

# HomeWork6

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## Section 1: Improving analysis code by writing functions

```
# Can you improve this analysis code?
library(bio3d)
# s1 <- read.pdb("4AKE") # kinase with drug
# s2 <- read.pdb("1AKE") # kinase no drug
# s3 <- read.pdb("1E4Y") # kinase with drug
# s1.chainA <- trim.pdb(s1, chain="A", elety="CA")
# s2.chainA <- trim.pdb(s2, chain="A", elety="CA")
# s3.chainA <- trim.pdb(s1, chain="A", elety="CA")
# s1.b <- s1.chainA$atom$b
# s2.b <- s2.chainA$atom$b
# s3.b <- s3.chainA$atom$b
# plotb3(s1.b, sse=s1.chainA, typ="l", ylab="Bfactor")
# plotb3(s2.b, sse=s2.chainA, typ="l", ylab="Bfactor")
# plotb3(s3.b, sse=s3.chainA, typ="l", ylab="Bfactor")

#improving copy paste issues:
s1 <- read.pdb("4AKE") # kinase with drug
```

B.

```
## Note: Accessing on-line PDB file
```

```
s2 <- read.pdb("1AKE") # kinase no drug
```

```
## Note: Accessing on-line PDB file
```

```
## PDB has ALT records, taking A only, rm.alt=TRUE
```

```
s3 <- read.pdb("1E4Y") # kinase with drug
```

```
## Note: Accessing on-line PDB file
```

```
s1.chainA <- trim.pdb(s1, chain="A", elety="CA")
```

```
s2.chainA <- trim.pdb(s2, chain="A", elety="CA")
```

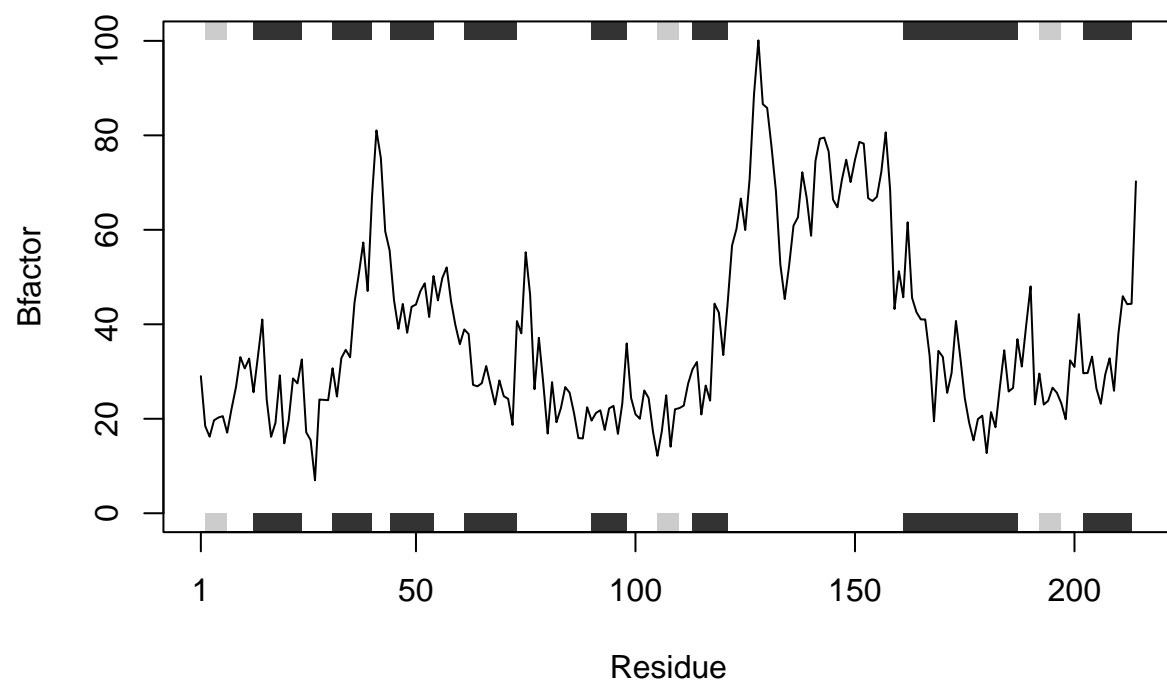
```
s3.chainA <- trim.pdb(s3, chain="A", elety="CA")
```

```
s1.b <- s1.chainA$atom$b
```

```
s2.b <- s2.chainA$atom$b
```

```
s3.b <- s3.chainA$atom$b
```

