

1 **The genetic architecture of target-site**
2 **resistance to pyrethroid insecticides in the**
3 **African malaria vectors *Anopheles gambiae***
4 **and *Anopheles coluzzii***

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16 **Abstract**

17 Resistance to pyrethroid insecticides is a major concern for malaria vector control be-
18 cause these compounds are used in almost all insecticide-treated bed-nets (ITNs), and
19 are also used for indoor residual spraying (IRS). Pyrethroids target the voltage-gated
20 sodium channel (VGSC), an essential component of the mosquito nervous system, but
21 substitutions in the amino acid sequence can disrupt the activity of these insecticides,
22 inducing a resistance phenotype. Here we use Illumina whole-genome sequence data
23 from phase 2 of the *Anopheles gambiae* 1000 Genomes Project (Ag1000G) to provide a
24 comprehensive account of genetic variation in the *Vgsc* gene in mosquito populations

from 13 African countries. In addition to three known resistance alleles, we describe 20 other non-synonymous nucleotide substitutions at appreciable population frequency, and map these variants onto a molecular model of the protein to investigate the likelihood of a pyrethroid resistance phenotype. Thirteen of these novel alleles were found to occur almost exclusively on haplotypes carrying the known L995F *kdr* resistance allele (L1014F in *Musca domestica* codon numbering) and may enhance or compensate for the L995F resistance phenotype. A novel mutation I1527T, adjacent to a predicted pyrethroid binding site, was found in tight linkage with V402L substitutions, similar to a combination of alleles found to cause pyrethroid resistance in several other insect species. We also analysed genetic backgrounds carrying resistance alleles, to determine which alleles have experienced recent positive selection, and to investigate the spread of resistance between species and geographical locations. We describe ten distinct haplotype groups carrying known *kdr* resistance alleles. Five of these groups are localised to a single geographical location, and five include haplotypes from different countries, in one case separated by over 3000 km, providing new information about the potential for the geographical spread of resistance. Markers are identified that could be used to design high-throughput, low-cost genetic assays for tracking the spread of pyrethroid resistance in the field. Our results demonstrate that the molecular basis of target-site pyrethroid resistance in malaria vectors is more complex than previously appreciated, and provide a foundation for the development of new genetic tools for insecticide resistance management.

46 **Introduction**

47 Pyrethroid insecticides have been the cornerstone of malaria prevention in Africa for almost
48 two decades [1]. Pyrethroids are currently used in all insecticide-treated bed-nets (ITNs),
49 and are used in indoor residual spraying (IRS) as well as in agriculture. Resistance to these
50 insecticides is now widespread in malaria vector populations across Africa [2]. The World
51 Health Organization (WHO) has published plans for insecticide resistance management
52 (IRM) that emphasise the need for improvements in both our knowledge of the molecular
53 mechanisms of resistance and our ability to monitor them in natural populations [3, 4].

54 The voltage-gated sodium channel (VGSC) is the physiological target of pyrethroid in-
55 secticides, and is integral to the insect nervous system. The sodium channel protein con-

sists of four homologous domains (DI-IV) each of which comprises six transmembrane segments (S1-S6) connected by intracellular and extracellular loops [5]. Pyrethroid molecules bind to this protein, stabilise the ion-conducting active state, and thus disrupt normal nervous system function, producing paralysis (“knock-down”) and death. However, amino acid substitutions at key positions within the protein alter the interaction with insecticide molecules, increasing the dose of insecticide required for knock-down, known as knock-down resistance or *kdr* [6, 5].

In the African malaria vectors *Anopheles gambiae* and *An. coluzzii*, three substitutions have been found to cause pyrethroid resistance. Two of these substitutions occur in codon 995¹, with L995F prevalent in West and Central Africa [7, 8], and L995S found in Central and East Africa [9, 8]. A third substitution, N1570Y, has been found in West and Central Africa and shown to increase resistance in association with L995F [11]. However, studies in other insect species have found a variety of other *Vgsc* substitutions inducing a resistance phenotype [12, 13, 5]. To our knowledge, no studies in malaria vectors have analysed genetic variation across the full *Vgsc* coding sequence, thus the molecular basis of pyrethroid target-site resistance has not been fully explored.

Basic information is also lacking about the spread of pyrethroid resistance in malaria vectors [3]. For example, it is not clear when, where or how many times pyrethroid target-site resistance has emerged. Geographical paths of transmission, carrying resistance alleles between mosquito populations, are also not known. Previous studies have found evidence that L995F occurs on several different genetic backgrounds, suggesting multiple independent outbreaks of resistance driven by this allele [14, 15, 16, 17]. However, these studies analysed only small gene regions in a limited number of mosquito populations, and therefore had limited resolution to make inferences about relationships between haplotypes carrying this allele. It has also been shown that the L995F allele spread from *An. gambiae* to *An. coluzzii* in West Africa [18, 19, 20, 21]. However, both L995F and L995S now have wide geographical distributions [8], and to our knowledge no attempts have been made to infer or track the geographical spread of either allele across Africa.

Here we report an in-depth analysis of genetic variation in the *Vgsc* gene, using whole-

¹Codon numbering is given here relative to transcript AGAP004707-RD as defined in the AgamP4.12 gene-set annotations. A mapping of codon numbers from AGAP004707-RD to *Musca domestica*, the system in which *kdr* mutations were first described [10], is given in Table 1.

85 genome Illumina sequence data from phase 2 of the *Anopheles gambiae* 1000 Genomes
86 Project (Ag1000G) [22]. The Ag1000G phase 2 resource includes data on nucleotide vari-
87 ation in 1,142 wild-caught mosquitoes sampled from 13 countries, with representation of
88 West, Central, Southern and East Africa, and of both *An. gambiae* and *An. coluzzii*.
89 We investigate variation across the complete gene coding sequence, and report popula-
90 tion genetic data for both known and novel non-synonymous nucleotide substitutions. We
91 then use haplotype data from the chromosomal region spanning the *Vgsc* gene to study
92 the genetic backgrounds carrying resistance alleles, investigate the geographical spread
93 of resistance between mosquito populations, and provide evidence for recent positive se-
94 lection. Finally, we explore ways in which variation data from Ag1000G can be used to
95 design high-throughput, low-cost genetic assays for surveillance of pyrethroid resistance,
96 with the capability to differentiate and track resistance outbreaks.

97 **Results**

98 ***Vgsc* non-synonymous nucleotide variation**

99 To identify variants with a potentially functional role in pyrethroid resistance, we ex-
100 tracted single nucleotide polymorphisms (SNPs) that alter the amino acid sequence of the
101 VGSC protein from the Ag1000G phase 2 data resource [22]. We then computed their
102 allele frequencies among 16 mosquito populations defined by species and country of ori-
103 gin. Alleles that confer resistance are expected to increase in frequency under selective
104 pressure, therefore we filtered the list of potentially functional variant alleles to retain
105 only those at or above 5% frequency in one or more populations (Table 1). The resulting
106 list comprises 23 variant alleles, including the known L995F, L995S and N1570Y resistance
107 alleles, and a further 20 alleles which prior to Ag1000G had not previously been described
108 in anopheline mosquitoes. We reported 12 of these novel alleles in our overall analysis of
109 the 765 samples in the Ag1000G phase 1 data resource [23], and we extend the analyses
110 here to incorporate SNPs which alter codon 531, 697, 1507, 1603 and two tri-allelic SNPs
111 affecting codons 402 and 490.

112 The 23 non-synonymous variants were located on a transmembrane topology map and on
113 a 3-dimensional homology model of the *Vgsc* protein. (Figure 1). The substitutions were

