## 1. Summary

The protein encoded by this gene is proinflammatory cytokine of the IL-1 family that is constitutively found as a precursor within the cytoplasm of a variety of cells including macrophages and keratinocytes with 193 amino acids. Inactivity of Il18 precursor is processed to its active form by caspase-1 and is capable of stimulating interferon gamma production and regulating both TH1 and TH2 responses. Alternatively spliced transcript variants encoding different isoforms have been found for this gene. This gene has been reviewed for its involvement in Coronavirus biology. We wish to clone in to pPiCZAC vector for expressing a recombinant hIl-18 protein using a yeast expression system.

## References

- (Kato, Z., Jee, J., Shikano, H., Mishima, M., Ohki, I., Ohnishi, H., Li, A., Hashimoto, K., Matsukuma, E., Omoya, K., Yamamoto, Y., Yoneda, T., Hara, T., Kondo, N. and Shirakawa, M. (2003) The structure and binding mode of interleukin-18. Nat Struct Biol, )
- 2. (Wei, H., Wang, D., Qian, Y., Liu, X., Fan, S., Yin, H. S. and Wang, X. (2014) Structural basis for the specific recognition of IL-18 by its alpha receptor. FEBS Lett, 588, 3838-43.)
- (Le, H., Spearman, P., Waggoner, S. N. and Singh, K. (2022) Ebola virus protein VP40 stimulates IL-12- and IL-18-dependent activation of human natural killer cells. JCI Insight, 7.)

## 2. Gene structure, from ensemble:

		20.84 kb							Forward stran		
112 144%	112.146Mb	112 149Mb	112.150Mb	112.15296	112 154V6 n73 1	112.156ИЪ	112 158Mb	112.160Mb	112 163%b		
< 1.18-202 - ENSTO00 protein coding	00524595										
< IL18-203 - ENST000 retained intron	00525547	0			< IL18-206 - ENSTROCCOS retained intron	D					
< IL18-201 - ENSTO00 protein coding	00280357	_	-					10-01			
< IL18-204 - ENST protein coding	T10000528832	-	_								
< NL18-205 - ENS protein coding C	70000533858 DS not defined		_0_	0		0					
< IL18-201 - ENSTO00 protein coding	00280357		-						V		

 From ensemble, 5' and 3' untranslated regions in yellow highlights

.....

1
CCTTTGCTCCCCTGGCGACTGCCTGGACAGTCAGCAAGGAATTGT
CTCCCAGTGCATTTT 60
61
GCCCTCCTGGCTGCCAACTCTGGCTGCTAAAGCGGCTGCCACCTG
CTCCACTCTACACAC 120

121

CTTCGGGAAGAGGAAAGGAACCTCAGACCTTCCAGATCGCTTCCT
CTCGCAACAAACTAT 180

181

TTGTCGCAGGAATAAAGATGGCTGCTGAACCAGTAGAAGACAAT
TGCATCAACTTTGTGG 240
......ATGGCTGCTGAACCAGTAGAAGACAATTGCATCAACT
TTGTGG 43

.....-M--A--A--E--P--V--E--D--N--C--I--N--F--V-- 14

241

CAATGAAATTTATTGACAATACGCTTTACTTTATAGCTGAAGATGATGAAAACCTGGAAT 300

44

CAATGAAATTTATTGACAATACGCTTTACTTTATAGCTGAAGATG ATGAAAACCTGGAAT 103

15 A--M--K--F--I--D--N--T--L--Y--F--I--A--E--D--D--E--N--L--E--

301

CA<mark>GAT</mark>TAC<mark>TTT</mark>GGC<mark>AAG</mark>CTT<mark>GAA</mark>TCT<mark>AAA</mark>TTA<mark>TCA</mark>GTC<mark>ATA</mark>AGA<mark>A</mark> ATTTGAATGACCAAG 360

104

CAGATTACTTTGGCAAGCTTGAATCTAAATTATCAGTCATAAGAA ATTTGAATGACCAAG 163

35 S--D--Y--F--G--K--L--E--S--K--L--S--V--I--R--N--L--N--D--Q--54

361

TTCTCTTCATTGACCAAGGAAATCGGCCTCTATTTGAAGATATGAC
TGATTCTGACTGTA 420

164

TTCTCTTCATTGACCAAGGAAATCGGCCTCTATTTGAAGATATGAC TGATTCTGACTGTA 223

55 V-L--F--I--D--Q--G--N--R--P--L--F--E--D--M--T--D--S--D--C--

421

GAGATAAT<mark>GCA</mark>CCC<mark>CGG</mark>ACC<mark>ATA</mark>TTT<mark>ATT</mark>ATA<mark>AGT</mark>ATG<mark>TAT</mark>AAA<mark>G</mark> ATAGCCAGCCTAGAG 480

224

75 R--D--N--A--P--R--T--I--F--I--I--S--M--Y--K--D--S--Q--P--R--94

481

GT<mark>ATG</mark>GCT<mark>GTA</mark>ACT<mark>ATC</mark>TCT<mark>GTG</mark>AAG<mark>TGT</mark>GAG<mark>AAA</mark>ATT<mark>TCA</mark>ACT<mark>C</mark> TCTCCTGTGAGAACA 540