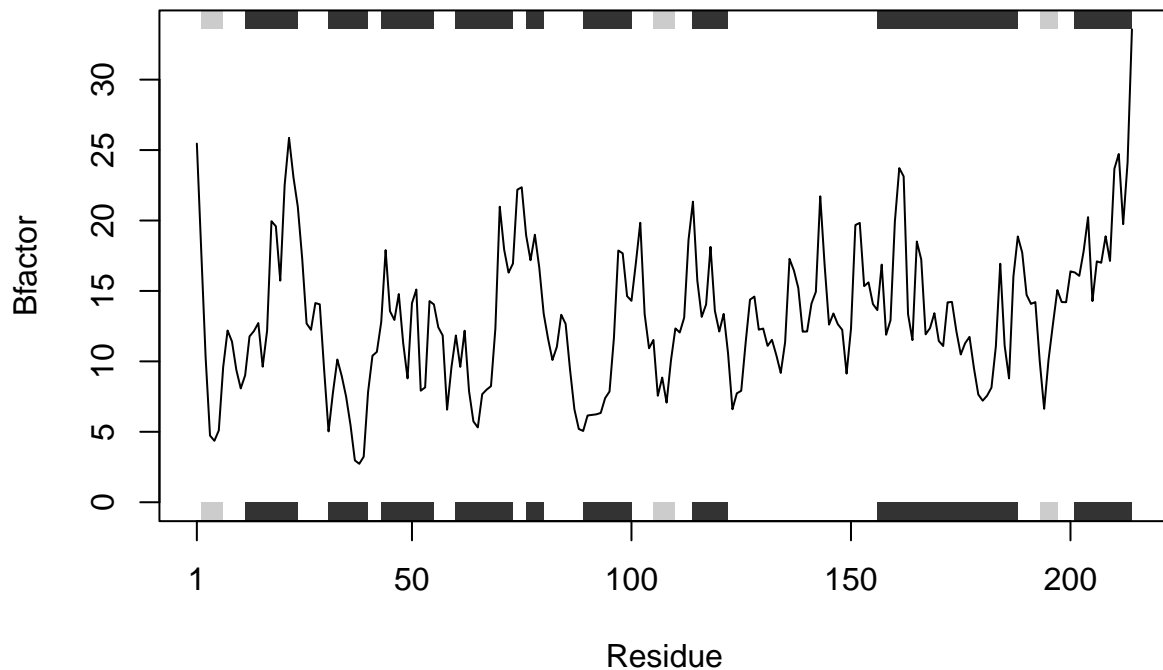
```
plotb3(s1.b, sse = s1.chainA, typ = "l", ylab = "Bfactor")
```



```
plotb3(s2.b, sse = s2.chainA, typ = "l", ylab = "Bfactor")
```



```
plotb3(s3.b, sse = s3.chainA, typ = "l", ylab = "Bfactor")
```