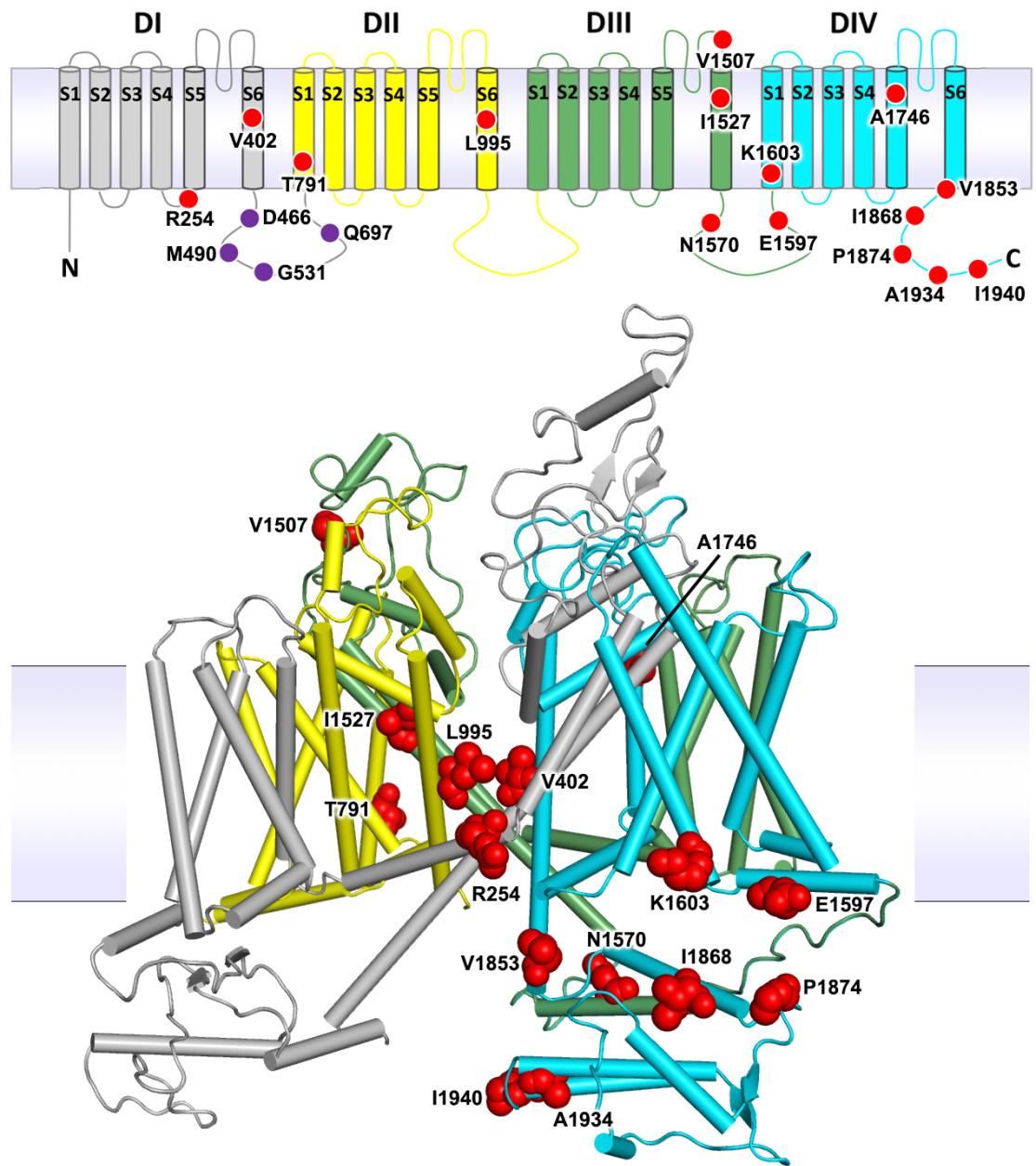


Figure 1. Voltage-gated sodium channel protein structure and non-synonymous variation. The *An. gambiae* voltage-gated sodium channel (AGAP004707-RD AgamP4.12) is shown as a transmembrane topology map (**top**) and as a homology model (**bottom**) in cartoon format coloured by domain. Variant positions are shown as red circles in the topology map and as red space-fill in the 3D model. Purple circles in the map show amino acids absent from the model due to the lack of modelled structure in this region.

114 found to be distributed throughout the channel, in all of the four internally homologous
 115 domains (DI-DIV), in S1, S5 and S6 membrane-spanning segments, in two of the intracellular
 116 loops connecting domains, and in the C-terminal tail. The S5 and S6 segments that
 117 form the central ion-conducting pore of the channel carry six of the eight segment substi-

tutions, including V402 and L995 which have been shown to produce insecticide resistance phenotypes [6, 5, 7, 8, 9]. Two substitutions are located on the DIII-DIV linker including the resistance conferring N1570 [11]. A further six substitutions are found concentrated in the protein's carboxyl tail (C-terminus), including two alternative substitutions at the resistance associated P1874 residue [24]. The DIII-DIV linker and the C-terminus segment interact in the closed-state channel and substitutions are found throughout this intracellular subdomain. Finally, there are four novel substitutions located on the DI-DII intracellular linker, but this region is missing from the model as it was not resolved in the cockroach Na_vPaS structure used as the model template [25].

The two known resistance alleles affecting codon 995 had the highest overall allele frequencies within the Ag1000G phase 2 cohort (Table 1). The L995F allele was at high frequency in populations of both species from West, Central and Southern Africa. The L995S allele was at high frequency among *An. gambiae* populations from Central and East Africa. Both of these alleles were present in *An. gambiae* populations sampled from Cameroon and Gabon. This included individuals with a heterozygous L995F/S genotype (50/297 individuals in Cameroon, 41/69 in Gabon). We calculated empirical p-values for these heterozygous genotype counts using the Dirichlet distribution and 1,000,000 Monte Carlo simulations. In Cameroon p=0.410 of simulations found higher proportions of heterozygous genotypes, however in Gabon this dropped to p=0.005, suggesting there may be a fitness advantage for mosquitoes carrying both alleles in some circumstances.

The N1570Y allele was present in Guinea *An. gambiae*, Ghana *An. gambiae*, Burkina Faso (both species) and Cameroon *An. gambiae*. This allele has been shown to substantially increase pyrethroid resistance when it occurs in combination with L995F, both in association tests of phenotyped field samples [11] and functional tests using *Xenopus* oocytes [26]. To study the patterns of association among non-synonymous variants, we used haplotypes from the Ag1000G phase 2 resource to compute the normalised coefficient of linkage disequilibrium (D') between all pairs of variant alleles (Figure 2). As expected, we found N1570Y in almost perfect linkage with L995F. Of the 20 novel non-synonymous alleles, 13 also occurred almost exclusively in combination with L995F (Figure 2). These included two variants in codon 1874 (P1874S, P1874L), one of which (P1874S) has previously been associated with pyrethroid resistance in the crop pest moth *Plutella xylostella*.

Table 1. Non-synonymous nucleotide variation in the voltage-gated sodium channel gene. AO=Angola; GH=Ghana; BF=Burkina Faso; CI=Côte d'Ivoire; GN=Guinea; GW=Guinea-Bissau; GM=Gambia; CM=Cameroon; GA=Gabon; UG=Uganda; GQ=Bioko; FR=Mayotte; KE=Kenya; *Ac*=*An. coluzzii*; *Ag*=*An. gambiae*. Species status of specimens from Guinea-Bissau, Gambia and Kenya is uncertain [22]. All variants are at 5% frequency or above in one or more of the 16 Ag1000G phase 2 populations, with the exception of 2,400,071 G>T which is only found in the CMAg population at 0.3% frequency but is included because another mutation is found at the same position (2,400,071 G>A) at >5% frequency and which causes the same amino acid substitution (M490I).

Position ¹	Variant			Population allele frequency (%)															
	Ag ²	Md ³	Domain ⁴	AOAc	GHAc	BFAc	CIAc	GNAc	GW	GM	CMAg	GHAg	BFAg	GNAg	GAAg	UGAg	GQAg	FRAg	KE
2,390,177 G>A	R254K	R261	IL45	0.0	0.009	0.0	0.0	0.0	0.0	0.0	0.313	0.0	0.0	0.0	0.203	0.0	0.0	0.0	0.0
2,391,228 G>C	V402L	V410	IS6	0.0	0.127	0.073	0.085	0.125	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,391,228 G>T	V402L	V410	IS6	0.0	0.045	0.06	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,399,997 G>C	D466H	-	LI/II	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.069	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,400,071 G>A	M490I	M508	LI/II	0.0	0.0	0.0	0.0	0.0	0.0	0.031	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.188
2,400,071 G>T	M490I	M508	LI/II	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.003	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,402,466 G>T	G531V	G549	LI/II	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.007	0.0	0.056	0.0	0.0
2,407,967 A>C	Q697P	Q724	LI/II	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.056	0.0	0.0
2,416,980 C>T	T791M	T810	IIS1	0.0	0.009	0.02	0.0	0.0	0.0	0.0	0.0	0.292	0.147	0.112	0.0	0.0	0.0	0.0	0.0
2,422,651 T>C	L995S	L1014	IIS6	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.157	0.0	0.0	0.0	0.674	1.0	0.0	0.0	0.76
2,422,652 A>T	L995F	L1014	IIS6	0.84	0.818	0.853	0.915	0.875	0.0	0.0	0.525	1.0	1.0	1.0	0.326	0.0	0.0	0.0	0.0
2,429,556 G>A	V1507I	-	IIIL56	0.0	0.0	0.0	0.0	0.125	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,429,617 T>C	I1527T	I1532	IIIS6	0.0	0.173	0.133	0.085	0.125	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,429,745 A>T	N1570Y	N1575	LIIV/IV	0.0	0.0	0.267	0.0	0.0	0.0	0.0	0.057	0.167	0.207	0.088	0.0	0.0	0.0	0.0	0.0
2,429,897 A>G	E1597G	E1602	LIIV/IV	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.065	0.062	0.0	0.0	0.0	0.0	0.0
2,429,915 A>C	K1603T	K1608	IVS1	0.0	0.055	0.047	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,430,424 G>T	A1746S	A1751	IVS5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.292	0.141	0.1	0.0	0.0	0.0	0.0	0.0
2,430,817 G>A	V1853I	V1858	COOH	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.542	0.049	0.062	0.0	0.0	0.0	0.0	0.0
2,430,863 T>C	I1868T	I1873	COOH	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.261	0.2	0.0	0.0	0.0	0.0	0.0
2,430,880 C>T	P1874S	P1879	COOH	0.0	0.027	0.207	0.345	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,430,881 C>T	P1874L	P1879	COOH	0.0	0.0	0.073	0.007	0.25	0.0	0.0	0.0	0.0	0.0	0.234	0.475	0.0	0.0	0.0	0.0
2,431,061 C>T	A1934V	A1939	COOH	0.0	0.018	0.107	0.465	0.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2,431,079 T>C	I1940T	I1945	COOH	0.0	0.118	0.04	0.0	0.0	0.0	0.0	0.067	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0

¹ Position relative to the AgamP3 reference sequence, chromosome arm 2L.

² Codon numbering according to *Anopheles gambiae* transcript AGAP004707-RD in geneset AgamP4.12.

³ Codon numbering according to *Musca domestica* EMBL accession X96668 [10].

⁴ Location of the variant within the protein structure. Transmembrane segments are named according to domain number (in Roman numerals) followed by 'S' then the number of the segment; e.g., 'IIS6' means domain two, transmembrane segment six. Internal linkers between segments within the same domain are named according to domain (in Roman numerals) followed by 'L' then the numbers of the linked segments; e.g., 'IL45' means domain one, linker between transmembrane segments four and five. Internal linkers between domains are named 'L' followed by the linked domains; e.g., 'LI/II' means the linker between domains one and two. 'COOH' means the internal carboxyl tail.

149 [24].

150 The abundance of high-frequency non-synonymous variants occurring in combination
151 with L995F is notable for two reasons. First, *Vgsc* is a highly conserved gene, expected
152 to be under strong functional constraint and therefore purifying selection, so any non-
153 synonymous variants are expected to be rare [12]. Second, in contrast with L995F, we did
154 not observe any high-frequency non-synonymous variants occurring in combination with
155 L995S. This contrast was clear when data on all variants within the gene were considered:
156 for haplotypes carrying the L995F allele, the ratio of non-synonymous to synonymous
157 nucleotide diversity π_N/π_S was 20.04 times higher than haplotypes carrying the wild-type
158 allele, but for those carrying L995S π_N/π_S was 0.5 times lower than haplotypes carrying the
159 wild-type allele. These results indicate that L995F has substantially altered the selective
160 regime for other amino acid positions within the protein. Secondary substitutions have
161 occurred and risen in frequency, suggesting that they are providing some further selective
162 advantage in the presence of insecticide pressure.

