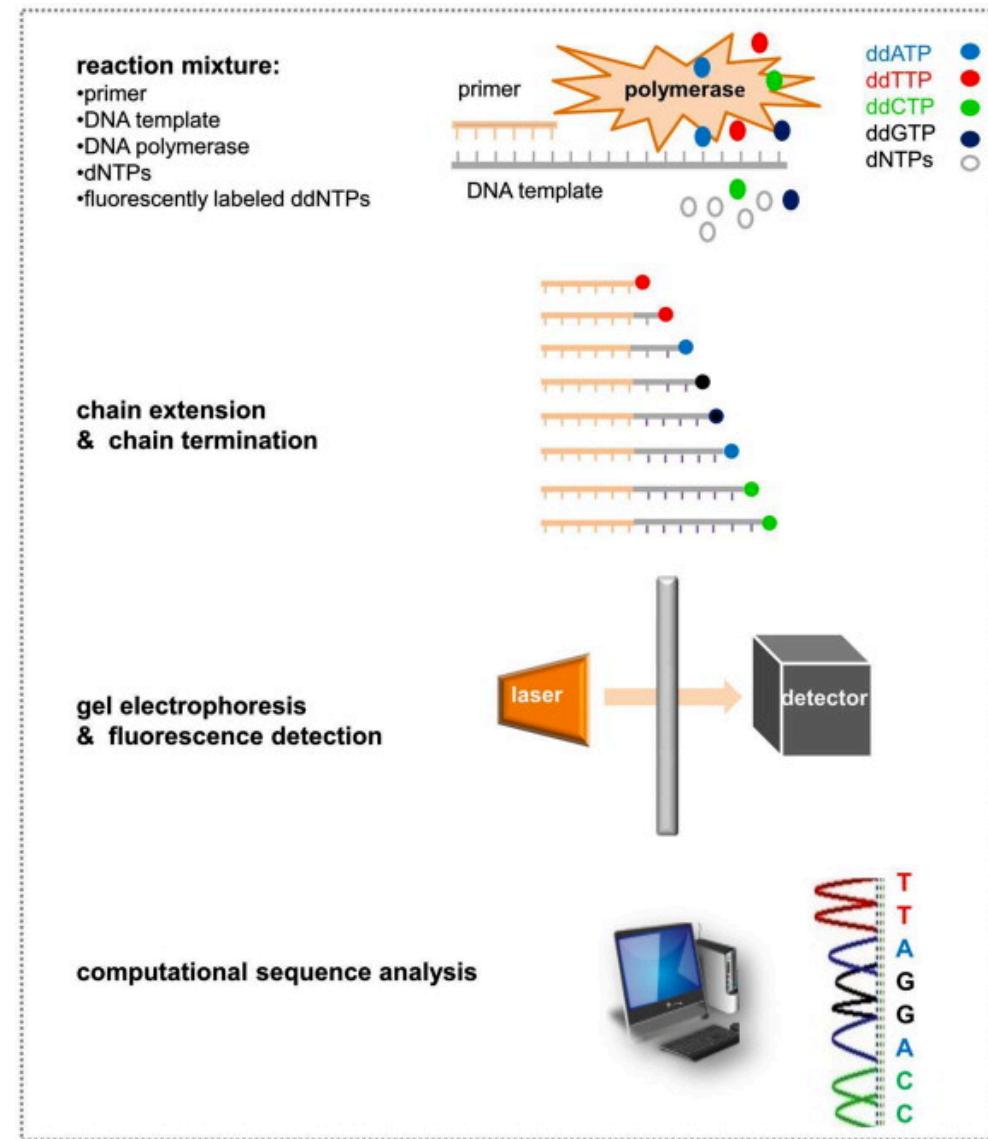
3/28/2017

In the beginning, there were sequencing gels...



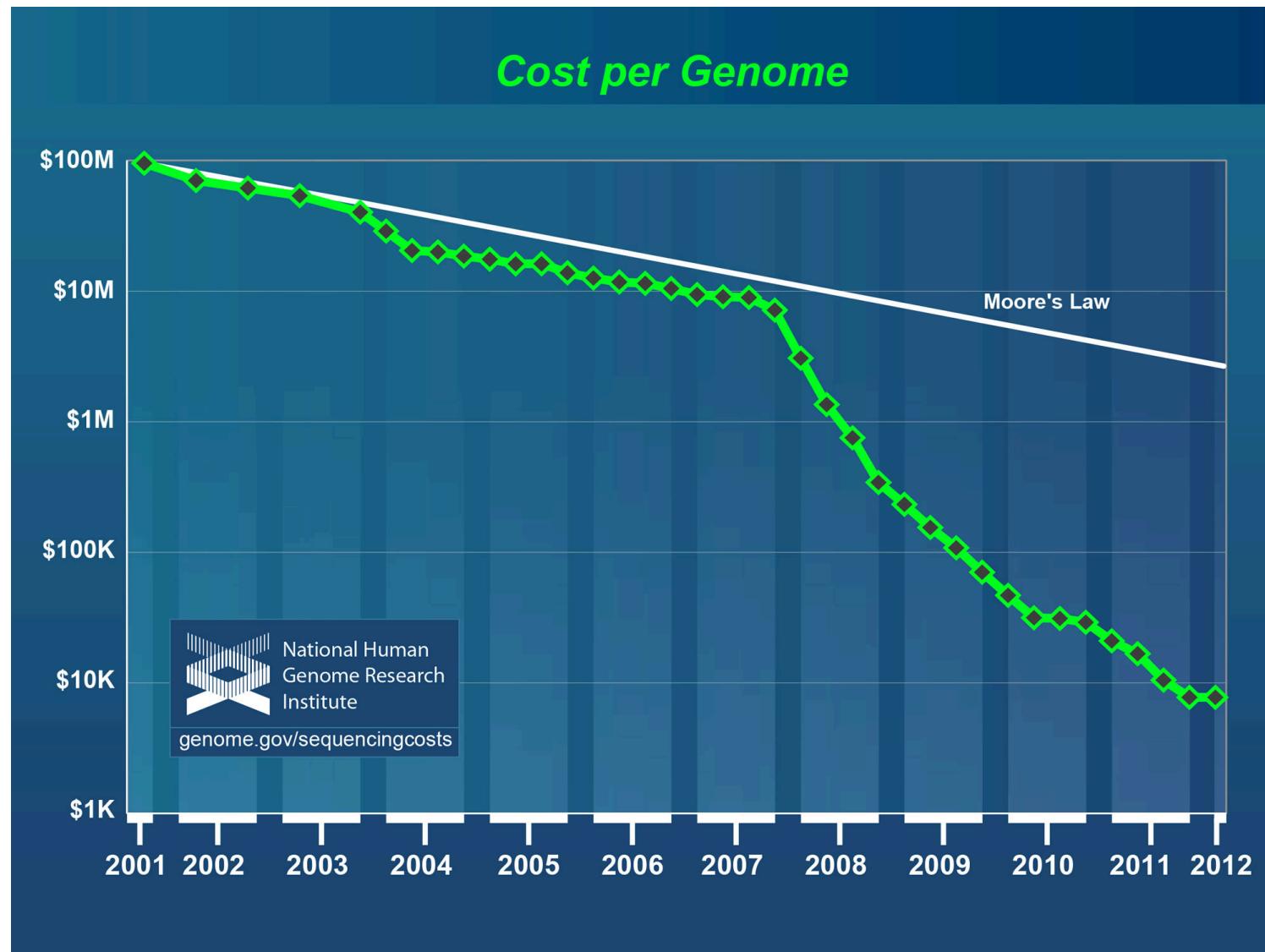
(via Victoria Schulman)

Then there was Sanger sequencing...



From P. Zhang, A. Seth, and H. Fernandes,
Pathobiology of Human Disease

... and then there was *Next Gen*



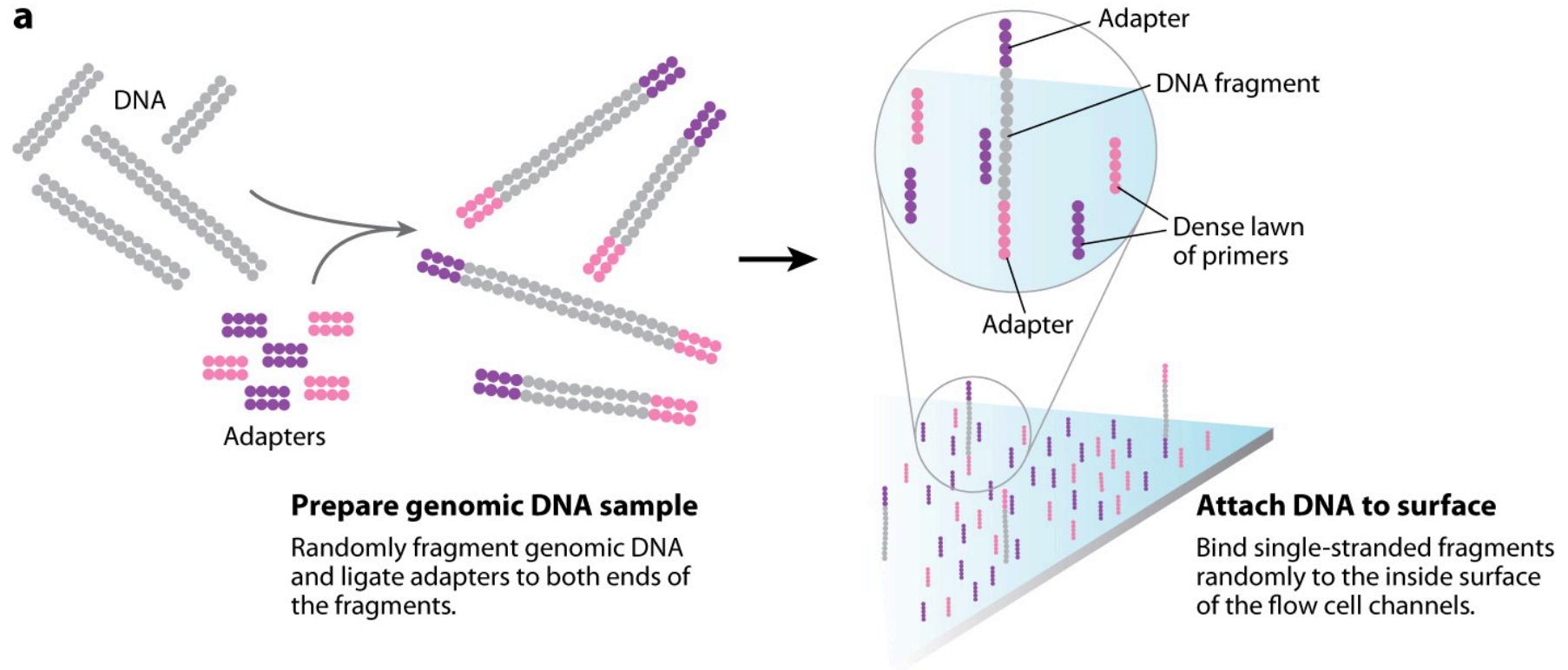
Outline

- Summary of NGS technologies (sequencing and applications)
- Introduction to NGS data analysis
- Commonly available databases
- Workflow integration and making use of existing NGS data

Outline

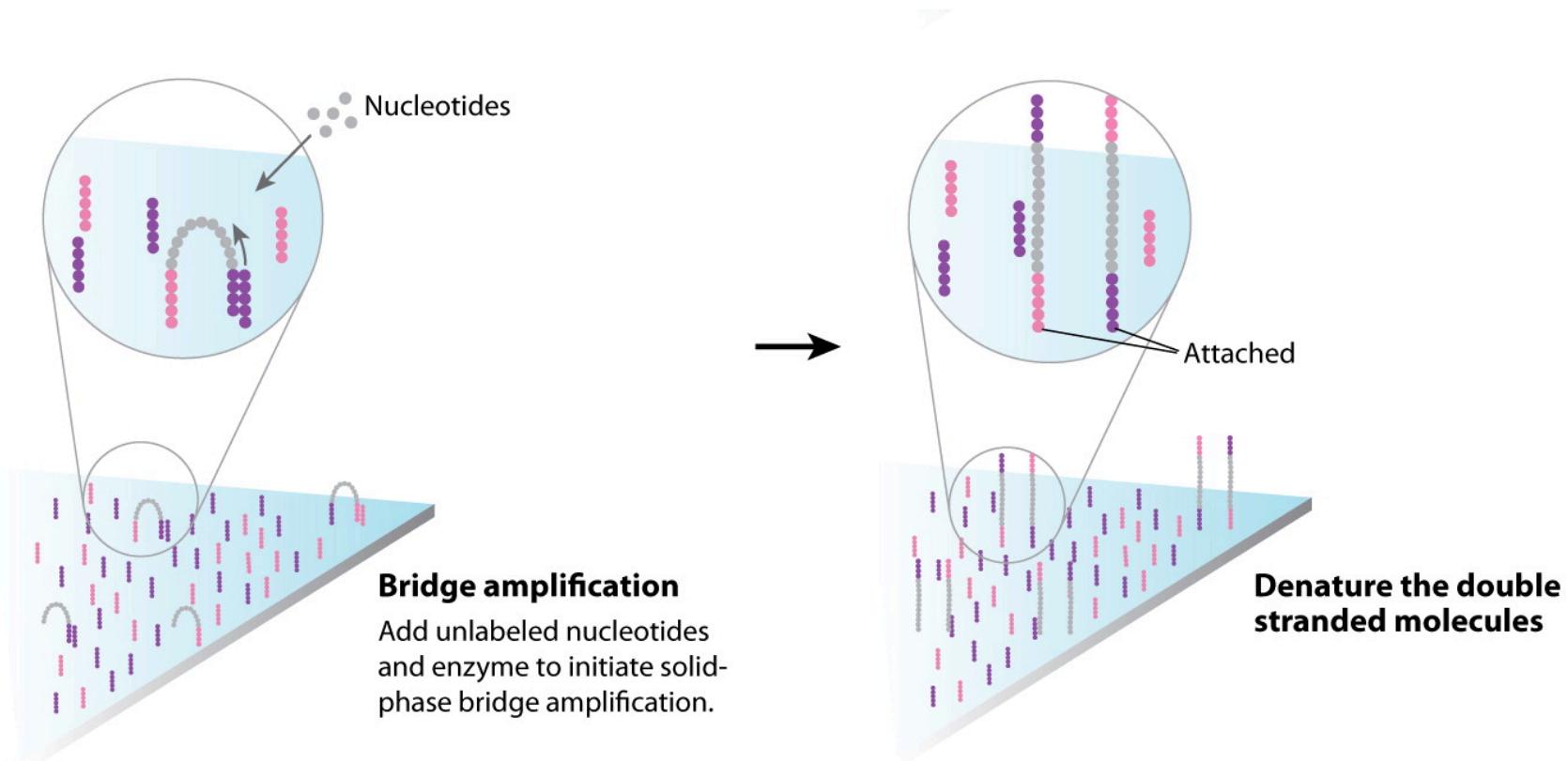
- **Summary of NGS technologies (sequencing and applications)**
- Introduction to NGS data analysis
- Commonly available databases
- Workflow integration and making use of existing NGS data

Illumina sequencing-by-synthesis



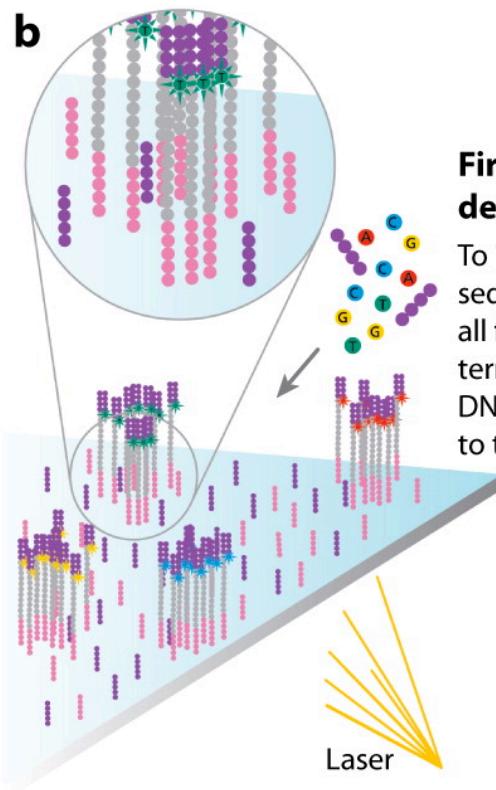
(Mardis, Ann. Rev. Genomics Hum. Genet., 2008)

Illumina sequencing-by-synthesis



(Mardis, Ann. Rev. Genomics Hum. Genet., 2008)

Illumina sequencing-by-synthesis



First chemistry cycle: determine first base

To initiate the first sequencing cycle, add all four labeled reversible terminators, primers, and DNA polymerase enzyme to the flow cell.

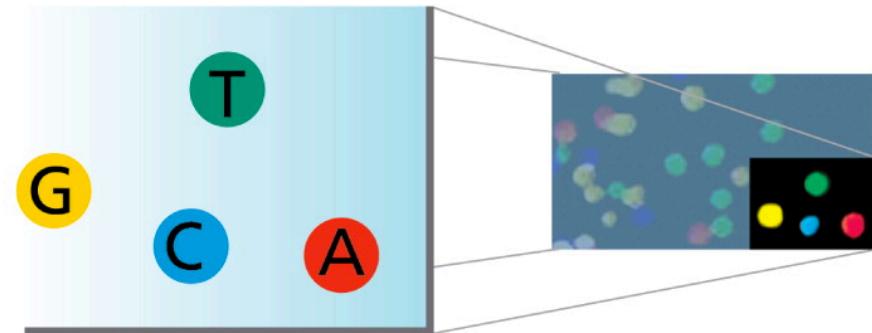


Image of first chemistry cycle

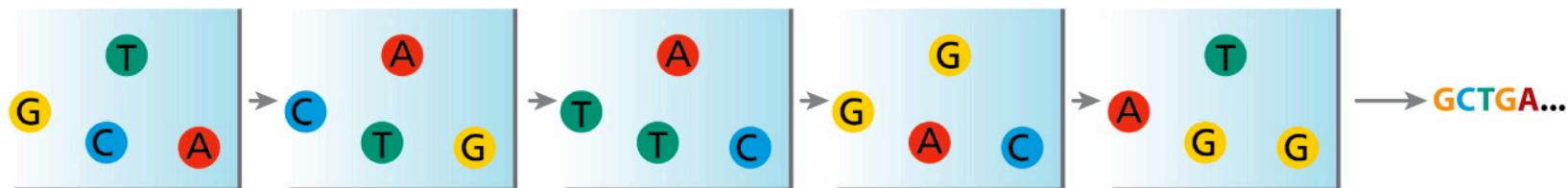
After laser excitation, capture the image of emitted fluorescence from each cluster on the flow cell. Record the identity of the first base for each cluster.