We want to create a function to be able to visualize protein-drug interactions from PDB data:

```
pr_dr <- function(file, chain, elmnt, fctr) {
  plot_colors <- c("red", "blue", "green")
  for (i in 1:length(file)) {
    s1 <- read.pdb(file[i])

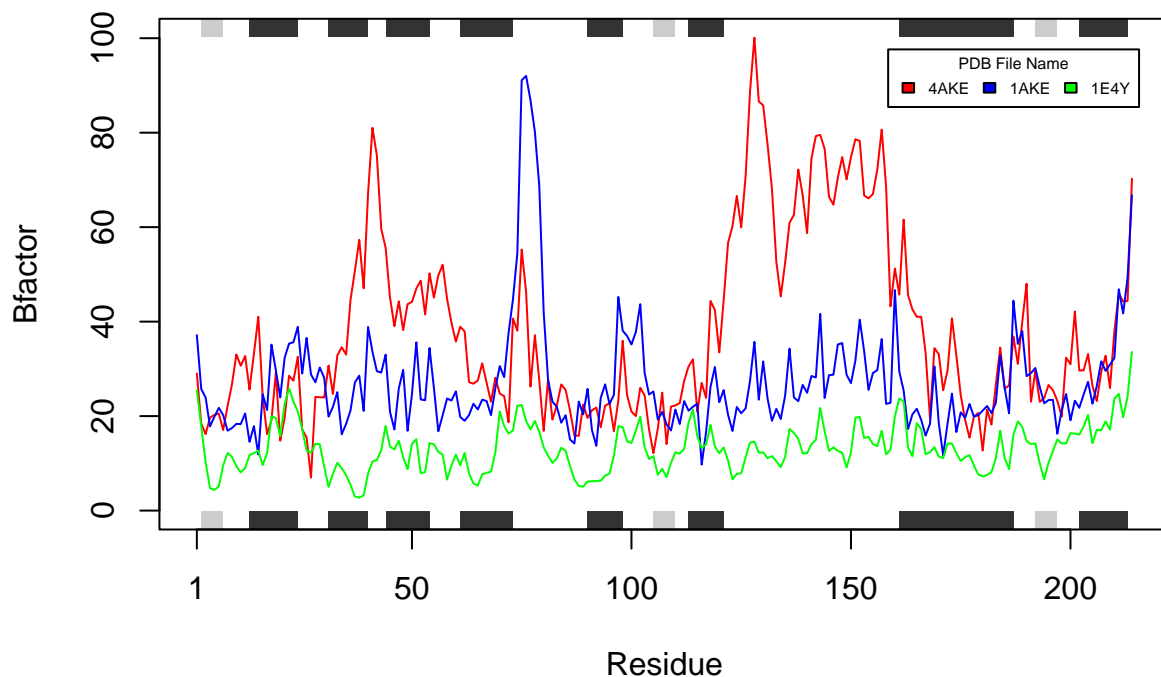
    s1.chain <- trim.pdb(s1, chain = chain, elety = elmnt)

    atom_df <- s1.chain$atom
    s1.fctr <- atom_df[, fctr]
    if (i == 1) {
      plotb3(s1.fctr, sse = s1.chain, typ = "l", ylab = paste(toupper(fctr), "factor", sep = ""), col = plot_colors[i])
    } else {
      lines(s1.fctr, col = plot_colors[i])
    }
  }
  legend("topright", title = "PDB File Name", file, fill = plot_colors, horiz=TRUE, cex = 0.5, inset = TRUE)
}

# Testing the function for theses 3 protein:
files <- c("4AKE", "1AKE", "1E4Y")
chains <- "A"
elements <- "CA"
factors <- "b"

pr_dr(files, chains, elements, factors)
```

```
## Note: Accessing on-line PDB file
## Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
## /var/folders/zx/ygntgn2j6lz4sh8bv9d1xsw00000gn/T//Rtmp1qwFKw/4AKE.pdb exists.
## Skipping download
## Note: Accessing on-line PDB file
## Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
## /var/folders/zx/ygntgn2j6lz4sh8bv9d1xsw00000gn/T//Rtmp1qwFKw/1AKE.pdb exists.
## Skipping download
## PDB has ALT records, taking A only, rm.alt=TRUE
## Note: Accessing on-line PDB file
## Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
## /var/folders/zx/ygntgn2j6lz4sh8bv9d1xsw00000gn/T//Rtmp1qwFKw/1E4Y.pdb exists.
## Skipping download
```



**Q1. What type of object is returned from the `read.pdb()` function?** This function returns information about the structure of the protein, its building block, and sequence of the protein from PDB

```
read.pdb("4AKE")
```

```
## Note: Accessing on-line PDB file
## Warning in get.pdb(file, path = tempdir(), verbose = FALSE):
## /var/folders/zx/ygntgn2j6lz4sh8bv9d1xsw00000gn/T//Rtmp1qwFKw/4AKE.pdb exists.
## Skipping download
##
```

```
## Call: read.pdb(file = "4AKE")
##
## Total Models#: 1
## Total Atoms#: 3459, XYZs#: 10377 Chains#: 2 (values: A B)
##
## Protein Atoms#: 3312 (residues/Calpha atoms#: 428)
## Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
## Non-protein/nucleic Atoms#: 147 (residues: 147)
## Non-protein/nucleic resid values: [ HOH (147) ]
##
## Protein sequence:
## MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
## DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
## VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG
## YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILGMRIILLGAPGA...<cut>...KILG
##
## + attr: atom, xyz, seqres, helix, sheet,
## calpha, remark, call
```