163 A novel allele, I1527T, was present in *An. coluzzii* from Ghana, Burkina Faso, Côte
164 d'Ivoire and Guinea. Codon 1527 occurs within trans-membrane segment IIIS6, imme-
165 diately adjacent to residues within a predicted binding site for pyrethroid molecules, thus
166 it is plausible that I1527T could alter pyrethroid binding [27, 5]. We also found that the
167 two variant alleles affecting codon 402, both of which induce a V402L substitution, were
168 in strong linkage with I1527T ($D' \geq 0.8$; Figure 2), and almost all haplotypes carrying
169 I1527T also carried a V402L substitution. Substitutions in codon 402 have been found in
170 a number of other insect species and shown experimentally to confer pyrethroid resistance
171 [5]. The species and geographical distribution of the I1527T+V402L alleles suggest they
172 arose in West African *An. coluzzii* and had not spread to other regions or to *An. gambiae*
173 at the time of sampling. The I1527T allele was present at lower frequency than L995F
174 in all of the West African *An. coluzzii* populations. L995F is known to have increased in
175 frequency in West African *An. coluzzii* [28] and thus could be replacing I1527T+V402L
176 in these populations. The four remaining novel alleles, Q697P, G531V and two separate
177 nucleotide substitutions causing M490I, did not occur in combination with any known
178 resistance allele and were almost exclusively private to a single population (Table 1).

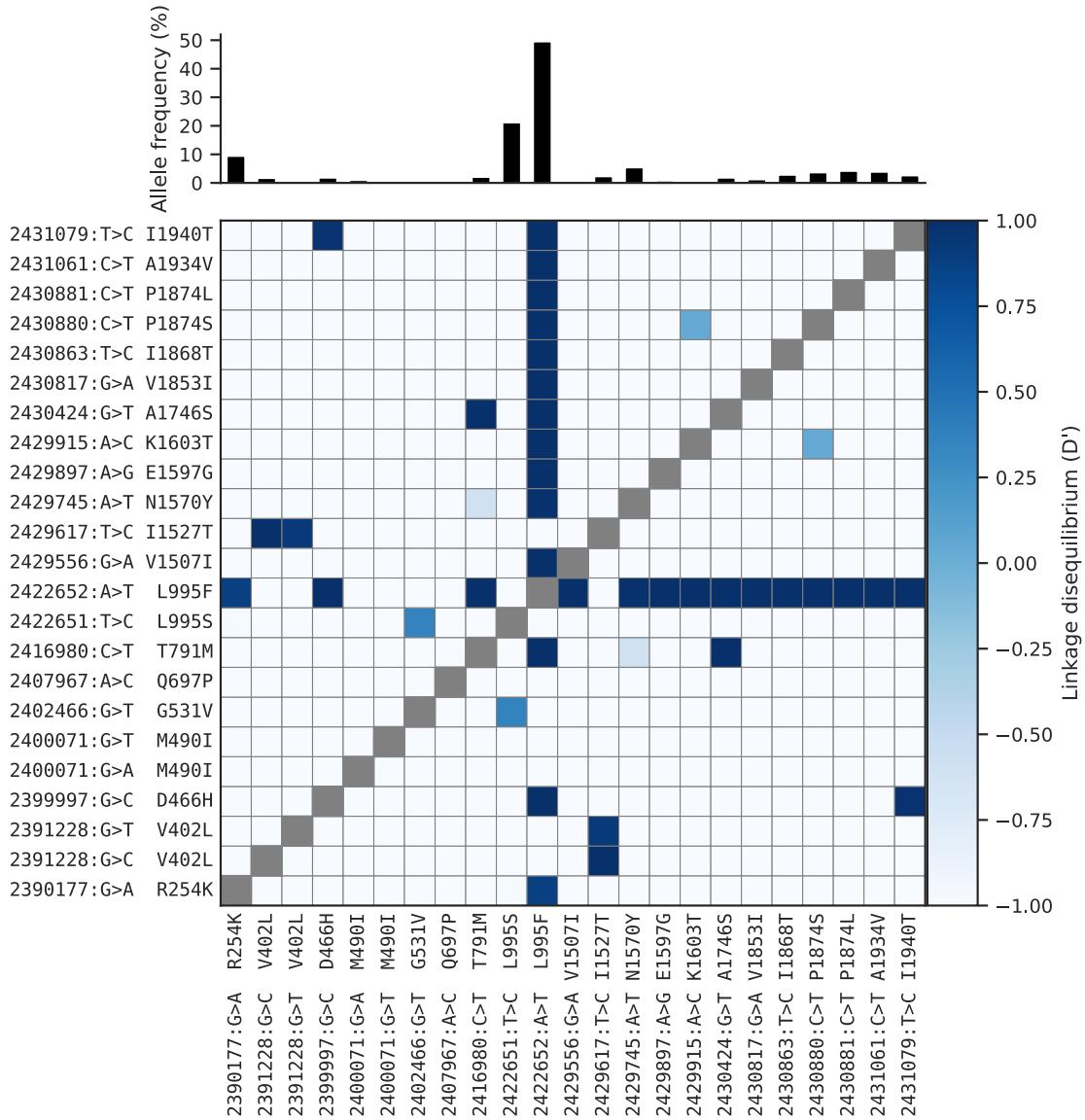


Figure 2. Linkage disequilibrium (D') between non-synonymous variants. A value of 1 indicates that two alleles are in perfect linkage, meaning that one of the alleles is only ever found in combination with the other. Conversely, a value of -1 indicates that two alleles are never found in combination with each other. The bar plot at the top shows the frequency of each allele within the Ag1000G phase 2 cohort. See Table 1 for population allele frequencies.

179 Genetic backgrounds carrying resistance alleles

180 The Ag1000G data resource provides a rich source of information about the spread of
 181 insecticide resistance alleles in any given gene, because data are not only available for
 182 SNPs in protein coding regions, but also SNPs in introns, flanking intergenic regions,
 183 and in neighbouring genes. These additional variants can be used to analyse the genetic
 184 backgrounds (haplotypes) on which resistance alleles are found. In our initial report of

185 the Ag1000G phase 1 resource [23], we used 1710 biallelic SNPs from within the 73.5 kbp
186 *Vgsc* gene (1607 intronic, 103 exonic) to compute the number of SNP differences between
187 all pairs of 1530 haplotypes derived from 765 wild-caught mosquitoes. We then used
188 pairwise genetic distances to perform hierarchical clustering, and found that haplotypes
189 carrying resistance alleles in codon 995 were grouped into 10 distinct clusters, each with
190 near-identical haplotypes. Five of these clusters contained haplotypes carrying the L995F
191 allele (labelled F1-F5), and a further five clusters contained haplotypes carrying L995S
192 (labelled S1-S5).

193 To further investigate genetic backgrounds carrying resistance alleles, we used the
194 Ag1000G phase 2 haplotype data from the *Vgsc* gene (2,284 haplotypes from 1,142 mosquitoes
195 [22]), to construct median-joining networks [29] (Figure 3). The network analysis improves
196 on hierarchical clustering by allowing for the reconstruction and placement of intermedi-
197 ate haplotypes that may not be observed in the data. It also allows for non-hierarchical
198 relationships between haplotypes, which may arise if recombination events have occurred
199 between haplotypes. We constructed the network up to a maximum edge distance of 2 SNP
200 differences, to ensure that each connected component captures a group of closely-related
201 haplotypes. The resulting network contained 5 groups containing haplotypes carrying
202 L995F, and a further 5 groups carrying L995S, in close correspondence with previous re-
203 sults from hierarchical clustering (96.8% overall concordance in assignment of haplotypes
204 to groups).

205 The haplotype network brings into sharp relief the explosive radiation of amino acid sub-
206 stitutions secondary to the L995F allele (Figure 3). Within the F1 group, nodes carrying
207 non-synonymous variants radiate out from a central node carrying only L995F, suggest-
208 ing that the central node represents the ancestral haplotype carrying just L995F which
209 initially came under selection, and these secondary variants have arisen subsequently as
210 new mutations. In F1 alone, 30 network edges (shown as red arrows - Figure 3) lead to
211 non-synonymous nodes. Many of the nodes carrying secondary variants are large, consis-
212 tent with positive selection and a functional role for these secondary variants as modifiers
213 of the L995F resistance phenotype. The F1 network also allows us to infer multiple intro-
214 gression events between the two species. The central (putatively ancestral) node contains
215 haplotypes from individuals of both species, as do nodes carrying the N1570Y, P1874L and