Before initiating the next chemistry cycle

The blocked 3' terminus and the fluorophore from each incorporated base are removed.

(Mardis, Ann. Rev. Genomics Hum. Genet., 2008)

Illumina sequencing-by-synthesis

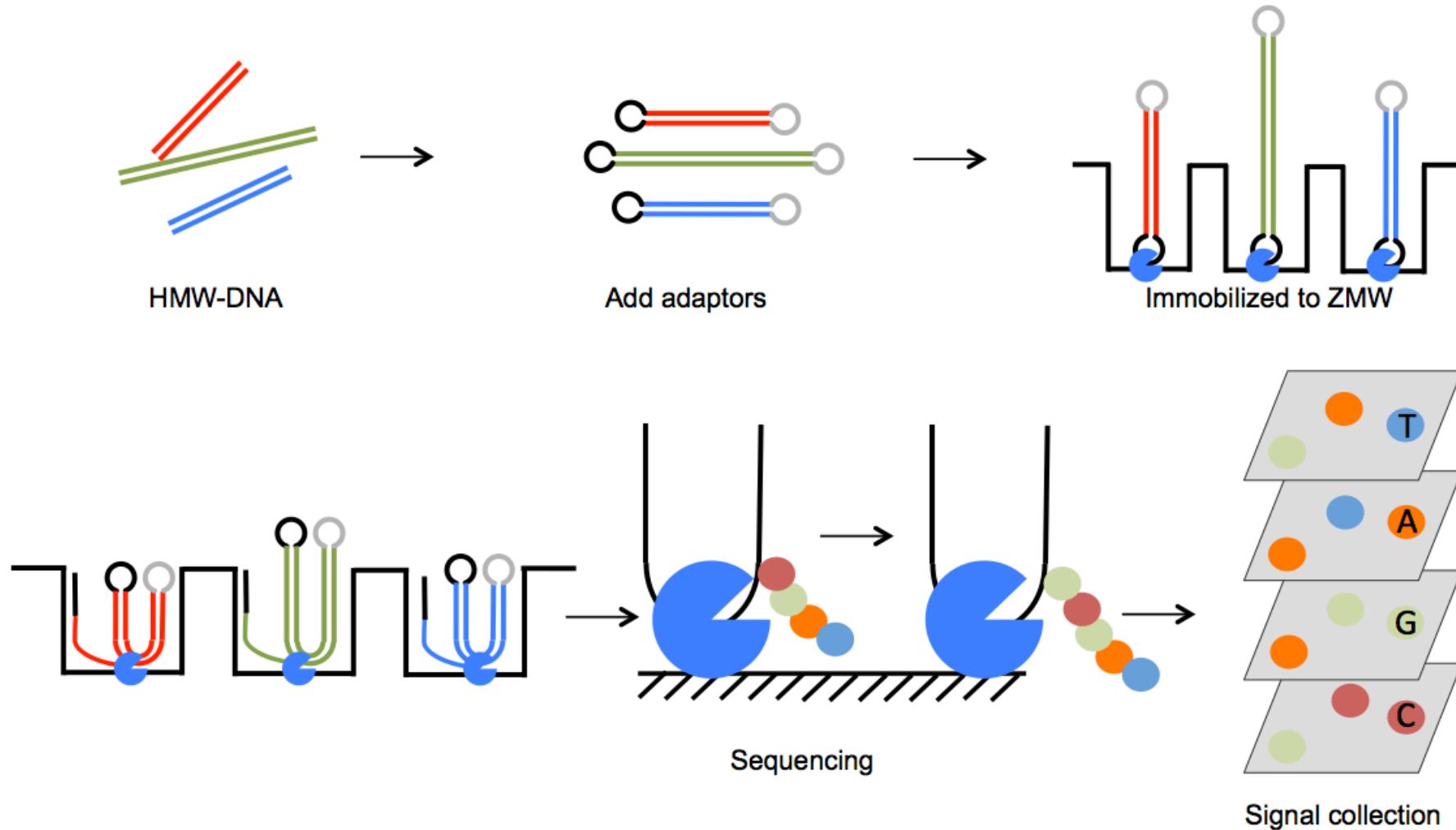


Sequence read over multiple chemistry cycles

Repeat cycles of sequencing to determine the sequence of bases in a given fragment a single base at a time.

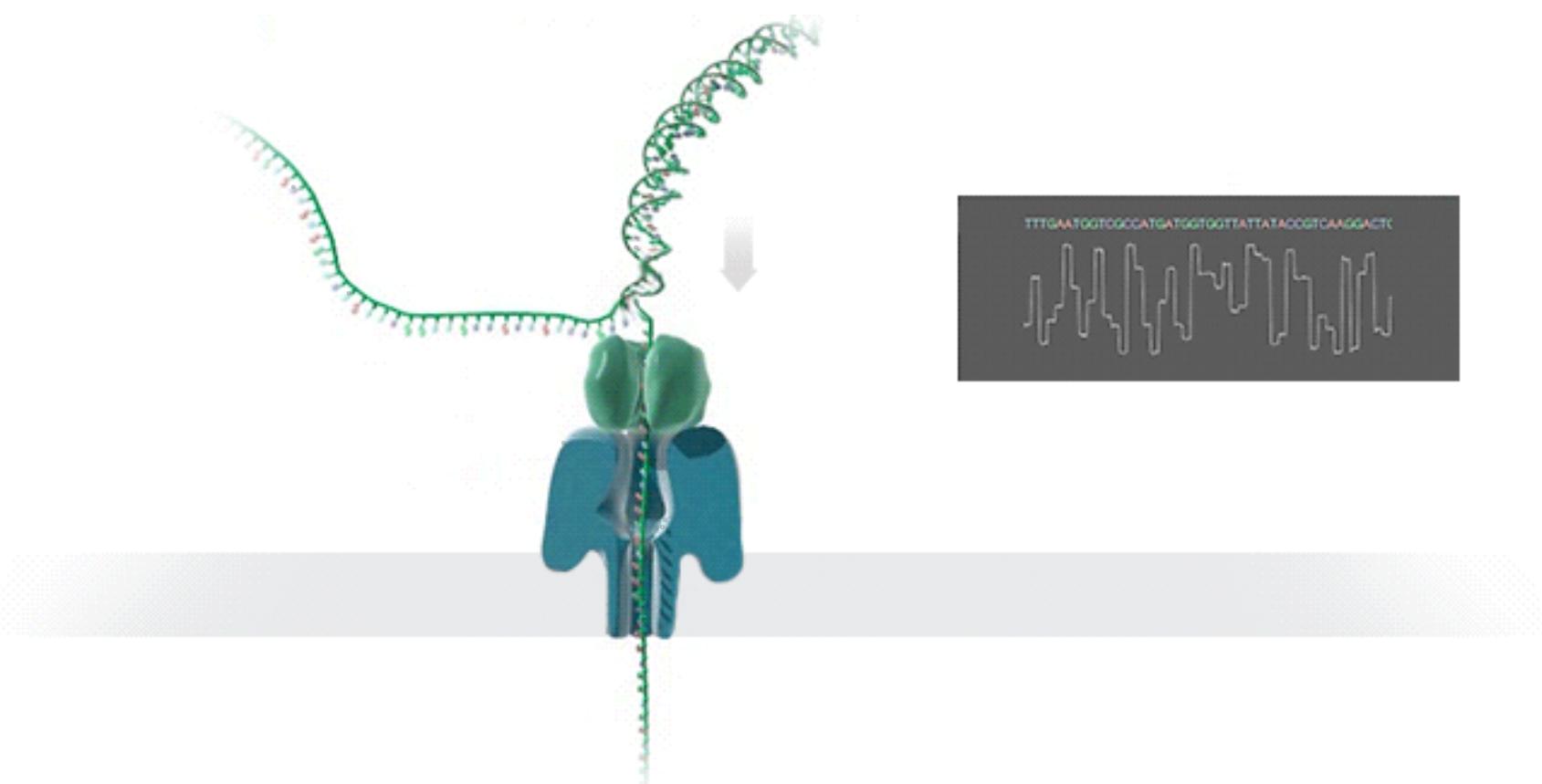
(Mardis, Ann. Rev. Genomics Hum. Genet., 2008)

PacBio SMRT Sequencing



(Image from 3402 Bioinformatics)

Nanopore sequencing



(Image via Oxford Nanopore)

Key considerations for NGS technologies

- Number of reads
- Quality of reads
- Length of reads
- Library preparation
- Cost

Key considerations for NGS technologies

	Illumina	PacBio	Nanopore
Number of reads	***	**	*
Quality of reads	**	*/***	***
Length of reads	*	**	***
Library preparation	Versatile, complex	Moderate	Simple

Library preparation for the Illumina platform

(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

Library preparation for the Illumina platform

TruSeq Universal Adapter:

5 AATGATA CGGCG ACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCT 3

TruSeq Indexed Adapter

5 GATCGGAAGAGCACACGTCTGA~~ACTCCAGTCAC~~-NNNNNN-ATCTCGTATGCCGTCTGCTTG 3

(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

Library preparation for the Illumina platform

TruSeq Universal Adapter:

5 AATGATAACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCT 3

TruSeq Indexed Adapter

5 GATCGGAAGAGCACACGTCTGAACTCCAGTCAC-NNNNNN-ATCTCGTATGCCGTCTTGCTTG 3

Anneal:

5 AATGATAACGGCGACCACCGAGATCTACACTCTTCCCTACACGAC -- GCTCTTCCGATC*T 3
3 GTTCGTCTTCTGCCGTATGCTCA(INDEX)CACTGACCTCAAGTCTGCACA -- CGAGAAGGCTAG*P 5

(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

Library preparation for the Illumina platform

After ligation:

LEFT OF INSERT

5 AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGAC--GCTCTTCCGATC*T 3
3 GTTCGTCTTCTGCCGTATGCTCTA(INDEX)CACTGACCTCAAGTCTGCACA--CGAGAAGGCTAG*P 5

RIGHT OF INSERT

5 P*GATCGGAAGAGC--ACACGTCTGAACCTCCAGTCAC(INDEX)ATCTCGTATGCCGTCTGCTTG 3
3 T*CTAGCCTTCTCG--CAGCACATCCCTTCTCACATCTAGAGCCACCAGCGGCATAGTAA 5

(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

Library preparation for the Illumina platform

PCR Primer 1.0

5 AATGATAACGGCGACCACCGAGATCTACACTCTTCCCTACACGA 3

PCR Primer 2.0

5 CAAGCAGAAGACGGCATACGAGAT 3

Universal Adapter:

5 AATGATAACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTTTCCGATCT 3

Indexing Adapter:

5 GATCGGAAGAGCACACGTCTGAACCTCCAGTCAC-NNNNNN-ATCTCGTATGCCGTCTTGCTTG 3

(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

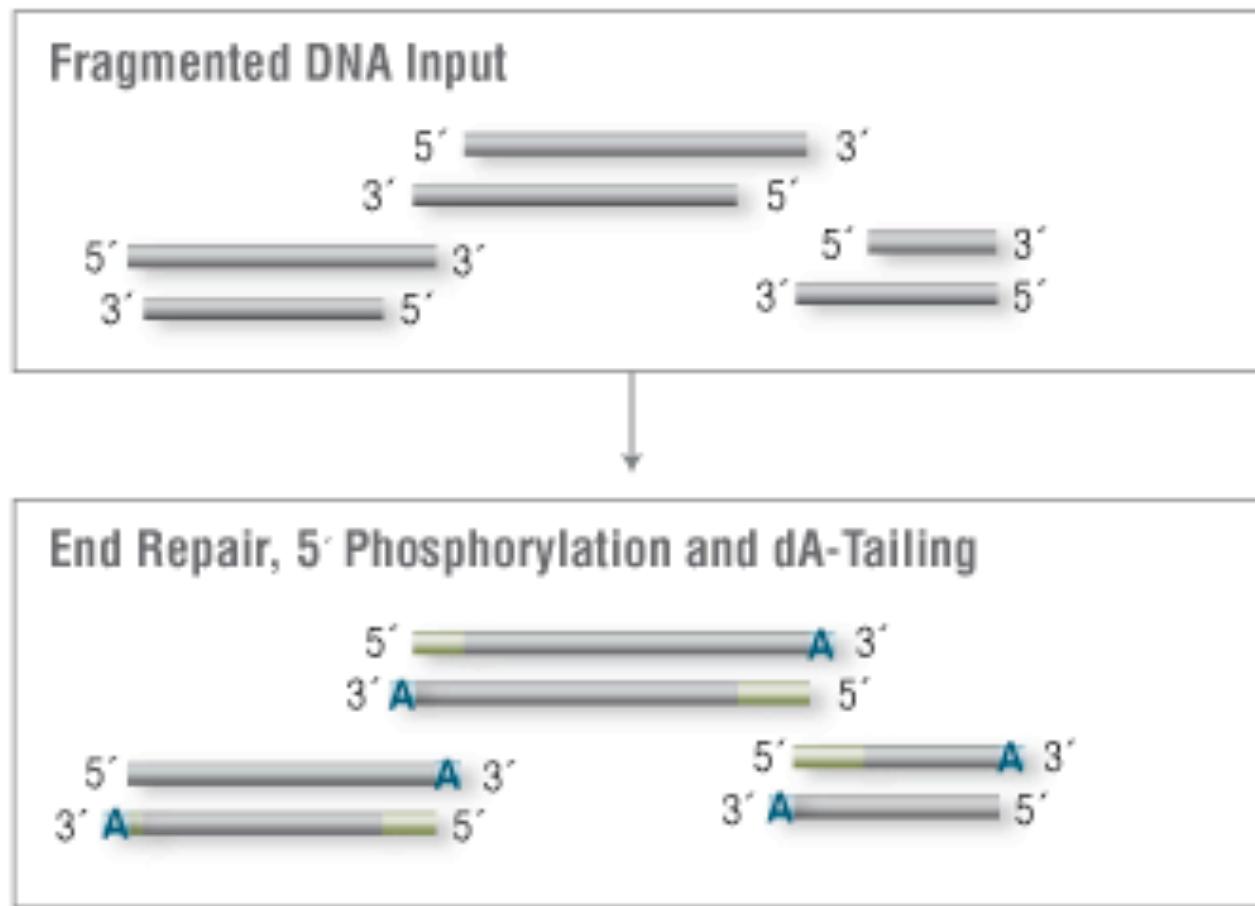
Library preparation for the Illumina platform



- **Universal Adapter**
- **DNA Fragment of Interest**
- **Indexed Adapter**
- **6 Base Index Region**

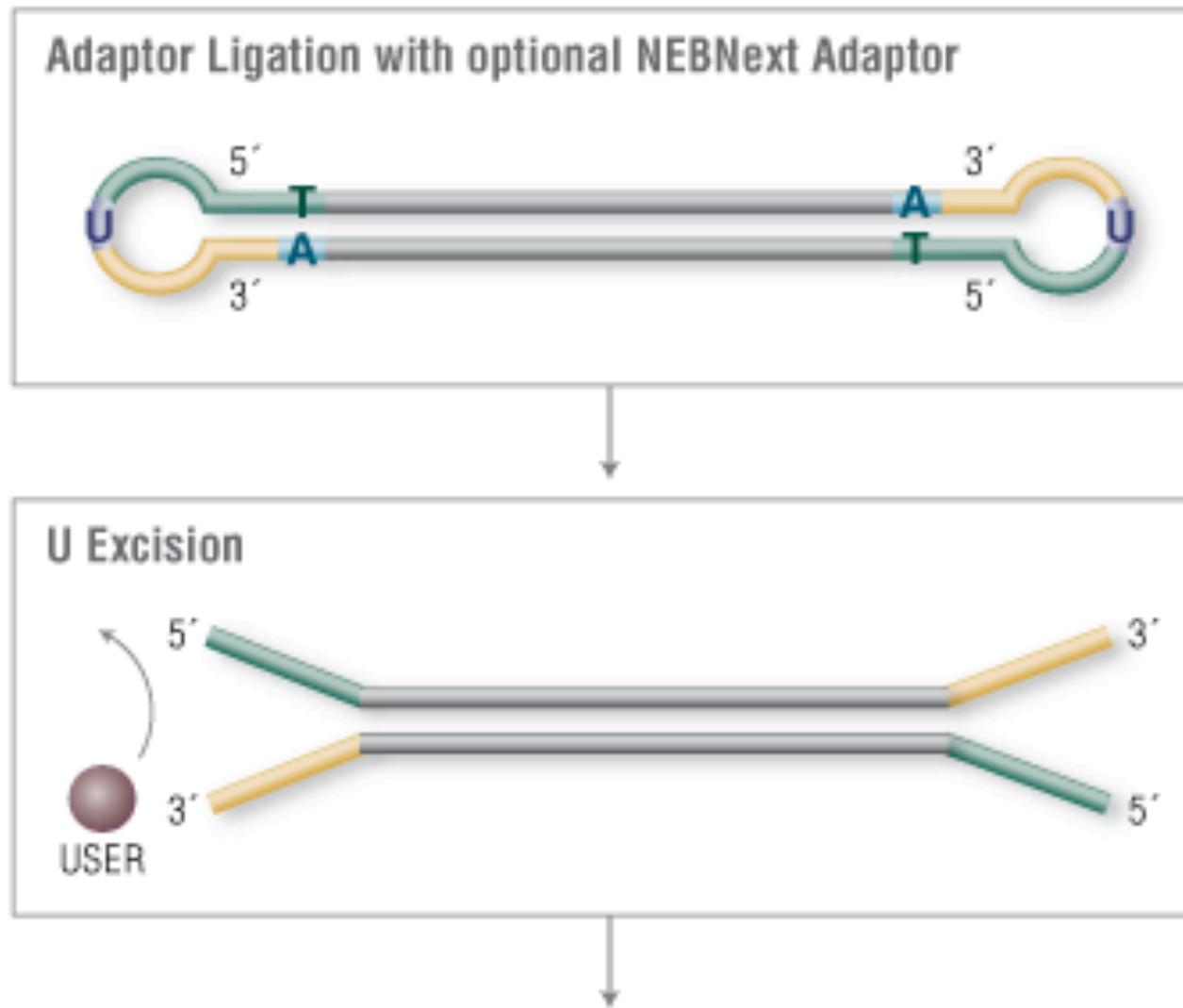
(From “Illumina TruSeq DNA Adapters De-Mystified” by James Schiemer)