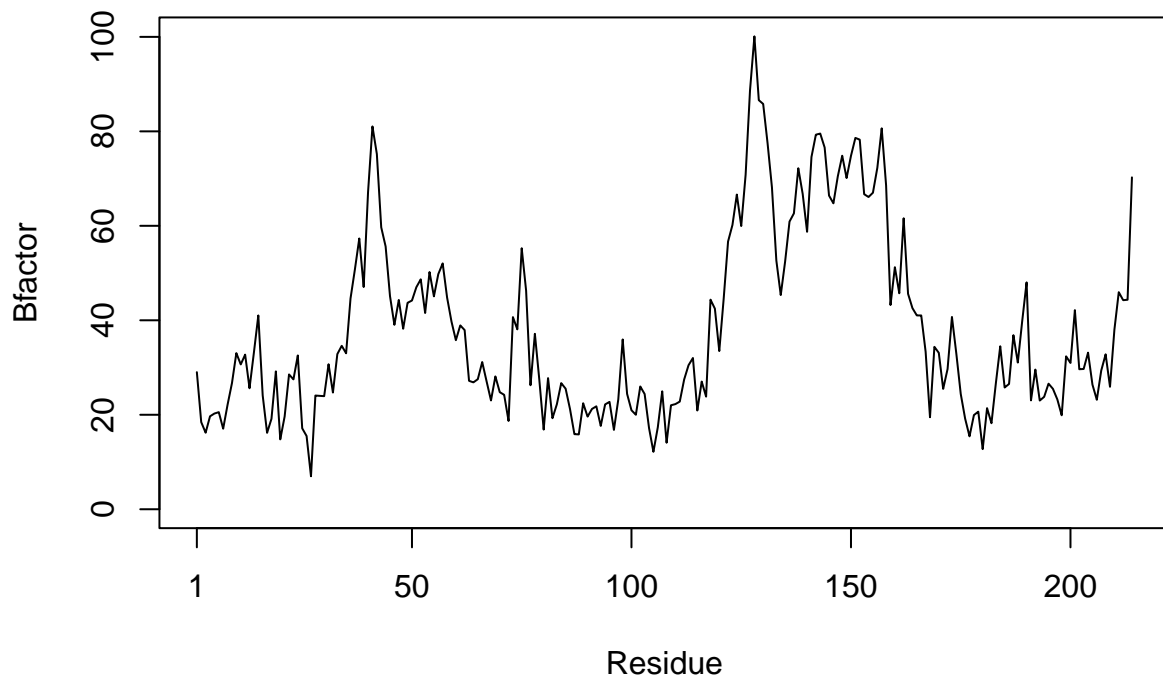
**Q2. What does the `trim.pdb()` function do?** This function trims the `pdb` object by selecting specific chains, residues, or atoms based on the provided parameters. In this case, `trim.pdb()` is used to extract a specific chain ("A") and element ("CA") from the `pdb` object.

```
trim.pdb(s1, chain="A", elety="CA")
```

```
##
## Call: trim.pdb(pdb = s1, chain = "A", elety = "CA")
##
## Total Models#: 1
## Total Atoms#: 214, XYZs#: 642 Chains#: 1 (values: A)
##
## Protein Atoms#: 214 (residues/Calpha atoms#: 214)
## Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
## Non-protein/nucleic Atoms#: 0 (residues: 0)
## Non-protein/nucleic resid values: [ none ]
##
## Protein sequence:
## MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
## DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
## VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG
## YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
##
## + attr: atom, helix, sheet, seqres, xyz,
## calpha, call
```

**Q3. What input parameter would turn off the marginal black and grey rectangles in the plots and what do they represent in this case?** To remove the black and grey rectangles, we need delete SSE(secondary structural element) from the chunk.

```
plotb3(s1.b, typ = "l", ylab = "Bfactor")
```



**Q4. What would be a better plot to compare across the different proteins?** a single plot with different colored lines representing each protein's B-factor trends which allows us to compare multiple proteins at the same time.

**Q5. Which proteins are more similar to each other in their B-factor trends. How could you quantify this?** From the clustering we can see that 1E4Y and 1AKE are the most similar ones.

```
bfactors <- rbind(s1.b, s2.b, s3.b)
dist_matrix <- dist(bfactors)
hclust_result <- hclust(dist_matrix)
plot(hclust_result)
```

## Cluster Dendrogram



```
dist_matrix
hclust (*, "complete")
```

Q6. How would you generalize the original code above to work with any set of input protein structures?

```
pr_dr <- function(file, chain, elmnt, fctr) {
  plot_colors <- c("red", "blue", "green")
  for (i in 1:length(file)) {
    s1 <- read.pdb(file[i])

    s1.chain <- trim.pdb(s1, chain = chain, elenty = elmnt)

    atom_df <- s1.chain$atom
    s1.fctr <- atom_df[, fctr]
    if (i == 1) {
      plotb3(s1.fctr, sse = s1.chain, typ = "l", ylab = paste(toupper(fctr), "factor", sep = ""), col = plot_colors[i])
    } else {
      lines(s1.fctr, col = plot_colors[i])
    }
  }
  legend("topright", title = "PDB File Name", file, fill = plot_colors, horiz=TRUE, cex = 0.5, inset = TRUE)
}
```