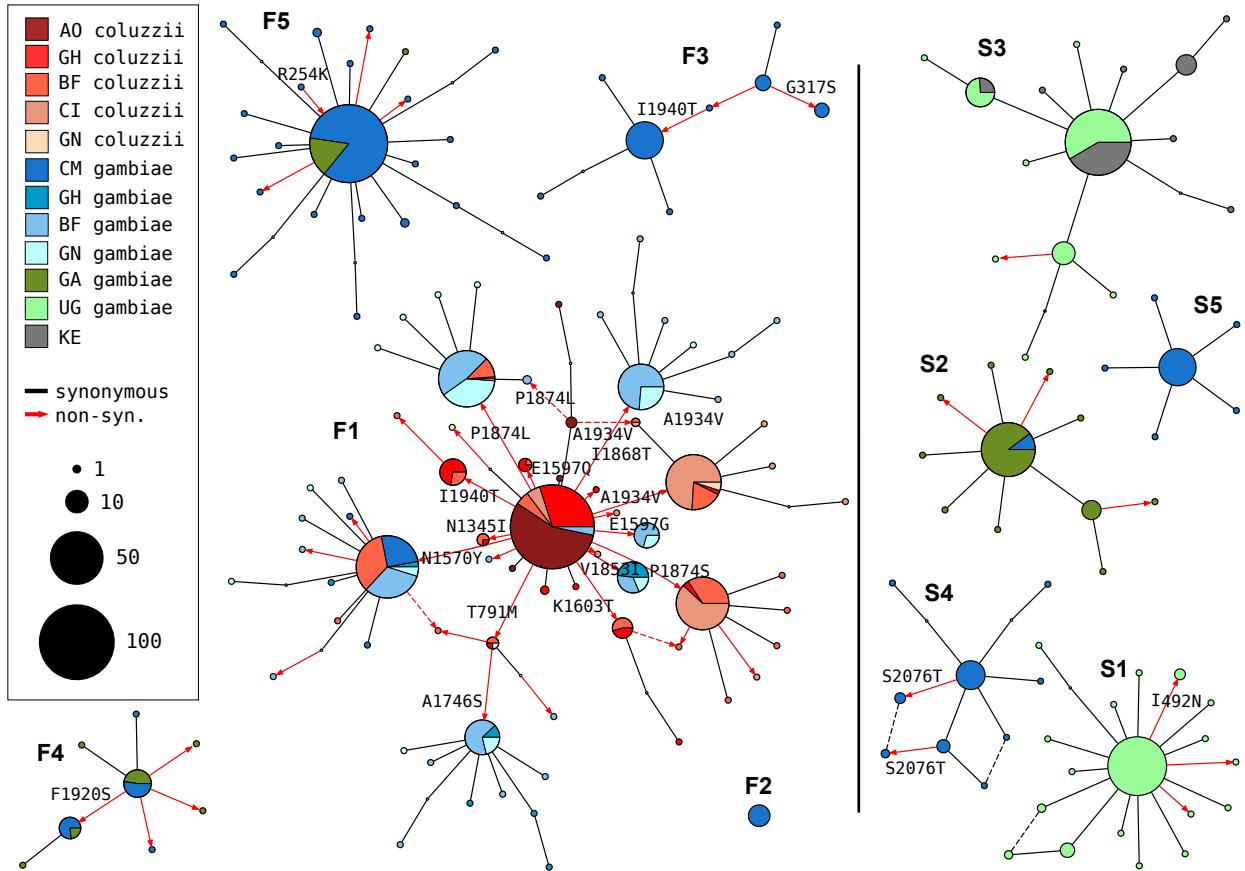


Figure 3. Haplotype networks. Median joining network for haplotypes carrying L995F (labelled F1-F5) or L995S variants (S1-S5) with a maximum edge distance of two SNPs. Labelling of network components is via concordance with hierarchical clusters discovered in [23]. Node size is relative to the number of haplotypes contained and node colour represents the proportion of haplotypes from mosquito populations/species - AO=Angola; GH=Ghana, BF=Burkina Faso; CI=Côte d'Ivoire; GN=Guinea; CM=Cameroon; GA=Gabon; UG=Uganda; KE=Kenya. Non-synonymous edges are highlighted in red and those leading to non-singleton nodes are labelled with the codon change, arrow head indicates direction of change away from the reference allele. Network components with fewer than three haplotypes are not shown.

216 T791M variants. This structure is consistent with an initial introgression of the ancestral
 217 F1 haplotype, followed later by introgressions of haplotypes carrying secondary mutations.
 218 The haplotype network also illustrates the contrasting levels of non-synonymous varia-
 219 tion between L995F and L995S. Within all of the L995S groups, only eight edges lead to
 220 non-synonymous nodes and all these nodes are small (low frequency variants), thus may
 221 be neutral or mildly deleterious variants that are hitch-hiking on selective sweeps for the
 222 L995S allele.

223 The F1 group contains haplotypes from mosquitoes of both species, and from mosquitoes
 224 sampled in six different countries (Angola, Burkina Faso, Cameroon, Côte d'Ivoire, Ghana,

225 Guinea) (Figure 4). The F4, F5 and S2 groups each contain haplotypes from both
226 Cameroon and Gabon. The S3 group contains haplotypes from both Uganda and Kenya.
227 The haplotypes within each of these five groups (F1, F4, F5, S2, S3) were nearly identi-
228 cal across the entire span of the *Vgsc* gene ($\pi < 4.5 \times 10^{-5} \text{ bp}^{-1}$). In contrast, diversity
229 among wild-type haplotypes was two orders of magnitude greater (Cameroon *An. gambiae*
230 $\pi = 1.4 \times 10^{-3} \text{ bp}^{-1}$; Guinea-Bissau $\pi = 5.7 \times 10^{-3} \text{ bp}^{-1}$). Thus it is reasonable to assume
231 that each of these five groups contains descendants of an ancestral haplotype that carried
232 a resistance allele and has risen in frequency due to selection for insecticide resistance.
233 Given this assumption, these groups each provide evidence for adaptive gene flow between
234 mosquito populations separated by considerable geographical distances.

235 Populations carrying *kdr* alleles were collected between the years 2009 and 2012, with the
236 exception of Gabon, which was collected in 2000. This temporal spread allows, albeit with
237 low-resolution, tracking of haplotypes through time. The spatially widespread F1 group
238 contains haplotypes from samples collected between 2009-2012 (Figure 4, [22]) (Figure
239 3). We still do not know how fast insecticide resistance alleles can travel between these
240 countries, but the large geographic spread suggests the F1 haplotype group originated some
241 considerable time before the earliest collection in 2009. Haplotype groups F4, F5 and S2,
242 all carry haplotypes from samples collected in Cameroon (2009) and Gabon (2000). These
243 observations demonstrate that, even in mosquito populations with high levels of genetic
244 diversity and large effective population size [23], nucleotide sequences carrying alleles under
245 strong selection can persist unchanged for almost a decade.

246 A limitation of both the hierarchical clustering and network analyses is that they rely
247 on genetic distances within a fixed genomic window from the start to the end of the
248 *Vgsc* gene. *Anopheles* mosquitoes undergo homologous recombination during meiosis in
249 both males and females, and any recombination events that occurred within this genomic
250 window could affect the way that haplotypes are grouped together in clusters or network
251 components. In particular, recombination events could occur during the geographical
252 spread of a resistance allele, altering the genetic background upstream and/or downstream
253 of the allele itself. An analysis based on a fixed genomic window might then fail to infer
254 gene flow between two mosquito populations, because haplotypes with and without a
255 recombination event could be grouped separately, despite the fact that they share a recent

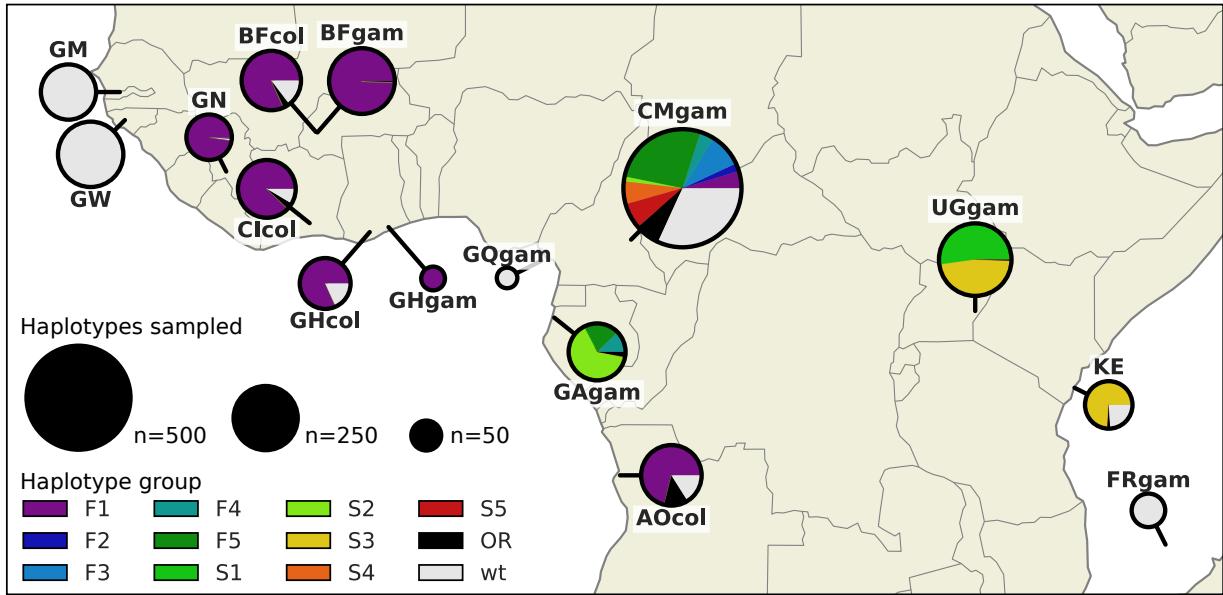


Figure 4. Map of haplotype frequencies. Each pie shows the frequency of different haplotype groups within one of the populations sampled. The size of the pie is proportional to the number of haplotypes sampled. The size of each wedge within the pie is proportional to the frequency of a haplotype group within the population. Haplotypes in groups F1-5 carry the L995F *kdr* allele. Haplotypes in groups S1-5 carry the L995S *kdr* allele. Haplotypes in group other resistant (OR) carry either L995F or L995S but did not cluster within any of the haplotype groups. Wild-type (*wt*) haplotypes do not carry any known resistance alleles.

common ancestor. To investigate the possibility that recombination events may have affected our grouping of haplotypes carrying resistance alleles, we performed a moving window analysis of haplotype homozygosity, spanning *Vgsc* and up to a megabase upstream and downstream of the gene (Supplementary Figures S1, S2). This analysis supported a refinement of our initial grouping of haplotypes carrying resistance alleles. All haplotypes within groups S4 and S5 were effectively identical on both the upstream and downstream flanks of the gene, but there was a region of divergence within the *Vgsc* gene itself that separated them in the fixed window analyses (Supplementary Figure S2). The 13.8 kbp region of divergence occurred upstream of codon 995 and contained 6 SNPs that were fixed differences between S4 and S5. A possible explanation for this short region of divergence is that a gene conversion event has occurred within the gene, bringing a segment from a different genetic background onto the original genetic background on which the L995S resistance mutation occurred.