Example of a full sequencing prep workflow

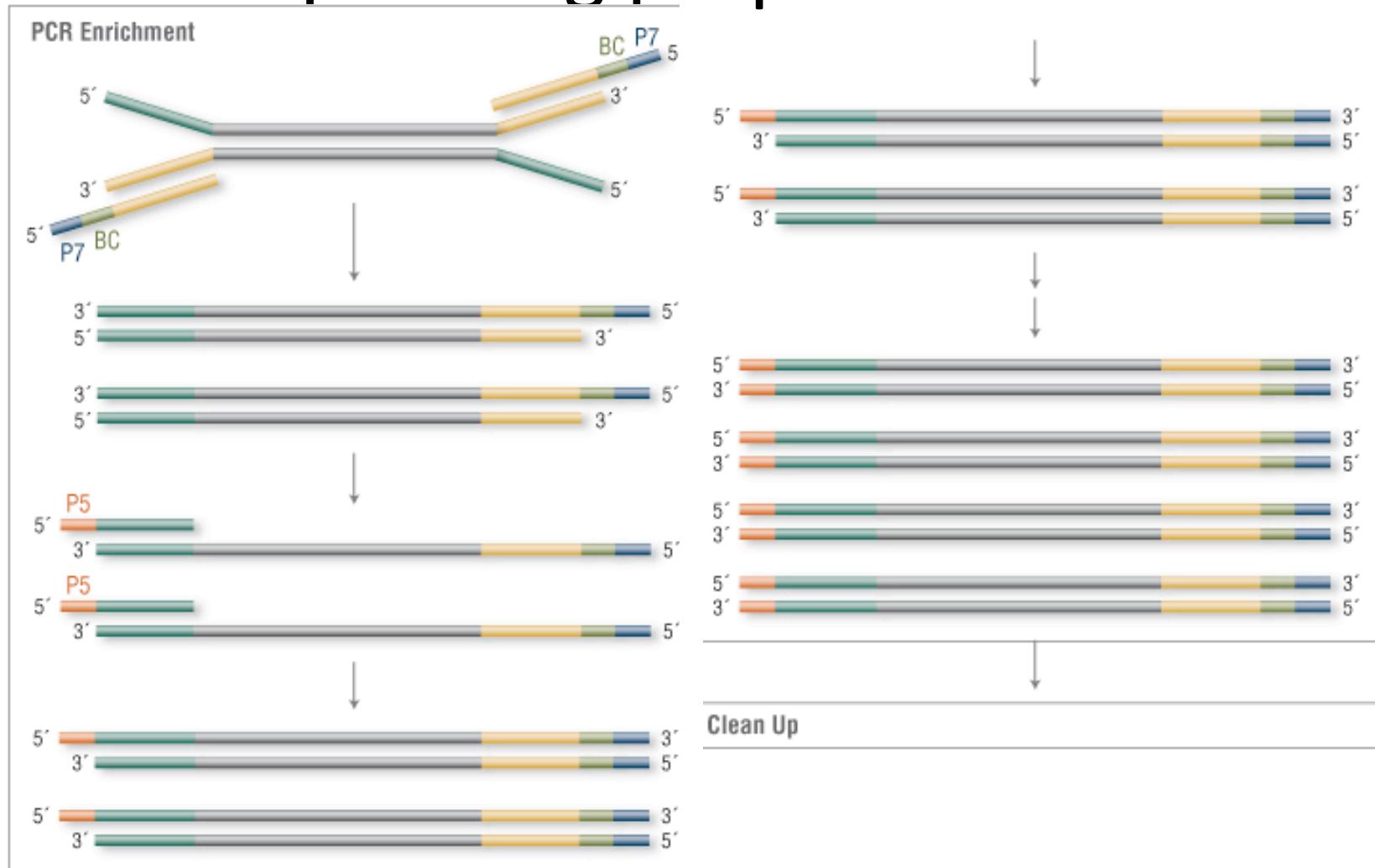


(Image from NEB)

Example of a full sequencing prep workflow



Example of a full sequencing prep workflow

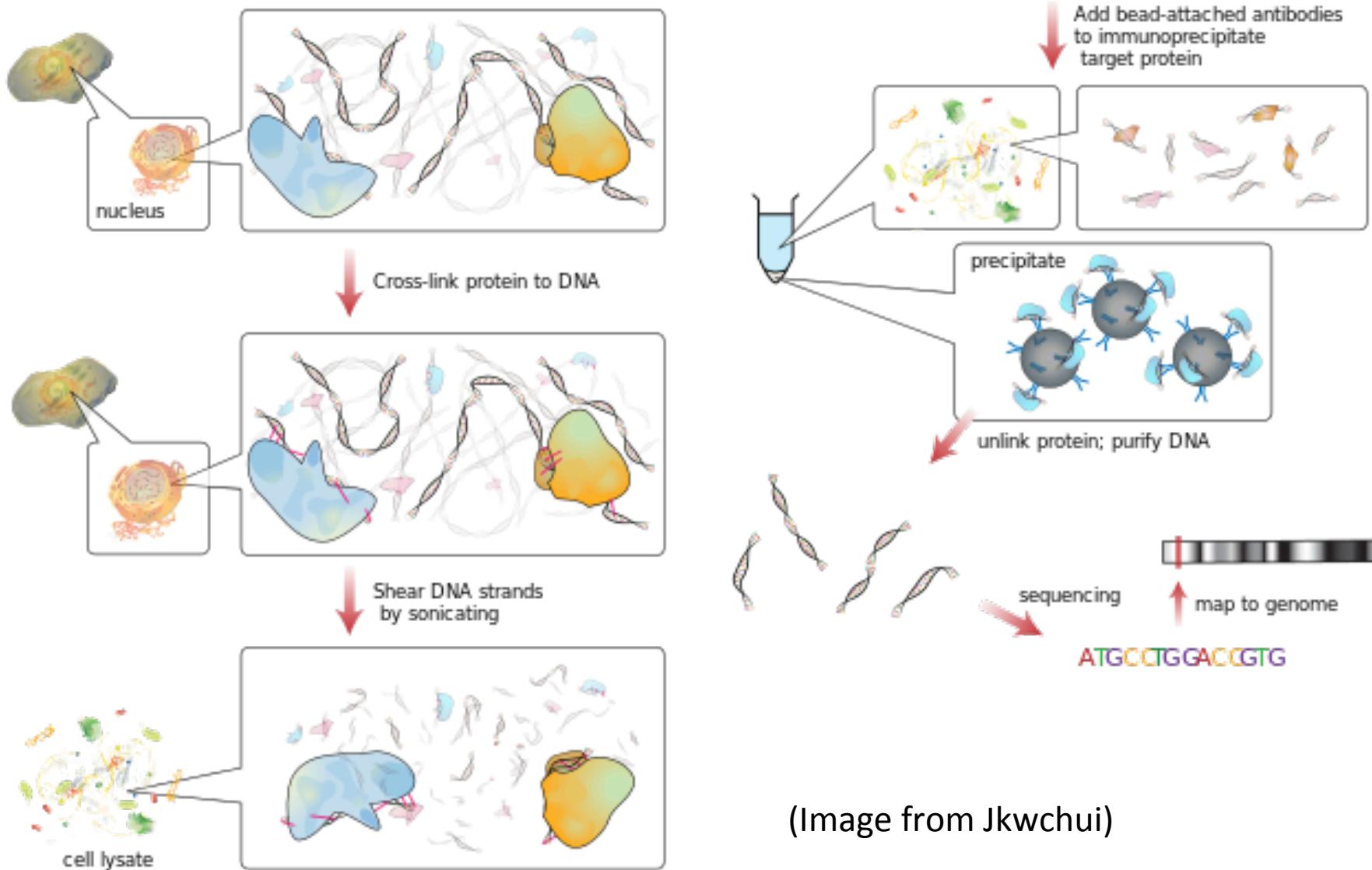


(Image from NEB)

Applications

- Genome sequencing (whole genome, mutations)
- RNA sequencing (transcript quantitation, transcriptome mapping)
- Finding protein-nucleic acid interaction (ChIP-seq, PAR-CLIP)
- Identifying methylation sites (bisulfite sequencing)

Applications



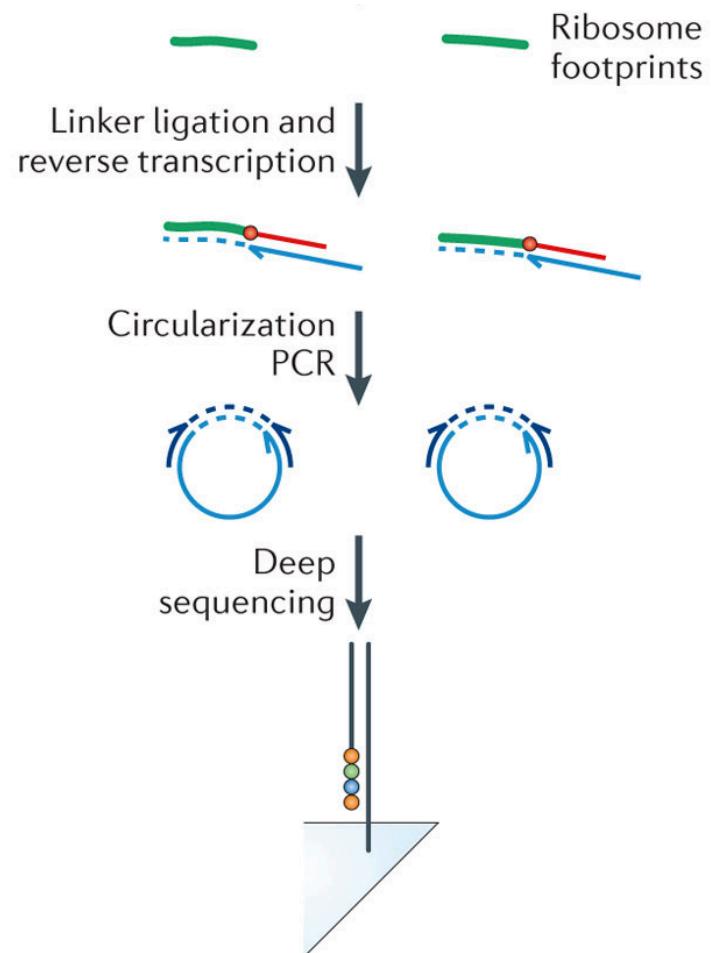
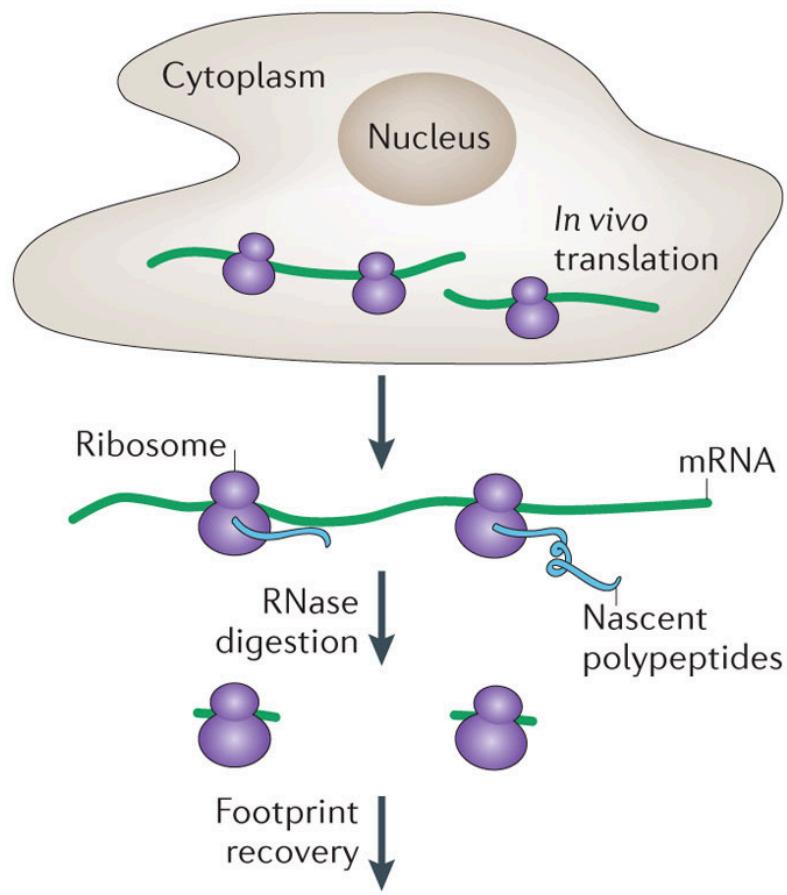
(Image from Jkwchui)

Applications

- Protein translation rates (ribosome profiling)
- Chromosomal conformations (Hi-C)
- Transcript stability (Bru-chase seq)
- Finding DNA-RNA hybrids (Drip-seq)
- Profiling chromatin accessibility (ATAC-seq, Mnase-seq)

And on and on...

Ribosome profiling

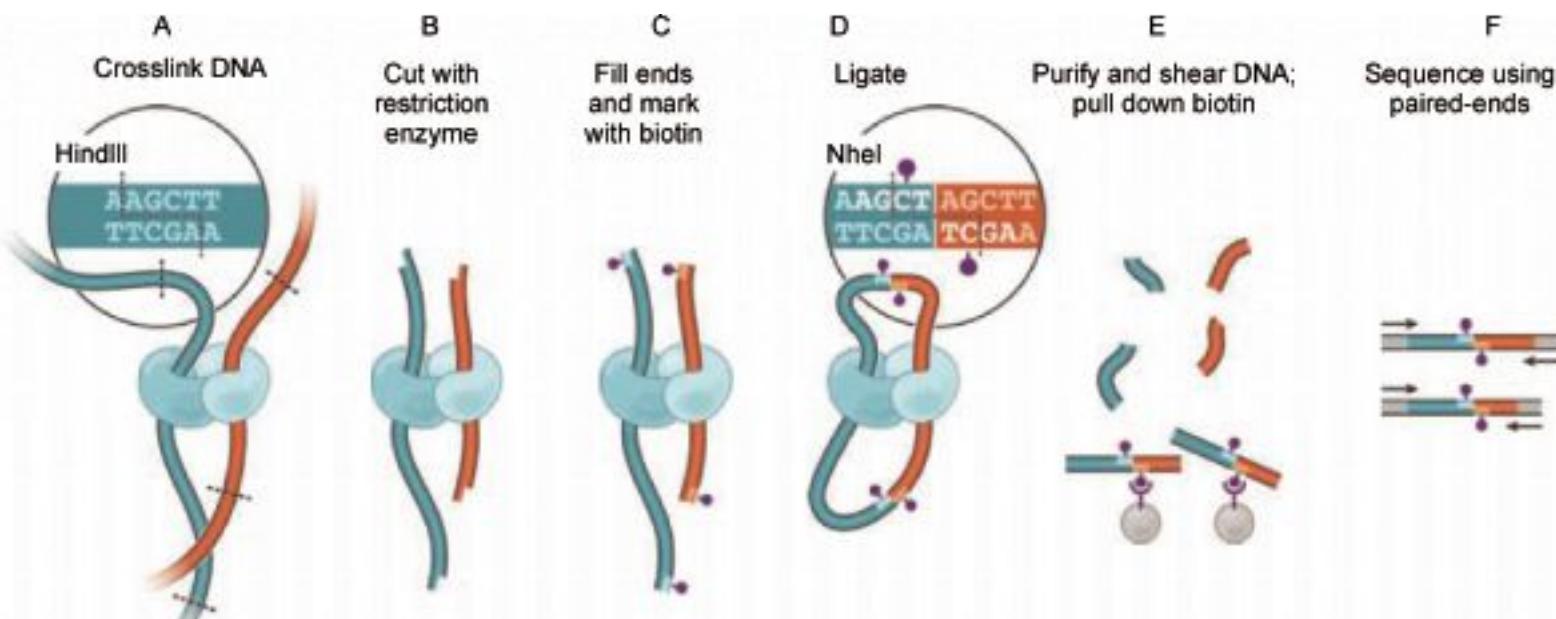


Applications

- Protein translation rates (ribosome profiling)
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And on and on...

Applications



(Lieberman-Aiden et al., Science, 2009)

Applications

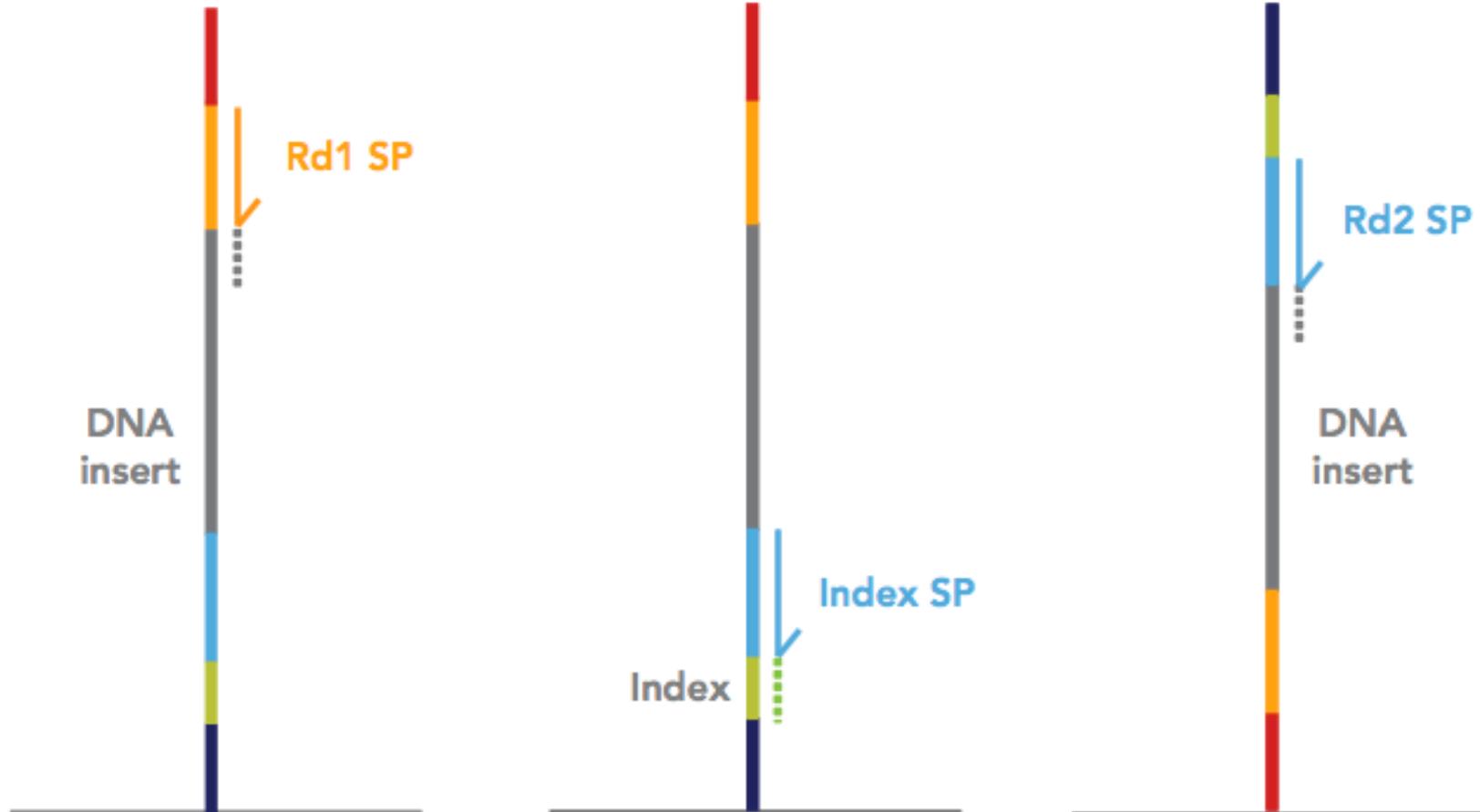
- Protein translation rates (ribosome profiling)
- Chromosomal conformations (Hi-C)
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And on and on...