284

GTATGGCTGTAACTATCTCTGTGAAGTGTGAGAAAATTTCAACTC TCTCCTGTGAGAACA 343

95 G--M--A--V--T--I--S--V--K--C--E--K--I--S--T--L--S--C--E--N--114

541 AA <mark>ATT</mark> ATT <mark>TCC</mark> TTT <mark>AAG</mark> GAA <mark>ATG</mark> AAT <mark>CCT</mark> CCT <mark>GAT</mark> AAC <mark>ATC</mark> AAG <mark>G</mark> ATACAAAAAGT <mark>GAC</mark> A 600
344 AAATTATTTCCTTTAAGGAAATGAATCCTCCTGATAACATCAAGG
ATACAAAAAGTGACA 403 115 KIISFKEMNPPDNIKDTKSD 134
601 TCATATTCTTTCAGAGAAGTGTCCCAGGACATGATAATAAGATGC
AATTTGAATCT <mark>TCA</mark> T 660 404
TCATATTCTTTCAGAGAAGTGTCCCAGGACATGATAATAAGATGC AATTTGAATCTTCAT 463
135 IIFFQRSVPGHDNKMQFESS 154
661 CA <mark>TAC</mark> GAA <mark>GGA</mark> TAC <mark>TTT</mark> CTA <mark>GCT</mark> TGT <mark>GAA</mark> AAA <mark>GAG</mark> AGAGACCTTT
TTAAACTCATTTTGA 720 464
CATACGAAGGATACTTTCTAGCTTGTGAAAAAGAGAGAGA
155 SYEGYFLACEKERDLFKLIL 174
721 AAAAAGAGGATGAATTGGGG <mark>GAT</mark> AGA <mark>TCT</mark> ATA <mark>ATG</mark> TTC <mark>ACT</mark> GTT
CAAAACGAAGACTAGC 780
aaaaagaggatgaattgggggatagatctataatgttca <mark>ctgtt</mark> <mark>Caaaacgaagactag</mark> . 582
175 KKEDELGDRSIMFTVQNED*193
781
TATTAAAATTTCATGCCGGGCGCAGTGGCTCACGCCTGTAATCCC AGCCCTTTGGGAGGC 840
841
TGAGGCGGGCAGATCACCAGAGGTCAGGTGTTCAAGACCAGCCT GACCAACATGGTGAAA 900
901
CCTCATCTCTACTAAAAATACAAAAAATTAGCTGAGTGTAGTGAC GCATGCCCTCAATCC 960
961 CAGCTACTCAAGAGGCTGAGGCAGGAGAATCACTTGCACTCCGG
and a second control of the control

AGGTAGAGGTTGTGGT 1020

......

1021	
GAGCCGAGATTGCACCATTGCGCTCTAGCCTGGGCAACAACA	GC
AAAACTCCATCTCAAA 1080	
1081 AAATAAAATAAATAAACAAATAAAAAATTCA	
1115	

Forward Primer: ATGGCTGCTGAACCAGTAGA Tm ~ 61 Reverse Primer: CTGTTCAAAACGAAGACTAG Tm ~57

## 4. PCR Protocol

The PCR provides a means of amplifying DNA sequences and can be used to generate microgram quantities of DNA and can amplify the DNA from a single cell into amounts sufficient for cloning or sequencing.

Tot clothing of sequencing.							
<u>Amount</u>	Component	Final Concentration					
μL	Water	31.5 μL					
μL	10x PCR Buffer (P2317)	1x					
1 μL	Deoxynucleotide Mix	200 μΜ					
μL	Forward primer	0.5 μΜ					
μL	Reverse primer	0.5 μΜ					
0.5 μL	Taq DNA Polymerase (D1806)	0.05 units/μL					
μL	Template DNA (typically 10 ng)	200 pg/μL					
μL	25 mM MgCl <sub>2</sub>	0.5 mM					
	Total reaction volume	50 μL					
Pcr Step	Temperature °C	<u>Duration</u>					
Initial Denaturation	94 °C	4 min					
30 cycles	94°C	1 min					
Anneal Primer	57°C	30 sec					
Final Extention	2°C	5 min					

- 5. The final step is to check the DNA sequence which involves synthesizing DNA sub-fragments of all possible lengths and separating them on a Agarose Gel Electrophoresis and subsequent Ethidium Bromide Staining and now we can read the sequence directly
  - Agarose (precast gels, powder, etc.)
  - Buffer such as MOPS-EDTA-sodium acetate, trisacetate-EDTA (TAE) or tris-borate-EDTA (TBE)
  - Gel loading solution and sample loading buffer for RNA
  - Electrophoresis stain or dye such as ethidium bromide.

We can now separate those fragments into four groups depending on the last base by using Sanger Sequencing Method, this method allow us to generate the fragments of the segment of DNA to be sequenced by using four different dideoxy analogs one for each four bases, allows the generation of four sets fragments in four separate reactions and now the reaction can be read off, starting at the bottom of gel and reading upwards.