269 **Positive selection for resistance alleles**

270 To investigate evidence for positive selection on non-synonymous alleles, we performed
271 an analysis of extended haplotype homozygosity (EHH) [30]. Haplotypes under recent
272 positive selection will have increased rapidly in frequency, thus have had less time to be
273 broken down by recombination, and should on average have longer regions of haplotype
274 homozygosity relative to wild-type haplotypes. We defined a core region spanning *Vgsc*
275 codon 995 and an additional 6 kbp of flanking sequence, which was the minimum required
276 to differentiate the haplotype groups identified via clustering and network analyses. Within
277 this core region, we found 18 distinct haplotypes at a frequency above 1% within the cohort.
278 These included core haplotypes corresponding to each of the 10 haplotype groups carrying
279 L995F or L995S alleles identified above, as well as a core haplotype carrying I1527T which
280 we labelled L1 (due to it carrying the the wild-type leucine codon at position 995). We also
281 found a core haplotype corresponding to a group of haplotypes from Kenya carrying an
282 M490I allele, which we labelled as L2. All other core haplotypes we labelled as wild-type
283 (*wt*). We then computed EHH decay for each core haplotype up to a megabase upstream
284 and downstream of the core locus (Figure 5).

285 As expected, haplotypes carrying the L995F and L995S resistance alleles all experience
286 a slower decay of EHH relative to wild-type haplotypes, supporting positive selection.
287 Previous studies have found evidence for different rates of EHH decay between L995F
288 and L995S haplotypes, suggesting differences in the timing and/or strength of selection
289 [16]. However, we found no systematic difference in the length of shared haplotypes when
290 comparing F1-5 (carrying L995F) against S1-5 (carrying L995S) (Supplementary Figure
291 S3). There were, however, some differences between core haplotypes carrying the same
292 allele. For example, shared haplotypes were significantly longer for S1 (median 1.006 cM,
293 95% CI [0.986 - 1.040]) versus other core haplotypes carrying L995S (e.g., S2 median
294 0.593 cM, 95% CI [0.589 - 0.623]; Supplementary Figure S3). Longer shared haplotypes
295 indicate a more recent common ancestor, and thus some of these core haplotypes may
296 have experienced more recent and/or more intense selection than others.

297 As sample collections took place over 12 years (2000-2012), it might be expected that
298 core haplotypes appearing earlier in our sampling would have smaller shared haplotypes

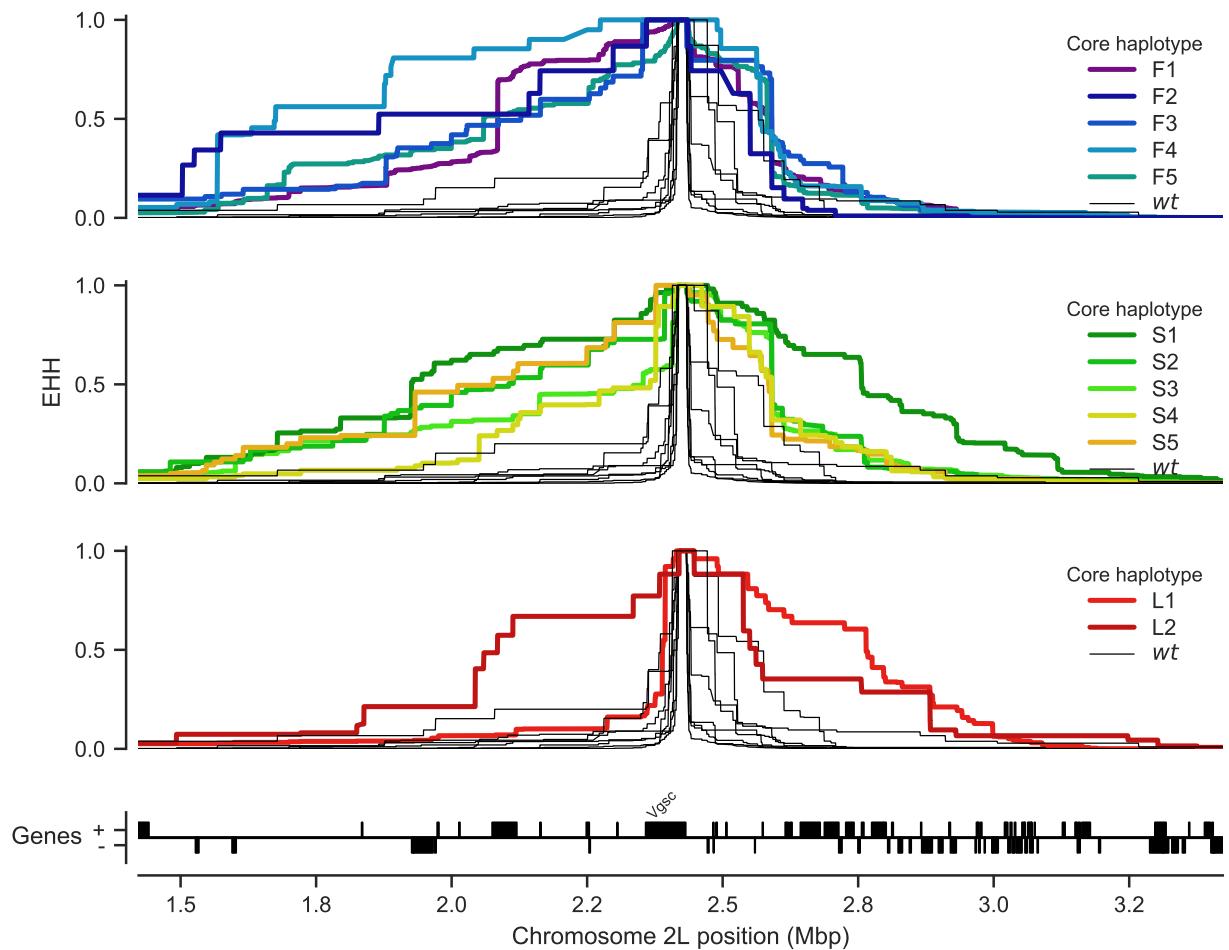


Figure 5. Evidence for positive selection on haplotypes carrying known or putative resistance alleles. Each panel plots the decay of extended haplotype homozygosity (EHH) for a set of core haplotypes centred on *Vgsc* codon 995. Core haplotypes F1-F5 carry the L995F allele; S1-S5 carry the L995S allele; L1 carries the I1527T allele; L2 carries the M490I allele. Wild-type (*wt*) haplotypes do not carry known or putative resistance alleles. A slower decay of EHH relative to wild-type haplotypes implies positive selection (each panel plots the same collection of wild-type haplotypes).

due to increased opportunity for recombination and mutation. However, no correlation was found between the year a core haplotype was first detected and the median length ($r(8)=0.03$, $p=0.93$, Supplementary Figure S3).

The L1 haplotype carrying I1527T+V402L exhibited a slow decay of EHH on the downstream flank of the gene, similar to haplotypes carrying L995F and L995S, indicating that this combination of alleles has experienced positive selection. EHH decay on the upstream gene flank was faster, being similar to wild-type haplotypes, however there were two separate nucleotide substitutions encoding V402L within this group of haplotypes, and a faster EHH decay on this flank is consistent with recombination events bringing V402L alleles

308 from different genetic backgrounds together with an ancestral haplotype carrying I1527T.
309 The L2 haplotype carrying M490I exhibited EHH decay on both flanks comparable to hap-
310 lotypes carrying known resistance alleles. This could indicate evidence for selection on the
311 M490I allele, but these haplotypes are derived from a Kenyan mosquito population where
312 there is evidence for a severe recent bottleneck [23], and there were not enough wild-type
313 haplotypes from Kenya with which to compare. Thus this signal may also be due to the
314 extreme demographic history of this population.

315 **Discussion**

316 **Cross-resistance between pyrethroids and DDT**

317 The VGSC protein is the physiological target of both pyrethroid insecticides and DDT [6].
318 The L995F and L995S alleles are known to increase resistance to both of these insecticide
319 classes [7, 9]. By 2012, over half of African households owned at least one pyrethroid
320 impregnated ITN and nearly two thirds of IRS programmes were using pyrethroids [2].
321 Pyrethroids were also introduced into agriculture in Africa prior to the scale-up of public
322 health vector control programmes, and continue to be used on a variety of crops such as
323 cotton [31]. DDT was used in Africa for several pilot IRS projects carried out during the
324 first global campaign to eradicate malaria, during the 1950s and 1960s [12]. DDT is still
325 approved for IRS use by WHO and remains in use in some locations, however within the
326 last two decades pyrethroid use has been far more common and widespread. DDT was also
327 used in agriculture from the 1940s, and although agricultural usage has greatly diminished
328 since the 1970s, some usage remains [32]. In this study we reported evidence of positive
329 selection on the L995F and L995S alleles, as well as the I1527T+V402L combination and
330 possibly M490I. We also found 14 other non-synonymous substitutions that have arisen in
331 association with L995F and appear to be positively selected. Given that pyrethroids have
332 dominated public health insecticide use for two decades, it is reasonable to assume that the
333 selection pressure on these alleles is primarily due to pyrethroids rather than DDT. It has
334 previously been suggested that L995S may have been initially selected by DDT usage [16].
335 However, we did not find any systematic difference in the extent of haplotype homozygosity
336 between these two alleles, suggesting that both alleles have been under selection over a

similar time frame. We did find some significant differences in haplotype homozygosity between different genetic backgrounds carrying resistance alleles, suggesting differences in the timing and/or strength of selection these may have experienced. However, there have been differences in the scale-up of pyrethroid-based interventions in different regions, and this could in turn generate heterogeneities in selection pressures. Nevertheless, it is possible that some if not all of the alleles we have reported provide some level of cross-resistance to DDT as well as pyrethroids, and we cannot exclude the possibility that earlier DDT usage may have contributed at least in part to their selection. The differing of resistance profiles to the two types of pyrethroids (type I, e.g., permethrin; and type II, e.g., deltamethrin) [33], will also affect the selection landscape. Further sampling and analysis will be required to investigate the timing of different selection events and relate these to historical patterns of insecticide use in different regions.