A few crucial concepts

- Single end vs. paired end reads
- “Coverage”
- Indexing
- FPKM, RPKM, TPM

A few crucial concepts



(Image from Illumina)

A few crucial concepts

- Single end vs. paired end reads
- “Coverage”
- Indexing
- FPKM, RPKM, TPM

Outline

- Summary of NGS technologies (sequencing and applications)
- **Introduction to NGS data analysis**
- Commonly available databases
- Workflow integration and making use of existing NGS data

Raw data: Fastq files

```
@K00135:141:HHJ3TBBXX:1:2228:1661:47383
CTCCTGTTCTTGTGGTTGCTGGGGCTCCAATAG
+
AAA-AAJFF-AFAAF7FFA-AA-77AJ<7A<-7
@K00135:141:HHJ3TBBXX:1:2228:5467:17685
GTATTTTAGTTCCATACACGCAAGAAGGAG
+
-A-<AF<<FFF<FJFA--<--77<<JF7-7<
```

Raw data: Fastq files

Read name	@K00135:141:HHJ3TBBXX:1:2228:1661:47383
Sequence	CTCCTGTTCTTGTGGTTGCTGGGGCTCCAATAG
Optional information	+
Quality scores	AAA-AAJFF-AFAAF7FFA-AA-77AJ<7A<-7
Read name	@K00135:141:HHJ3TBBXX:1:2228:5467:17685
Sequence	GTATTTTAGTTCCATACACGCAAGAAGGAG
Optional information	+
Quality scores	-A-<AF<<FFF<FJFA--<-77<<JF7-7<

Typical steps in analysis workflow

- Quality control
- Adapter clipping
- Quality trimming
- Alignment
- Analysis

Quality control

(Example: FastQC)

FastQC Report

Summary

- [Basic Statistics](#)
- [Per base sequence quality](#)
- [Per tile sequence quality](#)
- [Per sequence quality scores](#)
- [Per base sequence content](#)
- [Per sequence GC content](#)
- [Per base N content](#)
- [Sequence Length Distribution](#)
- [Sequence Duplication Levels](#)
- [Overrepresented sequences](#)
- [Adapter Content](#)
- [Kmer Content](#)

Basic Statistics

Measure	Value
Filename	good_sequence_short.txt
File type	Conventional base calls
Encoding	Illumina 1.5
Total Sequences	250000
Sequences flagged as poor quality	0
Sequence length	40
%GC	45

Per base sequence quality

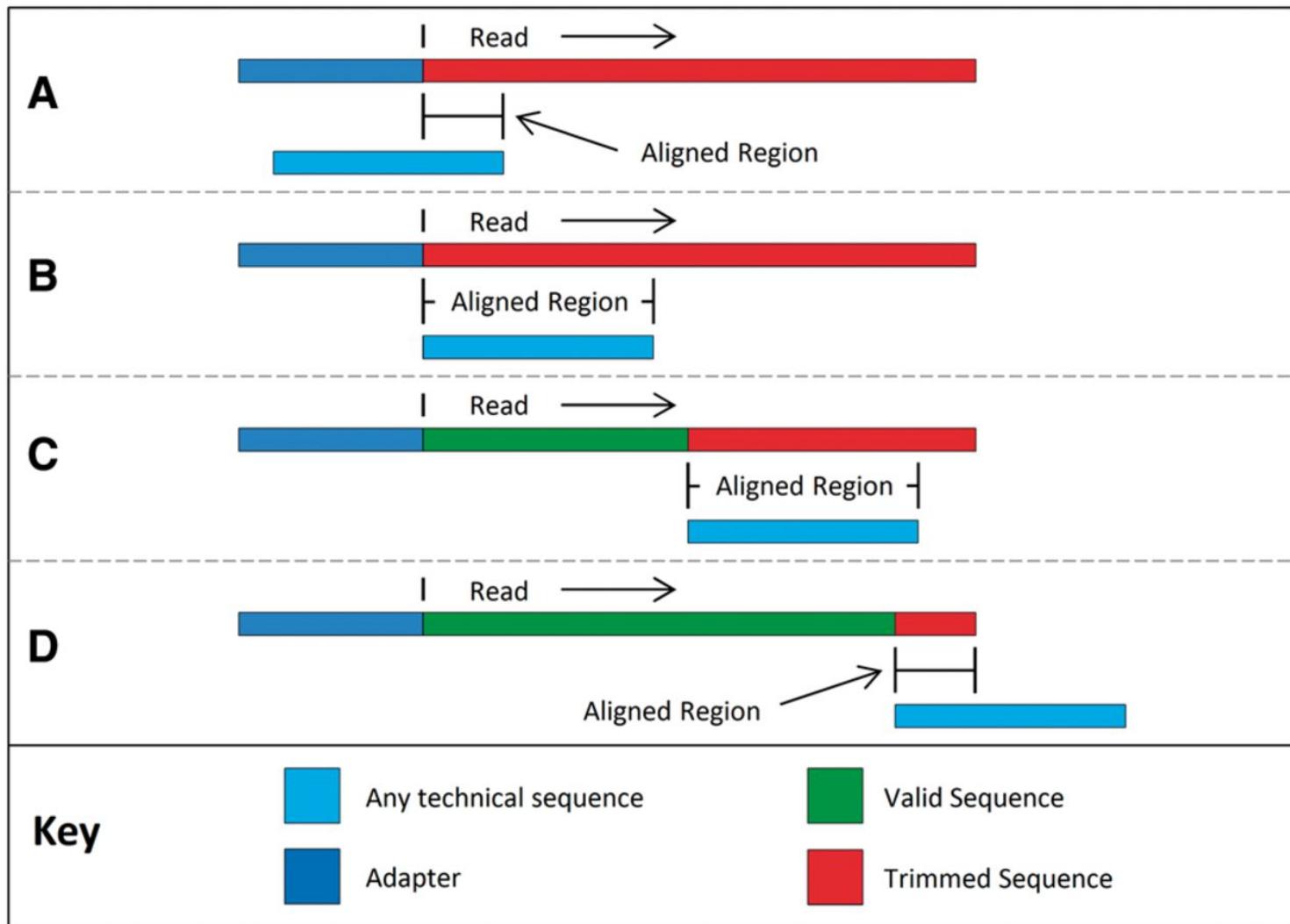
Quality scores across all bases (Illumina 1.5 encoding)

The plot displays the quality scores for each base in the sequence. The y-axis represents the quality score, ranging from 32 to 38. The x-axis represents the sequence positions. The plot shows a generally high quality score (around 37) with some minor fluctuations. A blue line represents the mean quality score, and green vertical bars represent the standard deviation.

(fastqc webpage)

Trimming/clipping

(fastxtools, cutadapt, trimmomatic)



Alignment

Examples: Bowtie, BWA, SOAP, STAR, HiSat, ...

(image from labtimes.org)

(or assembly...)

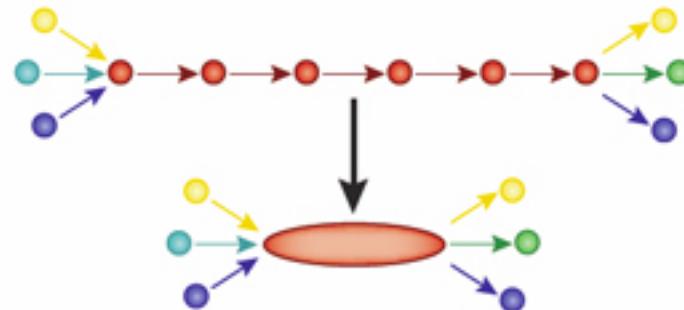
1. Fragment DNA and sequence



2. Find overlaps between reads

...AGCCTAGACCTACA**GGATGCGCGACACGT**
GGATGCGCGACACGTCGCATATCCGGT...

3. Assemble overlaps into contigs



4. Assemble contigs into scaffolds

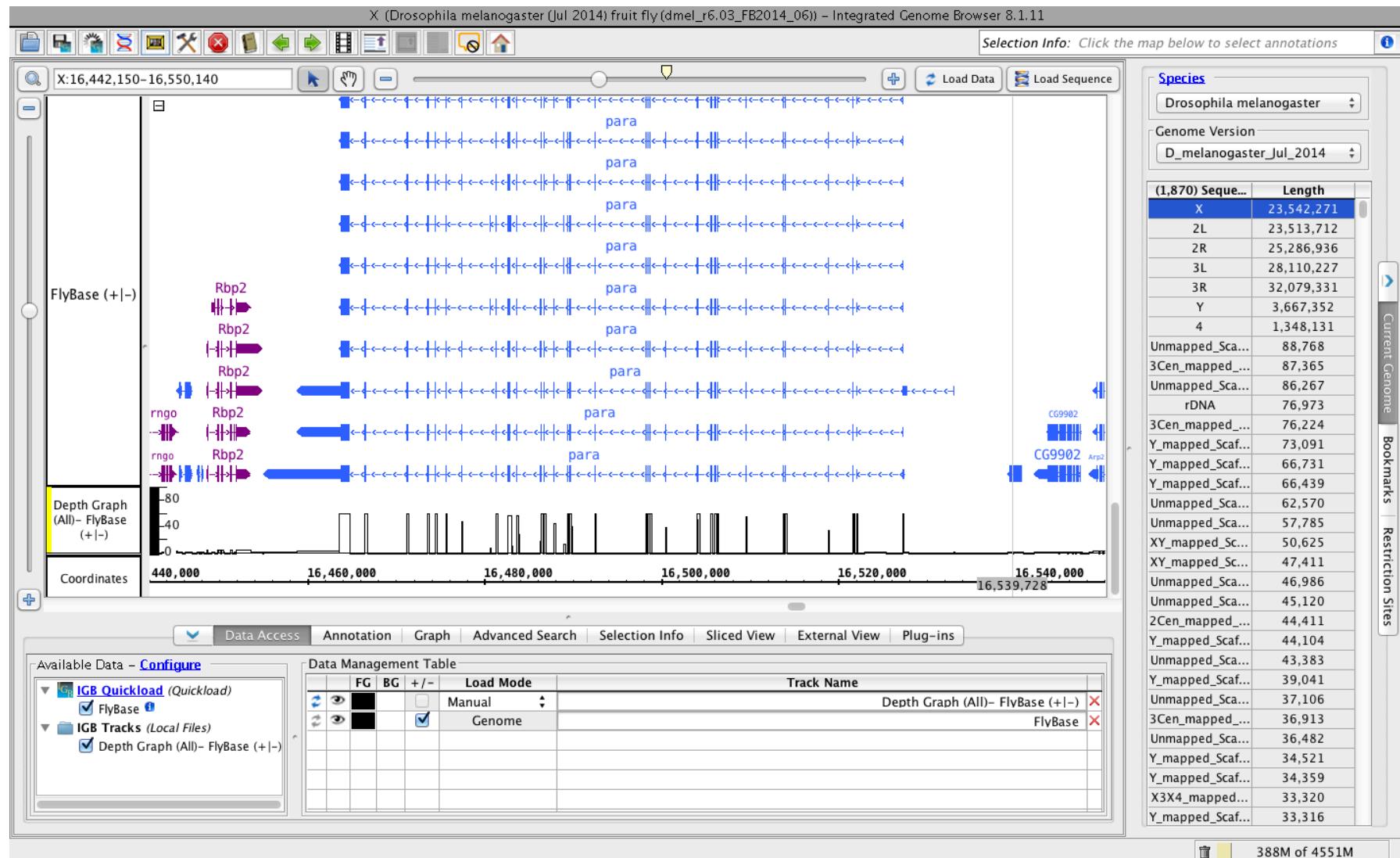


(image from Michael Schatz)

Examples:
ABySS, MIRA, SSAKE
(genome)

Cufflinks, Stringtie,
Trinity
(transcriptome)

Analysis



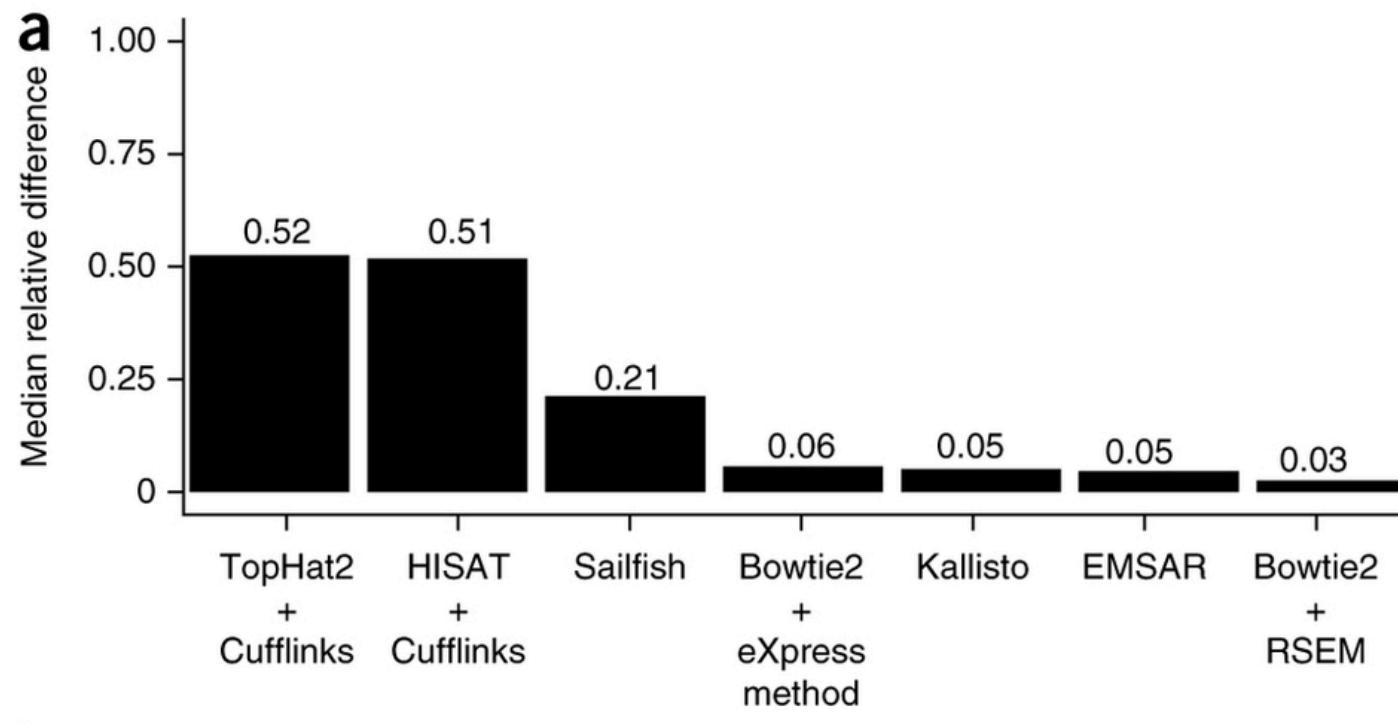
(IGB screenshot by Ann Loraine)

Common tasks

- Transcript quantitation
- Isoform calling
- Peak calling
- Gene set enrichment analysis
- Motif analysis
- Clustering/network inference

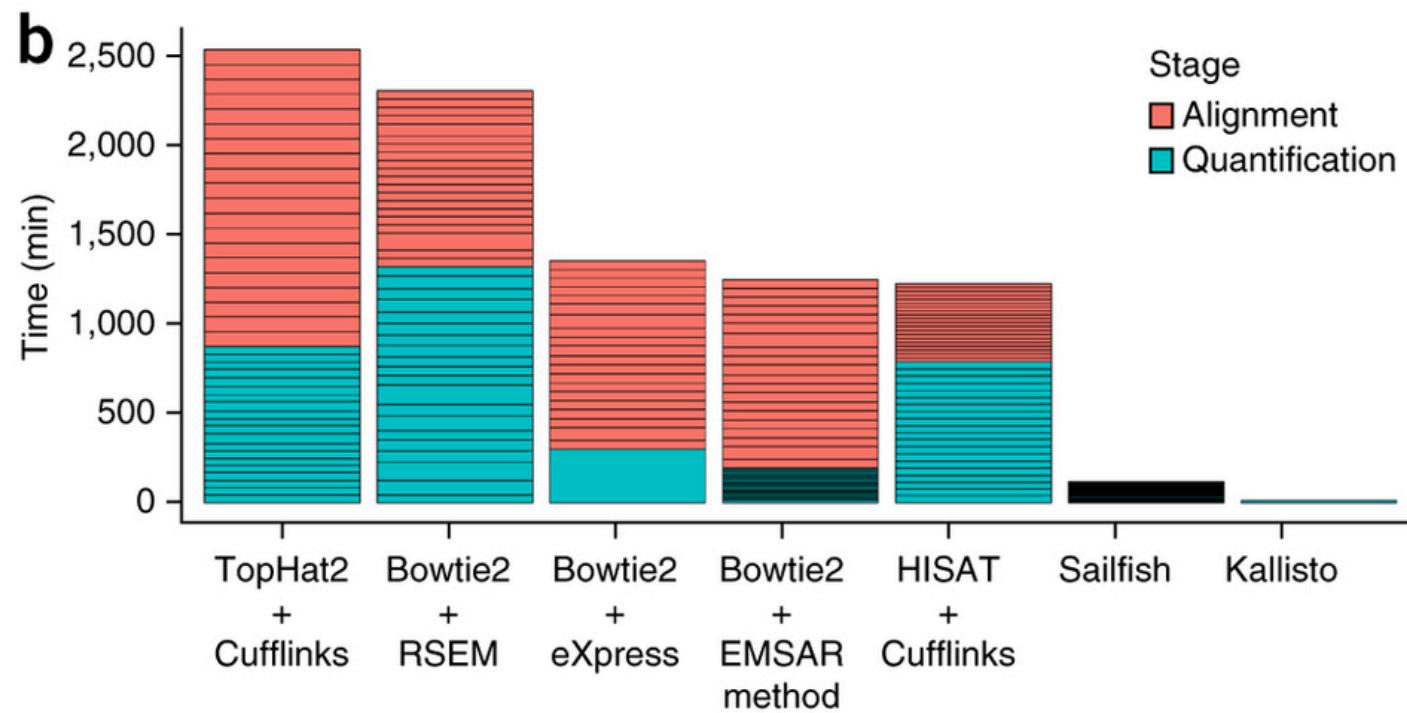
Pseudoalignment and fast RNA-seq workflows