Resistance phenotypes for novel non-synonymous variants

The non-synonymous variants are distributed throughout the channel protein but can be considered in terms of three clusters: (i) the transmembrane domain, (ii) the DI-II intracellular linker and (iii) the DIII-DIV/C-terminal subdomain. The pyrethroid binding site is located in the transmembrane domain between the IIS4-S5 linker and the IIS5, IIS6 and IIIS6 helices [34]. The I1527T substitution that we discovered in *An. coluzzii* mosquitoes from Burkina Faso occurs in segment IIIS6 and is immediately adjacent to two pyrethroid-sensing residues in this binding site [5]. It is thus plausible that pyrethroid binding could be altered by this substitution. The I1527T substitution (*M. domestica* codon 1532) has been found in *Aedes albopictus* [35], and substitutions in the nearby codon 1529 (*M. domestica* I1534T) have been reported in *Aedes albopictus* and in *Aedes aegypti* where it was found to be associated with pyrethroid resistance [5, 36, 37]. We found the I1527T allele in tight linkage with two alleles causing a V402L substitution (*M. domestica* V410L). Substitutions in codon 402 have been found in multiple insect species and are by themselves sufficient to confer pyrethroid resistance [5]. The fact that we find I1527T and V402L in such tight mutual association is intriguing because haplotypes carrying V402L alone should also have been positively selected and thus be present in one or more populations.

367 The V402 residue is located towards the middle of the IS6 helix. The L995F and L995S
368 substitutions occur at a similar position on the IIS6 helix. It was proposed these S6 sub-
369 stitutions confer resistance by allosterically modifying formation of the pyrethroid binding
370 site [34]. More recently the L995 *kdr* residue was speculated to form part of a second
371 pyrethroid binding site in the insect channel termed 'PyR2' [27, 38]. A major functional
372 effect of the L995F substitution is enhanced closed-state inactivation [39]. This contributes
373 to *kdr* resistance by reducing the number of channels that undergo activation, which is
374 the functional state that pyrethroids bind to with highest affinity [39]. Fast inactivation
375 involves movement of the DIV domain to form a receptor for the DIII-DIV linker fast in-
376 activation particle containing the 'MFM' sequence motif (equivalent to the 'IFM' motif in
377 mammals) [40, 5]. Recent eukaryotic sodium channel structures reveal that the DIII-DIV
378 linker is in complex with the C-terminal segment in the closed-state conformation but the
379 DIII-DIV linker appears to dissociate and bind in close proximity in the DIV S6 helix upon
380 transition to the inactivated state [25, 41]. It seems that binding of the DIII-DIV linker
381 pushes the DIV S6 helix forward to occlude the pore and produce the inactivated state
382 [41]. We suggest that substitutions located on the DIII-DIV linker and C-terminal tail may
383 perturb the conformation of this subdomain when it assembles in the closed-state channel
384 and may subsequently affect capture or release of the DIII-DIV linker from this complex.
385 The expected functional outcome would be altered channel inactivation, although whether
386 inactivation is enhanced or diminished and if this compensates for a deleterious effect of
387 L995F on channel function awaits elucidation. The N1570Y substitution on the DIII-DIV
388 linker has been functionally characterised but inactivation kinetics in the mutant channel
389 were found unaltered [26]. Pyrethroid sensitivity was also unaffected by N1570Y although
390 resistance was greatly enhanced in the N1570Y + L995F double mutant [26].

391 The final cluster of novel variants is located on the DI-DII intracellular linker. This
392 segment includes the novel M490I substitution that was found on the Kenyan L2 haplotypic
393 background potentially under selection. M490I did not occur in association with L995F or
394 any other non-synonymous substitutions. Although we were unable to model this region,
395 we speculate that the DI-DII linker passes under the DII S4-S5 linker and these regions
396 may interact, as was found in a bacterial sodium channel structure [42]. The structural
397 effects of DI-DII substitutions may be altered interactions with the DII S4-S5 linker, the

398 movement of which is critical for formation of the pyrethroid binding site [34, 43]. Overall,
399 there are a number of potential mechanisms by which a pyrethroid resistance phenotype
400 may arise and topology modelling reveals how many of the non-synonymous variants we
401 discover may be involved, though clearly much remains to be unravelled regarding the
402 molecular biology of pyrethroid resistance in this channel.

403 **Design of genetic assays for surveillance of pyrethroid resistance**

404 Entomological surveillance teams in Africa regularly genotype mosquitoes for resistance al-
405 leles in *Vgsc* codon 995, and use those results as an indicator for the presence of pyrethroid
406 resistance alongside results from insecticide resistance bioassays. They typically do not,
407 however, sequence the gene or genotype any other polymorphisms within the gene. Thus,
408 if there are other polymorphisms within the gene that cause or significantly enhance
409 pyrethroid resistance, these will not be detected. Also, if a codon 995 resistance allele is
410 observed, there is no way to know whether the allele is on a genetic background that has
411 also been observed in other mosquito populations, and thus no way to investigate whether
412 resistance alleles are emerging locally or being imported from elsewhere. Whole-genome
413 sequencing of individual mosquitoes clearly provides data of sufficient resolution to answer
414 these questions, and could be used to provide ongoing resistance surveillance. The cost of
415 whole-genome sequencing continues to fall, making it a practical tool for malaria vector
416 surveillance. However, to achieve substantial spatial and temporal coverage of mosquito
417 populations, it would also be necessary to develop targeted genetic assays for resistance
418 outbreak surveillance. Technologies such as amplicon sequencing [44] are already being
419 trialled on mosquitoes [45], these could scale to tens of thousands of samples at low cost
420 and could be implemented using existing platforms in national molecular biology facilities.

421 To facilitate the development of targeted genetic assays for surveillance of *Vgsc*-mediated
422 pyrethroid resistance, we have produced several supplementary data tables. In Supple-
423 mentary Table 1 we list all 82 non-synonymous variants found within the *Vgsc* gene in this
424 study, with population allele frequencies. In Supplementary Table 2 we list 756 biallelic
425 SNPs, within the *Vgsc* gene and up to 10 kbp upstream or downstream, that are poten-
426 tially informative regarding which haplotype group a resistance haplotype belongs to, and
427 thus could be used for tracking the spread of resistance. This table includes the allele

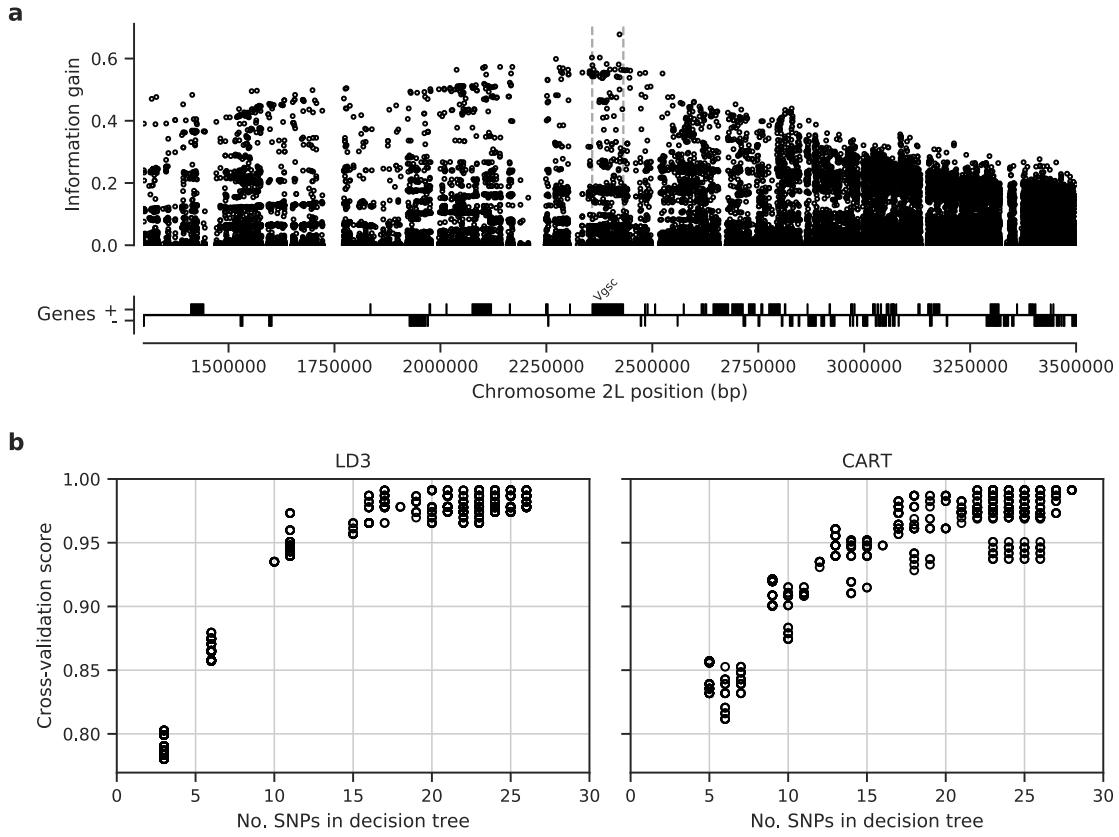


Figure 6. Informative SNPs for haplotype surveillance. **a**, Each data point represents a single SNP. The information gain value for each SNP provides an indication of how informative the SNP is likely to be if used as part of a genetic assay for testing whether a mosquito carries a resistance haplotype, and if so, which haplotype group it belongs to. **b**, Number of SNPs required to accurately predict which group a resistance haplotype belongs to. Each data point represents a single decision tree. Decision trees were constructed using either the LD3 (left) or CART (right) algorithm for comparison. Accuracy was evaluated using 10-fold stratified cross-validation.

frequency within each of the 10 haplotype groups defined here, to aid in identifying SNPs that are highly differentiated between two or more haplotype groups. We also provide Supplementary Table 3 which lists all 10,244 SNPs found within the *Vgsc* gene and up to 10 kbp upstream or downstream, which might need to be taken into account as flanking variation when searching for PCR primers to amplify a SNP of interest. To provide some indication for how many SNPs would need to be assayed in order to track the spread of resistance, we used haplotype data from this study to construct decision trees that could classify which of the 12 groups a given haplotype belongs to (Figure 6). This analysis suggested that it should be possible to construct a decision tree able to classify haplotypes with >95% accuracy by using 20 SNPs or less. In practice, more SNPs would be needed, to provide some redundancy, and also to type non-synonymous polymorphisms in

439 addition to identifying the genetic background. However, it is still likely to be well within
440 the number of SNPs that could be assayed in a single multiplex via amplicon sequenc-
441 ing. Thus it should be feasible to produce low-cost, high-throughput genetic assays for
442 tracking the spread of pyrethroid resistance. If combined with whole-genome sequencing
443 of mosquitoes at sentinel sites, this should also allow the identification of newly emerging
444 resistance outbreaks.