- Don't get exact alignments, just find transcripts compatible with each read
- Examples: kallisto, sailfish, salmon

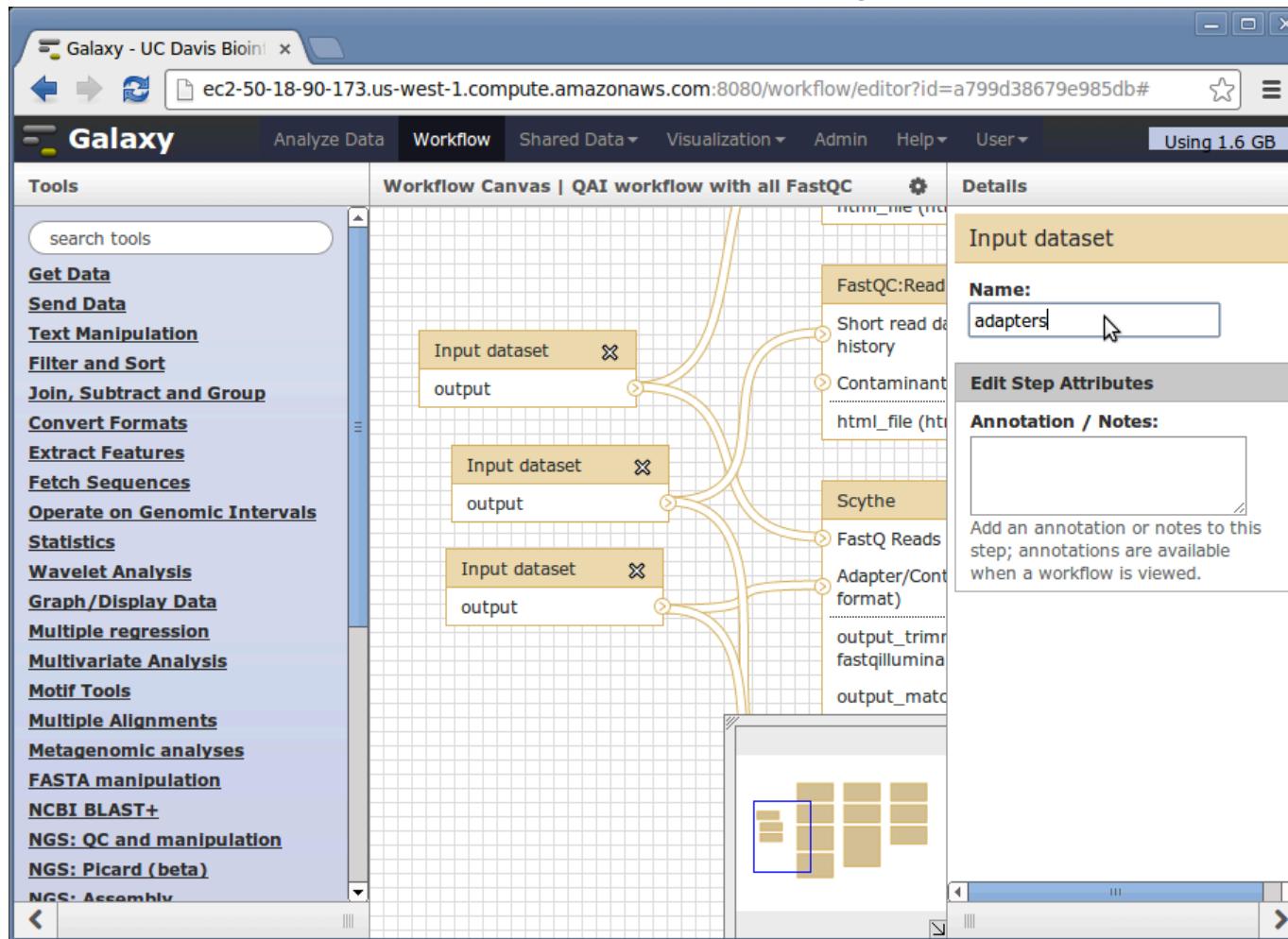


Pseudoalignment and fast RNA-seq workflows

- Don't get exact alignments, just find transcripts compatible with each read
- Examples: kallisto, sailfish, salmon



A unified graphical interface for NGS analysis



Galaxy: usegalaxy.org; image from UC Davis Bioinformatics Core

Outline

- Summary of NGS technologies (sequencing and applications)
- Introduction to NGS data analysis
- **Commonly available databases**
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GEO – the gene expression omnibus

www.ncbi.nlm.nih.gov/geo

NCBI Resources ▾ How To ▾

GEO Home Documentation ▾ Query & Browse ▾ Email GEO

petered My NCBI Sign Out

My GEO Submissions

Gene Expression Omnibus

GEO is a public functional genomics data repository supporting MIAME-compliant data submissions. Array- and sequence-based data are accepted. Tools are provided to help users query and download experiments and curated gene expression profiles.

 Gene Expression Omnibus

Keyword or GEO Accession

Getting Started	Tools	Browse Content
Overview	Search for Studies at GEO DataSets	Repository Browser
FAQ	Search for Gene Expression at GEO Profiles	DataSets: 4348
About GEO DataSets	Search GEO Documentation	Series:  82875
About GEO Profiles	Analyze a Study with GEO2R	Platforms: 17052
About GEO2R Analysis	GEO BLAST	Samples: 2018686
How to Construct a Query	Programmatic Access	
How to Download Data	FTP Site	

Information for Submitters

My GEO Submissions	Submission Guidelines	MIAME Standards
My GEO Profile	Update Guidelines	Citing and Linking to GEO
		Guidelines for Reviewers
		GEO Publications

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Example GEO dataset

The screenshot shows the NCBI GEO Accession Display page for dataset GSE32022. The top navigation bar includes links for HOME, SEARCH, SITE MAP, GEO Publications, FAQ, MIAME, and Email GEO. Below the navigation is a breadcrumb trail: NCBI > GEO > Accession Display. On the right, there are links for Contact, My submissions, and Logout. A search bar at the top allows users to filter by Scope (Self or All), Format (HTML or XML), Amount (Quick or Full), and GEO accession (GSE32022). A 'GO' button is also present.

Series GSE32022 UPDATE Query DataSets for GSE32022

Status	Public on Jun 01, 2012
Title	Characterization of transcriptional and fitness effects of a loss of function mutation in rho
Platform organism	Escherichia coli
Sample organism	Escherichia coli str. K-12 substr. MG1655
Experiment type	Expression profiling by genome tiling array Genome variation profiling by genome tiling array
Summary	In order to study the effects of a mutation to the transcriptional termination regulator Rho (referred to as rho*), we made use of expression microarrays to observe the direct and indirect effects of rho* on gene expression. In addition, we used arrays to map the fitness of strains from transposon mutagenized libraries under four conditions, showing that in each case the majority of genes with significant fitness effects were dependent on the genotype at rho.
Overall design	For expression arrays, we performed two-color microarrays comparing transcript levels in rho* and wild type cells during exponential growth in glucose minimal media. For selection experiments, transposon insertions were mapped through selective amplification of genomic regions adjacent to them. We then measured the fitness effects of insertions throughout the genome using two-color microarrays, comparing amplified DNA from a population grown under a selective condition of interest to an isogenic control population grown under a reference condition (glucose minimal media). All arrays were performed in duplicate, and the source material for the duplicates came from separate biological replicates.
Contributor(s)	Freddolino PL, Goodarzi H
Citation(s)	Freddolino PL, Goodarzi H, Tavazoie S. Fitness landscape transformation through a single amino acid change in the rho terminator. <i>PLoS Genet</i> 2012 May;8(5):e1002744. PMID: 22693458

Example GEO dataset

Platforms (1) [GPL10286](#) Escherichia coli whole-genome tiling array (2 x 105K)

Samples (9) [GSM794088](#) RNA

[≡ Less...](#)

[GSM794089](#) WT_AKG

[GSM794090](#) WT_CML

[GSM794091](#) WT_NADM

[GSM794092](#) WT_STP

[GSM794093](#) MUT_AKG

[GSM794094](#) MUT_CML

[GSM794095](#) MUT_NADM

[GSM794096](#) MUT_STP

Relations

BioProject [PRJNA147515](#)

Download family

Format

[SOFT](#) formatted family file(s)

[SOFT](#) [?](#)

[MINiML](#) formatted family file(s)

[MINiML](#) [?](#)

[Series Matrix File\(s\)](#)

[TXT](#) [?](#)

Supplementary file	Size	Download	File type/resource
GSE32022_RAW.tar	604.3 Mb	(http)(custom)	TAR (of TXT)

Raw data provided as supplementary file

Processed data provided as supplementary file

Example GEO dataset

Platforms (1) [GPL10286 Escherichia coli whole-genome tiling array \(2 x 105K\)](#)

Samples (9) [GSM794088 RNA](#)

[GSM794089 WT_AKG](#)
 [GSM794090 WT_CML](#)
 [GSM794091 WT_NADM](#)
 [GSM794092 WT_STP](#)
 [GSM794093 MUT_AKG](#)
 [GSM794094 MUT_CML](#)
 [GSM794095 MUT_NADM](#)
 [GSM794096 MUT_STP](#)

Relations

BioProject [PRJNA147515](#)

Download family **Format**

[SOFT formatted family file\(s\)](#) SOFT [?](#)

[MINiML formatted family file\(s\)](#) MINiML [?](#)

[Series Matrix File\(s\)](#) TXT [?](#)

Supplementary file	Size	Download	File type/resource
GSE32022_RAW.tar	604.3 Mb	(http)(custom)	TAR (of TXT)

Raw data provided as supplementary file

Processed data provided as supplementary file

| NLM | NIH | GEO Help | Disclaimer | Accessibility |

Example GEO dataset

NCBI  Gene Expression Omnibus

HOME SEARCH SITE MAP | GEO Publications | FAQ | MIAME | Email GEO | Contact: petefred | My submissions | Logout

Scope: Self Format: HTML Amount: Quick | GEO accession: GSM794088 | GO

Sample GSM794088 **UPDATE** Query DataSets for GSM794088

Status	Public on Jun 01, 2012
Title	RNA
Sample type	RNA

Channel 1

Source name	WT cells_mid-log-phase_M9t/glucose
Organism	Escherichia coli str. K-12 substr. MG1655
Characteristics	genotype/variation: WT growth media: M9t/glucose growth phase: mid-log
Growth protocol	For RNA samples, WT or rho* cells were grown to mid-log phase in M9t/glucose. WT or rho* transposon mutagenized libraries were grown overnight in the media indicated.
Extracted molecule	total RNA
Extraction protocol	Total RNA was extracted using total RNA purification kit (Norgen Biotek, Cat 17200).
Label	Cy5
Label protocol	A poly-A tail was added to the RNA samples using E. coli Poly(A) polymerase (NEB, M0276) for 15 minutes. Using an Agilent low input quick amp labeling kit, the rho* and WT samples were then labeled with Cy3 and Cy5, respectively.

Channel 2

Source name	rho* cells_mid-log-phase_M9t/glucose
Organism	Escherichia coli str. K-12 substr. MG1655
Characteristics	genotype/variation: rho* growth media: M9t/glucose growth phase: mid-log
Growth protocol	For RNA samples, WT or rho* cells were grown to mid-log phase in M9t/glucose. WT or rho* transposon mutagenized libraries were grown overnight in the media indicated.
Extracted molecule	total RNA
Extraction protocol	Total RNA was extracted using total RNA purification kit (Norgen Biotek, Cat 17200).

Example GEO dataset

Supplementary file	Size	Download	File type/resource
GSM794088_TAHG20110218_252456810085_S01_GE2-v5_95_Feb07_1_1.txt.gz	32.9 Mb	(ftp)(http)	TXT
GSM794088_TAHG20110218_252456810085_S01_GE2-v5_95_Feb07_1_2.txt.gz	32.8 Mb	(ftp)(http)	TXT
GSM794088_rna_lograt_zscore.txt.gz	1.3 Mb	(ftp)(http)	TXT

Raw

Raw

Processed

Raw data provided as supplementary file

Processed data provided as supplementary file

Navigating GEO datasets

- GPLXXXXX – Platform identifier
- GSEXXXXX – series of data sets (e.g., one paper)
- GSMXXXXX – One sample (may be one or more replicates, but should be same condition)
- GDSXXXXX – Curated data set with additional options available

Getting GEO expression data

Easiest: Use curated data sets

<https://www.ncbi.nlm.nih.gov/sites/GDSbrowser/>

The screenshot shows the NCBI GDS Browser interface. At the top, there is a search bar, a page size selector set to 20, and a navigation bar indicating 1 of 218 pages. Below this is a table of datasets:

DataSet	Title	Organism(s)	Platform	Series	Samples
GDS6082	Sendai virus infection effect on monocytic cell line: dose response	<i>Homo sapiens</i>	GPL10558	GSE67198	11
GDS6064	Arthritic tarsal joints induced by collagen: time course	<i>Mus musculus</i>	GPL6246	GSE61140	15
GDS6063	Influenza A effect on plasmacytoid dendritic cells	<i>Homo sapiens</i>	GPL10558	GSE68849	10
GDS6016	Transcription factor engrailed-2 loss-of-function model of autism spectrum disorder: hippocam...	<i>Mus musculus</i>	GPL7202	GSE51612	6
GDS6010	Influenza virus H5N1 infection of U251 astrocyte cell line: time course	<i>Homo sapiens</i>	GPL6480	GSE66597	18
GDS6000	High-fat diet effect on brown adipose tissue development	<i>Mus musculus</i>	GPL6887	GSE64718	33
GDS5948	Zipper-interacting protein kinase deficiency effect on coronary artery smooth muscle cells in vi...	<i>Homo sapiens</i>	GPL6244	GSE56819	6
GDS5914	YAP transcriptional regulator depletion effect on endothelial cells	<i>Homo sapiens</i>	GPL6244	GSE61989	12
GDS5913	SRPIN803 small molecule inhibitor of SRPK1 effect on retinal pigment epithelial cell line	<i>Homo sapiens</i>	GPL570	GSE62947	6
GDS5881	Nebulin deficiency effect on the soleus	<i>Mus musculus</i>	GPL6246	GSE70213	12

Below the table, a detailed record for DataSet GDS6177 is shown:

DataSet Record GDS6177: Expression Profiles Data Analysis Tools Sample Subsets			
Title:	Acute alcohol consumption effect on whole blood (control group): time course		
Summary:	Analysis of blood from subjects administered orange juice w/o alcohol. Blood collected at time points corresponding to collection times for the alcohol group in GDS4938. These results, together with those from GDS4938, provide insight into molecular response of blood during acute ethanol exposure.		
Organism:	<i>Homo sapiens</i>		
Platform:	GPL570: [HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array		
Citation:	Kupfer DM, White VL, Strayer DL, Crouch DJ et al. Microarray characterization of gene expression changes in blood during acute ethanol exposure. <i>BMC Med Genomics</i> 2013 Jul 25;6:26. PMID: 23883607		
Reference Series:	GSE20489	Sample count:	25
Value type:	transformed count	Series published:	2013/07/25

On the right side of the record details, there are links for "Cluster Analysis" (with a heatmap thumbnail) and "Download" (with links to various file formats: DataSet full SOFT file, DataSet SOFT file, Series family SOFT file, Series family MINIML file, Annotation SOFT file).

Getting GEO expression data

Easiest: Use curated data sets

<https://www.ncbi.nlm.nih.gov/sites/GDSbrowser/>

- [HLA-DRB4 - Acute alcohol consumption effect on whole blood \(control group\):](#)

1. [time course](#)

Annotation: **HLA-DRB4**, major histocompatibility complex, class II, DR beta 4

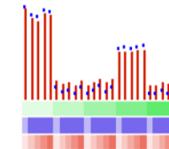
Organism: Homo sapiens

Reporter: **GPL570**, 215666_at (ID_REF), **GDS6177**, 3126 (Gene ID), **U70544**

DataSet type: Expression profiling by array, transformed count, 25 samples

ID: 132643760

[GEO DataSets](#) [Gene](#) [UniGene](#) [Profile neighbors](#)



- [TMEM176B - Acute alcohol consumption effect on whole blood \(control group\):](#)

2. [time course](#)

Annotation: **TMEM176B**, transmembrane protein 176B

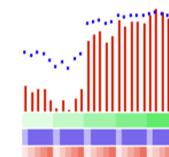
Organism: Homo sapiens

Reporter: **GPL570**, 220532_s_at (ID_REF), **GDS6177**, 28959 (Gene ID), **NM_014020**

DataSet type: Expression profiling by array, transformed count, 25 samples

ID: 132648617

[GEO DataSets](#) [Gene](#) [UniGene](#) [Profile neighbors](#) [Chromosome neighbors](#) [Homologene neighbors](#)



- [TMEM176A - Acute alcohol consumption effect on whole blood \(control group\):](#)

3. [time course](#)

Annotation: **TMEM176A**, transmembrane protein 176A

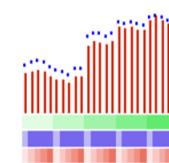
Organism: Homo sapiens

Reporter: **GPL570**, 218345_at (ID_REF), **GDS6177**, 55365 (Gene ID), **NM_018487**

DataSet type: Expression profiling by array, transformed count, 25 samples

ID: 132646431

[GEO DataSets](#) [Gene](#) [UniGene](#) [Profile neighbors](#) [Chromosome neighbors](#) [Homologene neighbors](#)



- [ZFP57 - Acute alcohol consumption effect on whole blood \(control group\): time](#)

4. [course](#)

Annotation: **ZFP57**, ZFP57 zinc finger protein

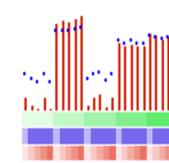
Organism: Homo sapiens

Reporter: **GPL570**, 231236_at (ID_REF), **GDS6177**, 346171 (Gene ID)

DataSet type: Expression profiling by array, transformed count, 25 samples

ID: 132659291

[GEO DataSets](#) [Gene](#) [UniGene](#) [Profile neighbors](#) [Chromosome neighbors](#) [Homologene neighbors](#)



Getting GEO expression data

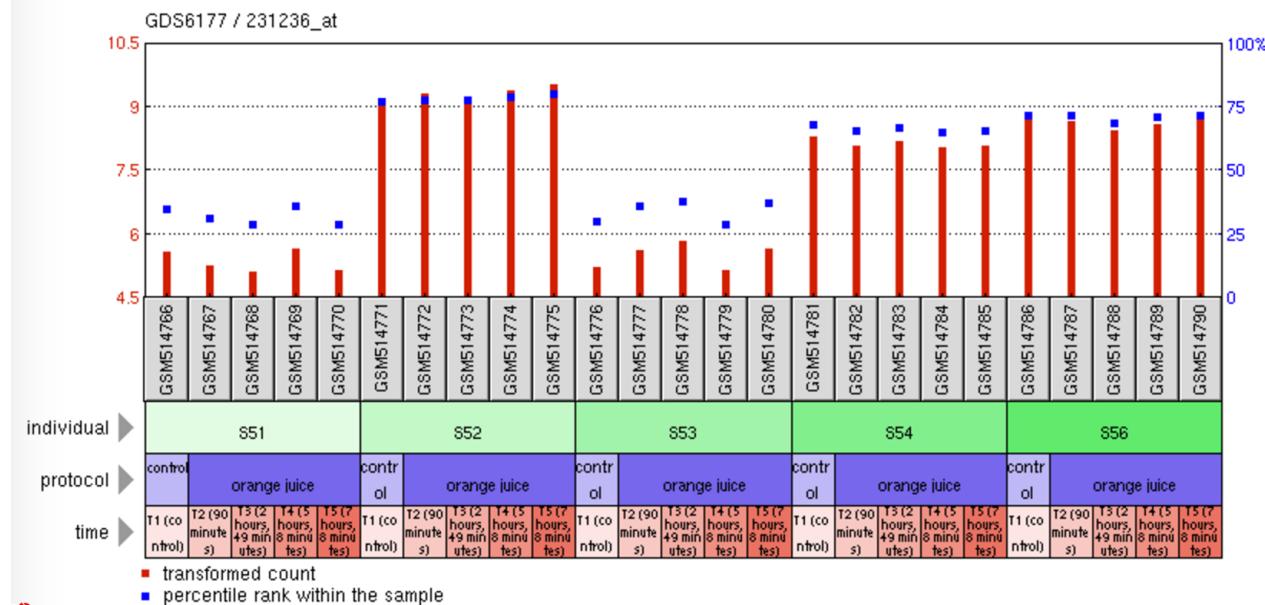
Easiest: Use curated data sets

<https://www.ncbi.nlm.nih.gov/sites/GDSbrowser/>

Profile GDS6177 / 231236_at

Title Acute alcohol consumption effect on whole blood (control group): time course

Organism Homo sapiens



Graph caption help

Sample	Title	Value	Rank
<u>GSM514766</u>	Blood_OJcontrol_T1_S51	5.61881	35
<u>GSM514767</u>	Blood_OJcontrol_T2_S51	5.28791	31
<u>GSM514768</u>	Blood_OJcontrol_T3_S51	5.13084	29

Getting GEO expression data

Otherwise: Get processed data from GSM pages...