445 Methods

446 Code

447 All scripts and Jupyter Notebooks used to generate analyses, figures and tables are avail-
448 able from the GitHub repository <https://github.com/malariagen/ag1000g-phase2-vgsc-report>.

449 Data

450 We used variant calls and phased haplotype data from the Ag1000G Phase 2 AR1 data re-
451 lease (<https://www.malariagen.net/data/ag1000g-phase-2-ar1>). Variant calls from
452 Ag1000G Phase 2 are also available from the European Nucleotide Archive (ENA; <http://www.ebi.ac.uk/ena>) under study PRJEB36277.

454 Data collection and processing

455 For detailed information on Ag1000G WGS sample collection, sequencing, variant call-
456 ing, quality control and phasing, see [23, 22]. In brief, *An. gambiae* and *An. coluzzii*
457 mosquitoes were collected from 33 sites in 13 countries across Sub-Saharan Africa: An-
458 gola, Bioko, Burkina Faso, Cameroon, Côte d'Ivoire, Gabon, The Gambia, Ghana, Guinea,
459 Guinea Bissau, Kenya, Mayotte and Uganda. From Angola and Côte d'Ivoire just *An.*
460 *coluzzii* were sampled, Burkina Faso, Ghana and Guinea had samples of both *An. gambiae*
461 and *An. coluzzii* and all other populations consisted of purely *An. gambiae*, except for
462 The Gambia, Guinea Bissau and Kenya where species status is uncertain [22]. Mosquitoes
463 were individually whole genome sequenced on the Illumina HiSeq 2000 platform, gener-
464 ating 100bp paired-end reads. Sequence reads were aligned to the *An. gambiae* AgamP3
465 reference genome assembly [46]. Aligned bam files underwent improvement, before variants

466 were called using GATK UnifiedGenotyper. Quality control included removal of samples
467 with mean coverage $\leq 14x$ and filtering of variants with attributes that were correlated
468 with Mendelian error in genetic crosses.

469 The Ag1000G variant data was functionally annotated using the SnpEff v4.1b software
470 [47]. Non-synonymous *Vgsc* variants were identified as all variants in AgamP4.12 transcript
471 AGAP004707-RD with a SnpEff annotation of “missense”. The *Vgsc* gene is known to
472 exhibit alternative splicing [6], however at the time of writing the *An. gambiae* gene
473 annotations did not include the alternative transcripts reported by Davies et al. We wrote
474 a Python script to check for the presence of variants that are synonymous according to
475 transcript AGAP004707-RD but non-synonymous according to one of the other transcripts
476 present in the gene annotations or in the set reported by Davies et al. Supplementary Table
477 1 includes the predicted effect for all SNPs that are non-synonymous in one or more of
478 these transcripts. None of the variants that are non-synonymous in a transcript other
479 than AGAP004707-RD were found to be above 5% frequency in any population.

480 For ease of comparison with previous work on *Vgsc*, pan Insecta, in Table 1 and Supple-
481 mentary Table 1 we report codon numbering for both *An. gambiae* and *Musca domestica*
482 (the species in which the gene was first discovered). The *M. domestica* *Vgsc* sequence
483 (EMBL accession X96668 [10]) was aligned with the *An. gambiae* AGAP004707-RD se-
484 quence (AgamP4.12 gene-set) using the Mega v7 software package [48]. A map of equiva-
485 lent codon numbers between the two species for the entire gene can be download from the
486 MalariaGEN website (https://www.malariagen.net/sites/default/files/content/blogs/domestica_gambiae_map.txt).

488 Haplotypes for each chromosome of each sample were estimated (phased) using using
489 phase informative reads (PIRs) and SHAPEIT2 v2.r837 [49], see [23] supplementary text
490 for more details. The SHAPEIT2 algorithm is unable to phase multi-allelic positions,
491 therefore the two multi-allelic non-synonymous SNPs within the *Vgsc* gene, altering codons
492 V402 and M490, were phased onto the biallelic haplotype scaffold using MVNcall v1.0 [50].
493 Lewontin’s *D'* [51] was used to compute the linkage disequilibrium (LD) between all pairs
494 of non-synonymous *Vgsc* mutations.

495 **Haplotype networks**

496 Haplotype networks were constructed using the median-joining algorithm [29] as imple-
497 mented in a Python module available from <https://github.com/malariagen/ag1000g-phase2-vgsc-report>
498 Haplotypes carrying either L995F or L995S mutations were analysed with a maximum edge
499 distance of two SNPs. Networks were rendered with the Graphviz library and a compos-
500 ite figure constructed using Inkscape. Non-synonymous edges were highlighted using the
501 SnpEff annotations [47].

502 **Positive selection**

503 Core haplotypes were defined on a 6,078 bp region spanning *Vgsc* codon 995, from chro-
504 mosome arm 2L position 2,420,443 and ending at position 2,426,521. This region was
505 chosen as it was the smallest region sufficient to differentiate between the ten genetic
506 backgrounds carrying either of the known resistance alleles L995F or L995S. Extended
507 haplotype homozygosity (EHH) was computed for all core haplotypes as described in [30]
508 using scikit-allel version 1.1.9 [52], excluding non-synonymous and singleton SNPs. Analy-
509 ses of haplotype homozygosity in moving windows (Supplementary Figs. S1, S2) and pair-
510 wise haplotype sharing (Supplementary Figure S3) were performed using custom Python
511 code available from <https://github.com/malariagen/ag1000g-phase2-vgsc-report>.

512 **Design of genetic assays for surveillance of pyrethroid resistance**

513 To explore the feasibility of indentifying a small subset of SNPs that would be sufficient
514 to identify each of the genetic backgrounds carrying known or putative resistance alleles,
515 we started with an input data set of all SNPs within the *Vgsc* gene or in the flanking
516 regions 20 kbp upstream and downstream of the gene. Each of the 2,284 haplotypes in
517 the Ag1000G Phase 2 cohort was labelled according to which core haplotype it carried,
518 combining all core haplotypes not carrying known or putative resistance alleles together as
519 a single "wild-type" group. Decision tree classifiers were then constructed using scikit-learn
520 version 0.19.0 [53] for a range of maximum depths, repeating the tree construction process
521 10 times for each maximum depth with a different initial random state. The classification
522 accuracy of each tree was evaluated using stratified 5-fold cross-validation.

523 **Homology modelling**

524 A homology model of the *An. gambiae* voltage-gated sodium channel (AGAP004707-RD
525 AgamP4.12) was generated using the 3.8 Å resolution structure of the *Periplaneta amer-*
526 *icana* sodium channel Na_vPaS structure (PDB code 5X0M) [25]. Sequences were aligned
527 using Clustal Omega [54]. 50 starting models were generated using MODELLER [55].
528 The internal scoring function of MODELLER was used to select 10 models, which were
529 visually inspected and submitted to the VADAR webserver [56] to assess stereochemistry
530 in order to select the best final model. Figures were produced using PyMOL (DeLano
531 Scientific, San Carlos, CA, USA).

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686 **Supplementary figures**

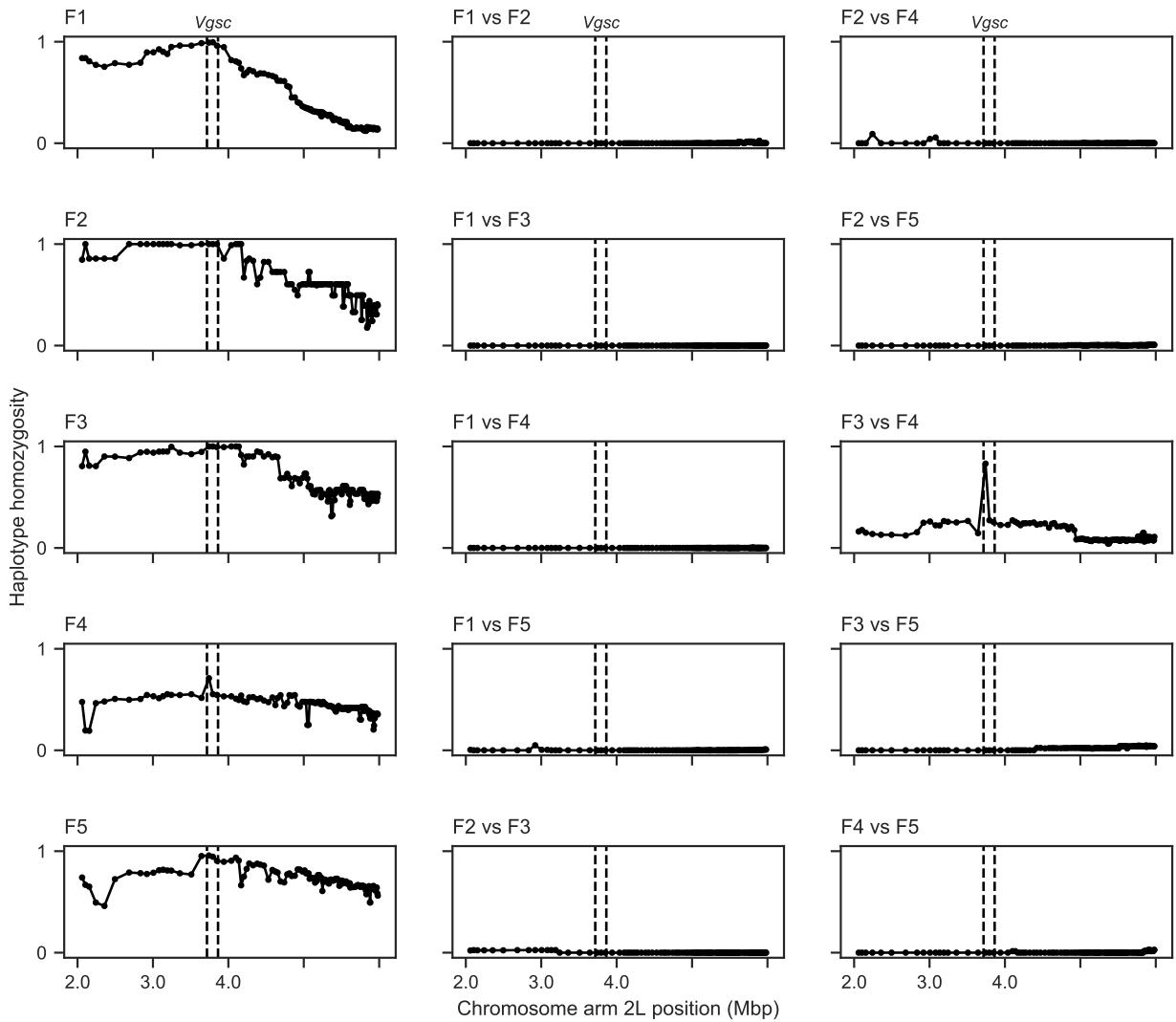


Figure S1. Windowed analysis of haplotype homozygosity for genetic backgrounds carrying the L995F allele. Each sub-plot shows the fraction of haplotype pairs that are identical within half-overlapping moving windows of 1000 SNPs. Each sub-plot in the left-hand column shows homozygosity for haplotype pairs within one of the haplotype groups identified by the network analysis. Sub-plots in the central and right-hand columns show homozygosity for haplotype pairs between two haplotype groups. If two haplotype groups are truly unrelated, haplotype homozygosity between them should be close to zero across the whole genome region. Dashed vertical lines show the location of the *Vgsc* gene.

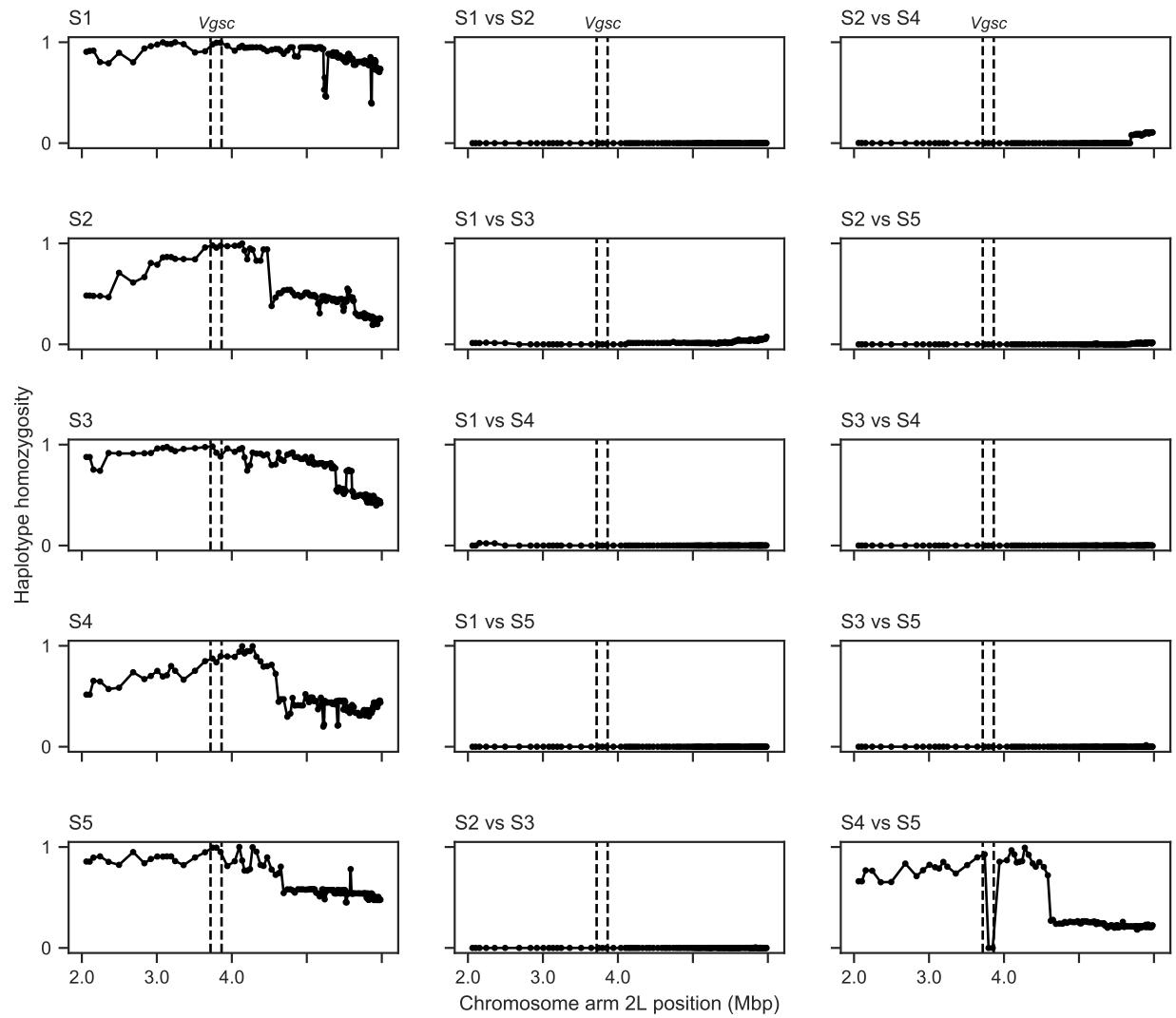


Figure S2. Windowed analysis of haplotype homozygosity for genetic backgrounds carrying the L995S allele. See Supplementary Figure S1 for explanation. Haplotype homozygosity is high between groups S4 and S5 on both flanks of the gene, indicating that haplotypes from both groups are in fact closely related.

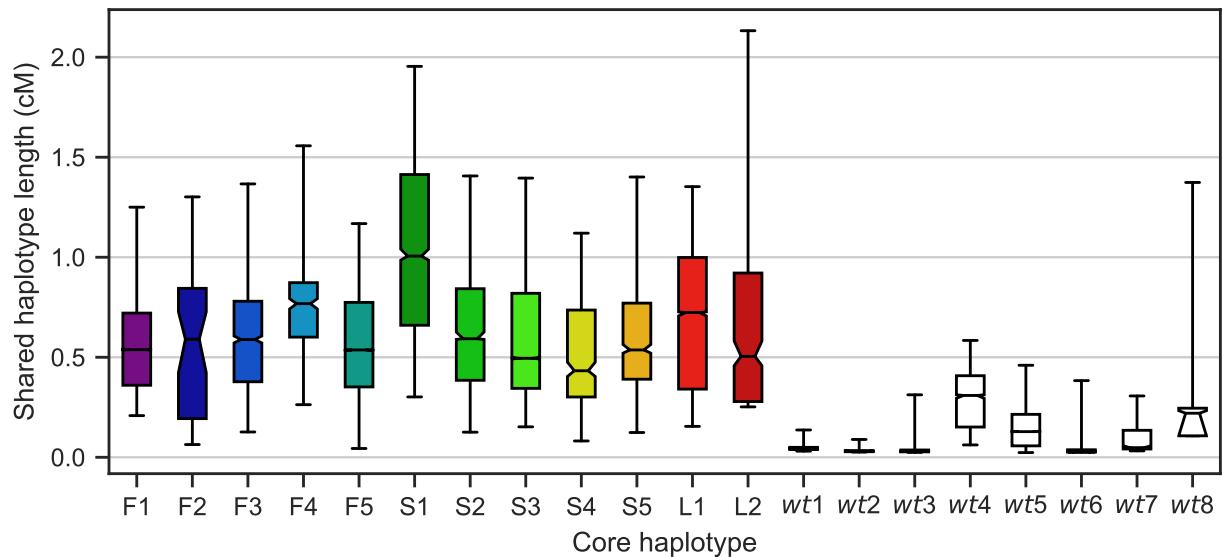


Figure S3. Shared haplotype length. Each bar shows the distribution of shared haplotype lengths between all pairs of haplotypes with the same core haplotype. For each pair of haplotypes, the shared haplotype length is computed as the region extending upstream and downstream from the core locus (*Vgsc* codon 995) over which haplotypes are identical at all non-singleton variants. The *Vgsc* gene sits on the border of pericentromeric heterochromatin and euchromatin, and we assume different recombination rates in upstream and downstream regions. The shared haplotype length is expressed in centiMorgans (cM) assuming a constant recombination rate of 2.0 cM/Mb on the downstream (euchromatin) flank and 0.6 cM/Mb on the upstream (heterochromatin) flank. Bars show the inter-quartile range, fliers show the 5-95th percentiles, horizontal black line shows the median, notch in bar shows the 95% bootstrap confidence interval for the median. Haplotypes F1-5 each carry the L995F resistance allele. Haplotypes S1-5 each carry the L995S resistance allele. Haplotype L1 carries the I1527T allele. Haplotype L2 carries the M490I allele. Wild-type (*wt*) haplotypes do not carry any known or putative resistance alleles.