Supplementary file	Size	Download	File type/resource
GSM794088_TAHG20110218_252456810085_S01_GE2-v5_95_Feb07_1_1.txt.gz	32.9 Mb	(ftp)(http)	TXT
GSM794088_TAHG20110218_252456810085_S01_GE2-v5_95_Feb07_1_2.txt.gz	32.8 Mb	(ftp)(http)	TXT
GSM794088_rna_lograt_zscore.txt.gz	1.3 Mb	(ftp)(http)	TXT

Raw data provided as supplementary file

Processed data provided as supplementary file

Raw

Raw

Processed



GEO isn't JUST about gene expression...

Platforms (1) [GPL17021](#) Illumina HiSeq 2500 (Mus musculus)

Samples (39) [GSM2143252](#) 114_FCX_120m_Control_input

[More...](#)

[GSM2143253](#) 115_FCX_120m_Treatment_input

[GSM2143254](#) 117_FCX_120m_Control_H3K27ac

This SubSeries is part of SuperSeries:

[GSE80345](#) Transcriptional regulatory dynamics set the stage for a coordinated metabolic and neural response to social threat in mice

Relations

BioProject [PRJNA320640](#)

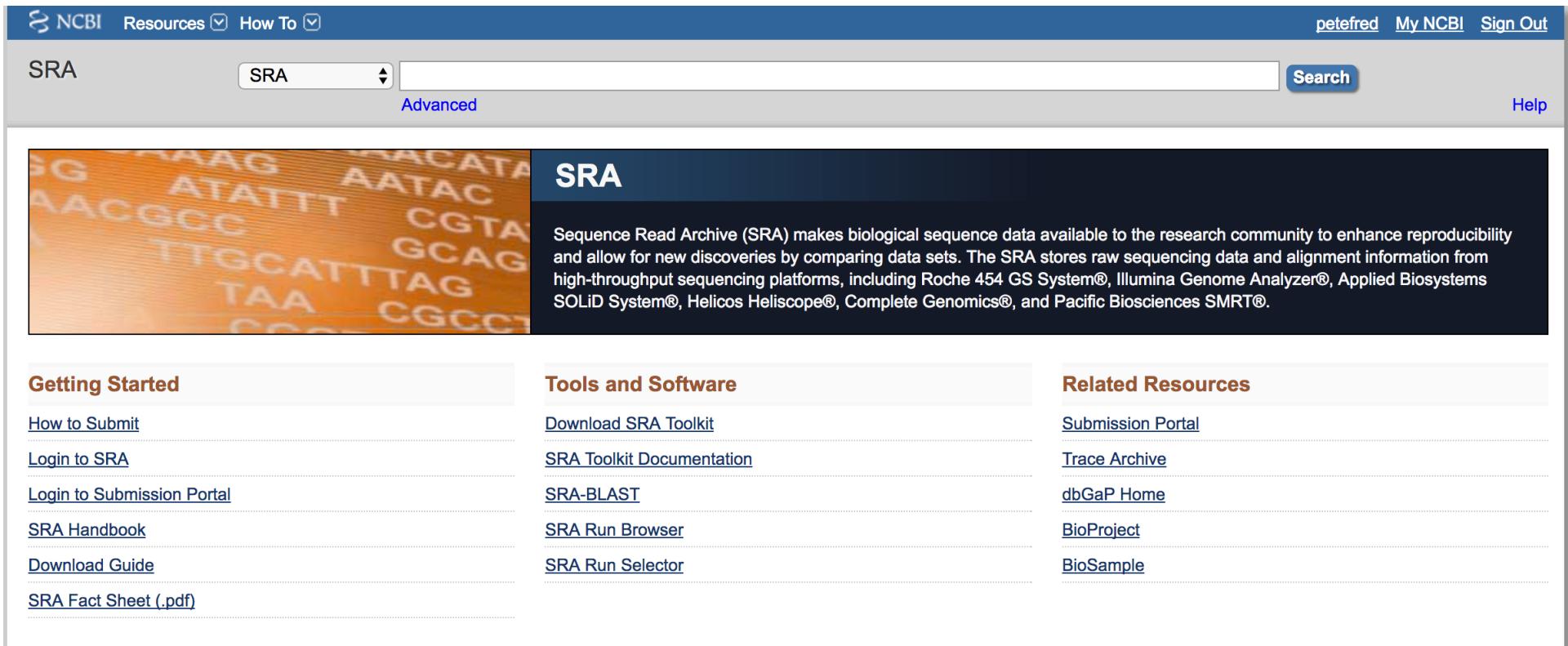
SRA [SRP074385](#)

Download family	Format
SOFT formatted family file(s)	SOFT <small>?</small>
MINiML formatted family file(s)	MINiML <small>?</small>
Series Matrix File(s)	TXT <small>?</small>

Supplementary file	Size	Download	File type/resource
GSE81122_114_FCX_120min_CK_input-117+118.ucsc.bigWig	133.5 Mb	(ftp) (http)	BIGWIG
GSE81122_115_FCX_120min_EX_input-119+120.ucsc.bigWig	163.6 Mb	(ftp) (http)	BIGWIG
GSE81122_117+118_FCX_120min_CK_1M_H3K27ac.ucsc.bigWig	317.5 Mb	(ftp) (http)	BIGWIG
GSE81122_119+120_FCX_120min_EX_1M_H3K27ac.ucsc.bigWig	242.3 Mb	(ftp) (http)	BIGWIG
GSE81122_121+122_Amy_CK_120min_1M_H3K27ac.ucsc.bigWig	351.1 Mb	(ftp) (http)	BIGWIG
GSE81122_125_Amy_CK_120min_0.1M_input.ucsc.bigWig	173.9 Mb	(ftp) (http)	BIGWIG
GSE81122_126+127_Amy_Exp_120min_1M_H3K27ac.ucsc.bigWig	352.7 Mb	(ftp) (http)	BIGWIG

Raw data available via the SRA

<https://www.ncbi.nlm.nih.gov/sra/>



The screenshot shows the NCBI SRA homepage. At the top, there's a navigation bar with links for NCBI, Resources, How To, petefred, My NCBI, and Sign Out. Below the navigation is a search bar with "SRA" selected in a dropdown, a search input field, a "Search" button, and an "Advanced" link. The main content area features a large image of a DNA sequence with various bases (A, T, C, G) highlighted in different colors. To the right of the image, the word "SRA" is displayed in large white letters. A descriptive text block explains the purpose of the SRA: "Sequence Read Archive (SRA) makes biological sequence data available to the research community to enhance reproducibility and allow for new discoveries by comparing data sets. The SRA stores raw sequencing data and alignment information from high-throughput sequencing platforms, including Roche 454 GS System®, Illumina Genome Analyzer®, Applied Biosystems SOLiD System®, Helicos Heliscope®, Complete Genomics®, and Pacific Biosciences SMRT®." Below this, there are three columns of links: "Getting Started" (How to Submit, Login to SRA, Login to Submission Portal, SRA Handbook, Download Guide, SRA Fact Sheet (.pdf)), "Tools and Software" (Download SRA Toolkit, SRA Toolkit Documentation, SRA-BLAST, SRA Run Browser, SRA Run Selector), and "Related Resources" (Submission Portal, Trace Archive, dbGaP Home, BioProject, BioSample).

Raw data available via the SRA

 Sequence Read Archive

Main Browse Search Download Submit Documentation Software Trace Archive Trace Assembly Trace BLAST

Studies Samples Analyses Run Browser Run Selector Provisional SRA

Massive transcriptional start site mapping of human fetal brain cells.

Identifiers:	SRA: DRP000023 BioProject: PRJDA34559 UT-MGS: DRP000023	Related SRA data
Study Type:	Transcriptome Analysis	Experiments: 1
Submission:	DRA000023	Runs: 4 (880.5Mbp; 2.5Gb)
Abstract:	Comprehensive identification and characterization of the transcriptional start sites of human genes were carried out. For this purpose, we used our TSS-Seq method, in which next gene sequencing technology and our full-length cDNA library technology, oligo-capping were combined.	
Description:	Although recent studies have revealed that the majority of human genes are subjected to regulation of alternative promoters (APs), the biological relevance of this phenomenon remains unclear. To enable more comprehensive TSS analysis in the respective cell types, we recently developed a method, combining oligo-capping with the massively parallel sequencing technology, Illumina GA. In this method, which we named TSS Seq, sequence adaptor which is necessary for Illumina GA sequencing is directly introduced to the cap site of the mRNA. By sequencing 36-48 sequence immediately downstream of the TSSs (TSS tags), it is possible to obtain precise positional information of transcriptional start sites (TSSs). In this paper, we used the TSS tag data accumulated from twelve different cell types and normal tissues in humans for the identification and characterization of the APs in human genes.	
Center Project:	Integrateve Transcriptome Analysis	
External Link:	DBTSS	

Raw data available via the SRA

Processed data often available through GEO link

 Sequence Read Archive

Main Browse Search Download Submit Documentation Software Trace Archive Trace Assembly Trace BLAST

Studies Samples Analyses Run Browser Run Selector Provisional SRA

Histone modification (H3K4me3 and H3K27me3) during vascular endothelial cell differentiation from mouse embryonic stem cells

Identifiers:	SRA: SRP099437 BioProject: PRJNA374539 GEO: GSE94828		Related SRA data Experiments: 20 Runs: 20 (22.1Gbp; 12.6Gb)
Study Type:	Other		
Submission:	SRA537391		
Abstract:	Although studies of the differentiation from mouse embryonic stem (ES) cells to vascular endothelial cells (ECs) provide an excellent model for investigating the molecular mechanisms underlying vascular development, temporal dynamics of gene expression and chromatin modifications have not been well studied. Herein, using transcriptomic and epigenomic analyses based on the H3K4me3 and H3K27me3 modifications at a genome-wide scale, we analyzed the EC differentiation steps from ES cells and crucial epigenetic modifications unique to ECs. We determined that Gata2, Fli1, Sox7, and Sox18 are master regulators of EC induced following expression of the hemangioblast commitment pioneer factor, Etv2. These master regulator gene loci were repressed by H3K27me3 under the mesoderm period, but rapidly transitioned to the histone modification switching from H3K27me3 to H3K4me3 after treatment with vascular		

Outline

- Summary of NGS technologies (sequencing and applications)
- Introduction to NGS data analysis
- Commonly available databases
- **Workflow integration and making use of existing NGS data**

Simplest: Download processed data
and view in spreadsheet

Simplest: Download processed data and view in spreadsheet

Example: gene_exp.diff from cufflinks

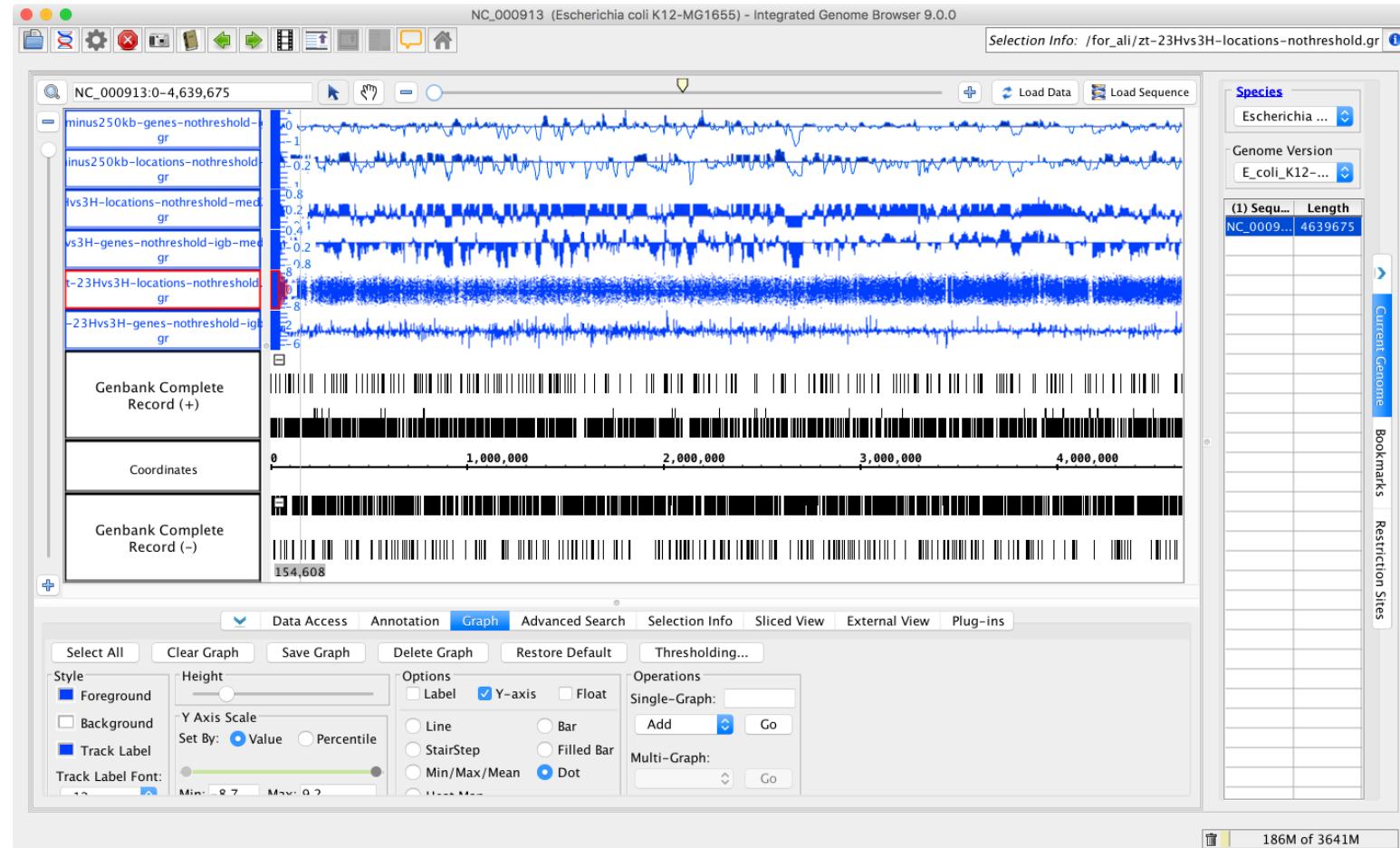
test_id	gene_id	gene	locus	sample_1	sample_2	status	value_1	value_2	log2(fold_change)	test_stat	p_value	q_value	sig
XLOC_00001	XLOC_00001	CG11023	2L: 7528-948	cupcake_sated	cupcake_hungry	OK	5.49313	0.206789	-4.7314	-1.90088	0.42235	0.593877	no
XLOC_00002	XLOC_00002	Ir21a	2L: 21918-25	cupcake_sated	cupcake_hungry	OK	2106.08	2913.7	0.468291	2.53576	0.85965	0.903137	no
XLOC_00031	XLOC_00031	dbr	2L: 67043-71	cupcake_sated	cupcake_hungry	OK	14.9389	16.8551	0.174119	0.230548	0.67695	0.781675	no
XLOC_00032	XLOC_00032	galectin	2L: 72387-76	cupcake_sated	cupcake_hungry	OK	112.48	85.6742	-0.39273	-0.67515	0.24695	0.423915	no
XLOC_00033	XLOC_00033	CG11374	2L: 76445-77	cupcake_sated	cupcake_hungry	OK	6.13796	14.0256	1.19223	0.705069	0.22855	0.416548	no
XLOC_00034	XLOC_00034	-	2L: 80193-80	cupcake_sated	cupcake_hungry	OK	0	402.552	inf	#NAME?	0.01965	0.158954	no

Simplest: Download processed data and view in spreadsheet

Example: gene_exp.diff from cufflinks

test_id	gene_id	gene	locus	sample_1	sample_2	status	value_1	value_2	log2(fold_change)	test_stat	p_value	q_value	sig
XLOC_002456	XLOC_002456	TepIV	2L: 19549792 -1955645	cupcake_sated	cupcake_hungry	OK	27.2788	0.762464	-5.16097	-3.36415	5.00E-05	0.004022	yes
XLOC_004017	XLOC_004017	dp	2L: 4479470- 4591963	cupcake_sated	cupcake_hungry	OK	1.66149	0.451918	-1.87835	-2.1873	0.00045	0.024242	yes
XLOC_008185	XLOC_008185	Cam	2R: 8146912- 8166208	cupcake_sated	cupcake_hungry	OK	2811.67	2026.02	-0.47278	-1.67712	0.0034	0.034841	yes

Genome Browsers



Allow loading and comparisons of various data sets with genomic features

Main candidates: IGB, IGV, UCSC Genome Browser

Genome Browsers



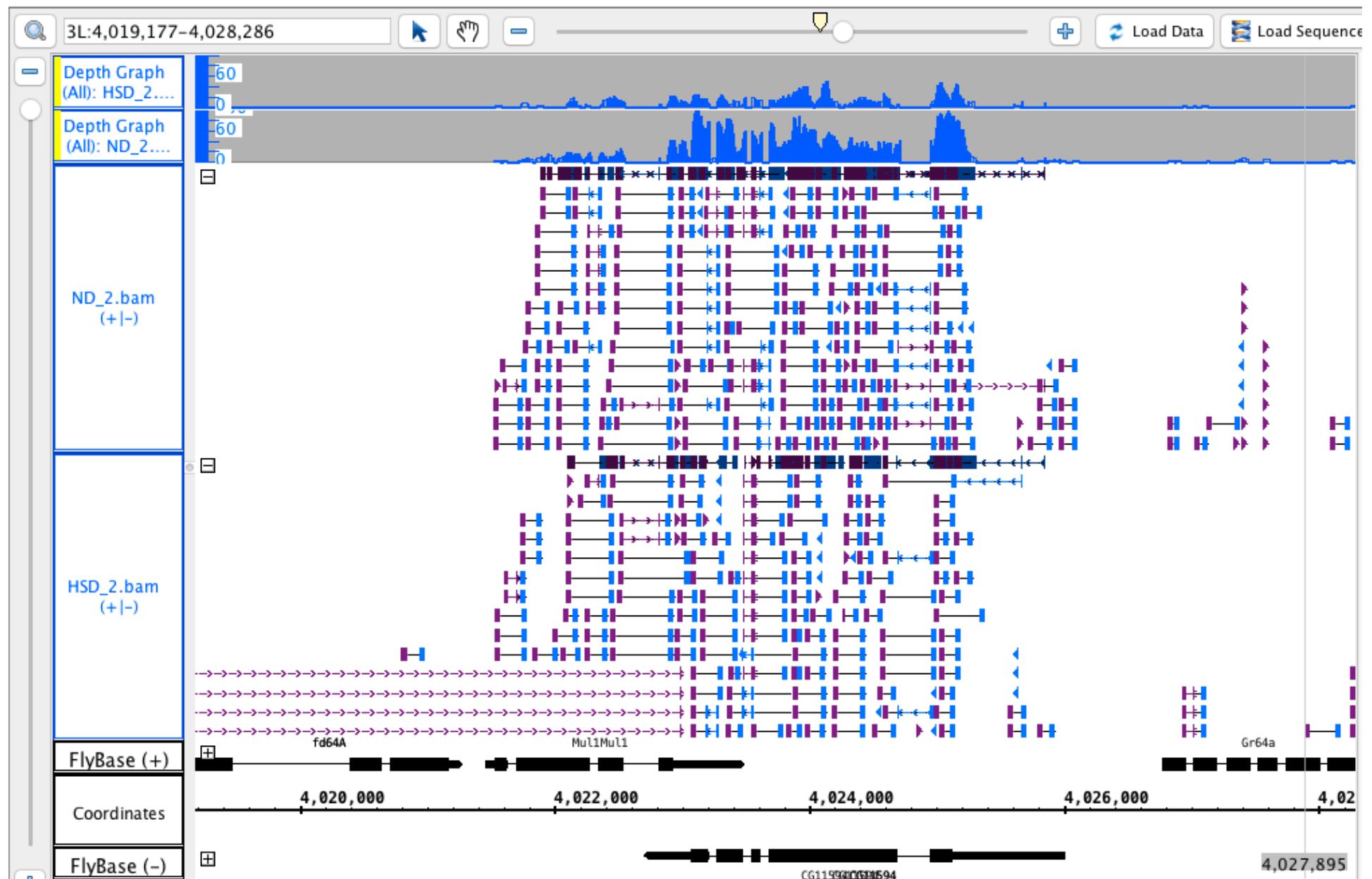
Analysis can scale from raw reads to highly processed functions of multiple data sets

Genome Browsers



Analysis can scale from raw reads to highly processed functions of multiple data sets

Genome Browsers



Analysis can scale from raw reads to highly processed functions of multiple data sets

Genome Browsers

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Scope: Self Format: HTML Amount: Quick GEO accession: GSE87509 GO

Series GSE87509 Query DataSets for GSE87509

Status	Public on Mar 16, 2017
Title	ChIP-seq of Atrophin in Drosophila S2 cells
Organism	Drosophila melanogaster
Experiment type	Genome binding/occupancy profiling by high throughput sequencing
Summary	Drosophila Atro mutants have a large range of phenotypes, including neurodegeneration, segmentation, patterning and planar polarity defects. Although Atro mutants have diverse phenotypes, little is known about Atro's binding partners and downstream targets. We present the first genomic analysis of Atro using ChIP-seq against endogenous Atro. These data sets will serve as a valuable resource for future studies on Atro.
Overall design	We performed three independent biological replicates of Atro ChIP-seq experiments in untreated S2 cells. A corresponding non-specific IgG control ChIP was performed with each Atro ChIP-seq and was used as a control.

Many GEO files are directly loadable

Genome Browsers

• • •

Download family	Format
SOFT formatted family file(s)	SOFT ?
MINiML formatted family file(s)	MINiML ?
Series Matrix File(s)	TXT ?

Supplementary file	Size	Download	File type/resource
SRP/SRP090/SRP090681		(ftp)	SRA Study
GSE87509_RAW.tar	34.5 Mb	(http) (custom)	TAR (of WIG)

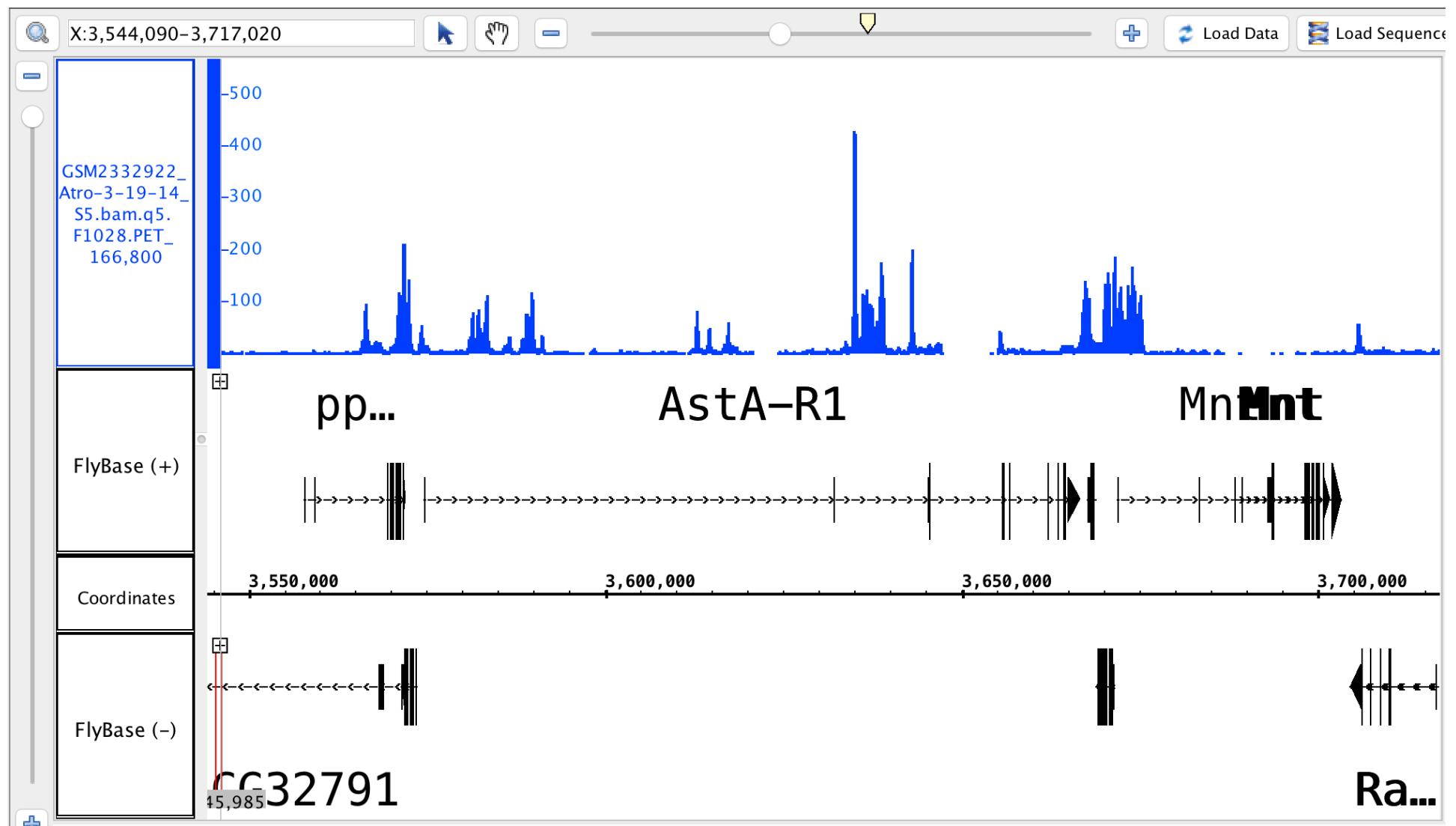
Raw data provided as supplementary file

Processed data provided as supplementary file



Many GEO files are directly loadable

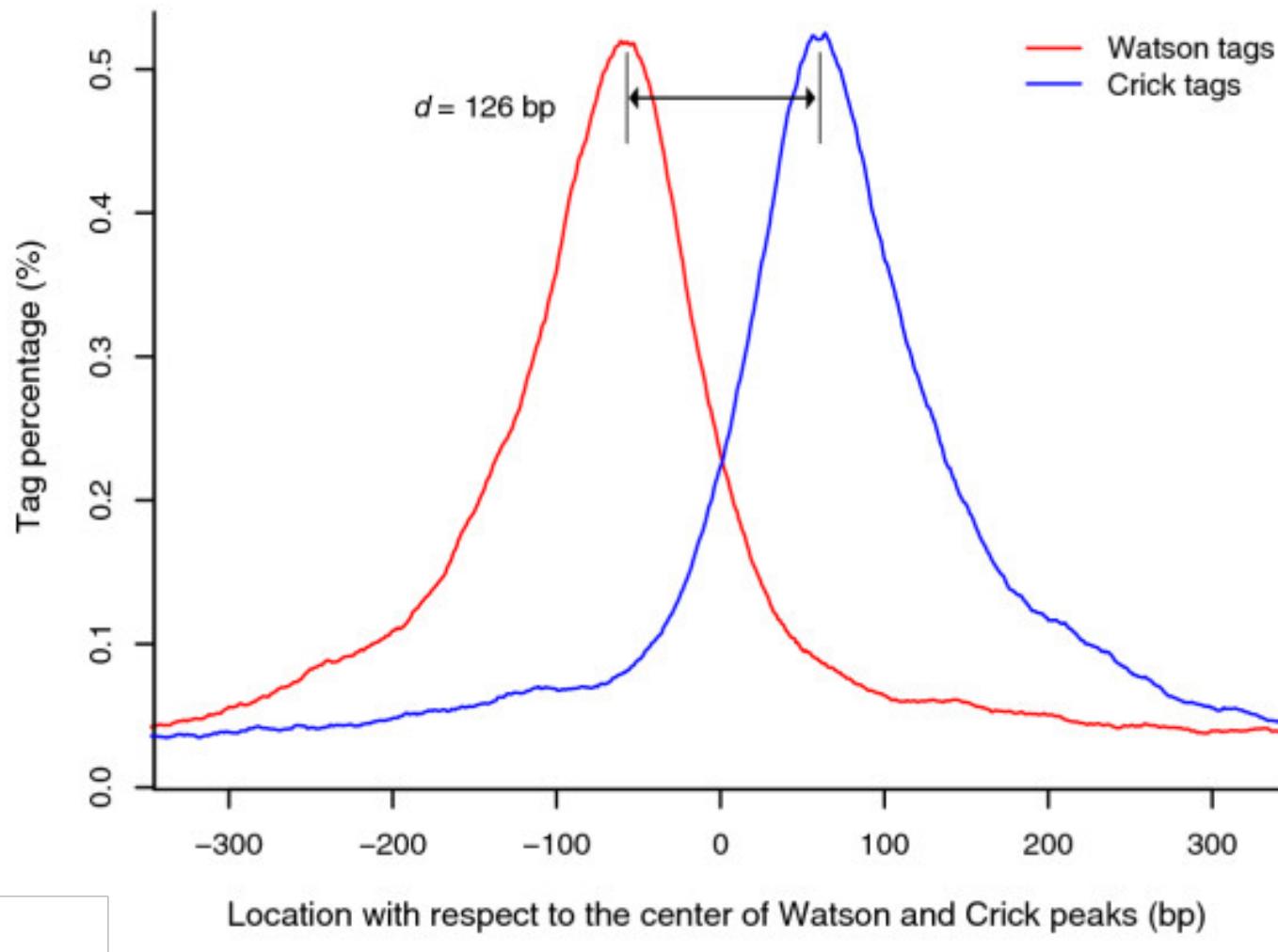
Genome Browsers



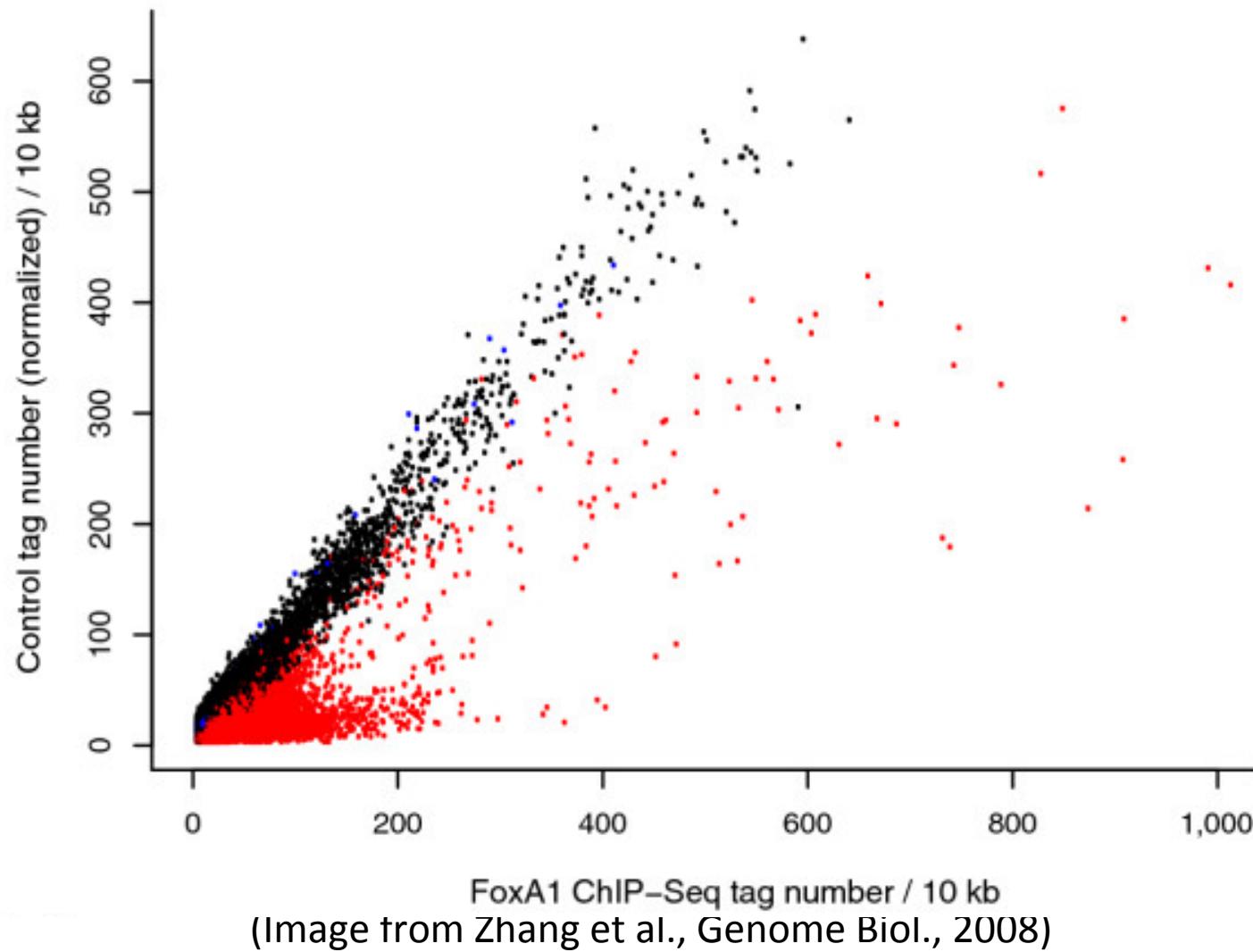
Many GEO files are directly loadable

Peak calling or differential calling are common tasks

Peak calling or differential calling are common tasks



Peak calling or differential calling are common tasks



So what do you do once you have peaks/expression calls/etc.?

- Direct inspection of known biological targets
- Literature-driven inference and hypothesis generation
- Gene set enrichment analysis
- Motif analysis
- Network inference

Gene set enrichment analysis

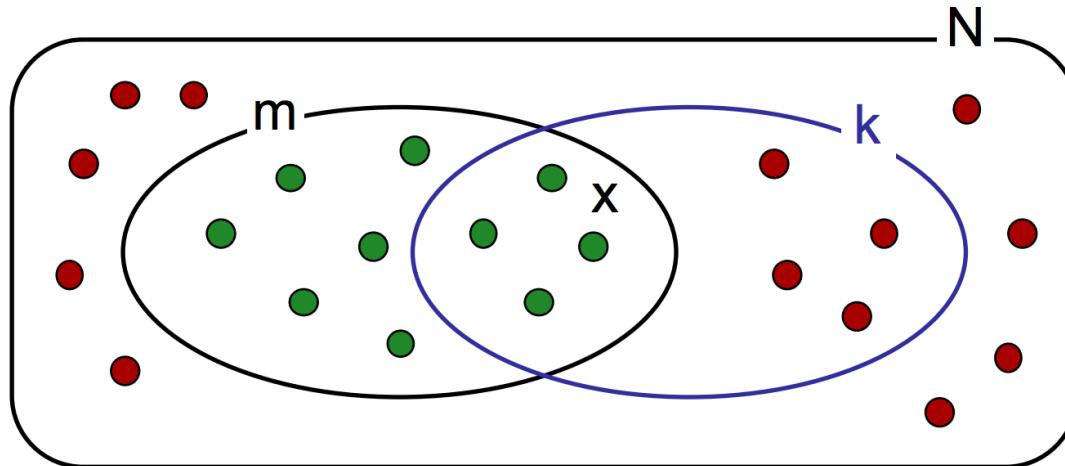
Identification of gene categories (e.g., GO terms)
that are correlated with another data set

Common Tools: GSEA, DAVID, iPAGE

Gene set enrichment analysis

Identification of gene categories (e.g., GO terms) that are correlated with another data set

Common Tools: GSEA, DAVID, iPAGE



N = total number of elements

m = number of marked elements

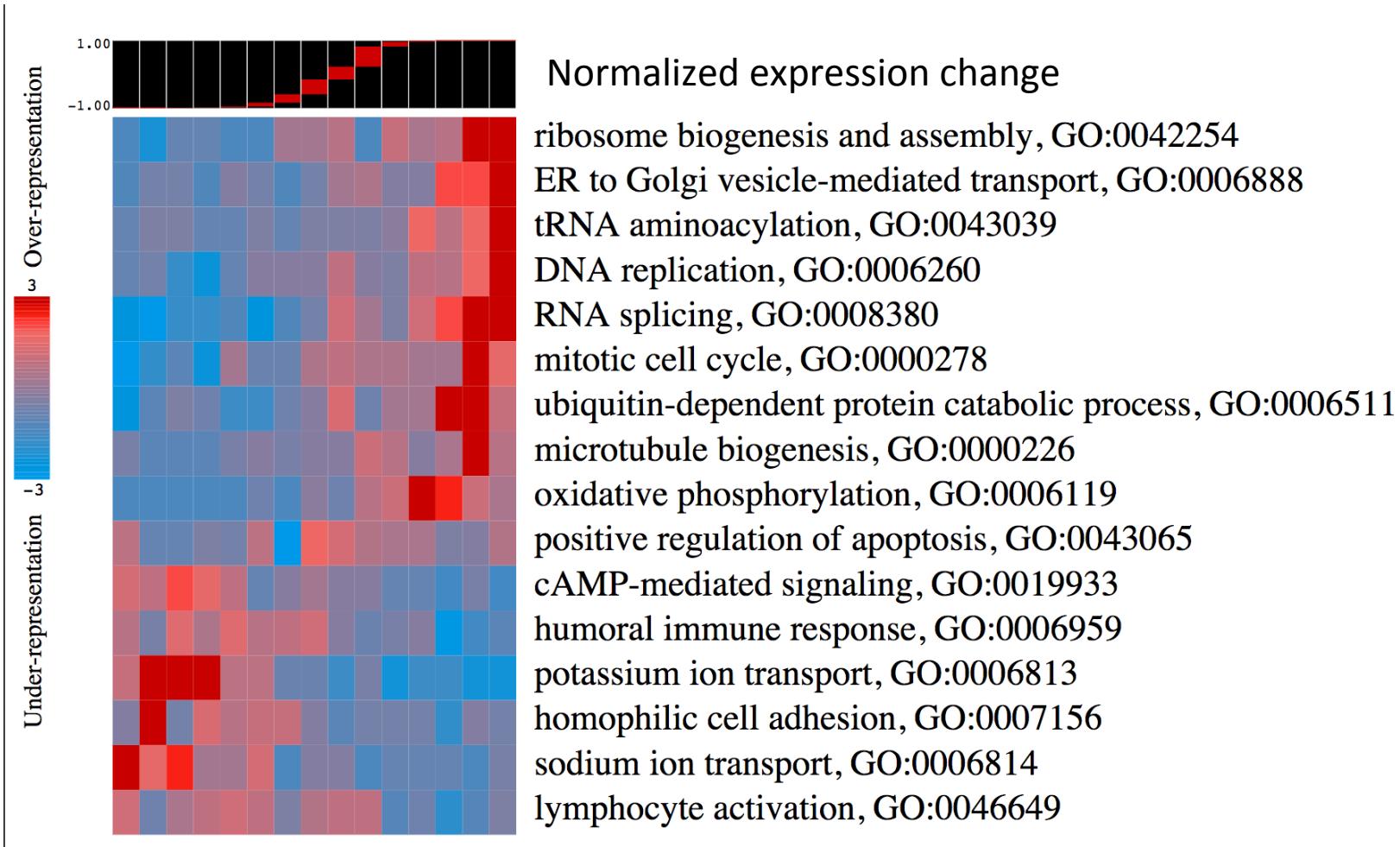
k = number of sampled elements

x = number of marked sampled elements

Gene set enrichment analysis

Identification of gene categories (e.g., GO terms) that are correlated with another data set

Example: Gene expression

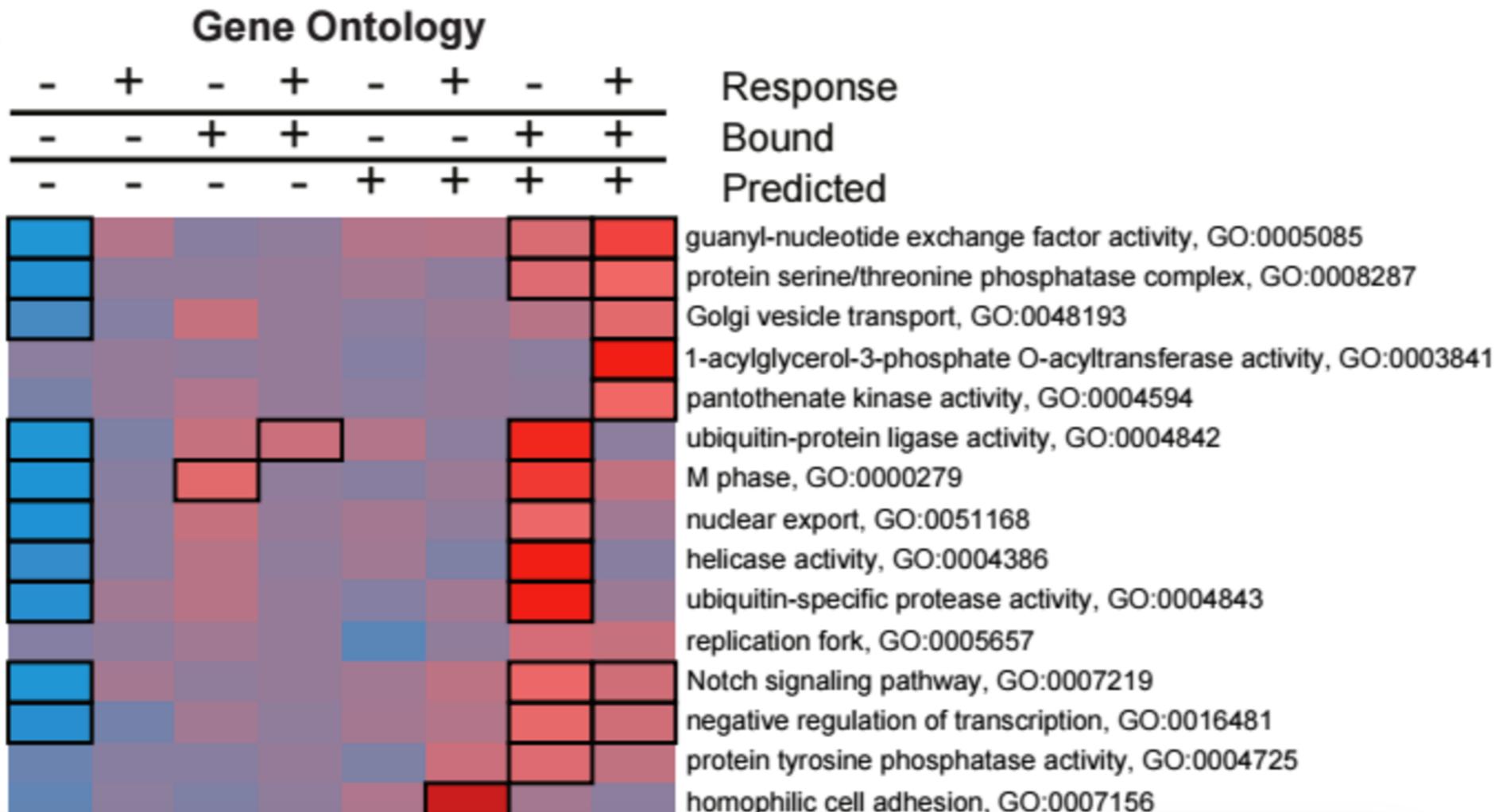


(From Goodarzi et al., Mol. Cell, 2009)

Gene set enrichment analysis

Identification of gene categories (e.g., GO terms) that are correlated with another data set

Example: Integration of data sets



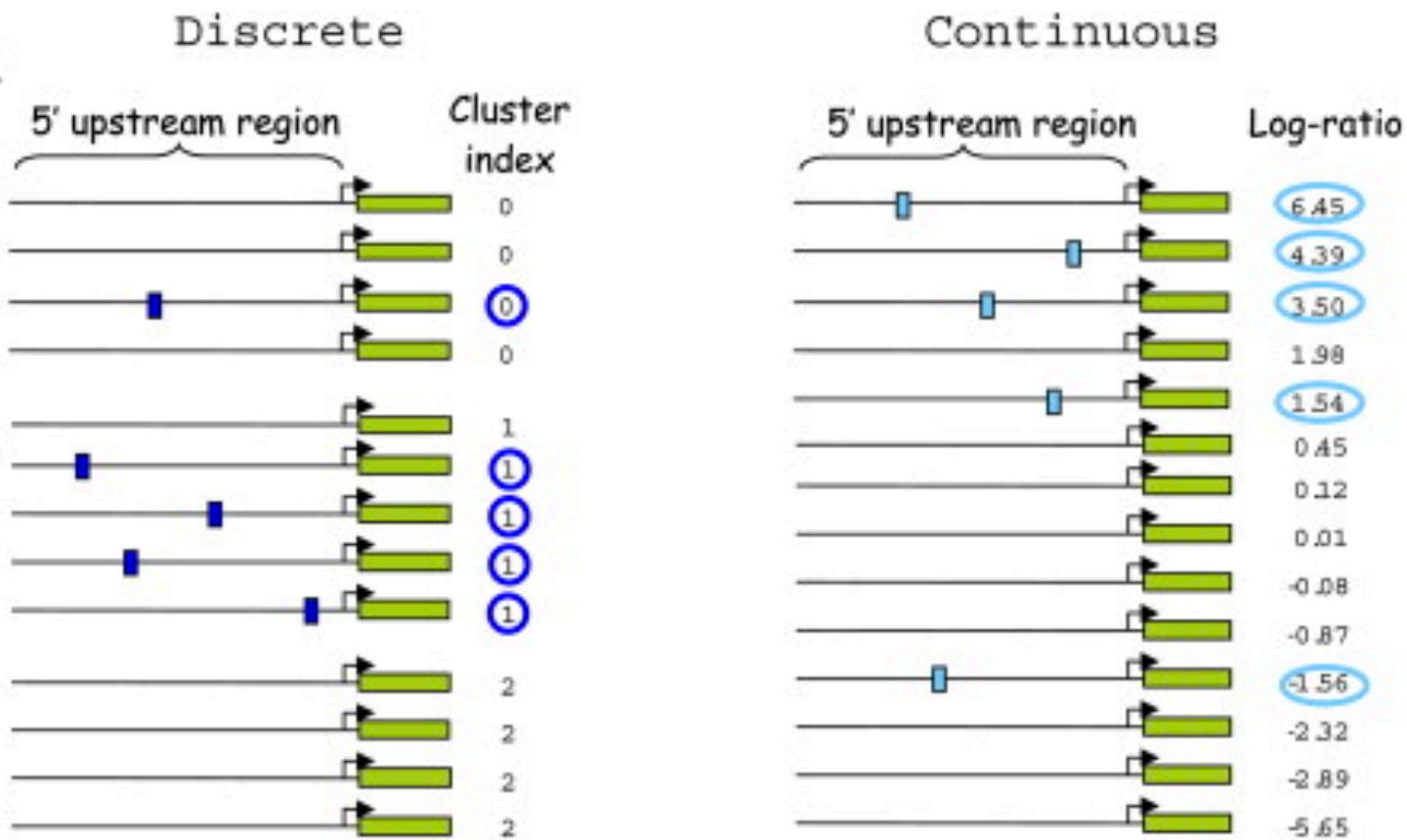
Motif analysis

Identify motifs (typically nucleic acid sequences) correlated with a data set of interest

Used in a variety of applications (RNA-seq, ChIP-seq, ribosome profiling, etc.)

Example tools: MEME suite, FIRE/TEISER, kmersvm

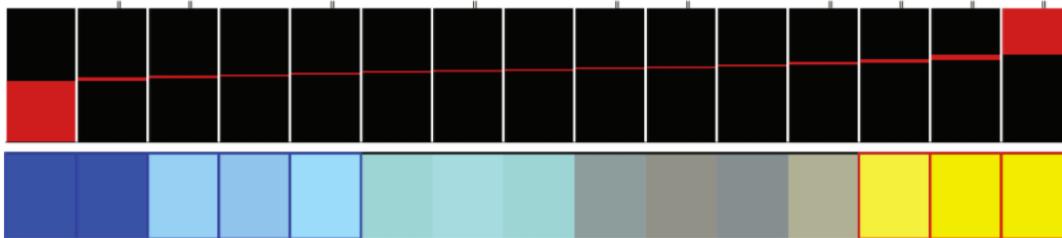
Motif analysis



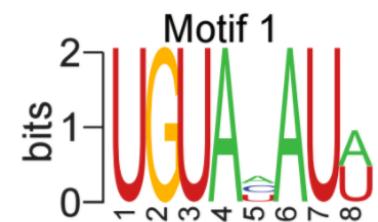
(Image from Elemento *et al.*, Mol. Cell 2007)

Motif analysis

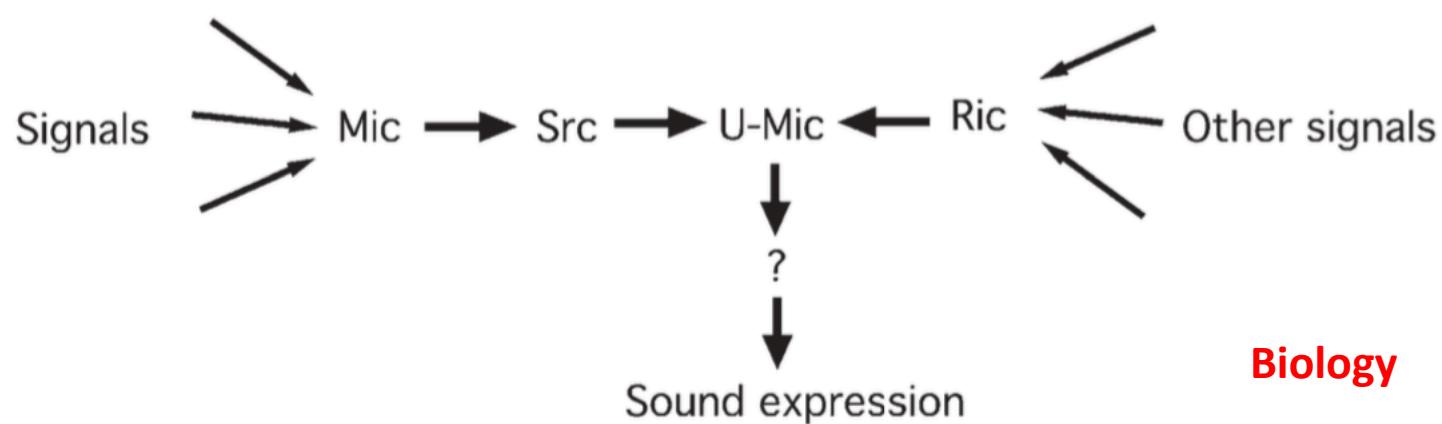
\log_2 fold change



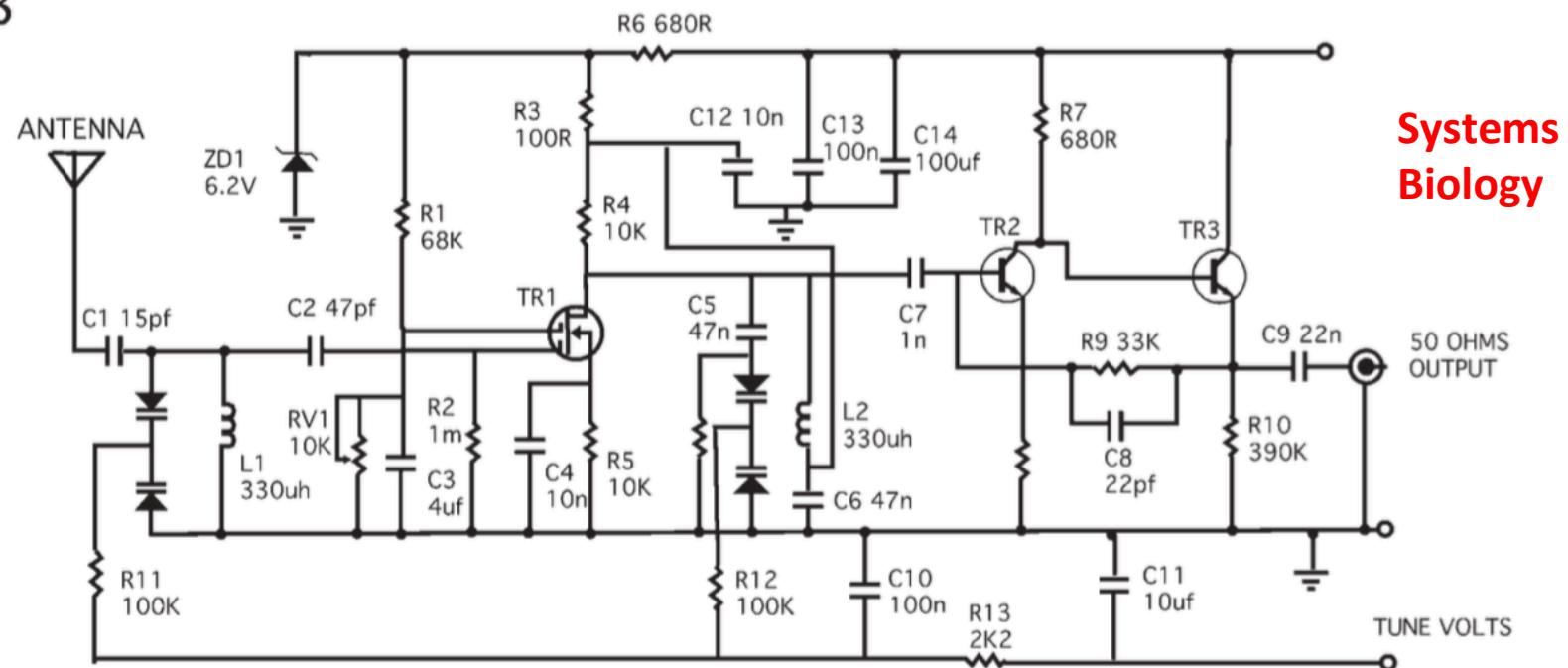
Motif 1



A



B



Lazebnik, Y. Cancer Cell 2002
(slides via Michael Wolfe)